Neurotherapeutics (2017) 14:1120–1133 DOI 10.1007/s13311-017-0546-7

# CrossMark

**ORIGINAL ARTICLE** 

# Chronic Paroxetine Treatment Prevents the Emergence of Abnormal Electroencephalogram Oscillations in Huntington's Disease Mice

Sandor Kantor<sup>1</sup> · Janos Varga<sup>1</sup> · Shreya Kulkarni<sup>1</sup> · A. Jennifer Morton<sup>1</sup>

Published online: 26 June 2017

© The Author(s) 2017. This article is an open access publication

**Abstract** Disturbance of rapid eye movement (REM) sleep appears early in both patients with Huntington's disease (HD) and mouse models of HD. Selective serotonin reuptake inhibitors are widely prescribed for patients with HD, and are also known to suppress REM sleep in healthy subjects. To test whether selective serotonin reuptake inhibitors can correct abnormal REM sleep and sleep-dependent brain oscillations in HD mice, we treated wild-type and symptomatic R6/2 mice acutely with vehicle and paroxetine (5, 10, and 20 mg/kg). In addition, we treated a group of R6/2 mice chronically with vehicle or paroxetine (20 mg/kg/day) for 8 weeks, with treatment starting before the onset of overt motor symptoms. During and after treatment, we recorded electroencephalogram/electromyogram from the mice. We found that both acute and chronic paroxetine treatment normalized REM sleep in R6/2 mice. However, only chronic paroxetine treatment prevented the emergence of abnormal low-gamma (25-45 Hz) electroencephalogram oscillations in R6/2 mice, an effect that persisted for at least 2 weeks after treatment stopped. Chronic paroxetine treatment also normalized REM sleep theta rhythm in R6/2 mice, but, interestingly, this effect was restricted to the treatment period. By contrast, acute paroxetine treatment slowed REM sleep theta rhythm in WT mice but had no effect on abnormal theta or low-gamma oscillations in R6/2

This article has been updated. The spelling of Electroencephalogram has been made consistent and the spacing in the legend to Fig. 6 was fixed.

**Electronic supplementary material** The online version of this article (doi:10.1007/s13311-017-0546-7) contains supplementary material, which is available to authorized users.

Department of Physiology, Development and Neuroscience, University of Cambridge, Cambridge, UK



mice. Our data show that paroxetine treatment, when initiated before the onset of symptoms, corrects both REM sleep disturbances and abnormal brain oscillations, suggesting a possible mechanistic link between early disruption of REM sleep and the subsequent abnormal brain activity in HD mice.

**Keywords** SSRI · Quantitative EEG · Gamma power · Theta oscillation · Biomarker

#### Introduction

Huntington's disease (HD) is an incurable neurodegenerative disorder characterized by motor, cognitive, and psychiatric disturbances, including depression [1]. Disrupted sleep and abnormal brain oscillations typically appear before the diagnostic motor signs, and may contribute to the cognitive impairment and depression seen in patients with HD [2-5]. One of the earliest and most consistent sleep findings in HD is the disturbance of rapid eye movement (REM) sleep. Patients with HD have longer REM sleep latencies, decreased REM sleep amount, and reduced electroencephalogram (EEG) theta oscillations during REM sleep [2, 4–8]. Similar to HD patients, mouse models of HD also have REM sleep disturbances early in the disease process [9-15]. In particular, we have shown previously that R6/2 mice exhibit a progressive increase in REM sleep amount during their active dark period, as well as a slowing of REM sleep EEG theta rhythm [9, 10]. R6/2 mice also develop abnormal low-gamma (25–45 Hz) oscillations in their sleep EEG, including REM sleep [9, 10], reflecting those changes seen in patients with early HD [2]. Similar sleep and EEG abnormalities have been found by other groups in both R6/2 [11, 16, 17] and R6/1 transgenic mice [12, 14, 15], as well as in the Q175 knock-in mouse model of HD [13, 18, 19]. REM sleep plays an important role in stabilizing and refining neuronal circuits in the developing brain [20–22]. Thus, an early disruption of REM sleep may lead to abnormal brain activity that is reflected in the disrupted sleep architecture and abnormal brain oscillations, which are seen not only in patients with HD, but also in mouse models of the disease.

In patients with HD, symptoms of depression are commonly treated with antidepressants, such as the tricyclic antidepressant amitriptyline or the selective serotonin reuptake inhibitor (SSRI) paroxetine [23, 24]. Most antidepressants (including amitriptyline and paroxetine) are known to alter sleep architecture, primarily by suppressing REM sleep [25]. Previously, we have shown that a single dose of amitriptyline normalizes REM sleep and supresses the abnormal EEG oscillations in R6/2 mice [10]. To test whether an antidepressant from a different class, but with similar REM-suppressing effect in healthy subjects, can normalize sleep-dependent brain oscillations in HD mice, we treated wild-type (WT) and R6/2 mice acutely with paroxetine and then monitored the changes in their sleep and EEG. Since changes in sleep and brain oscillatory activity precede most other symptoms in HD, we also treated a group of R6/2 mice chronically with paroxetine from an age that precedes the onset of overt motor symptoms by several weeks (presymptomatic stage). We wanted to see whether the suppression of REM sleep by paroxetine at an early presymptomatic stage of the disease can prevent the development of EEG abnormalities in HD mice. We found that chronic, but not acute, treatment with paroxetine not only normalized REM sleep, but also prevented the development of abnormal brain oscillations in R6/2 mice. Some of these changes persisted for at least 2 weeks after treatment stopped, suggesting that paroxetine treatment had an ameliorating effect on system changes underlying EEG abnormalities seen in HD mice.

## Methods

# **Animals and Housing Conditions**

All experiments were conducted under the authority of United Kingdom Animals (Scientific Procedures) Act 1986 and with the approval of University of Cambridge Animal Welfare and Ethical Review Panel, and are in compliance with the ARRIVE guidelines. R6/2 and WT littermate mice were taken from a colony established at the University of Cambridge (CBA  $\times$  C57/BL6 background). Genotyping and repeat length measurements were performed by Laragen (Los Angeles, CA, USA), as described previously [9]. R6/2 mice had a mean CAG repeat length of  $252\pm3$ . In the acute drug treatment study, 8 WT and 10 R6/2 male mice were used. Before the end of study, 1 WT and 1 R6/2 mouse lost their EEG/electromyogram (EMG) implants and were euthanized. In

addition, 1 R6/2 mouse died of its disease. In the chronic drug treatment experiment, 12 male R6/2 mice were used. One R6/2 mouse lost its EEG/EMG implant before the end of study and was euthanized.

## Surgery and EEG/EMG Recordings

We implanted each mouse with EEG and EMG electrodes under isoflurane anesthesia (1.5–2%) [10]. Briefly, we placed screw electrodes epidurally over the frontal (1.5 mm lateral and 1.0 mm anterior to bregma) and parietal (1.5 mm lateral and 1.0 mm anterior to lambda) cortices for frontoparietal EEG recordings. EMG signals were acquired by a pair of stainless steel spring wires inserted into the neck extensor muscles. At the time of surgery, mice were 9 to 10 weeks of age.

After surgery, mice were housed individually under standard conditions [10]. After a recovery period of 7 to 10 days, we connected the mice to recording cables and left them to acclimatize for an additional 3 to 4 days before recording their EEG/EMG signals. The mice then remained connected to the recording cable throughout the study.

EEG/EMG signals were amplified and filtered (EEG: 0.5–60 Hz; EMG: 10–100 Hz) by head-mounted preamplifiers and amplifiers (8202-DSL and 8206-SL, respectively; Pinnacle Technology, Lawrence, KS, USA), and recorded on a computer (Vital Recorder, Kissei Comtec, Matsumoto, Japan) after analog-to-digital conversion.

# **Drug Administration**

Acute Treatment

We treated WT and R6/2 mice acutely with 3 different doses of paroxetine (5, 10, or 20 mg/kg; Sigma-Aldrich, Gillingham, UK) or vehicle (0.9% saline). To test whether paroxetine can correct the abnormally increased REM sleep seen in R6/2 mice during the active dark period [9, 10], we treated the mice just before the onset of the dark period. Then we recorded their sleep—wake behavior for 24 h. The mice were between 12 and 14 weeks of age at the time of treatment. The mice were given vehicle or paroxetine by intraperitoneal injection in a volume of 10 ml/kg body weight. The different doses of paroxetine and its vehicle were given to the mice in a crossover design and in a randomized order, with 3 to 4 days between the treatments. The doses were chosen based on the literature [26–28] and our pilot experiments (data not shown).

## Chronic Treatment

We treated R6/2 mice daily for 8 weeks with either 20 mg/kg paroxetine (n = 6), or vehicle (n = 6), with treatment starting at 6 weeks of age. At the age of 9 to 10 weeks, we implanted the



mice with EEG/EMG electrodes. The mice were not treated with paroxetine on the day of surgery. We recorded sleep—wake behavior in the mice on several different occasions during treatment, and on 2 occasions after treatment had stopped. On EEG/EMG recording days, the mice received the treatment just before the onset of the dark period, and their sleep—wake behavior was recorded from the beginning of dark period for 24 h, as it was in the acute study. This allowed us to compare directly the effect of acute and chronic treatments on sleep and EEG parameters in R6/2 mice. On the days when no recordings were made during the chronic study, the mice were treated at random times between 8 a.m. and 4 p.m. during the day, to avoid any potential zeitgeber effect of treatments.

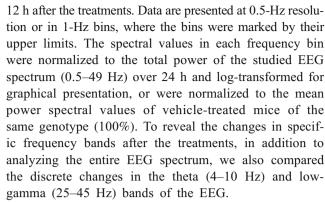
Successful EEG/EMG recordings with all four treatments were achieved in 6/8 WT mice in the acute drug study. Of the other 2 WT mice, 1 WT mouse received all treatments apart from the 5 mg/kg dose of paroxetine and 1 WT mouse received vehicle and the 20 mg/kg doses of paroxetine. Seven of 10 R6/2 mice received all the treatments in the acute drug experiment. Of the other 3 R6/2 mice, 2 received vehicle and all doses of paroxetine, apart from either the 5 mg/kg (1 mouse) or the 20 mg/kg (1 mouse) doses of the drug. One R6/2 mouse received vehicle and the 20 mg/kg dose of paroxetine.

In the chronic drug treatment experiment, successful EEG/EMG recordings were made in all 6 paroxetine-treated and 5/6 vehicle-treated R6/2 mice at all time points. One vehicle-treated R6/2 mouse had all the EEG/EMG recordings made from it, except the final recording that should have been done 2 weeks after drug treatment stopped.

# **Data Analysis and Statistics**

All signals were digitized at 256 Hz, digitally filtered (EEG: 0.5–60 Hz; EMG: 10–60 Hz), and semi-automatically scored as wake, non-REM (NREM) sleep, or REM sleep in 10-s epochs using SleepSign (Kissei Comtec, Matsumoto, Japan). Experienced scorers, blinded to treatment and genotype, visually inspected these preliminary scorings and made corrections when appropriate. We then measured the duration of bouts, counted the number of bouts, and calculated the time spent in each behavioral state during both dark and light periods. We also analyzed the probability of transitioning into REM sleep as a function of NREM sleep bout length, as described previously [9].

