

Ileal amino acids digestibility of sorghum in weaned piglets and growing pigs

G. Mariscal-Landín^{1,2†}, T. C. Reis de Souza³ and M. A. Avalos⁴

¹Centro Nacional de Investigación en Fisiología Animal – Instituto Nacional de Investigaciones Forestales Agrícolas y Pecuarias, México, Ajuchitlán Colón, 76280 Querétaro, México, USA; ²Facultad de Estudios Superiores Cuautitlán-UNAM, Campo 1-Edificio de Estudios de Posgrado, Av. 1 de Mayo s/n, Cuautitlán Izcalli, 54700 Edo, de México, México, USA; ³Facultad de Ciencias Naturales, Universidad Autónoma de Querétaro, México, Avenida de las Ciencias s/n, Colonia Juriquilla, Delegación Santa Rosa Jáuregui, 76230 Querétaro, México, USA; ⁴Master of Science Thesis, Facultad de Estudios Superiores Cuautitlán, Universidad Nacional Autónoma de México, USA

(Received 4 April 2009; Accepted 24 January 2010; First published online 4 March 2010)

The objective of the study was to determine the coefficients of ileal apparent digestibility (CIAD) of sorghum protein and amino acids (AA) in weaned piglets and growing pigs. Digestibility coefficients were estimated using the regression and difference methods for the weaned piglets; and the direct and difference methods for the growing pigs. To test the hypothesis that CP and AA digestibility of sorghum is lower in weaned piglets than in growing pigs, two experiments were conducted. In experiment one, 20 weaned piglets were fitted with a 'T' cannula at 21 days of age and were fed for 2 weeks one of five dietary treatments: a reference or control diet providing 200 g of CP/kg from casein (C) as the sole protein source, and four casein–sorghum (C–S) diets kept isoproteic to C by the appropriate adjustment of C and maize starch proportions; the amount of sorghum (S) in these diets was 135, 307, 460 and 614 g/kg. In experiment 2, fifteen castrated pigs weighing 57.8 ± 2.8 kg were used and randomly allotted to one of three dietary treatments: a reference casein–maize starch diet containing C as the sole protein source, a C–S diet, both diets containing 160 g of CP/kg, and a fortified S diet containing 68 g of CP/kg. In piglets the CIAD for CP and AA decreased linearly ($P < 0.05$) as the amount of S in the diet increased. The average ileal digestibility of AA from C was 0.858 ± 0.111 , and decreased to 0.663 ± 0.191 at the higher S level. The CIAD estimated using the regression or difference methods were similar for leucine, cysteine, glutamic acid, serine, alanine and tyrosine, and different for the other AA. In growing pigs the CIAD of protein and AA (except alanine and cysteine) were similar ($P > 0.05$) for the C and the C–S diets, but higher ($P < 0.05$) than those for the S diet. The CIAD for S obtained by the difference method were higher ($P < 0.05$) than those obtained using the direct method, except for lysine, isoleucine, valine, methionine, threonine and cysteine. The results indicate that except for lysine and cysteine, growing pigs' ability to digest AA and protein is superior than weaned piglets.

Keywords: amino acids, ileal digestibility, piglets, protein, sorghum

Implications

This study shows that the digestibility of sorghum (S) is lower in piglets than in growing pigs. The difference is greater for protein than for independent amino acids (AA) thus accuracy in dietary formulation and estimation of AA requirements for pigs at weaning is in need of critical improvement if the appropriate digestibility figures are to be used. The lower digestibility of S in piglets is possibly due to an exacerbated response of the gut of the piglets to the deleterious effect of tannins. For this reason S is not recommended in diets for starter pigs, particularly if it is rich in tannins.

Introduction

Sorghum (S) ranks fifth among the cereals produced worldwide (Bansal *et al.*, 2008). A characteristic of S is its huge variability in chemical composition and consequently in its nutritive value (Mossé *et al.*, 1988). Ramírez *et al.* (2005) analyzed 216 samples of S and obtained a wide range for the CP content (5.2% to 11.0%), NDF (6.3% to 22.9%) and tannins (0.02% to 5.06%). Tannins affect in a negative way the digestibility of nutrients in S (Brand *et al.*, 1990; Jondreville *et al.*, 2001; Mariscal-Landín *et al.*, 2004) and the efficiency with which pigs use its nutrients for growth (Kondos and Foale, 1983; Bell and Keith, 1989). However, all the experiments cited above were conducted in growing pigs; for that reason it is unknown whether the coefficients

† E-mail: mariscal.gerardo@inifap.gob.mx

of ileal apparent digestibility (CIAD) of protein and amino acids (AA) that have been determined in growing pigs are appropriate to pigs at weaning. Consequently, the aim of this study was to determine the CIAD of CP and AA in S, both for piglets at weaning and for growing pigs to test the hypothesis that CP and AA digestibility of S is lower in weaned piglets than in growing pigs.

Material and methods

Two experiments using weaned piglets and growing pigs of the same herd were conducted according to the guidelines of the International Guiding Principles for Biomedical Research Involving Animals, and the 'Mexican Official Standard for the Production, Protection and Use of Lab Animals' (Diario Oficial de la Federación, 2001).

