

Review Article

Commonalities in the Association between PPARG and Vitamin D Related with Obesity and Carcinogenesis

Borja Bandera Merchan,¹ Francisco José Tinahones,^{1,2} and Manuel Macías-González^{1,2}

¹Unidad de Gestión Clínica Endocrinología y Nutrición, Instituto de Investigación Biomédica de Málaga (IBIMA), Complejo Hospitalario de Málaga (Virgen de la Victoria), Universidad de Málaga, 29010 Malaga, Spain ²CIBER Pathophysiology of Obesity and Nutrition (CB06/03), 28029 Madrid, Spain

Correspondence should be addressed to Francisco José Tinahones; fjtinahones@hotmail.com and Manuel Macías-González; mmacias.manuel@gmail.com

Received 25 March 2016; Accepted 15 May 2016

Academic Editor: Daniele Fanale

Copyright © 2016 Borja Bandera Merchan et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The PPAR nuclear receptor family has acquired great relevance in the last decade, which is formed by three different isoforms (PPAR α , PPAR β/δ , and PPAR Y). Those nuclear receptors are members of the steroid receptor superfamily which take part in essential metabolic and life-sustaining actions. Specifically, PPARG has been implicated in the regulation of processes concerning metabolism, inflammation, atherosclerosis, cell differentiation, and proliferation. Thus, a considerable amount of literature has emerged in the last ten years linking PPARG signalling with metabolic conditions such as obesity and diabetes, cardiovascular disease, and, more recently, cancer. This review paper, at crossroads of basic sciences, preclinical, and clinical data, intends to analyse the last research concerning PPARG signalling in obesity and cancer. Afterwards, possible links between four interrelated actors will be established: PPARG, the vitamin D/VDR system, obesity, and cancer, opening up the door to further investigation and new hypothesis in this fascinating area of research.

1. Introduction

There are three subtypes of PPARG, known as PPARG1, PPARG2, and PPARG3. It has been established that PPARG2 leads in potency as a transcription factor [1]. PPARG performs its functions mainly through PPARG1 and PPARG2 [2]. Moreover, it shares lots of additional features with its other counterparts. Concerning that, the parallelism found between the PPARG system and the vitamin D/vitamin D receptor (VD/VDR) system will be further explored later on.

In order to modulate gene expression, the PPAR NRs family, and specifically the PPARG, after binding with either natural or synthetic ligands, heterodimerizes with the Retinoid X Receptor (RXR) as vitamin D receptor (VDR) does.

Later on, the complex PPARG-RXR translocates to the nucleus in order to get attached to PPREs (PPAR Response Elements), genome nucleotides sequences wherefrom the PPARs will coordinate the expression or repression of some genes involved in metabolism, immunity, differentiation, or cellular proliferation, to cite some [3–6].

Once in the nucleus, several molecules known as corepressors and coactivators, which show histone modifying activities by themselves [7], bind the PPARG-RXR complex, showing some control over the genetic expression-repression interplay. Some known corepressors are SMRT or NCOR. When it comes to coactivators, we can mention p300/CRRBbinding protein (CBP) or SRC/p160 [8]. Importantly, differential recruitment of coactivators implies different gene expression patterns [9], wherefrom it can be deduced that the corepressors and coactivators comprise another gene expression regulatory point which is worth studying. PPREs are normally found in the promoter of those genes, which is regulated by PPARG activity [3]. The direct nucleotide sequences which PPARG-RXR will be bound to are known as DR-1 motifs (direct hexanucleotide repeats) of PPRE [8]. Some PPARG target genes are those codifying CD36, FABP4 (Fatty Acid Binding Protein 4), adiponectin, or the CCAAT/enhancer binding protein α [10], all being genes involved in adipose tissue homeostasis. However, afar of its

adipose functions PPARG is also vital for development of some important organs such as heart and the placenta [11].

2. The PPARG Physiology

PPARG behaves as a transcription factor, as many other nuclear receptors (NRs) do. Then, it modulates the expression and repression of a myriad of genes involved in metabolic homeostasis, regulating energy expenditure and storage [12, 13]. Some PPARG target genes are those codifying CD36, FABP4 (Fatty Acid Binding Protein 4), adiponectin, or the CCAAT/enhancer binding protein α [10], all being genes involved in adipose tissue homeostasis. However, afar of its adipose functions PPARG is also vital for development of some important organs such as heart and the placenta [11]. Although most research on PPARG has been focused on its metabolic action, some of them are neurogenesis, osteogenesis, cancer, or cardiovascular disease [14]. Such pleiotropism of actions gives us a clue of the relevance of this transcription factor regarding health and disease. We know for instance that universal PPARG deletion and life are not compatible [11].

The considerable host of actions performed by PPARG can be compared to those of vitamin D and VDR [15], which has been implicated in neurologic disorders [16–18], autoimmune pathologies [19–21], cardiovascular disease [22], diabetes mellitus [23, 24], psoriasis [15] or infectious disease [25, 26], and, above all of what is mentioned, cancer [27, 28].

3. PPARG and Obesity

Much has been already written about PPARG signalling and its role in conditions such as obesity or diabetes. In obesity, PPARG orchestrates adipocyte maturation and differentiation, harmonising the role of many other players in that process [29]. Remarkably, it is the only known factor, which is completely necessary and sufficient for the adipocyte differentiation process to occur [11, 30]. This nuclear receptor acts, then, as a master regulator of adipogenesis.

In addition, it is widely known that PPARG has an important whole-body insulin-sensitizer role. For example, muscle-PPARG knocked-out mice are insulin resistant [31]. In adipose tissue, PPARG deletion leads to increases in bone mass, lipoatrophy, and insulin resistance (IR) [32]. In the same fashion, PPARG induces the proliferation of adipocytes progenitors into mature adipocytes and diminishes the osteoblasts population likewise [33].

The specific deletion of PPARG in liver conduces to IR and decrease of hepatic fat depots [34]. Even in macrophages, the presence of PPARG is important to keep adequate insulin sensitivity levels throughout the body [35, 36]. It is then easy to deduce that one of the main objectives of PPARG activity is the insulin sensitivity maintenance through different tissues.

Thiazolidinediones (TZD), a family of synthetic PPARG agonist widely used in diabetes treatment, show clear improvements in insulin sensitivity, enhanced adipocyte differentiation, reduction of leptin levels, and upregulation of adiponectin [37].

Contrary to the catabolic actions elicited by the PPAR α and PPAR δ , the PPARG is in charge of anabolic functions. As

we have already addressed, adipogenesis and lipid storage are some of them. Illustrating this, a high-fat feeding augments PPARG expression while fasting diminishes it [38].

Remarkably, PPARG performs different functions in metabolically sick rodents and metabolically healthy ones. In disease, PPARG activation seems to improve metabolic parameters, but in the healthy population its downregulation shows antiobesity effects [39].

In the same way, more different effects have been described in metabolic health and disease regarding PPARG expression. For instance, in healthy subjects a high-fat meal greatly induced the expression of PPARG while the same high-fat feeding diminished PPARG expression in a group of morbidly obese patients [40].

In like manner, an indirect correlation between IR and PPARG expression, measured by glucose status, HOMA-IR index, and insulin levels, can be set in morbidly obese persons, whose visceral adipose and muscle tissues show less PPARG expression as IR increases [40].

During placentation and intrauterine development, the PPARG gene methylation patterns could be altered by maternal nutrition, which actually exerts long-term effects upon the receptor status in the offspring, as indicated very recently by Lendvai et al. [41]. This is preliminary evidence about the early programming of our lifelong metabolism set points through nutritional inputs, which could easily leave us susceptible to obesity and metabolic disease in later stages of life.

4. PPARG and Cancer

PPARG is highly expressed in lung, prostate, colorectal, bladder, and breast tumours [42]. Furthermore, we can find in the literature compelling evidence for PPARG having antineoplastic actions in colon, prostate, breast, and lung cancers [43, 44], which happen to be the most prevalent forms of cancer in occident (Figure 1).

Solid evidence backs up that epigenetic events frequently found in cancer can hamper nuclear receptors responsiveness toward their ligands. In that respect, increased levels of corepressor NCOR in prostate cancer can silence the expression of target genes and constitute a potential epigenetic lesion, which selectively distorts the actions of PPARG/PPAR α [45].

In the same line, PPARG promoter methylation in colorectal carcinoma (CRC) is associated with poor prognosis [46]. This transcriptional silencing of PPARG is operated through HDAC1 (Histone Deacetylase 1), EZH2 (Enhancer of Zeste 2 Polycomb Repressive Complex 2 Subunit), and MeCP2 (Methyl CpG Binding Protein 2) recruitment, leading to repressive chromatin states that eventually increase cell proliferation and invasive potential [46]. Correspondingly, APC^{min/+} mice which have undergone PPARG genetic ablation demonstrate increased colon tumour growth [47].

In the literature, some mutations and variations in PPARG expression have been associated with cancer in our specie [48, 49]. Beyond that, its expression comprises an independent prognostic factor in CRC [50, 51].

Apart from epigenetics, we should not lose sight of the fact that metabolic syndrome, insulin resistance, obesity,

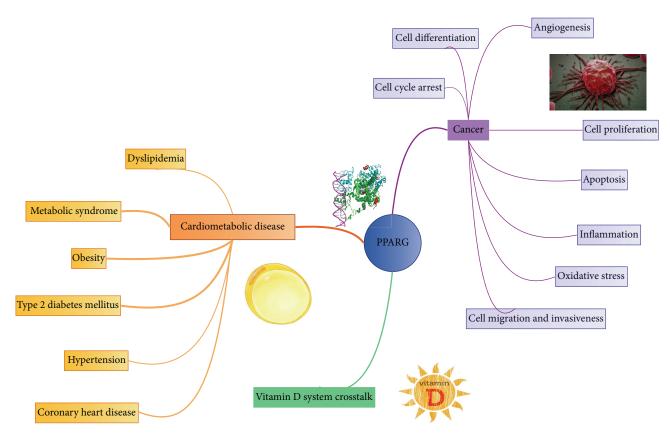


FIGURE 1: PPARG actions: PPARG plays an important role in cardiometabolic disease and cancer. The noteworthy crosstalk between vitamin D system and PPARG is also considered. Arrow's width exemplifies the level of consistency found in the literature regarding each association in the mind picture.

and inflammation, importantly interrelated conditions in which PPARG has modifying and regulatory actions, increase cancer risk [52–59], which adds weight to PPARG and cancer research (Figure 1).

There is some evidence linking PPARG agonist's actions to better cancer treatment responsiveness as well. PPARG agonist Rosiglitazone, in this phase II clinical trial, raised the radioiodine uptake in differentiated thyroid cancer [60].

IFN- β treated pancreatic cancer cells were more affected when Troglitazone was added to the therapy, showing synergistic effects between IFN- β and TGZ [61]. But it is necessary to be careful in some studies, in which PPARG agonist like Rosiglitazone acts as a great promoter of hydroxybutyl nitrosamine-induced urinary bladder cancers [62].

In the following paragraphs, we will review what we know about the specific molecular actions of PPARG in cancer biology. Cell cycle arrest, cell differentiation, angiogenesis, proliferation, invasiveness, migration capacity, apoptosis, inflammation, and oxidative stress should be evaluated.

4.1. Cell Cycle Arrests. Some evidence suggests that PPARG and its agonists have the ability to interfere with the cellular cycle and then, likely, with malignancies development.

In renal cell carcinoma, Troglitazone (TGZ) was able to induce G2/M cell cycle arrest via activation of p38 MAPK (Mitogen-Activated Protein Kinase) [63].

In human pancreatic cancer cells the same phenomenon is observed: PPARG is able to trigger cell cycle arrest of the malignant cells through activation by thiazolidinediones [64].

Through PPARG activation, its ligands increase the expression of the cyclin-dependent kinase inhibitors p21 [64, 65] and p27 [65–69], enhance the turnover of β -catenin, and downregulate the expression of cyclin D1 [70–74].

