

J Endocrinol Invest (2014) 37:875–909
DOI 10.1007/s40618-014-0119-0

POSITION STATEMENT

Italian Association of Clinical Endocrinologists (AME) position statement: a stepwise clinical approach to the diagnosis of gastroenteropancreatic neuroendocrine neoplasms

Franco Grimaldi · Nicola Fazio · Roberto Attanasio · Andrea Frasoldati · Enrico Papini · Francesco Angelini · Roberto Baldelli · Debora Berretti · Sara Bianchetti · Giancarlo Bizzarri · Marco Caputo · Roberto Castello · Nadia Cremonini · Anna Crescenzi · Maria Vittoria Davì · Angela Valentina D’Elia · Antongiulio Faggiano · Stefano Pizzolitto · Annibale Versari · Michele Zini · Guido Rindi · Kjell Öberg

Received: 10 February 2014 / Accepted: 29 March 2014 / Published online: 20 July 2014
© The Author(s) 2014. This article is published with open access at Springerlink.com

Keywords Neuroendocrine tumors · Diagnostic work-up · Markers · Imaging · Incidental findings · Non-functioning tumors · Carcinoid syndrome · Gastrinoma · Insulinoma · NET · NEC · NEN

Abbreviations

5-HIAA 5-Hydroxy-indolacetic acid
ACE Angiotensin-converting enzyme
ACTH Adrenocorticotropin
AJCC American Joint Committee on Cancer

AKT A protein-serine-threonine kinase that is activated by phosphorylation in response to growth factors or insulin
CD117 Antigen specific for the proto-oncogene *c-kit*
CD56 Antigen expressed by all lymphocytes
CD99 Cluster of differentiation
CDX-2 Transcription factor expressed specifically in gut epithelium
CEACAM1 Cell adhesion molecule
CEUS Contrast-enhanced US
CgA Chromogranin A
CK19 Cytokeratin 19
CS Carcinoid syndrome
CT Computerized tomography
DBE Double balloon enteroscopy

On behalf of AME.

Other members of AME oncologic endocrinology group are listed in the conclusions.

Franco Grimaldi and Nicola Fazio contributed equally as first authors.

F. Grimaldi (✉)
Endocrinology and Metabolic Disease Unit, Azienda Ospedaliero-Universitaria “S. Maria della Misericordia”, P.le S.M. della Misericordia, 15-33100, Udine, Italy
e-mail: franco.grimaldi@aliceposta.it

N. Fazio
Unit of Gastrointestinal and Neuroendocrine Tumors, European Institute of Oncology, Milan, Italy
e-mail: nicola.fazio@ieo.it

R. Attanasio
Endocrinology Service, Galeazzi Institute IRCCS, Milan, Italy
e-mail: roberto.serena@libero.it

A. Frasoldati · M. Zini
Endocrinology Unit, Arcispedale S. Maria Nuova IRCCS, Reggio Emilia, Italy
e-mail: andrea.frasoldati@asmn.re.it

M. Zini
e-mail: michele.zini@asmn.re.it

E. Papini
Endocrinology Unit, Regina Apostolorum Hospital, Albano Laziale, Rome, Italy
e-mail: papinie@gmail.com

F. Angelini · S. Bianchetti
Oncology and Hematology Unit, Regina Apostolorum Hospital, Albano Laziale, Rome, Italy
e-mail: angelini.f@libero.it

S. Bianchetti
e-mail: bianchetti.sara@gmail.com

R. Baldelli
Endocrinology Section, Regina Elena National Cancer Institute, Rome, Italy
e-mail: baldelli@ifo.it

D. Berretti
Gastroenterology Unit, Azienda Ospedaliero-Universitaria “S. Maria della Misericordia”, Udine, Italy
e-mail: berretti.debora@aoud.sanita.fvg.it

DOPA	Dihydroxyphenylalanine	H2RAs	Histamine H2-receptor antagonists
DOTA	1,4,7,10-Tetra-azacyclo-dodecane-tetraacetic acid	Her/2	A cell surface protein-tyrosine kinase receptor overexpressed in adenocarcinomas
DOTANOC	DOTA-Nal3-octreotide	HPF	High-power field
DOTATATE	DOTA-octreotate	HPLC	High-pressure liquid chromatography
DOTATOC	DOTA-edotreotide	HTP	Hydroxy-L-tryptophan
ECLomas	Enterochromaffin-like cell carcinoids	IHC	Immunohistochemistry
EGDS	Esophago-gastro-duodenoscopy	IRMA	Immunoradiometric assay
ELISA	Enzyme-linked immunosorbent assay	Ki-67	Nuclear antigen present only in the nuclei of cycling cells
ENETS	European Neuroendocrine Tumor Society	LoE	Level of evidence
ERCC-1	Excision repair cross-complementing	MANEC	Mixed adeno-neuroendocrine carcinoma
ERCP	Endoscopic-retrograde-cholangio-pancreatography	MAO	Monoamine oxidase
EUS	Endoscopic ultrasonography	MDCT	Multidetector CT
FDG	Fluoro-deoxy-glucose	MEN-1	Multiple endocrine neoplasms
FGF	Fibroblast growth factor	MGMT	Methylguanine-DNA methyltransferase
FNA	Fine needle aspiration	MIB-1	A monoclonal antibody used to detect Ki-67 antigen
FNB	Fine needle biopsy	MRI	Magnetic resonance imaging
FSG	Fasting serum gastrin	mTOR	Mammalian target of rapamycin
GEP	Gastroenteropancreatic	NEC	Neuroendocrine carcinoma
GERD	Gastroesophageal reflux disease	NEN	Neuroendocrine neoplasm
GI	Gastrointestinal	NET	Neuroendocrine tumor
GRADE	Grading of recommendations, assessment, development, and evaluation	NF	Non-functioning
hCG	Human chorionic gonadotropin	NF1	Neurofibromatosis type 1
hHAS-1	Human achaete-scute homolog 1	NIH	National Institutes of Health

G. Bizzarri

Diagnostic Imaging Unit, Regina Apostolorum Hospital, Albano Laziale, Rome, Italy
e-mail: bizzarri giancarlo@libero.it

M. Caputo

Dipartimento Servizi di Diagnosi e Cura, AUSL 22 Regione Veneto, Bussolengo, VR, Italy
e-mail: mcaputo@ulss22.ven.it

R. Castello

Medicina Interna ad indirizzo Endocrinologico, Azienda Ospedaliera Universitaria Integrata, Verona, Italy
e-mail: roberto.castello@ospedaleuniverona.it

N. Cremonini

Endocrinology Unit, Maggiore and Bellaria Hospital, Bologna, Italy
e-mail: nadia.cremonini@ausl.bo.it

A. Crescenzi

Pathology Unit, Regina Apostolorum Hospital, Albano Laziale, Rome, Italy
e-mail: crescenzianna@libero.it

M. V. Davì

Medicina Interna D, Azienda Ospedaliera Universitaria Integrata, Verona, Italy
e-mail: mariavittoria.davi@ospedaleuniverona.it

A. V. D'Elia

Genetic Service, Azienda Ospedaliero-Universitaria "S. Maria della Misericordia", Udine, Italy
e-mail: delia.angela@aoud.sanita.fvg.it

A. Faggiano

Department of Clinical Medicine and Surgery, Federico II University, Naples, Italy
e-mail: afaggian@unina.it

S. Pizzolitto

Pathology Unit, Azienda Ospedaliero-Universitaria "S. Maria della Misericordia", Udine, Italy
e-mail: pizzolitto.stefano@aoud.sanita.fvg.it

A. Versari

Nuclear Medicine Service, Arcispedale S. Maria Nuova IRCCS, Reggio Emilia, Italy
e-mail: versari.annibale@asmn.re.it

G. Rindi

Institute of Pathology, Policlinico A. Gemelli, Università Cattolica del Sacro Cuore, Rome, Italy
e-mail: guido.rindi@rm.unicatt.it

K. Öberg

Department of Endocrine Oncology, University Hospital, Uppsala, Sweden
e-mail: kjell.oberg@medsci.uu.se

NPV Negative predictive value
 NSE Neuron-specific enolase
 NSP-55 Neuroendocrine-specific protein
 OS Overall survival
 PAX-8 Paired box gene 8
 PET Positron emission tomography
 PGP.9.5 Pan neuronal marker protein in the Islets of Langerhans
 PIK3 Phosphoinositide-3-kinase
 PLGF Placental growth factor
 pNEN Pancreatic NEN
 PPIs Proton pump inhibitors
 PPV Positive predictive value
 PTEN Phosphatase and tensin homolog (mutated in multiple advanced cancers)
 RIA Radioimmunoassay
 SA Somatostatin analog
 SEER Surveillance, epidemiology and end results
 SPECT Single photon emission computed tomography
 SRS Somatostatin receptor scintigraphy
 SSTR Somatostatin subtype receptor
 TNM Tumor-node-metastases
 TSC Tuberos sclerosis complex
 TTF-1 Thyroid transcription factor 1
 UICC Union for International Cancer Control
 UPN Unknown primary NEN
 US Ultrasonography
 VCE Video-capsule endoscopy
 VHL von Hippel-Lindau disease
 VIP Vasoactive intestinal peptide
 ZES Zollinger–Ellison syndrome

2.1.4 Genetic assessment..... 881
 2.2 Laboratory assessment..... 882
 2.2.1 “Unspecific markers” 882
 2.2.1.1 Chromogranin A..... 882
 2.2.1.2 Other unspecific markers 883
 2.2.2 “Specific markers” 883
 2.2.2.1 5-HIAA 883
 2.2.2.2 Gastrin..... 883
 2.2.2.3 Insulin 884
 2.2.2.4 Other specific markers 884
 2.3 Imaging procedures 885
 2.3.1 Radiologic procedures..... 885
 2.3.1.1 Ultrasonography 885
 2.3.1.2 Multislice triple phase CT 885
 2.3.1.3 MRI..... 885
 2.3.2 Nuclear medicine procedures..... 886
 2.3.2.1 SSTR functional imaging..... 886
 2.3.2.2 PET with other tracers 886
 2.3.3 Endoscopic procedures..... 887
 2.3.3.1 Upper and lower gastrointestinal NENs..... 887
 2.3.3.2 Small-bowel NENs..... 887
 2.3.3.3 Pancreatic NENs 887
 3 A step-by-step multidisciplinary approach to clinical diagnosis 888
 3.1 Incidental finding..... 888
 3.1.1 GEP-NENs suspected at endoscopy 888
 3.1.2 GEP-NEN suspected at morphological (US/CT/MR) imaging 888
 3.1.3 GEP-NEN suspected after elevated serum CgA levels..... 889
 3.2 Symptomatic patient with symptoms due to GEP-NEN-related local effects..... 890
 3.2.1 When to suspect a GEP-NEN..... 890
 3.2.2 Work-up in the patient with local compressive symptoms..... 890
 3.2.2.1 Isolated abdominal pain 890
 3.2.2.2 Subocclusive picture 890
 3.2.2.3 Jaundice 891
 3.2.2.4 Gastrointestinal bleeding..... 891
 3.3 Symptomatic patient with syndromes 892
 3.3.1 Diarrhea and flushing..... 892
 3.3.1.1 Clinical approach: when to suspect a GEP-NEN 892
 3.3.1.2 Work-up in the patient with suspected carcinoid syndrome 893
 3.3.2 Resistant/relapsing ulcer disease 894
 3.3.2.1 Clinical approach: when to suspect a GEP-NEN 894
 3.3.2.2 Work-up in the patient with suspected gastrinoma 895
 3.3.3 Spontaneous hypoglycemia..... 896
 3.3.3.1 Clinical approach: when to suspect a GEP-NEN 896
 3.3.3.2 Work-up in the patient with suspected insulinoma 897
 3.4 Work-up in the patient with metastatic disease and unknown primary tumor 899
 3.5 When and how to stage a previously diagnosed GEP-NEN..... 899
 4 Conclusions..... 900
 5 References..... 901

Table of contents

1 Introduction..... 878
 1.1 Why this document..... 878
 1.2 Methodology 878
 1.3 Definitions..... 878
 1.4 Classification..... 878
 1.4.1 Grading assessment 879
 1.4.2 Pathologic staging 879
 2 Diagnostic tools..... 879
 2.1 Histology, cytology, immunohistochemistry, and molecular biology..... 879
 2.1.1 Morphologic criteria..... 879
 2.1.2 Immunohistochemistry and molecular biology techniques 880
 2.1.3 Working with the pathologist and his pathologic report..... 881

1 Introduction

1.1 Why this document

Neuroendocrine neoplasms (NENs) can arise almost throughout the entire body and share common morphological, ultrastructural, and immunohistochemical characteristics.

Neuroendocrine neoplasms are an emerging entity that can occur at any age, with the median age at diagnosis in the late fifth decade and an age-related incidence increase. About two-thirds involve the gastro-entero-pancreatic (GEP) tract and epidemiological studies show their increasing incidence [1]. In the last decades, the overall reported incidence of GEP-NENs increased from 1.0 to 5.25/100.000 persons/year, with a present estimated prevalence of 35/100.000 [1–10]. Physicians' awareness, endoscopic screening and increased sensitivity of diagnostic tools may at least in part explain this growing trend.

Most guidelines are focused on staging, treatment and follow-up of NENs. However, an appropriate clinical suspicion and a correct diagnostic work-up are critical starting points. A multidisciplinary approach, moreover, is crucial to provide a timely and integrated care. Hence, this document is neither a review, nor a guideline; rather, it is a clinical guide for a stepwise and integrated diagnostic work-up of GEP-NENs. Hopefully, this will result in a correct utilization of resources and optimization of the cost/benefit ratio.

1.2 Methodology

The grading of recommendations, assessment, development, and evaluation (GRADE) system was adopted for the present position statement [11–14]. Briefly, the GRADE system classifies evidence into four quality levels (high, moderate, low, or very low), and recommendations into two grades (strong or weak).

Whenever possible, the level of evidence (LoE) has been ranked as follows: very low (⊗○○○), low (⊗⊗○○), moderate (⊗⊗⊗○), and high (⊗⊗⊗⊗). “Very low quality” evidence corresponds to unsystematic clinical observations (case report, case series) or indirect evidence (e.g., surrogate end points); “low quality” evidence corresponds to observational studies or randomized controlled trials (RCT) with major limits; “moderate quality evidence” corresponds to RCTs with limitations or rigorous observational studies; and “high quality evidence” corresponds to well performed RCTs and strong evidence from unbiased observational studies [13].

We labeled as “recommendations” and “suggestions” the strong and weak recommendations, respectively. Each recommendation/suggestion is based on the quality of

supporting evidence, downgraded or upgraded according to adjunctive factors (e.g., inconsistency of results, indirectness of evidence, lack of precision and limited number of relevant publications downgrade the recommendation/suggestion; large effect size, narrow confidence intervals, clinically very significant end points upgrade the recommendation/suggestion), and the level of panel agreement [13].

1.3 Definitions

Neuroendocrine neoplasms neoplastic cells possess features of both neural and epithelial cells. Therefore, in line with the WHO classification, the term neuroendocrine will be adopted throughout this document [15].

WHO recommends the use of the term “neuroendocrine neoplasm” (NEN) to indicate low- to high-grade lesions. The term “neuroendocrine tumor” (NET) will be used throughout this document, due to its widespread diffusion, to indicate low- to intermediate-grade lesions and the term “neuroendocrine carcinoma” (NEC) to indicate high-grade lesions. Terms like “carcinoids” and the embryological classification of GEP-NENs in tumors of foregut (thymus, esophagus, lung, stomach, duodenum, pancreas), midgut (appendix, ileum, cecum, ascending colon) and hindgut (distal colon and rectum) will be avoided.

1.4 Classification

In the last 10 years WHO has repeatedly revised the pathologic classification of GEP-NENs (Table 1) [16].

According to the 2010 classification, **NET G1** includes the “carcinoids” or “well-differentiated tumors” of the

Table 1 WHO classifications of GEP-NENs

WHO 1980	WHO 2000	WHO 2010
I. Carcinoid	Well-differentiated endocrine tumor	Neuroendocrine tumors
	Well-differentiated endocrine carcinoma	NET G1 (Grade 1) NET G2 (Grade 2)
	Poorly differentiated endocrine carcinoma/ small-cell carcinoma	Neuroendocrine carcinoma NEC G3 (Grade 3): Large-cell NEC small-cell NEC
II. Mucocarcinoid	Mixed exocrine– endocrine carcinoma	Mixed adeno- neuroendocrine carcinoma (MANEC)
III. Mixed carcinoid- adenocarcinoma forms		
IV. Pseudotumor lesions	Tumor-like lesions	Hyperplastic and preneoplastic lesions

Table 2 Grading system for GEP-NENs (adapted from 19)

	Ki-67 index (%) ^a	Mitotic count/10 HPF ^b
NET G1	≤2	<2
NET G2	3–20	2–20
NET G3	>20	>20

^a Assessed by MIB-1 labeling in at least 2,000 tumor cells in high nuclear density (“hot spot”) areas

^b 10 HPF = 2 mm², at least 50 optical fields in high-density mitotic areas

1980 and 2000 WHO classifications. These tumors are usually indolent, but can occasionally behave as malignant.

NET G2 may be considered a “grey zone”, with heterogeneous behavior, and requires a tailored management.

NET (G3) is a malignant neoplasm with an aggressive clinical course.

MANEC has a malignant phenotype with features of both adenocarcinoma and NET. This definition requires the presence of at least 25 % of each component. Neuroendocrine cells are usually interspersed and the two populations may be identified only by immunohistochemistry (IHC). Less frequently, neuroendocrine cells may be grouped in distinct regions that are recognized by light microscopy.