Experiment 1: weaned piglets

A total of 20 piglets, Duroc × Landrace, from four litters, weaned at 17 ± 0.5 days of age and weighing 5.5 ± 0.7 kg, were assigned to one of five different treatment groups based on litter of origin and body weight (four piglets per treatment). Piglets were placed individually in metabolism cages provided with a self-feeder and a nipple watering device, in a room with a controlled temperature of $26 \pm 3^\circ\text{C}$. To have the piglets eating solid feed, but to alter as little as possible the enzymatic profile of suckling animals, from day 17 to day 20 of age all piglets were fed with a dry mash mixture composed of dry whole milk (80%) and maize starch (20%), three times daily, at 0800, 1300 and 1800 h. When the piglets were 21 days of age, a 'T' cannula was fitted in the terminal ileum (Reis de Souza *et al.*, 2000). After surgery, piglets from each treatment group received one of five treatments: a reference diet containing 200 g of CP/kg using casein (C) as the sole protein source, or one of four casein-sorghum (C-S) diets (Tables 1 and 2). The amount of S used in the C-S diets was 135, 307, 460 and 614 g/kg. To ensure isoproteic diets the appropriate amounts of C and maize starch were adjusted. In addition to the variable amounts of starch, the five diets contained the same amount of lactose, maize oil, calcium carbonate, dicalcium phosphate, salt and a mixture of vitamins and minerals. Calcium carbonate and dicalcium phosphate were used to adjust the Ca and P levels, and premixes of vitamins and trace elements were added to satisfy or exceed the recommended daily allowances (NRC, 1998). Chromic oxide was added at a rate of 3 g/kg as an indigestible marker.

Piglets were fed during 14 days with the same experimental diet, starting on day 22 according to the previously described schedule. The experimental period lasted 7 days each: 5 days for adaptation and the remaining 2 days for collection of ileal digesta. Piglets were given 150 g feed/day during the 1st week and 300 g/day during the 2nd week; the piglets had free access to water at all times.

Ileal digesta were collected using plastic bags (5×3 cm) containing 2 ml of 0.2 M HCl solution to block any bacterial activity. The bags were attached to the cannula using a

Table 1 Chemical composition of casein and sorghum as fed*

	Casein	Sorghum
Dry matter	904.2	905.1
Protein	903.7	77.7
Tannins		23.9
Essential amino acids		
Lysine	60.5	2.2
Arginine	31.6	3.5
Histidine	21.9	1.8
Leucine	83.8	10.4
Isoleucine	49.4	3.3
Valine	64.7	4.3
Phenylalanine	43.9	4.3
Methionine	25.9	1.5
Threonine	42.4	4.1
Non essential amino acids		
Aspartic acid	63.8	4.8
Glutamic acid	208.0	14.9
Serine	48.1	3.4
Glycine	14.6	3.1
Alanine	25.9	7.3
Tyrosine	47.9	2.8
Cysteine	5.3	2.1
Proline	92.4	7.74

*In g/kg of raw material. The values reported are measured.

rubber band and ileal digesta were collected six times per day, from 0800 to 2100 h on day 1, and from 0900 to 2200 h on day 2. During the first 2 hours of collection (0800 to 1000 on day 1 and 0900 to 1100 on day 2) the cannulae remained open. The cannulae were then alternately opened and closed for periods of 60 min, and one bagful of digesta was collected every 2nd hour. As the collecting bags were withdrawn, the samples of ileal digesta were transferred to a container for immediate freezing at -20°C until lyophilization.

Experiment 2: growing pigs

Fifteen castrated, Duroc × Landrace, pigs weighing 57.8 ± 2.8 kg were used for this experiment (five pigs by treatment). A 'T' cannula was fitted in the terminal ileum when the pigs weighed 45 kg. After surgery, the pigs were placed individually in metabolism cages provided with a self-feeder and a nipple watering device. Room temperature was maintained at $19 \pm 2^\circ\text{C}$. The post-surgery period lasted 14 days, during which the pigs received a growth diet that provided 160 g of CP/kg of feed twice daily (0800 and 1700 h). The amount fed was increased every day until the pigs reached the intake level observed before surgery. Pigs were randomly allotted to one of three treatments (Table 2), a reference C-maize starch diet containing C as the sole protein source, a C-S diet (both at about 160 g of CP/kg), and a fortified S diet containing 68 g of CP/kg. These diets contained fixed amounts of maize oil, sucrose, salt and a mixture of vitamins and minerals. Calcium carbonate and dicalcium phosphate were used to adjust the Ca and P levels, while a premix of vitamins and trace elements was added to satisfy or exceed the recommended daily allowances (NRC, 1998). Chromic oxide was included at

Table 2 Composition of experimental diets in g/kg of feed

Treatments*	Weaned piglets					Growing pigs		
	C	C-S-15	C-S-37	C-S-46	C-S-61	C	C-S-70	S
Sorghum		153.5	307.0	460.5	614.0		700.0	875.0
Casein	221.0	207.8	194.6	181.4	168.0	177.0	117.0	
Maize starch	561.0	420.7	280.4	140.1		580.6	58.0	
Lactose	126.3	126.3	126.3	126.3	126.3			
Sucrose						50.0	50.0	50.0
Maize oil	40.0	40.0	40.0	40.0	40.0	40.0	40.0	40.0
Maize straw						117.4		
Chromic oxide	3.0	3.0	3.0	3.0	3.0	3.0	3.0	3.0
Salt	3.8	3.8	3.8	3.8	3.8	3.5	3.5	3.5
Antibiotic premix [‡]	1.0	1.0	1.0	1.0	1.0			
Antioxidant [†]	0.2	0.2	0.2	0.2	0.2	0.2	0.2	0.2
Calcium carbonate	4.0	4.0	4.0	4.0	4.0	6.0	6.0	6.0
Dicalcium phosphate	35.0	35.0	35.0	35.0	35.0	20.0	10.0	13.0
Piglets premix [¶]	4.8	4.8	4.8	4.8	4.8			
Pig premix [§]						2.3	2.3	2.3
Chemical composition (g/kg of feed)								
NDF		16.6	33.2	49.7	66.3	75.6	75.6	94.5
ADF		12.7	25.4	38.1	50.8	48.4	57.9	72.4
CP	184.4	181.6	181.0	193.0	183.1	150.8	166.0	68.0

*Weaning piglets: C = casein diet; C-S-15 = casein-sorghum at 15%; C-S-37 = casein-sorghum at 37%; C-S-46 = casein-sorghum at 46%; C-S-61 = casein-sorghum at 61%. Growing pigs: C = casein diet; C-S-70 = casein-sorghum at 70%; S = sorghum.

