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# Bordonein-L, a new L-amino acid oxidase from *Crotalus durissus terrificus* snake venom: isolation, preliminary characterization and enzyme stability

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## Abstract

**Background:** *Crotalus durissus terrificus* venom (CdtV) is one of the most studied snake venoms in Brazil. Despite presenting several well known proteins, its L-amino acid oxidase (LAAO) has not been studied previously. This study aimed to isolate, characterize and evaluate the enzyme stability of bordonein-L, an LAAO from CdtV.

**Methods:** The enzyme was isolated through cation exchange, gel filtration and affinity chromatography, followed by a reversed-phase fast protein liquid chromatography to confirm its purity. Subsequently, its N-terminal amino acid sequence was determined by Edman degradation. The enzyme activity and stability were evaluated by a microplate colorimetric assay and the molecular mass was estimated by SDS-PAGE using periodic acid-Schiff staining and determined by mass spectrometry.

**Results:** The first 39 N-terminal amino acid residues exhibited high identity with other snake venom L-amino acid oxidases. Bordonein-L is a homodimer glycoprotein of approximately 101 kDa evaluated by gel filtration. Its monomer presents around 53 kDa estimated by SDS-PAGE and 58,702 Da determined by MALDI-TOF mass spectrometry. The enzyme exhibited maximum activity at pH 7.0 and lost about 50 % of its activity after five days of storage at 4 °C. Bordonein-L's activity was higher than the control when stored in 2.8 % mannitol or 8.5 % sucrose.

**Conclusions:** This research is pioneering in its isolation, characterization and enzyme stability evaluation of an LAAO from CdtV, denominated bordonein-L. These results are important because they increase the knowledge about stabilization of LAAOs, aiming to increase their shelf life. Since the maintenance of enzymatic activity after long periods of storage is essential to enable their biotechnological use as well as their functional studies.

**Keywords:** *Crotalus durissus terrificus*, L-amino acid oxidase, Rattlesnake, Enzyme activity, Enzyme stability, Chromatography, Snake venom, Yellow venom, Stabilization

## Background

L-amino acid oxidases (LAAOs) are enantioselective flavoenzymes that catalyze the stereospecific oxidative deamination of L-amino acids. An amino acid intermediate is hydrolyzed, releasing  $\alpha$ -keto acids and ammonia. Concomitantly, the reduced non-covalently bond cofactor – flavin mononucleotide (FMN) or flavin adenine

dinucleotide (FAD) – reoxidizes on molecular oxygen, producing hydrogen peroxide [1].

LAAOs are found in such diverse life forms as bacteria, marine organisms, fish, cyanobacteria, fungi, green algae, and snake venoms (SV) from the families Crotalidae, Elapidae and Viperidae [1–12].

SV-LAAOs are, in general, non-covalently bonded to FAD and their FAD-binding site shares sequential similarity with human monoamine oxidase, mouse interleukin 4-induced, bacterial and fungal LAAOs [1, 13]. SV-LAAOs usually constitute from 0.15 to 5 % of the snake venom protein, with some exceptions, such as the

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LAAO of *Bungarus caeruleus*, which represents 25 % of the total protein [14]. Several biological activities have been attributed to SV-LAAOs, including cytotoxicity, mild myonecrosis, apoptosis induction, induction and/or inhibition of platelet aggregation, as well as hemorrhagic, hemolytic, edematogenic, antibacterial, antiproliferative, antiparasitic and anti-HIV activities [14–25]. These activities are considered the result of the release of hydrogen peroxide, which produces oxidative stress [26]. However, the role of LAAOs in the venom has not been elucidated yet [26].

SV-LAAOs exhibit a wide range of isoelectric points (pI) from about 4.4 to 8.1, although it is unknown whether the different charges result in distinct pharmacological properties [13]. These enzymes prefer hydrophobic L-amino acids, because of substrate specificity related to side-chain binding sites [27].

LAAO activity is inhibited in the presence of ethylenediaminetetraacetic acid (EDTA), N-ethylmaleimide, phenylmethanesulfonyl fluoride (PMSF), glutathione and 1, 10-phenanthroline, since its cofactor is reduced under these conditions [14]. Furthermore, bivalent cations show different effects on LAAO activity. Manganese and calcium ions do not affect its specific activity. The LAAO from *C. adamanteus* requires  $Mg^{2+}$ , while those from *Lachesis muta* and *Bothrops brazili* are inhibited by  $Zn^{2+}$  [14].

The cytotoxic effect of BI-LAAO from *B. leucurus* venom was inhibited by about 25 % in the presence of catalase, an enzyme that cleaves hydrogen peroxide [17]. Additionally, the LAAOs of *Naja naja kaouthia* and *Calloselasma rhodostoma* venoms were inhibited by polyphenols from *Areca catechu* and *Quercus infectoria* extracts evaluated by *in vitro* tests [28]. Although the ethylacetate extract from *Azima tetracantha* leaves exerts an *in vitro* inhibitory activity on toxic enzymes from *B. caeruleus* and *Vipera russelli* venoms, LAAOs from neither venom was inhibited [29].

The LAAOs have shown maximum absorbance at 465 and 380 nm because of their bond with FAD [13]. Small changes in the absorption spectra of SV-LAAOs were observed after inactivation by freezing and thawing or modification of the ionic composition and pH conditions, indicating alterations in the microenvironment of the FAD cofactor [30]. Most of the studies in this area were published in the 1950s and 1960s [31–35]. One example is the inactivation of an LAAO isolated from *C. adamanteus* venom by high temperature and freezing. The higher the temperature or the pH of storage buffer, the higher the enzymatic inactivation, an inactivation that may be lower in the presence of chloride ions. On the other hand, at lower temperatures (freezing), the inactivation and storage buffer pH are inversely related. However, chloride ions were not able to prevent enzymatic inactivation in this case [31, 32]. Further studies showed that the inactivation of LAAOs causes changes in optical rotatory dispersion whereas the redox properties

of free flavin are similar to those of the inactive enzyme [33, 35]. The change in redox properties suggests the loss of most interactions between flavin and apoprotein. Raibekas and Massey [36] extracted the cofactor of the LAAO from *C. adamanteus* venom at pH 3.5, rebound it at pH 8.5 and restored the enzymatic activity in the presence of 50 % glycerol followed by dialysis at 4 °C against 0.1 M Tris–HCl buffer, pH 7.5, containing 0.1 M KCl [36].