The WHO 2010 classification strongly relies on tumor grading. Grading relates to the biological aggressiveness of the neoplasm, whereas differentiation indicates its similarity to the tissue of origin [15]. The clinical behavior of NENs may be basically predicted by their grading, staging, and evidence of hormonal syndromes. All these data should be collected and weighted to establish the prognosis and management of the patient.

1.4.1 Grading assessment

The grade of a tumor is the primary predictor of its clinical outcome. Grading is based on the proliferation rate of the tumor, as assessed by the Ki-67 cell labeling and by the mitotic count (number of mitosis × 10 high power fields—HPF) (Table 2) [15–23].

Visual estimates are currently used as the standard technique for evaluating both Ki-67 and the mitotic count [24, 25]. Several areas should be assessed within the tumor to reduce the risk of evaluation bias due to intratumoral heterogeneity. Densely stained regions (“hot spots”) should be preferentially evaluated. Results from these areas should be reported as a single percentage reflecting the highest identified count [16, 21, 22].

Potential pitfalls and limitations are:

- a. technical problems (e.g., tissue processing, differences in Ki-67 antibodies, etc.);

- b. intratumoral heterogeneity and sampling limitations (e.g., a single biopsy sample may not be representative of the tumor grade within the whole neoplastic mass) [24, 26];

- c. discordances:

- I. between the proliferative rate and the degree of differentiation (e.g., a morphologically well-differentiated NEN may exhibit a high proliferative rate);
- II. between the predictive value for prognosis and that for treatment response (e.g., Ki-67 is a reliable predictor of disease progression and overall survival (OS), but seems a less efficient predictor of response to medical treatment) [27].

1.4.2 Pathologic staging

GEP-NENs are staged according to tumor size, site of origin, and locoregional or distant spreading [21–23]. The staging information is integrated with the 2010 WHO classification to stratify the prognostic risk and optimize the therapeutic and follow-up strategies (Fig. 1).

We recommend the use of the 2010 WHO classification.

We recommend for staging the use of the AJCC-TNM 2009, and just for pancreas and appendix AJCC-TNM 2009 and ENETS-TNM 2006/07. The selected system should be specified in the pathologic report.

We recommend NEN classification and clinical actions be based on the less favorable data in case of conflicting findings.

2 Diagnostic tools

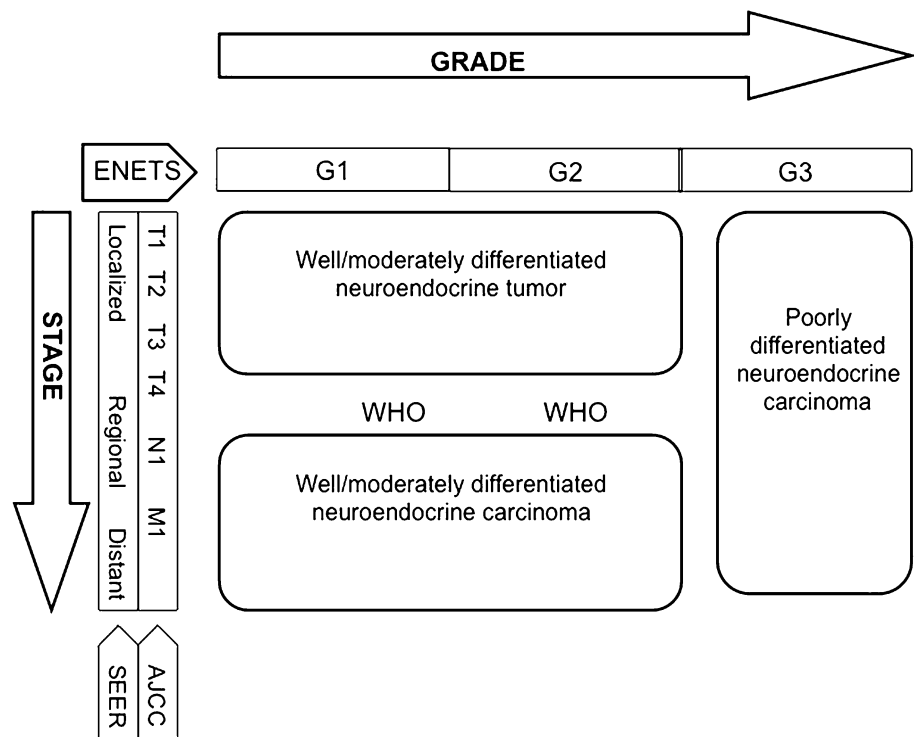
2.1 Histology, cytology, immunohistochemistry, and molecular biology

2.1.1 Morphologic criteria

Pathologic assessment is required for the diagnosis, classification and staging of NENs.

GEP-NENs present a broad architectural spectrum [28]. Well-differentiated tumors show an organoid pattern that ranges from solid nests to micro-macrotrabecular/gyriform pattern. A rich sinusoidal vascularity is usually observed. Stromal fibrosis, amyloid deposition, and calcification may be present. Necrosis can be present either as large infarct-

Fig. 1 Integrated pathologic and biologic classification (modified from 15)



like areas or as punctate foci in the center of neoplastic nests. Regardless of their growth pattern, NEN cells have a similar cytological appearance: small- to medium-size cells with round to oval shape and eosinophilic, lightly granular, cytoplasm. The nuclei are usually centrally placed, fairly uniform, with a finely dispersed (“salt and pepper”) chromatin pattern. Rarely, the neoplastic cells have a “plasmocytoid appearance” due to peripherally located nuclei. Nucleoli are usually inconspicuous or absent. Intracytoplasmic hyaline globules or nuclear pseudoinclusions may be seen.

High-grade NENs are composed of small or large-to-intermediate cells with high-grade features (marked nuclear atypia, multifocal necrosis, high mitotic index) and diffuse growth, sometimes with organoid feature resembling NEN.

The subgroup of GEP-NENs with Ki67 >20 % (and therefore G3 according to WHO 2010), but with a morphology of well-/moderately differentiated tumor should be considered low/intermediate rather than high-grade NENs [29].

Cytological specimens, which may be the only source of diagnostic material, pose some problems for clinical management. Cytology effectively separates high-grade NENs from low-grade NENs, but the distinction between low- and intermediate-grade NENs may be impossible. The diagnostic accuracy of aspiration techniques may be limited by the small sample size, the suboptimal

reproducibility and the risk of contamination from contiguous tissues.

Cytology is gaining a major role for the diagnosis in duodeno-pancreatic tumors. The endoscopic ultrasonography (EUS) fine needle aspiration (FNA) technique appears reliable, with a reported specificity of about 75 %, sensitivity of 87.5 %, accuracy of 89 %, positive predictive value (PPV) of 93 %, and negative predictive value (NPV) of 60 % [30–32].

2.1.2 Immunohistochemistry and molecular biology techniques

Neuroendocrine differentiation Synaptophysin (a small vesicle-associated marker) and Chromogranin A (CgA, a large secretory granule-associated marker) are useful IHC markers for the diagnosis of NENs. In NEC, the staining for both these markers is required to confirm the diagnosis, because CgA may be negative [15]. Routine IHC staining for peptide hormones and bioamines is not recommended. Other neuroendocrine markers, such as PGP.9.5, NSE, CD56, NSP-55, are of questionable specificity and clinical usefulness.

Prognostic markers proposed in addition to Ki-67 are CK19, CD117, CD99, p53, Her/2, CEACAM1, E-cadherin, β -catenin, hHAS-1, FGF13, PLGF, PAX-8, PTEN. None is presently recommended for clinical practice. The research of circulating tumor cells or the use of microRNAs is not indicated for routine use [33, 34].

Markers of primary site These markers may be a key for determining the unknown primary tumor in metastatic lesions. The most useful are [35]:

- TTF-1, indicative of pulmonary or thyroid origin;
- serotonin and CDX-2, indicative of intestinal origin;
- PAX-8 and histidine-decarboxylase, indicative of pancreatic origin;
- xenin, indicative of duodenal origin.

Markers predictive of response to specific treatments These biomarkers are not indicated for routine diagnostic practice. They include [19]:

- somatostatin receptors (SSTR)-2A (IHC determination at the cell membrane level), for planning the treatment with somatostatin analogs (SA);
- Akt/mTOR pathway molecules (PIK3, PTEN, TSC2), for treatment with everolimus;
- thymidylate synthase, for treatment with antifolates;
- ERCC-1, for treatment with platinum;
- topoisomerase II α , for treatment with etoposide;
- epigenetic events, as methylation of MGMT promoter, for treatment with alkylating agents.

- completeness of resection, distance of the surgical margins from the tumoral edge, depth of invasion;
- signs of malignancy (angiolymphatic and/or perineural invasion, necrosis, infiltration of the capsule and/or of gastrointestinal (GI) wall and/or surrounding tissues);
- number of examined lymph nodes, and number of lymph node metastases; presence of micrometastases; diameter of largest metastasis;
- presence of distant metastases, if demonstrated;
- functional activity (if appropriate).

The report should be concluded with the WHO diagnosis and classification of the lesion (NET G1–G2 or NEC G3) based on proliferative index (Ki-67 and/or mitotic count), and with the tumor stage (the staging system should be specified).

The minimum pathology data set for resected specimens (both primary and metastatic) should include [37]:

- site;
- diagnosis (e.g., pure neuroendocrine neoplasm);
- differentiation (i.e., well or poor);
- proliferation (i.e., G1 or G2 or G3).

We recommend routine IHC assessment of synaptophysin and CgA.

We suggest IHC assessment of peptide hormones or bioamines as optional in selected cases.

We recommend against routine use of other IHC markers in clinical practice.

We recommend histology as the diagnostic standard, cytology if histology is not available.

We recommend classification according to WHO 2010.

We recommend grading according to Ki-67 index and/or mitotic count.

We recommend staging according to AJCC/UICC TNM and ENETS.

2.1.3 Working with the pathologist and his pathologic report

The modality and timing of sampling techniques should be planned by a multidisciplinary team.

The pathologist should be provided with accurate clinical information including signs and symptoms, laboratory findings and imaging studies [36].

The ideal pathologic report should include:

- description of the macroscopic specimen;
- tumor size (three dimensions);
- description of cell features and histologic architecture;
- differentiation (well or poorly differentiated);
- IHC findings (CgA and synaptophysin routinely, SSTR2A when appropriate (e.g., when functional imaging for SSTR2 is negative));
- Ki-67 and mitotic count;

2.1.4 Genetic assessment

Approximately 5–10 % of GEP-NENs have a hereditary background as part of tumor susceptibility syndromes: multiple endocrine neoplasia type 1 (MEN-1), von Hippel-Lindau disease (VHL), neurofibromatosis type 1 (von Recklinghausen disease, NF1) and the tuberous sclerosis complex (TSC). All are inherited autosomal dominant disorders [38].

MEN-1 GEP-NENs are the second most common manifestation of MEN-1, reported in 30–70 % of cases in different series [mostly non-functioning (NF)] [39, 40]. A germ-line *MEN-1* mutation is identifiable in about 80–90 % of familial cases [41] and in about 42 % of sporadic cases [42]. Germline mutations arise de novo without any family history in approximately 10 % of patients [43]. *MEN-1*

mutation testing should be offered to index cases and to their first-degree relatives, even if asymptomatic [40]. Genetic counseling is recommended [40]. The family members who carry the *MEN-1* mutation require routine surveillance for early detection of endocrine tumors, whereas those who do not carry the mutation can be reassured. When molecular genetic testing is not available locally, patients highly suspected for MEN-1 should be addressed to a referral centers. No genotype/phenotype correlations have been demonstrated in MEN-1 syndrome [44, 45].

VHL Endocrine pancreatic NF tumors occur in 11–17 % of patients with VHL disease [46]. The penetrance of *VHL* mutations is almost complete by age 65 years [47]. Genetic testing detects mutations in virtually all affected individuals [48] and should be offered to all individuals with clinical evidence of VHL and to first-degree relatives. As ophthalmologic screening for those at risk for VHL disease begins before age five, molecular genetic testing is suggested also in young asymptomatic children [49, 50].

NF1 GEP-NENs occur in 1 % of the NF1 patients [51]. Half of affected individuals have NF1 as the result of a de novo mutation. The offspring of an affected individual is at a 50 % risk of inheriting the altered *NF1* gene, and the disease manifestations are extremely variable, even within the same family [52]. Molecular testing for NF1 is not usually recommended in the clinical practice: screening for NF1 mutations is useful only in individuals who do not completely fulfill the NIH diagnostic criteria.

TSC A few cases of pancreatic (p)NENs have been described in patients with TSC [53–55]. The diagnosis of TSC is usually based on clinical findings and mutations can be identified in approximately 85 % of individuals who meet the diagnostic criteria [56]. Two-thirds of affected individuals have TSC as the result of a de novo mutation.

We recommend germ-line DNA testing only in presence of a family history or clinical findings suggestive of MEN-1 or VHL. Genetic testing should include mutational screening and sequencing. A preliminary genetic counseling is needed.

We suggest the routine determination of serum calcium and PTH levels in patients with duodeno-pancreatic NEN as a first-line screening for MEN-1.

We recommend against routine somatic (tumor tissue) DNA testing.

2.2 Laboratory assessment

The determination of GEP-NENs serum markers should not be used as a first-line diagnostic tool whereas it is

appropriate for monitoring the response to treatment and for long-term follow-up [57, 58].

Serum markers should be determined after:

1. an established diagnosis or strong clinical suspicion of GEP-NEN;
2. exclusion of physiologic and pathologic confounding conditions.

NEN markers may be regarded as “unspecific” or “disease-specific”.

2.2.1 “Unspecific markers”

2.2.1.1 Chromogranin A Chromogranin A is a widely employed serum marker for GEP-NENs, but its use presents limitations [59]. CgA circulates under different antigenic forms and no universal calibration standard is available [60]. IRMA and RIA results may be considered roughly equivalent [61], but the reference intervals are variable and results obtained with different assays cannot be compared.

Chromogranin A level may be increased in a number of pathologic conditions (Table 3), and in healthy subjects after eating or physical exercise. Accordingly, CgA levels are highly variable in the general population [62], and may partially overlap between GEP-NEN patients and controls. Hence, CgA has a poor first-line diagnostic value [5, 60, 62–66].

Proton pump inhibitors (PPIs) increase (up to sevenfold) CgA levels. The effects of PPIs persist for several days after drug discontinuation. Therefore, CgA testing should be performed after an at least 2-week PPIs withdrawal [62, 67]. The effects of H₂-receptor antagonists (H₂RAs) on CgA are still controversial [68].

Table 3 Potential confounders causing CgA increase [64]

Neoplastic (other than GEP-NENs)
Breast cancer
Prostate cancer
Ovarian cancer
Hepatocarcinoma
Pancreas adenocarcinoma
Colon cancer
Non-neoplastic
Kidney or heart failure
Endocrine diseases (hyperthyroidism, hyperparathyroidism)
Local or systemic inflammatory disease
Chronic obstructive broncho-pulmonary disease
Gastro-enteric pathologies: chronic atrophic gastritis, pancreatitis, inflammatory bowel disease, cirrhosis, chronic hepatitis

Diagnostic accuracy of CgA depends on different variables:

- tumor burden (sensitivity 60–100 vs. 29–50 % in metastatic and localized disease, respectively) [62, 64, 69];
- type and site of tumor (sensitivity 96 vs. 75 % in functioning and NF tumors, respectively) [63, 70].

2.2.1.2 Other unspecific markers Neuron-specific enolase (NSE) is an enzyme found in neuroectoderm-derived cells. The presence of NSE has been reported in thyroid and prostate carcinomas, neuroblastomas, small-cell lung carcinomas, and pheochromocytomas. The clinical usefulness of this marker is hampered by its poor specificity [71]. NSE level is elevated in 30–50 % of patients with NEN, particularly those with poor differentiation. The combined determination of NSE and CgA may improve sensitivity in GEP-NEN diagnosis [72].

Pancreatic polypeptide (PP) is secreted by specialized pancreatic islet cells and inhibits gut motility and pancreatic exocrine secretion. PP has been proposed for the diagnosis and monitoring of NF pNENs, as its combination with CgA increases sensitivity up to 93 % [69]. Its routine use is not recommended due to the low diagnostic performance (sensitivity 63 % and specificity 81 %). PP levels may increase in old age, diarrhea, laxative abuse, gut inflammatory processes and chronic renal disease.

Beta subunit of human chorionic gonadotropin (hCG), a glycoprotein synthesized by the syncytiotrophoblastic cells of the placenta during pregnancy, may be increased in patients with pNENs [73], but has no use in every day practice.

As a whole, the clinical usefulness of the above reported markers is limited.

2.2.2 “Specific markers”

2.2.2.1 5-HIAA 5-HIAA, the main urinary metabolite of human serotonin, is determined by HPLC on 24 h urine samples. Results may be expressed as absolute values or as a ratio to creatinine excretion.

Some pre-analytical variables, mostly tryptophan/serotonin-rich foods and drugs, may interfere with serotonin metabolism (Table 4) [74]. These products should be avoided prior to urine collection, respectively, for at least 72 and 24 h [64].

The normal 5-HIAA urinary excretion ranges from 2 to 8 mg/day, but unspecific elevations (up to 30 mg/day) may be found in malabsorption syndromes, such as celiac and Whipple’s disease [75–77].