[‡]Antibiotic premix that provided per kilogram of PREMIX: Chlortetracycline 44 g/kg, Sodium sulfadimethylpyrimidine 44 g/kg and Penicillin G procaine 22 g/kg.

[†]ANTI-OX-HP, Compagnie Chimique d'Aquitaine.

[¶]Premix that provided per kilogram of piglet diet: vitamin A 10 200 IU, vitamin D 1980 IU, vitamin E 60 IU, vitamin K 1.20 mg, Choline 967 mg, Niacin 36 mg, Pantothenic Acid 17 mg, Riboflavin 7.2 mg, vitamin B₁₂ 40 µg, Thiamine 0.30 mg, Pyridoxine 0.31 mg, Biotin 0.08 mg, Folic Acid 0.75 mg, Cu 14.4 mg, I 0.96 mg, Fe 120 mg, Mn 36 mg, Se 0.3 mg, Zn 144 mg.

[§]Premix that provided per kilogram of growing pigs diet: vitamin A 6400 IU, vitamin D 1280 IU, vitamin E 30 IU, vitamin K 0.8 mg, Choline 375 mg, Niacin 24 mg, Pantothenic Acid 11.0 mg, Riboflavin 4.8 mg, vitamin B₁₂ 30 µg, Cu 8.4 mg, I 0.56 mg, Fe 70.0 mg, Mn 21.0 mg, Se 0.18 mg, Zn 84.0 mg.

3 g/kg as an indigestible marker. During the experimental period, the castrated pigs were fed 2.5 times their maintenance requirement of digestible energy (DE; 460 kJ of DE per kg of W^{0.75}) (INRA, 1984), and had free access to water.

The experimental period was as described for experiment 1, except for the following modifications. Ileal digesta were collected in plastic bags (11 × 5 cm) containing 10 ml of 0.2 M HCl. On day 1 the bags were attached to the cannulae using a rubber band at 0800 h and the ileal contents were collected from 0800 to 1000 h, and the cannulae remained open every other hour until 0900 h on day 2. On day 2, the cannulae remained open from 0900 to 1100 h, and every other hour until the end of the collection period (i.e. 0800 h on day 3).

At the end of both experiments, narcosis was induced in piglets and growing pigs with CO₂ inhalation, followed by euthanasia by exsanguination. A post-mortem examination was performed to verify the integrity of their small intestines.

Preparation of samples and chemical analysis

The samples of ileal digesta from experiments 1 and 2 were lyophilized and ground in a laboratory mill using a 0.5-mm mesh sieve (Arthur H. Thomas Co. Philadelphia, PA, USA). The following analyses were performed on the experimental diets and the samples of ileal digesta: dry matter (DM) and CP according to AOAC (2000) methods 934.01 and 976.05, respectively, and chromic oxide according to the method of

Fenton and Fenton (1979). Samples were prepared for determination of AA according to method 994.12 of the AOAC (2000). Samples were hydrolyzed at 110°C for 24 h in 6 M HCl. For assay of methionine and cysteine, oxidation with performic acid was carried out before the AA analysis, using reverse phase HPLC (1100 HPLC Hewlett Packard), according to the method described by Henderson *et al.* (2000). In addition, the level of tannins in S grain was determined according to the method of Price *et al.* (1978).

Data analysis

Estimation of the ileal apparent digestibility of protein and AA. The IAD coefficients for DM, CP and AA in the experimental diets were estimated using the following equation:

$$IAD_D = 1 - [(I_D \times A_F)/(A_D \times I_F)] \quad (1)$$

in which IAD_D is the coefficient of the ileal apparent digestibility (CIAD) of a nutrient in the diet; I_D is the concentration of the marker in the diet (mg/kg of DM); A_F is the concentration of the nutrient in the ileal digesta (mg/kg of DM); A_D is the concentration of the nutrient in the diet (mg/kg of DM); and I_F is the concentration of the marker in the ileal digesta (mg/kg of DM).

Estimation of the ileal apparent digestibility of S by the difference method. Using C as the basal feed ingredient, the

ileal apparent digestibility values for CP and AA in S were calculated by the difference method proposed by Fan and Sauer (1995):

$$IAD_A = [IAD_D - (D_B \times S_B)]/S_A \quad (2)$$

in which IAD_A is the coefficient of ileal apparent digestibility of a component in the assayed ingredient under the assumption of the additivity of digestible or indigestible components; IAD_D is the CIAD of protein or AA in the assayed diet; D_B is the coefficient of apparent digestibility of a component in the reference feed ingredient; S_B is the contribution of a nutrient in the reference ingredient to the assayed diet; and S_A is the contribution of a nutrient in the assayed ingredient to the assayed diet.