4.2. Differentiation. In vitro activation of PPARG by its ligands correlates with increased expression of carcinoembryonic antigen (CEA), E-cadherin, developmentally regulated GTP-binding protein 1 (DRG), alkaline phosphatase, or keratins, all of them being molecules expressed in well differentiated cells, opposing to the undifferentiated cell state commonly found in most cancers [48, 64, 75–77].

Tontonoz et al. gave us the first evidence about the effectiveness of PPARG ligands inducing differentiation in human cancer cells, concretely in liposarcoma cancer cells [75]. Again, in human liposarcoma, treatment with Troglitazone raised the level of differentiation of its cells [78].

More evidence that PPARG enhances terminal differentiation in cells is reviewed in papers of Grommes et al. and Koeffler, respectively [43, 44].

4.3. Angiogenesis. It is common knowledge that angiogenesis is a vital step in malignant development. The complex

process by which new vessels are formed, angiogenesis, has been feverishly studied as a new possible target in cancer treatment.

In vitro and in vivo angiogenesis-modulating functions have been described for PPARG [79]. In spite of that, differential effects regarding angiogenesis have been observed for PPARG in vitro and in vivo, showing either pro- or antiangiogenic actions dependent on cell context [80–85]. PPARG agonist can also enhance VEGF expression in cancer cells, as some studies reveal [86, 87].

The mechanisms deciding whether PPARG will act as a proangiogenic factor or as an antiangiogenic one are still elusive to us, but we believe that cellular context and environment are likely the controllers of such process.

4.4. *Proliferation*. Antiproliferative actions are also attributed to PPARG and its ligands. TZD, for example, has shown antiproliferative effects [88, 89].

Modulation of PPARG can have differential effects on carcinogenesis depending on the cellular microenvironment [90]. Therefore, depending on the cellular environment PPARG can behave as a proliferative or antiproliferative factor, as happened with angiogenesis.

Tumour cells are frequently in shortage of polyunsaturated fatty acids. Docosahexaenoic acid (DHA), a well-known ligand of the PPAR family, has been shown to reduce tumour proliferation in lung tumour cell cultures [91]. Along with that, DHA in breast cancer cells diminishes proliferation and increases apoptosis [92, 93].

In prostate cancer, PPARG ligand activation effect was assessed in a phase II clinical trial. The results showed a hampered cancer cell growth [94].

Eukaryotic initiation factor 2 is a target of inhibition for PPARG agonists (i.e., thiazolidinediones). Such factor inhibition, which is mediated in a PPARG-independent way, truncates the translation process [95].

In liposarcoma patients, treatment with Rosiglitazone increased the necessary time to double tumour volume in this clinical trial [96]. In other studies, however, Troglitazone (another member of the thiazolidinedione family) had low or no effects in prostate cancer [97] or breast or colorectal cancer [98, 99].

4.5. Apoptosis. The combined effect of an RXR agonist and Troglitazone curtailed gastric cancer cells proliferation in vitro by enhancing apoptotic mechanisms [100].

PPARG agonists increased the expression of PTEN [101– 105], BAX, BAD [106, 107], and the turnover of the FLICE inhibitory protein (FLIP) [108, 109], known for its antiapoptotic role.

Conversely, PPARG agonists can inhibit BCL-X_L and BCL-2 expression [107, 110], PI3K activity, and AKT phosphorylation [101, 111, 112] and restrain the activation of JUN N-terminal protein kinase [107]. It is worth mentioning that many of those actions were elicited in a PPARG-independent manner. The exact mechanisms by which these effects are performed are still unknown.

4.6. Inflammation. Nowadays, it is common knowledge in the scientific community that chronic inflammation promotes

cancer. The milieu found in chronic inflammation acts as a facilitator for carcinogenesis and cancer development [52, 113]. This has been shown in colorectal, liver, bladder, lung, and gastric neoplasms [114, 115] and investigated in several more. The range of processes in which inflammation partakes in carcinogenesis goes from cell growth and survival, metastasis and cell invasion, treatment response, angiogenesis, and tumour immunity [115, 116].

There is evidence of PPARG having anti-inflammatory activity in several cell lines [117, 118]. In models of experimentally induced colitis PPARG expressed in macrophages is capable of inhibiting inflammation [119].

It is widely known that some PPAR ligands such as omega-3 fatty acids EPA and DHA have anti-inflammatory properties. Those and other natural and synthetic ligands could be used in the future as chemopreventive agents in a vast range of conditions linked to inflammation, that is, cancer [105, 120, 121].

Activation of PPARG by its ligands reduces cytokines such as TNF α and NF- $\kappa\beta$ in monocytes, turning down the inflammatory milieu [120, 122].

The epigenetic process of sumoylation has been linked to PPARG transrepression of inflammation. After ligand activation, PPARG binds to a SUMO protein (Small Ubiquitinlike Modifier) and both join a nuclear corepressor complex, reducing the proinflammatory gene expression [123].

The NF- $\kappa\beta$ transcription factor has repeatedly been associated with tumour development and thriving [52]. Interacting with this factor, PPARG inhibits the genesis of proinflammatory molecules such as IL-6, TNF, and MCP1 through transrepression [3, 117].

Again, a word of caution must be said due to the seemingly tumour-promoting effects of PPARG found sporadically [124–127]. Therefore, it seems as if the effects carried on by the cell depend of cell context and environment. Environment is, usually, at the helm of cellular functions.

4.7. Oxidative Stress. PPARG has demonstrated an antioxidant effect [128, 129]. SOD (Superoxide Dismutase) expression might well be regulated by PPAR because a PPRE is found in the Cu/Zn-SOD promoter [40].

IR found in diabetes mellitus and metabolic disease is certainly correlated with increased oxidative stress, which eventually could lead to an increased risk of cancer through nongenomic carcinogenesis [130–133].

In macrophages, PPARG mediates some notable abilities: uptake and reverse transport of cholesterol, macrophage subtype specification (enhancing the M2 macrophage phenotype, which is associated with higher insulin sensitivity and lower inflammation levels), and anti-inflammation properties [36, 134, 135].

Postprandial hypertriglyceridemia is associated with lower PPARG expression in metabolic syndrome patients while in healthy subjects the same "insult" leads to overexpression of PPARG [136]. We could hypothesize that since the PPARG system is injured in the metabolically ill patients, after an oxidative stress insult (a high-fat feeding), it cannot respond, leaving us more susceptible to oxidative actions and its consequences (hypothesis coined as "*nuclear receptor* *exhaustion theory*"). In the healthy group, the PPARG would perfectly be capable of managing the lipid storage and would act as an oxidative stress buffer.

4.8. Cell Migration and Invasiveness. Less evidence is available with respect to invasiveness and PPARG. However, we should pay attention to some preliminary data.

The PPARG gene modulates the invasion of cytotrophoblast into uterine tissue, which could be a novel indicator of some invasion-related function of PPARG [137].

Going further, this study by Yoshizumi et al. showed how PPARG ligand thiazolidinedione (TZD) is able to inhibit growth and metastasis of HT-29 human colon cancer cells, via the induction of cell differentiation. The use of the TZD drives to Gl arrest, in association with a great increase in p21Waf-1, Drg-1, and E-cadherin expression [77].

Paradoxically, molecules with PPARG antagonist actions are able to inhibit invasiveness and proliferation of some cancer cell lines [26, 138–140]. Again, one nuclear receptor can exert one or just the opposite function depending on the cellular environment and ligand exposure.

5. Connecting the Dots: PPARG, Vitamin D System, Obesity, and Cancer

Often in biology and medicine research, we tend to focus on the individualities of separated molecules or molecule systems in order to explain their functions, forgetting the intermolecular communication, which is ever-present in every biological system. More frequent than not, that separateness gives us a rather limited perspective of the matter at hand. For instance, the interconnectedness of biology systems and the emerging properties of such interconnectedness should be further examined and taken into account.

The crosstalk between different NRs, the "dance" and messages they give one another, is recently becoming an exciting new area which will be explored. This is the case of the VDR/VD and the PPARG system, in which both have been shown to be involved in some relationship we do not utterly understand yet.

5.1. PPARG and VDR/VD System: Commonalities in Cancer. Noteworthy, great parallelism exists between PPARG and the VDR/VD system regarding its protective role in carcinogenesis. There are a vast number of studies describing the anticancer properties of vitamin D. The majority of them are brilliantly analysed in this review by Feldman et al. [28].

Vitamin D has been extensively associated with antiinflammatory actions [141–143], apoptotic mechanisms [144– 150], antiproliferative functions [151–159], prodifferentiation effects [160–166], antiangiogenic properties [167–171], a potential role-managing invasion and metastasis [172–184], microRNA modulation [185–189], and even some role in the Hedgehog signalling pathway modulation [190]. Remarkably, most of those actions have been attributed to PPARG signalling in a somewhat lesser extent, as reviewed in this work. Such similarity and overlap in anticancer actions are worth studying. Moreover, there is enough evidence to assert that epigenetic events can influence both PPARG and VDR/VD systems behaviour.

In this study, Fujiki et al. showed that in a diabetic mouse model PPARG promoter methylation levels are higher than those of the control mice [191], along with the possibility of methylation reversal when the animals were exposed to 5AZA (5'-aza-cytidine). At least three messages can be drawn from this study: (1) the PPARG system is susceptible to epigenetic regulation, (2) diabetes and other metabolic conditions could alter the PPARG epigenetic landscape and then disrupt its proper functioning, and (3) this disruption can be reversed by drug-induced changes or, likely, by lifestyle changes.

The vitamin D system is likewise susceptible to epigenetic regulation [192–195] and, interestingly, in cancer this epigenetic repression of the vitamin D system is almost always present [196–204], which compellingly leaves the door opened to the possibility of the same phenomena happening in the PPARG system.

In fact, PPARG promoter hypermethylation is a prognostic factor of adverse outcome in colorectal cancer [46, 205]. Higher levels of PPARG promoter methylation were found in advanced tumour stages while earlier stages showed lower methylation levels. This suggests that as happens with vitamin D, advanced cancer stages can epigenetically repress PPARG expression and then nullify its antineoplastic actions.

5.2. The PPARG/VDR Crosstalk: What an Interesting Conversation! Some studies have clearly shown the existence of some communication between PPARG and VD/VDR. Interestingly, potent VDRE (Vitamin D Response Elements) have been discovered in human PPAR δ promoter, which opens the door to VDR/VD influence over the PPAR system [206]. In the opposite direction, some studies have demonstrated the ability of PPARG to bind VDR and inhibit vitamin D-mediated transactivation [207]. This data might be an indicator of bidirectional or reciprocal actions of both systems influencing each other, which have deep implications and introduce new and interesting questions to ponder upon.

Even between PPAR subtypes some modulation of expression have been found: PPAR δ could repress PPAR α and PPARG gene expression [208], illustrating the complexity of PPAR system regulation.

In the adipocyte cell, the VD/VDR system has shown anti-PPARG activity, inhibiting its expression and then adipogenesis [209, 210], which is contradictory with the commonly found proadipogenesis effects of vitamin D [211], at least in human. The factors leading to either pro- or antiadipogenesis effects are completely uncharted.

In melanoma cell lines, administration of calcitriol and several PPAR ligands modified the expression of both PPARG and VDR, demonstrating again this intriguing connection [212]. Sertznig et al. conclude in this article that calcitriol and some PPAR ligands can inhibit proliferation of the human melanoma cell line MeWo [213].

5.3. PPARG and VD/VDR System: Metabolic Commonalities. We are about to discuss the metabolic effects of vitamin D

and their analogy with those of PPARG, establishing again the parallelism.

As contradictory as it seems, VDR or CYP27B1 knockedout mice show great fat mass loss [211] while obesity in humans is commonly associated with poor vitamin D plasmatic levels [214]. Actually, an indirect relationship between Body Mass Index (BMI) and 25OHD3 has been amply described in the literature [215].

In addition, low plasmatic vitamin D levels are associated with increased risk of type 2 diabetes mellitus (T2DM) independently of BMI [24] and with hypertension, dyslipidemia (DLP), and metabolic syndrome (MS) [216, 217]. Besides, vitamin D deficiency predisposes to diabetes in animal models, while its supplementation prevents the disease [214]. Concerning PPARG, we have extensively discussed before in the review its orchestrating actions regarding adipogenesis and adipocyte metabolism. Both calcitriol (the active form of vitamin D) and PPARG seem to oppose metabolic homeostasis disruption.

