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RP-HPLC Determination of Atomoxetine Hydrochloride in Bulk and Pharmaceutical Formulations

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Abstract: A reversed phase high performance liquid chromatographic (RP-HPLC) method was developed and subsequently validated for the determination of atomoxetine hydrochloride in bulk and pharmaceutical formulation. The separation was done by a PerkinElmer Brownlee analytical C8 column (260 mm x 4.6 mm, 5 μ m) using methanol: 50 mM KH_2PO_4 buffer (PH adjusted to 6.8 with 0.1 M NaOH), 80:20 v/v as an eluent. UV detection was performed at 270 nm at a flow rate 1.0 mL/min. The validation of the method was performed, and specificity, reproducibility, precision accuracy and ruggedness were confirmed. The correlation coefficient was found to be 0.997 for atomoxetine hydrochloride. The recovery was in the range of 99.94 to 100.98% and limit of quantification was found to be 5.707 $\mu\text{g/mL}$. The method is simple, rapid, selective and economical too and can be used for the routine analysis of drug in pharmaceutical formulations.

Keywords: Atomoxetine hydrochloride, RP-HPLC, Validation

Introduction

Atomoxetine is the first non-stimulant drug approved for the treatment of an attention-deficit hyperactivity disorder (ADHD). It is sold in the form of the hydrochloride salt of atomoxetine. It is a selective nor-adrenaline reuptake inhibitor^{1,2}. Chemically, it is (R)*n*-methyl-3-(2-methylphenoxy)-3-phenylpropylamine³. It is not official in any pharmacopoeias. Literature survey reveals that several methods have been reported like HPLC, HPLC-MS and HPTLC for its estimation in plasma and in capsule dosage form³⁻⁷. The present work reports simple, rapid, sensitive and economical RP-HPLC method with UV detection, useful for the routine analysis of atomoxetine hydrochloride in bulk and pharmaceutical

formulations. The method was validated by parameters such as linearity, accuracy, precision, robustness, stability and system suitability as per ICH guidelines and USP requirements^{8,9}. Suitable statistical tests were performed on validation data^{10,11}. The structure of atomoxetine hydrochloride³ is given in Figure 1.

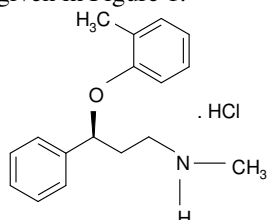


Figure 1. Structure of atomoxetine hydrochloride

Experimental

HPLC system is used for this study, the specifications are given below. Quantitative HPLC was performed on a isocratic high pressure liquid chromatography (PerkinElmer series-2000) equipped with a series 200 LC pump, 200 UV-VIS LC detector (Dual channel), Rheodyne valve (model 7725i) with 20 μ L fixed loop injector, PerkinElmer Brownlee analytical C8 column (260 mm x 4.6 mm, 5 μ m) and a software TotalChrom Workstation (version 6.3.1).

Reagents and chemicals

HPLC grade methanol and triple distilled water were procured from Lichrosolv-E. Merck (India) Ltd., Mumbai. Sodium hydroxide (A. R. grade) and potassium dihydrogen orthophosphate (A. R. grade) were obtained from allied chemical corporation, vadodara. Pure sample of drug was received as gift sun pharmaceuticals Pvt. Ltd., vadodara. Atomoxetine tablets (Axepta-40 mg) and capsules (Attentrol-25 mg) were procured from Intas Pharmaceutical Pvt. Ltd and sun pharmaceutical Pvt. Ltd., vadodara respectively. Triple distilled water was used to prepare all the solutions required for the method.

Preparation of buffer

50 mM KH_2PO_4 buffer was prepared by dissolving 6.8 g of potassium dihydrogen *ortho*- phosphate in 1000 mL of triple distilled water and pH was adjusted to 6.8 with 0.1 M NaOH.