Due to their participation in metabolic pathways involving nitrogen and their antimicrobial, antiviral and antitumor effects, SV-LAAOs are considered a promising biotechnological agent and a tool for investigating cellular processes [13, 14]. However, diverse conditional factors that can reduce the stability of biocatalysts – including temperature, pH, oxidative stress, the solvent, binding of metal ions or cofactors, and the presence of surfactants – limit the industrial use of enzymes [37, 38]. Working under operational conditions of enzyme stability, the process costs are reduced [37], since the enzyme is active when in use and keeps active over time [39].

Two reports have shown that the presence of univalent ions or substrates for LAAOs and analogues of the prosthetic group (competitive inhibitors) prevents the inactivation of some SV-LAAOs [32, 40]. However, no additional studies have addressed the use of additives to maintain LAAO activity, which is highly desirable for industrial applications.

The use of additives to maintain proteins in their active forms is widespread throughout the pharmaceutical industry. For example, cyclodextrins are employed as excipients in pharmaceutical formulations in order to avoid protein aggregations to keep the protein in its active form [41]. There is a huge diversity of additives that act as cryoprotectants. Sugars and polyols, such as sucrose and mannitol, respectively, are used as protein stabilizers since they are able to interact with protein through hydrogen bonds to replace the protein-water molecular interactions [42, 43]. Amino acids are also used as cryoprotectants [43]. Usually, adjuvants are employed at a percentage that ranges from 0.5 to 2 %, although higher concentrations have already been tested [44–46].

Therefore, this study isolated an LAAO from *C. durissus terrificus* venom (CdtV), denominated bordonein-L, and evaluated the effect of different additives (mannitol, sucrose, L-Lys and L-Gly) as cryoprotectants for the enzyme.

## Methods

### Isolation of bordonein-L

Cdt yellow venom from the Ribeirão Preto region (21° 10' 36" S, 47° 49' 15" W) was obtained from specimens kept in the central snake house (University of São Paulo, Ribeirão Preto, SP, Brazil), in accordance with the guidelines

of the Brazilian Institute of Environment and Renewable Natural Resources (IBAMA).

Desiccated CdtV (1 g) was purified through cation exchange chromatography, as described by Bordon et al. [47]. The CM5 fraction obtained in the first chromatographic step was fractionated on a HiPrep 16/60 Sephacryl S-100 HR column (1.6 × 60 cm, GE Healthcare, Sweden) equilibrated and eluted with 0.05 M sodium acetate buffer containing 0.15 M NaCl, pH 5.5, at a flow rate of 0.5 mL/min. The subfraction CM5S2 was applied on two 1-mL HiTrap Heparin HP columns (GE Healthcare) connected in a series equilibrated with 0.05 M sodium acetate buffer, pH 5.5. Adsorbed proteins were eluted using a step concentration gradient from 0 to 100 % of buffer B (1 M NaCl in the same buffer) at a 1.0 mL/min flow rate. To assess its purity degree, the peak H7 (LAAO bordonein-L) was submitted to RP-FPLC, as described by Bordon et al. [47].

#### Determination of proteins

Total proteins were determined by the 280/205 nm absorption method [48].

#### Determination of molecular mass

SDS-PAGE (10 %) was run according to the description of Laemmli [49]. The gel was stained with PlusOne Coomassie PhastGel Blue R-350 (GE Healthcare, Sweden) whereas periodic acid-Schiff (PAS) staining was employed to detect glycoproteins [50]. The hyaluronidase CdtHya1, a glycoprotein recently isolated from CdtV, was used as the control [47].

The molecular mass of bordonein-L was estimated by gel filtration chromatography on a Superdex 200 10/300GL column (GE Healthcare) calibrated with the following protein molecular mass standards: 12.4, 29, 66, 150 and 200 kDa (Sigma-Aldrich Co., United States). Blue dextran (2000 kDa, Sigma-Aldrich Co.) was used to determine the void volume. The column was equilibrated whereas the standards and the enzyme were eluted with the same buffer used on HiPrep 16/60 Sephacryl S-100 HR column. Each standard was filtered individually through the Superdex column and a calibration curve was constructed.

The molecular mass of bordonein-L was also analyzed by a MALDI-TOF mass spectrometer (Ultraflex II, Bruker Daltonics, Germany). MS spectrum was acquired in positive linear mode in the mass range 10,000-70,000 Da. TFA 0.1 % (10  $\mu$ L) was added to the lyophilized enzyme. This solution was mixed (1:1) with sinapinic acid (20 mg/mL in 50/50 0.2 % ACN/TFA, v/v); and 2  $\mu$ L of this mixture was spotted on a MALDI plate (384 positions) using the dried droplet method.

#### Bordonein-L sequencing and *in silico* analysis

The N-terminal of bordonein-L was determined by Edman degradation in an automated protein sequencer model PPSQ-33A (Shimadzu Co., Japan) and compared with sequences deposited in the Basic Local Alignment Search Tool (BLAST) [51]. The alignment was created by MultAlin Interface Page [52] and the figure was generated by ESPript [53] server.

#### LAAO activity

The LAAO activity of bordonein-L was performed through a microplate colorimetric assay according to modifications on the Kishimoto and Takahashi method [54]. Bordonein-L was incubated at 37 °C for 60 min with 0.002 M *o*-phenylenediamine (OPD) (Sigma-Aldrich Co.), 1 U/mL horseradish peroxidase (Sigma-Aldrich), 0.005 M L-Leucine (Sigma-Aldrich) and 0.05 M Tris-HCl buffer, pH 7.0. The reaction was stopped with 2 M H<sub>2</sub>SO<sub>4</sub> and the absorbance was measured at 492/630 nm. LAAO activity was also evaluated at different pH levels (5.0-9.0).

#### LAAO stability

The evaluation of LAAO stability was performed for 40 days at different concentration levels (1.4 %, 2.8 % and 8.5 %) of mannitol, sucrose, L-lysine and L-glycine, stored at 4 °C. Bordonein-L activity was also evaluated after being frozen (-20 °C) for a period of five days. The evaluation of enzymatic activity after lyophilization was performed as soon as this process was finished. The assays were carried out according to the LAAO activity assay previously described. Control consisted of bordonein-L in the absence of additives and storage at 4 °C. The enzyme was protected from light under all the tested conditions.