Estimation of the ileal apparent digestibility of S by the regression method. As defined by Mosenthin *et al.* (2000), in this method the basal and the assay feed ingredient are evaluated simultaneously. The basal and the assay feed ingredient provide the sole assay nutrients in the diet. The ingredients (C and S) were mixed at various graded levels. To perform the regression analyses the amount (g/kg) of nutrient furnished by C was utilized as the regressor using the REG procedure of SAS (Statistical analysis system, v 9.1, 2002):

$$y = a + bX, \quad (3)$$

where y is the predicted digestibility of a nutrient in the assay diet; the intercept a represents the digestibility of S nutrient (0 g/kg of C); b is the slope and represents the increase in digestibility due to an increase in the amount (g/kg) of the nutrient assay furnished by C; and X represents the amount (g/kg) of C in the assay diet.

Statistical analysis

The homogeneity of variance for all data was tested by the test of Levene using hovtest of SAS (version 9.1).

Data for the CIAD for CP or AA in experiment 1 (weaned piglets) were analyzed using the GLM procedure of SAS (version 9.1, 2002) for a split plot design, according to the following model:

$$Y_{ijkl} = \mu + T_i + A_k(T_i) + P_j + (P_j \times T_i) + e_{ijk}, \quad (4)$$

where Y_{ijkl} = dependent variable, μ = general mean, T_i = treatments 1, 2, 3, 4 and 5; A_k = animal as the whole plot unit (1, 2, 3, ... and 20), which was used as the error term to test the significance of treatment, P_j = collection period for the subplot 1, 2, and e_{ijk} = experimental error. Treatment means were compared using the Tukey method and the linear effect was tested using orthogonal contrast (Steel and Torrie, 1980).

The data for the CIAD for CP and AA in experiment 2 (growing pigs) were analyzed using the GLM procedure of SAS (version, v 9.1, 2002), for a randomized complete block (the block was a group of three animals, one per treatment used at the same time) design under the general model:

$$Y_{ij} = \mu + T_i + B_j + e_{ij}, \quad (5)$$

where Y_{ij} = is the dependent variable; μ = general mean; T_i = treatments 1, 2 and 3; B_j = block; 1, 2, 3, 4 and 5, and e_{ij} = experimental error. Treatment means were compared using the Tukey method.

Because both experiments were done at the same time under the same conditions with animals belonging to the same herd and with the same protein sources, the CIAD determined by the difference method in each experiment (treatment C–S-70 in growing pigs and treatment C–S-61 in weaned piglets) were combined and analyzed as a Completely Random Design (Steel and Torrie, 1980) using the GLM procedure of SAS (version 9.1, 2002). Treatment means were compared using the Tukey method.

Results

All the variables had a homogenous variance ($P > 0.05$), excepting valine, methionine and proline in weaned piglets and protein and alanine in growing pigs. Therefore, the digestibility values for these AA were subjected to angular transformation (Steel and Torrie, 1980), followed by statistical analysis as described above. Data are presented as back-transformed in Tables 5 and 6.

Experiment 1: weaned piglets

The CIAD are shown in Tables 3 and 4. The CIAD for CP and AA decreased linearly ($P < 0.05$) as the amount of S in the diet increased. The average ileal digestibility of AA from C was 0.858 ± 0.111 , and this decreased to 0.663 ± 0.191 when the percentage of S in the diet was 61. The greater decrease was observed in CP (from 0.859 to 0.633) and the smaller in lysine (from 0.932 to 0.895). The ileal digestibilities during the 1st and 2nd weeks after weaning were not different.

The CIAD estimated by the regression or difference methods were similar for leucine, cysteine, glutamic acid, serine, alanine and tyrosine and different for the other AA (Table 4). The lower digestibility obtained by the regression method were for protein, aspartic acid and proline (0.088, 0.130 and 0.169, respectively), and the higher were for lysine, histidine and methionine (0.588, 0.587 and 0.525, respectively). The capacity of the piglets to digest CP and AA from S was low.

Experiment 2: growing pigs

The results are shown in Tables 5 and 6. In general, the CIAD of protein and AA (except alanine and cysteine) were similar ($P > 0.05$) in the C and the C–S diets, and were higher ($P < 0.05$) than those of the S diet. The CIAD for S obtained by the difference method were higher ($P < 0.05$) than those obtained using the direct method, except for lysine, isoleucine, valine, methionine, threonine and cysteine. Using the direct method the AA with lower digestibility were glycine, arginine and proline (0.337, 0.470 and 0.518, respectively).

Comparison between weaned piglets v. growing pigs

The results are shown in Table 7. Excepting lysine and cysteine growing pigs digest better ($P < 0.01$) all the AA and protein than weaned piglets.

Table 3 Coefficients of ileal apparent digestibility of experimental diets in weaned piglets

