Stress-Tolerant Feedstocks for Sustainable Bioenergy Production on Marginal Land

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Abstract Given the mandated increases in fuel production from alternative sources, limited high-quality production land, and predicted climate changes, identification of stress-tolerant biomass crops will be increasingly important. However, existing literature largely focuses on the responses of a small number of crops to a single source of abiotic stress. Here, we provide a much-needed review of several types of stress likely to be encountered by biomass crops on marginal lands and under future climate scenarios: drought, flooding, salinity, cold, and heat. The stress responses of 17 leading biomass crops of all growth habits (e.g., perennial grasses, shortrotation woody crops, and large trees) are summarized, and we identify several that could be considered "all purpose" for multiple stress types. Importantly, we note that some of these crops are or could become invasive in some landscapes. Therefore, growers must take care to avoid dissemination of plants or propagules outside of cultivation.

Keywords Stress tolerance · Bioenergy · Feedstocks · Marginal land · Sustainability

Introduction

Production of second-generation biomass crops is growing in the USA, principally driven by the federal mandate [1] that requires novel feedstocks to offset greenhouse gas emissions from fossil fuels and minimize or avoid any negative impact on the global food supply. Thus, there has been much exploration into biomass crops that are capable of fast growth and high yields on land not suited to food production. Land that is unsuitable for traditional row crops, generally referred to as "marginal" land, may be appropriate for grazing or other uses [2], including biomass crop production. Use of marginal lands for bioenergy production could decrease land-use competition between energy and food crops and ameliorate land-use change associated with biomass crop expansion [3]. However, the fact that this land cannot economically support traditional crop production suggests it is suboptimal in some way and, therefore, may be associated with abiotic stress factors that must be overcome by any crop grown thereon. In addition, the definition of marginal land can also include land with slightly less severe abiotic stressors, potentially supporting some traditional row crops; however, even mild stress may cause those crops to perform suboptimally, causing farmers to operate at a loss, particularly if climate predictions increase the severity of abiotic stress. This paper explores abiotic stressors that may be encountered by biomass crops grown on marginal lands or under changing climate conditions and provides a breadth of options for selecting crops that can tolerate particular types of abiotic stress.

The Asia-Pacific Economic Cooperation (APEC) estimates that marginal lands make up approximately 400 million hectares across Asia, the Pacific Islands, Australia, and North America [4]. Other estimates put the global marginal land area anywhere from 1100 [5] to 6650 million hectares [2], depending on the parameters used to describe marginal (e.g., "nonfavored agricultural land," "abandoned or degraded cropland," or arid, forested, grassland, shrubland, or savanna habitats). The potential area available in the USA for cellulosic biomass crops and low-input, high-diversity native perennial mixtures ranges from 43 to 123 million hectares [5, 6]. The differences in these estimates reflect the inconsistencies in the usage of the term "marginal land," despite its common use in

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the bioenergy industry and literature [5, 7, 8]. Marginal lands are often described as degraded lands that are unfit for food production and/or of some ambiguously poor quality and are often termed unproductive [7]. Unproductive soils are characterized by unfavorable chemical and/or physical properties that limit plant growth and yield, including low water and nutrient storage capacity, high salinity, toxic elements, and poor texture [4, 9]. Further difficulties encountered in marginal landscapes include shallow soil depth due to erosion, poor drainage, low fertility, steep terrain, and unfavorable climate [2]. Despite the poor quality of marginal land and the potential problems it could present for its production, biomass is unlikely to be grown on high-quality land that is economically viable for traditional crops [7].

Climate change will exacerbate the issue of land marginalization and degradation [10, 11]. It has been predicted that negative impacts of climate change will increase over the next 25 years, with longer growing seasons (frost-free periods), increasing extreme precipitation events (both flooding and drought), fewer chilling hours, and a greater number of hot nights projected for most growing regions of the country [11]. Because these changes are projected to occur to a greater extent in certain regions of the country [11], crop production-potentially including biomass crops on marginal lands-may shift to novel regions where different stressors are present or growers may shift to different production systems. For example, to escape the predicted hot nights in the southeast, growers may shift to more northern regions where precipitation may be less frequent [11]. These changes willand already do-directly affect the physiology and reproductive development of many crop plants, including first generation biomass crops [12, 13]. Therefore, it will be important to select the most tolerant crops suitable for future climate scenarios [14].

In this work, we review and summarize the literature on the effects of three sources of abiotic stress that may be common to marginal lands-moisture, salinity, and temperature-and identify biomass crops that display tolerance to these types of stress. Our discussion is geared toward the USA but could be applied to similar circumstances in other regions. Although most plant matter has the potential to be converted into energy, we focus on the promising non-food crops identified in the US Department of Energy's Billion Ton Update [15] and in novel research programs that are investigating the next wave of potential biomass crops. Each of the crops in our report has been the subject of detailed study, investigating ecological, agronomic, physiological, or molecular responses to one or more stressors. Our goal is not to restate the detailed results of these studies, but instead to provide a comprehensive review of the effects of multiple stressors in the context of bioenergy production and a useful guide for growers to choose the best biomass crop for stressful growing conditions.

Moisture Stress

Framing the Issue

Water scarcity is the most limiting environmental factor to crop growth and yield [16] and is a major factor in the categorization of lands as "marginal" or unsuitable for crop production [17]. Drought is a prolonged period of time without significant precipitation, resulting in a shortage of water [18]. More frequent, severe, and longer-than-average duration of drought is predicted in the USA and globally due to changing climate conditions [11, 14, 19-21]. In addition to large-scale climatic factors, effects of drought can be exacerbated by local or regional factors including aridity, topography, or soil conditions [22]. Marginal lands are often characterized by sloped topography or eroded soils with little moisture-holding capacity [23]; therefore, the effects of drought-inducing climate changes have the potential to be stronger in these landscapes [10, 11]. For example, Lee et al. [24] observed that drought effects on biomass yield of perennial grasses varied according to topography, with greater effects at higher elevations.

Conversely, soils with poor drainage-and with the potential for flooding or waterlogging-are also typical of numerous marginal landscapes [23]. Many native Midwestern soils were poorly drained and unsuitable for annual crop production before an extensive network of tile drains were installed [23]. However, marginal land not in crop production is unlikely to have been tiled and therefore may be subject to problems associated with waterlogging in wet years or on wet sites. However, poor drainage is not only an issue in the Midwest. More than half of the freshwater wetlands in the USA have been drained or filled for agricultural use [25], and some have been or will be abandoned due to insufficient drainage [26]. Because rainfall events have become heavier and more frequent in recent years-and this trend is predicted to continue [11]—poorly drained soils can quickly become flooded or waterlogged.

Effects of Moisture Stress

Tissue expansion, dependent on cellular turgor pressure, is strongly tied to water availability, and plant growth is therefore limited by water deficits. This impacts germination, seedling growth, and stand establishment, and leads to reduced plant height, leaf area (both in size and number), and crop yield [27]. In adult plants, drought effects on shoot tissue expansion are generally more severe than in roots, which are less sensitive to water deficit stress [28]. Cellular dehydration inhibits photosynthesis under severe drought stress, resulting in reduced levels of carbon assimilation [29, 30]. Continued respiration under dry conditions can lead to a negative carbon balance in drought-affected plants [27]. This is due to the combined effects of decreased carbon assimilation overall and decreased translocation of photosynthates from leaves through phloem, which requires maintenance of cellular turgor [29]. Transpiration is inhibited in most plants under water deficit stress, due to stomatal closure and xylem cavitation, and this can lead to decreased nutrient uptake [27]. Prolonged periods of stomatal closure can also lead to heat stress (see Moisture Stress Tolerance Strategies below), as plants cannot employ transpirational cooling to reduce heat load. Reproductive structures, including both flowers and fruits, can also be severely affected by water deficit stress, due to reductions in the availability of photosynthates, inability to achieve turgor required for cellular expansion and tissue growth, and disruption in the activity of key enzymes [27]. Additional effects of water deficit have been described, but the above represent the most common impacts on plant growth and functioning.

Flooding primarily affects plants by reducing soil oxygen availability and therefore reducing root respiration [31]. Water replaces oxygen in soil pores, causing hypoxic or anoxic conditions that not only limit the diffusion of oxygen but also promotes the growth of anaerobic bacteria, which produce toxic compounds that could harm plants [29]. Many sensitive plants respond to waterlogged conditions by closing stomata, which reduces whole-plant water potential gradients and xylem transport [31, 32]. Further, waterlogged roots are unable to absorb nutrient ions, leading to nutrient deficiency symptoms in shoot tissue [29]. Similarly, the rate of photosynthate transport from leaves to roots can decrease by more than half within 30 min of flooding; this photosynthetic inhibition is not well understood, but it is thought to be linked to the toxic products of anaerobic respiration [32]. Plants sensitive to flooding are severely damaged within hours, with decreased growth, survival, and yield [29]. Plants with moderate flood tolerance are able to withstand anoxic conditions temporarily, although some can be damaged after multiple days of waterlogging. Strategies of highly flood-tolerant taxa will be discussed further below.

Moisture Stress Tolerance Strategies

True drought tolerance is the ability to grow, flower, and produce favorable yields under suboptimal water supply [27]. There are three major types of drought tolerance: drought escape, desiccation postponement or avoidance, and desiccation tolerance [29]. The life cycles of drought escapers are completed during wet seasons or while well watered [27, 29], and therefore, these plants do not necessarily possess physiological adaptations to drought stress. Desiccation postponers/ avoiders maintain tissue hydration under drought conditions, while plants tolerant of desiccation perform metabolic functions even while dehydrated [29]. Several morphological, molecular, and physiological mechanisms, including alteration of root/shoot ratios, osmotic adjustment, and production of dehydrin proteins, are important in drought tolerance and are covered elsewhere in comprehensive reviews [e.g., 33–37]. While we will not detail these mechanisms here, it is important to note that plants employing the three different photosynthetic pathways (C3, C4, and crassulacean acid metabolism (CAM)) show inherent drought tolerance and water use efficiency differences.

Stomatal closure helps to reduce the ratio of transpirationto-water uptake and delay dehydration [28]. Species with C4 and CAM photosynthetic pathways benefit from fixing a high rate of CO₂ with partially or fully closed stomata [38–40] and are generally considered more drought tolerant than most C3 species. Plants employing C4 photosynthesis also have higher water use efficiency (WUE), as well as increased biomass production in both ideal and drought conditions [41, 42]. The extensive root systems of perennial C4 grasses also maximize water uptake from the soil [43]. CAM plants take up CO₂ at night, making them better suited for chronically dry environments than C3 and C4 plants [44]. It has been estimated that CAM crops require only 20 % of the irrigation of the most efficient C3 and C4 crops [44].

Mechanisms for flood tolerance, including hormonal, molecular, and physiological mechanisms, have been reviewed in detail by a number of authors [e.g., 45–47]. Major morphological adaptations to flooding include production of aerenchyma tissue, enlargement of stem lenticels, and development of new roots [31]. Movement of oxygen to roots through aerenchyma not only allows root cells to reestablish aerobic respiration but also decreases toxic compounds outside of the root cortex through diffusion into soil and oxidation of reduced soil ions [32]. Formation of adventitous roots can compensate for decreased absorptive capacity or death and decay of existing roots and can lead to reopening of closed stomata [32].

Drought- and Flood-Tolerant Biomass Crops

As discussed previously, we searched for stress tolerance literature relating to several promising biomass crops being cultivated or developed in the USA currently. It is important to note that, as a consequence, our list of tolerant biomass crops should not be considered exhaustive. In addition, many of the available studies were designed to test relative tolerance among varieties or species. Thus, this analysis is not meant to provide absolutes or guarantee stress tolerance under every circumstance. However, our analysis still represents a more comprehensive review of multiple biomass crops and multiple stressors than currently exists. Table 1 details evidence of drought stress tolerance, and Table 2 details evidence of flooding tolerance in 17 biomass crops.

Of the 17 crops evaluated, six species were highly drought tolerant, and others had drought-tolerant species or genotypes