Preparation of atomoxetine hydrochloride (100 μ g/mL) standard stock solution

A 50 mg of standard atomoxetine hydrochloride was accurately weighed and transferred to a 50 mL volumetric flask. It was dissolved in 30 mL mobile phase. The flask was sonicated for 10 min. The flask was shaken and volume was made up to the mark with mobile phase to give a solution containing 1000 μ g/mL atomoxetine hydrochloride. From this solution, 2.5 mL was transferred to 25 mL volumetric flask. The volume was adjusted up to the mark with the mobile phase to give a solution containing 100 μ g/mL atomoxetine hydrochloride.

Determination of atomoxetine hydrochloride in pharmaceutical formulations

Sample preparation of atomoxetine hydrochloride in tablet dosage form

Twenty tablets were weighed and powdered. Powder equivalent to 25 mg of atomoxetine was transferred to 50 mL of volumetric flask containing 30 mL of mobile phase and sonicated for 10 min. The flask was shaken and volume was made up to the mark with mobile phase.

The flask was shaken and volume was made up to the mark with mobile phase to give a solution containing 500 $\mu\text{g/mL}$ atomoxetine Hydrochloride. The resulting solution was filtered through a 0.45 μm membrane filter. 2 mL of aliquot was taken and transferred to volumetric flask of 10 mL capacity and volume was made up to the mark with the mobile phase to give a solution containing 100 $\mu\text{g/mL}$ atomoxetine hydrochloride. From the above solution 2.5 mL of aliquot was taken and transferred to volumetric flask of 10 mL capacity and volume was made up to the mark with the mobile phase to give a solution containing 25 $\mu\text{g/mL}$ atomoxetine hydrochloride. This solution was used for the estimation of atomoxetine.

Sample preparation of atomoxetine hydrochloride in capsule dosage form

Twenty capsules were opened. Powder was taken and weighed. Sample solution was prepared as per the procedure described for sample preparation of amotoxetine hydrochloride tablet.

Chromatographic conditions

The contents of mobile phase were a mixture of methanol and 50 mM KH_2PO_4 buffer (pH adjusted to 6.8 with 0.1 M NaOH) in the ratio of 80:20v/v. It was filtered through 0.45 μm filter paper, sonicated for 10 minutes to degas the mixture. The flow rate of the mobile phase was maintained at 1.0 mL/min. The column temperature was set at 25 ± 1 $^\circ\text{C}$ and the detection was carried out by UV-Detector at a 226 nm wave length. The run time was set at 10 minutes and the volume of injection loop was 20 μL . Prior to the injection of the drug solution, the column was equilibrated for at least 30 minutes with the mobile phase flowing through the system. The data were acquired, stored and analysed with the software TotalChrom Workstation (version 6.3.1).

Calibration procedure

The calibration curve was plotted with five different concentrations (10-50 $\mu\text{g/mL}$) of the standard drug solution. Before injecting the solutions, the column was equilibrated for at least 30 minutes with the mobile phase flowing through the system. Six determinations were carried out for each dilution and graph was plotted for mean peak area response *versus* concentration of the drug. The linearity was evaluated by linear regression analysis.

Results and Discussion

The applied chromatographic conditions permitted a good resolution of atomoxetine hydrochloride in standard solution (A) and in sample solution for tablet (B) and capsule (C) (Figure 2). No drug decomposition was observed during the analysis. The HPLC method was validation for the parameters reported below.

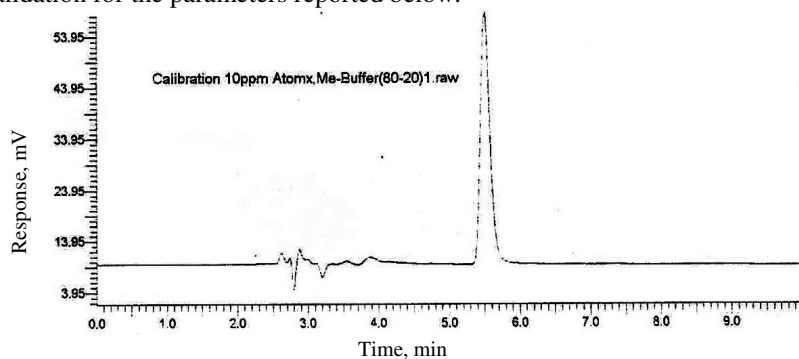


Figure 2. (A) Chromatogram of standard solution containing 10 $\mu\text{g/mL}$ atomoxetine hydrochloride using mobile phase methanol: 50 mM KH_2PO_4 buffer (pH = 6.8), 80: 20% v/v (Proposed Method).