Treatments*	C	C-S-15	C-S-37	C-S-46	C-S-61	s.e.m.	Period		s.e.m.
							1	2	
Dry matter ¹	0.894	0.852	0.791	0.806	0.703	0.0135	0.816	0.802	0.0085
Protein ¹	0.859	0.732	0.813	0.763	0.633	0.0180	0.760	0.761	0.0116
Essential amino acids									
Lysine ³	0.932	0.931	0.892	0.908	0.895	0.0147	0.910	0.913	0.0095
Arginine ²	0.771	0.704	0.762	0.747	0.582	0.0327	0.686	0.741	0.0210
Histidine ²	0.926	0.930	0.880	0.893	0.805	0.0227	0.890	0.883	0.0145
Leucine ¹	0.924	0.902	0.844	0.823	0.773	0.0126	0.849	0.857	0.0081
Isoleucine ¹	0.905	0.871	0.816	0.828	0.748	0.0112	0.832	0.835	0.0069
Valine ¹	0.924	0.890	0.842	0.843	0.771	0.0101	0.858	0.850	0.0065
Phenylalanine ¹	0.910	0.890	0.835	0.875	0.785	0.0104	0.854	0.864	0.0067
Methionine	0.941	0.927	0.878	0.880	0.810	0.0121	0.888	0.886	0.0077
Threonine ¹	0.874	0.815	0.765	0.753	0.693	0.0219	0.764	0.796	0.0141
Non-essential amino acids									
Aspartic acid ¹	0.865	0.838	0.774	0.784	0.686	0.0229	0.781	0.798	0.0147
Glutamic acid ¹	0.932	0.916	0.862	0.849	0.785	0.0108	0.866	0.872	0.0069
Serine ¹	0.855	0.812	0.758	0.774	0.673	0.0199	0.770	0.779	0.0128
Glycine ²	0.641	0.633	0.450	0.645	0.416	0.0362	0.558	0.555	0.0233
Alanine ¹	0.788	0.726	0.619	0.650	0.581	0.0251	0.666	0.680	0.0161
Tyrosine ¹	0.932	0.924	0.865	0.876	0.793	0.0091	0.875	0.881	0.0059
Cysteine	0.553	0.514	0.530	0.488	0.467	0.0772	0.485	0.536	0.0496
Proline ¹	0.907	0.865	0.771	0.814	0.705	0.0162	0.817	0.807	0.0104

*C = casein diet; C-S-15 = casein-sorghum at 15%; C-S-37 = casein-sorghum at 37%; C-S-46 = casein-sorghum at 46%; C-S-61 = casein-sorghum at 61%.

¹Linear effect ($P < 0.001$).

²Linear effect ($P < 0.01$).

³Linear effect ($P < 0.05$).

Table 4 Coefficients of ileal apparent digestibility of CP and AA in sorghum determined in weaned piglets using the regression or difference method

Method	Regression*							Difference method	CP or AA
	<i>a</i>	s.e.m.	Prob	<i>b</i>	s.e.m.	Prob	R^2 [†]	s.d.	Number (%) [#]
CP	0.088	0.083	0.29	0.042	0.005	0.001	0.67	0.146 ± 0.135	26
Essential AA									
Lysine	0.588	0.139	0.002	0.588	0.139	0.001	0.15	0.681 ± 0.174	11
Arginine	0.248	0.106	0.030	0.846	0.191	0.001	0.37	0.335 ± 0.141	31
Histidine	0.587	0.072	0.001	0.732	0.176	0.001	0.34	0.444 ± 0.288	25
Leucine	0.407	0.075	0.001	0.270	0.045	0.001	0.52	0.413 ± 0.136	30
Isoleucine	0.313	0.075	0.001	0.573	0.082	0.001	0.60	0.219 ± 0.140	21
Valine	0.352	0.075	0.001	0.038	0.056	0.001	0.62	0.169 ± 0.099	18
Phenylalanine	0.382	0.076	0.001	0.0576	0.090	0.001	0.55	0.444 ± 0.134	27
Methionine	0.525	0.042	0.001	0.0763	0.089	0.001	0.72	0.267 ± 0.167	19
Threonine	0.245	0.101	0.020	0.707	0.134	0.001	0.46	0.329 ± 0.125	28
Non essential AA									
Aspartic acid	0.130	0.132	0.330	0.563	0.111	0.001	0.44	0.167 ± 0.188	22
Glutamic acid	0.242	0.091	0.020	0.148	0.021	0.001	0.58	0.246 ± 0.177	19
Serine	0.210	0.090	0.030	0.661	0.105	0.001	0.55	0.207 ± 0.218	22
Glycine	0.271	0.093	0.010	1.353	0.405	0.002	0.25	0.187 ± 0.198	49
Alanine	0.392	0.056	0.001	0.695	0.130	0.001	0.46	0.364 ± 0.154	54
Tyrosine	0.217	0.071	0.005	0.744	0.071	0.001	0.72	0.239 ± 0.156	19
Cysteine	0.487	0.083	0.001	0.453	1.126	0.001	0.01	0.498 ± 0.205	61
Proline	0.169	0.082	0.050	0.392	0.049	0.001	0.69	0.207 ± 0.130	25

AA = amino acid; Prob = probability.

*Lineal regression according to equation: $Y = a + bX$, where the intercept *a* represent the digestibility of sorghum nutrient (0g of casein); *b* is the slope and represents the increase in digestibility due to an increase in the nutrient assay furnished by casein; X represent the g/kg of casein in the assay diet.

[†] R^2 = Coefficient of determination for the linear regression line.

[#]Percentage of nutrient furnished by sorghum.

