

Research Article

Impact Assessment of Cadmium Toxicity and Its Bioavailability in Human Cell Lines (Caco-2 and HL-7702)

Rukhsanda Aziz,¹ M. T. Rafiq,^{1,2} Jie Yang,³ Di Liu,¹ Lingli Lu,¹ Zhenli He,⁴ M. K. Daud,⁵ Tingqiang Li,¹ and Xiaoe Yang¹

¹ Ministry of Education Key, Laboratory of Environmental Remediation and Ecological Health, College of Environmental and Resource Sciences, Zhejiang University, Hangzhou 310058, China

² Department of Environmental Sciences, International Islamic University, Islamabad 44000, Pakistan

³ College of Architecture and Environment, Sichuan University, Chengdu 610065, China

⁴ Indian River Research and Education Center, Institute of Food and Agricultural Sciences, University of Florida, Fort Pierce, FL 34945, USA

⁵ Department of Biotechnology & Genetic Engineering, Kohat University of Science and Technology, Kohat 26000, Pakistan

Correspondence should be addressed to Tingqiang Li; litq@zju.edu.cn and Xiaoe Yang; xyang571@yahoo.com

Received 21 August 2013; Revised 23 October 2013; Accepted 9 November 2013; Published 16 February 2014

Academic Editor: Qaisar Mahmood

Copyright © 2014 Rukhsanda Aziz et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Cadmium (Cd) is a widespread environmental toxic contaminant, which causes serious health-related problems. In this study, human intestinal cell line (Caco-2 cells) and normal human liver cell line (HL-7702 cells) were used to investigate the toxicity and bioavailability of Cd to both cell lines and to validate these cell lines as *in vitro* models for studying Cd accumulation and toxicity in human intestine and liver. Results showed that Cd uptake by both cell lines increased in a dose-dependent manner and its uptake by Caco-2 cells ($720.15 \mu\text{g mg}^{-1}$ cell protein) was significantly higher than HL-7702 cells ($229.01 \mu\text{g mg}^{-1}$ cell protein) at 10 mg L^{-1} . A time- and dose-dependent effect of Cd on cytotoxicity assays (LDH release, MTT assay) was observed in both Cd-treated cell lines. The activities of antioxidant enzymes and differentiation markers (SOD, GPX, and AKP) of the HL-7702 cells were higher than those of Caco-2 cells, although both of them decreased significantly with raising Cd levels. The results from the present study indicate that Cd above a certain level inhibits cellular antioxidant activities and HL-7702 cells are more sensitive to Cd exposure than Caco-2 cells. However, Cd concentrations $<0.5 \text{ mg L}^{-1}$ pose no toxic effects on both cell lines.

1. Introduction

Cadmium (Cd) is one of the most concerned pollutants possessing high toxicity for both animals and plants [1]. The direct exposure of living beings, particularly human populations, to it may cause various health related problems such as Itai-Itai disease [2]. Cadmium is considered as a “guest” metal in Pb:Zn mining process and is widely dumped into the environment through various anthropogenic activities such as mining, smelting, and use of fertilizers [3]. It has been classified as a group I carcinogen by International Agency for Research in Cancer (IARC) and as a probable human carcinogen (group B1) by Environmental Protection Agency (EPA) [4]. Due to its toxic nature, it is now recognized that human exposure to Cd must either be stopped or minimized.

Cadmium salts, having greater solubility in water, can easily enter into our food chain through edible parts of the plant [5]. When grown in polluted soil, plants take up water, nutrients, and heavy metals that may then enter the food chain [6]. Khan et al. [7] found that Cd concentration in soils and food crops was above permissible limits in selected vegetables. They concluded that consuming such vegetables and food crops may result in Cd toxicity to both humans and grazing animals. Human beings are mostly exposed to Cd either by inhalation or ingestion, with oral intake being the major route. Understanding of the mechanisms of Cd absorption and circumventing the intestinal barrier is of prime interest. Available literature reports that, following oral exposure, Cd is absorbed in mammals preferentially in the duodenum and proximal jejunum [8]. After absorption Cd first reaches the

liver. In liver Cd induces the production of metallothionein. After continuous hepatocyte necrosis and apoptosis, Cd-metallothionein complexes are washed into sinusoidal blood. From here, some of the absorbed Cd enters into the enterohepatic cycle via secretion into the biliary tract in the form of Cd-glutathione conjugates. Enzymatically degraded to Cd-cysteine complexes in the biliary tract, Cd again enters the small intestine [9]. It is generally assumed that, after inhalation or ingestion of typical inorganic Cd compounds and after transferring across the alveolar membranes or intestinal wall, the major fraction of Cd initially accumulates in the liver [10]. *In vitro* and *in vivo* studies have depicted that low Cd concentrations ($<5.0 \mu\text{M}$) might enhance cell proliferation in a dose-dependent manner [11, 12], while its higher concentrations ($>5.0 \mu\text{M}$) could induce apoptosis in many tissues and organs [13].

Apoptosis is enhanced in cells in most cases upon Cd exposure [14]. Various assays such as LDH leakage, total protein contents, and the MTT are commonly employed for the detection of cytotoxicity or cell viability upon exposure to toxic substances. The LDH leakage assay measures the lactate dehydrogenase activity in the extracellular culture medium upon the loss of intracellular LDH and is an indicator of cell membrane damage [15]. 3-[4,5-Dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide (MTT) is a water soluble tetrazolium salt, which is converted to an insoluble purple formazan within the mitochondria. Formazan accumulates only in healthy cells and its reduction in stressed cells has been reported [16].