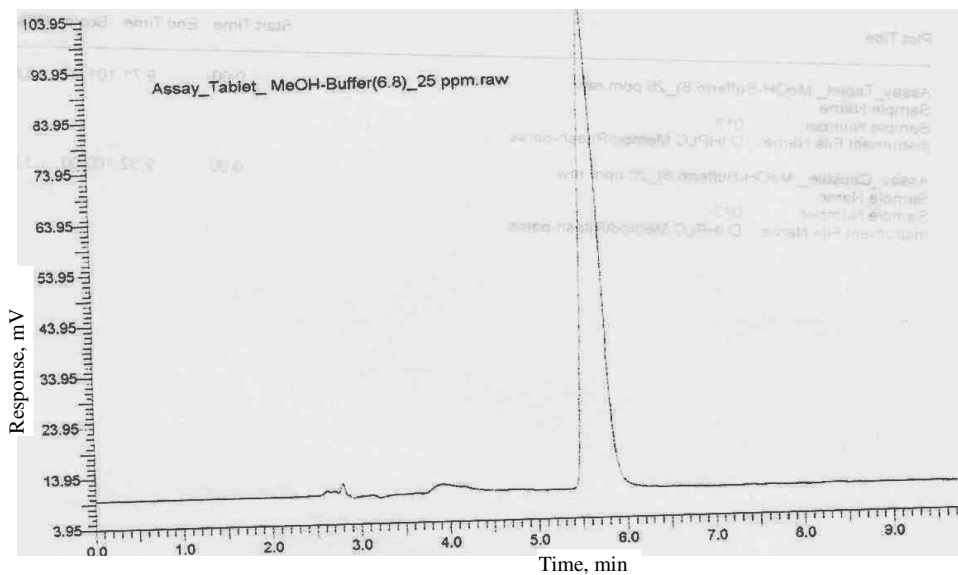


Figure 2. (B) Chromatogram of sample (tablet) solution containing 25 µg/mL atomoxetine hydrochloride using mobile phase methanol: 50 mM KH₂PO₄ buffer (pH = 6.8), 80 : 20% v/v.

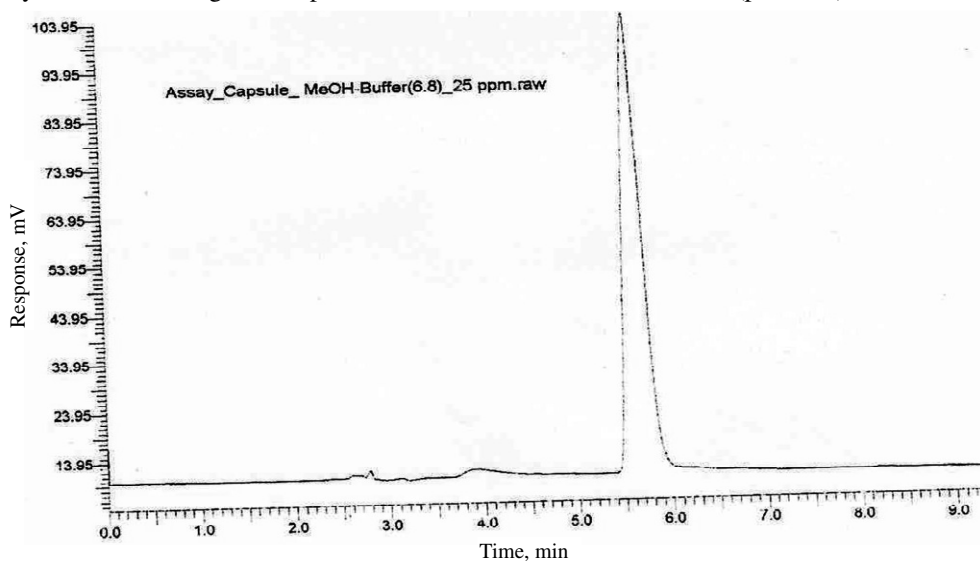


Figure 2. (C) Chromatogram of sample (capsule) solution containing 25 µg/mL atomoxetine hydrochloride using mobile phase methanol : 50 mM KH₂PO₄ buffer (pH = 6.8), 80 : 20% v/v.