To reveal the changes in the frequency content of the recorded signal, we performed a power spectral analysis of the EEG after the treatments. EEG power spectra were computed for artifact-free 2-s epochs by fast Fourier transformation, as described previously [10]. The values of consecutive 2-s EEG epochs in wake, NREM, and REM sleep were averaged over



To compare statistically the vigilance state parameters and normalized EEG data, we used multivariate analysis of variance with repeated measures and unpaired t-tests (Statistica 13; Statsoft, Tulsa, OK, USA). The results were considered statistically significant at p < 0.05. All results are expressed as means  $\pm$  SEM.

## **Results**

# Paroxetine Suppresses REM Sleep and Consolidates NREM Sleep

Acute treatment with paroxetine dose-dependently decreased the amount of REM sleep in both WT and R6/2 mice [drug effect:  $F_{(3,33)} = 20.26$ ; p < 0.01]. During the first 12 h, the amount of REM sleep was decreased by > 60% in both WT and R6/2 mice after the highest dose (20 mg/kg i.p.) of paroxetine (Table 1). The REM sleep-suppressing effect of paroxetine was restricted to the first 12 h in WT mice but extended into the second 12 h in R6/2 mice [drug × dark/light period interaction:  $F_{(3.33)} = 4.01$ ; p < 0.05 (Table 1)]. Paroxetine suppressed REM sleep by reducing its propensity in both genotypes, as shown by a reduction in the number of REM sleep bouts [drug effect:  $F_{(3,33)} = 21.79$ ; p < 0.01 (Table 1)]. The reduction in REM sleep propensity is also shown by the decreased probability of transitioning from NREM sleep into REM sleep in paroxetine-treated mice, irrespective of genotype or dark/light period [drug effect:  $F_{(1,14)} = 37.36$ ; p < 0.01(Fig. 1a-d)]. In WT mice, paroxetine also improved the maintenance of REM sleep, as indicated by longer REM sleep bouts after paroxetine treatment than after vehicle treatment [drug effect:  $F_{(3.30)} = 4.99$ ; p < 0.01 (Table 1)]. However, this effect of paroxetine depended on the dark/light period and could be seen during the light period only [drug × dark/light period interaction:  $F_{(3,30)} = 3.40$ ; p < 0.05 (Table 1)].

In addition to REM sleep suppression, paroxetine exerted a dose-dependent hypnotic effect in both WT and R6/2 mice [drug effect:  $F_{(3,33)} = 9.12$ ; p < 0.01 (Table 1)]. This hypnotic effect of paroxetine, shown by an increase in the amount of



Table 1 Vigilance state parameters in wild-type (WT) and R6/2 mice after vehicle or paroxetine treatment