Like any other environmental stress, Cd stress also causes oxidative stress in cultured cells due to the production of various reactive oxygen species (ROS) such as hydrogen peroxide, superoxide radical, and hydroxyl radical. Reactive oxygen species (ROS), normally produced during the aerobic metabolism, function as second messengers involved in many cellular functions. On the other hand, when ROS level increases because of oxidant treatments and defective antioxidant systems, these highly reactive compounds and radicals become dangerous toxic agents [17]. These ROS cause imbalance between oxidants and antioxidants of the cell tissues, which leads to the numerous degenerative diseases in humans. Either to avoid or to scavenge oxidative damage, mammalian cells possess inherent antioxidant machinery, which is comprised of superoxide dismutase (SOD), catalase, and glutathione peroxidase (GPx) [18]. However, there is scarce information regarding metabolic differences in oxidatively stressed intestinal cells compared to nonstressed cells. Due to its soluble nature, Cd can readily penetrate tissues after exposure and inhibit antioxidant enzymatic system [19] by affecting the cellular thiol redox balance [20], in particular with cellular glutathione. Several studies have shown that Cd indirectly generates ROS and consequently DNA, lipid, and protein oxidation in various cell lines [21].

The Caco-2 cell line is a human intestinal cell line and HL-7702 is normal human liver cell line that have been widely used as representative models of mammalian intestinal [22] and liver cells [23]. These cells have become useful tools for the study of uptake, toxicity, and transport of nutrients [22] and heavy metals [24]. The aims of this study were to (1)

determine the bioavailability of Cd in the Caco-2 cells and HL-7702 cells; (2) study the effect of Cd on the activities of antioxidant enzymes (SOD, GPx) and differentiation marker enzyme (AKP); and (3) assess the health risk of the Cd in human small intestine and liver.

2. Materials and Methods

2.1. Chemicals and Reagents. Cd(NO₃)₂ was purchased from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). Dulbecco's modified Eagle medium (DMEM with glucose 4.5 g L^{-1}), trypsin-EDTA, phosphate-buffered saline (PBS), fetal bovine serum (FBS), glutamine, nonessential amino acids, penicillin, and streptomycin were purchased from Gibco Life Technologies (Grand Island, NY). 3-[4,5-Dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide (MTT) was purchased from Sigma-Aldrich (St. Louis, MO, USA). SOD, LDH, AKP, and GSH-Px testing kits were obtained from Nanjing KeyGen Biotech. Co., Ltd. (Nanjing, China). All of the other chemicals used in this study were of analytical grade purchased from local chemical suppliers. All reagents were prepared with deionized water ($\leq 0.1 \mu\text{S/cm}$) using a Milli-Q system (Millipore, Billerica, MA). All laboratory glassware used in the experiments were soaked in 10% HNO₃ for 24 h and subsequently rinsed with deionized water and air-dried.

2.2. Preparation of the Cadmium Solutions. The stock solution of cadmium nitrate was made in UHQ water and sterilized using a 0.22 mm filter. A working solution in the corresponding media (1:10 v/v) was prepared to give the concentrations of 0.25, 0.5, 2, 4, 8, and 10 mg L^{-1} in the culture medium.

2.3. Cell Culture. Caco-2 cells and HL-7702 cells were obtained from the Institute of Biochemistry and Cell Biology (SIBS, CAS, Shanghai, China) and used in assays at passage 20–33. The Caco-2 and HL-7702 cells were normally cultured in 25 cm^2 flasks (Corning Inc., NY, USA) and maintained in high glucose (4.5 g L^{-1}) DMEM, supplemented with 10% (v/v) fetal bovine serum, 1% (v/v) nonessential amino acids, 4 mM L-glutamine, and 1% (v/v) antibiotic solution (penicillin-streptomycin). The cells were maintained at 37°C in an incubator (Heraeus, BB15, Germany) with 5% CO₂ and 95% relative humidity. After being 80% confluent, the cells were washed with phosphate-buffered saline (PBS) to remove any unattached cells. The attached cells were then harvested using a trypsin-ethylenediaminetetraacetic acid (EDTA) solution. The cells were seeded in 6-well plates at a density of approximately 10×10^4 cells/mL. The medium was changed every 2 days, and the cultures were maintained for 13 days to reach the stationary growth phase and to allow for maximal functional differentiation. Confluent cultures of differentiated cells were used for further studies and were exposed to known concentrations of Cd in the culture medium and incubated for 2 and 12 h at 13 days after seeding. Cytological and biochemical assays were carried out on the cell lysate and culture medium for determination of markers of cell damage.

2.4. Cd Uptake. To assess the uptake of $\text{Cd}(\text{NO}_3)_2$, Caco-2 cells and HL-7702 cells, cultured in 6-well plates, were exposed to different concentrations of $\text{Cd}(\text{NO}_3)_2$ solution (0.25, 0.5, 2, 4, 8, and 10 mg L^{-1}), for 2 h at 37°C . Then the cells were washed twice with ice cold PBS to remove extracellular bound Cd. The cell monolayer was lysed by the addition of 2 mL of deionized water in the well and then harvested. The lysate was put into the digestion tubes and digested with HNO_3 (5 mL) and H_2O_2 (1 mL). After cooling the resultant solutions were diluted to 10 mL using deionized water. The concentrations of Cd in the final solution were determined by ICP-MS (Agilent 7500a, Agilent Technologies, CA, USA) following a standard procedure.