The determination of 24-h urinary excretion of 5HIAA has a sensitivity of over 90 % and a specificity of 90 % for

Table 4 Drugs and foods interfering with 5-HIAA assay

False negative results

Acetylsalicylic acid
Phenothiazines: chlorpromazine, promethazine
Imipramine and MAO-inhibitors
ACTH
Ethanol
MethylDOPA and hydrazine derivatives
Ketoacids
LevoDOPA
Isoniazid, methenamine, gentisic and homogentisic acid
Streptozotocin
Heparin

False positive results

Acetaminophen, naproxen, phenacetin
Caffeine, nicotine
Coumaric acid
Diazepam
Ephedrine
Fluorouracil, melphalan
Phenobarbital
Phentolamine, reserpine, guaifenesin, mephesisin
Methamphetamine, Phenmetrazine
Methocarbamol
Mesalamine

Foods: bananas, avocados, kiwi, pineapples, peanuts, tomatoes, plums, eggplants, walnuts, pecans, coffee, tea, cocoa/ chocolate, vanilla, sweets, and cookies (sugar and marmalade are allowed)

full-blown carcinoid syndrome (CS, see “Box 2, Carcinoid syndrome”). Urinary 5-HIAA excretion in these patients is reportedly higher than 90 mg/day (up to 2,000 mg/day). The test sensitivity, however, is definitely lower in absence of clinical symptoms [77, 78].

There is a possibility to analyze plasma 5-HIAA which might replace urinary-5-HIAA in a future [79].

Various blood serotonin assays have been proposed, but their actual accuracy has not been established. False positives may occur due to several interfering factors, as the release of platelet serotonin or the previous ingestion of tryptophan/serotonin-rich foods [80]. Accordingly, serotonin determination is not recommended in clinical practice.

2.2.2.2 Gastrin Gastrin determination has a key role in the evaluation of patients with signs and symptoms suggestive of Zollinger–Ellison syndrome (ZES, see “Box 3, Gastrinoma”).

Various antigenic isoforms of gastrin circulate in the blood. Care must be taken because some commercial

Table 5 Main drugs and foods that may interfere in gastrin assay

False negative results
Acetylsalicylic acid
LevoDOPA
False positive results
Hypochlorhydria/achlorhydria due to chronic use of PPIs and H2RAs or chronic atrophic gastritis (often associated with pernicious anemia)
<i>Helicobacter pylori</i> infection
Gastric outlet obstruction
Renal failure
Antral G-cell syndromes
Short-bowel syndrome
Retained antrum

immunoassay kits detect only the gastrin-17 molecule [81] and may cause false positive results.

Hypergastrinemia is commonly defined as a fasting serum gastrin above 100 pg/mL. Simultaneous measurement of gastric pH on a single sample is needed to rule out secondary hypergastrinemia due to other causes. In achlorhydria, pernicious anemia or atrophic gastritis high gastrin levels are usually associated to high (i.e., >4) pH values. On the contrary, serum gastrin levels >1,000 pg/mL combined with a <2 gastric pH are virtually diagnostic of ZES. Falsely elevated gastrin levels may be due to a few drugs (Table 5) that should be discontinued at least 2 weeks before the test [77, 82–86].

In general, gastrin levels are higher in pancreatic than in duodenal NENs, and are proportional to tumor burden and in patients with metastatic disease, exceedingly high gastrin levels may be observed. However, the majority of patients with ZES show mildly elevated (e.g., 150–1,000 pg/mL) gastrin levels, partially overlapping those of patients with renal insufficiency, small-bowel resection, retained gastric antrum, or on potent antisecretory drugs [87]. When the diagnosis is equivocal, a secretin stimulation test is needed. A gastrin increase >120 pg/mL over basal level is considered diagnostic [88].

2.2.2.3 Insulin The occurrence of repeated symptomatic hypoglycemia (<60 mg/dL) is suspicious for insulinoma (see “Box 4”) in subjects without diabetes. The diagnosis is confirmed by the presence of non-suppressed insulin levels in presence of low glucose levels (see “Spontaneous hypoglycemia”). To rule out a spurious hypoglycemia, laboratory processing should not be delayed. In subjects with leucocytosis glucose determination should be repeated with a collection tube that contains an inhibitor of glycolysis.

In presence of an episode of spontaneous severe hypoglycemia with hyperinsulinism, the simultaneous measurement of serum C-peptide and beta-hydroxybutyrate is appropriate. If factitious hypoglycemia is suspected, urinary sulfonyleureas should be tested as well. The work-up may be completed with the measurement of serum proinsulin [89]. This test, even if not widely available, is diagnostic of insulinomas secreting immature forms of insulin.

In selected patients with endogenous hyperinsulinism, autoimmune hypoglycemia, suspected on the basis of negative imaging tests and coexistence of autoimmune disorders, should be ruled out with the determination of insulin autoantibodies [90].

If the patient is not hypoglycemic when observed, the association of severe hypoglycemia with non-suppressed insulin levels should be sought under the conditions in which hypoglycemia would be expected (see provocative testing, “Spontaneous hypoglycemia”) [89].

2.2.2.4 Other specific markers Glucagon: Glucagonoma is associated with serum glucagon concentrations higher than 500 pg/mL and a characteristic clinical syndrome (diabetes mellitus and cutaneous manifestations, such as migratory necrolytic erythema, nail dystrophies, stomatitis, etc.) [91]. Glucagon concentrations higher than 1,000 pg/mL are virtually diagnostic for the disease, but some patients may exhibit levels within the physiologically elevated range. Moderate elevations in serum glucagon may be caused by protracted fasting in normal subjects or by renal and hepatic failure, trauma, sepsis, pancreatitis, abdominal surgery, and Cushing’s syndrome.

Due to the fast degradation of glucagon in vitro, blood must be collected in test tubes containing aprotinin and should be rapidly delivered to the laboratory. Results obtained with different glucagon assays may profoundly differ, due to the different calibration standards and the variable cross-reactivity with glucagon isoforms.

Vasointestinal peptide (VIP): Vasointestinal peptide-secreting tumors cause the Verner–Morrison syndrome, characterized by variable combination of watery diarrhea (>700 mL/day even during fasting, with tea-colored, odorless stools), hypokalemia, achlorhydria, weight loss, metabolic acidosis, hypercalcemia, glucose intolerance, and flushing. The diagnosis is established by high-volume secretory diarrhea associated with VIP levels higher than 75 pg/mL (to be confirmed by a second RIA determination) [92, 93]. VIP blood concentration is, in fact, extremely low in healthy subjects. Commercial kits are available, but their use is usually limited to tertiary referral centers.

We recommend against the use of biochemical markers as the initial diagnostic step for potential GEP-NEN patients.

We recommend the determination of the appropriate biochemical marker only after the diagnosis or strong clinical suspicion of GEP-NEN. The panel of markers should take into account the clinical picture and local availability/expertise.

We recommend considering all possible clinical and analytical interfering factors in presence of elevated serum or urinary levels of GEP-NEN markers. The determination should be repeated, if possible, after their timely withdrawal.

We suggest PPIs discontinuation at least 2 weeks before CgA and gastrin measurements.

We recommend, after the finding of an elevated gastrin level, its repeated determination together with the assessment of gastric pH.

We recommend for the follow-up of the markers expressed by GEP-NENs a serial measurement with the same laboratory assay.

2.3 Imaging procedures

2.3.1 Radiologic procedures

2.3.1.1 Ultrasonography Transabdominal US is an inexpensive, safe, rapid and non-invasive tool. US accuracy is, however, operator dependent and its sensitivity is generally low (13–27 %), when compared with MultiDetector CT (MDCT) and magnetic resonance imaging (MRI) [94]. In case of pNEN, a mean 39 % US detection rate has been reported [95, 96].

Contrast-enhanced US (CEUS) enables identification of hypervascular lesions, even in case of fast-flow tumor circulation, as in NF pNENs. Therefore, CEUS is significantly superior to B-mode US both in the detection of NF pNENs and in the diagnosis of liver metastases, visualized as hyperenhancing non-homogeneous lesions [96–98], with a reported sensitivity of 82 % [99, 100]. US may help in defining complications of advanced disease (i.e., biliary stricture) and/or guide diagnostic or therapeutic procedures [101].

Endoscopic ultrasonography and EUS-guided FNA, a fundamental procedure for the diagnosis of pNENs [96, 102, 103], will be treated in “Pancreatic NENs”.

2.3.1.2 Multislice triple phase CT Multidetector CT is considered the first choice imaging modality for detection,

staging and follow-up of GEP-NENs. When compared to conventional CT, MDCT allows a markedly higher spatial and temporal resolution. MDCT sensitivity and specificity are increased due to multiphase scanning. Images should be acquired in precontrast, arterial, portal and equilibrium phases.

Non-functioning pNENs and NEN liver metastases typically appear as hypervascular lesions. In the evaluation of NF pNENs, the combination of arterial dominant-phase and portal venous-phase CT improves the detection of primary tumors and hepatic metastases [96].

Reported mean sensitivity and specificity of MDCT are 73 % (63–82 %) and 96 % (83–100 %) for pNENs, and 82 % (78–100 %) and 92 % (83–100 %) for liver metastases, respectively [104–106].

When a small ileum lesion is suspected, MDCT enterography can be performed by distending the small bowel with a large volume of neutral or low-attenuating oral contrast medium [107–109]. The reported sensitivity and specificity of MDCT enterography are variable, ranging from 50 to 85 % and from 25 to 97 %, respectively.

Due to radiation exposure, MDCT examination should be tailored, particularly in young people, to reduce the scanned volume and the number of phases.

2.3.1.3 MRI Like MDCT, MRI offers a high spatial and time resolution with the possibility of multiplanar acquisition and reconstruction and multiphase examination after contrast injection. Along with the absence of ionizing radiations, an advantage of MRI over MDCT is the intrinsic signal difference (contrast) between the neoplasm and the healthy parenchyma. This characteristic is increased with imaging sequences based on proton diffusion. If compared with MDCT, the major drawbacks of MRI are the higher cost, lower accessibility and longer scanning time. Furthermore, MRI is more dependent on patient cooperation. At MRI, GEP-NENs show the same enhancement characteristic described for MDCT. As for contrast medium, Gadolinium-based (Gd-EOB DTPA) agents (Primovist for MRI) should not be used in patients with advanced renal function impairment.

Magnetic resonance imaging demonstrates a particular sensitivity for liver, bone, soft-tissue, and central nervous system metastases [87, 95]. Multiphase CT scan and MRI have similar effectiveness in the detection of islet cell tumors if fat-saturated T1-weighted and delayed enhanced T1-weighted sequences are included.

In clinical practice, MRI should be used when MDCT does not offer clear-cut results or when contrast medium is contraindicated [95]. Due to the absence of radiation exposure, MRI is used, in association with US, either as a screening image modality in young patients or in long-term surveillance [110].

We recommend chest-abdomen MDCT as the routine morphologic imaging modality for the detection and staging of GEP-NENs.

We recommend MRI when the evaluation of bone and CNS is required. In all the other cases MRI should be used as a second-line imaging study, when MDCT is not conclusive or contraindicated.

We suggest CEUS or MRI for a better characterization of liver involvement.

2.3.2 Nuclear medicine procedures

2.3.2.1 SSTR functional imaging Up to 80 % of GEP-NENs express primarily SSTR2 and SSTR5: this feature enables imaging with SA compounds, labeled with radioactive tracers.

The most common radiopharmaceutical SA is ^{111}In -pentetreotide (commercially available as Octreoscan[®]) used for scintigraphy, SPECT and SPECT/CT [111, 112]. Modern hybrid acquisition systems as SPECT/CT allow a coregistration of functional and morphologic imaging, which improves the localization of lesions [113].

Due to its high affinity to SSTR2, Octreoscan[®] shows a higher detection rate of NEN lesions as compared to conventional imaging, with a sensitivity ranging from 67 to near 100 % [114–119].

Among other radiolabeled SA, ^{68}Ga -DOTA-D-Phe1-Tyr3-octreotide (DOTATOC) binds SSTR2 and SSTR5 with higher affinity than Octreoscan[®] [120]. In light of higher spatial resolution (3–5 mm) and better quantification of tracer uptake offered by PET in comparison with scintigraphy, PET and PET/CT scan with ^{68}Ga -DOTATOC have significant advantages over SRS imaging, particularly in organs with high physiologic uptake (e.g., liver) and in case of small lesions (<1.5 cm) [121–123]. Furthermore, ^{68}Ga -DOTATOC has proven to be superior to CT and bone scintigraphy in the detection of bone metastases from GEP-NENs [124].

Similar results have been obtained with PET imaging using other ^{68}Ga -labeled peptides (e.g., ^{68}Ga -DOTATATE and ^{68}Ga -DOTANOC) [125–130]. PET/CT with ^{68}Ga -labeled SA is quite effective, both in terms of diagnostic accuracy and impact on clinical management [131–134].

Accordingly, this imaging procedure is recommended for routine use [73]. PET/CT with ^{68}Ga -labeled SA is presently available at a limited number of institutions, but will hopefully become diffusely adopted worldwide in the next future (Table 6).

Clinical indications for nuclear imaging based on radiolabeled SA are [135]:

- primary tumor localization and staging;
- restaging (detection of residual, recurrent or progressive disease);
- SSTR status evaluation (patients with high positivity are more likely to respond to octreotide therapy);
- response to therapy monitoring;
- selection of patients eligible for peptide receptor radionuclide therapy.

As octreotide therapy can theoretically interfere with ^{111}In -pentetreotide uptake, a brief (1–2 months) withdrawal of long-acting SA or a transient switch to short-acting SA should be considered [135].

2.3.2.2 PET with other tracers ^{18}F -FDG-PET/CT has been traditionally thought to play a minor role in GEP-NENs imaging due to the expected low FDG uptake of low-grade GEP-NENs [136]. As FDG uptake is greater in high-grade tumors, ^{18}F -FDG-PET/CT has been proposed in patients with advanced, metastatic GEP-NENs with promising results [137, 138]. In addition, combined functional imaging with both ^{68}Ga -DOTATATE and ^{18}F -FDG may be useful for a more comprehensive tumor assessment in intermediate and high-grade tumors [125]. Two recent studies confirm that FDG-PET is a sensitive technique for staging GEP-NENs with high (≥ 10 –15 %) Ki-67 [139, 140]. As for other tumors, it has been suggested that FDG positivity points to a worse prognosis [141–143].

^{18}F - and ^{11}C -labeled amine precursors L-dihydroxyphenylalanine (DOPA) [144–148] and 5-hydroxy-L-tryptophan [146, 149, 150] have been utilized for PET imaging of GEP-NENs in a limited number of studies with promising results. A still investigational tool is ^{18}F -fluorothymidine PET that seems to provide non-invasive assessment of cell proliferation. Finally, there is the possibility of utilizing glucagon-like peptide-1 receptor imaging for the localization of insulinomas [151]. Clinical application of these radiopharmaceuticals is not for routine use and needs confirmation.

Table 6 Comparison between Octreoscan and Ga-DOTA-peptides

	Availability	Duration	Accuracy	NPV	PPV
^{111}In -pentetreotide (Octreoscan [®])	Widespread	2 days	++	++	+++
^{68}Ga -DOTA-conjugate peptides	Low	2 h	+++	+++	+++

We recommend the use of SSTR functional imaging for localization and staging of G1-G2 GEP-NENs.

We recommend PET/CT with ^{68}Ga -labeled SA as the procedure of choice. When not available, ^{111}In -pentetreotide (Octreoscan®) scintigraphy may be used.

We recommend against the routine use of ^{18}F -FDG PET/CT.

We suggest ^{18}F -FDG PET/CT for staging high grade (G3) and selected G2 GEP-NENs.

2.3.3 Endoscopic procedures

2.3.3.1 Upper and lower gastrointestinal NENs Upper gastrointestinal endoscopy (EGDS) with gastric biopsy is required for the detection of gastric NENs.

Esophago-gastro-duodenoscopy is the only recommended imaging procedure in small (<1 cm) enterochromaffin-like cell tumors (ECLomas). Type 1 and 2 gastric NENs generally present (in 65–77 % of cases) as small (<2 cm) multifocal polypoid mucosal protrusions in the body and/or fundus of the stomach. Type 3 tumors are usually solitary, ulcerated and larger than 2 cm. In addition to biopsies of the largest polyps, samples should be taken from the antrum (two biopsies) and body/fundus (four biopsies) [152, 153]. Regardless of the type of gastric NEN, EUS may help to determine the presence of tumor invasion of the gastric wall and it is recommended before the resection of polyps >1–2 cm in diameter. EUS is useful for the assessment of the regional lymph node involvement and for cyto-histologic confirmation by FNA [154].

Duodenal NENs are approached in the same manner, namely EGDS with biopsies and EUS [155, 156].

The majority of rectal NENs are diagnosed endoscopically. Most lesions present as polyps, which are completely removed by snare polypectomy, but their diagnosis may be established only after histologic evaluation. Full colonoscopic assessment is required to exclude concomitant colonic disease as part of staging, and the possibility of synchronous carcinoma must be excluded. EUS is very useful in assessing rectal NENs extension preoperatively and it accurately assesses tumor size, depth of invasion and perirectal lymph node metastases. Hence, EUS provides information critical for the choice of final treatment (endoscopic vs. surgical) [157, 158].