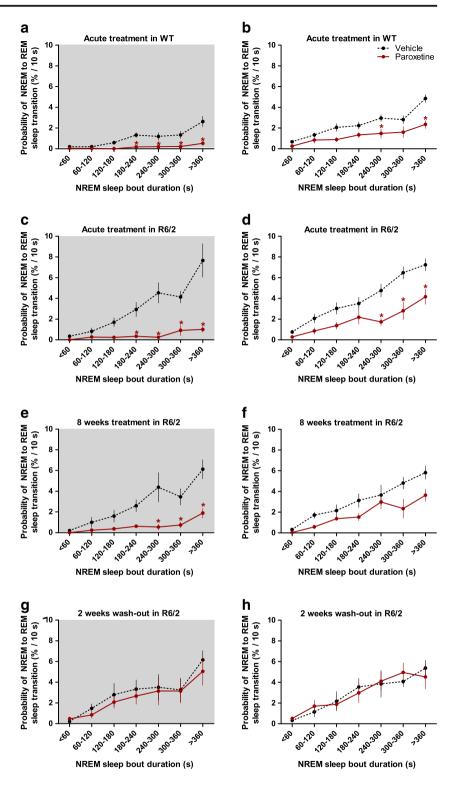
Operation         Achicle (n=8)         Singlégie (n=8)         ( nongègie (n=8))         ( volude (n=8))         ( nongègie (n=8))         ( nongèg	Genotype	WT				R6/2			
(min)  184.3 ± 7.5  184.4 ± 3.5  184.3 ± 7.5	Dose	Vehicle $(n=8)$	5  mg/kg  (n=6)	10  mg/kg  (n=7)	20  mg/kg  (n=8)	Vehicle $(n = 10)$	5  mg/kg  (n=8)	10  mg/kg  (n=9)	20  mg/kg  (n=9)
1843 ± 1.5   1871 ± 1.6   180 ± 1.5   1824 ± 1.0   11.5 ± 8.4   2.95 ± 1.2   2.95 ± 1.2   2.95 ± 1.0   2.95	Wake								
1800±205   397,1±168   2895±175*   298±445*   395,7±176   2555±120     1843±7.5   187,1±121   1796±5.9   1824±100   212.5±8.4   2092±8.5   2039±10.1     1843±7.5   187,1±121   1796±5.9   1824±100   212.5±8.4   2092±8.5   2039±10.1     1843±7.5   187,1±121   1796±5.9   1824±100   212.5±8.4   2092±8.5   2039±10.1     1843±7.5   187,1±121   1796±5.9   1824±100   212.5±8.4   2092±8.5   2039±10.1     1843±7.5   187,1±121   1796±5.9   1824±100   212.5±8.4   49±0.3   30±0.2     1900ms   27±0.4   2.7±0.5   24±0.3   211±0.1   26±0.2   30±0.2   30±0.3     1900ms   238±216   3897±189   4173±19.5*   411.7±16.0*   3218±17.5   3779±16.7   4106±13.2*     1000ms   338±216   3897±189   4173±19.5*   411.7±16.0*   33±0.2   34±0.2   34±0.2     1100±8.5   1110±9.3   49±0.4   47±0.3   30±0.2   34±0.2     1100±8.5   1110±9.3   49±0.4   47±0.3   444±3.*     1100±8.5   1110±9.3   483±2.5   444±3.*     1100±8.5   1110±9.3   483±2.5   444±3.*     1100±8.5   1110±9.3   483±2.5   444±3.*     1100±8.5   1110±9.3   483±2.5   444±3.*     1100±8.5   1110±9.3   41.7±0.2   11.7±0.2   11.2±0.1     1100±9.5   44.5±0.3   44.5±0.3     1100±9.5   44.5±0.3   44.5±0.3     1100±0.5   44.5±0.3	Total time (min)								
1843 ± 7.5   187.1 ± 12.1   179 6 ± 5.9   182.4 ± 10.0   212.5 ± 8.4   209.2 ± 8.5   205.9 ± 10.1   100 cmin)     cond	Dark period	$350.0 \pm 20.5$	$307.1\pm16.8$	$289.5 \pm 17.5 *$	$298.3 \pm 14.5 *$	$305.7\pm18.3$	$297.7 \pm 17.6$	$275.5 \pm 12.0$	$271.4 \pm 10.5$
rick         5.2±6.9         4.5±0.4         4.6±0.6         4.0±0.4         4.9±0.3         4.9±0.3           rick         5.2±6.9         4.5±0.4         4.5±0.4         4.6±0.6         4.0±0.4         3.0±0.2         3.0±0.3           rounts         7.8±6.9         2.4±0.3         2.1±0.1         2.6±0.2         3.0±0.2         3.0±0.3           rounts         7.8±6.11         7.0±6.9         7.08±6.1         7.08±6.1         7.08±6.0         5.8±2.6           rick         7.3±6.8         7.45±6.3         80.1±6.7         90.0±5.0         82.2±4.8         70.1±3.4         72.2±6.0           rick         3.3±6.8         7.45±6.3         417.3±19.5°         411.7±16.0°         82.2±4.8         70.1±3.4         72.2±6.0           rick         4.55±6.3         4.7±6.5         4.11.7±16.0°         82.2±4.8         70.1±3.4         72.2±6.0           rick         4.5±6.3         4.7±6.5         4.11.7±16.0°         351.8±17.5         377.9±16.7         410.6±13.2°           rick         3.9±0.2         4.2±0.3         4.9±0.4         4.7±0.3         3.3±0.2         5.5±0.4°         6.5±0.3           rick         3.9±0.2         4.2±0.3         4.9±0.4         4.7±0.3         3.0±0.2         3.8±0.	Light period	$184.3 \pm 7.5$	$187.1\pm12.1$	$179.6 \pm 5.9$	$182.4\pm10.0$	$212.5\pm8.4$	$209.2\pm8.5$	$205.9\pm10.1$	$217.8 \pm 8.8$
iod 2.7±0.9 4.5±0.4 4.0±0.4 4.0±0.4 4.0±0.3 30±0.3	Mean duration (mi	(u							
riod 2.7±0.4 2.7±0.5 2.4±0.3 2.1±0.1 2.6±0.2 30±0.2 30±0.3 bouns ind 79.5±6.8	Dark period	$5.2 \pm 0.9$	$4.5 \pm 0.4$	$4.2 \pm 0.4$	$4.6 \pm 0.6$	$4.0\pm0.4$	$4.9 \pm 0.3$	$4.9 \pm 0.3$	$4.9 \pm 0.7$
bounds         bounds           ried         78.4 ± 11.1         70.5 ± 3.4         71.9 ± 6.9         70.8 ± 6.1         70.1 ± 3.4         70.1 ± 3.4         70.2 ± 6.0           ried         73.3 ± 6.8         74.5 ± 6.3         80.1 ± 6.7         90.0 ± 5.0         82.2 ± 4.8         70.1 ± 3.4         72.2 ± 6.0           ried         45.3 ± 6.1.5         471.6 ± 5.7         471.7 ± 16.0*         351.8 ± 17.5         377.9 ± 16.7         410.6 ± 13.2*           ried         456.3 ± 8.4         454.6 ± 11.5         471.6 ± 5.7         472.5 ± 13.2         408.6 ± 7.8         417.2 ± 8.7         428.0 ± 8.3           ried         3.9 ± 0.2         4.2 ± 0.3         4.9 ± 0.4         4.7 ± 0.3         30.6 ± 0.2         3.5 ± 0.4         45.5 ± 0.3*           bounts         3.9 ± 0.2         4.2 ± 0.3         4.9 ± 0.4         4.7 ± 0.3         30.6 ± 0.2         3.5 ± 0.4*         45.5 ± 0.3*           ried         3.9 ± 0.2         4.2 ± 0.3         4.9 ± 0.4         4.7 ± 0.3         30.6 ± 0.2         3.5 ± 0.4*         45.5 ± 0.3*           ried         3.9 ± 0.2         4.2 ± 0.3         4.2 ± 0.3         3.0 ± 0.2         3.2 ± 0.4*         45.5 ± 0.3*         45.5 ± 0.3*           ried         3.9 ± 2.2         4.2 ± 0.3 <th< th=""><td>Light period</td><td><math display="block">2.7\pm0.4</math></td><td><math display="block">2.7\pm0.5</math></td><td><math display="block">2.4\pm0.3</math></td><td><math display="block">2.1\pm0.1</math></td><td><math display="block">2.6\pm0.2</math></td><td><math>3.0 \pm 0.2</math></td><td><math>3.0 \pm 0.3</math></td><td><math display="block">3.1\pm0.3</math></td></th<>	Light period	$2.7\pm0.4$	$2.7\pm0.5$	$2.4\pm0.3$	$2.1\pm0.1$	$2.6\pm0.2$	$3.0 \pm 0.2$	$3.0 \pm 0.3$	$3.1\pm0.3$
iod 73.3 ± 6.8 1.11 70.5 ± 3.4 71.9 ± 6.9 70.8 ± 6.1 79.8 ± 5.0 61.4 ± 4.6 56.8 ± 2.6 6 iod 73.3 ± 6.8 74.5 ± 6.3 80.1 ± 6.7 90.0 ± 5.0 82.2 ± 4.8 70.1 ± 3.4 72.2 ± 6.0 6 iod 73.3 ± 6.8 73.3 ± 6.8 73.3 ± 6.8 73.3 ± 6.8 73.3 ± 6.8 73.3 ± 6.2 70.1 ± 3.4 72.2 ± 6.0 6 ± 6.2 7 70.2 ± 6.0 7 70.2 ± 6.0 7 70.2 ± 6.0 7 70.2 ± 6.0 7 70.2 ± 6.0 7 70.2 ± 6.0 7 70.2 ± 6.0 7 70.2 ± 6.0 7 70.2 ± 6.0 7 70.2 ± 6.0 7 70.2 ± 6.0 ± 6.2	Number of bouts								
riod   73.3 ± 6.8   74,5 ± 6.3   80.1 ± 6.7   90.0 ± 5.0   82.2 ± 4.8   70.1 ± 3.4   72.2 ± 6.0      min	Dark period	$78.6\pm11.1$	$70.5 \pm 3.4$	$71.9 \pm 6.9$	$70.8\pm6.1$	$79.8 \pm 5.0$	$61.4 \pm 4.6$	$56.8 \pm 2.6$	$62.6 \pm 7.7$
(min)  ind 338.1±21.6 389.7±18.9 417.3±19.5* 411.7±16.0* 351.8±17.5 377.9±16.7 410.6±13.2*  ind 456.3±8.4 454.6±11.5 471.6±5.7 472.5±13.2 408.6±7.8 417.2±8.7 428.0±8.3  ind 3.9±0.3 5.3±0.2 6.0±0.7 6.0±0.6* 3.3±0.2 5.5±0.4* 6.5±0.3*  ind 3.9±0.2 4.2±0.3 4.9±0.4 4.7±0.3 3.0±0.2 38±0.2 4.5±0.3*  ind 3.9±0.2 4.2±0.3 4.9±0.4 4.7±0.3 3.0±0.2 38±0.2 4.5±0.3*  ind 3.9±0.2 74.8±4.9 74.1±6.8 71.5±5.9* 103.4±4.8* 137.1±7.0 112.1±5.0 98.9±7.5*  ind 31.9±5.2 23.2±2.9 13.2±3.0 10.0±2.9* 62.5±3.2 44.4±3.5* 86.1±3.8*  ind 1.3±0.1 1.4±0.1 1.7±0.2 1.7±0.2* 1.3±0.1 1.5±0.1 1.5±0.1 1.5±0.1 1.6±0.1 1.0±0.1*  ind 1.2±0.1 1.3±0.1 1.6±0.2 1.8±0.2* 1.3±0.1 1.5±0.1 1.4±0.1 1.5±0.1 1.0±0.1*  ind 23.4±3.5 16.7±1.6 83.2±2.5 64.2±1 50.0±3.5* 50.0±3.5* 56.5±0.* 56.5±0.*  ind 23.4±3.5 16.7±1.6 83.2±2.5 64.2±1 50.0±3.5* 56.5±0.* 56.5±0.*  ind 23.4±3.5 16.7±1.6 83.2±2.5 64.2±1 50.0±3.5* 56.5±0.* 56.5±0.*	Light period	$73.3 \pm 6.8$	$74.5 \pm 6.3$	$80.1\pm6.7$	$90.0 \pm 5.0$	$82.2\pm4.8$	$70.1 \pm 3.4$	$72.2\pm6.0$	$72.7 \pm 6.3$
eriod 338.1±21.6 389.7±18.9 417.3±19.5* 411.7±16.0* 351.8±17.5 377.9±16.7 410.6±13.2* artion (min) criod 3.9±0.3 5.3±0.2 6.0±0.7 4.7±5.13.2 408.6±7.8 417.2±8.7 428.0±8.3 artion (min) criod 3.9±0.2 4.2±0.3 4.9±0.4 4.7±0.3 3.3±0.2 5.5±0.4* 6.5±0.3* criod 3.9±0.2 4.2±0.3 4.9±0.4 4.7±0.3 3.0±0.2 3.8±0.2 4.2±0.3 4.9±0.4 4.7±0.8 3.3±0.2 3.8±0.2 4.2±0.3 4.9±0.4 4.7±0.8 71.5±5.9* 110.0±2.9* 110.0±2.9* 6.25±3.2 4.4±3.5* 6.3±2.2 6.2±2.2 78.3±3.3 6.8±2.2 6.2±2.2 1.3±0.1 1.3±0	NREM sleep								
eriod $338.1\pm21.6$ $389.7\pm18.9$ $417.3\pm19.5^*$ $411.7\pm16.0^*$ $351.8\pm17.5$ $377.9\pm16.7$ $410.6\pm13.2^*$ veriod $456.3\pm8.4$ $454.6\pm11.5$ $471.6\pm5.7$ $472.5\pm13.2$ $408.6\pm7.8$ $417.2\pm8.7$ $410.6\pm13.2^*$ veriod $3.9\pm0.3$ $5.3\pm0.2$ $6.0\pm0.6$ $6.0\pm0.6$ $3.3\pm0.2$ $5.5\pm0.4$ $6.5\pm0.3^*$ veriod $3.9\pm0.2$ $4.2\pm0.3$ $4.9\pm0.4$ $4.7\pm0.3$ $3.3\pm0.2$ $5.5\pm0.4^*$ $6.5\pm0.3^*$ veriod $3.9\pm0.2$ $4.2\pm0.3$ $4.9\pm0.4$ $4.7\pm0.3$ $3.0\pm0.2$ $3.8\pm0.2$ $6.5\pm0.3^*$ veriod $3.9\pm0.2$ $4.2\pm0.3$ $4.7\pm0.4$ $4.7\pm0.3$ $3.0\pm0.2$ $3.8\pm0.2$ $4.5\pm0.3$ veriod $11.90\pm8.5$ $111.0\pm9.3$ $98.9\pm7.5$ $10.0\pm2.9^*$ $62.5\pm3.2$ $44.4\pm3.5^*$ $44.4\pm3.5^*$ $44.4\pm3.5^*$ $44.4\pm3.5^*$ veriod $11.9\pm5.2$ $23.2\pm2.9$ $13.2\pm3.0$ $10.0\pm2.9^*$ $62.5\pm3.2$ $44.4\pm3.5^*$ $44.4\pm3.5^*$ veriod	Total time (min)								
eriod $456.3\pm 8.4$ $454.6\pm 11.5$ $471.6\pm 5.7$ $472.5\pm 13.2$ $408.6\pm 7.8$ $417.2\pm 8.7$ $428.0\pm 8.3$ eriod $3.9\pm 0.3$ $5.3\pm 0.2$ $6.0\pm 0.6$ * $3.3\pm 0.2$ $5.5\pm 0.4$ * $6.5\pm 0.3$ *           eriod $3.9\pm 0.2$ $4.2\pm 0.3$ $4.9\pm 0.4$ $4.7\pm 0.3$ $3.0\pm 0.2$ $3.8\pm 0.2$ $4.5\pm 0.3$ of bours         eriod $8.9\pm 10.6$ $74.8\pm 4.9$ $74.1\pm 6.8$ $71.5\pm 5.9$ * $108.7\pm 6.8$ $70.8\pm 4.9$ * $6.5\pm 0.3$ *           eriod $8.9\pm 10.6$ $74.8\pm 4.9$ $74.1\pm 6.8$ $71.5\pm 5.9$ * $108.7\pm 6.8$ $70.8\pm 4.9$ * $6.5\pm 0.3$ *           eriod $11.0\pm 8.5$ $111.0\pm 9.3$ $98.9\pm 7.5$ $10.3\pm 4.4$ * $13.7\pm 7.0$ $112.1\pm 5.0$ $112.1\pm 0.1$	Dark period	$338.1\pm21.6$	$389.7\pm18.9$	$417.3 \pm 19.5^*$	$411.7 \pm 16.0$ *	$351.8\pm17.5$	$377.9\pm16.7$	$410.6 \pm 13.2 *$	$426.7\pm10.8^*$