 Table 1
 Drought tolerance in 17 feedstocks with bioenergy potential

Taxa	Ps type	Tolerance level	Evidence of drought tolerance					
Agave spp.	CAM	High	Evolved in arid habitats [48] sunken stomata [48], elastic cells, shallow roots, osmotic adjustment, root isolation from dry soils [44]; CAM crops require 20 % of the irrigation of the most efficient C3 and C4 crops [44].					
Andropogon gerardii	C4	High	Greater allocation to roots, reduced allocation to flowering, more rapid leaf turnover, and more rapid recovery of photosynthesis after wilting, relative to <i>Sorghastrum nutans</i> [49]. Able to maintain carbon gain at lower water potentials than <i>Panicum virgatum</i> [50].					
Arundo donax	C3	Low	Not tolerant, but can survive drought [51, 52].					
Eucalyptus spp.	C3	High	Some taxa (e.g., <i>Eucalyptus rameliana</i> and <i>Eucalyptus pachyphylla</i>) occupy arid habitats with <350-mm annual precipitation [53]; Drought-tolerant genotypes of <i>Eucalyptus globulus</i> display multiple drought tolerance strategies, including more developed root system, smaller seeds with desiccation tolerance, great ABA content, and accumulation of proteins involved in stress tolerance [54].					
Helianthus annuus	C3	Nil	Not tolerant [55, 56], but certain cultivars and relatives could be used in breeding programs to increase tolerance (e.g., <i>Helianthus argophyllus</i> [57] and dwarf <i>Helianthus annuus</i> cultivars [55]).					
Jatropha curcas	C3	High	Drought avoidance through selective leaf abscission, decreases in Ps and WUE, maintaining above lethal water potential and ability to recover quickly [58].					
Miscanthus spp.	C4	Moderate	<i>Miscanthus</i> × <i>giganteus</i> leaf area and yield reduced under drought stress [59], but water availability does affect shoot production or plant height at the beginning of the growing season [60]. <i>Miscanthus sinensis</i> good drought tolerance [61] and could be included in future breeding programs to improve tolerance in <i>Miscanthus</i> hybrids [62].					
Panicum virgatum	C4	Moderate to high	Germination and biomass yield reduced under extreme drought [63], but after establishment, all plants survived at -4 MPa [64]. Generally very tolerant of moderate or even extreme drought [65] especial adequate rainfall occurs early in the growing season [66]. Upland cytotypes are more drought tolerar lowland types [67, 68].					
Pennisetum purpureum	C4	High	Requires good moisture during establishment, but acquires tolerance in later stages [69]; repeated drou stress did not affect biomass yields, but plant height was reduced after multiple drought treatments Several highly drought-tolerant genotypes could be used to further improve tolerance [71]					
Pinus spp.	C3	Varies	Drought-tolerant species: <i>Pinus bungeana</i> [72], <i>Pinus cembra</i> [72], <i>Pinus echinata</i> [72, 73], <i>Pinus elliottii</i> [72, 73] <i>Pinus flexilis</i> [72], <i>Pinus glabra</i> [73], <i>Pinus heldreichii</i> [72] <i>Pinus korariensis</i> [72], <i>Pinus mugo</i> [73], <i>Pinus nigra</i> [72], <i>Pinus palustris</i> [73], <i>Pinus rigida</i> [72], <i>Pinus sylvestris</i> [76, 77, <i>Pinus taeda</i> [72, 73] <i>Pinus thubergiana</i> [72], <i>Pinus virginiana</i> [73], <i>Pinus wallichiana</i> [72].					
Populus spp.	C3	Varies	<i>Populus euphratica</i> experiences drought-induced xylem cavitation [74], but calcium-dependent protein kinase confers drought tolerance [75]; transgenic <i>Populus deltoides</i> ("NL895") is drought tolerant [76]. Drought tolerance in <i>Populus nigra</i> varies among clones [77].					
Robinia pseudoacacia	C3	High	Listed as a drought-tolerant species in extension publications [76 77; tetraploid clones achieved greater biomass, WUE, and photosynthesis rate than common diploid clone [78]					
Saccharum spp.	C4	Varies	<i>Saccharum spontanaeum</i> cane yield, leaf area, plant height, and photosynthesis rate were reduced under drought [79], but genotypes Co 99004 and Co 99012 [79] and BOT-53, BOT-54, and BOT-6 [80] were more tolerant and productive; <i>Saccharum officinarum</i> root/shoot ratio, WUE, and rate of height growth were not affected by drought, but biomass, root factors, and stalk diameter were reduced. Genotypes 03-4-425 and Phill66-07 were more tolerant [81]. Transgenic sugarcane with introduction of AVP1 <i>Arabidopsis</i> gene was more tolerant [82].					
Salix spp.	C3	Varies	Salix cinerea [83], Salix elaeagnos [84], Salix gooddingii [85], and Salix matsudana "Navajo" [86] are more drought tolerant than other species.					
Sorghastrum nutans	C4	Varies	Sorghastrum nutans var. "Tejas" is better adapted for germination and early growth in dry conditions than "Cheyenne" or "Lometa" [87].					
Sorghum bicolor	C4	High	No negative effect on ethanol yield (actually increased EtOH yield if drought imposed during early flowering especially in hybrid DK-28E [88], drought tolerant through avoidance has deep and extensive root system—and true tolerance—osmotic adjustment allows stomata to stay open and metabolism to contin [89].					
Spartina pectinata	C4	Low	Not tolerant due to thin leaves, thin cuticle, and adaptation to wet conditions [90].					

within the genus (Table 1). Several of the most tolerant taxa were C4 perennial grasses, including *Andropogon gerardii*, *Pennisetum purpurem*, and *Sorghum bicolor*. For example, *Sorghum bicolor* is highly productive in dry African climates [109], likely due to the high WUE and large root systems found in many cultivars [110]. *Panicum virgatum* shows low

 Table 2
 Flooding tolerance in 17 feedstocks with bioenergy potential

Taxa Ps Toleranc type level			Evidence of flooding tolerance					
Agave spp.	CAM		N/A					
Andropogon gerardii	C4		N/A					
Arundo donax	C3	Moderate	Biomass decreased (~50 %) in flooded vs well-watered soil [51]; biomass and rhizome viability unaffected flooding [52].					
Eucalyptus spp.	C3	Varies	<i>Eucalyptus camaldulensis</i> is flood tolerant [53, 91, 92]; <i>Eucalyptus camaldulensis</i> > <i>Eucalyptus globulus</i> > <i>Eucalyptus obliqua</i> in level of flood tolerance [91], though even <i>Eucalyptus camaldulensis</i> showed some reductions in growth, biomass, and photoassimilate transport [92]; <i>Eucalyptus camaldulensis</i> requires seasonal flooding for regeneration [53].					
Helianthus annuus	C3		N/A					
Jatropha curcas	C3	Low	Flooding sensitive [93].					
Miscanthus spp.	C4	Moderate	Miscanthus × giganteus biomass and rhizome viability unaffected by flooding [52].					
Panicum virgatum	C4	Moderate	Seedlings emerged and established at all moisture conditions (-0.3 MPa to flooded). Transplants of lowland ecotypes performed as well in flooded conditions as in field capacity controls. Flooding reduced performance of upland ecotypes [64].					
Pennisetum purpureum	C4	Moderate	Compared with corn and mungbean (<i>Vigna radiata</i>), <i>Pennisetum purpureum</i> maintained higher stomatal and mesophyll conductance, allowing for continued CO2 assimilation under flood stress [94].					
Pinus spp.	C3	Varies	Almost 100 % of <i>Pinus elliotti</i> survived up to 40 days of flooding up to 60 cm [95]; <i>Pinus echinata, Pinus taeda, and Pinus rigida</i> var. <i>serotina</i> were all resistant to flooding. Standing water for 12 weeks produced slightly less growth relative to running water treatment and field capacity treatment [96]. Once established, <i>Pinus sylvestris</i> can withstand waterlogging for a long period (25 months in this study) [97].					
Populus spp.	C3	Varies	Populus deltoides × Populus nigra hybrid cv. "I-488" was more flood tolerant than other hybrids, especially with some pre-exposure to flooding [98]; Populus deltoides hybrid "Alton" was flood tolerant (least loss of leaf area and production of adventitious roots compared to other hybrids [99]. Populus deltoides "Alton" tolerates floods because stomatal conductance and root membrane integrity remain functional [100].					
Robinia pseudoacacia	C3	Low	Intolerant-did not survive continuous flooding during one growing season [101].					
Saccharum spp.	C4	Varies	Japanese sugarcane (<i>Saccharum</i> spp. var. "NiF 8") roots, leaves, stalks, sugar content (brix), and dry weight increased in response to flooding [102]; juice quality decreased from waterlogging and mean fiber content increased in waterlogged conditions. There were differences among sugarcane varieties, but sugar yield was reduced in waterlogged conditions across varieties [103].					
Salix spp.	C3	Varies	Salix petiolaris was least susceptible to flood-induced dieback, and Salix planifolia and Salix exigua were intermediately susceptible (Salix bebbiana and Salix discolor were most susceptible) [104]. In general, willow cover decreased on wetter transects over time (1993–2001), and increased on drier transects [104]. Salix elaeagnos established and survived in flooded conditions [84].					
Sorghastrum nutans	C4	Low	Compared with other native warm-season grasses tested and although pot studies suggested moderate flood tolerance, <i>Sorghastrum nutans</i> performance in flooded riparian sites was poor [109.					
Sorghum bicolor	C4	Moderate	Thirty days after seed germination, sorghum generally tolerates waterlogging (no effect on shoot growth). Some varieties form aerenchyma and adventitious roots [89].					
Spartina pectinata	C4	High	<i>Spartina pectinata</i> is flood tolerant, with little change in photosynthetic capacity in flooded conditions [105]. <i>Spartina pectinata</i> is the dominant species in low prairie where soils are too wet for switchgrass, maize and other grain, forage, and biofuel crops. Can also produce high biomass on well-drained soil on prime land and on coarse-textured soil on dry marginal land [106]. <i>Spartina pectinata</i> grew best under prolonged inundation (4 weeks), compared with alternating dry and wet conditions [107], and performed better than other warmseason grasses in riparian conditions [108].					

N/A no supporting evidence was found in literature databases

tolerance during establishment [64], possibly because of lower water use efficiency related to lower carbon assimilation during drought [111]. Although when drought was imposed after establishment, *Panicum virgatum* plants appeared to be drought hardy [64, 65]—particularly upland cytotypes [67, 68]. In addition, a long-term field evaluation of biomass crops

(34 herbaceous taxa assessed over 10 years across a wide range of soil and sites) indicated that *Panicum virgatum* and certain *Sorghum bicolor* varieties outperformed other crops under drought conditions [112]. As expected, the one CAM crop we included in our database, *Agave* spp., showed high drought tolerance due to several morphological and physiological adaptations to the arid climates in which it evolved [44, 48]. A number of woody crops were also drought tolerant, depending on genotype. These included several *Eucalyptus* species, *Jatropha curcas*, several *Pinus* species, *Populus* hybrids, *Robinia pseudoacacia*, and several *Salix* species (Table 1). Several studies identified particular genotypes that were more drought tolerant than the wild type (e.g., sugarcane genotype Co 99004) or identified congeners with higher tolerance than the target crop (e.g., *Helianthus argophyllus*) (Table 1). This information could potentially be used in future breeding programs to develop more tolerant biomass crops for marginal lands.

Several species and genotypes have evolved in wetland conditions and are moderately to strongly tolerant of flooded conditions (Table 2). For example, because Spartina pectinata is native to North American moist prairies, marshes, and drainage ways [113], it can be grown in soils that are too wet to grow corn, big bluestern, or switchgrass [114, 115], although lowland types of switchgrass are tolerant of flooded conditions [64, 68]. Other flood-tolerant crops include Eucalyptus camaldulensis, Miscanthus × giganteus, several hybrid Populus spp., Pinus elliotti, and Sorghum bicolor (Table 2). Eucalyptus camaldulensis formed adventitous roots and maintained moderate growth rates in flooded conditions [32], while the most tolerant Populus hybrids allocated more carbon to belowground structures, formed adventitious roots, and maintained stomatal function, net photosynthetic rate, and relative growth rate [116]. Again, novel genotypes of traditional crops showed greater flood tolerance than their parents, highlighting the potential for improvement in these and other taxa (Table 2). One example is a Japanese sugarcane (Saccharum spp.) hybrid which showed an increase in dry biomass in response to flooding. Continuous flooding is often deleterious to sugarcane growth [117], but because it is typically cultivated in wet tropical regions, it is important to develop varieties that are productive under flooded conditions.

Several of the focal biomass crops have not been the subject of study related to flooding tolerance (i.e., "N/A" in Table 2), but this lack of evidence should not be interpreted as flood sensitivity for these taxa.

Salt Stress

Framing the Issue

Salinity is a major environmental stressor affecting arid, semiarid, and irrigated land worldwide [118, 119] and contributing to the abandonment or marginalization of land [118]. Salinization can occur naturally, through aerosolization, deposition, or contact with sea salts in coastal locations [120], or through proximity to saline seeps, shallow water tables, and degradation of parent rock materials inland [121, 122]. Alternatively, so-called "secondary salinization" occurs anthropogenically, as a result of replacing deep-rooted native vegetation with shallow-rooted crops and pasture or from adding irrigation water to soils [120, 123]. Both actions can result in changes to water table depth, causing salts to accumulate in the root zone as excess water evaporates from the soil surface [120, 123]. While some dissolved salts can improve soil texture, an excess of salts, including sodium, can cause soil dispersion and reduced permeability [124]. Salt accumulation can render soil unsuitable for many traditional food crops by decreasing plant-available water and creating toxic cellular products [29].

More than 6 % of the global land area (>800 million hectares) [125] and at least 8.5 million hectare in the USA [124] are salt-affected. Secondary salinity, resulting from irrigation or land clearing, affects 20 [125] to 50 % [118, 126] of the irrigated land area globally. There is evidence that the land area affected by salinization is growing through anthropogenic causes [127] and due to changing hydrologic patterns related to climate change [128], resulting in an increasing proportion of marginal land. However, production of tolerant biomass crops on salt-affected soil could result in soil quality improvement and soil carbon sequestration [129].

Effects of Salt Stress

Effects of salinity on plant growth and physiology have been reviewed comprehensively elsewhere e. g [130-132], but common effects are discussed here. Under prolonged or severe salt stress, plants can experience negative developmental effects [126], from seed germination [133] and emergence [134] through maturation [135]. Dissolved salt ions in the soil solution can substantially reduce osmotic potential values (typical saline soil water potentials range between -1.6 and -10 MPa [136]), altering water potential gradients that drive water uptake and solute movement through plant tissues [133], resulting in decreased water uptake even when soils are wet. This functional reduction in water availability can lead to symptoms typical of drought-affected plants: reduced shoot and root growth rates, reduced leaf number, declines in stomatal conductance and photosynthesis rates, and damage or death of leaves [30, 130, 137-139]. In addition, salt ions can cause cellular toxicity, as well as disruption of normal membrane functioning, nutrient uptake, protein synthesis, and enzyme activation [29]. Secondary effects, including oxidative damage [140] and cell death, can also result from salt stress.

Salt Stress Tolerance Strategies

Halophytes ("salt-loving" plants) have specialized strategies for growth in saline conditions [130, 141–143]. These include succulence, which maintains water-to-salt ratios at acceptable levels as the overall cell volume increases, compartmentalization of salt ions into vacuoles or specialized salt glands on leaf surfaces, and efficient ion pumping mechanisms to exclude or remove ions from cytosol into plant apoplast [29]. However, halophytes are not the only plants that can tolerate salinity. Because saline soils are functionally similar to dry soils, plants with high water use efficiency (e.g., C4 and CAM plants) are predicted to perform well when exposed to salinity. In addition, many plants are capable of moderate levels of osmotic adjustment, in which salts accumulate in vacuoles to maintain cellular turgor and reestablish whole-plant water potential gradients. This adjustment takes place over a matter of hours to days [130], during which time growth is restricted and wilting may occur. More salt-tolerant plants go beyond compartmentalizing salts into vacuoles by excluding NaCl from xylem channels, actively exporting Na⁺ into the soil solution, and regulating K⁺ loss in cation channels [80]. Several comprehensive reviews offer additional information about salt tolerance mechanisms [e.g., [125, 144, 145].

Salt-Tolerant Biomass Crops

Some potential biomass crops, such as Pennisetum purpureum, show no more salinity tolerance than conventional agricultural crops [146, 147]. Thus, the current challenge is to find biomass crop species that can grow and maintain high yields on marginal salt-affected soils. Of the 17 crops evaluated, several species or genotypes were highly tolerant of salinity (Table 3). Deep-rooted perennial grasses are often recommended for drought- and salt-affected soils [180-183], and our literature search corroborated this recommendation. However, some of these grasses were more tolerant than others. For example, Andropogon gerardii, Arundo donax, and Spartina pectinata were highly salt tolerant, with the latter two classified as halophytes [152, 179]. In contrast, Pennisetum purpureum showed major reductions in shoot biomass in saline conditions [146], and $M. \times giganteus$ was only moderately salt tolerant [160]. Upland ecotypes of Panicum virgatum (e.g., "Blackwell," "Trailblazer," and "PV-1777") were among the more salt-tolerant cultivars [151, 164, 166, 184], although the upland ecotype, "Cave-in-Rock," was not tolerant at the seedling stage [167].