Linearity

The atomoxetine hydrochloride was chromatographed using the mobile phase and the linearity of peak area response versus concentration was studied from 10-50 µg/mL. A linear response was observed over the examined concentration range. The results are tabulated in the Table 1.

Table 1. Results of the data analysis for the quantitative determination of atomoxetine hydrochloride by RP-HPLC method

| Statistical parameters | Atomoxetine hydrochloride |
|--|---------------------------|
| Linear range, $\mu\text{g/mL}$ | 10-50 |
| Regression equation | $Y = 198625X + 136739$ |
| Correlation coefficient (r) | 0.997 |
| Limit of detection (LOD), $\mu\text{g/mL}$ | 1.883 |

Accuracy

The accuracy of the RP-HPLC method was assessed by adding a known amount (12.5, 25.0, and 37.5 $\mu\text{g/mL}$) of drug to a drug solution of known concentration (25 $\mu\text{g/mL}$) and subjecting the samples to the proposed method. The known amount of drug (12.5, 25.0 and 37.5 $\mu\text{g/mL}$) was also added to sample solutions of tablet and capsule, each containing 25 $\mu\text{g/mL}$ atomoxetine hydrochloride. The drug was estimated as the procedure described for the estimation of atomoxetine hydrochloride in the tablet and capsule formulations. In all the cases the recovery studies were replicated five times. The accuracy was expressed in terms of the recovery and calculated by multiplying the ratio of measured drug concentration with 100, so as to give the percentage recovery (Table 2 and 3).

Table 2. Recovery of the atomoxetine hydrochloride in tablet dosage form using proposed method.

| Amount of drug added, $\mu\text{g/mL}$ | Amount recovered, $\mu\text{g/mL}$, n=5 | % Recovery, n=5 |
|--|--|-----------------|
| 12.5 | 37.77 | 100.72 |
| 25.0 | 50.49 | 100.98 |
| 37.5 | 62.87 | 100.59 |

Table 3. Recovery of atomoxetine hydrochloride in capsule dosage form using proposed method.

| Amount of drug added, $\mu\text{g/mL}$ | Amount recovered, $\mu\text{g/mL}$, n=5 | % Recovery, n=5 |
|--|--|-----------------|
| 12.5 | 37.61 | 100.29 |
| 25.0 | 49.97 | 99.44 |
| 37.5 | 62.66 | 100.25 |

The method was found to be accurate with percentage recovery 100.59-100.98% and 99.94-100.29% for atomoxetine hydrochloride tablet and capsule respectively.

Precision

The precision of the assay was determined in terms of intra and inter day variation in the peak area for a set of drug solution (20, 30 and 40 $\mu\text{g/mL}$) assayed three times on the same day (intraday) and on three different days (interday). The intraday and interday variation in the peak ratio of the drug solution was calculated in terms of % RSD (Table 4).

Table 4. Interday and intraday precision for atomoxetine hydrochloride

| Atomoxetine hydrochloride concentration, $\mu\text{g/mL}$ | Concentration of atomoxetine hydrochloride ($\mu\text{g/mL}$) found on- | | | |
|---|---|--------|-----------------------|--------|
| | Interday | | Intraday | |
| | Mean area, n=3 | % RSD | Mean area, n=3 | % RSD |
| 20 | 695277.3 \pm 2738.8 | 0.3939 | 694119.8 \pm 1641.1 | 0.2364 |
| 30 | 1155453 \pm 1379.82 | 0.1194 | 1154234 \pm 1295.5 | 0.1122 |
| 40 | 1504489 \pm 2658.26 | 0.1766 | 1504140 \pm 3751.7 | 0.2494 |

The method was found to be precise with % RSD 0.11- 0.39 for inter-day (n=3) and % RSD 0.11- 0.24 for intraday (n=3) for atomoxetine hydrochloride. The method was found to be reproducible. The method was found to be specific as no interference observed when the drug was estimated in presence of excipients.