ation (min) eriod 3.9±0.3 5.3±0.2 6.0±0.7 6.0±0.6* 3.3±0.2 5.5±0.4* 6.5±0.3* eriod 3.9±0.2 4.2±0.3 4.9±0.4 4.7±0.3 3.0±0.2 3.8±0.2 4.5±0.3 4.5±0.3 beriod 3.9±0.2 4.2±0.3 4.9±0.4 4.7±0.3 3.0±0.2 3.8±0.2 4.5±0.3 4.5±0.3 eriod 8.99±10.6 74.8±4.9 74.1±6.8 71.5±5.9* 108.7±6.8 70.8±4.9* 63.7±2.1* eriod 119.0±8.5 111.0±9.3 98.9±7.5 103.4±4.8* 137.1±7.0 112.1±5.0 98.9±7.5* eriod 31.9±5.2 23.2±2.9 78.3±3.3 68.8±2.2 65.0±5.2 98.9±3.3 93.7±3.1 86.1±3.8 eriod 79.5±2.9 78.3±3.3 68.8±2.2 65.0±5.2 98.9±3.3 93.7±3.1 1.6±0.1 eriod 1.2±0.1 1.4±0.1 1.6±0.2 1.8±0.2* 1.2±0.1 1.4±0.1 1.5±0.1 1.4±0.1 1.5±0.1 eriod 2.3±3.5 16.7±1.6 83±2.5 64±2.1 50.0±4.5 69.5±3.3 50.0±3.5* 56.6±4.0*	Light period	$456.3 \pm 8.4$	$454.6\pm11.5$	$471.6\pm5.7$	$472.5 \pm 13.2$	$408.6 \pm 7.8$	$417.2\pm8.7$	$428.0 \pm 8.3$	$421.6\pm9.4$
eriod 3.9 ± 0.3 5.3 ± 0.2 6.0 ± 0.7 6.0 ± 0.6 * 3.3 ± 0.2 5.5 ± 0.4 * 6.5 ± 0.3 * 4.2 ± 0.3 4.2	Mean duration (mi	(u							
errind         3.9 ± 0.2         4.2 ± 0.3         4.9 ± 0.4         4.7 ± 0.3         3.0 ± 0.2         3.8 ± 0.2         4.5 ± 0.3           of bours         seriod         49.9 ± 10.6         74.8 ± 4.9         74.1 ± 6.8         71.5 ± 5.9*         108.7 ± 6.8         70.8 ± 4.9*         63.7 ± 2.1*           veriod         119.0 ± 8.5         111.0 ± 9.3         98.9 ± 7.5         103.4 ± 4.8*         137.1 ± 7.0         112.1 ± 5.0         98.9 ± 7.5*           erriod         31.9 ± 5.2         23.2 ± 2.9         13.2 ± 3.0         10.0 ± 2.9*         62.5 ± 3.2         44.4 ± 3.5*         98.9 ± 7.5*           erriod         79.5 ± 2.9         78.3 ± 3.3         68.8 ± 2.2         65.0 ± 5.2         98.9 ± 3.3         93.7 ± 3.1         86.1 ± 3.8           erriod         1.3 ± 0.1         1.7 ± 0.2         1.7 ± 0.2         1.3 ± 0.1         1.5 ± 0.1         1.5 ± 0.1           erriod         1.2 ± 0.1         1.3 ± 0.1         1.6 ± 0.2         1.8 ± 0.2*         98.9 ± 3.3         93.7 ± 3.1         1.5 ± 0.1           erriod         1.2 ± 0.1         1.3 ± 0.1         1.6 ± 0.2         1.8 ± 0.2*         1.2 ± 0.1         1.4 ± 0.1         1.5 ± 0.1           erriod         2.3.4 ± 3.5         6.3 ± 2.5         6.4 ± 2.1         50.0 ± 4.5 <td>Dark period</td> <td><math>3.9 \pm 0.3</math></td> <td><math display="block">5.3\pm0.2</math></td> <td><math>6.0 \pm 0.7</math></td> <td><math>6.0 \pm 0.6</math></td> <td><math display="block">3.3\pm0.2</math></td> <td><math display="block">5.5\pm0.4*</math></td> <td><math display="block">6.5\pm0.3*</math></td> <td><math>7.0 \pm 0.7*</math></td>	Dark period	$3.9 \pm 0.3$	$5.3\pm0.2$	$6.0 \pm 0.7$	$6.0 \pm 0.6$	$3.3\pm0.2$	$5.5\pm0.4*$	$6.5\pm0.3*$	$7.0 \pm 0.7*$
of bouts         Solution         Seried         71.5 ± 5.9*         108.7 ± 6.8         70.8 ± 4.9*         63.7 ± 2.1*           eriod         89.9 ± 10.6         74.8 ± 4.9         74.1 ± 6.8         71.5 ± 5.9*         108.7 ± 6.8         70.8 ± 4.9*         63.7 ± 2.1*           eriod         119.0 ± 8.5         111.0 ± 9.3         98.9 ± 7.5         103.4 ± 4.8*         137.1 ± 7.0         112.1 ± 5.0         98.9 ± 7.5*           eriod         31.9 ± 5.2         23.2 ± 2.9         13.2 ± 3.0         10.0 ± 2.9*         62.5 ± 3.2         44.4 ± 3.5*         33.9 ± 2.8*           eriod         79.5 ± 2.9         78.3 ± 3.3         68.8 ± 2.2         65.0 ± 5.2         98.9 ± 3.3         93.7 ± 3.1         86.1 ± 3.8           ation (min)         eriod         1.3 ± 0.1         1.7 ± 0.2         1.3 ± 0.1         1.5 ± 0.1         1.5 ± 0.1           eriod         1.2 ± 0.1         1.3 ± 0.1         1.6 ± 0.2         1.8 ± 0.2*         1.2 ± 0.1         1.4 ± 0.1         1.5 ± 0.1           prind         23.4 ± 3.5         66.4 ± 2.1         50.0 ± 4.5         50.0 ± 3.5         56.6 ± 4.0*	Light period	$3.9\pm0.2$	$4.2 \pm 0.3$	$4.9 \pm 0.4$	$4.7 \pm 0.3$	$3.0\pm0.2$	$3.8 \pm 0.2$	$4.5\pm0.3$	$4.9\pm0.5*$
eriod 89.9±10.6 74.8±4.9 74.1±6.8 71.5±5.9* 108.7±6.8 70.8±4.9* 63.7±2.1*  beriod 119.0±8.5 111.0±9.3 98.9±7.5 103.4±4.8* 137.1±7.0 112.1±5.0 98.9±7.5*  c (min)  eriod 31.9±5.2 23.2±2.9 13.2±3.0 10.0±2.9* 62.5±3.2 44.4±3.5* 33.9±2.8*  beriod 79.5±2.9 78.3±3.3 68.8±2.2 65.0±5.2 98.9±3.3 93.7±3.1 86.1±3.8  atton (min)  eriod 1.3±0.1 1.4±0.1 1.7±0.2 1.7±0.2 1.3±0.1 1.5±0.1 1.5±0.1  of bouts  eriod 23.4±3.5 16.7±1.6 83.±2.5 64.4±2.1 50.0±4.5 10.0±3.5* 21.8±1.2*  eriod 23.4±3.5 63.2±4.5 46.0±5.7* 39.9±6.4* 82.1±6.5 69.5±5.3 56.6±4.0*	Number of bouts								
eriod 119.0±8.5 111.0±9.3 98.9±7.5 103.4±4.8* 137.1±7.0 112.1±5.0 98.9±7.5*  etiod 31.9±5.2 23.2±2.9 13.2±3.0 10.0±2.9* 62.5±3.2 44.4±3.5* 33.9±2.8*  eriod 31.9±5.2 78.3±3.3 68.8±2.2 65.0±5.2 98.9±3.3 93.7±3.1 86.1±3.8  ation (min)  eriod 1.2±0.1 1.4±0.1 1.7±0.2 1.7±0.2 1.3±0.1 1.5±0.1 1.5±0.1  of bouts  eriod 23.4±3.5 64.4±2.1 50.0±4.5 60.5±5.3 30.0±3.5* 21.8±1.2*  eriod 23.4±3.5 63.2±4.5 46.0±5.7* 39.9±6.4* 82.1±6.5 69.5±5.3 56.6±4.0*	Dark period	$89.9\pm10.6$	$74.8 \pm 4.9$	$74.1 \pm 6.8$	$71.5 \pm 5.9*$	$108.7\pm6.8$	$70.8\pm4.9*$	$63.7 \pm 2.1 *$	$67.6 \pm 8.0 *$
eriod 31.9±5.2 23.2±2.9 13.2±3.0 10.0±2.9* 62.5±3.2 44.4±3.5* 33.9±2.8* beriod 79.5±2.9 78.3±3.3 68.8±2.2 65.0±5.2 98.9±3.3 93.7±3.1 86.1±3.8 ation (min) eriod 1.3±0.1 1.4±0.1 1.7±0.2 1.7±0.2 1.3±0.1 1.5±0.1 1.	Light period	$119.0\pm 8.5$	$111.0\pm9.3$	$98.9 \pm 7.5$	$103.4\pm4.8*$	$137.1 \pm 7.0$	$112.1 \pm 5.0$	$98.9 \pm 7.5 *$	$91.9\pm 8.4*$
31.9±5.2       23.2±2.9       13.2±3.0       10.0±2.9*       62.5±3.2       44.4±3.5*       33.9±2.8*         79.5±2.9       78.3±3.3       68.8±2.2       65.0±5.2       98.9±3.3       93.7±3.1       86.1±3.8         nin)       1.3±0.1       1.7±0.2       1.7±0.2       1.3±0.1       1.5±0.1       1.6±0.1         1.2±0.1       1.3±0.1       1.6±0.2       1.8±0.2*       1.2±0.1       1.4±0.1       1.5±0.1         23.4±3.5       16.7±1.6       8.3±2.5       6.4±2.1       50.0±4.5       30.0±3.5*       21.8±1.2*         68.3±3.2       63.2±4.5       46.0±5.7*       39.9±6.4*       82.1±6.5       69.5±5.3       56.6±4.0*	REM sleep								
31.9±5.2       23.2±2.9       13.2±3.0       10.0±2.9*       62.5±3.2       44.4±3.5*       33.9±2.8*         79.5±2.9       78.3±3.3       68.8±2.2       65.0±5.2       98.9±3.3       93.7±3.1       86.1±3.8         nin)       1.3±0.1       1.7±0.2       1.7±0.2       1.3±0.1       1.5±0.1       1.6±0.1         1.2±0.1       1.3±0.1       1.6±0.2       1.8±0.2*       1.2±0.1       1.4±0.1       1.5±0.1         23.4±3.5       16.7±1.6       8.3±2.5       6.4±2.1       50.0±4.5       30.0±3.5*       21.8±1.2*         68.3±3.2       63.2±4.5       46.0±5.7*       39.9±6.4*       82.1±6.5       69.5±5.3       56.6±4.0*	Total time (min)								
79.5±2.9       78.3±3.3       68.8±2.2       65.0±5.2       98.9±3.3       93.7±3.1       86.1±3.8         nin)       1.3±0.1       1.7±0.2       1.7±0.2       1.3±0.1       1.5±0.1       1.6±0.1         1.2±0.1       1.3±0.1       1.5±0.1       1.6±0.1       1.5±0.1         1.2±0.1       1.3±0.1       1.4±0.1       1.5±0.1         23.4±3.5       16.7±1.6       8.3±2.5       6.4±2.1       50.0±4.5       30.0±3.5*       21.8±1.2*         68.3±3.2       63.2±4.5       46.0±5.7*       39.9±6.4*       82.1±6.5       69.5±5.3       56.6±4.0*	Dark period	$31.9\pm5.2$	$23.2 \pm 2.9$	$13.2 \pm 3.0$	$10.0 \pm 2.9 *$	$62.5 \pm 3.2$	$44.4 \pm 3.5 *$	$33.9\pm2.8*$	$21.9\pm4.5*$
$\begin{array}{llllllllllllllllllllllllllllllllllll$	Light period	$79.5 \pm 2.9$	$78.3 \pm 3.3$	$68.8 \pm 2.2$	$65.0 \pm 5.2$	$98.9 \pm 3.3$	$93.7 \pm 3.1$	$86.1 \pm 3.8$	$80.7 \pm 3.4$ *
$1.3\pm0.1$ $1.4\pm0.1$ $1.7\pm0.2$ $1.7\pm0.2$ $1.3\pm0.1$ $1.5\pm0.1$ $1.5\pm0.1$ $1.6\pm0.1$ $1.2\pm0.1$ $1.3\pm0.1$ $1.6\pm0.2$ $1.8\pm0.2*$ $1.2\pm0.1$ $1.4\pm0.1$ $1.5\pm0.1$ $2.3.4\pm3.5$ $16.7\pm1.6$ $8.3\pm2.5$ $6.4\pm2.1$ $50.0\pm4.5$ $30.0\pm3.5*$ $21.8\pm1.2*$ $68.3\pm3.2$ $63.2\pm4.5$ $46.0\pm5.7*$ $39.9\pm6.4*$ $82.1\pm6.5$ $69.5\pm5.3$ $56.6\pm4.0*$	Mean duration (mi	_							
$1.2 \pm 0.1 \qquad 1.3 \pm 0.1 \qquad 1.6 \pm 0.2 \qquad 1.8 \pm 0.2 * \qquad 1.2 \pm 0.1 \qquad 1.4 \pm 0.1 \qquad 1.5 \pm 0.1$ $23.4 \pm 3.5 \qquad 16.7 \pm 1.6 \qquad 8.3 \pm 2.5 \qquad 6.4 \pm 2.1 \qquad 50.0 \pm 4.5 \qquad 30.0 \pm 3.5 * \qquad 21.8 \pm 1.2 * \qquad 68.3 \pm 3.2 \qquad 69.5 \pm 5.3 \qquad 56.6 \pm 4.0 * \qquad 21.8 \pm 1.2 * \qquad 21.8 \pm 1.$	Dark period	$1.3\pm0.1$	$1.4\pm0.1$	$1.7\pm0.2$	$1.7 \pm 0.2$	$1.3\pm0.1$	$1.5\pm0.1$	$1.6\pm0.1$	$1.5\pm0.2$
$23.4 \pm 3.5 \qquad 16.7 \pm 1.6 \qquad 8.3 \pm 2.5 \qquad 6.4 \pm 2.1 \qquad 50.0 \pm 4.5 \qquad 30.0 \pm 3.5 * \qquad 21.8 \pm 1.2 * \\ 68.3 \pm 3.2 \qquad 63.2 \pm 4.5 \qquad 46.0 \pm 5.7 * \qquad 39.9 \pm 6.4 * \qquad 82.1 \pm 6.5 \qquad 69.5 \pm 5.3 \qquad 56.6 \pm 4.0 *$	Light period	$1.2\pm0.1$	$1.3\pm0.1$	$1.6\pm0.2$	$1.8 \pm 0.2*$	$1.2\pm0.1$	$1.4 \pm 0.1$	$1.5\pm0.1$	$1.7\pm0.1$
$23.4 \pm 3.5 \qquad 16.7 \pm 1.6 \qquad 8.3 \pm 2.5 \qquad 6.4 \pm 2.1 \qquad 50.0 \pm 4.5 \qquad 30.0 \pm 3.5 * \qquad 21.8 \pm 1.2 * \\ 68.3 \pm 3.2 \qquad 63.2 \pm 4.5 \qquad 46.0 \pm 5.7 * \qquad 39.9 \pm 6.4 * \qquad 82.1 \pm 6.5 \qquad 69.5 \pm 5.3 \qquad 56.6 \pm 4.0 *$	Number of bouts								
$68.3 \pm 3.2$ $63.2 \pm 4.5$ $46.0 \pm 5.7$ * $39.9 \pm 6.4$ * $82.1 \pm 6.5$ $69.5 \pm 5.3$ $56.6 \pm 4.0$ *	Dark period	$23.4 \pm 3.5$	$16.7 \pm 1.6$	$8.3\pm2.5$	$6.4\pm2.1$	$50.0\pm4.5$	$30.0\pm3.5*$	$21.8\pm1.2*$	$15.7 \pm 3.9*$
	Light period	$68.3 \pm 3.2$	$63.2 \pm 4.5$	$46.0 \pm 5.7*$	$39.9 \pm 6.4*$	$82.1\pm6.5$	$69.5 \pm 5.3$	$56.6 \pm 4.0 *$	$51.4 \pm 6.2*$