Several salt-tolerant woody crops are also available (Table 3). For example, *Eucalyptus camaldulensis* cultivars "Silverton" and "Local" efficiently excluded or compartmentalized salts in saline and saline + hypoxic conditions [154]. *Pinus pinea* showed no growth reduction under saline conditions [168], and *Pinus banksiana* growth may have been stimulated by certain levels of salinity [169]. Among tree crops, short-rotation woody crop (SRWC) species have particularly strong bioenergy potential because of fast growth and high yields [185–188]. Salt-tolerant SRWC species include several poplar (*Populus* spp.) and willow (*Salix* spp.) hybrids and tetraploid *Robinia pseudoacacia* (Table 3). These hybrids and others identified in Table 3 highlight the possibility of breeding salinity tolerance into many of the biomass crops destined for production on marginal lands.

Temperature Stress

Framing the Issue

Temperature is a major factor governing plant growth and biomass production [189], and temperature extremes can cause severe abiotic stress and inhibit plant growth. Climate comparisons between the most recent decade and historical climates indicate unmistakable and consistently warming surface temperatures on a global scale and throughout much of the USA; however, some small regions in the southern USA are now experiencing cooler than average trends [190]. Irrigation and soil amendments in traditional production systems may offset some of the negative effects of heat and cold temperatures [191]. In order to avoid heat-related crop damage, some growers may also opt to shift production to alternate regions where summer temperatures are milder. However, moving perennial crops to more northern locations will introduce more extreme winter weather to crops that may be adapted to mild winter climates. In addition, thermal climate changes may impact crops grown on marginal lands to a greater extent than prime agricultural land [192]. With greater temperature fluctuations and movement of crops outside of their traditional production regions, it will be important to develop biomass crops that can tolerate temperature extremes.

Effects of Temperature Stress

All plant species are adapted to a range of optimal temperatures, but when they are subjected to temperatures outside that range, physiological, metabolic, and molecular changes occur to maintain homeostasis under suboptimal conditions [193]. If the plant experiences suboptimal temperatures for an extended period, these processes become more impaired and abnormal until temperatures reach lethal levels [193]. Both low and high temperatures can cause physiological stress symptoms and physical damage in plants. Low-temperature stress can be caused by both freezing (temperatures less than -1 °C) and chilling (0–18 °C). The injuries caused by low temperatures for both freezing and chilling can be seen within 48 to 72 h and may include phenotypic changes (e.g., wilting, reduced leaf expansion, chlorosis, and necrosis) [193]. Reproductive processes and structures are also severely affected by cold, which can lead to pollen and flower sterility [193]. Likewise, exposure to cold in the germination and establishment phases can lead to low germination rate, stunting of seedling growth, chlorosis, and reduced tillering in grasses [193]. On a physiological level,

 Table 3
 Salinity tolerance in 17 feedstocks with bioenergy potential

Taxa	Ps type	Tolerance level	Evidence of salinity tolerance				
Agave spp.	CAM	Low, but variable	<i>Agave parry</i> i var. <i>truncata</i> fresh weight, dry weight, and moisture content decreased as salinity levels increased [148]. <i>Agave sisalana</i> shoot and root growth was reduced at higher salinity levels. Height was also reduced but not as much as in some other species. Characterized as intermediately tolerant [149]. <i>Agave deserti</i> seedlings very sensitive to salinity [150].				
Andropogon gerardii	C4	High	Andropogon gerardii had the highest germination rates under increased salinity levels, compared with othe C4 grasses [151].				
Arundo donax	C3	High	High resilience to high-salinity tannery effluent wastewater [152]; high biomass production when group former salt evaporation pond, and when irrigated with water 90 % the salinity of seawater [153]. Des <i>Arundo donax</i> as a halophyte [153].				
Eucalyptus spp.	C3	Varies	Twenty species can grow in saline conditions (e.g., <i>Eucalyptus robusta</i> and <i>Eucalyptus camaldulensis</i>) [<i>Eucalyptus camaldulensis</i> "Silverton" and "Local" did better in salinity and saline + hypoxic treatment than <i>Eucalyptus tereticornis</i> , which was sensitive to salinity and hypoxia. <i>Eucalyptus camaldulensis</i> "Silverton" and "Local" use different strategies, with "Silverton" using tissue compartmentalization an "Local" using tissue exclusion [154].				
Helianthus annuus	C3	Varies	Several genotypes, particularly cytoplasmic male sterile (CMS) lines, performed well in high-salinity conditions [155]; some genotypes show evidence of salinity avoidance by excluding salts at the root level while resistant lines change biomass partitioning patterns [156].				
Jatropha curcas	C3	Moderate	Characterized as moderately tolerant [157] at salinity levels up to 100 mM NaCl [158]; salinity alone wa tolerated by <i>Jatopha curcas</i> , but salinity (100 mM NaCl) + heat was harmful to CO ₂ assimilation and membrane functioning [159].				
Miscanthus spp.	C4	Moderate	Salinity above 100 mM affected <i>Miscanthus</i> × <i>giganteus</i> growth, with rhizomes > roots > shoots in order increasing sensitivity (rhizomes least sensitive). Plants grown from larger rhizomes initially were less sensitive [160]. Some accessions of <i>Miscanthus sacchariflorus</i> were highly salt tolerant during germina and could be used to improve future hybrids [161]. Salt tolerance during germination was better in <i>Miscanthus floridulus</i> collected from lowland locations in Taiwan, suggesting the possibility that salt-tolerant germplasm exists and could be used for future hybrids [162].				
Panicum virgatum	C4	Moderate	Salinity had no effect on germination and survival under low-alkaline pH, but when salinity was con with higher pH, germination and survival were strongly reduced [163]. <i>Panicum virgatum</i> is mod tolerant of saline conditions, and cultivar "PV-1777" had the highest salinity tolerance for upland e in one study [151], but in others, "Blackwell" [164, 165] and "Trailblazer" [166] performed well salinity conditions. Compared with <i>Spartina pectinata, Panicum virgatum</i> "Cave-in-Rock" had lo germination (down 80 %) in high (300 mM) salinity levels, and less than 70 % of seedlings survived moderate salinity (100 mM) treatments [167].				
Pennisetum purpureum	C4	Low	Salinity inhibits hybrid pennisetum (<i>Pennisetum americanum × Pennisetum purpureum</i>) growth, photosynthesis, soluble sugar content, and more, but adverse effects were reduced by applications of nitrat up to 5 mmol/L [147]. The same hybrid can exclude salt from new leaves, but salinity levels of 100 mM results in shoot fresh and dry weight reductions of 50 % compared to controls [146].				
Pinus spp.	C3	Varies	<i>Pinus pinea</i> showed no significant reduction in growth when grown in 100 mM NaCL hydroponic solution [168]. Of two <i>Picea</i> and one <i>Pinus</i> species tested, the pine (<i>Pinus banksiana</i>) was least affected by salinity during emergence and may have even been stimulated by certain levels of salinity [169].				
Populus spp.	C3	Varies (some high)	 Populus × xiaozhannica cv. "Balizhuangyang" has high tolerance [170]. With respect to salt tolerance, Populus euphratica is "outstanding," handling up to 450 mM NaCl [171]. Growth was unaffected in Populus euphratica at low-moderate salinity (68 mM), and at 137 mM NaCl, 50 % of Populus deltoids ? Populus alba "M31" and 100 % of Populus alba "GuadalquivirF-21-40," Populus alba "GuadalquivirF-21-39," Populus alba "GuadalquivirF-21-38," and Populus euphratica (100 %) survived [172] 				
Robinia pseudoacacia	C3	High (in 4n)	Tetraploid black locust can withstand high levels of NaCl and Na ₂ SO ₄ to a greater extent than diploid b locust (e.g., salt injury not observed, no change in water or chlorophyll content, or photosynthesis rate intercellular CO ₂ concentration in 4n). Potentially adaptive changes in leaf anatomy were seen in tetrap type in response to salinity [173], with the tetraploid version being much more adaptable to salt stress the diploid [174].				
Saccharum spp.	C4	Varies	Transgenic salt- and drought-tolerant sugarcane, with longer and more profuse roots and the ability to withstand higher NaCl, were developed by introducing the AVP1 gene from <i>Arabidopsis</i> [82], and sugarcane variety "CP-4333" had the greatest salt tolerance limit at 15.51 dS/m. Characteristics such as pink and waxy-coated stems, large number and area of green leaves, greater root and shoot yield, high-tillering, and ratooning potential revealed positive correlation with salt tolerance and could be used as markers in future breeding programs [175].				

Table 3 (continued)

Taxa	Ps type	Tolerance level	Evidence of salinity tolerance				
Salix spp.	C3	Moderate	Most willow varieties tested in this study were able to tolerate moderately saline conditions (EC(e)≤5 dS/m). In addition, several varieties ("Alpha," "India," "Owasco," "Tully Champion," and "01X-268-015") showed no reduction in growth with severe salinity (EC(e)≤8.0 dS/m) [176].				
Sorghastrum nutans	C4	Varies	<i>Sorghastrum nutans</i> var. "Tejas" seeds appeared adapted for optimum germination at higher salt concentrations than "Lometa" or "Cheyenne," but seedlings of those varieties produced greater root and shoot growth at higher salt concentrations than Tejas [87].				
Sorghum bicolor	C4	Varies	One hundred genotypes were screened, and seven were salinity tolerant to 250 mM NaCl: "CSV 15," "ICSB 766," "NTJ 2," "ICSV 95030," "S 35," "ICSB 589," "ICSB 676" [177]. Sweet sorghum "Keller" was the most salt tolerant of three cultivars tested, with little reduction in stem yield and soluble carbohydrates [178].				
Spartina pectinata	C4	High	<i>Spartina pectinata</i> seeds germinated and seedlings survived in high salinity conditions (up to 500 mM NaCl). Under all salinity treatments, cordgrass produced more tillers and greater biomass than switchgrass, by exuding salt through salt glands [167]. <i>Spartina pectinata</i> has a level of tolerance to soil salinity that is higher than that of other tall warm-season grasses [106], and that is similar to halophytes [179], with an ability to maintain growth in salinity levels ranging from 2–20 dS/m [180].				

this damage can be caused by disruption of membrane and organelle functioning as fluid phosolipids become crystalline and dysfunctional in colder temperatures [193]. In addition, ice crystals can form in apoplastic space, physically damaging cells and causing dehydration through the movement of water out of cells down water potential gradients to join the extracellular ice [193, 194]. Cold exposure can also affect photosynthetic functioning, enzymatic activity [195], protein mechanics, and other metabolic processes [193].

High temperatures can affect plants directly through growth inhibition and indirectly through evaporative water loss [196]. Sensitive species can be affected when air temperatures exceed 35 °C, but tolerant species can withstand air temperatures approaching 65 °C [29, 196]. Most plant species, however, cannot survive for extended periods above 45 °C [29]. As in chilling and freezing stress, membrane stability can be affected by heat. In the case of heat, however, membranes can become excessively fluid, causing ion leakage and inhibition of photosynthesis, respiration, and other processes that involve membrane-embedded proteins and electron carriers [29, 30]. High temperature damage to heat-sensitive photosynthetic components (e.g., chlorophyll, thylakoid membranes, and photosystem II) can significantly affect photosynthetic function [30]. Moderate heat stress can inhibit photosynthesis and thus decrease productivity and yields [197, 198]. Photosynthesis is affected before respiration for most plants, meaning that the production of sugars stops before the demand for them does. This can result in the breakdown of stored sugars (e.g., in fruits, leading to decreased sweetness) [29]. Further, heat stress can significantly reduce ethanol yield of some fuel crops [88]. Additional problems associated with, and responses to, thermal stress are reviewed in greater depth elsewhere [e.g., 196, 197, 199-203].

Temperature Stress Tolerance Strategies

Both cold and heat tolerance can be induced in most species through gradual exposure to non-lethal temperatures. Chilling-resistant species overcome membrane fluidity problems by increasing the proportion of unsaturated relative to saturated fatty acids in the membrane [29, 204, 205], lowering the temperature at which membranes solidify. In addition, sucrose and other soluble sugars accumulate in cells and cell walls to lower the temperature at which freezing can occur and to restrict the growth of ice [29]. Some species, particularly woody taxa, are able to avoid cellular freezing until temperatures dip to -40 °C through the mechanism known as "deep supercooling" [206, 207]. This occurs because of an absence of ice nucleation sites within cells, though ice may form in extracellular spaces. Freezing-resistant species produce antifreeze proteins that halt the growth and spread of ice crystals in extracellular spaces [207, 208].