2.5. Cytotoxicity Assays

2.5.1. MTT Assay. Cytotoxicity induced by Cd was assessed by MTT assay. Caco-2 cells and HL-7702 cells were seeded at a density of 5×10^4 cells/well in a 96-well plate and incubated for 24 h. Cells were exposed to increasing concentrations of Cd (6 wells per concentration group plus 1 control group) for 2 h or 12 h at 37°C in an atmosphere of 5% CO_2 in air. Control groups consist of cells in media (minus chemical), which are processed identically and incubated simultaneously as treated groups. After various time intervals (2 and 12 h), medium was removed and replaced with $50 \mu\text{L}$ MTT solution (2 mg mL^{-1}) for another 4 h after which medium was removed. The medium is then replaced with $100 \mu\text{L}$ dimethyl sulfoxide (DMSO), agitated for 5 min at room temperature. Finally, the absorbance was measured at 552 nm using a microplate reader (Bio-Rad-680, Bio-Rad, USA). Cell viability is expressed as a percentage of the control group. Cell MTT response (% control) was calculated from the equation:

$$\% \text{ control} = \frac{\text{Absorbance}_{\text{treatment}}}{\text{Absorbance}_{\text{control}}} \times 100\%. \quad (1)$$

2.5.2. Lactate Dehydrogenase (LDH) Release. Cytotoxicity induced by Cd was assessed by lactate dehydrogenase (LDH) leakage into the culture medium. The activity of LDH in the medium was determined using a commercially available kit from Jiancheng Biochemical Co., Ltd. (Nanjing, China). The LDH assay is based on the conversion of lactate to pyruvate in the presence of LDH with parallel reduction of NAD to NADH. The change in the absorbance was recorded at 440 nm using a microplate spectrophotometer system (Bio-Rad-680, Bio-Rad, USA). Cell LDH release (% control) was calculated from the equation:

$$\% \text{ control} = \frac{(\text{U LDH/mg cell protein})_{\text{treatment}}}{(\text{U LDH/mg cell protein})_{\text{control}}} \times 100\%. \quad (2)$$

2.6. Enzyme Assays

2.6.1. Glutathione Peroxidase (GPx). The cell glutathione peroxidase (GPx) activity was measured using the GPx cellular activity assay kit from Jiancheng Biochemical Co.,

Ltd. (Nanjing, China). This kit uses an indirect method, based on the oxidation of reduced glutathione (GSH) to oxidized glutathione (GSSG) catalyzed by GPx, which is then coupled with recycling GSSG back to GSH utilizing glutathione reductase and NADPH. The decrease in NADPH at 412 nm during oxidation of NADPH to NADP is indicative of GSH-Px activity. GSH-Px activity was expressed in terms of international units per mg of soluble cell proteins.

2.6.2. Superoxide Dismutase (SOD). The cell superoxide dismutase (SOD) activity was measured in Caco-2 cells and HL-7702 cells using the SOD cellular activity assay kit from Jiancheng Biochemical Co., Ltd. (Nanjing, China) based on the competition between pyrogallol oxidation by superoxide radicals and superoxide dismutation by SOD. The absorbance was measured at 550 nm using a microplate reader (Bio-Rad-680, Bio-Rad, USA). The SOD's one unit activity is defined as the amount of the enzyme required to prohibit the rate of pyrogallol autooxidation by 50%. SOD activity was expressed as international units per mg of soluble cell proteins.

2.6.3. Alkaline Phosphatase. The alkaline phosphatase activity was measured by the alkaline phosphatase testing kit. This assay is based on measuring the alkaline phosphatase activity by monitoring the color change as paranitrophenol phosphate which is colorless is converted to paranitrophenol + phosphate which is yellow. Activity was determined using a microplate reader (Bio-Rad-680, Bio-Rad, USA) at 520 nm.

2.7. Protein Assay. Bradford [25] assay was performed in order to measure cell protein in Caco-2 cells and HL-7702 cells. Its absorbance was measured at 595 nm with a spectrophotometer (VersaMax, Molecular Devices, Sunnyvale, CA, USA).

2.8. Statistical Analyses. Data were analyzed using the SPSS 18.0 (SPSS Inc., Chicago, USA). Results are presented as mean \pm S.D. (standard deviation). Analysis of variance (ANOVA) was performed with the least significant difference (LSD) to compare means of different Cd concentrations with control. For comparison of the different cell lines, univariate ANOVA with LSD post hoc test was used [26]. Means were considered to be significantly different if P values were <0.05 .

3. Results and Discussion

In recent years, heavy metal pollution, particularly cadmium, lead, arsenic, and chromium [1], has threatened living beings, causing adverse effects on human health such as renal and testicular dysfunction and pulmonary problems. In the present study, the cytotoxic effects of Cd on a normal human liver cell line (HL-7702 cells) and human intestinal cell line (Caco-2 cells) were evaluated by studying Cd uptake by both cell lines, cytotoxicity assays, ROS scavenger enzymes, and differentiation marker enzyme (AKP).

3.1. Cellular Cd Uptake. Cd accumulation in both cell lines was found significantly different due to difference in the

origin of cell lines. The Cd uptake by both cell lines is shown in Figure 1. Dose-dependent rise in Cd accumulation in Caco-2 cells and HL-7702 cells could be noticed ranging from 52.3 to 720.1 and from 11.3 to 229.0 $\mu\text{g mg}^{-1}$ of cellular protein, respectively. Increasing Cd levels in the incubation medium were significantly ($P < 0.05$) correlated with rising cellular Cd levels. It was also observed that the uptake of the Cd at 10 mg L^{-1} by Caco-2 cells ($720.15 \mu\text{g mg}^{-1}$ cell protein) is higher as compared to HL-7702 cells ($229.01 \mu\text{g mg}^{-1}$ cell protein). Our results are in agreement with Templeton [27] who reported that Cd was absorbed from the gastrointestinal tract and was then taken up by the liver, the first organ after absorption which accumulates high Cd concentrations. Cd accumulation by the Caco-2 cells was augmented with the increasing Cd concentrations in the culture medium [28]. There can be various reasons regarding Cd uptake and its differential absorption in different organs of human body. Concentration-dependent Cd uptake in our present experiment might be due to its chemical and physical properties related to essential metals such as iron (Fe), zinc (Zn), or calcium (Ca). That is why Cd can be transported and taken up into the cells by a process referred to as “ionic and molecular mimicry” [29]. Intestinal absorption of Cd is characterized by a rapid rate of metal accumulation within the intestinal mucosa and a low rate of diffusive transfer into circulatory system [9]. In the present study we also found that the intestinal cells (Caco-2 cells) accumulated higher quantity of Cd as compared to the liver cells (HL-7702). There is a two-step process for the absorptive Cd movement from intestinal lumen into enterocytes: the first step consists of a nonspecific binding of Cd into the luminal plasma membrane and the second step consists of a transport across the luminal plasma membrane into enterocytes [30]. Much of the Cd absorbed in the intestines is delivered first to the liver through portal circulation, bound mainly to albumin, where it is taken up from the sinusoidal capillaries to the hepatocytes [31].