Total time spent in each state, mean duration, and number of bouts during the dark (active) and light (passive) periods. Data are shown as mean  $\pm$  SEM. \* p < 0.05 vs vehicle treatment of the same genotype (Bonferroni post-test)

NREM = non-rapid eye movement; REM = rapid eye movement



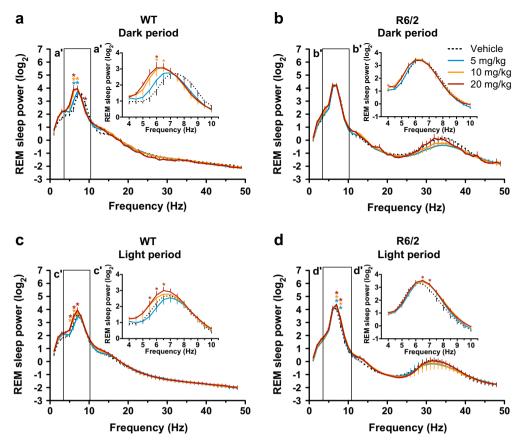
Fig. 1 Paroxetine decreases the probability of entering into rapid eye movement (REM) sleep from a non-REM (NREM) sleep episode. The probability of transitioning into REM sleep as a function of NREM sleep bout length as shown in (a, b) wildtype (WT) and (c-h) R6/2 mice during dark and light periods after (a-d) acute or (e, f) an 8-weeklong chronic treatment with vehicle (dashed line) or paroxetine (20 mg/kg i.p.; solid line), as well as (g, h) after a washout period of 2 weeks postchronic treatments. The absolute probability of transitioning from NREM into REM sleep for each 10-s epoch of NREM sleep was calculated and then the weighted average probability for bins of increasing duration (<60, 60–120, 120–180, 180-240, 240-300, 300-360, and > 360 s) was presented as group mean  $\pm$  SEM. The dark period is shown as shaded area. \*p < 0.05 vs vehicle treatment (Bonferroni post-test)



NREM sleep, was restricted to the first 12 h of treatment in both WT and R6/2 mice [drug × dark/light period interaction:  $F_{(3,33)} = 8.93$ ; p < 0.01 (Table 1)]. Paroxetine also consolidated NREM sleep by increasing the mean duration and reducing the number of NREM sleep bouts in both genotypes [drug effects:  $F_{(3,33)} = 23.62$  (p < 0.01) and  $F_{(3,33)} = 17.36$ 

(p < 0.01), respectively (Table 1)]. The paroxetine-induced changes in NREM sleep bout duration were restricted to the dark period in WT mice but continued into the light period in R6/2 mice [drug × dark/light period interaction:  $F_{(3,33)} = 5.57$ ; p < 0.01 (Table 1)]. In WT mice, paroxetine also decreased the amount of wakefulness, but this effect was limited to the first





**Fig. 2** Paroxetine reduces the frequency of rapid eye movement (REM) sleep theta oscillations in wild-type (WT) mice close to the one seen in R6/2 mice. Changes in relative power values of the electroencephalogram (EEG) spectra during REM sleep as shown in (a, c) WT and (b, d) R6/2 mice during the (a, b) dark and (c, d) light period after vehicle or paroxetine (5,

10, and 20 mg/kg i.p.) treatment. Enlarged images of relative EEG power values in the theta band (4–10 Hz) outlined by the box are shown in the insets. Data are shown as mean  $\pm$  SEM in (a–d) 1-Hz bins or (a'–d') at 0.5-Hz resolution. \*p < 0.05 vs vehicle treatment (Bonferroni post-test)

12 h after drug administration [drug × dark/light period interaction:  $F_{(3,33)} = 4.55$ ; p < 0.01 (Table 1)].

## Paroxetine Slows Down REM Sleep Theta Rhythm

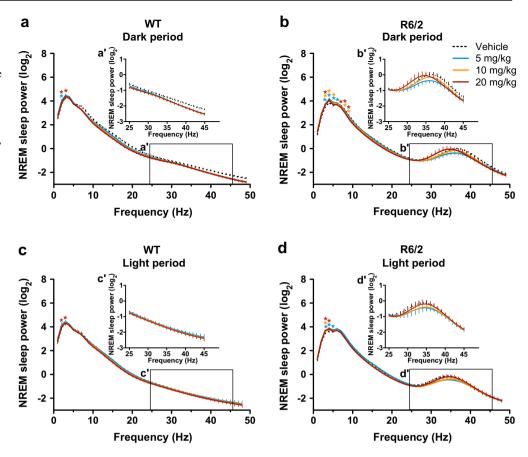
In WT mice, the acute effect of paroxetine treatment on REM sleep EEG spectra was restricted to the peak frequencies of the theta band (5–7 Hz) during both dark and light periods [drug × frequency interactions:  $F_{(144,576)} = 2.42 \ (p < 0.01)$  and  $F_{(144,720)} = 2.47$  (p < 0.01), respectively (Fig. 2a, c)]. Specifically, paroxetine shifted the peak frequency of REM sleep theta oscillation from 7 Hz to 6 Hz in WT mice during the dark period [drug × frequency interaction:  $F_{(36,144)} = 2.47$ ; p < 0.01(Fig. 2a')]. Interestingly, this is similar to the pathological theta peak frequency seen in symptomatic R6/2 mice (6-6.5 Hz in vehicle-treated mice; Fig. 2b'). The paroxetine-induced slow down of REM sleep theta rhythm persisted throughout the light period in WT mice [drug × frequency interaction:  $F_{(36.180)}$  = 1.85; p < 0.01 (Fig. 2c')]. In R6/2 mice, however, paroxetine had no further effect on the already-slowed EEG theta rhythm seen during the dark period (Fig. 2b, b'). Interestingly, paroxetine improved REM sleep EEG spectra in R6/2 mice during the second 12 h [drug effect:  $F_{(3.18)} = 6.09$ ; p < 0.01 (Fig. 2d)]. The rhythm of REM sleep theta oscillation became slightly faster during the light period in R6/2 mice after paroxetine treatment than after vehicle treatment [6.5 Hz vs 6.0 Hz; drug × frequency interaction:  $F_{(36,216)} = 1.79$ ; p < 0.01 (Fig. 2d')]. Furthermore, paroxetine increased the power of slow oscillations (2–6 Hz) in the NREM sleep EEG of WT mice during both dark and light periods [drug × frequency interactions:  $F_{(144.720)} = 1.53$ (p < 0.01) and  $F_{(144,720)} = 1.27$  (p < 0.05), respectively (Fig. 3a, c)]. A similar increase in EEG slow oscillations was seen in R6/2 mice during NREM sleep after acute paroxetine treatment during both dark and light periods [drug × frequency interactions:  $F_{(144,864)} = 4.03$  (p < 0.01) and  $F_{(144.864)} = 2.88 \ (p < 0.01)$ , respectively (Fig. 3b, d)]. Acute paroxetine treatment, however, had no effect on the abnormal low-gamma EEG oscillations seen in R6/2 mice (Fig. 3b', d').

# **Chronic Treatment With Paroxetine Normalizes REM Sleep in HD Mice**

In both acute and chronic studies, vehicle-treated R6/2 mice had an increased propensity for REM sleep during the dark



Fig. 3 Acute paroxetine treatment had no effect on the abnormal low-gamma electroencephalogram (EEG) oscillations in R6/2 mice. Changes in relative power values of EEG spectra during non-rapid eye movement (NREM) sleep as shown in (a, c) wild-type (WT) and (b, d) R6/2 mice during the (a, b) dark and (c, d) light period after vehicle or paroxetine (5, 10, and 20 mg/ kg i.p.) treatment. Enlarged images of relative EEG power values in the low-gamma band (25–45 Hz) outlined by the box are shown in the insets. Data are shown as mean  $\pm$  SEM in 1-Hz bins. \*p < 0.05 vs vehicle treatment (Bonferroni post-test)



period compared to that of WT mice receiving acute vehicle treatment. This is shown by the doubled amount of REM sleep, increased number of REM sleep bouts, and an increased probability of entering REM sleep from NREM sleep (Tables 1 and 2; Fig. 1). By contrast, the propensity for REM sleep was normalized in R6/2 mice treated chronically with paroxetine (20 mg/kg/day i.p.). This is shown by the decreased amount of REM sleep during the dark period seen in R6/2 mice that was significant after 7 weeks of paroxetine treatment [drug effect:  $F_{(1,9)} = 5.30$ ; p < 0.05 (see Fig. 4a)]. Eight weeks of treatment with paroxetine resulted in a 42% decrease in REM sleep amount and a 60% decrease in the number of REM sleep bouts in R6/2 mice during the dark period [drug × dark/light period interactions:  $F_{(1.10)} = 8.47$ (p < 0.05) and  $F_{(1,10)} = 8.50$  (p < 0.05), respectively (Table 2)]. Furthermore, the probability of entering REM sleep in the dark period gradually increased across the duration of NREM sleep in vehicle-treated R6/2 mice. By contrast, R6/2 mice treated with paroxetine had a much lower probability of entering REM sleep at any time during a NREM sleep episode if that was longer than 240 s [drug × NREM sleep bout duration interaction:  $F_{(6,60)} = 5.69$ ; p < 0.01 (Fig. 1e)]. Interestingly, chronic treatment with paroxetine corrected REM sleep abnormalities in R6/2 mice during the dark period but had little effect on REM sleep parameters during the light period (Table 2; Fig. 1f). Only the mean duration of REM sleep bouts changed during both dark and light periods in paroxetine-treated R6/2 mice. REM sleep bouts became longer [drug effect:  $F_{(1,10)}$  = 19.54; p < 0.01 (Table 2)], suggesting that R6/2 mice had a better consolidated REM sleep after chronic paroxetine treatment than after vehicle treatment. Paroxetine did not change any other sleep—wake parameters measured (Table 2). Furthermore, none of the changes in REM sleep parameters induced by paroxetine persisted after treatment stopped (Table 2; Fig. 1g, h, and Fig. 4a).