Another paradoxical event is found in the fact that in humans calcitriol enhances adipogenesis while in mice the same hormone diminishes it via downregulation of C/EBP β mRNA and upregulation of CBFA2T1 (a corepressor) [218, 219]. With reference to PPARG, it enhances adipogenesis [10].

In human subcutaneous preadipocytes, calcitriol elicits actions impressively similar to those of PPARG in adipocyte maturation and differentiation. For instance, calcitriol is able to increase the expression of the enzyme Fatty Acid Synthase (FASN) increasing lipogenesis in like manner as PPARG [210].

The *storage capacity theory* introduces the idea that lipid storage capacity and the ability of PPARG to manage the processes leading to lipid storage are limited. As to that, when the organism reaches a lipid level threshold lipotoxicity shows up, PPARG is no more capable of lipid handling, and the harmful hormonal environment of obesity starts to spread through the organism [220].

Transferring the same concept of "nuclear receptor exhaustion" to VD/VDR anticancer actions we could establish a parallelism. It has shown that the VD/VDR is epigenetically downregulated in late cancer stages but overexpressed or normally expressed in early stages [221, 222]. As the aforementioned studies show, in those later stages epigenetic downregulation of the VD system molecules occurs, leaving it unable to exert its antineoplastic functions properly. Is obesity, as cancer does with vitamin D, acting as a negative epigenetic driver when it comes to PPARG signalling? That could answer why in most morbidly obese patients expression of PPARG is greatly lower in comparison to healthy subjects.

Accordingly, PPARG1 and PPARG2 expression in visceral adipose tissue (VAT) from morbidly obese (MO) subjects is significantly downregulated when compared to metabolically healthy subjects [223]. Not only that, in insulin resistant MO subjects PPARG expression is even lower [220] compared with noninsulin resistant MO patients, whichever interestingly correlates with the lower vitamin D levels found in MO with IR compared to their insulin sensitive counterparts [24]. Somehow, the metabolic impairment caused by insulin resistance is able to deteriorate both PPARG and VD/VDR system. The underlying mechanism behind this deterioration should be further studied.

A disrupted VDR/VD system leads mice to loss of fat deposits and great increase of energy expenditure. In relation to that, $VDR^{-/-}$ mice increase the expression of UCP1 (uncoupling protein 1 or Thermogenin) twenty-five-fold [211], with the consequent energy consumption. Is vitamin D, along with PPARG, an energy-conserving and metabolic homeostasis-maintaining hormone?

However, adipose tissue is not the only one affected by disruption of the VD system. A shortage of calcitriol in rats was related with increased skeletal muscle ubiquitination and loss of total muscle mass [224]. On the PPARG side, its activation through TZD in growing pigs increased muscle fiber oxidative capacity independently of fiber type [225]. Overexpression of PPAR δ in mice almost doubles the animal endurance and exercise capacity [226]. We should not lose sight of the important role the muscle has in obesity and metabolic disease pathogenesis, being a potential target for calcitriol and PPAR modulating actions.

Taken all data together, the vitamin D system seems to team up with PPARG in order to maintain proper metabolic homeostasis. Notwithstanding, in some occasions this love relationship breaks apart and both partners seem to bother one another in ways that we utterly ignore but, likely, have something to do with epigenetic regulation.

6. Conclusions

The PPARG transcription factor has been classically associated with metabolic homeostasis and lipid storage functions. Recently, newfound anticancer actions are assigned to this nuclear receptor.

However, its anticancer actions are not always consistent; in some studies some oncogenic effects have been described. We believe that cellular environment is the guiding factor behind PPARG actions and cells are controlled "from outside in." In alignment with this, the PPARG and other nuclear receptors would only be "cellular effectors," carriers of outside messages of health or disease.

When a "disease threshold" is reached, in either obesity or cancer, PPARG and VDR expression, respectively, diminishes. However, in early stages of those diseases, the expression of those nuclear receptors is higher than normal. Derived from these observations, we have coined the socalled "*nuclear receptor exhaustion theory*," by which, in an early disease stage, nuclear receptors PPARG and VDR counterbalance the harmful effects that obesity and cancer exert upon the organism, their expression being high. However, sadly, if disease progresses, it generates epigenetic silencing mechanisms upon both transcription factors, whose expression decreases radically. This silencing leaves us increasingly susceptible to disease. The positive side is that through drugs or, better yet, lifestyle changes reversal of epigenetic changes is possible.

There is an exciting function overlap between PPARG and VDR/VD system, both of which wield oncoprotective and metabolic actions. Actually, parallel metabolic and anticancer actions are described in the literature, suggesting that they team up to keep at bay those diseases. Maybe the detailed

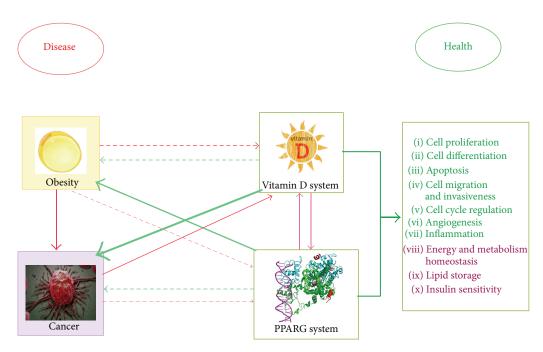


FIGURE 2: The four players: this figure shows the interrelation between the four players: obesity, cancer, vitamin D system, and PPARG. Red arrow: harmful effects, which contribute to disease. Green arrow:positive effects, which contribute to health. Disease perpetuates itself damaging both nuclear receptors. Arrow's width is in proportion with the strength and consistency of each association found in the literature. Dashed line: yet-to-be determined, preliminary, or hypothetical effects. Continuous line: in vitro/in vivo demonstrated effects. Right box: in green, actions mainly attributed to vitamin D, and in purple, actions classically attributed to PPARG. However, it is known that both agents exert every action illustrated in this box, in a higher or lower extent.

study of this overlap could give us clues in respect to the molecular pathogenesis of important conditions as metabolic disease and cancer. Further study in this new area is necessary to elucidate those questions.

Obesity, a first-order problem in our society, is linked with increased risk of cancer incidence and progression. The debatable factors behind this risk are an increment in oxidative stress, chronic inflammation, poorer vitamin D status, hormone misbalance, and, arguably, PPARG silencing through unknown mechanisms. As we know, PPARG and the vitamin D system play conjunctly a yet-to-elucidate role in cancer, so it is not surprising at all that their hypothetical epigenetic repression in obesity could be another mechanism linking this metabolic disorder to malignancies.

It has been shown, both in vitro and in vitro, that the tumours are capable of epigenetically silencing both the vitamin D and the PPARG system. This silencing could lead to the deterioration of their anticancer and metabolic actions.

Finally, a worse known crosstalk between the two NRs exists. Its usefulness, purpose, and message are (almost) utterly unexplored to us and should be studied more diligently. The interrelation, reciprocity, and interdependence of all four actors examined here might be the starting point of new fascinating research linking epigenetic signalling and two of the most hurtful diseases of our time (Figure 2).

Additional Points

Design is literature review across preclinical studies, descriptive studies, analytic studies, and reference lists of selected studies. The author focused mainly on systematic and narrative reviews. Data sources are medline (Pubmed), Jábega 2.0 (Málaga University Search Engine Software), Gerión search engine, and screening of citations and references. Regarding eligibility criteria, we focused on papers published in magazines considered to be in the first impact factor quartile without restrictions regarding publishing date. Keywords are PPARG; Obesity; Transcription factor; Vitamin D; Calcitriol; Vitamin D Receptor; Epigenetics; Nuclear Receptor; Cancer; Methylation.

Competing Interests

The authors declare that there are no competing interests.

Acknowledgments

This study was supported by "*Centros de Investigación En Red*" (CIBER, CB06/03/0018), the "Instituto de Salud Carlos III" (ISCIII), and grants from ISCIII (PII1/01661) and from Consejería de Innovacion, Ciencia y Empresa de la Junta de Andalucía (PII1-CTS-8181) and cofinanced by the European Regional Development Fund (FEDER). M. Macías-González was recipient of the Nicolas Monarde program from the Servicio Andaluz de Salud, Junta de Andalucía, Spain (C0029-2014).

References

 J. N. Feige, L. Gelman, L. Michalik, B. Desvergne, and W. Wahli, "From molecular action to physiological outputs: peroxisome proliferator-activated receptors are nuclear receptors at the crossroads of key cellular functions," *Progress in Lipid Research*, vol. 45, no. 2, pp. 120–159, 2006.

- [2] Y. Zhu, C. Qi, J. R. Korenberg et al., "Structural organization of mouse peroxisome proliferator-activated receptor γ (mPPARγ) gene: alternative promoter use and different splicing yield two mPPARγ isoforms," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 92, no. 17, pp. 7921– 7925, 1995.
- [3] J. M. Peters, Y. M. Shah, and F. J. Gonzalez, "The role of peroxisome proliferator-activated receptors in carcinogenesis and chemoprevention," *Nature Reviews Cancer*, vol. 12, no. 3, pp. 181–195, 2012.
- [4] A. Rogue, C. Spire, M. Brun, N. Claude, and A. Guillouzo, "Gene expression changes induced by PPAR γ agonists in animal and human liver," *PPAR Research*, vol. 2010, Article ID 325183, 16 pages, 2010.
- [5] T. M. Willson, P. J. Brown, D. D. Sternbach, and B. R. Henke, "The PPARs: from orphan receptors to drug discovery," *Journal* of Medicinal Chemistry, vol. 43, no. 4, pp. 527–550, 2000.
- [6] M. Schupp and M. A. Lazar, "Endogenous ligands for nuclear receptors: digging deeper," *Journal of Biological Chemistry*, vol. 285, no. 52, pp. 40409–40415, 2010.
- [7] S. C. Wu and Y. Zhang, "Minireview: role of protein methylation and demethylation in nuclear hormone signaling," *Molecular Endocrinology*, vol. 23, no. 9, pp. 1323–1334, 2009.
- [8] S. Sugii and R. M. Evans, "Epigenetic codes of PPARγ in metabolic disease," *FEBS Letters*, vol. 585, no. 13, pp. 2121–2128, 2011.
- [9] Y. Kodera, K.-I. Takeyama, A. Murayama, M. Suzawa, Y. Masuhiro, and S. Kato, "Ligand type-specific interactions of peroxisome proliferator-activated receptor *γ* with transcriptional coactivators," *The Journal of Biological Chemistry*, vol. 275, no. 43, pp. 33201–33204, 2000.
- [10] P. Tontonoz and B. M. Spiegelman, "Fat and beyond: the diverse biology of PPARy," *Annual Review of Biochemistry*, vol. 77, pp. 289–312, 2008.
- [11] Y. Barak, M. C. Nelson, E. S. Ong et al., "PPARy is required for placental, cardiac, and adipose tissue development," *Molecular Cell*, vol. 4, no. 4, pp. 585–595, 1999.
- [12] J. Berger and D. E. Moller, "The mechanisms of action of PPARs," *Annual Review of Medicine*, vol. 53, no. 1, pp. 409–435, 2002.
- [13] E. Boitier, J.-C. Gautier, and R. Roberts, "Advances in understanding the regulation of apoptosis and mitosis by peroxisomeproliferator activated receptors in pre-clinical models: relevance for human health and disease," *Comparative Hepatology*, vol. 2, article 3, 2003.
- [14] J.-H. Kim, J. Song, and K. W. Park, "The multifaceted factor peroxisome proliferator-activated receptor γ (PPAR γ) in metabolism, immunity, and cancer," *Archives of Pharmacal Research*, vol. 38, no. 3, pp. 302–312, 2015.
- [15] S. Nagpal, S. Na, and R. Rathnachalam, "Noncalcemic actions of vitamin D receptor ligands," *Endocrine Reviews*, vol. 26, no. 5, pp. 662–687, 2005.
- [16] N. J. Groves, J. J. McGrath, and T. H. J. Burne, "Vitamin D as a neurosteroid affecting the developing and adult brain," *Annual Review of Nutrition*, vol. 34, pp. 117–141, 2014.
- [17] J. W. London, "Low Vitamin D is linked to faster cognitive decline in older adults," *The British Medical Journal*, vol. 351, Article ID h4916, 2015.