When ample water is available, most plants are able to cool leaves through evaporative/transpirational cooling [197, 209]. However, when stomata close to prevent water loss in dry conditions, heat damage can occur. Plants adapted to hot climates have evolved morphological adaptations to minimize heat load, including pubescent, vertically oriented, or light-colored leaves [197, 209]. In many plants, increases in temperature initiate translation of heat shock proteins (HSPs), which serve to prevent and repair misfolding of other proteins and facilitate proper cellular functioning at high temperatures [200, 210]. At the whole-plant level, synthesis of HSPs increases tolerance of temperatures that could otherwise be lethal [29, 210]. Although HSPs protect cells against damage, the heat shock response increases the rate of maturation in crops and can decrease yields [211]. Further, the heat shock

Таха	Ps type	Tolerance level	Evidence of cold tolerance				
Agave spp.	CAM	Varies	Species native to higher elevations and latitudes where subfreezing temperatures regularly occur during winter include <i>Agave utahensis</i> , <i>Agave parryi</i> , <i>Agave havardiana</i> , <i>Agave neomexicana</i> , and <i>Agave lechuguilla</i> . These species or subspecies can survive with no discernible damage to -28 °C or lower [213].				
Andropogon gerardii	C4	Moderate	The base temp for germination in <i>Andropogon gerardii</i> was among the lowest for warm-season grasses tested, depending on cultivar. "Bison" had the lowest base temp of all species tested, at 2.6 °C, but "Niagara" was one of the higher ones at 5.0 °C. Bison showed some chilling sensitivity symptoms (purpling and partial wilting) during cold exposure, but electrolyte damage was lower for Bison than other cultivars [214].				
Arundo donax	C3	Moderate	Cold resistance of five grasses evaluated was, in order: <i>Arundo donax</i> (-21.3 °C) > <i>Arundo</i> sp. (-12.05 °C <i>Echinochloa crusgalli</i> (-1.98 °C) > <i>Pennisetum purpureum</i> (0.69 °C) > <i>Pennisetum</i> sp. (0.18 °C). Study evaluated semi-lethal temperatures (LT 50) [215].				
Eucalyptus spp.	C3	Varies	Some small trees or shrubs (e.g., <i>Eucalyptus coccifera</i> or <i>Eucalyptus pauciflora</i> ssp. <i>niphophila</i>) are adapted to subalpine conditions [53]. <i>Eucalyptus occidentalis</i> showed low cold tolerance, with foliage death due to frost occurring at -4 °C [216] A freeze-tolerance gene has been introduced into <i>Eucalyptus urograndis</i> elite clone EH1, and resulting trees are tolerant to freezing temps to -8 °C [185].				
Helianthus annuus	C3	Low	Carbon assimilate translocation was inhibited in cold (13 °C) grown sunflower, and photosynthesis rate was lower than in warm (30 °C) grown sunflowers [217]. Eighteen germplasm lines were found to be frost resistant, and two (NDCMS-1B and NDLR-2) escaped frost damage by flowering early [218].				
Jatropha curcas	C3	Low	Very low tolerance [157], but chilling tolerance can be induced in seedlings if exposed to a 5 °C chilling shock followed by a recovery period at optimal temperatures (26 °C) [219].				
Miscanthus spp.	C4	Moderate	The lethal temperature at which 50 % (LT50) of <i>Miscanthus</i> × <i>giganteus</i> rhizomes were killed was -3.4 which can be problematic especially during first winter. In <i>Miscanthus sinensis</i> , LT50 was -6.5 °C [2] <i>Miscanthus</i> × <i>giganteus</i> shows unusual cold tolerance for a C ₄ species [60]. <i>Miscanthus sinensis</i> grow where Tmin is down to -11 °C [221].				
Panicum virgatum	C4	Moderate	The base temp for germination in <i>Panicum virgatum</i> was among the lowest for warm-season grasses tested, depending on cultivar. "Dakota" had one of the lowest base temperatures of all grasses, at 2.79 °C, but the other three switchgrass cultivars ranged from 4.5 to 7.3 °C with Cave-in-Rock (CIR) the highest. No symptoms were seen during chilling treatment, but some after recovery. Leaf damage ranged from 40 to 56 % across cultivars, with CIR having the highest leaf area damage of all warm-season grasses tested. Electrolyte leakage was lowest in "Dakota" compared with all other grasses tested, and CIR was close to the highest [214]. Lowland cytotypes are particularly susceptible to cold winter conditions, as they are adapted to southern latitudes [222, 223]. <i>Panicum virgatum</i> shows unusual tolerance to cold night temps for a C ₄ grass [224].				
Pennisetum purpureum	C4	Low	Growth rate and productivity of <i>Pennisetum purpureum</i> was greater than corn at chilling temps, but chilling reduced leaf extension, leaf area, and chlorophyll content in <i>Pennisetum purpureum</i> . Roots were more resistant to chilling than shoots [225]. The semi-lethal (LT50) temperature for <i>Pennisetum</i> spp. (0.18 °C) was the highest of all grasses tested [215].				
Pinus spp.	C3	High	Some pines are frost hardy to -70 °C, including <i>Pinus sylvestris</i> [226]. Conifers are among the most cold-tolerant of vascular plants, with twigs of some pine species withstanding temps as low as -196 °C [202].				
Populus spp.	C3	High	A calcium-dependent protein kinase gene in <i>Populus euphratica</i> confers drought and cold stress tolerance [75]. <i>Populus deltoides</i> ssp. <i>monilifera</i> survived cooling to -70 °C [227].				
Robinia pseudoacacia	C3	High	Withstands cold and freezing temperatures by increasing fatty acid concentration [228] and protein synthesis in bark cells [229, 230], and produces glycoproteins to prevent ice crystal formation in cells. Hungarian cultivars "Penzesdombi" and "Kiscsalai" are comparatively frost tolerant [231, 232]. Stem dieback has been reported in cold conditions, and frost can decrease growth rate and height.				
Saccharum spp.	C4	Varies	Greatest ration cold tolerance was identified in <i>Saccharum spontanaeum</i> genotypes IND 81–144, IND 81–80, IND 81–165, and MPTH 97–216, and these were more tolerant than the most tolerant commercial variety [233].				
Salix spp.	C3	High	Salix matsudana, especially variety "Navajo," is extremely cold hardy [86]. With pretreatment (hardening), tropical willows were able to withstand cold temperatures (to -30 °C), while a northern willow (<i>Salix sieboldiana</i>), was able to withstand temperatures of -50 °C and survived immersion in liquid N at -196 °C after hardening for 2 weeks in cold temperatures [234].				
Sorghastrum nutans	C4	Moderate	The base temperature for germination for <i>Sorghastrum nutans</i> was low to midrange (2.8 to 4.5 °C) among the warm-season grasses tested. Chilling symptoms were seen during chilling treatment for "Tomahawk," and in "Holt" after temperatures returned to normal. Leaf damage ranged from 30 % in Tomahawk (the lowest across all grasses) to 41 % for "Holt." Electrolyte leakage was among the lowest in "Tomahawk" for all grasses [214]. <i>Sorghastrum nutans</i> is not particularly cold tolerant, but var. "Lometa" had greater percent germination (24 %) at the low temperature treatment (5–15 °C) than other varieties (7–17 %) [87].				

 Table 4
 Cold tolerance in 17 feedstocks with bioenergy potential. N/A indicates no supporting evidence was found in literature databases

Table 4 (continued)						
Taxa	Ps type	Tolerance level	Evidence of cold tolerance			
Sorghum bicolor	C4	Low	<i>Sorghum bicolor</i> is sensitive to cold stress at all stages of development and typically is planted 3–5 weeks later than other annual crops to avoid inhibition of germination, emergence, and crop establishment [235]. Chilling (2–8 °C) for 1 to 8 days inhibited <i>Sorghum bicolor</i> growth and nitrogen uptake during exposure, and the ability to recover was greater in warmer and shorter chilling treatments [236]. Eight advanced breeding lines and one recombinant inbred line showed early emergence, higher biomass (30 days after emergence), and relatively earlier flowering than other lines under cold temperatures (14 °C) [237].			
Spartina pectinata	C4	High	<i>Spartina pectinata</i> is known to have one of the most northerly distributions among C4 grasses [238]. Natural populations are found in the boreal forest of NW Canada where July mean minimum temperatures were greater than 7.5 °C [239], and cultivar CWNC was recommended for production in Canada due to early spring growth [240]. <i>Spartina pectinata</i> cells showed relatively limited injury under freezing conditions, possibly because, as a salt-marsh grass, it is salt tolerant and this may confer greater inherent cell freezing resistance [241].			

response can halt the synthesis of other proteins [210] and cause oxidative stress [212]. Many heat-tolerant plants are able to maintain higher photosynthetic rates and membrane stability by increasing the proportion of saturated and monounsaturated fatty acids and maintain overall tissue water balance through osmotic adjustment [197]. A number of additional physiological changes occur in response to heat in tolerant taxa, including hormonal changes, increases in protective pigments, and synthesis of secondary metabolites. These are detailed, along with molecular tolerance mechanisms, in several comprehensive reviews [e.g., 196, 197, 199–201].

Heat- and Cold-Tolerant Biomass Crops

Several woody biomass crops are naturally cold tolerant (Table 4), as many of them evolved in cold climates, including alpine or boreal ecosystems. For example, the phenomenon of cellular "supercooling" is common among conifer species, and some Pinus species can survive temperatures as low as -196 °C [202]! Other cold-hardy woody crops include Populus [75, 227] and Salix spp. [86, 234] (Table 4), with Robinia pseudoacacia introduced into several Canadian provinces (http://plants.usda.gov/core/profile?symbol=rops). Moreover, in unpublished University of Illinois research, several hundred black locust genotypes survived the abnormally cold 2013-2014 winter in Urbana, IL (40.0645 N, -88.2078 W), when average January and February temperatures were 5.5 and 5.7 °C lower than 30-year averages; furthermore, all trees grew productively the following season (T. Voigt, personal observation). Several herbaceous biomass crops tolerate cold conditions, as well. These include Spartina pectinata [239, 241], as well as Andropogon gerardii cv. "Bison" [214], Panicum virgatum cv. "Dakota" [214] and other upland cytotypes [222], Miscanthus sinensis [242] and, to a lesser extent, $M. \times giganteus$ [60] (Table 4). Commonly cultivated Panicum virgatum cultivars "Alamo," "Cave-inRock," and "Kanlow" were sensitive or moderately sensitive to low temperatures [243]. Although many *Agave* species are associated with warm desert ecosystems, a number of *Agave* spp. that evolved in high elevations (e.g., *Agave utahensis* and *Agave parryi*) are able to withstand temperatures down to – 28 °C. In addition, cold-tolerant genotypes of subtropical and tropical biomass crops *Eucalyptus* spp. and *Saccharum* spp. have been developed (Table 4), indicating the possibility for cold tolerance to be improved in future breeding programs for these and other crops.

Because C4 and CAM species have inherent mechanisms to resist heat stress, it makes sense to consider biomass crops with these photosynthetic pathways (see Table 5). Agave species (CAM) can withstand temperatures between 57 and 65 °C because of thick cuticle, low absorbance of short-wave radiation, and deployment of heat shock proteins [244, 260]. A number of C4 and highly efficient C3 perennial grasses are heat tolerant, including Andropogon gerardii, Arundo donax, Miscanthus sinensis, and some Sorghastrum nutans and Saccharum varieties (Table 5). Panicum virgatum cultivars exhibit variable heat tolerance, with lowland cytotypes generally performing better in warm, southern climates [222]. For example, several Panicum virgatum genotypes show intermediate (cv. "Alamo" and others) to high tolerance (cv. "Summer") to heat stress during germination [243] and thus may be good candidates for production on marginal lands in warm regions. Commonly grown cultivars "Cave-in-Rock" and "Kanlow" were heat sensitive [243]. Heat-tolerant woody species include Jatropha curcas, and Eucalyptus occidentalis and others, Pinus densiflora and others, Populus euphratica, Robinia pseudoacacia, and Salix nigra (Table 5). Many of these crops evolved in hot climates, but others have been improved through breeding for greater heat tolerance. For example, a heat tolerant Saccharum spp. (CP-4333) has been developed and shows rapid recovery following heat stress [256 Table 5].

Taxa	Ps type	Tolerance level	Evidence of heat tolerance				
Agave spp.	CAM	High	<i>Agave americana</i> produced the greatest levels of heat shock proteins in response to heat stress compared to three other moderately and highly heat tolerant species, protecting photosynthetic functioning [244].				
Andropogon gerardii	C4	High	Andropogon gerardii photosynthetic components can tolerate high (35 °C+) temperatures, particularly under elevated CO ₂ [245].				
Arundo donax	C3	High	<i>Arundo donax</i> rhizome fragments can produce shoots and roots in controlled conditions up to 41 ° (R. Tayyar, L. Quinn, and J. Holt, unpublished data).				
Eucalyptus spp.	C3	High	Several species (including <i>Eucalyptus deglupta</i> , <i>Eucalyptus pellita</i> , <i>Eucalyptus occidentalis</i> , and <i>Eucalyptus urophylla</i>) appear to be adapted to hot environments, based on conditions typical in the native range [53, 216]. <i>Eucalyptus occidentalis</i> leaves resist heat damage to 51.8 °C [216].				
Helianthus annuus	C3	Low	High leaf temperatures (40–45 °C) were detrimental to physiological traits including photosyntheti rate, transpiration rate, and stomatal conductance [246]. Some genotypes can reduce heat load b changing leaf inclination, with the most tolerant genotypes angling leaves upward. These were able to maintain lower leaf temperatures and membrane leakage [247].				
Jatropha curcas	C3	High	<i>Jatropha curcas</i> has a high tolerance for heat [157]. A period of heat treatment was favorable for young <i>Jatropha curcas</i> plants, unless heat was combined with salinity [159].				
Miscanthus spp.	C4	Moderate	Heat shock genes have been identified in <i>Miscanthus sinensis</i> and could be used to improve future hybrids [248].				
Panicum virgatum	C4	Moderate	Lowland cytotypes of <i>Panicum virgatum</i> , adapted to southern climates, may be more heat tolerant than upland types [223]. Even though plant height and total biomass decreased under heat stress [249], a climate modeling paper shows that <i>Panicum virgatum</i> yields could increase under warmer climate scenarios (3 to 8 °C), due to extended growing season and limited cold stress [211].				
Pennisetum	C4		N/A				
purpureum Pinus spp.	C3	High	Pinus sylvestris (20-year trees) increased in diameter earlier in the season and stopped growing la when exposed to elevated temperatures (compared with ambient temperatures), and diameter v 26 % greater in elevated temperatures vs ambient temperatures over the 3-year study [250]. Elevated temperatures did not significantly alter net photosynthesis across the native range of <i>Pinus taeda</i> [251]. <i>Pinus densiflora</i> relative growth rate and dry matter yield increased in responted to higher temps (30 vs 25 °C) [252].				
Populus spp.	C3	Varies	<i>Populus euphratica</i> is tolerant to extreme temperatures, via proteins related to lipid biogenesis, cytoskeleton structure, sulfate assimilation, thiamine and hydrophobic amino acid biosynthesis, and nuclear transport. Photosynthesis is maintained by decreasing photosystem (PSII) abundance and increasing PSI contribution to linear electron flow [253].				
Robinia pseudoacacia	C3	Moderate	Virginia Tech extension publication lists black locust as a heat tolerant tree, but recommends caution because of invasive tendencies [254]. Stem dieback was associated with hot, dry conditions in Oklahoma plantings [255].				
Saccharum spp.	C4	Varies	A heat-tolerant variety of sugarcane (CP-4333) recovered more quickly from heat stress than a heat-sensitive variety, due to leaf rolling (decreased water loss) and rapid reversal of this effect during recovery [256].				
Salix spp.	C3	Varies	Salix arctica responded negatively to simulated heat waves and did not recover its cold tolerance when normal (arctic) temperatures were reimposed [257]. Salix phylicifolia, which naturally occurs in or near natural hot springs in Iceland, had higher photosynthesis rates in hotter soils than in cool soils away from the hot springs [258]. Salix nigra plants treated with 40 °C hot water showed higher values both for photosynthesis rates and stomatal conductance than untreated plants [259].				
Sorghastrum nutans	C4	Moderate	<i>Sorghastrum nutans</i> var. "Llano" and "Lometa" had a higher percent germination at the high temperature treatment (30–40 °C) than other varieties. Optimal temperatures for these were 10–30 °C ("Llano") and 15–30 °C ("Lometa") [87].				
Sorghum bicolor	C4	Low	Heat stress significantly reduced glucose release and EtOH yield from hybrid DK-28E, especially during seed-filling stages [88].				
Spartina pectinata	C4	Moderate	Relative <i>Spartina alterniflora</i> heat shock protein production in response to heat stress increased photosynthetic thermotolerance [244].				