Robustness

As defined by the ICH, the robustness of an analytical procedure describes to its capability to remain unaffected by small and deliberate variations in method parameters. Robustness was performed by small variation in the chromatographic conditions and found to be unaffected by small variations like $\pm 2\%$ variation in volume of mobile phase composition, ± 0.1 mL/min variation in flow rate of mobile phase and ± 0.1 variation in pH.

Limit of detection and limit of quantification

The parameters LOD and LOQ were determined on the basis of signal to noise ratio. LOD and LOQ were calculated by the method which was based on the standard deviation of the response and the slope (S) of the calibration curve and found to be 1.883 and 5.707 $\mu\text{g/mL}$ respectively (Table 1).

Ruggedness

The method was also found to be rugged as there was no change in absorbance up to 48 h of preparation of solution in methanol.

Reproducibility

The method was found to be reproducible as there was no significant difference when sample concentration (25 $\mu\text{g/mL}$) was estimated using two different instruments, result of *t*-test obtained was 1.74 (< 4.30) (Table 5).

Table 5. Reproducibility data for atomoxetine hydrochloride (25 $\mu\text{g/mL}$)

| Instrument 1 Peak area \pm S.D (n=3) | Instrument 2 Peak area \pm S.D (n=3) | Result of <i>t</i> -test * | Inference |
|---|---|----------------------------|---------------------------|
| 902084.06 \pm 1062.06 | 907530.83 \pm 886.98 | 1.74 | No significant difference |

*At 95% confidence interval, (*t*-Tabulated = 4.3)

Repeatability

The repeatability for atomoxetine hydrochloride was determined by taking atomoxetine hydrochloride sample (tablet) solution (25 $\mu\text{g/mL}$), assayed six times (n=6) and RSD was found to be 0.0036 (Table 6).

Table 6. Repeatability of atomoxetine hydrochloride sample (tablet) solution (25 $\mu\text{g/mL}$)

| Concentration of atomoxetine hydrochloride solution | 25 $\mu\text{g/mL}$ |
|---|---------------------|
| Mean Peak Area | 901079.1 |
| Std. Dev. | 33.17 |
| RSD | 0.0036 |

Assay for marketed dosage forms *i.e.* tablet and capsule was also performed (Table 7). The tablet (Axepta) and capsule (Attentrol) were found to contain 98.99% and 100.45% of the labelled amount respectively.

Table 7. Assay results of marketed dosage forms

| Formulations | Actual concentration | Amount obtained | % purity |
|---------------------|----------------------|------------------|----------|
| | $\mu\text{g/mL}$ | $\mu\text{g/mL}$ | |
| Tablet (Axepta) | 25 | 24.69 | 98.99 |
| Capsule (Attentrol) | 25 | 25.33 | 100.45 |

Conclusion

The proposed reversed phase high performance liquid chromatography method has been evaluated over the linearity, precision, accuracy, reproducibility, robustness, ruggedness and proved to be convenient and effective for the routine analysis of atomoxetine hydrochloride in pharmaceutical formulations. The measured signal was shown to be precise, accurate and linear over the concentration range tested (10-50 $\mu\text{g/mL}$) with a correction coefficient of 0.997. The recovery was in the range of 99.94-100.98% for atomoxetine hydrochloride. Limit of quantification for atomoxetine hydrochloride was found to be 5.707 $\mu\text{g/mL}$. Thus, the proposed method is simple, rapid, accurate, selective, repeatable and requires simple sample preparation procedure. Moreover, an utilisation of methanol: buffer (80: 20) as a mobile phase makes this method cost effective.

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