Table 5 Coefficients of ileal apparent digestibility of experimental diets in growing pigs

Treatments*	C	C-S-70	S	s.e.m.	Probability [†]
DM	0.775	0.808	0.780	0.0146	0.267
CP	0.859 ^a	0.813 ^a	0.543 ^b	0.0190	0.001
Essential AA					
Lysine	0.996 ^a	0.926 ^a	0.647 ^b	0.0477	0.002
Arginine	0.841 ^a	0.760 ^a	0.468 ^b	0.0474	0.002
Histidine	0.889 ^a	0.892 ^a	0.714 ^b	0.0219	0.001
Leucine	0.934 ^a	0.902 ^a	0.800 ^b	0.0139	0.001
Isoleucine	0.921 ^a	0.883 ^a	0.704 ^b	0.0291	0.002
Valine	0.916 ^a	0.863 ^a	0.647 ^b	0.0246	0.003
Phenylalanine	0.940 ^a	0.927 ^a	0.791 ^b	0.0254	0.006
Methionine	0.926 ^a	0.883 ^a	0.590 ^b	0.0363	0.001
Threonine	0.839	0.764	0.628	0.0502	0.056
Non essential AA					
Aspartic acid	0.894 ^a	0.841 ^a	0.617 ^b	0.0304	0.001
Glutamic acid	0.916 ^a	0.892 ^a	0.742 ^b	0.0172	0.001
Serine	0.849 ^a	0.827 ^a	0.625 ^b	0.0439	0.011
Glycine	0.632 ^a	0.586 ^a	0.337 ^b	0.0584	0.015
Alanine	0.771	0.784	0.711	0.0243	0.084
Tyrosine	0.951 ^a	0.943 ^a	0.743 ^b	0.0164	0.001
Cysteine	0.417	0.578	0.675	0.0837	0.235
Proline	0.888 ^a	0.818 ^a	0.511 ^b	0.0368	0.001

DM = dry matter; AA = amino acid.

*C = casein diet; C-S-70 = casein-sorghum at 70%; S = sorghum.

[†]Means with different superscript (a,b) differ at probability level shown in the sixth column.

Table 6 Coefficients of ileal apparent digestibility of CP and AA in sorghum determined in growing pigs using the difference or direct method

Treatments*	Difference	Direct	s.e.m.	Probability [†]
CP	0.718 ^a	0.545 ^b	0.065	0.001
Essential AA				
Lysine	0.592	0.668	0.0743	0.488
Arginine	0.630	0.470	0.0542	0.066
Histidine	0.897 ^a	0.718 ^b	0.0404	0.012
Leucine	0.858 ^a	0.801 ^b	0.0151	0.026
Isoleucine	0.782	0.711	0.0386	0.221
Valine	0.722	0.649	0.0332	0.152
Phenylalanine	0.904 ^a	0.791 ^b	0.0240	0.009
Methionine	0.753	0.608	0.0815	0.239
Threonine	0.625	0.638	0.0578	0.881
Non essential AA				
Aspartic acid	0.717 ^a	0.618 ^b	0.0245	0.039
Glutamic acid	0.833 ^a	0.744 ^b	0.0194	0.010
Serine	0.771 ^a	0.628 ^b	0.0425	0.040
Glycine	0.548 ^a	0.337 ^b	0.0622	0.040
Alanine	0.792 ^a	0.712 ^b	0.0227	0.036
Tyrosine	0.918 ^a	0.744 ^b	0.0326	0.004
Cysteine	0.649	0.681	0.0777	0.776
Proline	0.670 ^a	0.518 ^b	0.0421	0.0314

AA = amino acid.

[†]Means with different superscript (a,b) differ at probability level shown in the fifth column.

Table 7 Comparison of ileal digestibility coefficients between weaned piglets and growing pigs

	Weaned piglets		s.e.m.	Probability [†]
	C-S-61*	C-S-70 [†]		
CP	0.146 ^a	0.718 ^b	0.0426	0.001
Essential AA				
Lysine	0.681	0.592	0.0764	0.429
Arginine	0.335 ^a	0.630 ^b	0.0534	0.010
Histidine	0.444 ^a	0.897 ^b	0.0969	0.010
Leucine	0.413 ^a	0.858 ^b	0.0437	0.001
Isoleucine	0.219 ^a	0.782 ^b	0.0618	0.001
Valine	0.169 ^a	0.722 ^b	0.0387	0.001
Phenylalanine	0.444 ^a	0.904 ^b	0.0445	0.001
Methionine	0.267 ^a	0.753 ^b	0.0813	0.010
Threonine	0.329	0.625 ^b	0.0520	0.010
Non essential AA				
Aspartic acid	0.167 ^a	0.717 ^b	0.0541	0.001
Glutamic acid	0.246 ^a	0.833 ^b	0.0334	0.001
Serine	0.207 ^a	0.771 ^b	0.0776	0.001
Glycine	0.187 ^a	0.548 ^b	0.0703	0.010
Alanine	0.364 ^a	0.792 ^b	0.0493	0.001
Tyrosine	0.239 ^a	0.918 ^b	0.0564	0.001
Cysteine	0.498	0.649	0.0915	0.272
Proline	0.207 ^a	0.670 ^b	0.0544	0.001

AA = amino acid.

*C-S-61 = casein-sorghum at 61%.

[†]C-S-70 = casein-sorghum at 70%.

[†]Means with different superscript (a,b) differ at probability level shown in the fifth column.

Discussion

The digestibility of S is affected by tannins, and by the presence of α -kaffirin, a S storage protein, which is resistant to pepsin (Oria *et al.*, 1995). The low digestibility of proline and glycine could be attributable to the capacity of tannins to stimulate the synthesis of a proline-rich protein (Jansman, 1993; Haslam, 1998); this protein may be acidic, basic or glycosylated (Bennic, 1982), and may contain 40% proline and 20% glycine. It acts as the first line of defense against dietary tannins (Mehansho *et al.*, 1985). Its secretion responds to the ability of tannins to precipitate proteins, and it shows a particular preference for combination with proline (Hagerman and Butler, 1981; Mitaru *et al.*, 1984; Kim *et al.*, 2004). In the case of arginine, Charlton *et al.* (2002) reported that this AA strengthens the interaction between proteins and tannins forming an additional site of interaction.