 $N\!/\!A$ no supporting evidence was found in literature databases

Additional Considerations in Evaluating Stress-Tolerant Biomass Crops

Multiple Stressors

While research into genetically modifying biofuel crops to enhance abiotic stress tolerance may expand the area suitable for cultivation [64], breeders will need to anticipate the combined stressors that are likely to occur in many marginal production systems. Different combinations of stressors may cause conflicting responses [261], but there are species that are well adapted to multiple stressors. For example, xerohalophytes are specialized halophytes (salt-tolerant species) that are found in dry conditions (e.g., Salsola kali [262]). Conversely, most true halophytes are adapted to wetlands and therefore have adaptations to withstand inundated and saline soils (e.g., Spartina alterniflora [263]). Some of these may be suitable for improvement as energy crops on marginal land. In contrast, multiple stressors often cause damage to growing plants, even if the plant is tolerant of a particular type of environmental stress. For example, Jatropha curcas, a highly heat-tolerant species, suffers more from the combination of salinity and heat stress than from either stressor alone [159]. Therefore, it will be important to identify the prevailing stressor(s) in the marginal area under production and choose the most tolerant biomass crops. It will also be important for producers to be aware of the possibility of reduced yield even among the most tolerant crops in years when multiple stressors occur (e.g., low rainfall years in saline conditions). Further, this review focused on a narrow, but physiologically important, set of stressors. We acknowledge that

 Table 6
 Summary table of all species and all stressors

additional stress factors will influence biomass crop productivity on marginal land. Some of these, like nutrient deficiencies, can be ameliorated with available agronomic management practices.

Several of the crops we have highlighted are suitable for a number of stressful conditions. These will be discussed further in the "Conclusions" section.

Invasiveness

The ability to produce high biomass yields under unfavorable growing conditions is correlated with invasiveness, and several authors have cautioned against the use of non-native and potentially invasive biomass crops [264-270]. Some of the crops mentioned in this review have been evaluated as highrisk species and have received attention from environmental groups and invasion ecologists (e.g., Arundo donax, seedbearing Miscanthus spp., Jatropha curcas, Pennisetum purpureum) [265, 267, 268, 271-275]. Therefore, these and other high invasion-risk crops should only be chosen when they can be grown and transported with strict containment procedures in place [276] and when state and federal regulations allow their introduction and cultivation [277, 278]. Other crops in this review, however, are either US natives or have been evaluated as low-risk for invasion in the USA [279]. The authors encourage the choice of native biomass crops, but note that most "native" species are only native to a specific region of the USA and can, in fact, be "weedy" or invasive outside of the native range (e.g., Robinia pseudoacacia, which is native to small areas in the Piedmont and Missouri regions, but has invaded and naturalized throughout the continental USA)

Таха	Ps type	Drought	Flooding	Salinity	Heat	Cold
Agave spp.	CAM	High	No data	Varies	High	Varies
Andropogon gerardii	C4	High	No data	High	High	Moderate
Arundo donax	C3	Low	Moderate	High	High	Moderate
Eucalyptus spp.	C3	High	Varies	Varies	High	Varies
Helianthus annuus	C3	Nil	No data	Varies	Low	No data
Jatropha curcas	C3	High	Low	Moderate	High	Low
Miscanthus spp.	C4	Moderate	Moderate	Moderate	Moderate	Moderate
Panicum virgatum	C4	Low	Moderate	Moderate	Moderate	Moderate
Pennisetum purpureum	C4	High	No data	Low	No data	Low
Pinus spp.	C3	Varies	Varies	Varies	High	High
Populus spp.	C3	Varies	Varies	Varies	Varies	High
Robinia pseudoacacia	C3	High	No data	High (in 4n)	High	No data
Saccharum spp.	C4	Varies	Varies	Varies	Varies	Moderate
Salix spp.	C3	Varies	Varies	Moderate	Varies	High
Sorghastrum nutans	C4	Varies	No data	Varies	Moderate	Moderate
Sorghum bicolor	C4	High	Moderate	Varies	Low	No data
Spartina pectinata	C4	Low	High	High	Moderate	High

[273]. In addition, some native species can be pests within their native regions. For example, *Helianthus annuus* is native to the entire continental USA but is a regulated noxious weed in Iowa due to its negative impacts on agriculture [277]. As such, it will be important for producers to choose biomass crops that are native and/or low-risk in the production region [279]. Growers can consult online databases to determine invasion risk [280–282] or choose from a recently released list of low-risk biomass crops [279].

Conclusions

Predicted climate changes will increase the likelihood of abiotic stress throughout the country, including various combinations of multiple stressors. For example, heat waves are predicted to become more intense throughout the country while precipitation is expected to increase in the northern USA and to decrease in the southwest [11]. Our literature review has revealed several "all purpose" biomass crops that are moderately or highly tolerant of multiple environmental stressors (Table 6). For example, Andropogon gerardii, Eucalyptus spp., Miscanthus spp., Panicum virgatum, Pinus spp., Populus spp., Robinia pseudoacacia, and Spartina pectinata were shown to be moderately or highly tolerant of four or more stress types. For particular growing conditions such as some hot and dry areas, growers could choose among Agave americana, Andropogon gerardii, Jatropha curcas, Miscanthus sinensis, Pinus sylvestris, Pinus taeda, Populus euphratica, or Robinia pseudoacacia. Many wet and saline environments could likely support Arundo donax, Eucalyptus camaldulensis (particularly Eucalyptus camaldulensis "Silverton" and "Local"), Miscanthus × giganteus, Panicum virgatum "Trailblazer," Sorghum bicolor varieties, and Spartina pectinata. Genera such as Pinus and Populus comprise a host of species that are adaptable to different stressors and combinations of stressors.

As previously mentioned, our list is not exhaustive and, in some cases, is based on studies that assessed relative—not absolute—stress tolerance, but it represents a much more comprehensive biomass crop selection guide for growers than currently exists. Based on this review, growers could choose from a variety of plant types representing a variety of industrial uses from ethanol (e.g., *Miscanthus* spp.) to combustion (e.g., *Pinus* spp.), depending on their preferences and the capabilities of local processing plants. In addition, we have indicated here that a number of biomass crops have already been improved for greater stress tolerance, and we assume that breeding programs will continue to develop additional stress-tolerant crops. Therefore, it appears that there will be a number of options available for marginal lands now and into the increasingly stressful future [105].

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References

- United States Congress (2007) Energy Independence and Security Act of 2007. Title II - Subtitle A - Renewable Fuel Standard. https:// www.govtrack.us/congress/bills/110/hr6/text. Accessed 23 Dec 14
- FAO (1997) Report of the study on CGIAR research priorities for marginal lands. Consultative Group on International Agricultural Research, Food and Agriculture Organization of the United Nations; Available at: http://www.fao.org/Wairdocs/TAC/X5784E/ x5784e02.htm. Accessed 18 Jun 14
- 3. Tilman D et al (2009) Beneficial biofuels—the food, energy, and environment trilemma. Science 325:270
- Milbrandt A, Overend RP (2009) Assessment of biomass resources from marginal lands in APEC economies. Energy Working Group, NREL/TP-6A2-46209. http://www.nrel.gov/docs/fy10osti/46209. pdf. Accessed 18 Jun 14
- 5. Cai X, Zhang X, Wang D (2011) Land availability for biofuel production. Environ Sci Technol 45:334
- Campbell JE, Lobell DB, Genova RC, Field CB (2008) The global potential of bioenergy on abandoned agriculture lands. Environ Sci Technol 42:5791–5794. doi:10.1021/es800052w
- 7. Shortall OK (2013) "Marginal land" for energy crops: exploring definitions and embedded assumptions. Energ Policy 62:19
- Richards BK, Stoof CR, Cary I, Woodbury PB (2014) Reporting on marginal lands for bioenergy feedstock production—a modest proposal. Bioenerg Res 7:1060
- Blake A, Clarke M, Stuart-Street A (2012) Resource management technical report 379: changing land use on unproductive soils N. E. A. R. S. P. 3, Ed., (Department of Agriculture and Food)
- Sivakumar MVK, Stefanski R (2007) In: MVK Sivakumar, N Ndiangui (eds) Climate and land degradation.p 105–135
- Hatfield J, et al (2014) Agriculture. In: JM Melillo, TC Richmond, GW Yohe (eds) Climate change impacts in the United States: the third national climate assessment. U.S. Global Change Research Program, Washington, D.C, p 150–174
- Schlenker W, Roberts MJ (2009) Nonlinear temperature effects indicate severe damages to U.S. crop yields under climate change. P Natl Acad Sci USA
- Lobell DB, Schlenker W, Costa-Roberts J (2011) Climate trends and global crop production since 1980. Science 333, 616
- 14. Oliver RJ, Finch JW, Taylor G (2009) Second generation bioenergy crops and climate change: a review of the effects of elevated atmospheric CO2 and drought on water use and the implications for yield. GCB Bioenerg 1:97
- U.S. Department of Energy. (2011) Billion-Ton Update: Biomass Supply for a Bioenergy and Bioproducts Industry. Page 227 pp in R. D. Perlack and B. J. Stokes, editors. ORNL/TM-2011/224. Oak Ridge National Laboratory, Oak Ridge, TN
- 16. Begg JE, Turner NC (1976) Crop water deficits. Adv Agron 28:161
- USDA NRCS (2014) Prime and other important farmlands definitions. http://www.nrcs.usda.gov/wps/portal/nrcs/detail/pr/soils/? cid=nrcs141p2 037285. Accessed 18 Jun 14
- Hale MG, Orcutt DM (1987) The physiology of plants under stress. John Wiley & Sons Chichester, UK

- Bates BC, Kundzewicz ZW, Wu S, Palutikof JP (2008) Climate change and water (technical paper of the intergovernmental panel on climate change). IPCC Secretariat, Geneva
- Cook ER, Woodhouse CA, Eakin CM, Meko DM, Stahle DW (2004) Long-term aridity changes in the western United States. Science 306, 1015
- Woodhouse CA, Meko DM, MacDonald GM, Stahle DW, Cook ER (2010) A 1,200-year perspective of 21st century drought in southwestern North America. P Natl Acad Sci USA 107:21283
- Zhang J (2004) Risk assessment of drought disaster in the maizegrowing region of Songliao Plain, China. Agr Ecosys Environ 102:133
- Gopalkrishnan G, Negri MC, Snyder SW (2011) A novel framework to classify marginal land for sustainable biomass feedstock production. J Environ Qual 40:1593
- 24. Lee D, Owens VN, Boe A, Koo B-C (2009) Biomass and seed yields of big bluestem, switchgrass, and intermediate wheatgrass in response to manure and harvest timing at two topographic positions. GCB Bioenerg 1:171
- USDA NRCS (2014) Wetlands http://www.nrcs.usda.gov/wps/ portal/nrcs/main/national/water/wetlands/. Accessed 7 May 14
- Rosenzweig C, Tubiello FN, Goldberg R, Mills E, Bloomfield J (2002) Increased crop damage in the US from excess precipitation under climate change. Glob Environ Chang 12:197
- Farooq M, Wahid A, Kobayashi N, Fujita D, Basra SMA (2009) In: Lichtfouse E, Navarrete M, Debaeke P, Véronique S, Alberola C (eds) Sustainable agriculture. Springer, Netherlands, pp 153–188
- Maroco JP, Pereira JS, Manuela Chaves M (2000) Growth, photosynthesis and water-use efficiency of two C4 Sahelian grasses subjected to water deficits. J Arid Environ 45:119
- 29. Taiz L, Zeiger E (2006) Plant physiology, 4th edn. Sinauer, Sunderland, MA
- Ashraf M, Harris PJC (2013) Photosynthesis under stressful environments: an overview. Photosynthetica 51:163
- Caudle KL, Maricle BR (2012) Effects of flooding on photosynthesis, chlorophyll fluorescence, and oxygen stress in plants of varying flooding tolerance. Trans Kansas Acad Sci 115:5
- Kozlowski TT (1984) Plant-responses to flooding of soil. Bioscience 34:162
- Hanin M et al (2011) Plant dehydrins and stress tolerance: versatile proteins for complex mechanisms. Plant Signal Behav 6:1503
- Bray EA (1993) Molecular responses to water deficit. Plant Physiol 103:1035
- Ingram J, Bartels D (1996) The molecular basis of dehydration tolerance in plants. Annu Rev Plant Physio 47:377
- Ludlow MM, Muchow RC (1990) A critical evaluation of traits for improving crop yields in water-limited environments. Adv Agron 43:107
- Turner N, Jones M (1980) In: NC Turner, PJ Kramer (eds) Adaptation of plants to water and high temperature stress. Wiley, New York, p 87–103
- Vogel K, Jung H (2001) Genetic modification of herbaceous plants for Feed and Fuel. Crit Rev Plant Sci 20:15
- Long SP, ZHU XG, Naidu SL, Ort DR (2006) Can improvement in photosynthesis increase crop yields? Plant Cell Environ 29:315
- 40. Kern M (2002) Food, feed, fibre, fuel and industrial products of the future: challenges and opportunities Understanding the strategic potential of plant genetic engineering. J Agron Crop Sci 188:291
- Waller S, Lewis J (1979) Occurrence of C3 and C4 photosynthetic pathways in North American grasses. J Range Manage 32:12–28. doi:10.2307/3897378
- 42. Simacek T (2008) Growing alternative energy crops in West Central Minnesota. (University of Minnesota West Central Research and Outreach Center, Morris, MN, 2008). http:// renewables.morris.umn.edu/biomass/documents/Simacek-GrowingAlternativeEnergyCropsInWestCentralMinnesotaPaper. pdf. Accessed 18 Jun 14