- [18] M. McCarthy, "Study supports link between low vitamin D and dementia risk," *BMJ*, vol. 349, Article ID g5049, 2014.
- [19] T. Koizumi, Y. Nakao, T. Matsui et al., "Effects of corticosteroid and 1,24R-dihydroxy-vitamin D₃ administration on lymphoproliferation and autoimmune disease in MRL/MPlpr/lpr mice," *International Archives of Allergy and Applied Immunology*, vol. 77, no. 4, pp. 396–404, 1985.
- [20] S. Gregori, N. Giarratana, S. Smiroldo, M. Uskokovic, and L. Adorini, "A 1α ,25-dihydroxyvitamin D₃ analog enhances regulatory T-cells and arrests autoimmune diabetes in NOD mice," *Diabetes*, vol. 51, no. 5, pp. 1367–1374, 2002.
- [21] M. Tsuji, K. Fujii, T. Nakano, and Y. Nishii, " 1α -hydroxyvitamin D₃ inhibits Type II collagen-induced arthritis in rats," *FEBS Letters*, vol. 337, no. 3, pp. 248–250, 1994.
- [22] A. Manousopoulou, N. M. Al-Daghri, S. D. Garbis, and G. P. Chrousos, "Vitamin D and cardiovascular risk among adults with obesity: a systematic review and meta-analysis," *European Journal of Clinical Investigation*, vol. 45, no. 10, pp. 1113–1126, 2015.
- [23] B. J. Boucher, W. G. John, and K. Noonan, "Hypovitaminosis D is associated with insulin resistance and beta cell dysfunction," *The American Journal of Clinical Nutrition*, vol. 80, no. 6, pp. 1666–1667, 2004.
- [24] M. Clemente-Postigo, A. Muñoz-Garach, M. Serrano et al., "Serum 25-hydroxyvitamin D and adipose tissue vitamin D receptor gene expression: relationship with obesity and type 2 diabetes," *The Journal of Clinical Endocrinology & Metabolism*, vol. 100, no. 4, pp. E591–E595, 2015.
- [25] G. Bakdash, T. M. M. van Capel, L. M. K. Mason, M. L. Kapsenberg, and E. C. de Jong, "Vitamin D3 metabolite calcidiol primes human dendritic cells to promote the development of immunomodulatory IL-10-producing T cells," *Vaccine*, vol. 32, no. 47, pp. 6294–6302, 2014.
- [26] K. Takahashi, "Influence of bacteria on epigenetic gene control," *Cellular and Molecular Life Sciences*, vol. 71, no. 6, pp. 1045–1054, 2014.
- [27] Y. Ma, C. S. Johnson, and D. L. Trump, Mechanistic Insights of Vitamin D Anticancer Effects, 2016.
- [28] D. Feldman, A. V. Krishnan, S. Swami, E. Giovannucci, and B. J. Feldman, "The role of vitamin D in reducing cancer risk and progression," *Nature Reviews Cancer*, vol. 14, no. 5, pp. 342–357, 2014.
- [29] A. G. Cristancho and M. A. Lazar, "Forming functional fat: a growing understanding of adipocyte differentiation," *Nature Reviews Molecular Cell Biology*, vol. 12, no. 11, pp. 722–734, 2011.
- [30] E. D. Rosen and B. M. Spiegelman, "Molecular regulation of adipogenesis," *Annual Review of Cell and Developmental Biology*, vol. 16, pp. 145–171, 2000.
- [31] A. L. Hevener, W. He, Y. Barak et al., "Muscle-specific Pparg deletion causes insulin resistance," *Nature Medicine*, vol. 9, no. 12, pp. 1491–1497, 2003.
- [32] F. Wang, S. E. Mullican, J. R. DiSpirito, L. C. Peed, and M. A. Lazar, "Lipoatrophy and severe metabolic disturbance in mice with fat-specific deletion of PPARy," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 46, pp. 18656–18661, 2013.
- [33] K. Won Park, D. S. Halperin, and P. Tontonoz, "Before they were fat: adipocyte progenitors," *Cell Metabolism*, vol. 8, no. 6, pp. 454–457, 2008.
- [34] K. Matsusue, M. Haluzik, G. Lambert et al., "Liver-specific disruption of PPARy in leptin-deficient mice improves fatty

liver but aggravates diabetic phenotypes," *Journal of Clinical Investigation*, vol. 111, no. 5, pp. 737–747, 2003.

- [35] A. L. Hevener, J. M. Olefsky, D. Reichart et al., "Macrophage PPARγ is required for normal skeletal muscle and hepatic insulin sensitivity and full antidiabetic effects of thiazolidinediones," *Journal of Clinical Investigation*, vol. 117, no. 6, pp. 1658– 1669, 2007.
- [36] J. I. Odegaard, R. R. Ricardo-Gonzalez, M. H. Goforth et al., "Macrophage-specific PPARy controls alternative activation and improves insulin resistance," *Nature*, vol. 447, no. 7148, pp. 1116–1120, 2007.
- [37] Z. Ament, M. Masoodi, and J. L. Griffin, "Applications of metabolomics for understanding the action of peroxisome proliferator-activated receptors (PPARs) in diabetes, obesity and cancer," *Genome Medicine*, vol. 4, no. 4, article 32, 2012.
- [38] A. Vidal-Puig, M. Jimenez-Liñan, B. B. Lowell et al., "Regulation of PPAR γ gene expression by nutrition and obesity in rodents," *The Journal of Clinical Investigation*, vol. 97, no. 11, pp. 2553– 2561, 1996.
- [39] M. U. Imam, M. Ismail, H. Ithnin, Z. Tubesha, and A. R. Omar, "Effects of germinated brown rice and its bioactive compounds on the expression of the peroxisome proliferatoractivated receptor gamma gene," *Nutrients*, vol. 5, no. 2, pp. 468– 477, 2013.
- [40] E. Garcia-Fuentes, M. Murri, L. Garrido-Sanchez et al., "PPARγ expression after a high-fat meal is associated with plasma superoxide dismutase activity in morbidly obese persons," *Obesity*, vol. 18, no. 5, pp. 952–958, 2010.
- [41] Á. Lendvai, M. J. Deutsch, T. Plösch, and R. Ensenauer, "The peroxisome proliferator-activated receptors under epigenetic control in placental metabolism and fetal development," *American Journal of Physiology-Endocrinology And Metabolism*, vol. 310, no. 10, pp. E797–E810, 2016.
- [42] M. J. Campbell, C. Carlberg, and H. P. Koeffler, "A role for the PPARγ in cancer therapy," *PPAR Research*, vol. 2008, Article ID 314974, 17 pages, 2008.
- [43] C. Grommes, G. E. Landreth, and M. T. Heneka, "Antineoplastic effects of peroxisome proliferator-activated receptor γ agonists," *Lancet Oncology*, vol. 5, no. 7, pp. 419–429, 2004.
- [44] H. P. Koeffler, "Peroxisome proliferator-activated receptor γ and cancers," *Clinical Cancer Research*, vol. 9, no. 1, pp. 1–9, 2003.
- [45] S. Battaglia, O. Maguire, J. L. Thorne et al., "Elevated NCOR1 disrupts PPAR α/γ signaling in prostate cancer and forms a targetable epigenetic lesion," *Carcinogenesis*, vol. 31, no. 9, pp. 1650–1660, 2010.
- [46] M. Pancione, L. Sabatino, A. Fucci et al., "Epigenetic silencing of peroxisome proliferator-activated receptor γ is a biomarker for colorectal cancer progression and adverse patients' outcome," *PLoS ONE*, vol. 5, no. 12, Article ID e14229, 2010.
- [47] C. A. McAlpine, Y. Barak, I. Matise, and R. T. Cormier, "Intestinal-specific PPARγ deficiency enhances tumorigenesis in Apc^{Min/+} mice," *International Journal of Cancer*, vol. 119, no. 10, pp. 2339–2346, 2006.
- [48] P. Sarraf, E. Mueller, W. M. Smith et al., "Loss-of-function mutations in PPARγ associated with human colon cancer," *Molecular Cell*, vol. 3, no. 6, pp. 799–804, 1999.
- [49] D. Capaccio, A. Ciccodicola, L. Sabatino et al., "A novel germline mutation in Peroxisome Proliferator-Activated Receptor γ gene associated with large intestine polyp formation and dyslipidemia," *Biochimica et Biophysica Acta-Molecular Basis of Disease*, vol. 1802, no. 6, pp. 572–581, 2010.

- [50] M. Pancione, N. Forte, L. Sabatino et al., "Reduced β-catenin and peroxisome proliferator-activated receptor-γ expression levels are associated with colorectal cancer metastatic progression: correlation with tumor-associated macrophages, cyclooxygenase 2, and patient outcome," *Human Pathology*, vol. 40, no. 5, pp. 714–725, 2009.
- [51] S. Ogino, K. Shima, Y. Baba et al., "Colorectal cancer expression of peroxisome proliferator-activated receptor γ (PPARG, PPAR γ) is associated with good prognosis," *Gastroenterology*, vol. 136, no. 4, pp. 1242–1250, 2009.
- [52] A. Mantovani, P. Allavena, A. Sica, and F. Balkwill, "Cancerrelated inflammation," *Nature*, vol. 454, no. 7203, pp. 436–444, 2008.
- [53] S. Tsugane and M. Inoue, "Insulin resistance and cancer: epidemiological evidence," *Cancer Science*, vol. 101, no. 5, pp. 1073–1079, 2010.
- [54] I. Bilic, "Obesity and cancer," *Periodicum Biologorum*, vol. 116, no. 4, pp. 355–359, 2014.
- [55] M. J. Khandekar, P. Cohen, and B. M. Spiegelman, "Molecular mechanisms of cancer development in obesity," *Nature Reviews Cancer*, vol. 11, no. 12, pp. 886–895, 2011.
- [56] A. G. Renehan, "Obesity and cancer," in *Adipose Tissue in Health and Disease*, pp. 369–384, 2010.
- [57] E. E. Calle and R. Kaaks, "Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms," *Nature Reviews Cancer*, vol. 4, no. 8, pp. 579–591, 2004.
- [58] K. Y. Wolin, K. Carson, and G. A. Colditz, "Obesity and cancer," *Oncologist*, vol. 15, no. 6, pp. 556–565, 2010.
- [59] C. A. Gonzá Lez Svatetz and A. Goday Arnó, "Obesidad y cáncer," *Medicina Clínica*, vol. 145, no. 1, pp. 24–30, 2015.
- [60] E. Kebebew, M. Peng, E. Reiff et al., "A phase II trial of rosiglitazone in patients with thyroglobulin-positive and radioiodinenegative differentiated thyroid cancer," *Surgery*, vol. 140, no. 6, pp. 960–967, 2006.
- [61] G. Vitale, S. Zappavigna, M. Marra et al., "The PPAR-γ agonist troglitazone antagonizes survival pathways induced by STAT-3 in recombinant interferon-β treated pancreatic cancer cells," *Biotechnology Advances*, vol. 30, no. 1, pp. 169–184, 2012.
- [62] R. A. Lubet, S. M. Fischer, V. E. Steele, M. M. Juliana, R. Desmond, and C. J. Grubbs, "Rosiglitazone, a PPAR γ agonist: potent promoter of hydroxybutyl(butyl)nitrosamine-induced urinary bladder cancers," *International Journal of Cancer*, vol. 123, no. 10, pp. 2254–2259, 2008.
- [63] M. Fujita, T. Yagami, M. Fujio et al., "Cytotoxicity of troglitazone through PPARγ-independent pathway and p38 MAPK pathway in renal cell carcinoma," *Cancer Letters*, vol. 312, no. 2, pp. 219– 227, 2011.
- [64] A. Elnemr, T. Ohta, K. Iwata et al., "PPARgamma ligand (thiazolidinedione) induces growth arrest and differentiation markers of human pancreatic cancer cells," *International Journal* of Oncology, vol. 17, no. 6, pp. 1157–1164, 2000.
- [65] H. Koga, S. Sakisaka, M. Harada et al., "Involvement of p21^{WAF1/Cip1}, p27^{Kip1}, and p18^{INK4c} in troglitazone-induced cellcycle arrest in human hepatoma cell lines," *Hepatology*, vol. 33, no. 5, pp. 1087–1097, 2001.
- [66] F. Chen and L. E. Harrison, "Ciglitazone-induced cellular antiproliferation increases p27 kip1 protein levels through both increased transcriptional activity and inhibition of proteasome degradation," *Cellular Signalling*, vol. 17, no. 7, pp. 809–816, 2005.
- [67] F. Chen, E. Kim, C.-C. Wang, and L. E. Harrison, "Ciglitazoneinduced p27 gene transcriptional activity is mediated through

Sp1 and is negatively regulated by the MAPK signaling pathway," *Cellular Signalling*, vol. 17, no. 12, pp. 1572–1577, 2005.