The CIAD of protein and AA from S in growing pigs are in the range of values reported previously (Lin *et al.*, 1987; Mariscal-Landín, 1992; Mariscal-Landín *et al.*, 2004).

The lower CIAD values obtained with the direct method than with the difference method has been reported and criticized in pigs by Fan and Sauer (1995). This is the reason why values for standardized ileal digestibility are now commonly used in diet formulation.

The ileal digestibility coefficient of protein from S obtained in piglets by either the regression or the difference method

was low (0.088 and 0.146, respectively). The low protein digestibility could be caused by the negative effect of tannins or the presence of α -kaffirin; and be aggravated by the lower capacity of weaned piglets to digest vegetable protein when compared with growing pigs (Mariscal-Landín *et al.*, 2008). For this reason the use of highly digestible proteins is recommended in starter feeds (Makkink *et al.*, 1994; Shon *et al.*, 1994).

Even if the average digestibility of AA from S was low (0.339 or 0.319 by the regression or difference method, respectively), however, all the AA had a higher digestibility than protein. The lower protein digestibility could be due to a high non-amino nitrogen content in the digesta, probably provided by the amino sugars (50% to 80% of the molecular weight) of mucin, a glycoprotein with a high content of *N*-acetyl glucosamine and *N*-acetyl galactosamine (Deplancke and Gaskins, 2001). Mucin is the principal component of the mucus layer in the gastrointestinal tract (Corfield and Shukla, 2003; Montagne *et al.*, 2004). The mucus layer functions as an interface between the external environment (the intestinal lumen) and epithelial cells (enterocytes); its principal task is the protection of the epithelium. An increase in the loss of endogenous protein (enzymes and mucins) has been reported in pigs that are fed S (Jondreville *et al.*, 2001), as well an increase of mucin in the feces of mice fed S containing high levels of tannins (Sell *et al.*, 1985).

Ileal digestibility coefficients are lower in piglets than in growing pigs, as shown by our results. The greater difference was observed for protein because the CIAD in the piglets was only 27% of that observed in the growing pigs. Lysine was the AA that was most digestible in piglets, independent of method utilized for analysis; this could be because this was the AA with the lowest proportion furnished by S (11% of dietary lysine); consequently the impact of the lysine furnished by S was negligible. In fact, this is a limitation of the used methods (regression and difference), because the proportion of AA (11% of lysine to 61% of cysteine) and protein (26%) furnished by S was low. Due to the low digestive capacity of piglets that limits the use of one plant ingredient as a sole protein source these are the best methods available to measure the ileal digestibility of raw materials in weaned piglets.

Conclusions

The protein and AA of S are less digestible in piglets than in growing pigs. In piglets the ileal digestibility of protein is lower than the average ileal digestibility of AA. The coefficients estimated in growing pigs using the direct method were lower than those estimated by the difference method.

Acknowledgements

The authors acknowledge Benito Mar-Botello, MSc, for surgical procedures; Mariela Camacho, MSc for amino acid analysis and chemistry and Ericka Ramirez for general laboratory analyses. And also to the National Council for Science and Technology of

Mexico (CONACYT) for the financial support given to Research Project G13972

References

- AOAC 2000. Official methods of analysis. Association of official analytical chemists, 17th edition. AOAC, Arlington, VA, USA.
- Bansal S, Mishra A, Tomar A, Sharma S, Khanna VK and Garg GK 2008. Isolation and temporal endosperm expression of α -kaffirin gene of grain sorghum (*Sorghum bicolor* L. moench) var. M 35-1 for introgression analysis of transgene. *Journal of Cereal Science* 48, 808–815.
- Bell JM and Keith MO 1989. Factors affecting the digestibility by pigs of energy and protein in wheat, barley and sorghum diets supplemented with canola meal. *Animal Feed Science and Technology* 24, 253–265.
- Bennic A 1982. Salivary proline rich proteins. *Molecular and Cellular Biochemistry* 45, 83–89.
- Brand TS, Badenhorst HA and Siebrits FK 1990. The use of pigs both intact and with ileo-rectal anastomosis to estimate the apparent and true digestibility of amino acids in untreated, heat-treated and thermal-ammoniated high-tannin grain sorghum. *South African Journal of Animal Science* 20, 223–228.
- Charlton AJ, Baxter NJ, Khan ML, Moir AJG, Haslam E, Davies AP and Williamson MP 2002. Polyphenol/peptide binding and precipitation. *Journal of Agricultural and Food Chemistry* 50, 1593–1601.
- Corfield AP and Shukla AK 2003. Mucins: vital components of the mucosal defensive barrier. *Genomic Proteomic Technology* 3, 20–23.
- Deplancke B and Gaskins HR 2001. Microbial modulation of innate defense: goblet cells and the intestinal mucus layer. *The American Journal of Clinical Nutrition* 73, 1131S–1141S.
- Diario Oficial de la Federación 2001. Especificaciones técnicas para la producción, cuidado y uso de los animales de laboratorio. Norma Oficial Mexicana NOM-062-ZOO-1999. Diario Oficial De La Federación, Miércoles 2 de Agosto. Segunda Sección.
- Fan MZ and Sauer WC 1995. Determination of ileal apparent amino acid digestibility in barley and canola meal for pigs with the direct, difference, and regression methods. *Journal of Animal Science* 73, 2364–2374.
- Fenton TW and Fenton M 1979. An improved procedure for determination of chromic oxide in feed and feces. *Canadian Journal of Animal Science* 59, 631–634.
- Hagerman AE and Butler LG 1981. The specificity of proanthocyanidin-protein interactions. *The Journal of Biological Chemistry* 256, 4494–4497.
- Haslam E 1998. Practical polyphenolics: from structure to molecular recognition and physiological action. Cambridge University Press, Cambridge.
- Henderson JH, Ricker RD, Bidlingmeyer BA and Woodward C 2000. Rapid, accurate and reproducible HPLC analysis of amino acids. Amino acid analysis using Zorbax Eclipse AAA columns and the Agilent 1100 HPLC. *Agilent Technologies Part No.5980-1193E*, 10pp.
- INRA 1984. L'alimentation des animaux monogastriques: porc, lapin, volailles. Institut National de la Recherche Agronomique, Paris, France.
- Jansman AJM 1993. Tannins in feedstuffs for simple-stomached animals. *Nutrition Research Reviews* 6, 209–236.
- Jondreville C, van den Broecke J, Gatel F, Grosjean F, van Cauwenbergh S and Sève B 2001. Ileal digestibility of amino acids and estimates of endogenous amino acid losses in pigs fed wheat, triticale, rye, barley, maize and sorghum. *Animal Research* 50, 119–134.
- Kim H, House WA and Miller DD 2004. Habitual tea consumption protects against the inhibitory effects of tea on iron adsorption rats. *Nutrition Research* 24, 383–393.
- Kondos AC and Foale MA 1983. Comparison of the nutritional value of low and medium tannin sorghum grains for pigs. *Animal Feed Science and Technology* 8, 85–90.
- Lin FD, Knabe DA and Tanksley TD 1987. Apparent digestibility of amino acids, gross energy and starch in corn, sorghum, wheat, barley, oats groats and wheat middlings for growing pigs. *Journal of Animal Science* 64, 1655–1663.
- Makkink CA, Berntsen PJM, op den Kamp BML, Kemp B and Verstegen WA 1994. Gastric protein breakdown and pancreatic enzyme activities in response to two different dietary protein sources in newly weaned pigs. *Journal of Animal Science* 72, 2843–2850.
- Mariscal-Landín G 1992. Facteurs de variation de l'utilisation digestive des acides aminés chez le porc. PhD. Thesis Rennes 1 University, 134pp.