- Xu Q, Huang B (2001) Morphological and physiological characteristics associated with heat tolerance in creeping bentgrass. Crop Sci. doi:10.2135/cropsci2001.411127x
- 44. Borland AM, Griffiths H, Hartwell J, Smith JAC (2009) Exploiting the potential of plants with crassulacean acid metabolism for bioenergy production on marginal lands. J Exp Bot 60: 2879–2896
- Armstrong W, Brandle R, Jackson MB (1994) Mechanisms of flood tolerance in plants. Acta Bot Neerl 43:307
- Blom CWPM, Voesenek LACJ (1996) Flooding: the survival strategies of plants. Trends Ecol Evol 11:290–295
- Pradhan C, Mohanty M (2013) Submergence Stress: Responses and adaptations in crop plants. In: GR Rout, AB Das, (eds) Molecular stress physiology of plants, p 331–357. Springer, India
- Davis SC, Dohleman FG, Long SP (2011) The global potential for Agave as a biofuel feedstock. GCB Bioenerg 3:68
- Swemmer AM, Knapp AK, SmithMD (2006) Growth responses of Two dominant C4 grass species to altered water availability. Int J Plant Sci 167:1001
- Knapp AK (1985) Effect of fire and drought on the ecophysiology of *Andropogon gerardii* and *Panicum virgatum* in a tall grass prairie. Ecology 66:1309
- Lambert AM, Dudley TL, Robbins J (2014) Nutrient enrichment and soil conditions drive productivity in the large-statured invasive grass Arundo donax. Aquat Bot 112:16
- 52. Mann J, Kyser G, DiTomaso J, Barney J (2013) Assessment of above and belowground vegetative fragments as propagules in the bioenergy crops *Arundo donax* and *Miscanthus* × giganteus. Bioenerg Res 6:688
- Rejmanek M, Richardson DM (2010) In: Simberloff D, Rejmanek M (eds) Encyclopedia of biological invasions. University of California Press, Los Angeles
- Valdes AE et al (2013) Drought tolerance acquisition in *Eucalyptus globulus* (Labill.): a research on plant morphology, physiology and proteomics. J Proteomics 79:263–276
- 55. Hussain S, Ali A, Ibrahim M, Saleem MF, Bukhsh M (2012) Exogenous application of abscisic acid for drought tolerance in sunflower (*Helianthus annuus* L.): a review. J Anim Plant Sci 22: 806–826
- 56. Sobrado MA, Turner NC (1986) Photosynthesis Dry matter accumulation and distribution in the wild sunflower helianthus petiolaris and the cultivated sunflower helianthus annuus as influenced by water deficits. Oecologia 69:181
- Milton EF, Goolsby EW, Donovan LA (2013) Cultivated *Helianthus annuus* differs from two wild relatives in germination response to simulated drought stress. Helia 36:35
- Fini A, Bellasio C, Pollastri S, Tattini M, Ferrini F (2013) Water relations, growth, and leaf gas exchange as affected by water stress in *Jatropha curcas*. J Arid Environ 89:21
- Clifton-Brown JC, Lewandowski I (2000) Water use efficiency and biomass partitioning of three different *Miscanthus* genotypes with limited and unlimited water supply. Ann Bot 86:191
- Zub HW, Brancourt-Hulmel M (2010) Agronomic and physiological performances of different species of *Miscanthus*, a major energy crop. Agron Sustain Dev 30:201
- Chen M, Hou X, Fan X, Wu J, Pan Y (2013) Drought tolerance analysis of *Miscanthus sinensis* 'Gracillimu' seedlings. Acta Prataculturae Sinica 22:184–189
- 62. Stewart JR et al (2009) The ecology and agronomy of *Miscanthus sinensis*, a species important to bioenergy crop development, in its native range in Japan: a review. GCB Bioenerg 1:126
- 63. Tucker SS, Craine JM, Nippert JB (2011) Physiological drought tolerance and the structuring of tallgrass prairie assemblages. Ecosphere 2:art. 48
- 64. Barney JN et al (2009) Tolerance of switchgrass to extreme soil moisture stress: ecological implications. Plant Sci 177:724

- Sanderson MA et al (1999) Switchgrass cultivars and germplasm for biomass feedstock production in Texas. Bioresource Technol 67: 209–219
- Lee DK, Boe A (2005) Biomass production of switchgrass in central South Dakota. Crop Sci 45:2583
- 67. Stroup JA, Sanderson MA, Muir JP, McFarland MJ, Reed RL (2003) Comparison of growth and performance in upland and lowland switchgrass types to water and nitrogen stress. Bioresource Technol 86:65
- Porter CL (1966) An analysis of variation between upland and lowland switchgrass *Panicum virgatum* L. in Central Oklahoma. Ecology 47:980–992
- 69. Hegarty A (1957) Elephant grass Queensland Agric J 83:119
- Purbajanti ED, Anwar S (2012) Wydiati, F. Kusmiyati, Drought stress effect on morphology characters, water use efficiency, growth and yield of guinea and napier grasses. Int Res J Plant Sci 3:47
- Ponsens J, Basweti E, Hanson J (2010) Paper presented at the Tropentag 2010: International Research on Food Security, Natural Resource Management, and Rural Development, Zurich
- 72. Evans E (2014) Drought tolerant trees. NC State University Horticultural Sciences Cooperative Extension publication. http:// www.ces.ncsu.edu/depts/hort/consumer/quickref/trees/ droughttolerant.html. Accessed 23 Apr 14
- Coder KD (1999) Tree selection for drought resistance. The University of Georgia School of Forest Resources Extension Publication FOR 99-008. 4/1999. http://warnell.forestry.uga.edu/ SERVICE/LIBRARY/for99-008/for99-008.pdf. Accessed 23 Apr 14
- 74. Hukin D, Cochard H, Dreyer E, Le Thiec D, Bogeat-Triboulot MB (2005) Cavitation vulnerability in roots and shoots: does *Populus euphratica* Oliv., a poplar from arid areas of Central Asia, differ from other poplar species? J Exp Bot 56:2003
- 75. Chen JH, Xue B, Xia XL, Yin WL (2013) A novel calciumdependent protein kinase gene from *Populus euphratica*, confers both drought and cold stress tolerance. Biochem Bioph Res Co 441:630
- 76. Zhang L et al (2013) Transgenic poplar "NL895" expressing CpFATB gene shows enhanced tolerance to drought stress. Acta Physiol Plant 35:603
- Regier N et al (2009) Drought tolerance of two black poplar (*Populus nigra* L.) clones: contribution of carbohydrates and oxidative stress defence. Plant Cell Environ 32:1724–1736
- Zhang Z, Sheng Y-B, Luo X-F (2010) Effects of water stress on biomass and photosynthetic characteristics of tetraploid black locust (*Robinia pseudoacacia* L.) clones. For Res 23:920
- Gomathi R, Vasantha S, Alarmelu A, Shanthi RM (2011) Field evaluation of elite sugarcane clones (*Saccharum officinarum* L.) for drought tolerance. J Sugarcane Res 1:55–62
- Munawarti A, Taryono, Semiarti E, Holford P, Sismindari (2013) Tolerance of accessions of glagah (*Saccharum spontaneum*) to drought stress and their accumulation of proline. Am J Agricultural Biol Sci 8:1–11
- Jangpromma N, Thammasirirak S, Jaisil P, Songsri P (2012) Effects of drought and recovery from drought stress on above ground and root growth, and water use efficiency in sugarcane (Saccharum officinarum L.). Aust J Crop Sci 6
- Kumar T, Uzma, Khan MR, Abbas Z, Ali GM (2014) Genetic improvement of sugarcane for drought and salinity stress tolerance using arabidopsis vacuolar pyrophosphatase (AVP1) gene. Mol Biotechnol 56:199–209
- Zhivotovsky OP, Kuzovkina YA (2010) Response of two Salix L. species to water deficit. J Environ Horti 28:63
- 84. Francis RA, Gurnell AM, Petts GE, Edwards PJ (2005) Survival and growth responses of Populus nigra *Salix elaeagnos* and *Alnus incana* cuttings to varying levels of hydric stress. For Ecol Manag 210:291

- Stella JC, Battles JJ (2010) How do riparian woody seedlings survive seasonal drought? Oecologia 164:579
- Martin C (2014) Virtual library of Phoenix landscape plants: Salix matsudana. http://www.public.asu.edu/~camartin/plants/Plant html files/salixmatsudana.html. Accessed 30 Apr 2014
- Fulbright TE (1988) Effects of temperature, water potential and sodium-chloride on indiangrass germination. J Range Manage 41: 207
- Ananda N, Vadlani PV, Prasad P (2011) Evaluation of drought and heat stressed grain sorghum (*Sorghum bicolor*) for ethanol production. Ind Crop Prod 33:779
- Tari I, Laskay G, Takacs Z, Poor P (2013) Response of sorghum to abiotic stresses: a review. J Agron Crop Sci 199:264
- 90. Maricle BR, Koteyeva NK, Voznesenskaya EV, Thomasson JR, Edwards GE (2009) Diversity in leaf anatomy, and stomatal distribution and conductance, between salt marsh and freshwater species in the C-4 genus Spartina (Poaceae). New Phytol 184:216
- Blake T, Reid D (1981) Ethylene Water relations and tolerance to waterlogging of three *Eucalyptus* species. Funct plant biol 8:497
- 92. Kogawara S, Yamanoshita T, Norisada M, Masumori M, Kojima K (2006) Photosynthesis and photoassimilate transport during root hypoxia in *Melaleuca cajuputi*, a flood-tolerant species, and in *Eucalyptus camaldulensis*, a moderately flood-tolerant species. Tree Physiol 26:1413
- Gimeno V et al (2012) Physiological and morphological responses to flooding with fresh or saline water in *Jatropha curcas*. Environ Exp Bot 78:47
- 94. Araki T, Mai Thi Phuong N, Kubota F (2012) Specific feature in photosynthetic response of kenaf (*Hibiscus cannabinus* L.) to flooding stress. Environ Control Biol 50:127–134
- 95. Shi W et al (2010) Comparative test of flood tolerance of *Pinus elliottii* and other 4 trees in the Taihu Lake beach. Jiangsu For Scie Technol 37
- Hunt FM (1951) Effects of flooded soil on growth of pine seedlings. Plant Physiol 26:363–368
- Mukassabi TA, Polwart A, Coleshaw T, Thomas PA (2012) How long can young Scots pine seedlings survive waterlogging? Trees-Struct Func 26:1641
- Bejaoui Z, Albouchi A, Lamhamedi MS, Abassi M, El Aouni MH (2012) Adaptation and morpho-physiology of three *Populus deltoides* Marsh. x *P-nigra* L. clones after preconditioning to prolonged waterlogging. Agroforest Sys 86:433
- Luquez VMC, Achinelli FG, Cortizo S (2012) Evaluation of flooding tolerance in cuttings of Populus clones used for forestation at the Paraná River Delta, Argentina. South Forests. doi:10.2989/ 20702620.2012.686214
- 100. Rodriguez ME, Doffo G, Achinelli F, Bartoli C, Luquez VMC (2011) Characterization of responses to flooding and post flooding recovery in two *Populus deltoides* clones: physiological and biochemical aspects. BMC Proceedings. doi:10.1186/1753-6561-5-S7-P94
- Broadfoot WM, Williston HL (1973) Flooding effects on southern forests. J Forest 71:584–587
- 102. Tetsushi H, Karim MA (2007) Flooding tolerance of sugarcane in relation to growth, physiology, and root structure. South Pacific Stud 28:9
- 103. Rinki K, Singh VP, Navnit K (2013) Effect of waterlogging on quality constraints of sugarcane varieties. Environ Ecol 31
- 104. Timoney KP, Argus G (2006) Willows, water regime, and recent cover change in the Peace–Athabasca Delta. Ecoscience 13:308
- 105. Waring EF, Maricle BR (2012) Photosynthetic variation and carbon isotope discrimination in invasive wetland grasses in response to flooding. Environ Exp Bot 77:77
- 106. Lee DK, Boe A, Owens V, Gonzalez-Hernandez J, Rayburn AL (2011) Developing prairie cord grass (*Spartina pectinata*) as a new bioenergy crop. Aspect Appl Biol 197–201

- 107. Miller RC, Zedler JB (2003) Responses of native and invasive wetland plants to hydroperiod and water depth. Plant Ecol 167:57
- Skinner RH, Zobel RW, van der Grinten M, Skaradek W (2009) Evaluation of native warm-season grass cultivars for riparian zones. J Soil Water Conserv 64:413
- Rooney WL, Blumenthal J, Bean B, Mullet JE (2007) Designing sorghum as a dedicated bioenergy feedstock. Biofuels Bioprod Biorefin 1(147)
- 110. Zegada-Lizarazu W, Zatta A, Monti A (2012) Water uptake efficiency and above- and belowground biomass development of sweet sorghum and maize under different water regimes. Plant Soil 351:47–60
- 111. Wagle P, Kakani VG (2014) Growing season variability in evapotranspiration, ecosystem water use efficiency, and energy partitioning in switchgrass. Ecohydrology 7:64
- 112. Wright L, Turhollow A (2010) Switchgrass selection as a model bioenergy crop: a history of the process. Biomass Bioenerg 34:851
- 113. Barkworth ME, Anderton LK, Capels KM, Long S, Piep MB (2007) Manual of grasses for North America. Utah State University Press, Logan
- 114. Weaver JE, Fitzpatrick T (1932) Ecology and relative importance of the dominants of tall-grass prairie. Bot Gaz. doi:10.1086/334244
- 115. Weaver JE (1954) North American prairie. Johnson, Lincoln, NE
- 116. Guo XY, Huang ZY, Xu AC, Zhang XS (2011) A comparison of physiological, morphological and growth responses of 13 hybrid poplar clones to flooding. Forestry 84:1
- 117. Ray JD, Sinclair TR, Glaz B (2010) Sugarcane response to high water tables and intermittent flooding. J of Crop Improv 24
- 118. Flowers TJ, Yeo AR (1995) Breeding for salinity resistance in crop plants-where next? Aust J Plant Physiol 22:875
- 119. Bot AJ, Nachtergaele FO, Young A (2000) Land resource potential and constraints at regional and country levels (Food and Agricultural Organization (FAO) of the United Nations, Rome). ftp://ftp.fao.org/agl/agll/docs/wsr.pdf. Accessed 19 Jun 14
- 120. Sim L (2012) In: Lawn J (ed) A guide to managing and restoring wetlands in Western Australia. Department of Environment and Conservation, Perth
- Lobell D et al (2010) Regional-scale assessment of soil salinity in the Red River Valley using multi-year MODIS EVI and NDVI. J Environ Qual 39:35
- 122. Tober D, Duckwitz W, Sieler S (2007) Plant materials for saltaffected sites in the northern Great Plains (USDA-NRCS Plant Materials Center, Bismarck, ND). http://www.nrcs.usda.gov/ Internet/FSE_PLANTMATERIALS/publications/ndpmctn7094. pdf. Accessed 23 Jun 14
- 123. Abrol IP, Yadav JSP, Massoud FI (1988) Salt-affected soils and their management. FAO Soils Bulletin 39. Food and Agriculture Organization of the United Nations, Rome, 1988, vol. 39
- 124. Pearson K (2009) The basics of salinity and sodicity effects on soil physical properties. Montana State University Extension Service. http://waterquality.montana.edu/docs/methane/basics_highlight. shtml . Accessed 23 Dec 14
- 125. Munns R, Tester M (2008) Mechanisms of Salinity Tolerance. Annu Rev Plant Biol 59:651
- 126. Pitman MG, Läuchli A (2002) Global impact of salinity and agricultural ecosystems. In: A Lauchli, U Luttge, (eds) Salinity: environment-plants-molecules. Springer Netherlands, p 3–20.
- 127. Ghassemi F, Jakeman AJ, Nix HA (1995) Salinisation of land and water resources: human causes, extent, management and case stud ies (UNSW Press, Sydney, Australia, and CAB International, Wallingford, UK)
- 128. Varallyay G (2010) The impact of climate change on soils and on their water management. Agron Res 8:385
- 129. Wicke B et al (2011) The global technical and economic potential of bioenergy from salt-affected soils. Energ Environ Sci 4:2669
- Munns R (2002) Comparative physiology of salt and water stress. Plant Cell Environ 25:239