# **Chronic Paroxetine Treatment Prevents EEG Abnormalities in HD Mice**

At 14 weeks of age, the peak frequency of REM sleep theta oscillations was 6.5 Hz in vehicle-treated R6/2 mice during both dark and light periods (Fig. 5a', c'). That is 0.5 to 1 Hz slower than the peak frequency of the REM sleep theta oscillation (7–7.5 Hz) seen in WT mice after acute vehicle treatment (Fig. 2a', c'). Paroxetine normalized REM sleep EEG spectra in chronically treated R6/2 mice during both dark and light periods [drug effects:  $F_{(1,10)} = 23.36 \ (p < 0.01)$  and  $F_{(1,10)} = 17.68 \ (p < 0.01)$ , respectively (Fig. 5a, c)]. In R6/2 mice, REM sleep theta rhythm became more robust (with an increase in power between 7 and 11 Hz) after paroxetine



**Table 2** Vigilance state parameters in R6/2 mice after chronic treatment with vehicle or paroxetine

Recording	8 weeks of treatment		2 weeks of washout	
Treatment	Vehicle $(n = 6)$	Paroxetine $(n = 5)$	Vehicle $(n = 6)$	Paroxetine $(n = 6)$
Wake				
Total time (min)				
Dark period	$277.9 \pm 7.1$	$271.0 \pm 25.2$	$312.1 \pm 24.5$	$308.8\pm22.5$
Light period	$227.3\pm15.1$	$219.9 \pm 16.3$	$234.2\pm49.6$	$297.6 \pm 38.5$
Mean duration (mi	in)			
Dark period	$3.3 \pm 0.4$	$3.4\pm0.7$	$4.2\pm0.6$	$3.9\pm0.5$
Light period	$2.9\pm0.3$	$3.0\pm0.4$	$3.0\pm0.6$	$3.3\pm0.5$
Number of bouts				
Dark period	$90.5\pm10.5$	$91.5 \pm 14.3$	$79.2 \pm 7.8$	$82.8 \pm 9.1$
Light period	$83.0 \pm 9.8$	$77.8 \pm 6.4$	$77.8 \pm 7.0$	$94.0\pm10.0$
NREM sleep				
Total time (min)				
Dark period	$369.4 \pm 11.3$	$407.1\pm26.1$	$336.3 \pm 19.9$	$354.7\pm25.2$
Light period	$397.9 \pm 13.1$	$409.7 \pm 13.7$	$396.5 \pm 40.1$	$351.9 \pm 30.1$
Mean duration (mi	in)			
Dark period	$3.3 \pm 0.2$	$4.4\pm0.4$	$3.3\pm0.2$	$3.7 \pm 0.6$
Light period	$3.4\pm0.3$	$4.4\pm0.3$	$3.6 \pm 0.3$	$3.1\pm0.4$
Number of bouts				
Dark period	$114.2 \pm 6.9$	$97.2 \pm 13.9$	$105.2\pm10.1$	$106.0\pm12.6$
Light period	$118.8 \pm 8.3$	$95.2 \pm 9.7$	$110.0 \pm 7.8$	$118.8 \pm 13.5$
REM sleep				
Total time (min)				
Dark period	$72.7 \pm 8.9$	$41.9 \pm 6.5*$	$71.6 \pm 8.9$	$56.6 \pm 8.1$
Light period	$94.8 \pm 6.7$	$90.4 \pm 3.1$	$89.2 \pm 12.8$	$70.6 \pm 13.8$
Mean duration (mi	in)			
Dark period	$1.5\pm0.0$	$2.3 \pm 0.2^*$	$1.5\pm0.2$	$1.4\pm0.1$
Light period	$1.5\pm0.1$	$2.1\pm0.2^*$	$1.5\pm0.1$	$1.4\pm0.1$
Number of bouts				
Dark period	$48.0 \pm 5.8$	$19.0 \pm 3.6 *$	$50.0\pm7.8$	$42.5 \pm 7.5$
Light period	$63.2 \pm 4.8$	$44.8 \pm 5.4$	$60.6 \pm 9.4$	49.5 ± 9.1

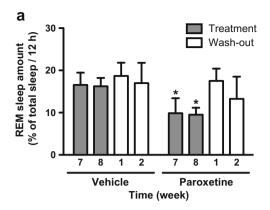
Total time spent in each state, mean duration, and number of bouts during the dark (active) and light (passive) periods after 8 weeks of treatment with vehicle or paroxetine (20 mg/kg/day i.p.) or after a 2-week washout period. Data are shown as mean  $\pm$  SEM. \* $p < 0.05 \ vs$  vehicle-treated group of the same recording day (Bonferroni posttest)

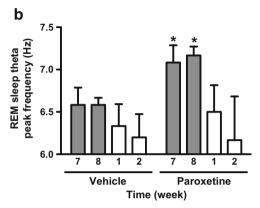
treatment than after vehicle treatment during both dark and light periods [drug effects:  $F_{(1,10)} = 5.50 \ (p < 0.05)$  and  $F_{(1,10)} = 5.66 \ (p < 0.05)$ , respectively (Fig. 5b, d)]. REM sleep theta rhythm also became faster (with a peak at 7 Hz) in R6/2 mice after paroxetine treatment than after vehicle treatment during both the dark and light periods [drug × frequency interactions:  $F_{(12,120)} = 8.80 \ (\underline{p} < 0.01)$  and  $F_{(12,120)} = 7.47 \ (p < 0.01)$ , respectively (Fig. 5a', c')]. REM sleep theta rhythm was already faster in paroxetine-treated than in vehicle-treated R6/2 mice after 7 weeks of treatment [time × drug interaction:  $F_{(3,27)} = 3.70$ ;  $p < 0.05 \ (Fig. 4b)$ ]. Two weeks after the treatment was finished, the drug-induced differences in theta rhythm between the groups were reduced (Fig. 4b and

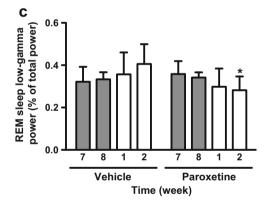
Fig. 5e–h). Specifically, the rhythm of REM sleep theta was still slightly faster in paroxetine-treated R6/2 mice than in vehicle-treated mice during the light period [6.5 vs 6 Hz; drug × frequency interaction:  $F_{(12,108)} = 1.90$ ; p < 0.05 (Fig. 5g')] but not during the dark period 2 weeks after treatment stopped (Fig. 5e' and Fig. 4b).

As the disease progressed, an abnormal low-gamma EEG activity emerged in vehicle-treated but not in paroxetine-treated R6/2 mice during both REM and NREM sleep [time  $\times$  drug interaction:  $F_{(3,27)} = 9.58$  (p < 0.01); drug effect:  $F_{(1,9)} = 7.78$  (p < 0.05), respectively (Fig. 4c, d)]. After 8 weeks of paroxetine treatment, R6/2 mice had significantly fewer abnormal low-gamma oscillations in their NREM sleep EEG









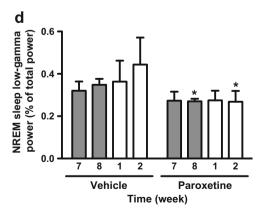


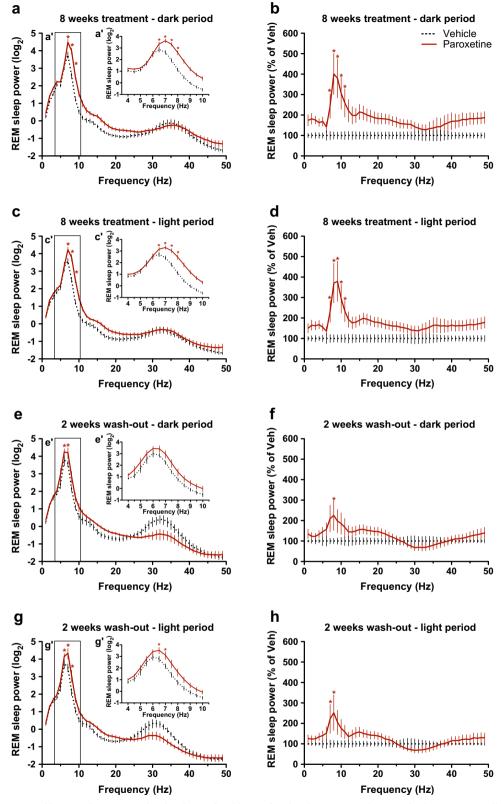
Fig. 4 Chronic paroxetine treatment normalizes rapid eye movement (REM) sleep, as well as theta and low-gamma electroencephalogram (EEG) oscillations in R6/2 mice. Changes in (a) the amount of REM sleep, (b) REM sleep theta peak frequency, (c) REM sleep low-gamma power (25–45 Hz), and (d) non-REM (NREM) sleep low-gamma power is shown in vehicle- and paroxetine (20 mg/kg/day)-treated R6/2 mice during the dark period after 7 and 8 weeks of treatment (closed bars), as well as 1 and 2 weeks after treatment stopped ("washout", open bars). Theta peak frequency was defined as the frequency value (0.5-Hz resolution) within the theta range (4–10 Hz) with the highest EEG power value. Data are shown as mean ± SEM. \*p < 0.05 vs vehicle treatment at the corresponding time (unpaired t-test)</p>

than vehicle-treated mice during both dark and light periods [drug × frequency interactions:  $F_{(20.200)} = 2.72$  (p < 0.01) and  $F_{(20,200)} = 4.01 \ (p < 0.01)$ , respectively (Fig. 6a', c')]. The suppression of low-gamma EEG activity during NREM sleep persisted for at least 2 weeks after treatment stopped in R6/2 mice [time × drug interaction:  $F_{(3,27)} = 6.10$ ; p < 0.01(Fig. 4d)]. Specifically, as the disease progressed, vehicletreated R6/2 mice developed an abnormally increased lowgamma EEG activity during NREM sleep (with a peak frequency at 33–35 Hz) during both dark and light periods that was 2-fold higher than the low-gamma activity seen in paroxetine-treated R6/2 mice [drug × frequency interactions:  $F_{(20,180)} = 4.76 \ (p < 0.01)$  and  $F_{(20,180)} = 6.50 \ (p < 0.01)$ , respectively (Fig. 6e', g')]. Chronic paroxetine treatment also prevented the development of abnormal low-gamma EEG oscillations in R6/2 mice during REM sleep, an effect that persisted for at least 2 weeks after treatment stopped, during both dark and light periods [drug × frequency interactions:  $F_{(20,180)} = 2.60 \ (p < 0.01)$  and  $F_{(20,180)} = 2.55 \ (p < 0.01)$ , respectively (Fig. 5e, g)].