3.2. Effect of Cadmium on LDH Release and Mitochondrial Activity (MTT). The results obtained from the cytotoxicity assays indicate that there are differences between the two cell lines concerning their sensitivity to Cd exposure. HL-7702 cells appear to be more sensitive as indicated by the LDH and MTT assays (Figures 2 and 3). LDH release assay is an important method to assess cell membrane stability [13]. As shown in Figures 2(a) and 2(b), there was an increase in LDH release when Caco-2 cells and HL-7702 cells were exposed to different concentrations for 2 and 12 h. Increased concentrations and time of Cd exposure resulted in a significant increase of LDH release in culture media. At Cd concentrations greater than 0.5 mg L^{-1} , a significant ($P < 0.05$) increase in LDH release was observed for both cell lines, as compared with their respective controls. It was found that Cd dosage increased the LDH release from 99% to 178.5% in Caco-2 cells and from 100% to 198.4% in HL-7702 cells at the incubation time of 2 h. After 12 h, this increase was much larger at 10 mg L^{-1} both in Caco-2 cells (264.6%) and HL-7702 cells (250.7%). In terms of LDH release, for both cell lines, Cd levels and exposure time significantly correlate

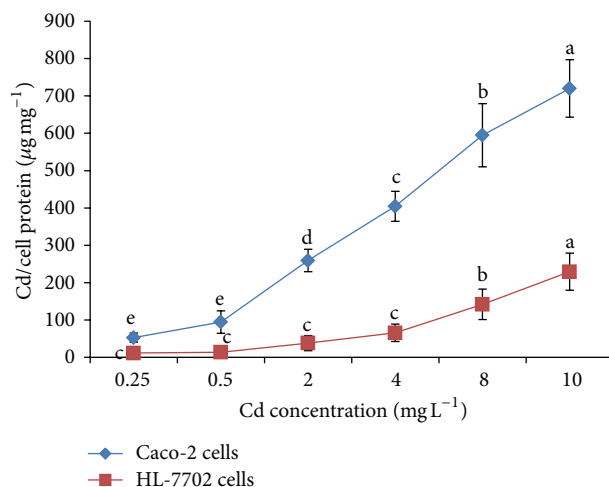


FIGURE 1: Bioavailability of cadmium (Cd) by HL-7702 and Caco-2 cells after 2 h. Values are presented as mean \pm S.D.; $n = 3$. Different letters indicate significant differences at $P < 0.05$ by the least significant difference (LSD) test.

($P < 0.05$) with each other. The large increase in LDH release might also be due to cell rounding and loss of adherence found at the concentration of 10 mg L^{-1} Cd. This noticeable increase of LDH release might be due to the increase of necrotic cell death as scavenging enzymes could be depleted by relatively high concentrations of Cd. Morphological changes (loss of adherence and cell rounding) based on 10 mg L^{-1} Cd might be associated with loss of intercellular contacts and break down of desmosomes [26].

The MTT assay is mainly based on the enzymatic conversion of MTT in the mitochondria. It has been suggested that Cd disrupts mitochondrial function both *in vivo* [32] and *in vitro* [33]. Apoptosis mediated by mitochondria may be relevant in metal-induced cell death. Mitochondrial activity was significantly decreased by increasing concentrations and time of Cd exposure to both cells (Figures 3(a), and 3(b)). MTT assay was very low ($<3\%$) in both cell lines treated with $0\text{--}0.5 \text{ mg L}^{-1}$ Cd. A significant decrease in MTT assay, compared to control cells, was noted in Caco-2 cells and HL-7702 cells subjected to Cd for 2 and 12 h. Initial cytotoxicity was observed for both cell lines at 2 mg L^{-1} Cd. Cadmium dosage decreased cell viability by 84% and 44% in Caco-2 cells and by 75% and 38% in HL-7702 cells after 2 h while the percentage further decreased after 12 h in both Caco-2 cells (74% and 26%) and HL-7702 cells (60% and 24%) under different concentrations of Cd (2 mg L^{-1} and 10 mg L^{-1}), as compared to their respective controls. These results indicated that Cd was able to induce cytotoxicity in a concentration- and time-dependent manner in the Caco-2 cells and HL-7702 cells. Similar findings were also reported by Nemmiche et al. [34] and Fotakis and Timbrell [15]. Cadmium toxicity primarily resulted from the binding of Cd to thiol groups in mitochondria, causing mitochondrial dysfunction and subsequent injury. Furthermore, Cd may enter mitochondria through Ca uniporter and interact with thiol groups