#### Statistical analysis

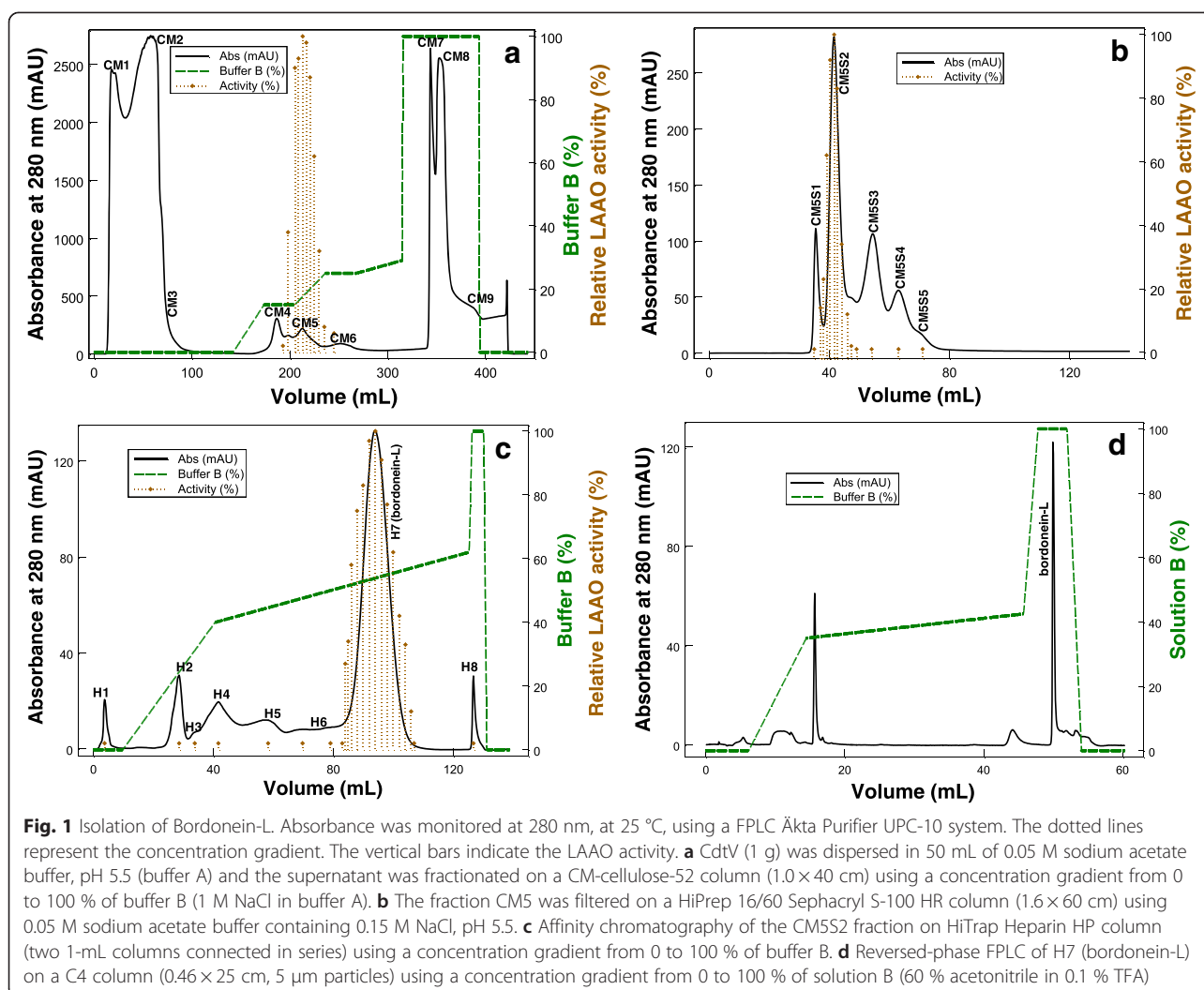
LAAO activity data were expressed as mean  $\pm$  standard error of mean (SEM). The analysis of variance (ANOVA) test was employed to evaluate data on LAAO activity in the presence of additives and to compare lyophilized, frozen and LAAO at 4 °C (five days), whereas the *t* test was utilized to compare LAAO stability after freezing versus already lyophilized. They were statistically significant when  $p < 0.05$ .

## Results

#### Isolation of bordonein-L

Bordonein-L was purified in three chromatographic steps: cation exchange, molecular exclusion and affinity chromatography.

LAAO activity was detected in the CM5 fraction (vertical bars, Fig. 1a) eluted from CM-cellulose-52 column. This fraction corresponds to 1.8 % of the total protein (Table 1). The CM5 fraction was applied on a HiPrep 16/60 Sephacryl



S-100 HR column and LAAO activity was detected in the CM5S2 fraction (Fig. 1b), which was submitted to affinity chromatography on a HiTrap Heparin HP column. Thus, pure LAAO (peak H7), denominated bordonein-L, was obtained (Fig. 1c). The pure enzyme represents 48.3 % of the total activity and 0.5 % of the total protein of the venom (Table 1). Bordonein-L was then applied on a C4 column

(Fig. 1d) and the main peak was submitted to Edman degradation.

#### Determination of molecular mass

SDS-PAGE under non-reducing conditions indicated that the peak H7 (bordonein-L) showed a high degree of purity while its monomer presented around 53 kDa

**Table 1** Specific activity and recovery of active fractions eluted during the purification procedure of bordonein-L

Fraction	Total protein (mg) <sup>a</sup>	Protein recovery (%)	One unit of LAAO activity (mg) <sup>b</sup>	Specific activity (U/mg) <sup>c</sup>	Total LAAO activity (U)	Yield (%)	Relative activity
CdtV	556.0	100.0	14.29	0.07	38.9	100.0	1.0
CM5	10.3	1.8	0.29	3.47	35.7	91.8	49.6
CM5S2	4.3	0.8	0.14	6.96	29.9	76.9	99.4
H7 (bordonein-L)	2.7	0.5	0.14	6.96	18.8	48.3	99.4

<sup>a</sup>Total protein quantified by absorbance method 280/205 nm (SCOPES, 1974)

<sup>b</sup>LAAO activity unit (U): amount of protein (mg) able to release 1.0 μmol of H<sub>2</sub>O<sub>2</sub> per minute

<sup>c</sup>Specific activity: amount of H<sub>2</sub>O<sub>2</sub> (μmol) released per minute per mg of protein

(Fig. 2a), versus 56 kDa under reducing conditions (data not shown). Periodic acid-Schiff (PAS) staining evidenced that bordonein-L is a glycoprotein (Fig. 2b). The molecular mass of 58,702 Da was determined by MALDI-TOF (linear positive mode) mass spectrometry (Fig. 2c). Gel filtration under non-reducing conditions revealed a protein of approximately 101 kDa (Fig. 2d), indicating that bordonein-L is a dimer protein.