Mariscal-Landín, Reis de Souza and Avalos

- Mariscal-Landín G, Avellaneda JH, Reis de Souza TC, Aguilera A, Borbolla GA and Mar B 2004. Effect of tannins in sorghum on amino acid ileal digestibility and on trypsin (E.C.2.4.21.4) and chymotrypsin (E.C.2.4.21.1) activity of growing pigs. *Animal Feed Science and Technology* 117, 245–264.
- Mariscal-Landín G, Reis de Souza TC, Parra SJE, Aguilera BA and Mar BB 2008. Ileal digestibility of protein and amino acids from canola meal in weaned piglets and growing pigs. *Livestock Science* 116, 53–62.
- Mehansho H, Clements S, Sheares BT, Smith S and Carlson DM 1985. Induction of proline-rich glycoprotein synthesis in mouse salivary glands by isoproterenol and by tannins. *The Journal of Biological Chemistry* 260, 4418–4423.
- Mitaru BN, Reichert RD and Blair R 1984. The binding of dietary protein by sorghum tannins in the digestive tract of pigs. *The Journal of Nutrition* 114, 1787–1796.
- Montagne L, Piel C and Lallès JP 2004. Effect of diet on mucin kinetics and composition: nutrition and health implications. *Nutrition Reviews* 62, 105–114.
- Mosenthin R, Sauer WC, Blank R, Huisman J and Fan MZ 2000. The concept of digestible amino acids in diet formulation for pigs. *Livestock Production Science* 64, 265–280.
- Mossé J, Huet JC and Baudet J 1988. The amino acid composition of whole sorghum grain in relation to its nitrogen content. *Cereal Chemistry* 65, 271–277.
- NRC 1998. Nutrient requirements of swine, 10th edition. National Academy Press, Washington, DC.
- Oria PM, Hamaker BR and Smith S 1995. Resistance of sorghum α , β and γ kafirins to pepsin digestion. *Journal of Agricultural and Food Chemistry* 43, 2148–2153.
- Price ML, Steve VS and Butler LC 1978. A critical evaluation of the vanillin reaction as an assay for tannin in sorghum grain. *Journal of Agricultural and Food Chemistry* 26, 1214–1218.
- Ramírez RE, Anaya EAM and Mariscal LG 2005. Predicción de la composición química del grano de sorgo mediante espectroscopia de reflectancia en el infrarrojo cercano (NIRS). *Técnica Pecuaria en México* 43, 1–11.
- Reis de Souza TC, Mar BB and Mariscal LG 2000. Canulación de cerdos posdestete para pruebas de digestibilidad ileal: desarrollo de una metodología. *Técnica Pecuaria en México* 38, 143–150.
- SAS. Statistical Analysis Systems Institute. User's guide, version 9.1. 2002. SAS Institute Inc., Cary, NC, USA.
- Sell DR, Reed WM, Chrisman CL and Rogler JC 1985. Mucin excretion and morphology of the intestinal tract as influenced by sorghum tannins. *Nutrition Reports International* 31, 1369–1374.
- Shon KS, Maxwell CV, Southern LL and Buchanam DS 1994. Improved soybean protein sources for early-weaned pigs: II. Effects on ileal amino acid digestibility. *Journal of Animal Science* 72, 631–637.
- Steel RGD and Torrie JH 1980. Principles and procedures of statistics. A biometrical approach, 2nd edition. McGraw-Hill, New York.