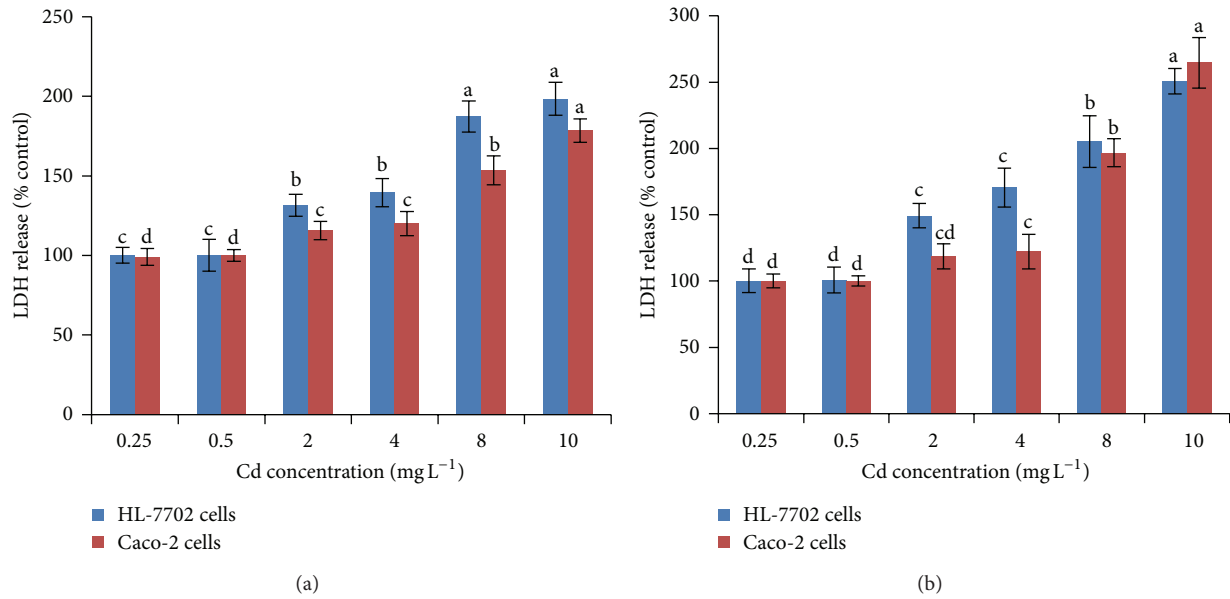


FIGURE 2: Effect of Cd on LDH release in Caco-2 and HL-7702 cells. Multiple range tests were conducted for Caco-2 and HL-7702 cells at different Cd levels after 2 h (a) and 12 h (b). Different letters indicate significant differences at $P < 0.05$ by the least significant difference (LSD) test. Values are presented as mean \pm S.D.; $n = 3$.

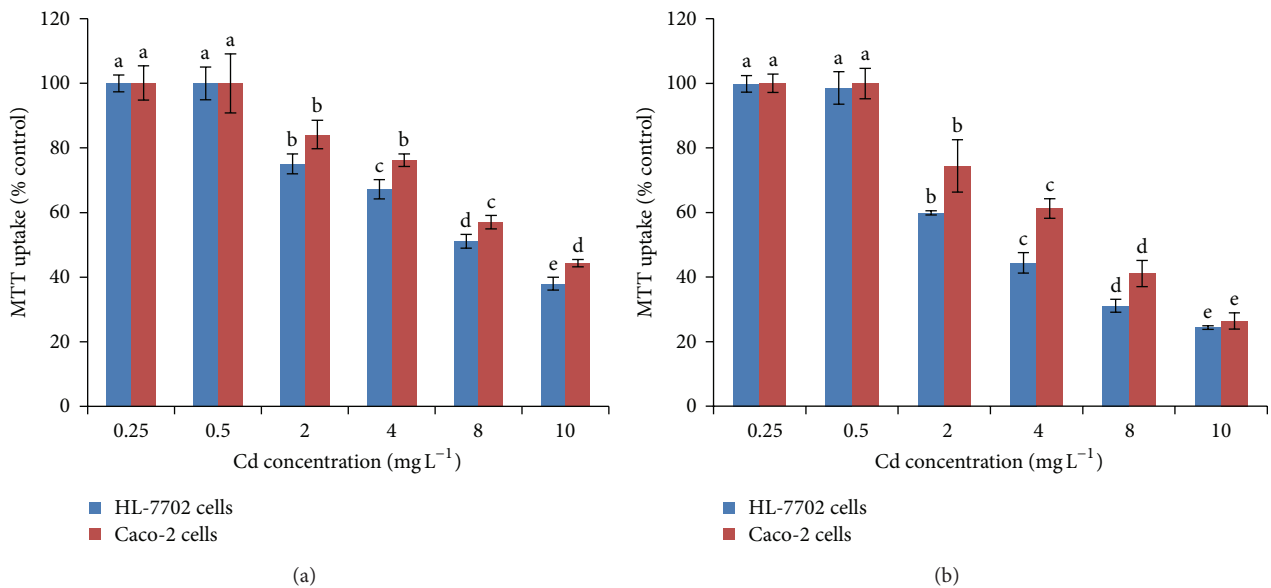


FIGURE 3: Effect of Cd on MTT assay in Caco-2 and HL-7702 cells. Multiple range tests were conducted for Caco-2 and HL-7702 cells at different Cd levels after 2 h (a) and 12 h (b). Different letters indicate significant differences at $P < 0.05$ by the least significant difference (LSD) test. Values are presented as mean \pm S.D.; $n = 3$.

of adenine nucleotide translocator (ANT) to induce the Cytochrome c release and apoptosis. Xie et al. [23] also reported that Cd-induced apoptosis was dependent on Cd dose and exposure time in both normal (HL-7702) and tumor cells (Raji cells). Furthermore they [23] observed that Cd-induced Ca elevation was attenuated in HL-7702 cells co-incubated with a Ca chelator. Therefore, Cd-induced apoptosis was mediated by the release of Ca from intracellular Ca storage but not an influx of extracellular Ca.

3.3. Effect of Cd on Antioxidant Enzymes Activity. Cadmium promotes an early oxidative stress and thus contributes to the development of serious pathological conditions because of its long retention in some tissues [35]. Physiologically, SOD is an important defense enzyme, which converts O₂ to H₂O₂ and thus protects against superoxide-induced damage [36]. GSH-Px, in particular, is highly dependent on glutathione concentration. The antioxidant enzymes (SOD, GPx) activities in Caco-2 cells and HL-7702 cells exposed to different

TABLE 1: Activities of antioxidant enzymes (SOD, GPX) and differentiation marker enzyme (AKP) in Caco-2 cells and HL-7702 cells at different Cd levels (after 2 hr exposure).