2.3.3.2 Small-bowel NENs Direct visualization of small-bowel NENs may be obtained by standard colonoscopy if the tumor is prolapsed through the ileocecal valve into the colon, or if intubation of the ileum via the ileocecal valve is performed. Newer modalities to investigate the proximal parts of the ileum or the jejunum include video-capsule endoscopy (VCE) and enteroscopy. Small-scale studies have reported successful detection of occult small-bowel NENs by VCE where other techniques have failed. It is advisable to use a dissolvable “patency” capsule to avoid capsule “retention” within strictures. Major VCE limitations are as follows: (a) precise localization of the tumor is not usually possible; (b) in case of predominantly extraluminal GEP-NEN, the evaluation of the tumor cannot be accurate; and (c) cost and operating time. VCE revealed a sensitivity of 60 % and a specificity of 100 % as compared to CT enteroclysis [107, 159].

In selected cases, double balloon enteroscopy (DBE) seems to be a valuable method. It allows histologic confirmation by luminal biopsy and accurate preoperative localization by tumor marking with ink injection. A 33 % diagnostic yield of DBE for primary tumor detection in patients with metastatic or suspected GEP-NEN has been reported [160].

2.3.3.3 Pancreatic NENs Endoscopic ultrasonography is an effective tool to identify pNENs, which typically appear as well-defined hypoechoic, hypervascular masses. Cystic change, calcifications, and necrosis are common in large tumors. EUS-guided FNA (or biopsy, FNAB) is useful to confirm the diagnosis of pNEN. EUS sensitivity is quite high (79–100 %) with a PPV close to 100 % [161–164]. The accuracy decreases in case of lesions located in the pancreatic tail [165]. While EUS shows a higher sensitivity than cross sectional imaging in the diagnosis of small, multiple pNENs in MEN-1 or VHL syndromes, its accuracy in the detection of small duodenal tumor is controversial. The combination of dual-phase thin-section multidetector CT and EUS has been reported as the most accurate procedure to detect insulinomas [166]. EUS plus FNA is highly cost-effective when used early in the preoperative work-up, reducing the need for additional invasive tests [167, 168]; complication rate is quite low (<1 %) [168]. A close correlation between aspiration cytology and the final histology after resection has been demonstrated [169]. EUS is thus useful in the preoperative setting as it provides information that significantly influences the therapeutic planning [170].

We recommend endoscopy with biopsy in gastro-duodenal and colorectal NENs. If an ileal involvement is suspected, colonoscopy should possibly be extended to terminal ileum.

We recommend EUS to locally stage the disease before resection of gastric, duodenal and rectal polypoid lesions.

We suggest VCE and/or DBE as second-line tools for the diagnosis of small bowel NENs.

We recommend EUS plus FNA for the diagnosis of suspected pNENs.

3 A step-by-step multidisciplinary approach to clinical diagnosis

The suspicion of GEP-NEN can be raised in four different scenarios: (1) incidental finding either in a totally asymptomatic patient or in a patient with symptoms unrelated to GEP-NEN; (2) symptomatic patient with GEP-NEN-related local effects, (3) syndromes, and (4) metastases from unknown primary GEP-NEN. The first two scenarios are typical of NF GEP-NENs.

3.1 Incidental finding

GEP-NENs are often suspected following incidental imaging (e.g., US, CT, MRI) or endoscopic findings, in patients without signs or symptoms related to GEP-NEN [1, 3, 6].

The patient should be checked for minor GI complains (diarrhea, constipation, peptic disease, gastroesophageal reflux), any palpable mass, skin and metabolic signs/

symptoms, possibly suggesting a functioning syndrome. An accurate clinical history of the patient’s family should also be collected to confirm or rule out a hereditary syndrome [6].

3.1.1 GEP-NENs suspected at endoscopy

Incidental diagnosis of GEP-NENs often follows the histologic examination of polypoid lesions found during endoscopic procedures in an asymptomatic patient. Otherwise, gastroduodenal and colorectal NENs may be suspected in case of single or multifocal polypoid mucosal protrusions [152, 155, 158], even though no endoscopic finding is highly specific of NEN.

An endoscopic biopsy of the suspected lesion is mandatory. In case, the endoscopic biopsy is either not feasible or non-diagnostic, morphologic imaging studies should be programmed as the second step. Image-guided or laparoscopic biopsy should be discussed by the multidisciplinary team. Functional imaging could subsequently be performed as a complementary staging-prognostic tool.

No lab tests are indicated in the diagnostic work-up. The finding of hypergastrinemia, achlorhydria, macrocytic anemia, B₁₂ deficiency and/or intrinsic factor antibodies may be useful to categorize a gastric NEN (Fig. 2).

3.1.2 GEP-NEN suspected at morphological (US/CT/MR) imaging

This incidental finding is usually related to primary pancreatic tumor or liver metastases from a GEP-NEN.

A pNEN might be suspected in case of hypoechoic, hypervascular, and/or well-defined lesions at US/CEUS and of enhancing hypervascular lesions at CT scan or MRI. Cystic changes, calcifications, and necrosis are frequently observed in large lesions [171].

Fig. 2 Diagnostic flow-chart for GEP-NEN suspected at endoscopy

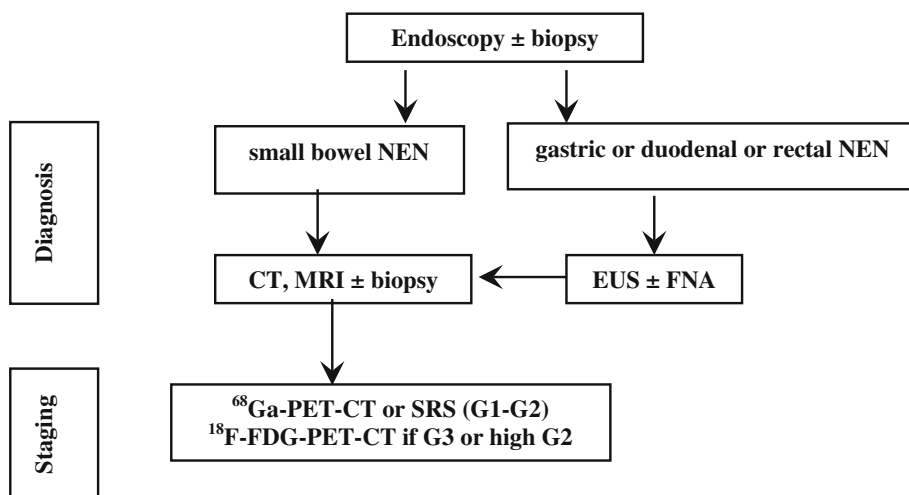
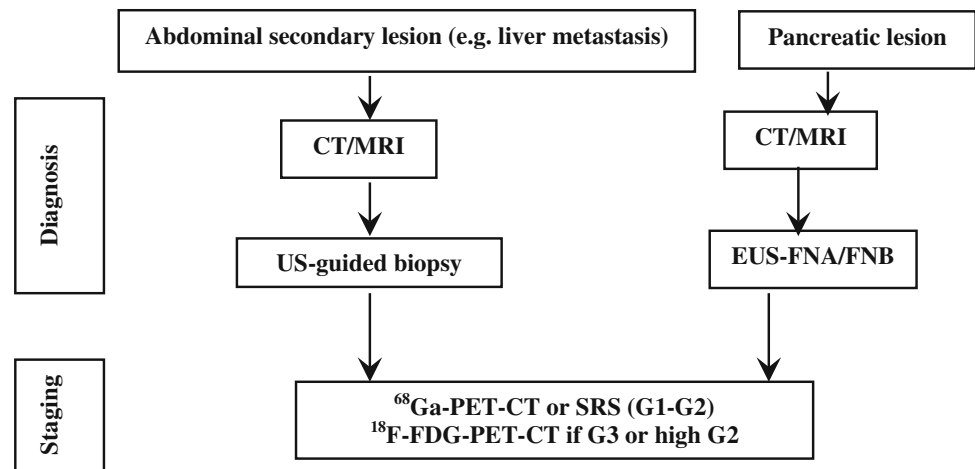


Fig. 3 Diagnostic flow-chart for GEP-NEN suspected at morphological imaging



False positives, especially in case of US imaging, like hemangiomas, hepatocellular and pancreatic carcinomas, intraductal pancreatic mucinous tumors, adenomas and metastasis from other tumors [94, 95, 97–101] should be ruled out by the multidisciplinary team.

A histologic/cytological specimen should possibly be obtained [96, 172].

Once the diagnosis of GEP-NEN is pathologically confirmed, proceed to morphologic and functional staging (see below, “When and how to stage a previously diagnosed GEP-NEN”). If biopsy is unfeasible or inconclusive, a second imaging technique (e.g., EUS, CEUS, liver-specific contrast-enhanced MRI, etc.) should be performed according to local expertise and availability [6].

Metastatic lesion(s) from occult primary may require a specific work-up (see below, “Work-up in the patient with metastatic disease and unknown primary tumor”).

No lab tests are recommended in the diagnostic work-up. Nevertheless, elevated 5-HIAA urinary excretion is highly specific of GEP-NEN liver metastases and may, therefore, be a strong diagnostic clue in case of a non-diagnostic biopsy. In patients with pNENs, the occurrence of subclinical, vague functional signs/symptoms possibly indicating a functional syndrome should always be carefully checked. Accordingly, specific hormonal assays may be required in selected cases (Fig. 3).

We recommend biopsy as the first diagnostic step in all lesions suspected for GEP-NEN.
We recommend diagnostic work-up to be routinely discussed within a NEN multidisciplinary team.
We recommend against the use of laboratory assays or functional imaging as a first-line diagnostic procedure.

3.1.3 GEP-NEN suspected after elevated serum CgA levels

Chromogranin A must never be considered a first-line diagnostic test. Nevertheless, NEN suspicion may occasionally be driven by the finding of elevated serum CgA levels, measured on the basis of unspecific symptoms or signs.

Before proceeding to imaging/endoscopic studies, all factors affecting CgA levels must thoroughly be ruled out (see “Table 3”). A second CgA determination is always required for confirmation. In patients on PPI treatment, serum CgA should be repeated after a two-week PPI withdrawal.

If CgA levels are confirmed elevated in absence of confounding factors, transabdominal US should be performed. A further diagnostic work-up should be discussed by a multidisciplinary team or a referral center should be involved (Fig. 4).

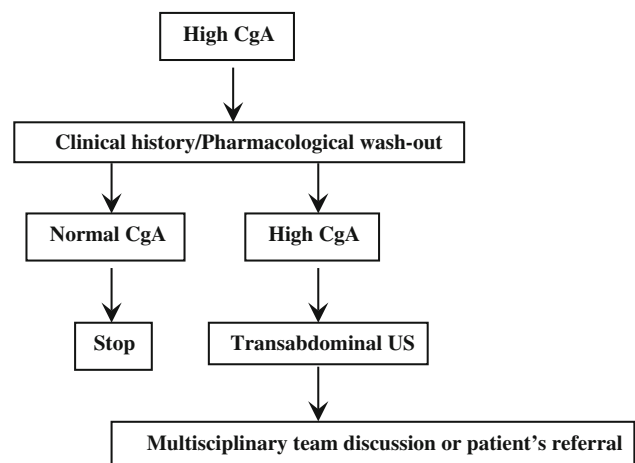


Fig. 4 Diagnostic flow-chart for NEN suspected after high CgA

We recommend careful exclusion of all potentially interfering factors in patients with elevated serum CgA levels and no previous NEN diagnosis.

We suggest transabdominal US as first step in case of confirmed CgA increase.

We recommend discussion of further work-up in a multidisciplinary team, involving a referral center when required.

3.2 Symptomatic patient with symptoms due to GEP-NEN-related local effects

3.2.1 When to suspect a GEP-NEN

Non-functioning GEP-NENs (Box 1) may become symptomatic when they compress or invade adjacent structures or when they metastasize. The suspicion of GEP-NEN might be raised by suggestive imaging findings (see above) and/or by the apparently slow progression of the disease [73]. Lab findings (e.g., frankly elevated CgA levels in absence of confounding factors) may reinforce the suspicion. As previously stated, only pathology (cytological or histologic characterization), however, will establish the diagnosis [6].

Box 1

Non-functioning GEP-NENs

Definition: NF GEP-NENs are tumors that do not show symptoms related to hormonal hypersecretion. Intracellular hormones or peptides may be demonstrated by IHC, but they are either not secreted, or secreted in quantities unable to elicit a clinical syndrome and/or in an inactive form [3]. Clinical presentation of NF GEP-NENs depends upon the site of origin and metastases. They can be incidentally discovered when asymptomatic due to the widespread use of diagnostic imaging [1, 3]. Clinical presentations according to the site of origin are listed below.

Pancreas: Up to 60% of pNENs is NF. Most NF pNENs are well differentiated. Annual incidence is 1.8 and 2.6 per million in females and males, respectively [3]. NF pNEN were traditionally diagnosed late in the course of the disease, with metastases in 46 to 73% of cases, but presently the number of incidentally found small lesions is steeply increasing. Presenting symptoms and signs are abdominal pain (35–78%), weight loss (20–35%), anorexia and nausea (45%), intra-abdominal hemorrhage (4–20%), jaundice (17–50%), and a palpable

mass (7–40%) [96, 172]. NF pNEN may occur in familiar syndromes such as MEN-1, VHL, and TSC.

Gastrointestinal: NENs are frequently detected during a screening program or an imaging exam performed to search the primary tumor in an asymptomatic but metastatic patient [1, 3]. Alternatively, a common clinical presentation is abdominal pain that may be caused by gastro-intestinal dysmotility or obstruction (associated or not to nausea, vomiting or constipation), or by bacterial overgrowth. Less common symptoms and signs are jaundice, weight loss, fatigue, fever and bleeding (massive or dripping). Clinical presentation of appendiceal NEN may mimic acute appendicitis [1, 3]. Obstructive symptoms are typical of small bowel, whereas minor bleeding is frequent in rectal disease [6, 73, 173].

3.2.2 Work-up in the patient with local compressive symptoms

A detailed history and complete physical examination are required.

Abdominal pain is the most common presenting symptom of NF GEP-NENs and may be related to the primary tumor or metastatic lesions [1, 3]. Pain localization and characteristics should be carefully examined. Four different scenarios can be distinguished.

3.2.2.1 Isolated abdominal pain A persistent and oppressive upper-abdominal pain may signal a pancreatic or retroperitoneal mass (pattern 1a) [96, 172], while a discontinuous cramping pain usually refers to an intestinal origin (pattern 1b) [73, 173]. In the former case, a radiological imaging should be performed first, followed by endoscopy/EUS as second step for pancreatic and duodenal lesions. In the latter case, endoscopy is recommended [73, 173]. A cytologic/histologic sampling should be obtained whenever possible (Fig. 5) [96, 172].

An ill-defined and diffuse abdominal pain (pattern 1c) can also be related to liver or nodal metastases. Abdominal US followed by a whole-body CT scan and a US-guided biopsy should be performed (Fig. 5).

3.2.2.2 Subocclusive picture It may be due to a large, often metastatic, ileal NEN and/or peritoneal carcinomatosis. Depending on the severity of the clinical picture, a direct abdomen-X-ray and/or an endoscopy could be performed [73, 173]. If an extrinsic obstruction is suspected, then an abdomen CT scan should be performed. If a peritoneal carcinomatosis is suspected, a transit evaluation water-soluble contrast medium X-ray could be useful (Fig. 6). If possible, histological specimens should be obtained through endoscopy. If not, a US/CT-guided biopsy

Fig. 5 Diagnostic flow-chart for GEP-NEN suspected after pattern 1a and 1b

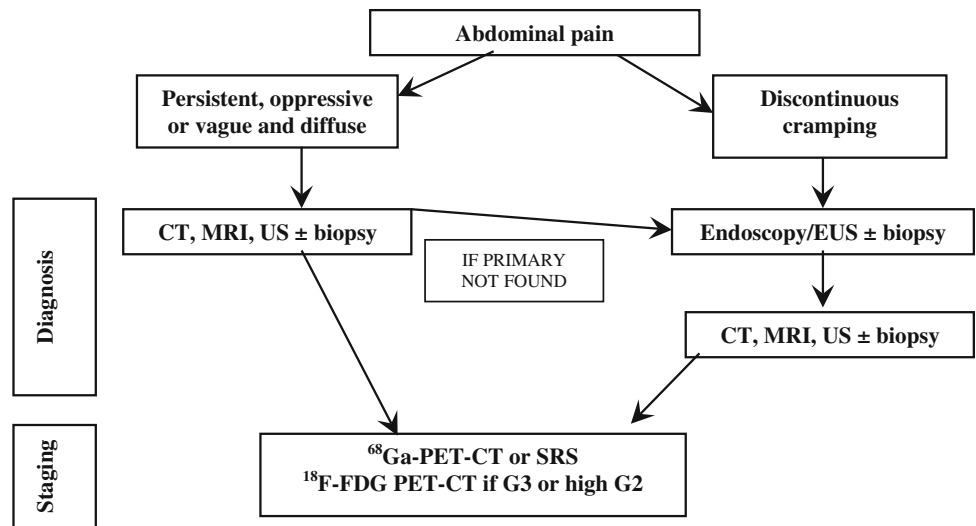
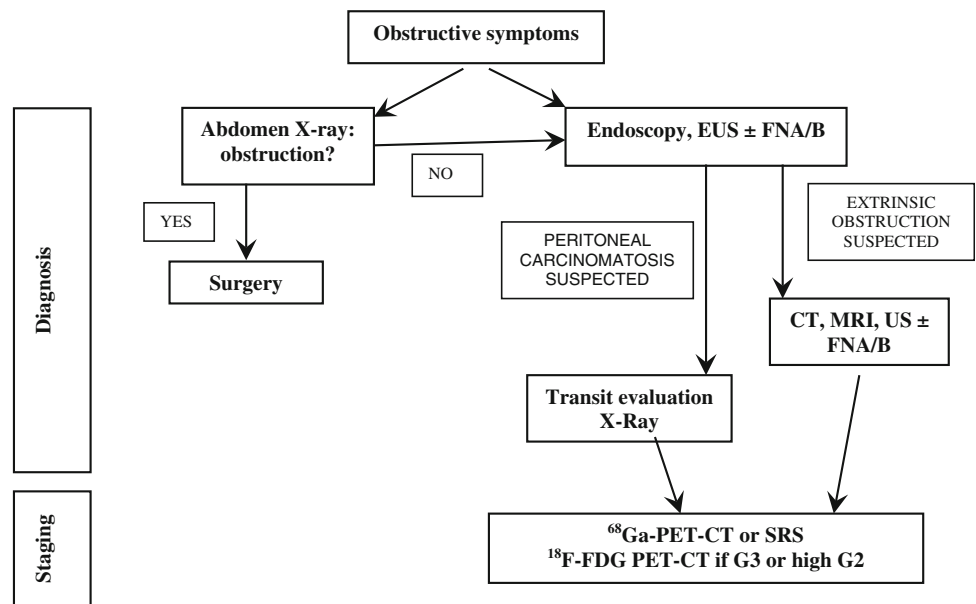


Fig. 6 Diagnostic flow-chart for GEP-NEN suspected after subocclusive picture



of the liver or other site lesions or laparoscopy-guided biopsy should be discussed in a multidisciplinary team.