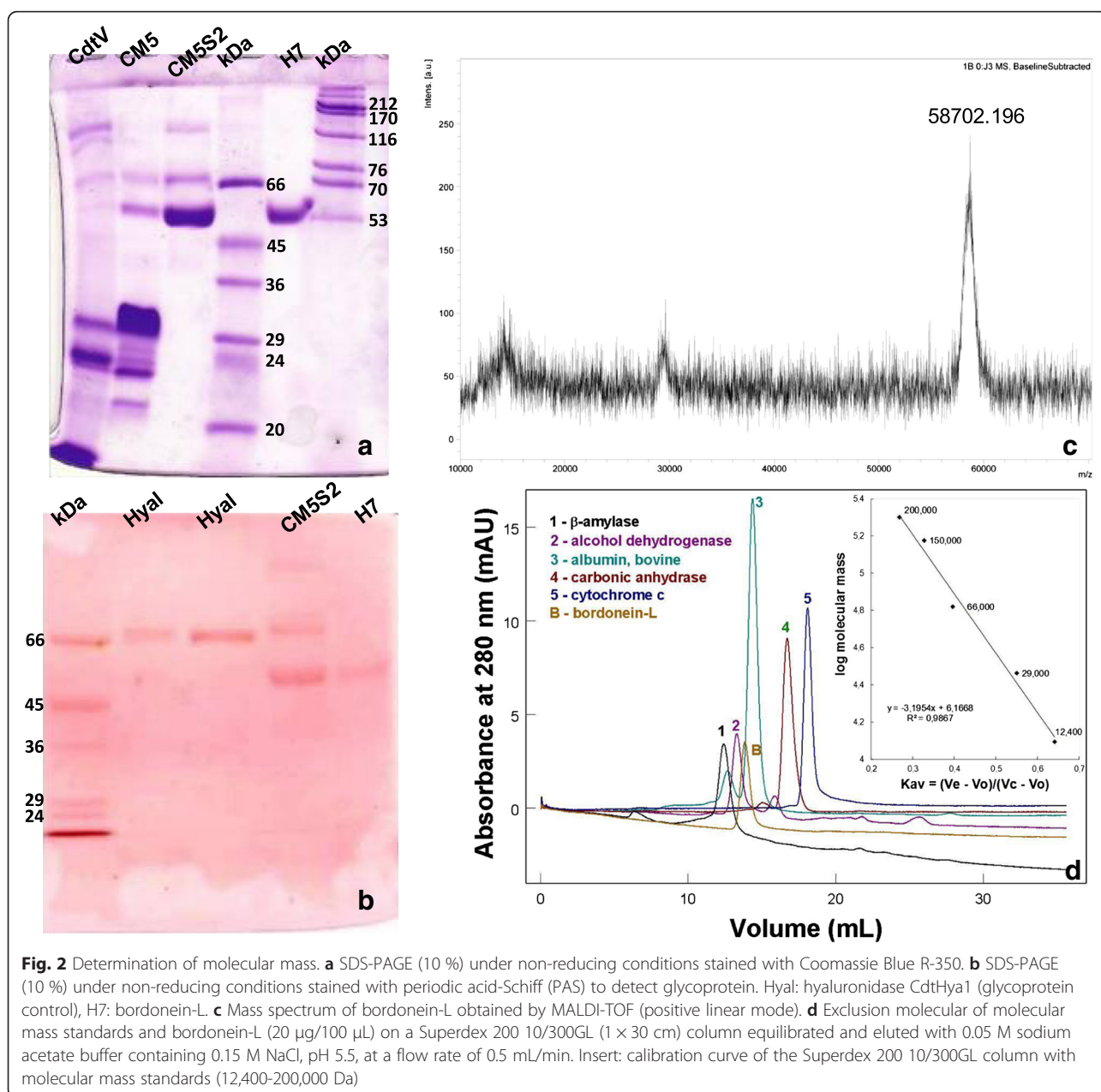
#### *In silico* assays

The sequence of the first 39 N-terminal amino acid residues from bordonein-L was determined by Edman degradation and appears in the UniProt Knowledgebase

under the accession number C0HJE7. This primary sequence exhibited high identity with other SV-LAAOs of the genus *Crotalus* (Fig. 3).

#### LAAO activity and stability

Bordonein-L showed an optimum pH of 7.0 (Fig. 4) and lost around 50 % of its activity in the first five days of storage at 4 °C (Fig. 5a-e). The frozen bordonein-L did not show enzymatic activity after lyophilization (Fig. 5a). Low activity (5 %) was also seen after thawing (Fig. 5a). Furthermore, LAAO activity was statistically significant when freezing and lyophilization were compared (Fig. 5a). L-lysine and L-glycine were not able to avoid



	1	10	20	30	40	50	60
F8S0Z5_adamanteus	MNVFFMFSLFLAALGS	CAH	DRNPLEE	CFRET	DYEEFLEIA	KNGLT	ATSNPKRVVIVGAG
P56742_atrox	MNVFFMFSLFLAALGS	CAH	DRNPLEE	CFRET	DYEEFLEIA	KNGLT	ATSNPKRVVIVGAG
O93364_adamanteus	MNVFFMFSLFLAALGS	CAH	DRNPLEE	CFRET	DYEEFLEIA	KNGLT	ATSNPKRVVIVGAG
T1DJZ4_horridus	MNVFFMFSLFLAALGS	CAH	DRNPLEE	CFRET	DYEEFLEIA	KNGLT	ATSNPKRVVIVGAG
K9N7B7_cumanensis	.....	S	CADD	DRNPLEE	CFRET	DYEEFLEIA	RNGLT
P0C2D2_cascavella	.....	A	DRNPLEE	Q	CFRET	DYEEFLEIA	RNGLT
C0HJE7_bordonein_L	.....	A	DRNPLEE	CFRET	DYEEFLEIA	RNGLT	VTSNPKRVVIVGAG

**Fig. 3** Multiple sequential alignment of snake venom L-amino acid oxidases from the genus *Crotalus*. Initial N-terminal of bordonein-L [Swiss-Prot: C0HJE7, bottom] and LAAOs from crotalic venoms: *C. adamanteus* [Swiss-Prot: F8S0Z5, O93364], *C. atrox* [Swiss-Prot: P56742], *C. horridus* [Swiss-Prot: T1DJZ4], *C. d. cumanensis* [Swiss-Prot: K9N7B7 – fragment] and *C. d. cascavella* [Swiss-Prot: P0C2D2 – fragment]. The highly conserved residues in bordonein-L are highlighted in black. The amino acid residues in red indicate low consensus. Cys residues are shaded in blue. The alignment and figure were generated by the servers MultAlin [52] and ESPript [53], respectively

the loss of activity at the tested concentrations (Fig. 5d and e). Bordonein-L activity was decreased when stored in 2.8 % mannitol, but during the course of the determined period of time (20 days), it was higher than the control. The enzymatic activity was the same as the control in the presence of other mannitol concentrations (1.4 % and 8.5 %) (Fig. 5b). On the other hand, 8.5 % sucrose kept bordonein-L more active than control during the first 20 days. Other tested sucrose concentrations were unable to keep bordonein-L more active than the control in the same time period (Fig. 5c).