- Bernstein L (1975) Effects of Salinity and Sodicity on Plant Growth. Annu Rev Phytopathol 13:295
- 132. Sudhir P, Murthy SDS (2004) Effects of salt stress on basic processes of photosynthesis. Photosynthetica 42:481–486
- 133. Almansouri M, Kinet JM, Lutts S (2001) Effect of salt and osmotic stresses on germination in durum wheat (*Triticum durum* Desf.). Plant Soil 231:243
- 134. Khajeh-Hosseini M, Powell A, Bingham I (2003) The interaction between salinity stress and seed vigour during germination of soyabean seeds. Seed Sci Technol 31:715
- 135. Adam P (1990) Saltmarsh ecology. Cambridge University Press, Cambridge
- 136. Ungar IA (1977) The relationship between soil water potential and plant water potential in two inland halophytes under field conditions. Bot Gaz 138:498
- 137. Wang Y, Nii N (2000) Changes in chlorophyll, ribulose bisphosphate carboxylase-oxygenase, glycine betaine content, photosynthesis and transpiration in Amaranthus tricolor leaves during salt stress. J Hortic Sci Biotech 75:623
- Rivelli AR, Lovelli S, Perniola M (2002) Effects of salinity on gas exchange, water relations and growth of sunflower (Helianthus annuus). Funct Plant Biol 29:1405
- Kerstiens G, Tych W, Robinson MF, Mansfield TA (2002) Sodium related partial stomatal closure and salt tolerance of *Aster tripolium*. New Phytol 153:509
- 140. Gossett DR, Millhollon EP, Lucas M (1994) Antioxidant response to NaCl stress in salt-tolerant and salt-sensitive cultivars of cotton. Crop Sci 34:706
- 141. Flowers T, Troke P, Yeo A (1977) The mechanism of salt tolerance in halophytes. Annu Rev Plant Physio 28:89
- 142. Ashraf M, Wu L (1994) Breeding for salinity tolerance in plants. Crit Rev Plant Sci 13:17
- 143. Glenn EP, Brown JJ, Blumwald E (1999) Salt tolerance and crop potential of halophytes. Crit Rev Plant Sci 18:227
- 144. Greenway H, Munns R (1980) Mechanisms of salt tolerance in nonhalophytes. Annu Rev Plant Physio 31:149
- Maas EV, Hoffman GJ (1977) Crop salt tolerance—current assessment. J Irrig Drain Div–ASCE 103:115–134
- 146. Wang D, Yuan F, Wang B-S, Chen M (2012) Response of plant biofuel hybrid Pennisetum to NaCl stress and its salinity threshold. Chinese J of Plant Ecol 36:572
- 147. Chen M et al (2013) Elevated N supply reduces the inhibition of growth and photosynthesis caused by salinity in the bioenergy plant hybrid Pennisetum. In: AWang, LK Che, R Dong, G Zhao (eds) Applied energy technology, pts 1 and 2, vol. 724–725. Trans Tech, Stafa-Zurich, p 443–451
- Schuch UK, Kelly JJ (2007) Salinity tolerance of cacti and succulents. Hortscie 42:940
- El-Bagoury OH, El-Agroudy MH, Shenouda MA (1993) Effect of salinity levels on growth of six plant species. Egypt J Agron 18:129
- Nobel PS, BerryWL (1985) Element responses of agaves.AmJ Bot 72:686
- 151. Schmer MR, Xue Q, Hendrickson JR (2012) Salinity effects on perennial, warm-season (C-4) grass germination adapted to the northern Great Plains. Can J Plant Sci 92:873
- 152. Calheiros CSC et al (2012) Toxicity of high salinity tannery wastewater and effects on constructed wetland plants. Int J Phytoremediation 14:669
- 153. Williams CMJ et al (2009) Use of poor quality water to produce high biomass yields of giant reed (Arundo donax L.) on marginal lands for biofuel or pulp/paper. In: H Jaenicke, J Ganry, I Hoeschle-Zeledon, R Kahane (eds) Acta horticulturae
- 154. Nasim M et al (2009) Different eucalyptus species show different mechanisms of tolerance to salinity and salinity x hypoxia. J Plant Nutr 32:1427

- 155. Nagarathna TK, Shadakshari YG, Puttarangaswami KT (2012) Evaluation of different sunflower (*Helianthus annuus* L.) genotypes for sodium chloride induced salinity. Asian J Bio Sci 7:182–184
- 156. Rauf S, Shahzad M, Teixeira da Silva JA, Noorka IR (2012) Biomass partitioning and genetic analyses of salinity tolerance in sunflower (*Helianthus annuus* L.). J Crop Sci Biotechnol 15
- 157. Garcia-Almodovar C et al (2014) Improving the tolerance of *Jatropha curcas* L. plants to abiotic stresses. CAB Reviews 9:0–10
- 158. HishidaMet al (2013) Differential responses of Jatropha species on growth and physiological parameters to salinity stress at seedling plant stage. Commun Soil Sci Plant Anal 44:2820
- 159. Silva EN et al (2013) Contrasting physiological responses of *Jatropha curcas* plants to single and combined stresses of salinity and heat. J Plant Growth Regul 32:159
- 160. Plazek A et al (2014) Tolerance of Miscanthus x giganteus to salinity depends on initial weight of rhizomes as well as high accumulation of potassium and proline in leaves. Ind Crops Prod 52:278
- 161. Zong J, Gao Y, Chen J, Nie D, Liu J (2013) Assessment of salinity tolerance of *Miscanthus scchariflora* germplasm during germination period. Acta Agrestia Sinica 21
- 162. Hsu FH (1990) Effects of salt stress on germination of *Miscanthus* species and the physiological response to salt stress. J Taiwan Livest Res 23
- 163. Liu Yet al (2014) Synergistic and antagonistic effects of salinity and pH on germination in switchgrass (*Panicum virgatum* L.). Plos One. doi:10.1371/journal.pone.0085282
- 164. Harper J, Spooner AE (1983) Establishment of selected herbaceous species on acid bauxite minesoils. In: DH Graves (ed) 1983 Symposium on surface mining, hydrology, sedimentology and reclamation. Lexington, KY, pp. 413–417
- 165. Rodgers CS, Anderson RC (1995) Plant growth inhibition by soluble salts in sewage sludge-amended mine spoils. J Environ Qual 24: 627
- 166. Carson MA, Morris AN (2012) Germination of Panicum virgatum cultivars in a NaCl gradient. Bios 83:90
- 167. Kim S, Rayburn AL, Voigt T, Parrish A, Lee DK (2012) Salinity effects on germination and plant growth of prairie cordgrass and switchgrass. Bioenerg Res 5:225–235
- 168. Khaldi A, Ben Ammar R, Woo SY, Akrimi N, Zid E (2011) Salinity tolerance of hydroponically grown *Pinus pinea* L. seedlings. Acta Physiol Plantarum 33:765
- 169. Croser C, Renault S, Franklin J, Zwiazek J (2001) The effect of salinity on the emergence and seedling growth of *Picea mariana*, *Picea glauca*, and *Pinus banksiana*. Environ Pollut 115:9
- 170. Yin J et al (2004) Field test of saline resistant transgenic *Populus* × xiaozhannica cv. "Balizhuangyang". J Northeast For Uni 32:23–25
- 171. Chen S, Polle A (2010) Salinity tolerance of Populus. Plant Biol 12: 317
- 172. Sixto H, Grau JM, Alba N, Alia R (2005) Response to sodium chloride in different species and clones of genus *Populus* L. Forest 78:93
- 173. Meng F-J, Wang Q-Y, Wang J-Z, Li S-Y, Wang J-J (2008) Salt resistance of tetraploid *Robinia pseudoacacia*. Zhiwu Shengtai Xuebao 32:654
- 174. Wang ZM, Wang MY, Liu L, Meng FJ (2013) Physiological and proteomic responses of diploid and tetraploid black locust (*Robinia pseudoacacia* L.) subjected to salt stress. Int J Mol Sci 14:20299
- 175. Wahid A, Rao AUR, Rasul E (1997) Identification of salt tolerance traits in sugarcane lines. Field Crop Res 54:9
- 176. Hangs RD, Schoenau JJ, Van Rees KCJ, Steppuhn H (2011) Examining the salt tolerance of willow (*Salix* spp.) bioenergy species for use on salt-affected agricultural lands. Can J Plant Sci 91:509
- 177. Krishnamurthy L, Serraj R, Hash CT, Dakheel AJ, Reddy BVS (2007) Screening sorghum genotypes for salinity tolerant biomass production. Euphytica 156:15

- 178. Almodares A, Hadi MR, Ahmadpour H (2008) Sorghum stem yield and soluble carbohydrates under different salinity levels. Afr J Biotechnol 7:4051
- 179. Warren RS, Baird LM, Thompson AK (1985) Salt tolerance in cultured-cells of spartina-pectinata. Plant Cell Rep 4:84
- 180. Montemayor MB, Price JS, Rochefort L, Boudreau S (2008) Temporal variations and spatial patterns in saline and waterlogged peat fields: 1. Survival and growth of salt marsh graminoids. Environ Exp Bot 62:333
- Alderson J, Sharp WC (1995) Grass varieties in the United States. CRC, Boca Raton
- 182. Lewandowski I, Scurlock JMO, Lindvall E, Christou M (2003) The development and current status of perennial rhizomatous grasses as energy crops in the US and Europe. Biomass Bioenerg 25:335
- 183. Chen Z et al (2007) Potassium and sodium relations in salinised barley tissues as a basis of differential salt tolerance. Funct Plant Biol 34:150
- 184. Rodgers HL, Day FP, Atkinson R (2004) Root dynamics in restored and naturally regenerated Atlantic white cedar wetlands. Restor Ecol 12:401
- 185. Hinchee M et al (2009) Freeze-tolerant *Eucalyptus* as a renewable feedstock for industrial applications. In Vitro Cell Dev Biol— Animal 45:S24
- 186. Johnson JM et al (2007) Biomass-bioenergy crops in the United States: a changing paradigm. Am J Plant Sci Biotech 1:1–28
- 187. Rédei K, Osváth-Bujtás Z, Veperdi I (2008) Black locust (*Robinia pseudoacacia* L.) improvement in Hungary: a review. Acta Silv Lign Hung 4:127–132
- Nieminen K, Robischon M, Immanen J, Helariutta Y (2012) Towards optimizing wood development in bioenergy trees. New Phytologist 194:46
- 189. Boyer JS (1982) Plant productivity and environment. Science 218:443
- 190. Walsh J et al (2014) Climate Change Impacts in the United States: The Third National Climate Assessment, J. M. Melillo, T. C. Richmond, G. W. Yohe, Eds. (U.S. Global Change Research Program, 2014), pp. 19–67
- 191. Evans LT (1996) Crop evolution, adaptation and yield. Cambridge University Press, Cambridge
- 192. Jarvis A, Ramirez J, Anderson B, Leibing C, Aggarwal PK (2010) Scenarios of climate change within the context of agriculture. In: MP Reynolds (ed) Climate change and crop production. CAB International, Wallingford UK, p 9–37
- 193. Yadav SK (2010) Cold stress tolerance mechanisms in plants. A review Agronomy for Sust Dev 30:515
- 194. Bracale M, Coraggio I (2003) Cellular responses and molecular strategies for the adaptation to chilling and freezing stresses in plants. In: LS Di Toppi, B Pawlik Skowronska (eds) Abiotic stresses in plants. Kluwer, Boston, MA
- 195. Smallwood M, Bowles DJ (2002) Plants in a cold climate. Philos Trans R Soc Lond B Biol Sci 357:831
- 196. Hall AE (2010) Crop responses to environment. CRC, Boca Raton, FL
- 197. Bita CE, Gerats T (2013) Plant tolerance to high temperature in a changing environment: scientific fundamentals and production of heat stress-tolerant crops. Front in Plant Sci 4:273
- 198. Carmo-Silva AE, Salvucci ME (2012) The temperature response of CO2 assimilation, photochemical activities and Rubisco activation in Camelina sativa, a potential bioenergy crop with limited capacity for acclimation to heat stress. Planta 236:1433
- Wahid A, Gelani S, AshrafM, FooladMR (2007) Heat tolerance in plants: an overview. Environ Exp Bot 61:199–223
- 200. Kotak S et al (2007) Complexity of the heat stress response in plants. Curr Opin Plant Biol 10:310–316
- SungD-Y, Kaplan F, Lee K-J, Guy CL (2003) Acquired tolerance to temperature extremes. Trends Plant Sci 8:179–187
- 202. Sakai A, Larcher W (1987) Frost survival of plants. Responses and adaptation to freezing stress. Springer, New York, NY