3.2.2.3 Jaundice This clinical presentation points to the involvement of the liver, biliary tract or pancreas. Liver function and structure should be assessed by blood tests and US, to rule out the obstruction of the biliary tract. Compressive effects of lymphadenopathies or a pancreatic mass may cause an extra-hepatic tract dilatation, whereas liver metastases are more likely related to an intra-hepatic tract dilatation [96, 172]. In case of obstructive jaundice, a cholangio-MRI and endoscopic-retrograde-cholangio-pancreatography (ERCP) can be considered. Cytology by means of brushing or histology can be obtained through ERCP. Whole-body CT scan and endoscopy should be

used to define the primary site of the tumor and for staging purpose (Fig. 7).

3.2.2.4 Gastrointestinal bleeding It can be related to the compressive and infiltrating effects of a tumor mass. Bleeding can be massive (hematemesis, melena and rectal bleeding) or dripping and occult. Blood tests, iron assessment and endoscopy must be performed. Massive bleeding always requires hospitalization and may require angiography [73, 173]. In case of lesions located in the stomach-duodenum or in terminal ileum-colon tract, a histologic diagnosis may be obtained through biopsy during EGDS or ileo-colonoscopy. If upper and lower endoscopy is negative, enteroscopy, enteroCT/MRI, VCE should be discussed in the multidisciplinary team according to the local

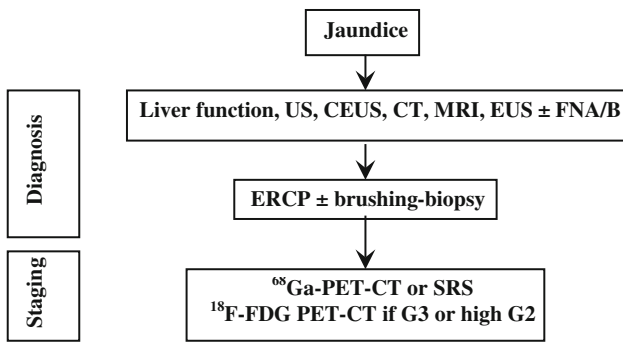


Fig. 7 Diagnostic flow-chart for GEP-NEN suspected after jaundice

availability and expertise (Fig. 8). For lesions located in the small bowel, a surgical diagnostic/therapeutic approach should be considered

We recommend to consider GEP-NEN as possible causes of isolated abdominal pain, subocclusive symptoms, jaundice or gastrointestinal bleeding.
We recommend to obtain a histologic or cytological diagnosis.
We recommend a specific diagnostic work-up, according to clinical presentation.

3.3 Symptomatic patient with syndromes

3.3.1 Diarrhea and flushing

3.3.1.1 Clinical approach: when to suspect a GEP-NEN

The patient with diarrhea and flushing should raise the suspicion of CS (Box 2).

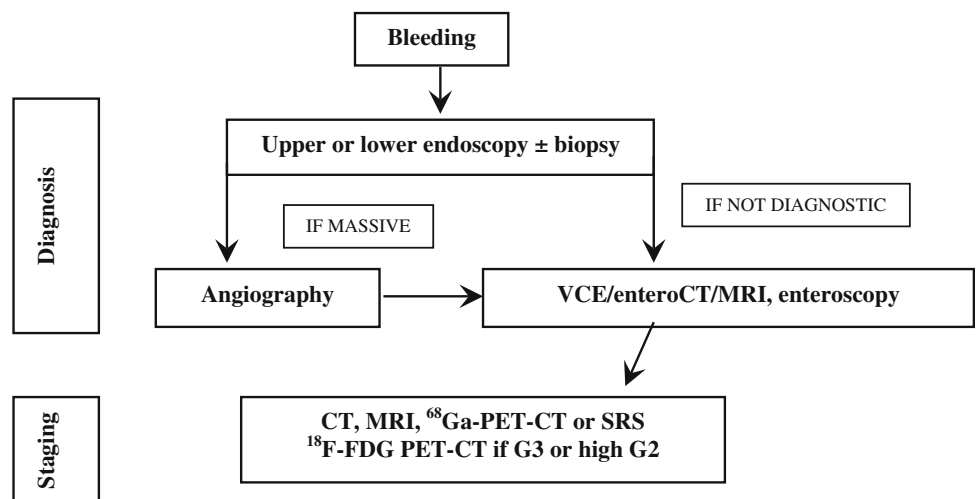
Carcinoid syndrome diagnosis may be difficult. A detailed history and complete physical examination are must. Symptoms may be under-reported by patients or be attributed to other, more common GI disorders. Differential diagnoses include irritable/inflammatory bowel diseases, microscopic colitis, food intolerance/allergy, bacterial overgrowth, celiac disease, hypersecretory states (i.e., gastrinoma, see “Resistant/relapsing ulcer disease”), chronic pancreatitis, other neoplastic (i.e., colon carcinoma, lymphoma) and non-neoplastic conditions (asthma, anxiety, alcoholism) [6].

Diarrhea in patients with CS is chronic, predominantly secretory, does not change with fasting, and is associated with fluid and electrolyte imbalance. A detailed history of the diarrhea and specific questioning about other possible manifestations of CS (i.e., facial flushing) are required. The stools are usually watery and result from intestinal hypermotility and hypersecretion. Nocturnal diarrhea is generally considered as characteristic of CS. The incomplete response to antidiarrhoic treatment should raise the suspicion of possible CS [174].

Flushing is the most common symptom in CS. Eating, emotion, alcohol, and exercise may worsen flushing. The face, neck and upper trunk usually turn pink to red in color and the skin is characteristically dry. Flushing may also be associated with transient hypotension and bronchoconstriction. Other causes of flushing/sweating disorders to be considered are [175]:

- pheochromocytoma, menopause, ZES, and medullary thyroid carcinoma (intermittent flushing);
- alcoholism, polycythemia, mitral stenosis, and Cushing’s syndrome (constant flushing).

Fig. 8 Diagnostic flow-chart for GEP-NEN suspected after GI bleeding



We recommend considering CS in patients with chronic diarrhea and/or flushing. The additional presence of abdominal pain and wheezing strengthens the suspicion.

Box 2

Carcinoid syndrome

CS is classified as typical or atypical, accounting for 95% and 5% of total cases, respectively [176,177,178]. Typical CS occurs in about 15–20% of patients with jejuno-ileal NENs, with liver metastases. In less than 5%, it can be caused by retroperitoneal or ovarian metastases that release serotonin or tachykinin, bypass the liver and enter the systemic circulation [179, 180, 181, 182]. These so-called “functioning carcinoids” exhibit a variable clinical presentation, due to the type of secreted bioactive substances (serotonin, tachykinins, kallikreins, and prostaglandins). Typical CS may present with cutaneous flushing (face, neck, upper chest), GI hypermotility with pain (intermittent and crampy, described as dull, achy and colicky, and not relieved by defecation), telangiectasia, peripheral edema, wheezing, cyanosis, pellagra, and right-sided heart failure caused by cardiac valve abnormalities. Symptoms may occur spontaneously or be triggered by alcohol intake, serotonin-rich foods, and exercise [182]. Atypical CS is associated to overproduction of histamine and is characterized by prolonged flushing, bronchoconstriction and hypotension [178]. Wheezing might suggest asthma that can be identified by lung function tests. Carcinoid crisis is an extreme and life-threatening expression of the CS, induced by the massive release of amines into the circulation following anesthesia, interventional procedures or medication [183]. Main features of carcinoid crisis are: hypotension, rarely hypertension, tachycardia, bronchial wheezing, and central nervous system dysfunction [184]. Carcinoid heart disease affects 10–20% of the patients at presentation. CS causes a thickening of the heart valves, impairing their proper function, resulting in insufficiency. Heart failure typically involves the right-side valves. Signs and symptoms include fatigue and shortness of breath during physical activity and peripheral edema in 1 out of 5 patients. Up to 50% of deaths in CS are due to heart failure [185, 186].

3.3.1.2 Work-up in the patient with suspected carcinoid syndrome Before proceeding to the work-up, other causes of flushing with or without diarrhea must be excluded (Table 7) [187]. To this aim it could be useful a 2 to 4-week detailed self-recording of the flushing and diarrhea episodes.

Since symptoms associated with CS can be triggered by alcohol intake and serotonin-rich foods [188–190], the patient should follow an exclusion diet for at least 3 days before starting urinary collection for 5-HIAA and should avoid for at least 24 h (or according to half-life) drugs that affect this test (see Table 4).

Biochemical testing: Urinary excretion of 5-HIAA is the most useful test in patients with typical CS due to jejuno-ileal NENs. Atypical CS is induced by gastroduodenal and bronchial NENs that only rarely secrete serotonin because they lack DOPA-decarboxylase, the enzyme that converts 5-hydroxytryptophan into serotonin [191]. These tumors may thus produce 5-hydroxytryptophan and histamine instead of serotonin, but no assay for urinary 5-hydroxytryptophan is commercially available, whereas histamine assays are limited to very few centers.

5-HIAA testing is highly sensitive (up to 90%) and specific (85–90%) for the diagnosis of CS. In patients with CS 5-HIAA levels are usually at least twice as high as the upper normal limit. They may reflect the tumor burden and are rarely normal in patients with CS [57, 73, 76, 77, 192–194]. Attention must always be paid to factors causing falsely high or low levels (see Table 4).

Serum serotonin determination is not recommended because it may vary considerably according to activity and stress levels [73]. CgA is poorly specific whereas NSE has no diagnostic role [77, 193, 195].

Imaging procedures: Carcinoid syndrome is most frequently due to a NEN in the small bowel associated with liver metastases [196]. Therefore liver assessment examinations should be firstly performed, including US/CEUS (useful to drive biopsy), CT and MRI (superior to CT for small lesions) [95, 197–200].

The type of work-up aimed to the detection of the primary tumor (see “[Work-up in the patient with metastatic disease and unknown primary tumor](#)”) and to rule out atypical situations should be discussed in a multidisciplinary panel, also taking into account the possible surgical resection.

Functional imaging studies (SRS, or when available ^{68}Ga -DOTA-peptide-PET) may help in localizing the

Table 7 Differential diagnosis of flushing

Drugs	All vasodilators, calcium channel blockers, morphine and other opiates, etc.
Menopause	Associated with sweating
Mastocytosis	Flushing lasting longer than CS, may be accompanied by headache, dyspnea, palpitations, abdominal pain and diarrhea
Medullary thyroid carcinoma	Associated with diarrhea in patients with advanced disease
Pheochromocytoma	Rare, but it may occur after a paroxysm of hypertension, tachycardia and palpitations and is preceded by pallor

primary tumor and small metastases, and as a predictive factor for somatostatin receptor driven therapies. Combination of SRS or PET with CT increases the sensitivity [117].

In case of persistently negative results of morphological and functional studies, the primary tumor may be located by intraoperative palpation [73].

Transthoracic echocardiography should be performed at diagnosis of CS and then annually to detect any right-sided fibrosis involving tricuspid and pulmonary valves [201] (Fig. 9).

We recommend to rule out other causes of flushing and diarrhea (history and physical examination) before proceeding with work-up of CS.

We recommend to perform urinary 5-HIAA and serum CgA plus abdomen US as first step if CS is clinically suspected.

We recommend against routine use of other circulating biomarkers.

We recommend contrast-enhanced abdominal CT scan or MRI plus SSTR-related imaging when first step studies are positive.

If first step studies are negative, **we suggest** considering other diagnoses, atypical CS or very rare non-metastatic NENs in a multidisciplinary team, involving a referral center when required.

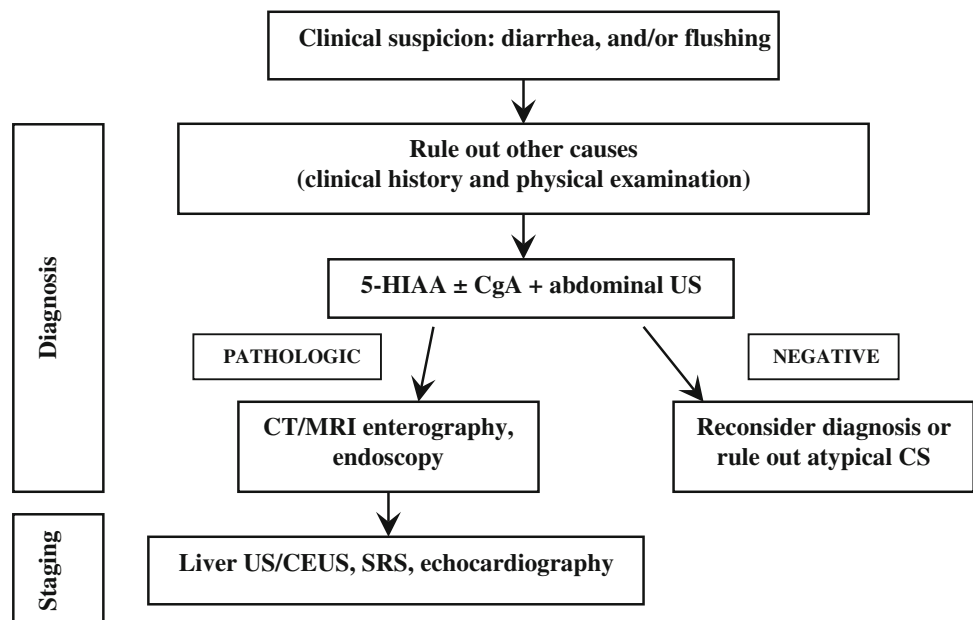
We recommend echocardiography for each patient with CS.

3.3.2 Resistant/relapsing ulcer disease

3.3.2.1 Clinical approach: when to suspect a GEP-NEN ZES (Box 3) is characterized by gastric acid hypersecretion resulting in severe peptic disease and gastroesophageal reflux disease (GERD) [202–205].

The majority of ZES patients presents with a single duodenal ulcer, peptic symptoms, GERD symptoms or ulcer complications and diarrhea. Multiple ulcers or ulcers in unusual locations are a less frequent presenting feature than in the past [8, 84, 85, 87, 202, 204–208]. With the widespread use of gastric antisecretory drugs, particularly PPIs and H2RAs, symptoms may be masked. The diagnosis is most often suggested by a long history of peptic ulcer disease or GERD symptoms or their recurrence after treatment [83–85, 206, 208]. This delay may postpone the diagnosis of gastrinoma to a higher stage of the disease.

Fig. 9 Diagnostic flow-chart for suspected carcinoid syndrome



We recommend considering ZES (marker of gastrinoma) in case of:

- Recurrent, severe or familial peptic ulcer disease;
- Or peptic ulcer disease:
 - without *Helicobacter pylori* or other risk factors;
 - associated with severe GERD;
 - resistant to treatment or associated with complications (perforation, penetration, bleeding);
 - associated with endocrinopathies or diarrhea (which promptly resolved under PPIs);
 - with prominent gastric folds at endoscopy;
- MEN-1.

We recommend MEN-1 be suspected in patients with refractory peptic ulcer disease or a confirmed ZES.

Box 3 **Gastrinoma**

Gastrinoma is a functioning GEP-NEN, usually located in the duodenum or pancreas that secretes gastrin and causes a clinical syndrome known as ZES.

The incidence of gastrinomas is 0.5–2/million population/year. Gastrinoma is one of the most common functioning GEP-NEN in the general population [8] and occurs in 25–40 % of subjects with MEN-1 [207, 209]. ZES occurs at an earlier age (mean 32–35 years) in patients with MEN-1 than in those with sporadic disease [204, 207, 209].

Pancreatic gastrinomas may occur in any portion of the pancreas, while duodenal gastrinomas are predominantly found in the first part of the duodenum including the bulb [210, 211]. At surgery, 70–85 % of gastrinomas are found in the right upper quadrant (duodenal and pancreatic head area), the so-called “gastrinoma triangle” [210, 211, 212].

The main symptoms classically associated to ZES are due to gastric acid hypersecretion and are represented by abdominal pain (75–98 % of the cases), diarrhea (30–73 %), heartburn (44–56 %), bleeding (44–75 %), nausea/vomiting (12–30 %), and weight loss (7–53%) [85, 204, 205].