Parameters	Cell lines	Cd concentration (mg L ⁻¹)						
		0	0.25	0.5	2	4	8	10
GPx (U mg ⁻¹ cell protein)	Caco-2 cells	198.7 ± 0.7 ^a	190.9 ± 2.6 ^b	183.9 ± 3.2 ^c	133.7 ± 0.8 ^d	70.1 ± 1.1 ^e	48.09 ± 0.9 ^f	30.3 ± 0.7 ^g
	HL-7702 cells	306.9 ± 2.3 ^a	300.2 ± 8.7 ^{ab}	295.6 ± 5.3 ^b	207.2 ± 1.1 ^c	144.9 ± 0.8 ^d	100.3 ± 0.6 ^e	60.4 ± 0.6 ^f
SOD (U mg ⁻¹ cell protein)	Caco-2 cells	26.2 ± 1.8 ^a	25.1 ± 3 ^a	22.91 ± 2.4 ^{ab}	21.8 ± 0.5 ^{bc}	21.7 ± 0.9 ^{bc}	19.0 ± 2 ^{cd}	15.9 ± 0.3 ^d
	HL-7702 cells	26.0 ± 1.8 ^a	24.5 ± 0.6 ^a	24.0 ± 0.2 ^a	21.4 ± 1.0 ^b	20.03 ± 2.0 ^b	15.2 ± 0.5 ^c	11.1 ± 1.1 ^d
AKP (%Ck)	Caco-2 cells	100 ± 0.1 ^a	97.6 ± 3.3 ^b	95.3 ± 3.6 ^b	83.1 ± 0.7 ^c	79.41 ± 0.7 ^d	75.0 ± 1.0 ^e	66.1 ± 0.6 ^f
	HL-7702 cells	100 ± 0.4 ^a	95.5 ± 5.1 ^b	90.7 ± 3.7 ^c	65.32 ± 0.9 ^d	45.97 ± 1.0 ^e	28.31 ± 1.3 ^f	27.01 ± 1.0 ^f

Multiple comparison tests were for Caco-2 cells and HL-7702 cells at different Cd levels and different letters indicate significant differences at $P < 0.05$ as calculated by the least significant difference (LSD) test. Values are presented as mean ± S.D.; $n = 3$.

concentrations of Cd for 2 h are shown in Table 1. Significant changes in antioxidant enzymes activity were observed in both cell lines with the increase in Cd level, as compared with controls. Cd had no significant influence ($P > 0.05$) on SOD activity in both cell lines when the concentration was below 0.5 mg L⁻¹. However, the SOD activity was significantly decreased ($P < 0.05$) when the concentration of Cd was more than 0.5 mg L⁻¹. SOD activity of Caco-2 cells and the HL-7702 cells was decreased by 61.1% and 42.7%, respectively at 10 mg L⁻¹ Cd level, as compared to their respective controls. The results of GPx activity showed a similar trend to SOD activity. The results showed that GPx activity in Cd-treated Caco-2 cells and HL-7702 cells ranged from 96% to 15.3% and 97.8% to 19.7%, respectively at 0.25 mg L⁻¹ and 10 mg L⁻¹ from control. Our results showed that antioxidant enzyme activities influenced by Cd in both cell lines differed significantly from their controls. In response to Cd stress, the activities of SOD and GPx in HL-7702 cells were considerably higher than those of Caco-2 cells.

According to our findings, the major toxic effects of increasing doses of Cd concentration involve decreased antioxidant enzyme levels. We presume that Cd also enters the mitochondria and inhibits the activities of many enzymes by binding to their -SH groups or by inhibiting the protein synthesis [37]. Cadmium binding to cysteine in reduced glutathione (GSH) results in the inactivation of GSH-Px, which, therefore, fails to convert H₂O₂ to water [38, 39]. Moreover, there is an increasing evidence that Cd interacts with Se and disrupts GSH-Px activity. This probably is the consequence of the intracellular accumulation of ROS with subsequent development of tissue injury.

3.4. Effect of Cd on Differentiation Marker Enzyme (AKP). Cadmium exposure had a significant ($P < 0.05$) effect on AKP activity in Caco-2 cells and HL-7702 cells (Table 1). AKP activity was highest in Caco-2 cells (97.6%) and HL-7702 cells (95.5%) when treated with 0.25 mg L⁻¹ Cd, while at 10 mg L⁻¹ its activity was decreased by 66.1% and 27.01%, respectively.

Our results showed that differentiation marker enzyme (AKP) was significantly decreased in both cell lines after 2 h Cd exposure. The inhibition of alkaline phosphatase activity has been already reported by others [40–42]. Also, El-Demerdash et al. [43] revealed that Cd *in vitro* caused

significant inhibition of AKP activity in human plasma. The decrease in AKP activity may be due to changes in the permeability of plasma membrane in addition to changes in the balance between synthesis and degradation of enzyme [40]. Also, Lakshmi et al. [44] reported that the inhibition of AKP may be due to the breakdown of the membrane transport system and an inhibitory effect on cell growth and proliferation.

4. Conclusions

This study concludes that Caco-2 and HL-7702 cell models are in good agreement with the *in vivo* experiment after oral ingestion of Cd: low rate being transferred from the lumen into the body, higher accumulation in the intestinal cells than the liver cells. These results demonstrated that Cd is able to induce oxidative damage and apoptosis in both cell lines when its concentration exceeds 0.5 mg L⁻¹. Finally, our findings suggest that HL-7702 cells are more sensitive to Cd exposure than Caco-2 cells at the same concentration and exposure duration.

Conflict of Interests

Authors declare that they have no conflict of interests.

Acknowledgments

This study was supported by the Ministry of Science and Technology of China (Grant no. 2012AA101405), Ministry of Environmental Protection of China (Grant no. 2011467057) and the Fundamental Research Funds for the Central Universities of China.

References

- [1] M. K. Daud, S. Ali, M. T. Variath et al., "Differential physiological, ultra-morphological and metabolic responses of cotton cultivars under cadmium stress," *Chemosphere*, vol. 93, no. 10, pp. 2593–2602, 2013.
- [2] G. C. Lalor, "Review of cadmium transfers from soil to humans and its health effects and Jamaican environment," *Science of the Total Environment*, vol. 400, no. 1–3, pp. 162–172, 2008.