- [68] W. Motomura, T. Okumura, N. Takahashi, T. Obara, and Y. Kohgo, "Activation of peroxisome proliferator-activated receptor gamma by troglitazone inhibits cell growth through the increase of p27KiP1 in human. Pancreatic carcinoma cells," *Cancer Research*, vol. 60, no. 19, pp. 5558–5564, 2000.
- [69] A. Itami, G. Watanabe, Y. Shimada et al., "Ligands for peroxisome proliferator-activated receptor γ inhibit growth of pancreatic cancers both in vitro and in vivo," *International Journal of Cancer*, vol. 94, no. 3, pp. 370–376, 2001.
- [70] H. Lapillonne, M. Konopleva, T. Tsao et al., "Activation of peroxisome proliferator-activated receptor gamma by a novel synthetic triterpenoid 2-cyano-3,12-dioxooleana-1,9-dien-28oic acid induces growth arrest and apoptosis in breast cancer cells," *Cancer Research*, vol. 63, no. 18, pp. 5926–5939, 2003.
- [71] J.-W. Huang, C.-W. Shiau, Y.-T. Yang et al., "Peroxisome proliferator-activated receptor γ-independent ablation of cyclin D1 by thiazolidinediones and their derivatives in breast cancer cells," *Molecular Pharmacology*, vol. 67, no. 4, pp. 1342–1348, 2005.
- [72] C. Qin, R. Burghardt, R. Smith, M. Wormke, J. Stewart, and S. Safe, "Peroxisome proliferator-activated receptor γ agonists induce proteasome-dependent degradation of cyclin D1 and estrogen receptor α in MCF-7 breast cancer cells," *Cancer Research*, vol. 63, no. 5, pp. 958–964, 2003.
- [73] C. Wang, M. Fu, M. D'Amico et al., "Inhibition of cellular proliferation through IκB kinase-independent and peroxisome proliferator-activated receptor γ-dependent repression of cyclin D1," *Molecular and Cellular Biology*, vol. 21, no. 9, pp. 3057–3070, 2001.
- [74] F. Yin, S. Wakino, Z. Liu et al., "Troglitazone inhibits growth of MCF-7 breast carcinoma cells by targeting G1 cell cycle regulators," *Biochemical and Biophysical Research Communications*, vol. 286, no. 5, pp. 916–922, 2001.
- [75] P. Tontonoz, S. Singer, B. M. Forman et al., "Terminal differentiation of human liposarcoma cells induced by ligands for peroxisome proliferator-activated receptor γ and the retinoid X receptor," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 1, pp. 237–241, 1997.
- [76] R. A. Gupta, J. A. Brockman, P. Sarraf, T. M. Willson, and R. N. DuBois, "Target genes of peroxisome proliferator-activated receptor γ in colorectal cancer cells," *The Journal of Biological Chemistry*, vol. 276, no. 32, pp. 29681–29687, 2001.
- [77] T. Yoshizumi, T. Ohta, I. Ninomiya et al., "Thiazolidinedione, a peroxisome proliferator-activated receptor-gamma ligand, inhibits growth and metastasis of HT-29 human colon cancer cells through differentiation-promoting effects," *International Journal of Oncology*, vol. 25, no. 3, pp. 631–639, 2004.
- [78] G. D. Demetri, C. D. M. Fletcher, E. Mueller et al., "Induction of solid tumor differentiation by the peroxisome proliferatoractivated receptor-γ ligand troglitazone in patients with liposarcoma," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 96, no. 7, pp. 3951–3956, 1999.
- [79] A. Margeli, G. Kouraklis, and S. Theocharis, "Peroxisome proliferator activated receptor-γ (PPAR-γ) ligands and angiogenesis," *Angiogenesis*, vol. 6, no. 3, pp. 165–169, 2003.
- [80] F. Biscetti, E. Gaetani, A. Flex et al., "Selective activation of peroxisome proliferator-activated receptor (PPAR)α and PPARγ induces neoangiogenesis through a vascular endothelial growth factor-dependent mechanism," *Diabetes*, vol. 57, no. 5, pp. 1394–1404, 2008.

- [81] K. Chu, S.-T. Lee, J.-S. Koo et al., "Peroxisome proliferatoractivated receptor-γ-agonist, rosiglitazone, promotes angiogenesis after focal cerebral ischemia," *Brain Research*, vol. 1093, no. 1, pp. 208–218, 2006.
- [82] P.-H. Huang, M. Sata, H. Nishimatsu, M. Sumi, Y. Hirata, and R. Nagai, "Pioglitazone ameliorates endothelial dysfunction and restores ischemia-induced angiogenesis in diabetic mice," *Biomedicine and Pharmacotherapy*, vol. 62, no. 1, pp. 46–52, 2008.
- [83] D. Bishop-Bailey and T. Hla, "Endothelial cell apoptosis induced by the peroxisome proliferator- activated receptor (PPAR) ligand 15-deoxy-Δ12,14-prostaglandin J2," *Journal of Biological Chemistry*, vol. 274, no. 24, pp. 17042–17048, 1999.
- [84] X. Xin, S. Yang, J. Kowalski, and M. E. Gerritsen, "Peroxisome proliferator-activated receptor γ ligands are potent inhibitors of angiogenesis *in vitro* and *in vivo*," *The Journal of Biological Chemistry*, vol. 274, no. 13, pp. 9116–9121, 1999.
- [85] A. Margeli, G. Kouraklis, and S. Theocharis, "Peroxisome proliferator activated receptor-γ (PPAR-γ) ligands and angiogenesis," *Angiogenesis*, vol. 6, no. 3, pp. 165–169, 2003.
- [86] S. Fauconnet, I. Lascombe, E. Chabannes et al., "Differential regulation of vascular endothelial growth factor expression by peroxisome proliferator-activated receptors in bladder cancer cells," *Journal of Biological Chemistry*, vol. 277, no. 26, pp. 23534– 23543, 2002.
- [87] V. Chintalgattu, G. S. Harris, S. M. Akula, and L. C. Katwa, "PPAR-γ agonists induce the expression of VEGF and its receptors in cultured cardiac myofibroblasts," *Cardiovascular Research*, vol. 74, no. 1, pp. 140–150, 2007.
- [88] R. M. Bambury, G. Iyer, and J. E. Rosenberg, "Specific PPAR gamma agonists may have different effects on cancer incidence," *Annals of Oncology*, vol. 24, no. 3, article 854, 2013.
- [89] M. Terrasi, V. Bazan, S. Caruso et al., "Effects of PPARy agonists on the expression of leptin and vascular endothelial growth factor in breast cancer cells," *Journal of Cellular Physiology*, vol. 228, no. 6, pp. 1368–1374, 2013.
- [90] H. A. Elrod and S.-Y. Sun, "PPARy and apoptosis in cancer," PPAR Research, vol. 2008, Article ID 704165, 12 pages, 2008.
- [91] A. Trombetta, M. Maggiora, G. Martinasso, P. Cotogni, R. A. Canuto, and G. Muzio, "Arachidonic and docosahexaenoic acids reduce the growth of A549 human lung-tumor cells increasing lipid peroxidation and PPARs," *Chemico-Biological Interactions*, vol. 165, no. 3, pp. 239–250, 2007.
- [92] I. J. Edwards, I. M. Berquin, H. Sun et al., "Differential effects of delivery of omega-3 fatty acids to human cancer cells by lowdensity lipoproteins versus albumin," *Clinical Cancer Research*, vol. 10, no. 24, pp. 8275–8283, 2004.
- [93] H. Sun, I. M. Berquin, R. T. Owens, J. T. O'Flaherty, and I. J. Edwards, "Peroxisome proliferator-activated receptor γmediated up-regulation of syndecan-1 by n-3 fatty acids promotes apoptosis of human breast cancer cells," *Cancer Research*, vol. 68, no. 8, pp. 2912–2919, 2008.
- [94] E. Mueller, M. Smith, P. Sarraf et al., "Effects of ligand activation of peroxisome proliferator-activated receptor γ in human prostate cancer," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 20, pp. 10990–10995, 2000.
- [95] S. S. Palakurthi, H. Aktas, L. M. Grubissich, R. M. Mortensen, and J. A. Halperin, "Anticancer effects of thiazolidinediones are independent of peroxisome proliferator-activated receptor γ and mediated by inhibition of translation initiation," *Cancer Research*, vol. 61, no. 16, pp. 6213–6218, 2001.

- [96] G. Debrock, V. Vanhentenrijk, R. Sciot, M. Debiec-Rychter, R. Oyen, and A. Van Oosterom, "A phase II trial with rosiglitazone in liposarcoma patients," *British Journal of Cancer*, vol. 89, no. 8, pp. 1409–1412, 2003.
- [97] M. R. Smith, J. Manola, D. S. Kaufman et al., "Rosiglitazone versus placebo for men with prostate carcinoma and a rising serum prostate-specific antigen level after radical prostatectomy and/or radiation therapy," *Cancer*, vol. 101, no. 7, pp. 1569–1574, 2004.
- [98] H. J. Burstein, G. D. Demetri, E. Mueller, P. Sarraf, B. M. Spiegelman, and E. P. Winer, "Use of the peroxisome proliferatoractivated receptor (PPAR) γ ligand troglitazone as treatment for refractory breast cancer: a phase II study," *Breast Cancer Research and Treatment*, vol. 79, no. 3, pp. 391–397, 2003.
- [99] M. H. Kulke, G. D. Demetri, N. E. Sharpless et al., "A phase II study of troglitazone, an activator of the PPARy receptor, in patients with chemotherapy-resistant metastatic colorectal cancer," *Cancer Journal*, vol. 8, no. 5, pp. 395–399, 2002.
- [100] Y. Liu, Z.-A. Zhu, S.-N. Zhang et al., "Combinational effect of PPARγ agonist and RXR agonist on the growth of SGC7901 gastric carcinoma cells in vitro," *Tumor Biology*, vol. 34, no. 4, pp. 2409–2418, 2013.
- [101] B. Farrow and B. M. Evers, "Activation of PPARγ increases PTEN expression in pancreatic cancer cells," *Biochemical and Biophysical Research Communications*, vol. 301, no. 1, pp. 50–53, 2003.
- [102] W. Zhang, N. Wu, Z. Li, L. Wang, J. Jin, and X.-L. Zha, "PPARγ activator rosiglitazone inhibits cell migration via upregulation of PTEN in human hepatocarcinoma cell line BEL-7404," *Cancer Biology and Therapy*, vol. 5, no. 8, pp. 1008–1014, 2006.
- [103] R. E. Teresi, C.-W. Shaiu, C.-S. Chen, V. K. Chatterjee, K. A. Waite, and C. Eng, "Increased PTEN expression due to transcriptional activation of PPARγ by Lovastatin and Rosiglitazone," *International Journal of Cancer*, vol. 118, no. 10, pp. 2390–2398, 2006.
- [104] S. Y. Lee, G. Y. Hur, K. H. Jung et al., "PPAR-γ agonist increase gefitinib's antitumor activity through PTEN expression," *Lung Cancer*, vol. 51, no. 3, pp. 297–301, 2006.
- [105] L. Patel, I. Pass, P. Coxon, C. P. Downes, S. A. Smith, and C. H. Macphee, "Tumor suppressor and anti-inflammatory actions of PPARy agonists are mediated via upregulation of PTEN," *Current Biology*, vol. 11, no. 10, pp. 764–768, 2001.
- [106] T. Zander, J. A. Kraus, C. Grommes et al., "Induction of apoptosis in human and rat glioma by agonists of the nuclear receptor PPARy," *Journal of Neurochemistry*, vol. 81, no. 5, pp. 1052–1060, 2002.
- [107] M.-A. Bae and B. J. Song, "Critical role of c-Jun N-terminal protein kinase activation in troglitazone-induced apoptosis of human HepG2 hepatoma cells," *Molecular Pharmacology*, vol. 63, no. 2, pp. 401–408, 2003.
- [108] Y. Kim, N. Suh, M. Sporn, and J. C. Reed, "An inducible pathway for degradation of FLIP protein sensitizes tumor cells to TRAILinduced apoptosis," *Journal of Biological Chemistry*, vol. 277, no. 25, pp. 22320–22329, 2002.
- [109] K. Schultze, B. Böck, A. Eckert et al., "Troglitazone sensitizes tumor cells to TRAIL-induced apoptosis via down-regulation of FLIP and survivin," *Apoptosis*, vol. 11, no. 9, pp. 1503–1512, 2006.
- [110] C.-W. Shiau, C.-C. Yang, S. K. Kulp et al., "Thiazolidenediones mediate apoptosis in prostate cancer cells in part through inhibition of Bcl-xL/Bcl-2 functions independently of PPARy," *Cancer Research*, vol. 65, no. 4, pp. 1561–1569, 2005.