## **Discussion**

We show that chronic treatment with paroxetine, when started at a presymptomatic stage of disease, prevented sleep and EEG abnormalities in the R6/2 mouse model of HD. Control R6/2 mice, treated with vehicle, exhibited an abnormally increased REM sleep amount (particularly during the dark, when they are normally active), slowed REM sleep theta rhythm, and abnormal low-gamma oscillations in their sleep EEG. None of these abnormalities could be seen in R6/2 mice treated chronically with paroxetine, during either dark or light period. Two weeks after treatment stopped, the beneficial effect of paroxetine on REM sleep and REM sleep theta rhythm had largely disappeared. On the one hand, suppression of abnormal EEG gamma activity persisted in the R6/2 mice for at least 2 weeks after the last paroxetine treatment. On the other hand, acute treatment with paroxetine normalized the abnormal increase in REM sleep seen during the active period and consolidated NREM sleep but had no effect on abnormal theta

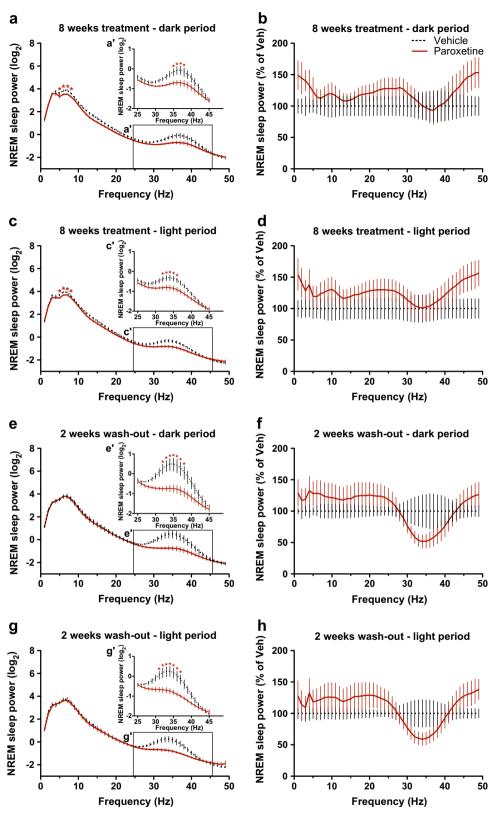




**Fig. 5** Chronic treatment with paroxetine prevents the slowdown of rapid eye movement (REM) sleep theta rhythm in R6/2 mice. Changes in relative power values of electroencephalogram (EEG) spectra during REM sleep as shown in R6/2 mice during the (a, b, e, f) dark and (c, d, g, h) light period after (a–d) an 8-week-long chronic vehicle (dashed line) or paroxetine (20 mg/kg/day i.p.; solid line) treatment, as well as (e–h) after a washout period of 2 weeks

after chronic treatment. The spectral values were normalized to (a, c, e, g) the total power of the studied EEG spectrum or to (b, d, f, h) the mean power spectral values of vehicle-treated mice. Enlarged images of relative EEG power values in the theta band (4–10 Hz) outlined by the box are shown in the insets. Data are shown as mean  $\pm$  SEM in (a, c, e, g) 1-Hz bins or at (a', c', e', g') 0.5 Hz resolution. \*p < 0.05 vs vehicle treatment (Bonferroni post-test)





**Fig. 6** Chronic paroxetine treatment prevents abnormal low gamma electroencephalogram (EEG) oscillation in R6/2 mice. Changes in relative power values of EEG spectra during non-rapid eye movement (NREM) sleep are shown in R6/2 mice during the dark (a, b, e, f) and light (c, d,g, h) period after an 8-week-long vehicle (dashed line) or paroxetine (20 mg/kg/day i.p.; solid line) treatment (a–d), as well as after a washout period after the chronic

treatment ended of 2 weeks duration (e–h). The spectral values were normalized to the total power (a, c, e, g) of the studied EEG spectrum or to the mean power (b, d, f, h) spectral values of vehicle treated mice. Enlarged images of relative EEG power values in the low-gamma band (25–45Hz) outlined by the box are shown in the insets. Data are shown as mean  $\pm$  SEM in 1-Hz bins. \*p < 0.05 vs vehicle treatment (Bonferroni post-hoc test).



or gamma oscillations in R6/2 mice. In WT mice, acute paroxetine treatment slowed REM sleep theta rhythm to the level seen in symptomatic R6/2 mice, suggesting that 5-hydroxytryptamine (serotonin; 5-HT) neurotransmission is already altered in HD mice. Our data show that paroxetine corrects both disrupted sleep and abnormal brain oscillations in HD mice when prophylactic treatment is initiated before the onset of symptoms.

Patients with HD show disturbed sleep and abnormal brain oscillations early in the disease process, with decreased REM sleep time being one of the most consistent findings [2, 5, 6, 8]. Mice are nocturnal, and HD mice, including the R6/2 mice studied here, already show an abnormal increase in REM sleep amount during the night at presymptomatic stage of the disease [9–13]. REM sleep is generated primarily by brain stem neurons [29, 30]. 5-HT inhibits REM sleep [31], and paroxetine treatment increases the extracellular level of 5-HT [32]. In our study, acute treatment with paroxetine suppressed REM sleep in both WT and R6/2 mice. We previously obtained similar results using the 5-HT/noradrenaline reuptake inhibitor amitriptyline [10]. The REM sleep-suppressing effect of antidepressants is well documented [25]. However, in humans most antidepressants (including paroxetine) suppress REM sleep early in the treatment, and this effect gradually diminishes after repeated administration of the drug [25]. In R6/2 mice, however, paroxetine prevented the abnormal increase in REM sleep amount for at least 8 weeks during treatment. Whether this is a consequence of an enhanced 5-HT neurotransmission or is due to changes in REM sleep-controlling circuits needs further investigation, particularly because drugs may have markedly different effects on the neurologically normal and HD brain.

In patients with HD, EEG theta oscillations are slowed during quiet wakefulness [33], and have decreased power during REM sleep [2, 7]. Symptomatic R6/2 mice in our study had slowed EEG theta oscillations during REM sleep, which is in accord with previous findings in R6/2 [9, 11], R6/1 [14, 15], and Q175 [13] mice. EEG theta oscillations in rodents are thought to have a hippocampal origin [34, 35], and 5-HT neurons play a key role in modulating hippocampal theta oscillations [36]. Here in WT mice, acute treatment with paroxetine reduced the frequency of REM sleep theta rhythm to a level seen in symptomatic R6/2 mice. A similar decrease in reticular-elicited theta frequency has been shown in Sprague-Dawley rats after SSRI fluoxetine treatment [37]. However, paroxetine had no further effect on the already slowed REM sleep theta rhythm in R6/2 mice, suggesting an already altered 5-HT neurotransmission in these mice. Increased signaling through 5-HT<sub>6</sub> receptors could account for the abnormal theta rhythm seen in HD mice as it has been shown that activation of these receptors decreases hippocampal theta frequency [38]. Although there is no direct evidence of an altered 5-HT signaling in HD, the diminishing of depressive symptoms in patients with late-stage HD [1] and the decrease in 5-HT-

dependent behavioral despair in symptomatic R6/2 mice [39] both suggest compensatory changes in brain 5-HT neurotransmission. It should be noted, however, that alterations in neurotransmitter systems other than 5-HT may also underlie the changes we see.

Abnormal EEG gamma oscillations have been reported in several neuropsychiatric disorders, including schizophrenia [40], depression [41], and Alzheimer's disease [42]. Abnormal EEG gamma oscillations have been found in both patients with early-HD and in presymptomatic R6/2 mice [2, 9–11]. Although the functional significance of abnormal highfrequency EEG oscillations in these neuropsychiatric disorders is unknown, gamma oscillations have attracted a lot of attention in recent years because of the role they play in feature binding [43], object representation [44], and selective attention [45]. Gamma oscillations are generated in the cortex and hippocampus [46, 47]. Since basal ganglia is severely affected in HD, insufficient inhibition of cortical activity by cortical projecting neurons of basal ganglia could account, at least in part, for the abnormal low-gamma EEG oscillations seen in HD [10]. Both cortex and hippocampus express high levels of brain-derived neurotrophic factor (BDNF); a protein that promotes the maintenance and survival of neurons mainly through its tyrosine kinase (Trk) B receptor [48]. BDNF modulates gamma oscillations [49], and BDNF-TrkB signaling is impaired in HD [50]. Chronic (but not acute) treatment with the SSRI paroxetine or fluoxetine, increases BDNF expression and improves disease symptoms in HD mice [27, 28, 51, 52]. Here, chronic treatment with paroxetine suppressed abnormal low-gamma EEG oscillations in R6/2 mice. This is similar to the suppression of EEG gamma activity seen in R6/2 mice after acute amitriptyline treatment [10]. Interestingly, amitriptyline acts on TrkA and TrkB receptors as agonist and has potent neurotrophic activity [53]. Thus, we hypothesize that the correction of abnormal low-gamma EEG oscillations in R6/2 mice by chronic paroxetine treatment may be due, at least in part, to an increase in brain neurotrophic activity. Although instrumental REM sleep deprivation also increases BDNF expression in rodents [54, 55], it needs to be further investigated with regard to whether the paroxetine-induced suppression of REM sleep results in increased neurotrophic activity in the HD mouse brain.

Our data show that paroxetine treatment, when initiated before the onset of symptoms, corrects both REM sleep disturbances and abnormal brain oscillations in HD mice. This suggests a possible mechanistic link between early disruption of REM sleep and abnormal brain oscillations in HD mice. Since abnormal sleep and EEG changes are likely to be correlates of altered brain function in HD, correcting these abnormalities might also be reflected in improvements in HD symptoms other than sleep and sleep-dependent brain oscillations.



**Acknowledgments** We thank Drs Zhiguang Zheng, Koliane Ouk, and Szilvia Vas, and Ms Juliet Aungier, for their technical assistance. This work was supported by a grant from CHDI Foundation, Inc.