At presentation, >97 % of patients have an elevated fasting serum gastrin (FSG) level, 87–90 % have marked gastric acid hypersecretion (basal acid output >15 mEq/h) and 100 % have a gastric acid pH <2 [202, 213].

The rate of malignancy is high with liver metastases in 30–40 % of cases [214].

hyperparathyroidism in MEN-1 [40], serum calcium and PTH are the first step to rule out the diagnosis.

Biochemical testing: Fasting serum gastrin is an excellent screening test (>98 % sensitivity). False positive conditions should always be excluded (Table 5). The diagnosis of ZES requires inappropriately elevated FSG levels in association with a >15 mEq/h (>5 mEq/h in gastrectomized patients) basal acid output or in association with a gastric pH <2.0. Under these conditions, FSG >1,000 pg/mL means a certain diagnosis of ZES. On the contrary, a gastric pH >2.0 virtually excludes ZES [84]. In subjects under chronic therapy with PPIs these drugs have to be withdrawn for at least 1 week [84, 216], although the optimal wash-out time for PPIs should be longer (4 weeks). H2RAs exert a less pronounced suppression of gastric acid output than PPIs [217, 218]. In case of subjects on PPIs who are at risk of bleeding ulcer, diarrhea with dehydration or hypokalemia, these drugs may be replaced with H2RAs for at least 1 week under medical supervision [219, 220].

Secretin test (2 U/kg rapid infusion), a gastrin provocative test, may be performed in controversial cases [77, 221]. Withdrawal of antacid and anticholinergic drugs (12 h), and of PPIs (1 week) is recommended [222]. The secretin test is positive when a >120 pg/mL increase of FSG over the basal value is found (sensitivity 94 %, specificity 100 %) [88, 223]. Calcium stimulation test (5 mg/kg body weight per hour, infused over 3 h, increase >395 pg/mL over the basal FSG as cut-off) may alternatively be used. However, it is hampered by lower sensitivity, specificity and higher side effects [88]. Gastric acid secretion stimuli are no longer performed [203].

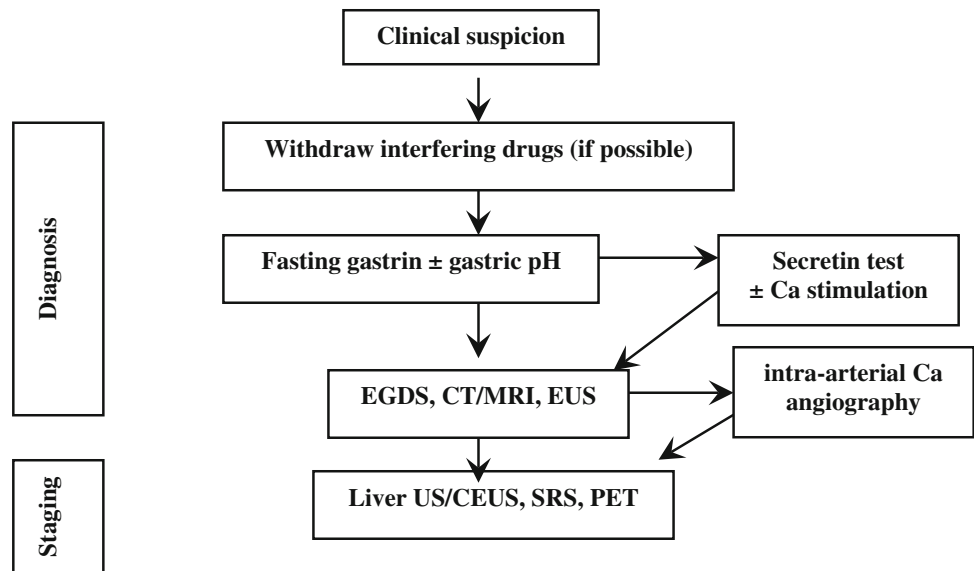
Imaging: After biochemical diagnosis, EGDS is required. In ZES, peptic ulcer disease is found distally to the duodenal bulb within the descending part of the duodenum or even further distally within the jejunum. Peptic ulcers frequently occur in groups indicating some substantial acid hypersecretion [84].

The following imaging procedures may be used to localize the primary tumor, determine the extent of the disease, evaluate indication to surgery, and assess response

3.3.2.2 Work-up in the patient with suspected gastrinoma History and clinical examination are the first steps in the diagnosis of ZES. The use of acetylsalicylic acid and other non-steroidal anti-inflammatory drugs, which can mimic a ZES picture, should be ruled out [215].

Multiple endocrine neoplasms should be considered in all patients with ZES, especially in case of familial or personal history of endocrine disease, kidney stones, other NENs [88, 207]. Due to high penetrance of primary

Fig. 10 Diagnostic flow-chart for suspected gastrinoma



to treatments [88]: (1) contrast-enhanced CT and/or MRI, EUS; (2) functional imaging (SRS, PET); and (3) selective intra-arterial calcium injection angiography [84, 88].

Accurate localization of the tumor can result in complete surgical resection, decreased rate of developing lymph node metastases, and increasing survival [88, 222, 224–226] (Fig. 10).

We recommend fasting serum gastrin as the first step in patients with clinical suspicion of ZES.

We recommend exclusion of all other causes of hypergastrinemia before proceeding with the diagnostic work-up.

We suggest secretin test when the diagnosis of ZES is unclear/controversial.

We recommend morphological and functional imaging in all patients with biochemically established ZES.

3.3.3 Spontaneous hypoglycemia

3.3.3.1 Clinical approach: when to suspect a GEP-NEN Hypoglycemia (plasma glucose <60 mg/dL on a venous blood sample) is an uncommon clinical problem in non-diabetic adults. The presence of symptoms reinforces the clinical relevance of this finding because some normal subjects may have an asymptomatic low glucose level after prolonged fasting. Symptoms may be due to sympathoadrenal activation (“adrenergic symptoms”, i.e., sweating, shakiness, tachycardia, anxiety, hunger) and/or neurogly-

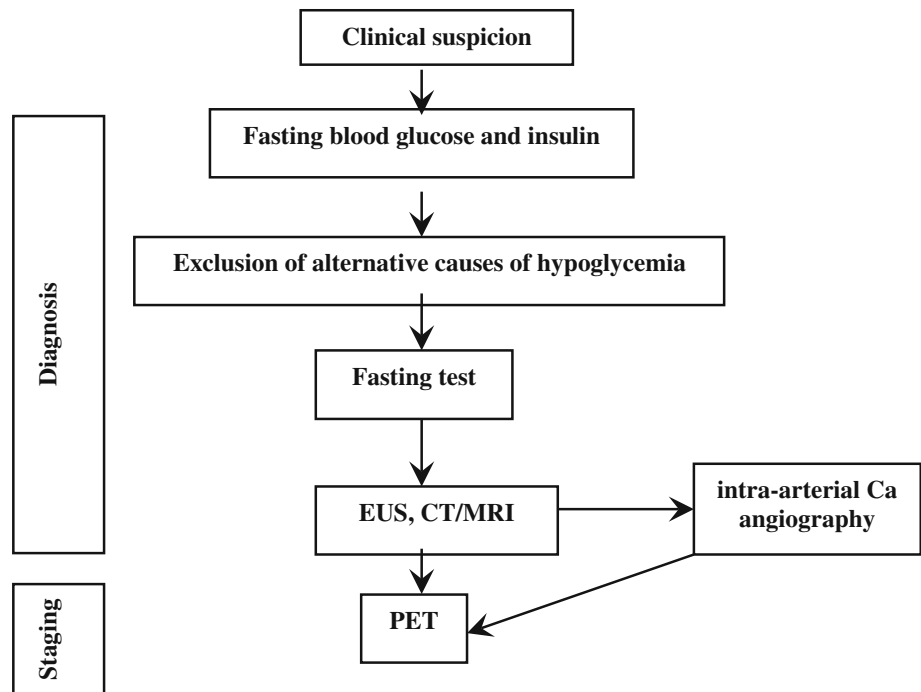
copenia (weakness, dizziness, inappropriate behavior, altered concentration, confusion, blurred vision and, in extreme cases, coma and death) [227–229]. Symptoms may present at a variable glucose level (generally as low as <55–60 mg/dL) [227, 228, 230, 231].

Hypoglycemia may be due to several conditions beyond insulin-secreting tumors [232] (Table 8).

Insulinoma (Box 4) should be strongly suspected in presence of the Whipple triad, which occurs in about 75 % of patients and combines (1) symptoms of hypoglycemia, (2) low blood sugar concurrent with symptoms, and (3) reversal of symptoms after glucose administration [233]. Neuroglycopenic symptoms usually dominate the clinical picture so that insulinoma may be misdiagnosed with cognitive impairment, psychiatric illnesses or seizure disorders. Frequently, the occurrence of bizarre behavior or confusion states is more precisely described by concerned relatives or friends than by the patient himself. Adrenergic and neuroglycopenic symptoms may coexist, especially in the early phase of the disease. A detailed description of pure adrenergic symptoms, however, is more specific of a “functional hypoglycemia”.

Hypoglycemic symptoms occur most frequently at night and/or early morning and, anyway, in a protracted fasting state. Yet, the occurrence of post-prandial hypoglycemia does not exclude an insulinoma [234, 235]. Symptoms can be worsened by exercise, alcohol, hypocaloric diet, and by concomitant clinical conditions or use of drugs (see above) [236, 237]. Weight gain occurs in 20–40 % of patients that may develop overweight because of hyperinsulinism.

Fig. 11 Diagnostic flow-chart for suspected insulinoma



We recommend the immediate determination of blood glucose and insulin in any subject with acute change in mental status, especially if recurrent.

We recommend the exclusion of all alternative causes of hypoglycemia before starting a work-up for insulinoma.

We suggest considering insulinoma as a probable cause in patients with predominant neuroglycopenic symptoms, recurrent fasting hypoglycemia, and weight gain.

Box 4
Insulinoma [236, 237]
Insulinoma is a NEN arising from insulin-secreting cells in pancreatic islets. Other hormones and metabolites (gastrin, ACTH, glucagon, hCG, somatostatin, and 5-HIAA) may be also secreted from this neoplasm.
About 90 % of insulinomas are benign. In rare cases neither a single nor multiple tumors can be identified and the syndrome depends on diffuse beta-cell hyperplasia. In malignant forms with liver metastases, a 16–26 months survival is to be expected. Only 5 % of all insulinomas are associated with MEN-1; in case of multiple insulinomas (near 10%), MEN-1 prevalence raises to 50 %.

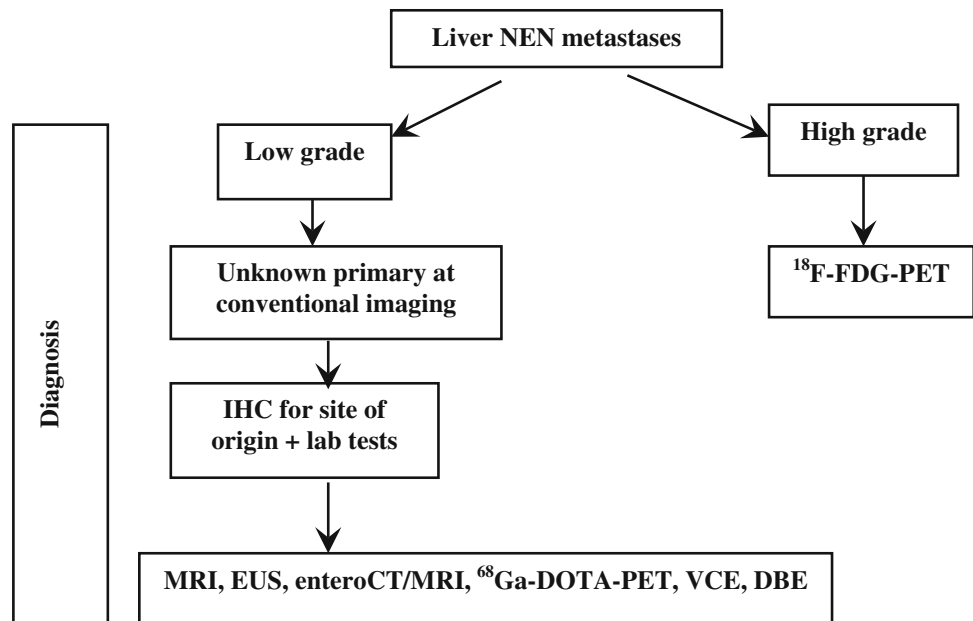
Table 8 Differential diagnosis of hypoglycemia

Drugs	Insulin, oral hypoglycemic drugs Quinine, pentamidine, indomethacin, lithium More rarely: ACE-inhibitors, levofloxacin, trimethoprim-sulfamethoxazole, and heparin
Excessive alcohol consumption	Block of stored glucose release
Liver, kidney or heart failure	Depletion of substrates required for gluconeogenesis
Long-term starvation (anorexia nervosa)	Depletion of substrates required for gluconeogenesis
Non-islet cell tumors	Excessive production of IGF-II that causes the use of too much glucose
Gastric surgery (post-gastric bypass)	Accelerated transit and malabsorption
Hypoadrenalism and hypopituitarism	Deficiency of hormones that regulate glucose production
Insulin autoimmune hypoglycemia	

3.3.3.2 *Work-up in the patient with suspected insulinoma* Biochemical assessment: Symptoms and/or signs suggesting hypoglycemia combined with a ≤ 55 mg/dL

(3.0 mmol/L) plasma glucose, a ≥ 3.0 μ U/mL (18 pmol/L) plasma insulin, a ≥ 0.6 ng/mL (0.2 nmol/L) C-peptide, and a ≥ 5.0 pmol/L proinsulin indicate endogenous hyperinsulinism [227, 228, 231]. Exogenous insulin-induced hypoglycemia is always associated with low levels of C-peptide. In patients with insulinoma, proinsulin corresponds to about 70 % of insulin immunoreactivity, whereas it is normally limited to 20 %.

Fig. 12 Diagnostic flow-chart in the patient with metastatic disease and unknown primary tumor



Blood and urine assays for sulfonylureas will detect factitious hypoglycemia caused by these drugs. Pituitary and adrenal function tests are useful to rule out hypoadrenalism and hypopituitarism [227, 228, 230, 238].

Provocative tests: Biochemical diagnosis is based on lack of suppression of endogenous insulin secretion by hypoglycemia [239] and inappropriately elevated insulin level during hypoglycemia is the diagnostic key point. In 95 % of cases, the diagnosis is achieved only during prolonged fasting (up to 72 h) inducing symptomatic hypoglycemia [240]. The test should be performed on inpatients under close supervision and with regular control of glycaemia and mental status. A ≥ 3 $\mu\text{U/mL}$ (≥ 18 pmol/L) insulin value, in the presence of glucose level < 55 mg/dL has recently been proposed as diagnostic cut-off [223]. Plasma β -hydroxybutyrate levels ≤ 2.7 mmol/L may confirm the diagnosis, demonstrating the suppressive effect of insulin on ketogenesis even during a protracted fasting [231].

At the end of the 72-h fasting test, in the absence of hypoglycemia, the use of stimulation tests was proposed [231]. Stimulation tests, e.g., tolbutamide, glucagon or calcium, are not recommended because they may induce a prolonged and refractory hypoglycemic condition, but long-term fasting can be finished after 72 h with bicycle test.

Imaging: In all patients with a confirmed biochemical diagnosis, imaging is indicated to localize the tumor [241].

Since 80 % of insulinomas are < 2 cm in size, they are frequently missed by high-resolution transabdominal US (50 % sensitivity), while EUS is more sensitive (77 %) and should be preferred [242]. Helical or multislice CT and

MRI offer a comparable (82–94 %), but incomplete, sensitivity [243, 244]. Selective arteriography has an 82 % sensitivity and a 95 % specificity.

Due to small size and/or lack of SSTR2 expression in 50 % of insulinoma [151], SSTR-related imaging plays a minor role than morphological imaging. DOPA-PET has been proposed as an alternative tool [245]. Radiolabelling with ^{111}In -labeled glucagon-like peptide-1 receptors agonist (^{111}In -DOTA-exendin-4) is a promising technique, still not routinely used [246].

Arteriography combined with selective calcium stimulation: Calcium is able to stimulate insulin release from neoplastic tissue, but not from normal islets. Hence, the catheterization of the arterial branches of the celiac system and the measurement of insulin in the blood sampled from hepatic veins during selective intra-arterial calcium injection localize the pancreatic area nesting the tumors in 88–100 % of cases [34, 247, 248]. This test is cumbersome, expensive and poorly available. Accordingly, it should be reserved only to selected, biochemically proved cases with negative imaging studies.

In spite of all the above reported diagnostic techniques, only 60–70 % of patients have a successful preoperative localization. In patients with less threatening symptoms that are fairly controlled by medical treatment a close surveillance may be advisable. In severely symptomatic cases, the use of intraoperative US and the pancreatic exploration conducted by an experienced surgeon identifies more than 90 % of the insulin-secreting tumors [242, 249] (Fig. 11).

We recommend the simultaneous evaluation of blood glucose, insulin and C-peptide to detect endogenous hyperinsulinemia in all patients with spontaneous hypoglycemia.

We recommend a prolonged fasting test (up to 72 h) in patients referred for a previous hypoglycemic episode who are free of symptoms at the moment of medical examination.

We recommend against the use of stimulation tests for the diagnosis of insulinoma.

We recommend the use of localization tests (CT/CEUS and EUS) only after the biochemical diagnosis of insulinoma is established.