- [3] M. K. Daud, Y. Sun, M. Dawood et al., "Cadmium-induced functional and ultrastructural alterations in roots of two transgenic cotton cultivars," *Journal of Hazardous Materials*, vol. 161, no. 1, pp. 463–473, 2009.
- [4] J. C. Merrill, J. J. P. Morton, and S. D. Soileau, "Metals: cadmium," in *Principles and Methods of Toxicology*, A. W. Hayes, Ed., pp. 665–667, Taylor and Francis, London, UK, 2001.
- [5] M. K. Daud, M. T. Variath, S. Ali et al., "Cadmium-induced ultramorphological and physiological changes in leaves of two transgenic cotton cultivars and their wild relative," *Journal of Hazardous Materials*, vol. 168, no. 2-3, pp. 614–625, 2009.
- [6] P. N. Williams, M. Lei, G. Sun et al., "Occurrence and partitioning of cadmium, arsenic and lead in mine impacted paddy rice: Hunan, China," *Environmental Science and Technology*, vol. 43, no. 3, pp. 637–642, 2009.
- [7] M. U. Khan, R. N. Malik, and S. Muhammad, "Human health risk from heavy metal via food crops consumption with wastewater irrigation practices in Pakistan," *Chemosphere*, vol. 93, no. 10, pp. 2230–2238, 2013.
- [8] O. Andersen, J. B. Nielsen, J. A. Sorensen, and L. Scherrebeck, "Experimental localization of intestinal uptake sites for metals (Cd, Hg, Zn, Se) *in vivo* in mice," *Environmental Health Perspectives*, vol. 102, no. 3, pp. 199–206, 1994.
- [9] R. K. Zalups and S. Ahmad, "Molecular handling of cadmium in transporting epithelia," *Toxicology and Applied Pharmacology*, vol. 186, no. 3, pp. 163–188, 2003.
- [10] B. Elsenhans, G. J. Strugala, and S. G. Schäfer, "Small-intestinal absorption of cadmium and the significance of mucosal metallothionein," *Human and Experimental Toxicology*, vol. 16, no. 8, pp. 429–434, 1997.
- [11] R. Arriazu, J. M. Pozuelo, N. Henriques-Gil et al., "Immunohistochemical study of cell proliferation, Bcl-2, p53, and caspase-3 expression on preneoplastic changes induced by cadmium and zinc chloride in the ventral rat prostate," *Journal of Histochemistry and Cytochemistry*, vol. 54, no. 9, pp. 981–990, 2006.
- [12] D. Huang, Y. Zhang, Y. Qi, C. Chen, and W. Ji, "Global DNA hypomethylation, rather than reactive oxygen species (ROS), a potential facilitator of cadmium-stimulated K562 cell proliferation," *Toxicology Letters*, vol. 179, no. 1, pp. 43–47, 2008.
- [13] P. Yang, H. Chen, J. Tsai, and L. Lin, "Cadmium induces Ca^{2+} -dependent necrotic cell death through calpain-triggered mitochondrial depolarization and reactive oxygen species-mediated inhibition of nuclear factor- κ B activity," *Chemical Research in Toxicology*, vol. 20, no. 3, pp. 406–415, 2007.
- [14] C. Shih, W. Ko, J. Wu et al., "Mediating of caspase-independent apoptosis by cadmium through the mitochondria-ROS pathway in MRC-5 fibroblasts," *Journal of Cellular Biochemistry*, vol. 91, no. 2, pp. 384–397, 2004.
- [15] G. Fotakis and J. A. Timbrell, "Role of trace elements in cadmium chloride uptake in hepatoma cell lines," *Toxicology Letters*, vol. 164, no. 2, pp. 97–103, 2006.
- [16] M. V. Berridge and A. S. Tan, "Characterization of the cellular reduction of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT): subcellular localization, substrate dependence, and involvement of mitochondrial electron transport in MTT reduction," *Archives of Biochemistry and Biophysics*, vol. 303, no. 2, pp. 474–482, 1993.
- [17] M. R. Ruocco, F. Cecere, A. Iuliano et al., "Diclofenac-induced apoptosis in the neuroblastoma cell line SH-SY5Y: possible involvement of the mitochondrial superoxide dismutase," *Journal of Biomedicine and Biotechnology*, vol. 2010, Article ID 801726, 11 pages, 2010.
- [18] S. S. Baker and R. D. Baker, "Antioxidant enzymes in the differentiated Caco-2 cell line," *In Vitro Cellular and Developmental Biology*, vol. 28, no. 9-10, pp. 643–647, 1992.
- [19] S. Chater, T. Douki, A. Favier, M. Sakly, and H. Abdelmelek, "Changes in antioxidant status and biochemical parameters after orally cadmium administration in females rats," *Acta Biologica Hungarica*, vol. 60, no. 1, pp. 79–88, 2009.
- [20] Y. Nzengue, R. Steiman, C. Garrel, E. Lefèbvre, and P. Guiraud, "Oxidative stress and DNA damage induced by cadmium in the human keratinocyte HaCaT cell line: role of glutathione in the resistance to cadmium," *Toxicology*, vol. 243, no. 1-2, pp. 193–206, 2008.
- [21] Y. Wang, J. Fang, S. S. Leonard, and K. M. K. Rao, "Cadmium inhibits the electron transfer chain and induces reactive oxygen species," *Free Radical Biology and Medicine*, vol. 36, no. 11, pp. 1434–1443, 2004.
- [22] V. Meunier, M. Bourrie, Y. Berger, and G. Fabre, "The human intestinal epithelial cell line Caco-2; pharmacological and pharmacokinetic applications," *Cell Biology and Toxicology*, vol. 11, no. 3-4, pp. 187–194, 1995.
- [23] Z. Xie, Y. Zhang, A. Li, P. Li, W. Ji, and D. Huang, "Cd-induced apoptosis was mediated by the release of Ca^{2+} from intracellular Ca storage," *Toxicology Letters*, vol. 192, no. 2, pp. 115–118, 2010.
- [24] C. Jumarie, P. G. C. Campbell, A. Berteloot, M. Houde, and F. Denizeau, "Caco-2 cell line used as an *in vitro* model to study cadmium accumulation in intestinal epithelial cells," *Journal of Membrane Biology*, vol. 158, no. 1, pp. 31–48, 1997.
- [25] M. M. Bradford, "A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein dye binding," *Analytical Biochemistry*, vol. 72, no. 1-2, pp. 248–254, 1976.
- [26] B. Zödl, M. Zeiner, M. Sargazi et al., "Toxic and biochemical effects of zinc in Caco-2 cells," *Journal of Inorganic Biochemistry*, vol. 97, no. 4, pp. 324–330, 2003.
- [27] D. M. Templeton, "Biomedical aspects of trace element speciation," *Fresenius' Journal of Analytical Chemistry*, vol. 363, no. 5-6, pp. 505–511, 1999.
- [28] Y. Ui-Jeong, Y. So-Ra, C. Jae-Hwan et al., "Water spinach (*Ipomoea aquatica* Forsk.) reduced the absorption of heavy metals in an *in vitro* bio-mimicking model system," *Food and Chemical Toxicology*, vol. 50, pp. 3862–3866, 2012.
- [29] D. A. Vesey, "Transport pathways for cadmium in the intestine and kidney proximal tubule: focus on the interaction with essential metals," *Toxicology Letters*, vol. 198, no. 1, pp. 13–19, 2010.
- [30] E. C. Foulkes, "Transport of toxic heavy metals across cell membranes," *Proceedings of the Society for Experimental Biology and Medicine*, vol. 223, no. 3, pp. 234–240, 2000.
- [31] N. J. DelRaso, B. D. Foy, J. M. Gearhart, and J. M. Frazier, "Cadmium uptake kinetics in rat hepatocytes: correction for albumin binding," *Toxicological Sciences*, vol. 72, no. 1, pp. 19–30, 2003.
- [32] E. A. Belyaeva, V. V. Glazunov, and S. M. Korotkov, "Cyclosporin A-sensitive permeability transition pore is involved in Cd^{2+} -induced dysfunction of isolated rat liver mitochondria: doubts no more," *Archives of Biochemistry and Biophysics*, vol. 405, no. 2, pp. 252–264, 2002.
- [33] J. Pourahmad and P. J. O'Brien, "A comparison of hepatocyte cytotoxic mechanisms for Cu^{2+} and Cd^{2+} ," *Toxicology*, vol. 143, no. 3, pp. 263–273, 2000.