## Discussion

There are 78 and 51 known primary sequences of SV-LAAOs deposited in the NCBI and UniProt databanks, respectively. However, the LAAO from *Crotalus durissus terrificus* venom (CdtV), one of the most studied snake venoms in Brazil, had not been assessed previously.

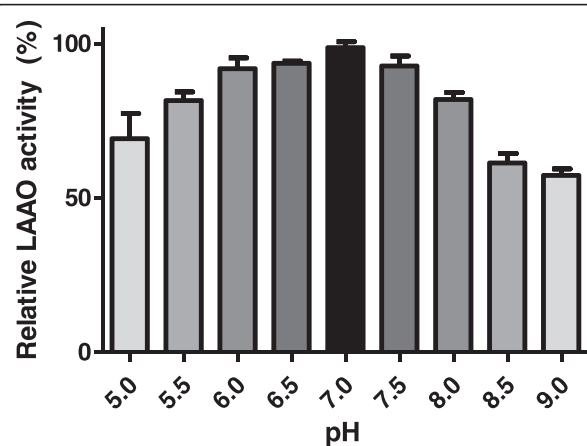
This is the first report of an LAAO from CdtV, denominated bordonein-L. The enzyme was isolated in three chromatographic steps and represented 0.5 % of the soluble venom protein. The specific activity for the

soluble venom was 0.07 against 6.96 for bordonein-L, representing a 99.4-fold purification. The fractionation of 1 g of CdtV yielded only 2.7 mg of bordonein-L, a yield approximately four fold lower than the one obtained from the purification of 1 g of *C. adamanteus* venom [36]. However, its recovery is within the range from 0.15 to 5 % of the total protein observed in other snake venoms [14]. Significant differences in activity and protein concentration are observed even in snake venoms from the same species and region, as recently reported for the Cdt venom from the Botucatu region (SP, Brazil) [55].

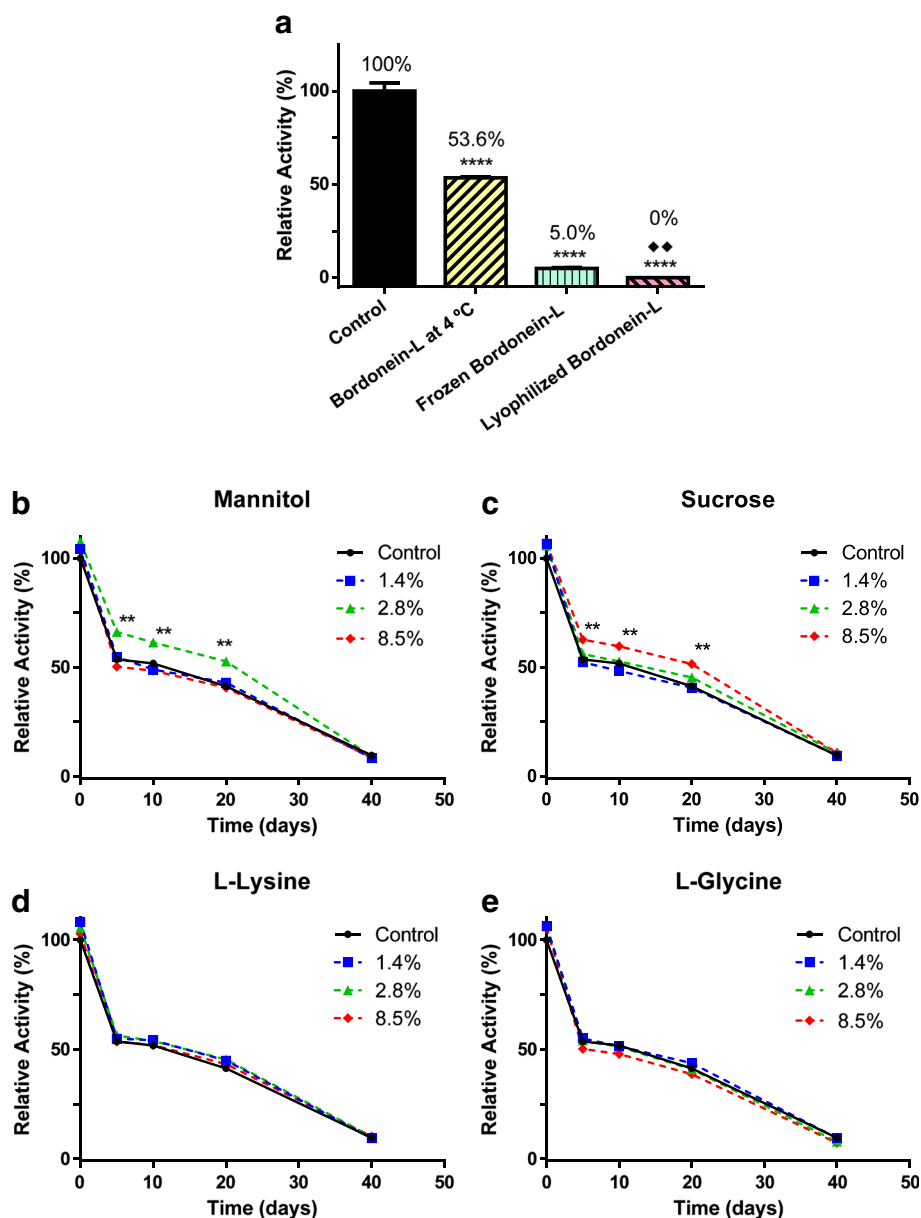
Bordonein-L is a homodimeric glycoprotein. Molecular sieve chromatography under non-reducing conditions revealed a protein of approximately 101 kDa, while its mass was estimated at about 53 kDa by SDS-PAGE and 58,702 Da by mass spectrometry. SV-LAAOs are usually homodimeric FAD-binding glycoproteins with a molecular mass of around 110–150 kDa when measured by gel filtration under non-denaturing conditions and around 50–70 kDa when assayed by SDS-PAGE under reducing and non-reducing conditions [13]. Our results indicate that bordonein-L is a non-covalently associated homodimer, as reported for most SV-LAAOs.