3.4 Work-up in the patient with metastatic disease and unknown primary tumor

Unknown primary NEN (UPN) is a condition of metastatic histologic or cytological confirmed NEN without evidence of a primary site after a first diagnostic work-up, including chest-abdomen CT scan, SRS, and upper and lower endoscopy.

The frequency of well-differentiated UPNs ranges from 9 to 19 % [250, 251]. The presence of liver metastases largely influences prognosis in all types of NENs and is dependent on primary tumor site, tumor extent (T-stage), and histologic differentiation (NET vs. NEC). Reported survival rate at 5 years of G1–G2 small intestinal and pancreatic NENs in the SEER database is 54 and 27 %, respectively [252]. Furthermore, survival is reportedly worse in UPN patients as compared to patients with liver metastases whose primary NEN is known [253]. In liver metastatic patients survival rate is influenced by the presence of obstructive symptoms or symptoms related to peptide secretion.

The evaluation of a patient with UPN should encompass a detailed clinical history, including family history to identify affected relatives and a patient's increased risk for endocrine tumors (i.e., MEN type 1 or 2), laboratory and radiographic studies [254].

Histologic preparations should be reevaluated by IHC to guide the search for the primary tumor: TTF-1 (pulmonary or medullary thyroid carcinoma), CDX-2 (intestinal), PAX-8, histidine-decarboxylase (pancreatic), xenin (duodenal), gastrin (occult gastrinoma), and PP/glucagon (pancreatic) [38, 255]. Biochemical work-up may include 5-HIAA, gastrin, and other locally available tumor markers [256].

It has been recently reported that most UPNs are derived from pancreas and small bowel [257]. Accordingly, further

investigations for localizing the primary site in well-differentiated NENs might include abdomen MRI, EUS, enteroCT/MR, ⁶⁸Ga-PET, VCE, DBE to be shared within a multidisciplinary team according to clinics, local availability and expertise [124, 258, 259]. In NECs ¹⁸F-FDG-PET may be useful (Fig. 12)

We recommend biopsy at the metastatic site with histologic and IHC examination as a first step to confirm NEN diagnosis.

We recommend a detailed clinical history to elicit signs or symptoms that could point to the primary site (carcinoid syndrome, asthma, diarrhea, etc.), as well as a complete family history.

In all cases of unknown primary of low grade metastatic liver NEN after a conventional imaging, including chest-abdomen CT scan, upper and lower endoscopy and SRS, **we recommend** an IHC for site of origin, and imaging according to results.

3.5 When and how to stage a previously diagnosed GEP-NEN

Evaluation of disease extension has a pivotal role in treatment planning.

Pre-treatment staging should include morphologic and functional imaging. Morphological imaging is required for all GEP-NENs, irrespectively of their grade. SSTR-based functional imaging (SRS or ⁶⁸Ga-DOTA-peptide-PET) should be used for low-/intermediate-grade GEP-NENs (WHO 2010 G1-G2), whereas ¹⁸F-FDG-PET should be preferentially used in G3 GEP-NENs and in some G2 cases.

For morphologic staging, a chest-abdomen-pelvis multidetector CT or a chest basal CT plus abdomen-pelvis MRI should be used [87]. For functional staging, SRS using ¹¹¹In-pentetreotide (Octreoscan®) is presently regarded as the gold standard. However, if available, ⁶⁸Ga-DOTA-peptide-PET with simultaneous CT should be preferred to SRS. In facts, PET lacks the anatomic details required for therapeutic stratification (surgical planning or dose calculation for radioembolization with radiolabeled microspheres). Recently, MRI with liver-specific contrast combined with ⁶⁸Ga-DOTA-peptides-PET has been reported to be more accurate than PET-CT to detect GEP-NEN hepatic metastases [260].

¹⁸F-DOPA-PET-CT and ¹¹C-5HTP-PET-CT are promising tools. Their use might be considered if results of SRS or ⁶⁸Ga-DOTA-peptides-PET are negative [261].

Gastric NENs In small (<1 cm) type 1 and type 2 tumors, EGDS is usually the only recommended imaging

procedure [153]. Tumor invasiveness through the gastric wall must be evaluated with EUS study: it is recommended before resection for polyps >1 cm in diameter. EUS is also useful in assessing regional lymph nodes involvement, and allows histological confirmation by FNA. Type 1 tumors do not require either abdomen multislice CT or MRI, or SRS/⁶⁸Ga-DOTA-peptides-PET; these imaging studies should be performed for type 2 and type 3 neoplasm staging.

Duodenal NENs EUS is useful before resection of polypoid lesions; multislice abdomen CT or MRI should be performed to assess local and distant disease extension. In patients with local advanced neoplasm and/or liver metastases, bone scan and MRI of spine and pelvis should be performed [153].

Jejuno-ileum NENs Chest-abdomen-pelvis CT scan or chest basal CT scan and abdomen-pelvis MRI, SRS or ⁶⁸Ga-DOTA-peptides-PET should be performed looking for distant metastases [73]. Liver CT scan should be performed by multislice and multiphase technique. Colonoscopy should be performed to rule out synchronous colorectal carcinoma.

Colorectal NENs Chest-abdomen-pelvis multislice CT should be carried out. Endoanal/rectal US is very useful for assessing preoperatively the depth of tumor invasion in the rectal wall and regional lymph node involvement [173].

NF pNENs For morphologic staging a multislice/multiphase CT or fat-saturated T1-weighted and delayed enhanced T1-weighted MRI can be performed and EUS with biopsy [262]. Afterwards, SRS or ⁶⁸Ga-DOTA-peptides-PET should be performed.

We recommend contrast enhanced chest-abdomen-pelvis multidetector CT scan or basal chest CT scan plus abdomen-pelvis MRI in pre-treatment staging of GEP-NENs.

We recommend ⁶⁸Ga-DOTA-peptides-PET-CT for functional staging of well-differentiated GEP-NENs, or, if not available, ¹¹¹In-pentetreotide (Octreoscan®) scintigraphy.

If SSTR-related imaging is negative, **we suggest** further functional staging with alternative radiotracers.

We suggest ¹⁸F-FDG-PET-CT for staging of selected G3 and high G2 GEP-NENs.

We recommend EUS study for local staging of >1 cm gastric, duodenal, and rectal polypoid NEN lesions.

4 Conclusions

The management of patients with GEP-NENs poses a significant challenge to clinicians from the very start of the diagnostic work-up. The wide heterogeneity of disease presentation, with a majority of asymptomatic patients and poorly specific clinical pictures may account for a delay in definite diagnosis and appropriate treatment. The present document has, therefore, been drawn with the purpose of offering a practical guide to physicians facing the suspicion of GEP-NENs, in light of the available clinical evidence and experience. Of course, many questions are still to be fully answered and many others still to be addressed in the near future, as we move forward to new promising techniques and diagnostic tools. For these reasons, in spite of its goal as a state-of-the-art update, our document has not been conceived as the repository of the “ultimate truth” in the field of GEP-NENs diagnosis. Instead, much attention has been devoted to the logical framework, which should back up the clinical reasoning. Furthermore, the diagnosis of GEP-NENs is heavily based on the contribution of a wide range of know-how and skills provided by different specialists. The core team may include a varying combination of different specialists, according to the local expertise and facilities; nevertheless, the pathologist plays a key role in the diagnosis and classification of GEP-NENs, because his/her information is critical to guide the prognosis and treatment planning. Hence, a multidisciplinary team model is recommended as the best opportunity to reach an accurate, safe and cost-effective diagnosis, likely to improve the outcome of patients with GEP-NENs.

In conclusion, the Italian Association of Clinical Endocrinologists (AME) hopes the present Position Statement will constitute an effective tool in helping the clinical management of patients with GEP-NENs. Further implementations and updates of this document will follow as new evidence and progress in the field emerge.

Other members of AME oncologic endocrinology group Giorgio Borretta, Cuneo; Renato Cozzi, Milan; Giuseppe Francia, Verona; Rinaldo Guglielmi, Albano Laziale; Gabriele Luppi, Modena; Salvatore Monti, Rome; Silvia Nasoni, Albano Laziale; Micaela Pellegrino, Cuneo; Anna Pia, Turin; Sara Pusceddu, Milan; Valeria Ramundo, Naples; Francesco Scavuzzo, Naples; Alessandro Scoppola, Rome; Ettore Seregni, Milan; Francesca Spada, Milan; Laura Tonutti, Udine; Vincenzo Toscano, Rome; Maria Chiara Zatelli, Ferrara.

Conflict of interest The authors declare they have no conflict of interest.

Open Access This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

References

- Modlin IR, Lye KD, Kidd M (2003) A 5-decade analysis of 13,715 carcinoid tumors. *Cancer* 97:934–959 (LoE ⊗⊗⊗)
- Hauso O, Gustafsson BI, Kidd M, Waldum HL, Drozdov I, Chan AKC, Modlin IM (2008) Neuroendocrine tumor epidemiology. Contrasting Norway and north America. *Cancer* 113:2655–2664 (LoE ⊗⊗⊗)
- Yao JC, Hassan M, Phan A, Dagohoy C, Leary C, Mares JE, Abdalla EK, Fleming JB, Vauthey JN, Rashid A, Evans DB (2008) One hundred years after ‘carcinoid’: epidemiology of and prognostic factors for neuroendocrine tumors in 35,825 cases in the United States. *J Clin Oncol* 26:3063–3072 (LoE ⊗⊗⊗)
- Garcia-Carbonero R, Capdevila J, Crespo-Herrero G, Díaz-Pérez JA, Martínez Del Prado MP, Alonso Orduña V, Sevilla-García I, Villabona-Artero C, Beguiristain-Gómez A, Llanos-Muñoz M, Marazuela M, Alvarez-Escola C, Castellano D, Vilar E, Jiménez-Fonseca P, Teulé A, Sastre-Valera J, Benavent-Viñuelas M, Monleon A, Salazar R (2010) Incidence, patterns of care and prognostic factors for outcome of gastroenteropancreatic neuroendocrine tumors (GEP-NETs): results from National Cancer Registry of Spain (RGETNE). *Ann Oncol* 21:1794–1803 (LoE ⊗⊗⊗)
- Lawrence B, Gustafsson BJ, Kidd M, Pavel M, Svejda B, Modlin IM (2011) The clinical relevance of chromogranin A as a biomarker for gastroenteropancreatic neuroendocrine tumors. *Endocrinol Metab Clin North Am* 40:111–134
- Öberg K, Knigge U, Kwekkeboom D, Perren A, ESMO Guidelines Working Group (2012) Neuroendocrine gastro-entero-pancreatic tumors: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 23(Suppl 7):124–130
- Cho M-Y, Kim JM, Sohn JH, Kim MJ, Kim KM, Kim WH, Kim H, Kook MC, Park Y, Lee JH, Chang H, Jung ES, Kim HK, Jin SY, Choi JH, Gu MJ, Kim S, Kang MS, Cho CH, Park do MI, Kang YK, Kim YW, Yoon SO, Bae HI, Joo M, Moon WS, Kang DY, Chang SJ (2012) Current trends of the incidence and pathological diagnosis of gastroenteropancreatic neuroendocrine tumors (GEP-NETs) in Korea 2000–2009: multicentre study. *Cancer Res Treat* 44:157–165
- Faggiano A, Ferolla P, Grimaldi F, Campana D, Manzoni M, Davì MV, Bianchi A, Valcavi R, Papini E, Giuffrida D, Ferone D, Fanciulli G, Arnaldi G, Franchi GM, Francia G, Fasola G, Crinò L, Pontecorvi A, Tomassetti P, Colao A, Natural history of gastro-entero-pancreatic and thoracic neuroendocrine tumors (2012) Data from a large prospective and retrospective Italian epidemiological study: the NET management study. *J Endocrinol Invest* 35:817–823 (LoE ⊗⊗⊗)
- Lepage C, Bouvier AM, Faivre J (2013) Epidemiology of malignant digestive neuroendocrine tumours. *Eur J Endocrinol* 168:R77–R83
- Niederle MB, Hackl M, Kaserer K, Niederle B (2010) Gastroenteropancreatic neuroendocrine tumours: the current incidence and staging based on the WHO and European Neuroendocrine Tumour Society classification: an analysis based on prospectively collected parameters. *Endocr Relat Cancer* 17:909–918 (LoE ⊗⊗⊗)
- GRADE Working Group website. <http://www.grade-workinggroup.org> (accessed 21 January 2014)
- Atkins D, Best D, Briss PA, Eccles M, Falck-Ytter Y, Flottorp S, Guyatt GH, Harbour RT, Haugh MC, Henry D, Hill S, Jaeschke R, Leng G, Liberati A, Magrini N, Mason J, Middleton P, Mrukowicz J, O’Connell D, Oxman AD, Phillips B, Schünemann HJ, Edejer T, Varonen H, Vist GE, GRADE Working Group (2004) Grading quality of evidence and strength of recommendations. *Br Med J* 328:1490–1497
- Swiglo BA, Murad MH, Schünemann HJ, Kunz R, Vigersky RA, Guyatt GH, Montori VM (2008) A case for clarity, consistency, and helpfulness: state-of-the-art clinical practice guidelines in endocrinology using the Grading of Recommendations, Assessment, Development, and Evaluation System. *J Clin Endocrinol Metab* 93:666–673
- Guyatt GH, Oxman AD, Vist GE, Kunz R, Falck-Ytter Y, Alonso-Coello P, Schünemann HJ, GRADE Working Group (2008) GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *Br Med J* 336:924–926
- Klimstra DS, Modlin IR, Coppola D, Lloyd RV, Suster S (2010) The pathologic classification of neuroendocrine tumors. A review of nomenclature, grading, and staging systems. *Pancreas* 39:707–712
- Bosman F, Carneiro F, Hruban R, Theise ND (2010) WHO classification of tumours of the digestive system, 4th edn. IARC Press, Lyon
- Singh S, Rowsell C, Ingbe N, et al (2010) The role of Ki-67 in the prognosis and management of neuroendocrine (NET) patients in a multidisciplinary cancer center. ASCO Gastrointestinal Cancers Symposium. Abstract no. 266
- Singh S, Feinberg Y, Rowsell C, et al (2011) Significance and reliability of pathologic marker Ki-67 in patients with neuroendocrine cancers. *J Clin Oncol* 29(suppl 4):abstr 264
- Klimstra DS (2013) Pathology reporting of neuroendocrine tumors: essential elements for accurate diagnosis, classification, and staging. *Semin Oncol* 40:23–36
- Jann H, Roll S, Couvelard A, Hentic O, Pavel M, Müller-Nordhorn J, Koch M, Röcken C, Rindi G, Ruszniewski P, Wiedenmann B, Pape UF (2011) Neuroendocrine tumors of midgut and hindgut origin: tumor-node-metastasis classification determines clinical outcome. *Cancer* 117:3332–3341 (LoE ⊗⊗⊗)
- Rindi G, Kloppel G, Alhman H, Caplin M, Couvelard A, de Herder WW, Eriksson B, Falchetti A, Falconi M, Komminoth P, Körner M, Lopes JM, McNicol AM, Nilsson O, Perren A, Scarpa A, Scoazec JY, Wiedenmann B, all other Frascati Consensus Conference participants, European Neuroendocrine Tumor Society (ENETS) (2006) TNM staging of foregut (neuro)endocrine tumors: a consensus proposal including a grading system. *Virchows Arch* 449:395–401
- Rindi G, Kloppel G, Couvelard A, Komminoth P, Körner M, Lopes JM, McNicol AM, Nilsson O, Perren A, Scarpa A, Scoazec JY, Wiedenmann B (2007) TNM staging of hindgut (neuro)endocrine tumors: a consensus proposal including a grading system. *Virchows Arch* 451:757–762
- Sobin LH, Gospodarowicz M, Wittekind C (2009) TNM classification of malignant tumours. 7th edn. Wiley Blackwell, Oxford
- Yang Z, Tang LH, Klimstra DS (2011) Effect of tumor heterogeneity on the assessment of Ki-67 labeling index in well-differentiated neuroendocrine tumors metastatic to the liver: implications for prognostic stratification. *Am J Surg Pathol* 35:853–860 (LoE ⊗⊗⊗)
- Tang LH, Gonen M, Hedvat C, Modlin I, Klimstra DS (2012) Objective quantification of the Ki-67 proliferation index in neuroendocrine tumors of the gastroenteropancreatic system. *Am J Surg Pathol* 36:1761–1770 (LoE ⊗⊗⊗)
- Kahn MS, Luong TV, Watkins J, Toumpanakis C, Caplin ME, Meyer T (2013) A comparison of Ki-67 and mitotic count as