- [111] K. Y. Kim, S. S. Kim, and H. G. Cheon, "Differential anti-proliferative actions of peroxisome proliferator-activated receptor-γ agonists in MCF-7 breast cancer cells," *Biochemical Pharmacology*, vol. 72, no. 5, pp. 530–540, 2006.
- [112] K.-H. Yan, C.-J. Yao, H.-Y. Chang, G.-M. Lai, A.-L. Cheng, and S.-E. Chuang, "The synergistic anticancer effect of troglitazone combined with aspirin causes cell cycle arrest and apoptosis in human lung cancer cells," *Molecular Carcinogenesis*, vol. 49, no. 3, pp. 235–246, 2010.
- [113] F. Colotta, P. Allavena, A. Sica, C. Garlanda, and A. Mantovani, "Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability," *Carcinogenesis*, vol. 30, no. 7, pp. 1073–1081, 2009.
- [114] T. Atsumi, R. Singh, L. Sabharwal et al., "Inflammation amplifier, a new paradigm in cancer biology," *Cancer Research*, vol. 74, no. 1, pp. 8–14, 2014.
- [115] N. Gagliani, B. Hu, S. Huber, E. Elinav, and R. A. Flavell, "The fire within: microbes inflame tumors," *Cell*, vol. 157, no. 4, pp. 776–783, 2014.
- [116] C. I. Diakos, K. A. Charles, D. C. McMillan, and S. J. Clarke, "Cancer-related inflammation and treatment effectiveness," *The Lancet Oncology*, vol. 15, no. 11, pp. e493–e503, 2014.
- [117] C. K. Glass and K. Saijo, "Nuclear receptor transrepression pathways that regulate inflammation in macrophages and T cells," *Nature Reviews Immunology*, vol. 10, no. 5, pp. 365–376, 2010.
- [118] D. S. Straus and C. K. Glass, "Anti-inflammatory actions of PPAR ligands: new insights on cellular and molecular mechanisms," *Trends in Immunology*, vol. 28, no. 12, pp. 551–558, 2007.
- [119] Y. M. Shah, K. Morimura, and F. J. Gonzalez, "Expression of peroxisome proliferator-activated receptor-γ in macrophage suppresses experimentally induced colitis," *American Journal of Physiology—Gastrointestinal and Liver Physiology*, vol. 292, no. 2, pp. G657–G666, 2007.
- [120] D. S. Straus, G. Pascual, M. Li et al., "15-Deoxy- $\Delta^{12,14}$ prostaglandin J₂ inhibits multiple steps in the NF- κ B signaling pathway," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 9, pp. 4844–4849, 2000.
- [121] R. Kapadia, J.-H. Yi, and R. Vemuganti, "Mechanisms of antiinflammatory and neuroprotective actions of PPAR-gamma agonists," *Frontiers in Bioscience*, vol. 13, no. 5, pp. 1813–1826, 2008.
- [122] A. M. Sharma and B. Staels, "Review: peroxisome proliferatoractivated receptor gamma and adipose tissue—understanding obesity-related changes in regulation of lipid and glucose metabolism," *The Journal of Clinical Endocrinology & Metabolism*, vol. 92, no. 2, pp. 386–395, 2007.
- [123] G. Pascual, A. L. Fong, S. Ogawa et al., "A SUMOylationdependent pathway mediates transrepression of inflammatory response genes by PPAR-γ," *Nature*, vol. 437, no. 7059, pp. 759– 763, 2005.
- [124] A. M. Lefebvre, I. Chen, P. Desreumaux et al., "Activation of the peroxisome proliferator-activated receptor γ promotes the development of colon tumors in C57BL/6J-APCMin/+ mice," *Nature Medicine*, vol. 4, no. 9, pp. 1053–1057, 1998.
- [125] M. V. Pino, M. F. Kelley, and Z. Jayyosi, "Promotion of colon tumors in C57BL/6J-APC(min)/+ mice by thiazolidinedione PPARy agonists and a structurally unrelated PPARy agonist," *Toxicologic Pathology*, vol. 32, no. 1, pp. 58–63, 2004.
- [126] E. Saez, P. Tontonoz, M. C. Nelson et al., "Activators of the nuclear receptor PPARγ enhance colon polyp formation," *Nature Medicine*, vol. 4, no. 9, pp. 1058–1061, 1998.

- [127] K. Yang, K.-H. Fan, S. A. Lamprecht et al., "Peroxisome proliferator-activated receptor γ agonist troglitazone induces colon tumors in normal C57BL/6J mice and enhances colonic carcinogenesis in Apc1638 N/+ Mlh1+/- double mutant mice," *International Journal of Cancer*, vol. 116, no. 4, pp. 495–499, 2005.
- [128] Z. Bagi, A. Koller, and G. Kaley, "PPARγ activation, by reducing oxidative stress, increases NO bioavailability in coronary arterioles of mice with Type 2 diabetes," *American Journal of Physiology—Heart and Circulatory Physiology*, vol. 286, no. 2, pp. H742–H748, 2004.
- [129] I. Inoue, S.-I. Goto, T. Matsunaga et al., "The ligands/activators for peroxisome proliferator-activated receptor α (PPAR α) and PPAR γ increase Cu²⁺,Zn²⁺-superoxide dismutase and decrease p22phox message expressions in primary endothelial cells," *Metabolism: Clinical and Experimental*, vol. 50, no. 1, pp. 3–11, 2001.
- [130] F. J. Tinahones, M. Murri-Pierri, L. Garrido-Sánchez et al., "Oxidative stress in severely obese persons is greater in those with insulin resistance," *Obesity*, vol. 17, no. 2, pp. 240–246, 2009.
- [131] S. O. Olusi, "Obesity is an independent risk factor for plasma lipid peroxidation and depletion of erythrocyte cytoprotectic enzymes in humans," *International Journal of Obesity and Related Metabolic Disorders*, vol. 26, no. 9, pp. 1159–1164, 2002.
- [132] F. Giacco and M. Brownlee, "Oxidative stress and diabetic complications," *Circulation Research*, vol. 107, no. 9, pp. 1058– 1070, 2010.
- [133] J. W. Baynes, "Role of oxidative stress in development of complications in diabetes," *Diabetes*, vol. 40, no. 4, pp. 405–412, 1991.
- [134] J. M. Olefsky and C. K. Glass, "Macrophages, inflammation, and insulin resistance," *Annual Review of Physiology*, vol. 72, no. 1, pp. 219–246, 2010.
- [135] K. J. Moore, M. L. Fitzgerald, and M. W. Freeman, "Peroxisome proliferator-activated receptors in macrophage biology: friend or foe?" *Current Opinion in Lipidology*, vol. 12, no. 5, pp. 519– 527, 2001.
- [136] M. Macias-Gonzalez, F. Cardona, M. Queipo-Ortuño, R. Bernal, M. Martin, and F. J. Tinahones, "PPARy mRNA expression is reduced in peripheral blood mononuclear cells after fat overload in patients with metabolic syndrome," *Journal of Nutrition*, vol. 138, no. 5, pp. 903–907, 2008.
- [137] M. Bilban, P. Haslinger, J. Prast et al., "Identification of novel trophoblast invasion-related genes: heme oxygenase-1 controls motility via peroxisome proliferator-activated receptor γ," *Endocrinology*, vol. 150, no. 2, pp. 1000–1013, 2009.
- [138] J. D. Burton, M. E. Castillo, D. M. Goldenberg, and R. D. Blumenthal, "Peroxisome proliferator-activated receptor-γ antagonists exhibit potent antiproliferative effects versus many hematopoietic and epithelial cancer cell lines," *Anti-Cancer Drugs*, vol. 18, no. 5, pp. 525–534, 2007.
- [139] K. L. Schaefer, K. Wada, H. Takahashi et al., "Peroxisome proliferator-activated receptor γ inhibition prevents adhesion to the extracellular matrix and induces anoikis in hepatocellular carcinoma cells," *Cancer Research*, vol. 65, no. 6, pp. 2251–2259, 2005.
- [140] M. A. Lea, M. Sura, and C. Desbordes, "Inhibition of cell proliferation by potential peroxisome proliferator-activated receptor (PPAR) gamma agonists and antagonists," *Anticancer Research*, vol. 24, no. 5A, pp. 2765–2771, 2004.
- [141] L. Nonn, L. Peng, D. Feldman, and D. M. Peehl, "Inhibition of p38 by vitamin D reduces interleukin-6 production in normal

prostate cells via mitogen-activated protein kinase phosphatase 5: implications for prostate cancer prevention by vitamin D," *Cancer Research*, vol. 66, no. 8, pp. 4516–4524, 2006.