The sequence of the first 39 N-terminal amino acid residues of bordonein-L exhibited identity with other SV-LAAOs, since the amino-terminal region is highly conserved. A high degree of similarity (>84 %) has been described among the primary sequences of SV-LAAOs even when comparing distinct genera [14].

Bordonein-L exhibited more than 80 % of relative activity in the pH range from 5.5 to 8.0, showing maximum activity at pH 7.0. Other SV-LAAOs show an active conformation at a pH ranging from 5.5 to 7.5, being inactivated at extremely basic pHs [34]. We observed an approximately 50 % loss of LAAO activity in the first five days of storage at 4 °C, almost complete inactivation after freezing and thawing, and total inactivation after lyophilization. The activity of the LAAO isolated from *C. adamanteus*, which shares high sequence identity with bordonein-L, is also greatly decreased by freezing



**Fig. 4** pH profile of LAAO activity. *Crotalus durissus terrificus* crude soluble venom, horseradish peroxidase, OPD and L-leucine were incubated in different 0.05 M buffers, at different pHs (5.0 to 9.0), for 60 min at 37 °C



**Fig. 5** Bordonein-L stability. **a** Evaluation of stability after five days at  $-20^{\circ}\text{C}$  and  $4^{\circ}\text{C}$  and as soon as the lyophilization was finished. The stability was also evaluated for 40 days in the presence of **(b)** mannitol, **(c)** sucrose, **(d)** L-lysine and **(e)** L-glycine. All samples were kept protected from light. Each point represents the mean  $\pm$  S.E.M. ( $n = 3$ ) at each additive concentration (\*\* $p < 0.0001$  compared to the respective control using one-way ANOVA test). Each bar represents the mean  $\pm$  S.E.M. ( $n = 3$ ) at  $4^{\circ}\text{C}$ , freezing and lyophilization conditions (\*\*\*\* $p < 0.0001$  when  $4^{\circ}\text{C}$ , freezing and lyophilization was compared to control and when freezing and lyophilization compared to  $4^{\circ}\text{C}$  using one-way ANOVA test;  $\blacklozenge p < 0.05$  when freezing and lyophilization were compared to each other using the  $t$  test)

[31, 32]. Other SV-LAAOs presented similar results [13]. Therefore, we suggest that bordonein-L be kept at  $4^{\circ}\text{C}$  and near neutral pH to avoid its inactivation.

In relation to the stability of bordonein-L, L-glycine and L-lysine did not prevent the loss of enzymatic activity during the 40 days of storage at  $4^{\circ}\text{C}$ , probably because they are not able to effectively interact with the active site in contrast to the hydrophobic L-amino acids

and competitive inhibitors. L-glycine is the smallest amino acid and this small size may hinder its interaction with the catalytic site of bordonein-L. On the other hand, the amino acid L-lysine presents high polarity and the presence of polar groups might disrupt hydrophobic interactions. Hydrophobic L-amino acids, e.g. L-leucine, were not tested in this study as cryoprotectants because they are usually the preferred substrates of LAAOs

whereas changes in the amino acid concentration would occur due to their concomitant oxidation during the activity assay, which would prevent the correct quantification of the LAAO activity [32].

Bordonein-L's activity was higher than the control during the first 20 days when stored in 2.8 % mannitol or 8.5 % sucrose. At those concentrations, mannitol and sucrose interacted with bordonein-L through hydrogen bonds, which probably stabilized the enzyme by replacing the water molecular interactions, as reported for other proteins [42, 43]. However, after 40 days of storage, bordonein-L lost almost all of its activity even in the presence of additives. The rapid loss of activity (around 50 %) in the first five days and activity loss even in the presence of additives lead us to speculate that an alteration in the cofactor, such as oxidation or reduction, and/or changes in the catalytic site are responsible for the loss of LAAO activity since they may hinder the interaction among flavin, protein and substrate. The reduction of enzymatic activity as a result of FAD loss or conformational alterations was reported in other LAAOs [30, 33, 35]. Some conformational changes at the catalytic site were also suggested for gyroxin, another enzyme isolated from *CdtV*, whose catalytic efficiency was decreased in the presence of  $Mn^{2+}$  and  $Cu^{2+}$  [56].

The incorporation of additives to improve the stabilization of enzymes is the oldest and one of the most reliable enzyme stabilization methods, being employed in the most marketed enzyme formulations [57]. Since LAAOs are considered a promising biotechnological agent and a tool to investigate cellular processes, the retention of its enzymatic activity over time is essential [13, 14].

## Conclusions

An LAAO, denominated bordonein-L, was isolated from *CdtV* and presented higher enzymatic activity than the control when stored in 2.8 % mannitol or 8.5 % sucrose. These results may help the search for new additives to be used in stabilizing the LAAO, with the objective of increasing the shelf life of the enzyme.

## Abbreviations

ANOVA: Analysis of variance; BLAST: Basic local alignment search tool; *CdtV*: *Crotalus durissus terrificus* venom; EDTA: Ethylenediaminetetraacetic acid; FAD: Flavin adenine dinucleotide; FMN: Flavin mononucleotide; LAAO: L-amino acid oxidase; MALDI-TOF: Matrix assisted laser desorption ionization time of flight; OPD: O-phenylenediamine; PAS: Periodic acid-Schiff; pI: Isoelectric point; PMSF: Phenylmethanesulfonyl fluoride; RP-FPLC: Reversed-phase fast protein liquid chromatography; SDS-PAGE: Sodium dodecyl sulphate polyacrylamide gel electrophoresis; SEM: Standard error of mean; SV: Snake venom.

## Competing interests

The authors declare that there are no competing interests.

## Authors' contributions

KCFB worked on the isolation and characterization of the enzyme and drafted the manuscript. GAW carried out the enzyme stability assays and

helped in drafting the manuscript. HC participated in the design of the study and analysis of the results. ECA is the corresponding author and designer of the research. All authors read and approved the final manuscript.