**Open Access** This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

#### References

- Paulsen JS, Nehl C, Hoth KF, et al. Depression and stages of Huntington's disease. J Neuropsychiatry Clin Neurosci 2005;17: 496-502.
- Lazar AS, Panin F, Goodman AO, et al. Sleep deficits but no metabolic deficits in premanifest Huntington's disease. Ann Neurol 2015;78:630-648.
- Aziz NA, Anguelova GV, Marinus J, Lammers GJ, Roos RA. Sleep and circadian rhythm alterations correlate with depression and cognitive impairment in Huntington's disease. Parkinsonism Relat Disord 2010;16:345-350.
- Goodman AO, Rogers L, Pilsworth S, et al. Asymptomatic sleep abnormalities are a common early feature in patients with Huntington's disease. Curr Neurol Neurosci Rep 2011;11:211-217.
- Arnulf I, Nielsen J, Lohmann E, et al. Rapid eye movement sleep disturbances in Huntington disease. Arch Neurol 2008;65:482-488.
- Piano C, Losurdo A, Della Marca G, et al. Polysomnographic findings and clinical correlates in Huntington disease: a cross-sectional cohort study. Sleep 2015;38:1489-1495.
- 7. Piano C, Mazzucchi E, Bentivoglio AR, et al. Wake and sleep EEG in patients with Huntington Disease: an eLORETA study and review of the literature. Clin EEG Neurosci 2017;48:60-71.
- Silvestri R, Raffaele M, De Domenico P, et al. Sleep features in Tourette's syndrome, neuroacanthocytosis and Huntington's chorea. Neurophysiol Clin 1995;25:66-77.
- Kantor S, Szabo L, Varga J, Cuesta M, Morton AJ. Progressive sleep and electroencephalogram changes in mice carrying the Huntington's disease mutation. Brain 2013;136:2147-2158.
- Kantor S, Varga J, Morton AJ. A single dose of hypnotic corrects sleep and EEG abnormalities in symptomatic Huntington's disease mice. Neuropharmacology 2016;105:298-307.
- Fisher SP, Black SW, Schwartz MD, et al. Longitudinal analysis of the electroencephalogram and sleep phenotype in the R6/2 mouse model of Huntington's disease. Brain 2013;136:2159-2172.
- Lebreton F, Cayzac S, Pietropaolo S, Jeantet Y, Cho YH. Sleep physiology alterations precede plethoric phenotypic changes in R6/1 Huntington's disease mice. PLOS ONE 2015;10: e0126972.
- Fisher SP, Schwartz MD, Wurts-Black S, et al. Quantitative electroencephalographic analysis provides an early-stage indicator of disease onset and progression in the zQ175 knock-in mouse model of Huntington's disease. Sleep 2016;39:379-391.
- Pignatelli M, Lebreton F, Cho YH, Leinekugel X. "Ectopic" theta oscillations and interictal activity during slow-wave state in the R6/ 1 mouse model of Huntington's disease. Neurobiol Dis 2012;48: 409-417.
- Jeantet Y, Cayzac S, Cho YH. Beta oscillation during slow wave sleep and rapid eye movement sleep in the electroencephalogram of a transgenic mouse model of Huntington's disease. PLOS ONE 2013;8:e79509.

- Hong SL, Cossyleon D, Hussain WA, Walker LJ, Barton SJ, Rebec GV. Dysfunctional behavioral modulation of corticostriatal communication in the R6/2 mouse model of Huntington's disease. PLOS ONE 2012;7:e47026.
- Callahan JW, Abercrombie ED. Relationship between subthalamic nucleus neuronal activity and electrocorticogram is altered in the R6/2 mouse model of Huntington's disease. J Physiol 2015;593: 3727-3738.
- Loh DH, Kudo T, Truong D, Wu Y, Colwell CS. The Q175 mouse model of Huntington's disease shows gene dosage- and age-related decline in circadian rhythms of activity and sleep. PLOS ONE 2013;8:e69993.
- Nagy D, Tingley FD, 3rd, Stoiljkovic M, Hajos M. Application of neurophysiological biomarkers for Huntington's disease: evaluating a phosphodiesterase 9A inhibitor. Exp Neurol 2015;263:122-131.
- Lopez J, Roffwarg HP, Dreher A, Bissette G, Karolewicz B, Shaffery JP. Rapid eye movement sleep deprivation decreases long-term potentiation stability and affects some glutamatergic signaling proteins during hippocampal development. Neuroscience 2008;153:44-53.
- Shaffery JP, Lopez J, Roffwarg HP. Brain-derived neurotrophic factor (BDNF) reverses the effects of rapid eye movement sleep deprivation (REMSD) on developmentally regulated, long-term potentiation (LTP) in visual cortex slices. Neurosci Lett 2012;513:84-88.
- 22. Shaffery JP, Sinton CM, Bissette G, Roffwarg HP, Marks GA. Rapid eye movement sleep deprivation modifies expression of long-term potentiation in visual cortex of immature rats. Neuroscience 2002;110:431-443.
- Ross CA, Tabrizi SJ. Huntington's disease: from molecular pathogenesis to clinical treatment. Lancet Neurol 2011;10:83-98.
- Adam OR, Jankovic J. Symptomatic treatment of Huntington disease. Neurotherapeutics 2008;5:181-197.
- Wilson S, Argyropoulos S. Antidepressants and sleep: a qualitative review of the literature. Drugs 2005;65:927-947.
- Smith MI, Piper DC, Duxon MS, Upton N. Effect of SB-243213, a selective 5-HT<sub>(2C)</sub> receptor antagonist, on the rat sleep profile: a comparison to paroxetine. Pharmacol Biochem Behav 2002;71: 599-605.
- Duan W, Guo Z, Jiang H, et al. Paroxetine retards disease onset and progression in Huntingtin mutant mice. Ann Neurol 2004;55:590-594.
- Sillaber I, Panhuysen M, Henniger MS, et al. Profiling of behavioral changes and hippocampal gene expression in mice chronically treated with the SSRI paroxetine. Psychopharmacology (Berl) 2008;200:557-572.
- Brown RE, Basheer R, McKenna JT, Strecker RE, McCarley RW. Control of sleep and wakefulness. Physiol Rev 2012;92:1087-1187.
- Weber F, Chung S, Beier KT, Xu M, Luo L, Dan Y. Control of REM sleep by ventral medulla GABAergic neurons. Nature 2015;526: 435-438.
- Monti JM. Serotonin control of sleep-wake behavior. Sleep Med Rev 2011;15:269-281.
- David DJ, Bourin M, Jego G, Przybylski C, Jolliet P, Gardier AM. Effects of acute treatment with paroxetine, citalopram and venlafaxine in vivo on noradrenaline and serotonin outflow: a microdialysis study in Swiss mice. Br J Pharmacol 2003;140:1128-1136.
- Bylsma FW, Peyser CE, Folstein SE, Folstein MF, Ross C, Brandt J. EEG power spectra in Huntington's disease: clinical and neuropsychological correlates. Neuropsychologia 1994;32:137-150.
- Gerbrandt LK, Lawrence JC, Eckardt MJ, Lloyd RL. Origin of the neocortically monitored theta rhythm in the curarized rat. Electroencephalogr Clin Neurophysiol 1978;45:454-467.



- 35. Kantor S, Jakus R, Molnar E, et al. Despite similar anxiolytic potential, the 5-hydroxytryptamine 2C receptor antagonist SB-242084 [6-chloro-5-methyl-1-[2-(2-methylpyrid-3-yloxy)-pyrid-5-yl carbamoyl] indoline] and chlordiazepoxide produced differential effects on electroencephalogram power spectra. J Pharmacol Exp Ther 2005;315:921-930.
- Jackson J, Dickson CT, Bland BH. Median raphe stimulation disrupts hippocampal theta via rapid inhibition and state-dependent phase reset of theta-related neural circuitry. J Neurophysiol 2008:99:3009-3026
- Munn RG, McNaughton N. Effects of fluoxetine on hippocampal rhythmic slow activity and behavioural inhibition. Behav Pharmacol 2008;19:257-264.
- Ly S, Pishdari B, Lok LL, Hajos M, Kocsis B. Activation of 5-HT<sub>6</sub> receptors modulates sleep-wake activity and hippocampal theta oscillation. ACS Chem Neurosci 2013;4:191-199.
- Ciamei A, Detloff PJ, Morton AJ. Progression of behavioural despair in R6/2 and Hdh knock-in mouse models recapitulates depression in Huntington's disease. Behav Brain Res 2015;291:140-146.
- Farzan F, Barr MS, Levinson AJ, et al. Evidence for gamma inhibition deficits in the dorsolateral prefrontal cortex of patients with schizophrenia. Brain 2010;133:1505-1514.
- Tekell JL, Hoffmann R, Hendrickse W, Greene RW, Rush AJ, Armitage R. High frequency EEG activity during sleep: characteristics in schizophrenia and depression. Clin EEG Neurosci 2005;36: 25-35.
- van Deursen JA, Vuurman EF, Verhey FR, van Kranen-Mastenbroek VH, Riedel WJ. Increased EEG gamma band activity in Alzheimer's disease and mild cognitive impairment. J Neural Transm (Vienna) 2008;115:1301-1311.
- Engel AK, Fries P, Singer W. Dynamic predictions: oscillations and synchrony in top-down processing. Nat Rev Neurosci 2001;2:704-716.
- Gruber T, Muller MM, Keil A. Modulation of induced gamma band responses in a perceptual learning task in the human EEG. J Cogn Neurosci 2002;14:732-744.

- Fell J, Fernandez G, Klaver P, Elger CE, Fries P. Is synchronized neuronal gamma activity relevant for selective attention? Brain Res Brain Res Rev 2003;42:265-272.
- 46. Dine J, Genewsky A, Hladky F, et al. Local optogenetic induction of fast (20-40 Hz) pyramidal-interneuron network oscillations in the in vitro and in vivo CA1 hippocampus: modulation by CRF and enforcement of perirhinal theta activity. Front Cell Neurosci 2016;10:108.
- Middleton SJ, Racca C, Cunningham MO, et al. High-frequency network oscillations in cerebellar cortex. Neuron 2008;58:763-774
- Conner JM, Lauterborn JC, Yan Q, Gall CM, Varon S. Distribution of brain-derived neurotrophic factor (BDNF) protein and mRNA in the normal adult rat CNS: evidence for anterograde axonal transport. J Neurosci 1997;17:2295-2313.
- Huang Y, Morozov A. Hippocampal deletion of BDNF gene attenuates gamma oscillations in area CA1 by up-regulating 5-HT<sub>3</sub> receptor. PLOS ONE 2011;6:e16480.
- Zuccato C, Cattaneo E. Role of brain-derived neurotrophic factor in Huntington's disease. Prog Neurobiol 2007;81:294-330.
- Martinez-Turrillas R, Del Rio J, Frechilla D. Sequential changes in BDNF mRNA expression and synaptic levels of AMPA receptor subunits in rat hippocampus after chronic antidepressant treatment. Neuropharmacology 2005;49:1178-1188.
- Grote HE, Bull ND, Howard ML, et al. Cognitive disorders and neurogenesis deficits in Huntington's disease mice are rescued by fluoxetine. Eur J Neurosci 2005;22:2081-2088.
- Jang SW, Liu X, Chan CB, et al. Amitriptyline is a TrkA and TrkB receptor agonist that promotes TrkA/TrkB heterodimerization and has potent neurotrophic activity. Chem Biol 2009;16:644-656.
- Datta S, Knapp CM, Koul-Tiwari R, Barnes A. The homeostatic regulation of REM sleep: a role for localized expression of brainderived neurotrophic factor in the brainstem. Behav Brain Res 2015;292:381-392.
- Ravassard P, Hamieh AM, Malleret G, Salin PA. Paradoxical sleep: a vigilance state to gate long-term brain plasticity? Neurobiol Learn Mem 2015;122:4-10.