- [142] B.-Y. Bao, J. Yao, and Y.-F. Lee, "1α, 25-dihydroxyvitamin D₃ suppresses interleukin-8-mediated prostate cancer cell angiogenesis," *Carcinogenesis*, vol. 27, no. 9, pp. 1883–1893, 2006.
- [143] J. Moreno, A. V. Krishnan, S. Swami, L. Nonn, D. M. Peehl, and D. Feldman, "Regulation of prostaglandin metabolism by calcitriol attenuates growth stimulation in prostate cancer cells," *Cancer Research*, vol. 65, no. 17, pp. 7917–7925, 2005.
- [144] N. Wagner, K.-D. Wagner, G. Schley, L. Badiali, H. Theres, and H. Scholz, "1,25-Dihydroxyvitamin D3-induced apoptosis of retinoblastoma cells is associated with reciprocal changes of Bcl-2 and bax," *Experimental Eye Research*, vol. 77, no. 1, pp. 1–9, 2003.
- [145] S. Kizildag, H. Ates, and S. Kizildag, "Treatment of K562 cells with 1,25-dihydroxyvitamin D3 induces distinct alterations in the expression of apoptosis-related genes BCL2, BAX, BCLXL, and p21," *Annals of Hematology*, vol. 89, no. 1, pp. 1–7, 2010.
- [146] M. Peterlik, W. B. Grant, and H. S. Cross, "Calcium, vitamin D and cancer," *Anticancer Research*, vol. 29, no. 9, pp. 3687–3698, 2009.
- [147] G. E. Weitsman, R. Koren, E. Zuck, C. Rotem, U. A. Liberman, and A. Ravid, "Vitamin D sensitizes breast cancer cells to the action of H_2O_2 : mitochondria as a convergence point in the death pathway," *Free Radical Biology and Medicine*, vol. 39, no. 2, pp. 266–278, 2005.
- [148] A. Ravid and R. Koren, "The role of reactive oxygen species in the anticancer activity of vitamin D," in *Vitamin D Analogs* in Cancer Prevention and Therapy, vol. 164 of Recent Results in Cancer Research, pp. 357–367, Springer, Berlin, Germany, 2003.
- [149] P. De Haes, M. Garmyn, H. Degreef, K. Vantieghem, R. Bouillon, and S. Segaert, "1,25-Dihydroxyvitamin D3 inhibits ultraviolet B-induced apoptosis, Jun kinase activation, and interleukin-6 production in primary human keratinocytes," *Journal of Cellular Biochemistry*, vol. 89, no. 4, pp. 663–673, 2003.
- [150] R. Riachy, B. Vandewalle, E. Moerman et al., "1,25-Dihydroxyvitamin D₃ protects human pancreatic islets against cytokineinduced apoptosis via down-regulation of the Fas receptor," *Apoptosis*, vol. 11, no. 2, pp. 151–159, 2006.
- [151] O. Flores, Z. Wang, K. E. Knudsen, and K. L. Burnstein, "Nuclear targeting of cyclin-dependent kinase 2 reveals essential roles of cyclin-dependent kinase 2 localization and cyclin E in vitamin D-mediated growth inhibition," *Endocrinology*, vol. 151, no. 3, pp. 896–908, 2010.
- [152] S. S. Jensen, M. W. Madsen, J. Lukas, L. Binderup, and J. Bartek, "Inhibitory effects of 1α,25-dihydroxyvitamin D₃ on the G₁–S phase-controlling machinery," *Molecular Endocrinology*, vol. 15, no. 8, pp. 1370–1380, 2001.
- [153] G. Hager, J. Kornfehl, B. Knerer, G. Weigel, and M. Formanek, "Molecular analysis of p21 promoter activity isolated from squamous carcinoma cell lines of the head and neck under the influence of 1,25(OH)₂ vitamin D3 and its analogs," *Acta Oto-Laryngologica*, vol. 124, no. 1, pp. 90–96, 2004.
- [154] P. A. Hershberger, R. A. Modzelewski, Z. R. Shurin, R. M. Rueger, D. L. Trump, and C. S. Johnson, "1,25-dihydroxycholecalciferol (1,25-D3) inhibits the growth of squamous cell carcinoma and down-modulates p21(Waf1/Cip1) in vitro and in vivo," *Cancer Research*, vol. 59, no. 11, pp. 2644–2649, 1999.
- [155] K. Colston, M. J. Colston, and D. Feldman, "1,25-Dihydroxyvitamin D₃ and malignant melanoma: the presence of receptors

and inhibition of cell growth in culture," *Endocrinology*, vol. 108, no. 3, pp. 1083–1086, 1981.

- [156] B. J. Boyle, X.-Y. Zhao, P. Cohen, and D. Feldman, "Insulinlike growth factor binding protein-3 mediates 1α,25dihydroxyvitamin D3 growth inhibition in the LNCaP prostate cancer cell line through p21/WAF1," *The Journal of Urology*, vol. 165, no. 4, pp. 1319–1324, 2001.
- [157] J. Welsh, "Cellular and molecular effects of vitamin D on carcinogenesis," *Archives of Biochemistry and Biophysics*, vol. 523, no. 1, pp. 107–114, 2012.
- [158] J. N. P. Rohan and N. L. Weigel, "1 α ,25-dihydroxyvitamin D₃ reduces c-Myc expression, inhibiting proliferation and causing G₁ accumulation in C4-2 prostate cancer cells," *Endocrinology*, vol. 150, no. 5, pp. 2046–2054, 2009.
- [159] X. Wang, S. Pesakhov, A. Weng et al., "ERK 5/MAPK pathway has a major role in 1α,25-(OH)2 vitamin D3-induced terminal differentiation of myeloid leukemia cells," *The Journal of Steroid Biochemistry and Molecular Biology*, vol. 144, pp. 223–227, 2014.
- [160] N. Pendás-Franco, J. M. González-Sancho, Y. Suárez et al., "Vitamin D regulates the phenotype of human breast cancer cells," *Differentiation*, vol. 75, no. 3, pp. 193–207, 2007.
- [161] E. Gocek and G. P. Studzinski, "Vitamin D and differentiation in cancer," *Critical Reviews in Clinical Laboratory Sciences*, vol. 46, no. 4, pp. 190–209, 2009.
- [162] M. Liu, M.-H. Lee, M. Cohen, M. Bommakanti, and L. P. Freedman, "Transcriptional activation of the Cdk inhibitor p21 by vitamin D₃ leads to the induced differentiation of the myelomonocytic cell line U937," *Genes & Development*, vol. 10, no. 2, pp. 142–153, 1996.
- [163] F. Pereira, M. J. Larriba, and A. Muñoz, "Vitamin D and colon cancer," *Endocrine-Related Cancer*, vol. 19, no. 3, pp. R51–R71, 2012.
- [164] H. G. Pálmer, M. J. Larriba, J. M. García et al., "The transcription factor SNAIL represses vitamin D receptor expression and responsiveness in human colon cancer," *Nature Medicine*, vol. 10, no. 9, pp. 917–919, 2004.
- [165] K. K. Deeb, D. L. Trump, and C. S. Johnson, "Vitamin D signalling pathways in cancer: potential for anticancer therapeutics," *Nature Reviews Cancer*, vol. 7, no. 9, pp. 684–700, 2007.
- [166] S. Upadhyay, A. Verone, S. Shoemaker et al., "1,25-dihydroxyvitamin D_3 (1,25(OH)₂ D_3) signaling capacity and the epithelialmesenchymal transition in non-small cell lung cancer (NSCLC): implications for use of 1,25(OH)₂D3 in NSCLC treatment," *Cancers*, vol. 5, no. 4, pp. 1504–1521, 2013.
- [167] R. Fukuda, B. Kelly, and G. L. Semenza, "Vascular endothelial growth factor gene expression in colon cancer cells exposed to prostaglandin E2 is mediated by hypoxia-inducible factor 1," *Cancer Research*, vol. 63, no. 9, pp. 2330–2334, 2003.
- [168] N. I. Fernandez-Garcia, H. G. Palmer, M. Garcia et al., " 1_{α} ,25-Dihydroxyvitamin D3 regulates the expression of Id1 and Id2 genes and the angiogenic phenotype of human colon carcinoma cells," *Oncogene*, vol. 24, no. 43, pp. 6533–6544, 2005.
- [169] M. J. Levine and D. Teegarden, "1α,25-dihydroxycholecalciferol increases the expression of vascular endothelial growth factor in C3H10T1/2 mouse embryo fibroblasts," *The Journal of Nutrition*, vol. 134, no. 9, pp. 2244–2250, 2004.
- [170] R. Lin, N. Amizuka, T. Sasaki et al., "1α,25-dihydroxyvitamin D3 promotes vascularization of the chondro-osseous junction by stimulating expression of vascular endothelial growth factor and matrix metalloproteinase 9," *Journal of Bone and Mineral Research*, vol. 17, no. 9, pp. 1604–1612, 2002.

- [171] M. Grundmann, M. Haidar, S. Placzko et al., "Vitamin D improves the angiogenic properties of endothelial progenitor cells," *American Journal of Physiology—Cell Physiology*, vol. 303, no. 9, pp. C954–C962, 2012.
- [172] M. J. Campbell, E. Elstner, S. Holden, M. Uskokovic, and H. P. Koeffler, "Inhibition of proliferation of prostate cancer cells by a 19-nor- hexafluoride vitamin D3 analogue involves the induction of p21(waf1) p27(kip1) and E-cadherin," *Journal of Molecular Endocrinology*, vol. 19, no. 1, pp. 15–27, 1997.
- [173] Y. Ma, W.-D. Yu, B. Su et al., "Regulation of motility, invasion, and metastatic potential of squamous cell carcinoma by 1α,25dihydroxycholecalciferol," *Cancer*, vol. 119, no. 3, pp. 563–574, 2013.
- [174] J.-W. Hsu, S. Yasmin-Karim, M. R. King et al., "Suppression of prostate cancer cell rolling and adhesion to endothelium by 1α,25-dihydroxyvitamin D₃," *The American Journal of Pathol*ogy, vol. 178, no. 2, pp. 872–880, 2011.
- [175] J. M. González-Sancho, M. Alvarez-Dolado, and A. Muñoz, "1,25-dihydroxyvitamin D3 inhibits tenascin-C expression in mammary epithelial cells," *FEBS Letters*, vol. 426, no. 2, pp. 225– 228, 1998.
- [176] V. Sung and D. Feldman, "1,25-Dihydroxyvitamin D3 decreases human prostate cancer cell adhesion and migration," *Molecular* and Cellular Endocrinology, vol. 164, no. 1-2, pp. 133–143, 2000.
- [177] H.-W. Lo, S.-C. Hsu, W. Xia et al., "Epidermal growth factor receptor cooperates with signal transducer and activator of transcription 3 to induce epithelial-mesenchymal transition in cancer cells via up-regulation of TWIST gene expression," *Cancer Research*, vol. 67, no. 19, pp. 9066–9076, 2007.
- [178] N. J. Sullivan, A. K. Sasser, A. E. Axel et al., "Interleukin-6 induces an epithelial-mesenchymal transition phenotype in human breast cancer cells," *Oncogene*, vol. 28, no. 33, pp. 2940– 2947, 2009.
- [179] Y. Wu, J. Deng, P. G. Rychahou, S. Qiu, B. M. Evers, and B. P. Zhou, "Stabilization of snail by NF-κB is required for inflammation-induced cell migration and invasion," *Cancer Cell*, vol. 15, no. 5, pp. 416–428, 2009.
- [180] K.-C. Chiang, C.-N. Yeh, J.-T. Hsu et al., "The vitamin D analog, MART-10, represses metastasis potential via downregulation of epithelial-mesenchymal transition in pancreatic cancer cells," *Cancer Letters*, vol. 354, no. 2, pp. 235–244, 2014.
- [181] K.-C. Chiang, S.-F. Kuo, C.-H. Chen et al., "MART-10, the vitamin D analog, is a potent drug to inhibit anaplastic thyroid cancer cell metastatic potential," *Cancer Letters*, vol. 369, no. 1, pp. 76–85, 2015.
- [182] K. Koli and J. Keski-Oja, "1α,25-dihydroxyvitamin D₃ and its analogues down-regulate cell invasion-associated proteases in cultured malignant cells," *Cell Growth and Differentiation*, vol. 11, no. 4, pp. 221–229, 2000.
- [183] B.-Y. Bao, S.-D. Yeh, and Y.-F. Lee, "1alpha,25-dihydroxyvitamin D3 inhibits prostate cancer cell invasion via modulation of selective proteases," *Carcinogenesis*, vol. 27, no. 1, pp. 32–42, 2006.
- [184] A. A. Ajibade, J. S. Kirk, E. Karasik et al., "Early growth inhibition is followed by increased metastatic disease with vitamin D (Calcitriol) treatment in the TRAMP model of prostate cancer," *PLoS ONE*, vol. 9, no. 2, Article ID e89555, 2014.
- [185] S. Alvarez-Díaz, N. Valle, G. Ferrer-Mayorga et al., "MicroRNA-22 is induced by vitamin D and contributes to its antiproliferative, antimigratory and gene regulatory effects in colon cancer cells," *Human Molecular Genetics*, vol. 21, no. 10, pp. 2157–2165, 2012.