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## References

- Costa TR, Burin SM, Menaldo DL, de Castro FA, Sampaio SV. Snake venom L-amino acid oxidases: an overview on their antitumor effects. *J Venom Anim Toxins incl Trop Dis*. 2014;20:23.
- Stumpf PK, Green DE. L-amino acid oxidase of *Proteus vulgaris*. *J Biol Chem*. 1944;153:387–99.
- Geueke B, Hummel W. A new bacterial L-amino acid oxidase with a broad substrate specificity: purification and characterization. *Enzyme Microb Technol*. 2002;31(1–2):77–87.
- Coudert M, Vandecasteele JP. Characterization and physiological function of a soluble L-amino-acid oxidase in *Corynebacterium*. *Arch Microbiol*. 1975;102(2):151–3.
- Tong H, Chen W, Shi W, Qi F, Dong X. SO-LAAO, a novel L-amino acid oxidase that enables *Streptococcus oligofermentans* to outcompete *Streptococcus mutans* by generating  $H_2O_2$  from peptone. *J Bacteriol*. 2008;190(13):4716–21.
- Yang HC, Johnson PM, Ko KC, Kamio M, Germann MW, Derby CD, et al. Cloning, characterization and expression of escapin, a broadly antimicrobial FAD-containing L-amino acid oxidase from ink of the sea hare *Aplysia californica*. *J Exp Biol*. 2005;208(Pt 18):3609–22.
- Palenik B, Morel FMM. Comparison of cell-surface L-amino-acid oxidases from several marine-phytoplankton. *Mar Ecol Prog Ser*. 1990;59:195–201.
- Kitani Y, Kikuchi N, Zhang G, Ishizaki S, Shimakura K, Shiomi K, et al. Antibacterial action of L-amino acid oxidase from the skin mucus of rockfish *Sebastes schlegelii*. *Comp Biochem Physiol B Biochem Mol Biol*. 2008;149(2):394–400.
- Kasai K, Ishikawa T, Komata T, Fukuchi K, Chiba M, Nozaka H, et al. Novel L-amino acid oxidase with antibacterial activity against methicillin-resistant *Staphylococcus aureus* isolated from epidermal mucus of the flounder *Platichthys stellatus*. *FEBS J*. 2010;277:453–65.
- Bockholt R, Masepohl B, Kruff V, Wittmann-Liebold B, Pistorius EK. Partial amino acid sequence of an L-amino acid oxidase from the cyanobacterium *Synechococcus PCC6301*, cloning and DNA sequence analysis of the *aoxA* gene. *Biochim Biophys Acta*. 1995;1264(3):289–93.
- Nuutinen JT, Marttinen E, Soliymani R, Hilden K, Timonen S. L-Amino acid oxidase of the fungus *Hebeloma cylindrosporum* displays substrate preference towards glutamate. *Microbiology*. 2012;158(Pt 1):272–83.
- Piedras P, Pineda M, Muñoz J, Cárdenas J. Purification and characterization of an L-amino-acid oxidase from *Chlamydomonas reinhardtii*. *Planta*. 1992;188(1):13–8.
- Du XY, Clemetson KJ. Snake venom L-amino acid oxidases. *Toxicon*. 2002;40(6):659–65.