- Thomashow MF (1999) Plant cold acclimation: freezing tolerance genes and regulatory mechanisms. Annu Rev Plant Physiol Plant Mol Biol 50:571
- 204. Williams JP, Khan MU, Mitchell K, Johnson G (1988) The effect of temperature on the level and biosynthesis of unsaturated fatty acids in diacylglycerols of Brassica napus leaves. Plant Physiol 87:904
- 205. Palta JP, Whitaker BD, Weiss LS (1993) Plasma membrane lipids associated with genetic variability in freezing tolerance and cold acclimation of Solanum species. Plant Physiol 103:793
- 206. Burke M, Gusta L, Quamme H, Weiser C, Li P (1976) Freezing and injury in plants. Annu Rev Plant Physiol 27:507
- 207. Wisniewski M, Gusta L, Neuner G (2014) Adaptive mechanisms of freeze avoidance in plants: a brief update. Environ Exp Bot 99: 133–140
- Marentes E, Griffith M, Mlynarz A, Brush RA (1993) Proteins accumulate in the apoplast of winter rye leaves during cold-acclimation. Physiol Plant 87:499
- Barbour MG, Burk JH, Pitts WD (1987) Terrestrial plant ecology. Benjamin/Cummings, Menlo Park
- Vierling E (1991) The roles of heat shock proteins in plants. Annu Rev Plant Biol 42:579
- 211. Brown RA, Rosenberg NJ, Hays CJ, Easterling WE, Mearns LO (2000) Potential production and environmental effects of switchgrass and traditional crops under current and greenhouse-altered climate in the central United States: a simulation study. Agr Ecosyst Environ 78:31
- 212. Panchuk II, Volkov RA, Schöffl F (2002) Heat stress-and heat shock transcription factor-dependent expression and activity of ascorbate peroxidase in Arabidopsis. Plant Physiol 129:838
- 213. Irish M (2002) Growing agaves in cold climates. Cactus Succ J 74
- 214. Madakadze IC, Stewart KA, Madakadze RM, Smith DL (2003) Base temperatures for seedling growth and their correlation with chilling sensitivity for warm-season grasses. Crop Sci 43:874
- 215. Lin X et al (2013) Cold-tolerance of 5 species of Juncao under low temperature stress. Acta Prataculturae Sinica 22
- Zohar Y, Waisel Y, Karschon R (1981) Heat and cold resistance of eucalyptus-occidentalis endl leaves and its relationship to soil-water conditions. Aust J Ecol 6:79
- 217. Paul MJ, Lawlor DW, Driscoll SP (1990) The effect of temperature on photosynthesis and carbon fluxes in sunflower and rape. J Exp Bot 41:547
- 218. Bajaj RK, Dhillon SK (2008) Evaluation of sunflower germplasm for frost resistance. Crop Improv 35:55
- 219. Li ZG, Yuan LX, Wang QL, Ding ZL, Dong CY (2013) Combined action of antioxidant defense system and osmolytes in chilling shock-induced chilling tolerance in *Jatropha curcas* seedlings. Acta Physiol Plant 35:2127
- 220. Clifton-Brown JC, Lewandowski I (2000) Overwintering problems of newly established *Miscanthus* plantations can be overcome by identifying genotypes with improved rhizome cold tolerance. New Phytologist 148:287
- 221. Quinn L D et al (2011) Environmental tolerances of *Miscanthus* sinensis in invasive and native populations. BioEnergy Research. doi:10.1007/s12155-011-9163-1
- 222. Casler MD, Vogel KP, Taliaferro CM, Wynia RL (2004) Latitudinal adaptation of switchgrass populations. Crop Sci 44:293
- Casler MD et al (2007) Latitudinal and longitudinal adaptation of switchgrass populations. Crop Sci 47:2249
- 224. Ku SB, Edwards GE, Smith D (1978) Photosynthesis and nonstructural carbohydrate concentration in leaf blades of panicum-virgatum as affected by night temperature. Can J Bot 56:63
- 225. Al-Shoaibi AA (2007) Growth response of elephant grass (*Pennisetum purpureum*) and *Zea mays* to chilling temperature. Bioscie, Biotechnol Res Asia 4

- 226. Sakai S (1983) Comparative study on freezing resistance of conifers with special reference to cold adaptation and its evolutive aspects. Can J Bot 61
- 227. Friedman JM, Roelle JE, Gaskin JF, Pepper AE, Manhart JR (2008) Latitudinal variation in cold hardiness in introduced Tamarix and native *Populus*. Evol Appl 1:598
- 228. Smith A (1992) In: J Hanover, KMiller, S Plesko (eds) Proceedings: International Conference on Black Locust: Biology, Culture, Utilization. Department of Forestry. Michigan State University, East Lansing, MI, p 208–217
- 229. Siminovitch D (1963) Evidence from increase in ribonucleic acid and protein synthesis in autumn for increase in protoplasm during the frost-hardening of black locust bark cells. Can J Bot 41:1301
- 230. Siminovitch D, Rheaume B, Pomeroy K, Lepage M (1968) Phospholipid, protein, and nucleic acid increases in protoplasm and membrane structures associated with development of extreme freezing resistance in black locust tree cells. Cryobiology 5:202
- 231. Keresztesi B (1988) The black locust. Forestry monograph series of the agricultural Science department of the Hungarian Academy of Sciences. Akadémiai Kiadó, Budapest, Hungary
- Keresztesi B (1983) Breeding and cultivation of black locust, Robinia pseudoacacia, in Hungary. For Ecol Manag 6:217
- 233. Hale AL, Viator RP, Veremis JC (2014) Identification of freeze tolerant Saccharum spontaneum accessions through a pot-based study for use in sugarcane germplasm enhancement for adaptation to temperate climates. Biomass Bioenerg 61:53
- Sakai A (1970) Freezing resistance in willows from different climates. Ecology 51:485
- 235. Maulana F (2011) Kansas State University
- Ercoli L, Mariotti M, Masoni A, Arduini I (2004) Growth responses of sorghum plants to chilling temperature and duration of exposure. Eur J Agron 21:93
- 237. Kapanigowda MH et al (2013) Analyses of sorghum Sorghum bicolor (L.) Moench lines and hybrids in response to early-season planting and cool conditions. Can J Plant Sci 93:773–784
- 238. Potter L, Bingham M, Baker M, Long S (1995) The potential of two perennial C4 grasses and a perennial C4 sedge as ligno-cellulosic fuel crops in NW Europe Crop establishment and yields in E England. Ann Bot 76:513
- Schwarz AG (1989) Physiological ecology of northern populations of C 4 grasses. Dissertation Abstracts International. B, Sciences and Engineering 50, 2257B
- 240. Madakadze IC, Coulman BE,McElroy AR, Stewart KA, Smith DL (1998) Evaluation of selected warm-season grasses for biomass production in areas with a short growing season. Bioresource Technol 65:1–12
- Bartolo ME, Wallner SJ, Ketchum RE (1987) Comparison of freezing tolerance in cultured plant cells and their respective protoplasts. Cryobiology 24:53
- 242. Clifton-Brown JC et al (2001) Performance of 15 *Miscanthus* genotypes at five sites in Europe. Agro J 93:1013
- 243. Seepaul R, Macoon B, Reddy KR, Baldwin B (2011) Switchgrass (*Panicum virgatum* L.) intraspecific variation and thermotolerance classification using in vitro seed germination assay. Am J Plant Sci 2:134
- 244. Shakeel SN, Ul Haq N, Heckathorn S, Luthe D (2012) Analysis of gene sequences indicates that quantity not quality of chloroplast small HSPs improves thermotolerance in C4 and CAM plants. Plant Cell Rep 31:1943
- 245. Kakani VG, Surabhi GK, Reddy KR (2008) Photosynthesis and fluorescence responses of C(4) plant *Andropogon gerardii* acclimated to temperature and carbon dioxide. Photosynthetica 46:420
- 246. Kalyar T, Rauf S, Teixeira da Silva JA, Haidar S, Iqbal Z (2013) Utilization of leaf temperature for the selection of leaf gas-exchange traits to induce heat resistance in sunflower (*Helianthus annuus* L.). Photosynthetica 51:419

- 247. Kalyar T, Rauf S, Teixeira da Silva JA, Shahzad M (2014) Handling sunflower (*Helianthus annuus* L.) populations under heat stress. Arch Agron Soil Sci 60
- 248. Sharmin SA et al (2013) Mapping the leaf proteome of *Miscanthus sinensis* and its application to the identification of heat-responsive proteins. Planta 238:459
- 249. Li Y-F, Wang Y, Tang Y, Kakani V, Mahalingam R (2013) Transcriptome analysis of heat stress response in switchgrass (*Panicum virgatum* L.). BMC Plant Biol 13:153
- 250. Peltola H, Kilpelainen A, Kellomaki S (2002) Diameter growth of Scots pine (*Pinus sylvestris*) trees grown at elevated temperature and carbon dioxide concentration under boreal conditions. Tree Physiol 22:963
- 251. Wertin TM, McGuire MA, Teskey RO (2010) The influence of elevated temperature, elevated atmospheric CO2 concentration and water stress on net photosynthesis of loblolly pine (*Pinus taeda* L.) at northern, central and southern sites in its native range. Glob Chang Biol 16:2089
- 252. Ultra VU Jr, Han S-H, Kim D-H, Aggangan NS (2012) Changes in growth, photosynthetic parameters, carbon and nitrogen uptake of *Pinus densiflora* S. et Z. under elevated temperature and CO2 level. *Asia*. Life Sci 21:523
- 253. Ferreira S et al (2006) Proteome Profiling of *Populus euphratica* Oliv upon heat stress. Ann Bot 98:361
- 254. Appleton B et al (2009) Trees for hot sites. Virginia Cooperative Extension Publication 420-024. http://pubs.ext.vt.edu/430/430-024/ 430-024_pdf.pdf. Accessed 29 Apr 2014
- 255. Tauer CG (2007) Performance of a wide-ranging collection of black locust seed sources in western Oklahoma. Tree Planters' Notes 52:26
- 256. Sadia G, Abdul W, Muhammad A, Muhammad A, Islam ud D (2008) Changes in growth and leaf water status of sugarcane (*Saccharum officinarum*) during heat stress and recovery. Int J Agri Biol 10:191–195
- 257. Marchand FL, Kockelbergh F, Van De Vijver B, Beyens L, Nijs I (2006) Are heat and cold resistance of arctic species affected by successive extreme temperature events? New Phytol 170:291
- 258. Medori M, Michelini L, Nogues I, Loreto F, Calfapietra C (2012) The impact of root temperature on photosynthesis and isoprene emission in three different plant species. Sci World J. doi:10.1100/2012/525827
- 259. McLeod KW, Donovan LA, Stumpff NJ, Sherrod KC (1986) Biomass, photosynthesis and water use efficiency of woody swamp species subjected to flooding and elevated water temperature. Tree Physiol 2:341
- 260. Lujan R et al (2009) Small heat-shock proteins and leaf cooling capacity account for the unusual heat tolerance of the central spike leaves in *Agave tequilana* var. Weber Plant Cell Environ 32:1791
- 261. Mittler R (2006) Abiotic stress, the field environment and stress combination. Trends Plant Sci 11:15
- 262. Ben Rejeb K et al (2013) Evaluation of the Cd2+ phytoextraction potential in the xerohalophyte Salsola kali L. and the impact of EDTA on this process. Ecol Eng 60:309–315
- 263. Subudhi PK, Baisakh N (2011) Spartina alterniflora Loisel, a halophyte grass model to dissect salt stress tolerance. In Vitro Cell Dev Biol—Plant 47:441–457
- 264. Davis AS et al (2010) Screening bioenergy feedstock crops to mitigate invasion risk. Front Ecol Environ 8:533

- 265. Quinn LD, Allen DJ, Stewart JR (2010) Invasiveness potential of *Miscanthus sinensis*: implications for bioenergy production in the U.S. GCB Bioenergy 2:310–320
- 266. Raghu S et al (2006) Adding biofuels to the invasive species fire? Science 313:1742
- 267. Barney JN, DiTomaso JM (2008) Nonnative species and bioenergy: Are we cultivating the next invader? Bioscience 58:64
- 268. Gordon DR, Tancig KJ, Onderdonk DA, Gantz CA (2011) Assessing the invasive potential of biofuel species proposed for Florida and the United States using the Australian Weed Risk Assessment. Biomass Bioenerg 35:74
- 269. ISAC (2009) Biofuels: cultivating energy, not invasive species (National Invasive Species Council. http://www.invasivespecies. gov/global/ISAC/ISAC_whitepapers.html. Accessed 11 Jan 2012
- 270. GISP (2007) Global Invasive Species Programme white paper pp. http://www.gisp.org/whatsnew/docs/biofuels.pdf. Accessed on 20 Jun 2012
- 271. Quinn LD, Holt JS (2008) Ecological correlates of invasion by *Arundo donax* in three southern California riparian habitats. Biol Invasions 10:591
- 272. Dougherty R F, Quinn L D, Endres A B, Voigt T B, Barney J N (2014) Natural history survey of the ornamental grass *Miscanthus sinensis* in the introduced range. Invas Plant Sci Mana 7:113–120. http://dx.doi.org/10.1614/IPSM-D-13-00037.1
- 273. Quinn LD (2014) In: LD Quinn, DP Matlaga, JN Barney (eds) Bioenergy and biological invasions: ecological, agronomic and policy perspectives on minimising risk. CABI, Oxfordshire
- 274. Buddenhagen CE, Chimera C, Clifford P (2009) What would invasive feedstock populations look like? Perspectives from existing invasions. Assessing Biofuel Crop Invasiveness: a case study. PLos ONE 4:e5261
- 275. Glaser A, Glick P (2012) Growing risk: Addressing the invasive potential of bioenergy feedstocks. National Wildlife Federation, Washington, DC
- 276. Barney JN (2012) Best Management Practices for Bioenergy Crops: reducing the invasion risk. Virginia Cooperative Extension Publication PPWS-8P
- 277. Quinn LD, Barney JN, McCubbins JSN, Endres AB (2013) Navigating the "noxious" and "invasive" regulatory landscape: suggestion for improved regulation. Bioscience 63:124
- 278. Quinn L D et al (2014) Resolving regulatory uncertainty: legislative language for potentially invasive bioenergy feedstocks. GCB Bioenergy. doi:10.1111/gcbb.12216
- 279. Quinn LD, Gordon DR, Glaser A, Lieurance D, Flory SL (2014) Bioenergy feedstocks at low risk for invasion in the U.S.: a "white list" approach. Bioenerg Res. doi:10.1007/s12155-014-9503-z
- PIER (2013) US Forest Service, Pacific Island Ecosystems at Risk (PIER). http://www.hear.org/pier/. Accessed 16 Apr 2013
- IFAS Invasive Plant Working Group (2013) IFAS Assessment of non-native plants in Florida's natural areas. http://plants.ifas.ufl.edu/ assessment/. Accessed 1 Feb 2014
- 282. USDA APHIS, (2014) Noxious Weeds Program Weed Risk Assessments Database, US Department of Agriculture Animal and Plant Health Inspection Service. http://www.aphis.usda.gov/ planthealth/fnw. Accessed 21 Oct 2014