- [186] S. K. R. Padi, Q. Zhang, Y. M. Rustum, C. Morrison, and B. Guo, "MicroRNA-627 mediates the epigenetic mechanisms of vitamin d to suppress proliferation of human colorectal cancer cells and growth of xenograft tumors in mice," *Gastroenterology*, vol. 145, no. 2, pp. 437–446, 2013.
- [187] X. Wang, E. Gocek, C.-G. Liu, and G. P. Studzinski, "MicroRNAs181 regulate the expression of p27Kip1 in human myeloid leukemia cells induced to differentiate by 1,25-dihydroxyvitamin D3," *Cell Cycle*, vol. 8, no. 5, pp. 736–741, 2009.
- [188] E. Gocek, X. Wang, X. Liu, C.-G. Liu, and G. P. Studzinski, "MicroRNA-32 upregulation by 1,25-dihydroxyvitamin D 3in human myeloid leukemia cells leads to bim targeting and inhibition of AraC-induced apoptosis," *Cancer Research*, vol. 71, no. 19, pp. 6230–6239, 2011.
- [189] H.-J. Ting, J. Messing, S. Yasmin-Karim, and Y.-F. Lee, "Identification of microRNA-98 as a therapeutic target inhibiting prostate cancer growth and a biomarker induced by vitamin D," *The Journal of Biological Chemistry*, vol. 288, no. 1, pp. 1–9, 2013.
- [190] M. F. Bijlsma, C. A. Spek, D. Zivkovic, S. Van De Water, F. Rezaee, and M. P. Peppelenbosch, "Repression of smoothened by patched-dependent (pro-)vitamin D3 secretion," *PLoS Biol*ogy, vol. 4, no. 8, pp. 1397–1410, 2006.
- [191] K. Fujiki, F. Kano, K. Shiota, and M. Murata, "Expression of the peroxisome proliferator activated receptor gamma gene is repressed by DNA methylation in visceral adipose tissue of mouse models of diabetes," *BMC Biology*, vol. 7, article 38, 2009.
- [192] S. A. Abedin, C. M. Banwell, K. W. Colston, C. Carlberg, and M. J. Campbell, "Epigenetic corruption of VDR signalling in malignancy," *Anticancer Research*, vol. 26, no. 4, pp. 2557–2566, 2006.
- [193] J. Höbaus, I. S. Fetahu, M. Khorchide, T. Manhardt, and E. Kallay, "Epigenetic regulation of the 1,25-dihydroxyvitamin D3 24-hydroxylase (CYP24A1) in colon cancer cells," *Journal of Steroid Biochemistry and Molecular Biology*, vol. 136, no. 1, pp. 296–299, 2013.
- [194] J. Höbaus, D. M. Hummel, U. Thiem et al., "Increased copynumber and not DNA hypomethylation causes overexpression of the candidate proto-oncogene CYP24A1 in colorectal cancer," *International Journal of Cancer*, vol. 133, no. 6, pp. 1380–1388, 2013.
- [195] D. Karolchik, G. P. Barber, J. Casper et al., "The UCSC genome browser database: 2014 update," *Nucleic Acids Research*, vol. 42, no. 1, pp. D764–D770, 2014.
- [196] R. Marik, M. Fackler, E. Gabrielson et al., "DNA methylationrelated vitamin D receptor insensitivity in breast cancer," *Cancer Biology & Therapy*, vol. 10, no. 1, pp. 44–53, 2010.
- [197] C. A. Godman, R. Joshi, B. R. Tierney et al., "HDAC3 impacts multiple oncogenic pathways in colon cancer cells with effects on Wnt and vitamin D signaling," *Cancer Biology & Therapy*, vol. 7, no. 10, pp. 1570–1580, 2014.
- [198] H. Zhu, X. Wang, H. Shi et al., "A genome-wide methylation study of severe vitamin D deficiency in African American adolescents," *Journal of Pediatrics*, vol. 162, no. 5, pp. 1004– 1009.el, 2013.
- [199] H. Shi, P. S. Yan, C.-M. Chen et al., "Expressed CpG island sequence tag microarray for dual screening of DNA hypermethylation and gene silencing in cancer cells," *Cancer Research*, vol. 62, no. 11, pp. 3214–3220, 2002.
- [200] B. Novakovic, M. Sibson, H. K. Ng et al., "Placenta-specific methylation of the vitamin D 24-hydroxylase gene. Implications

for feedback autoregulation of active vitamin D levels at the fetomaternal interface," *The Journal of Biological Chemistry*, vol. 284, no. 22, pp. 14838–14848, 2009.

- [201] H. Shi, J. Guo, D. J. Duff et al., "Discovery of novel epigenetic markers in non-Hodgkin's lymphoma," *Carcinogenesis*, vol. 28, no. 1, pp. 60–70, 2007.
- [202] M. Wjst, I. Heimbeck, D. Kutschke, and K. Pukelsheim, "Epigenetic regulation of vitamin D converting enzymes," *Journal of Steroid Biochemistry and Molecular Biology*, vol. 121, no. 1-2, pp. 80–83, 2010.
- [203] J.-Y. Hsu, D. Feldman, J. E. McNeal, and D. M. Peehl, "Reduced 1α -hydroxylase activity in human prostate cancer cells correlates with decreased susceptibility to 25-hydroxyvitamin D_3 -induced growth inhibition," *Cancer Research*, vol. 61, no. 7, pp. 2852–2856, 2001.
- [204] M. Tannour-Louet, S. K. Lewis, J.-F. Louet et al., "Increased expression of CYP24A1 correlates with advanced stages of prostate cancer and can cause resistance to vitamin D3-based therapies," *The FASEB Journal*, vol. 28, no. 1, pp. 364–372, 2014.
- [205] L. Sabatino, A. Fucci, M. Pancione, and V. Colantuoni, "PPARG epigenetic deregulation and its role in colorectal tumorigenesis," *PPAR Research*, vol. 2012, Article ID 687492, 12 pages, 2012.
- [206] T. W. Dunlop, S. Väisänen, C. Frank, F. Molnár, L. Sinkkonen, and C. Carlberg, "The human peroxisome proliferator-activated receptor δ gene is a primary target of 1 α ,25-dihydroxyvitamin D3 and its nuclear receptor," *Journal of Molecular Biology*, vol. 349, no. 2, pp. 248–260, 2005.
- [207] F. Alimirah, X. Peng, L. Yuan et al., "Crosstalk between the peroxisome proliferator-activated receptor γ (PPAR γ) and the vitamin D receptor (VDR) in human breast cancer cells: PPAR γ binds to VDR and inhibits 1 α ,25-dihydroxyvitamin D3 mediated transactivation," *Experimental Cell Research*, vol. 318, no. 19, pp. 2490–2497, 2012.
- [208] Y. Shi, M. Hon, and R. M. Evans, "The peroxisome proliferatoractivated receptor δ , an integrator of transcriptional repression and nuclear receptor signaling," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 5, pp. 2613–2618, 2002.
- [209] R. J. Wood, "Vitamin D and adipogenesis: new molecular insights," *Nutrition Reviews*, vol. 66, no. 1, pp. 40–46, 2008.
- [210] C. J. Narvaez, K. M. Simmons, J. Brunton, A. Salinero, S. V. Chittur, and J. E. Welsh, "Induction of STEAP4 correlates with 1,25-dihydroxyvitamin D3 stimulation of adipogenesis in mesenchymal progenitor cells derived from human adipose tissue," *Journal of Cellular Physiology*, vol. 228, no. 10, pp. 2024–2036, 2013.
- [211] R. Bouillon, G. Carmeliet, L. Lieben et al., "Vitamin D and energy homeostasis—of mice and men," *Nature Reviews Endocrinology*, vol. 10, no. 2, pp. 79–87, 2014.
- [212] P. Sertznig, T. Dunlop, M. Seifert, W. Tilgen, and J. Reichrath, "Cross-talk between Vitamin D Receptor (VDR)- and Peroxisome Proliferator-activated Receptor (PPAR)-signaling in melanoma cells," *Anticancer Research*, vol. 29, no. 9, pp. 3647– 3658, 2009.
- [213] P. Sertznig, M. Seifert, W. Tilgen, and J. Reichrath, "Peroxisome proliferator-activated receptor (PPAR) and vitamin D receptor (VDR) signaling pathways in melanoma cells: promising new therapeutic targets?" *Journal of Steroid Biochemistry and Molecular Biology*, vol. 121, no. 1-2, pp. 383–386, 2010.
- [214] T. L. Van Belle, C. Gysemans, and C. Mathieu, "Vitamin D and diabetes: the odd couple," *Trends in Endocrinology and Metabolism*, vol. 24, no. 11, pp. 561–568, 2013.

- [215] K. S. Vimaleswaran, D. J. Berry, C. Lu et al., "Causal relationship between obesity and vitamin D status: bi-directional mendelian randomization analysis of multiple cohorts," *PLoS Medicine*, vol. 10, no. 2, Article ID e1001383, 2013.
- [216] J. Auwerx, R. Bouillon, and H. Kesteloot, "Relation between 25-hydroxyvitamin D3, apolipoprotein A-I, and high density lipoprotein cholesterol," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 12, no. 6, pp. 671–674, 1992.
- [217] S. Kayaniyil, S. B. Harris, R. Retnakaran et al., "Prospective association of 25(OH)D with metabolic syndrome," *Clinical Endocrinology*, vol. 80, no. 4, pp. 502–507, 2014.
- [218] Y. Yoon, "Anti-adipogenic effects of 1,25-dihydroxyvitamin D3 are mediated by the maintenance of the wingless-type MMTV integration site/β-catenin pathway," *International Journal of Molecular Medicine*, vol. 30, no. 5, pp. 1219–1224, 2012.
- [219] J. M. Blumberg, I. Tzameli, I. Astapova, F. S. Lam, J. S. Flier, and A. N. Hollenberg, "Complex role of the vitamin D receptor and its ligand in adipogenesis in 3T3-L1 cells," *The Journal of Biological Chemistry*, vol. 281, no. 16, pp. 11205–11213, 2006.
- [220] M. MacIas-Gonzalez, I. Moreno-Santos, J. M. García-Almeida, F. J. Tinahones, and E. Garcia-Fuentes, "PPARy2 protects against obesity by means of a mechanism that mediates insulin resistance," *European Journal of Clinical Investigation*, vol. 39, no. 11, pp. 972–979, 2009.
- [221] J. B. Rawson, Z. Sun, E. Dicks et al., "Vitamin D intake is negatively associated with promoter methylation of the Wnt antagonist gene DKK1 in a large group of colorectal cancer patients," *Nutrition and Cancer*, vol. 64, no. 7, pp. 919–928, 2012.
- [222] J. B. Rawson, M. Manno, M. Mrkonjic et al., "Promoter methylation of Wnt antagonists DKK1 and SFRP1 is associated with opposing tumor subtypes in two large populations of colorectal cancer patients," *Carcinogenesis*, vol. 32, no. 5, pp. 741–747, 2011.
- [223] J. Hoffstedt, P. Arner, G. Hellers, and F. Lönnqvist, "Variation in adrenergic regulation of lipolysis between omental and subcutaneous adipocytes from obese and non-obese men," *Journal of Lipid Research*, vol. 38, no. 4, pp. 795–804, 1997.
- [224] M. Bhat, R. Kalam, S. Syh Qadri, S. Madabushi, and A. Ismail, "Vitamin D deficiency-induced muscle wasting occurs through the ubiquitin proteasome pathway and is partially corrected by calcium in male rats," *Endocrinology*, vol. 154, no. 11, pp. 4018– 4029, 2013.
- [225] G. J. Hausman, S. P. Poulos, T. D. Pringle, and M. J. Azain, "The influence of thiazolidinediones on adipogenesis in vitro and in vivo: potential modifiers of intramuscular adipose tissue deposition in meat animals," *Journal of Animal Science*, vol. 86, no. 14, pp. E236–E243, 2008.
- [226] Y.-X. Wang, C.-L. Zhang, R. T. Yu et al., "Regulation of muscle fiber type and running endurance by PPARδ," *PLoS Biology*, vol. 2, no. 10, article e294, 2004.





The Scientific World Journal



Research and Practice









Computational and Mathematical Methods in Medicine

Behavioural Neurology





Oxidative Medicine and Cellular Longevity