- [34] S. Nemmiche, D. Chabane-Sari, M. Kadri, and P. Guiraud, "Cadmium chloride-induced oxidative stress and DNA damage in the human Jurkat T cell line is not linked to intracellular trace elements depletion," *Toxicology in Vitro*, vol. 25, no. 1, pp. 191–198, 2011.
- [35] D. Bagchi, M. Bagchi, S. J. Stohs et al., "Free radicals and grape seed proanthocyanidin extract: importance in human health and disease prevention," *Toxicology*, vol. 148, no. 2-3, pp. 187–197, 2000.
- [36] J. M. Matés, "Effects of antioxidant enzymes in the molecular control of reactive oxygen species toxicology," *Toxicology*, vol. 153, no. 1–3, pp. 83–104, 2000.
- [37] M. Waisberg, P. Joseph, B. Hale, and D. Beyersmann, "Molecular and cellular mechanisms of cadmium carcinogenesis," *Toxicology*, vol. 192, no. 2-3, pp. 95–117, 2003.
- [38] U. R. Acharya, M. Mishra, J. Patro, and M. K. Pan, "Effect of vitamins C and E on spermatogenesis in mice exposed to cadmium," *Reproductive Toxicology*, vol. 25, no. 1, pp. 84–88, 2008.
- [39] F. M. Maiorino, R. Brigelius-Flohé, K. D. Aumann, A. Roveri, D. Schomburg, and L. Flohé, "Diversity of glutathione peroxidases," *Methods in Enzymology*, vol. 252, pp. 38–53, 1995.
- [40] S. V. Rana, S. Rekha, and V. Seema, "Protective effects of few antioxidants on liver function in rats treated with cadmium and mercury," *Indian Journal of Experimental Biology*, vol. 34, no. 2, pp. 177–179, 1996.
- [41] F. M. El-Demerdash, M. I. Yousef, F. S. Kedwany, and H. H. Baghdadi, "Cadmium-induced changes in lipid peroxidation, blood hematology, biochemical parameters and semen quality of male rats: protective role of vitamin E and β -carotene," *Food and Chemical Toxicology*, vol. 42, no. 10, pp. 1563–1571, 2004.
- [42] F. M. El-Demerdash and E. I. Elagamy, "Biological effects in *Tilapia nilotica* fish as indicators of pollution by cadmium and mercury," *International Journal of Environmental Health Research*, vol. 9, no. 3, pp. 143–156, 1999.
- [43] F. M. El-Demerdash, M. I. Yousef, and E. I. Elagamy, "Influence of paraquat, glyphosate, and cadmium on the activity of some serum enzymes and protein electrophoretic behavior (in vitro)," *Journal of Environmental Science and Health B*, vol. 36, no. 1, pp. 29–42, 2001.
- [44] R. Lakshmi, R. Kundu, E. Thomas, and A. P. Mansuri, "Mercuric chloride induced inhibition of acid and alkaline phosphatase activity in the kidney of Mudskipper, *Boleophthalmus dentatus*," *Acta Hydrochimica et Hydrobiologica*, vol. 3, pp. 341–344, 1991.



Hindawi

Submit your manuscripts at
<http://www.hindawi.com>