14. Izidoro LFM, Sobrinho JC, Mendes MM, Costa TR, Grabner AN, Rodrigues VM, et al. Snake venom L-amino acid oxidases: trends in pharmacology and biochemistry. *Biomed Res Int*. 2014; doi.org/10.1155/2014/196754.
15. Bregge-Silva C, Nonato MC, de Albuquerque S, Ho PL, de Azevedo IL J, Vasconcelos Diniz MR, et al. Isolation and biochemical, functional and structural characterization of a novel L-amino acid oxidase from *Lachesis muta* snake venom. *Toxicon*. 2012;60(7):1263–76.
16. Souza DH, Eugenio LM, Fletcher JE, Jiang MS, Garratt RC, Oliva G, et al. Isolation and structural characterization of a cytotoxic L-amino acid oxidase from *Agkistrodon contortrix latincinctus* snake venom: preliminary crystallographic data. *Arch Biochem Biophys*. 1999;368(2):285–90.
17. Naumann GB, Silva LF, Silva L, Faria G, Richardson M, Evangelista K, et al. Cytotoxicity and inhibition of platelet aggregation caused by an L-amino acid oxidase from *Bothrops leucurus* venom. *Biochim Biophys Acta*. 2011;1810(7):683–94.
18. Alves RM, Antonucci GA, Paiva HH, Cintra AC, Franco JJ, Mendonça-Franqueiro EP, et al. Evidence of caspase-mediated apoptosis induced by L-amino acid oxidase isolated from *Bothrops atrox* snake venom. *Comp Biochem Physiol A Mol Integr Physiol*. 2008;151(4):542–50.
19. Li R, Zhu S, Wu J, Wang W, Lu Q, Clemetson KJ. L-amino acid oxidase from *Naja atra* venom activates and binds to human platelets. *Acta Biochim Biophys Sin (Shanghai)*. 2008;40(1):19–26.
20. Ali SA, Stoeva S, Abbasi A, Alam JM, Kayed R, Faigle M, et al. Isolation, structural, and functional characterization of an apoptosis-inducing L-amino acid oxidase from leaf-nosed viper (*Eristocophis macmahoni*) snake venom. *Arch Biochem Biophys*. 2000;384(2):216–26.
21. Wei JF, Yang HW, Wei XL, Qiao LY, Wang WY, He SH. Purification, characterization and biological activities of the L-amino acid oxidase from *Bungarus fasciatus* snake venom. *Toxicon*. 2009;54(3):262–71.
22. Vargas Muñoz LJ, Estrada-Gomez S, Nuñez V, Sanz L, Calvete JJ. Characterization and cDNA sequence of *Bothriechis schlegelii* L-amino acid oxidase with antibacterial activity. *Int J Biol Macromol*. 2014;69:200–7.
23. Lee ML, Chung I, Fung SY, Kanthimathi MS, Hong TN. Antiproliferative activity of king cobra (*Ophiophagus hannah*) venom L-amino acid oxidase. *Basic Clin Pharmacol Toxicol*. 2014;114(4):336–43.
24. Yun Z, Yongtang Z, Yujie Z. Application of snake venom L-amino acid oxidase in preparing AIDS treating medicine. *PAT - CN1526445*.
25. Zhang YJ, Wang JH, Lee WH, Wang Q, Liu H, Zheng YT, et al. Molecular characterization of *Trimeresurus stejnegeri* venom L-amino acid oxidase with potential anti-HIV activity. *Biochem Biophys Res Commun*. 2003;309(3):598–604.
26. Fox JW. A brief review of the scientific history of several lesser-known snake venom proteins: L-amino acid oxidases, hyaluronidases and phosphodiesterases. *Toxicon*. 2013;62:75–82.
27. Ponnudurai G, Chung MC, Tan NH. Purification and properties of the L-amino acid oxidase from Malayan pit viper (*Calloselasma rhodostoma*) venom. *Arch Biochem Biophys*. 1994;313(2):373–8.
28. Leanpolchareanchai J, Pithayanukul P, Bavovada R. Anti-necrosis potential of polyphenols against snake venoms. *Immunopharmacol Immunotoxicol*. 2009;31(4):556–62.
29. Janardhan B, Shrikanth VM, Mirajkar KK, More SS. *In vitro* screening and evaluation of antivenom phytochemicals from *Azima tetracantha* Lam. leaves against *Bungarus caeruleus* and *Vipera russelli*. *J Venom Anim Toxins incl Trop Dis*. 2014;20:12.
30. Massey V, Curti B. On the reaction mechanism of *Crotalus adamanteus* L-amino acid oxidase. *J Biol Chem*. 1967;242(6):1259–64.
31. Curti B, Massey V, Zmudka M. Inactivation of snake venom L-amino acid oxidase by freezing. *J Biol Chem*. 1968;243:2306–14.
32. Kearney EB, Singer TP. The L-amino acid oxidases of snake venom. III. Reversible inactivation of L-amino acid oxidases. *Arch Biochem Biophys*. 1951;33(3):377–96.
33. Wellner D. Evidence for conformational changes in L-amino acid oxidase associated with reversible inactivation. *Biochemistry*. 1966;5(5):1585–91.
34. Coles CJ, Edmondson DE, Singer TP. Reversible inactivation of L-amino acid oxidase. Properties of the three conformational forms. *J Biol Chem*. 1977;252(22):8035–9.
35. Soltysik S, Byron CM, Einarsdottir GH, Stankovich MT. The effects of reversible freezing inactivation and inhibitor binding on redox properties of L-amino-acid oxidase. *Biochim Biophys Acta*. 1987;911(2):201–8.
36. Raibekas AA, Massey V. Glycerol-induced development of catalytically active conformation of *Crotalus adamanteus* L-amino acid oxidase *in vitro*. *Proc Natl Acad Sci U S A*. 1996;93(15):7546–51.
37. van den Burg B, Eijnsink VG. Selection of mutations for increased protein stability. *Curr Opin Biotechnol*. 2002;13(4):333–7.
38. Eijnsink VGH, Gaseidnes S, Borchert TV, van den Burg B. Directed evolution of enzyme stability. *Biomol Eng*. 2005;22(1–3):21–30.
39. Drago GA, Gibson TD. Enzyme stability and stabilisation: applications and case studies. In: Hofman M, Thonart P, editors. *Engineering and Manufacturing for Biotechnology*. Netherlands: Springer; 2002. p. 361–76. Series: Focus on Biotechnology.
40. Kearney EB, Singer TP. The inactivation of L-amino acid oxidase by inorganic phosphate and arsenate. *Arch Biochem*. 1949;21(1):242–5.
41. Serno T, Geidobler R, Winter G. Protein stabilization by cyclodextrins in the liquid and dried state. *Adv Drug Deliv Rev*. 2011;63(13):1086–106.
42. Allison SD, Chang B, Randolph TW, Carpenter JF. Hydrogen bonding between sugar and protein is responsible for inhibition of dehydration-induced protein unfolding. *Arch Biochem Biophys*. 1999;365(2):289–98.
43. Arakawa T, Prestrelski SJ, Kenney WC, Carpenter JF. Factors affecting short-term and long-term stabilities of proteins. *Adv Drug Deliv Rev*. 2001;46(1–3):307–26.
44. Namaldi A, Calik P, Uludag Y. Effects of spray drying temperature and additives on the stability of serine alkaline protease powders. *Dry Technol*. 2006;24(11):1495–500.
45. Xie GF, Timasheff SN. Mechanism of the stabilization of ribonuclease A by sorbitol: preferential hydration is greater for the denatured than for the native protein. *Protein Sci*. 1997;6(1):211–21.
46. Liu W, Wang DQ, Nail SL. Freeze-drying of proteins from a sucrose-glycine excipient system: effect of formulation composition on the initial recovery of protein activity. *AAPS Pharm Sci Tech*. 2005;6(2):E150–7.
47. Bordon KC, Perino MG, Giglio JR, Arantes EC. Isolation, enzymatic characterization and anti-edematogenic activity of the first reported rattlesnake hyaluronidase from *Crotalus durissus terrificus* venom. *Biochimie*. 2012;94(12):2740–8.
48. Scopes RK. Measurement of protein by spectrophotometry at 205 nm. *Anal Biochem*. 1974;59(1):277–82.
49. Laemmli UK. Cleavage of structural proteins during the assembly of the head of bacteriophage-T4. *Nature*. 1970;227:680–5.
50. Doerner KC, White BA. Detection of glycoproteins separated by nondenaturing polyacrylamide gel electrophoresis using the periodic acid-schiff stain. *Anal Biochem*. 1990;187(1):147–50.
51. Basic Local Alignment Search Tool. <http://blast.ncbi.nlm.nih.gov/Blast.cgi>.
52. Corpet F. Multiple sequence alignment with hierarchical-clustering. *Nucleic Acids Res*. 1988;16(22):10881–90.
53. Gouet P, Courcelle E, Stuart DI, Metz F. ESPript: analysis of multiple sequence alignments in PostScript. *Bioinformatics*. 1999;15(4):305–8.
54. Kishimoto M, Takahashi T. A spectrophotometric microplate assay for L-amino acid oxidase. *Anal Biochem*. 2001;298(1):136–9.
55. Lourenço Jr A, Zorzella Creste CF, de Barros LC, dos Santos LD, Pimenta DC, Barraviera B, et al. Individual venom profiling of *Crotalus durissus terrificus* specimens from a geographically limited region: crotonamine assessment and captivity evaluation on the biological activities. *Toxicon*. 2013;69:75–81.
56. Barros LC, Soares AM, Costa FL, Rodrigues VM, Fuly AL, Giglio JR, et al. Biochemical and biological evaluation of gyroxin isolated from *Crotalus durissus terrificus* venom. *J Venom Anim Toxins incl Trop Dis*. 2011;17(1):23–33 [[http://www.scielo.br/scielo.php?pid=S1678-91992011000100004&script=sci\\_arttext](http://www.scielo.br/scielo.php?pid=S1678-91992011000100004&script=sci_arttext)]
57. Iyer PV, Ananthanarayan L. Enzyme stability and stabilization - Aqueous and non-aqueous environment. *Process Biochem*. 2008;43(10):1019–32.