- prognostic markers for metastatic pancreatic and midgut neuroendocrine neoplasms. *Br J Cancer* 108:1838–1845 (LoE ⊗⊗○○)
27. Sorbye H, Welin S, Langer SW, Westermarck LW, Holt N, Osterlund P, Duelund S, Hofslie E, Guren MG, Ohrling K, Birkemayer E, Thiis-Evensen E, Biagini M, Gronbaek H, Soveri ML, Olsen IH, Federspiel B, Assmus J, Janson ET, Knigge U (2012) Predictive and prognostic factors for treatment and survival in 305 patients with advanced gastrointestinal neuroendocrine carcinoma (WHO G3): the NORDIC NEC study. *Ann Oncol* 11:1–9 (LoE ⊗⊗○○)
 28. Chetty R (2008) An overview of practical issues in the diagnosis of gastroenteropancreatic neuroendocrine pathology. *Arch Pathol Lab Med* 132:1285–1289
 29. Vélazoudom-Céphise FL, Duvillard P, Foucan L, Hadoux J, Chougnet CN, Leboulleux S, Malka D, Guigay J, Goere D, Debaere T, Caramella C, Schlumberger M, Planchard D, Elias D, Ducreux M, Scoazec JY, Baudin E (2013) Are G3 ENETS neuroendocrine neoplasms heterogeneous? *Endocr Relat Cancer* 20:649–657 (LoE ⊗⊗○○)
 30. Petrone M, Testoni SGG, Carrara S, Balzano G, Manzoni M, Doglioni C, Testoni PA, Arcidiacono PA (2012) EUS-Guided FNA for proliferative rate in pancreatic neuroendocrine tumors: a single center experience over a 11-year period. *JOP* 13(5 Suppl):569
 31. Larghi A, Capurso G, Carnuccio A, Ricci R, Alfieri S, Galasso D, Lugli F, Bianchi A, Panzuto F, De Marinis L, Falconi M, Delle Fave G, Doglietto GB, Costamagna G, Rindi G (2011) KI-67 grading of nonfunctioning pancreatic neuroendocrine tumors on histologic samples obtaining by EUS-guide fine-needle tissue acquisition: a prospective study. *Gastrointest Endosc* 76:570–577 (LoE ⊗⊗○○)
 32. Iqbal S, Friedel D, Gupta M, Ogden L, Stavroulos SN (2012) Endoscopic-ultrasound-guided fine needle aspiration and the role of the cytopathologist in solid pancreatic lesion diagnosis. *Pathol Res Intern* 2012:1–17
 33. Ramage J, Ahmed A, Ardill J, Bax N, Breen DJ, Caplin ME, Corrie P, Davar J, Davies AH, Lewington V, Meyer T, Newell-Price J, Poston G, Reed N, Rockall A, Steward W, Thakker RV, Toubanakis C, Valle J, Verbeke C, Grossman AB, UK and Ireland Neuroendocrine Tumour Society (2012) Guidelines for the management of gastroenteropancreatic neuroendocrine (including carcinoid) tumours (NETs). *Gut* 61:6–32
 34. Salazar R, Wiedenmann B, Rindi G, Ruzsniawski P (2012) ENETS 2011 consensus guidelines for the management of patients with digestive neuroendocrine tumours: an update. *Neuroendocrinology* 95:71–73
 35. Bellizzi A (2013) Assigning site of origin in metastatic neuroendocrine neoplasms: a clinically significant application of diagnostic immunohistochemistry. *Adv Anat Pathol* 20:285–314
 36. Metz DC, Choi J, Strasberg J, Heaney AP, Howden CW, Klimstra D, Yao JC (2012) A rationale for multidisciplinary care in treating neuroendocrine tumors. *Curr Opin Endocrinol Diabetes Obes* 19:306–313
 37. Klimstra DS, Modlin I, Adsay NV, Chetty R, Deshpande V, Gönen M, Jensen RT, Kidd M, Kulke MH, Lloyd RV, Moran C, Moss SF, Oberg K, O'Toole D, Rindi G, Robert ME, Suster S, Tang LH, Tzen CY, Washington MK, Wiedenmann B, Yao J (2010) Pathology reporting of neuroendocrine tumors: application of the Delphic consensus process to the development of a minimum pathology data set. *Am J Surg Pathol* 34:300–313
 38. Anlauf M, Garbrecht N, Bauersfeld J, Schmitt A, Henopp T, Komminoth P, Heitz PU, Perren A, Klöppel G (2007) Hereditary neuroendocrine tumors of the gastroenteropancreatic system. *Virchows Arch* 451(Suppl 1):S29–S38
 39. Starker LF, Carling T (2008) Molecular genetics of gastroenteropancreatic neuroendocrine tumors. *Curr Opin Oncol* 21:29–33
 40. Thakker RV, Newey PJ, Walls GV, Bilezikian J, Bralle H, Ebeling PR, Melmed S, Sakurai A, Tonelli F, Brandi ML, Endocrine Society (2012) Clinical practice guidelines for multiple endocrine neoplasia type 1 (MEN1). *J Clin Endocrinol Metab* 97:2990–3011
 41. Brandi ML, Gagel RF, Angeli A, Bilezikian JP, Beck-Peccoz P, Bordi C, Conte-Devolx B, Falchetti A, Gheri RG, Libroia A, Lips CJ, Lombardi G, Mannelli M, Pacini F, Ponder BA, Raue F, Skogseid B, Tamburrano G, Thakker RV, Thompson NW, Tomassetti P, Tonelli F, Wells SA, Marx SJ (2001) Guidelines for diagnosis and therapy of MEN type 1 and type 2. *J Clin Endocrinol Metab* 86:5658–5671
 42. Ellard S, Hattersley AT, Brewer CM, Vaidya B (2005) Detection of an *MEN1* gene mutation depends on clinical features and supports current referral criteria for diagnostic molecular genetic testing. *Clin Endocrinol* 62:169–175 (LoE ⊗⊗○○)
 43. Bassett JHD, Forbes SA, Pannett AAJ, Lloyd SE, Christie PT, Wooding C, Harding B, Besser GM, Edwards CR, Monson JP, Sampson J, Wass JAH, Wheeler MH, Thakker RV (1998) Characterization of mutations in patients with multiple endocrine neoplasia type I. *Am J Hum Genet* 62:232–244
 44. Lemos MC, Thakker RV (2008) Multiple endocrine neoplasia type 1 (MEN1): analysis of 1336 mutations reported in the first decade following identification of the gene. *Hum Mutat* 29:22–32
 45. Wautot V, Vercherat C, Lespinasse J, Chambe B, Lenoir GM, Zhang CX, Porchet N, Cordier M, Beroud C, Calender A (2002) Germline mutation profile of MEN1 in multiple endocrine neoplasia type 1: search for correlation between phenotype and the functional domains of the MEN1 protein. *Hum Mutat* 20:35–47
 46. Corcos O, Couvelard A, Giraud S, Vullierme MP, O'Toole Dermot, Rebours V, Stievenart JL, Penforis A, Niccoli-Sire P, Baudin E, Sauvanet A, Levy P, Ruzsniawski P, Richard S, Hammel P (2008) Endocrine pancreatic tumors in von Hippel-Lindau disease: clinical, histological, and genetic features. *Pancreas* 37:85–93
 47. Maher ER, Iselius L, Yates JR, Littler M, Benjamin C, Harris R, Sampson J, Williams A, Ferguson-Smith MA, Morton N (1991) Von Hippel-Lindau disease: a genetic study. *J Med Genet* 28:443–447
 48. Lonsler RR, Glenn GM, Walther M, Chew EY, Libutti SK, Linehan WM, Oldfield EH (2003) von Hippel-Lindau disease. *Lancet* 361:2059–2067
 49. Rasmussen A, Nava-Salazar S, Yescas P, Alonso E, Revuelta R, Ortiz I, Canizales-Quinteros S, Tusie-Luna MT, Lopez-Lopez M (2006) von Hippel-Lindau disease germline mutations in Mexican patients with cerebellar hemangioblastoma. *J Neurosurg* 104:389–394 (LoE ⊗⊗○○)
 50. American Society of Clinical Oncology (2003) Policy statement update: genetic testing for cancer susceptibility. *J Clin Oncol* 21:2397–2406
 51. Dayal Y, Tallberg KA, Nunnemacher G, DeLellis RA, Wolfe HJ (1986) Duodenal carcinoids in patients with and without neurofibromatosis. A comparative study. *Am J Surg Pathol* 10:348–357 (LoE ⊗○○○)
 52. Williams VC, Lucas J, Babcock MA, Gutmann DH, Korf B, Maria BL (2009) Neurofibromatosis type 1 revisited. *Pediatrics* 123:124–133
 53. Davoren PM, Epstein MT (1992) Insulinoma complicating tuberous sclerosis. *J Neurol Neurosurg Psychiatry* 55:1209
 54. Francelanci P, Diomedici-Camassei F, Purificato C, Santorelli FM, Giannotti A, Dominici C, Inserra A, Boldrini R (2003) Malignant pancreatic endocrine tumor in a child with tuberous sclerosis. *Am J Surg Pathol* 27:1386–1389

55. van Slegtenhorst M, de Hoogt R, Hermans C, Nellist M, Janssen B, Verhoef S, Lindhout D, van den Ouweland A, Halley D, Young J, Burley M, Jeremiah S, Woodward K, Nahmias J, Fox M, Ekong R, Osborne J, Wolfe J, Povey S, Snell RG, Cheadle JP, Jones AC, Tachataki M, Ravine D, Sampson JR, Reeve MP, Richardson P, Wilmer F, Munro C, Hawkins TL, Sepp T, Ali JB, Ward S, Green AJ, Yates JR, Kwiatkowska J, Henske EP, Short MP, Haines JH, Jozwiak S, Kwiatkowski DJ (1997) Identification of the tuberous sclerosis gene TSC1 on chromosome 9q34. *Science* 277:805–808
56. Sancak O, Nellist M, Goedbloed M, Elfferich P, Wouters C, Maat-Kievit A, Zonnenberg B, Verhoef S, Halley D, van den Ouweland A (2005) Mutational analysis of the TSC1 and TSC2 genes in a diagnostic setting: genotype–phenotype correlations and comparison of diagnostic DNA techniques in tuberous sclerosis complex. *Eur J Hum Genet* 13:731–741
57. Ardill JE, Eriksson B (2003) The importance of the measurement of circulating markers in patients with neuroendocrine tumours of the pancreas and gut. *Endocr Relat Cancer* 10:459–462
58. Ardill JES, O’Dorisio TM (2010) Circulating biomarkers in neuroendocrine tumors of the enteropancreatic tract: application to diagnosis, monitoring disease, and as prognostic indicators. *Endocrinol Metab Clin North Am* 40:777–790
59. Stridsberg M, Öberg K, Li Q, Engström U, Lundqvist G (1995) Measurements of chromogranin A, chromogranin B (secretogranin I), chromogranin C (secretogranin II) and pancreastatin in plasma and urine from patients with carcinoid tumours and endocrine pancreatic tumours. *J Endocrinol* 144:49–59 (LoE ⊗○○○)
60. Stridsberg M, Eriksson B, Öberg K, Janson ET (2003) A comparison between three commercial kits for chromogranin A measurements. *J Endocrinol* 177:337–341 (LoE ⊗○○○)
61. Zatelli MC, Torta M, Leon A, Ambrosio MR, Gion M, Tomassetti P, De Braud F, Delle Fave G, Dogliotti L, Degli Uberti EC, Italian CromaNet Working Group (2007) Chromogranin A as a marker of neuroendocrine neoplasia: an Italian Multicenter Study. *Endocr Relat Cancer* 14:473–482 (LoE ⊗○○○)
62. Granberg D, Stridsberg M, Seensalu R, Eriksson B, Lundqvist G, Öberg K, Skogseid B (1999) Plasma chromogranin A in patients with multiple endocrine neoplasia type 1. *J Clin Endocrinol Metab* 84:2712–2717 (LoE ⊗○○○)
63. Campana D, Nori F, Piscitelli L, Morselli-Labate AM, Pezzilli R, Corinaldesi R, Tomassetti P (2007) Chromogranin A: is it a useful marker of neuroendocrine tumors? *J Clin Oncol* 25:1967–1973 (LoE ⊗○○○)
64. Modlin IM, Gustafsson BI, Moss SF, Pavel M, Tsolakis AV, Kidd M (2010) Chromogranin A—biological function and clinical utility in neuro endocrine tumor disease. *Ann Surg Oncol* 17:2427–2443
65. Singh S, Law C (2012) Chromogranin A: a sensitive biomarker for the detection and post-treatment monitoring of gastroenteropancreatic neuroendocrine tumors. *Expert Rev Gastroenterol Hepatol* 6:313–334
66. Marotta V, Nuzzo V, Ferrara T, Zuccoli A, Masone M, Nocerino L, Del Prete M, Marciello F, Ramundo V, Lombardi G, Vitale M, Colao A, Faggiano A (2012) Limitations of Chromogranin A in clinical practice. *Biomarkers* 17:186–191 (LoE ⊗○○○)
67. Mosli HH, Dennis A, Kocha W, Asher LJ, Van Uum SH (2012) Effect of short-term proton pump inhibitor treatment and its discontinuation on chromogranin A in healthy subjects. *J Clin Endocrinol Metab* 97:E1731–E1735 (LoE ⊗○○○)
68. Korse CM, Muller M, Taal BG (2011) Discontinuation of proton pump inhibitors during assessment of chromogranin A levels in patients with neuroendocrine tumours. *Br J Cancer* 105:1173–1175 (LoE ⊗○○○)
69. Nikou GC, Marinou K, Thomakos P, Papageorgiou D, Sanzani V, Nikolaou P, Kosmidis C, Moulakakis A, Mallas E (2008) Chromogranin A levels in diagnosis, treatment and follow-up of 42 patients with non-functioning pancreatic endocrine tumours. *Pancreatol* 8:510–519 (LoE ⊗○○○)
70. Panzuto F, Severi C, Cannizzaro R, Falconi M, Angeletti S, Pasquali A, Corleto VD, Annibale B, Buonadonna A, Pederzoli P, Delle Fave G (2004) Utility of combined use of plasma levels of chromogranin A and pancreatic polypeptide in the diagnosis of gastrointestinal and pancreatic endocrine tumors. *J Endocrinol Invest* 27:6–11 (LoE ⊗○○○)
71. Seregini E, Ferrari L, Stivanello M, Dogliotti L (2000) Laboratory tests for neuroendocrine tumours. *Q J Nucl Med* 44:22–41
72. Korse CM, Taal BG, Vincent A, van Velthuysen ML, Baas P, Buning-Kager JC, Linders TC, Bonfrer JM (2012) Choice of tumour markers in patients with neuroendocrine tumours is dependent on the histological grade. A marker study of chromogranin A, neuron specific enolase, progastrin-releasing peptide and cytokeratin fragments. *Eur J Cancer* 48:662–671 (LoE ⊗○○○)
73. Shah T, Srirajakanthan R, Bhogal M, Toubanakis C, Meyer T, Noonan A, Witney-Smith C, Amin T, Bhogal P, Sivathasan N, Warner B, Hochhauser D, Caplin ME (2008) Alpha-fetoprotein and human chorionic gonadotrophin-beta as prognostic markers in neuroendocrine tumour patients. *Br J Cancer* 99:72–77 (LoE ⊗○○○)
74. Pape UF, Perren A, Niederle B, Gross D, Gress T, Costa F, Arnold R, Denecke T, Plöckinger U, Salazar R, Grossman A, Barcelona Consensus Conference participants (2012) ENETS Consensus Guidelines for the management of patients with neuroendocrine neoplasms from the jejunum-ileum and the appendix including goblet cell carcinomas. *Neuroendocrinology* 95:135–156
75. Kema IP, de Vries EG, Muskiet FA (1995) Measurement of 5-HIAA in urine. *Ann Clin Biochem* 32:102–104
76. Kema IP, de Vries EG, Muskiet FA (2000) Clinical chemistry of serotonin and metabolites. *J Chromatogr B Biomed Sci Appl* 747:33–48
77. O’Toole D, Grossman A, Gross D, Delle Fave G, Barkmanova J, O’Connor J, Pape UF, Plöckinger U (2009) ENETS consensus guidelines for the standards of care in neuroendocrine tumors: biochemical markers. *Neuroendocrinology* 90:194–202
78. Sjoblom SM (1988) Clinical presentation and prognosis of gastrointestinal carcinoid tumours. *Scand J Gastroenterol* 23:779–787
79. Tellez MR, Mamikunian G, O’Dorisio TM, Vinik AI, Woltering EA (2013) A single fasting plasma 5-HIAA value correlates with 24-hour urinary 5-HIAA values and other biomarkers in midgut neuroendocrine tumors (NETs). *Pancreas* 42:405–410 (LoE ⊗○○○)
80. Allen KR, Degg TJ, Anthony DA, Fitzroy-Smith D (2007) Monitoring the treatment of carcinoid disease using blood serotonin and plasma 5-hydroxyindoleacetic acid: three case examples. *Ann Clin Biochem* 44:300–307 (LoE ⊗○○○)
81. Rehfeld JF, Gingras MH, Bardram L, Hilsted L, Goetze JP, Poitras P (2011) The Zollinger-Ellison syndrome and mismeasurement of gastrin. *Gastroenterology* 140:1444–1453
82. Dhillon WS, Jayasena CN, Lewis CJ, Martin NM, Tang KC, Meeran K, Todd JF (2006) Plasma gastrin measurement cannot be used to diagnose a gastrinoma in patients on either proton pump inhibitors or histamine type-2 receptor antagonists. *Ann Clin Biochem* 43:153–155 (LoE ⊗○○○)
83. Arnold R (2007) Diagnosis and differential diagnosis of hypergastrinemia. *Wien Klin Wochenschr* 119:564–569
84. Banasch M, Schmitz F (2007) Diagnosis and treatment of gastrinoma in the era of proton pump inhibitors. *Wien Klin Wochenschr* 119:573–578

85. Osefo N, Ito T, Jensen RT (2009) Gastric acid hypersecretory states: recent insights and advances. *Curr Gastroenterol Rep* 11:433–441
86. Rehfeld JF, Bardram L, Hilsted L, Poitras P, Goetze JP (2012) Pitfalls in diagnostic gastrin measurements. *Clin Chem* 58:831–836
87. Berna MJ, Hoffmann KM, Serrano J, Gibril F, Jensen RT (2006) Serum gastrin in Zollinger-Ellison syndrome. Prospective study of fasting serum gastrin in 309 patients from the National Institutes of Health and comparison with 2,229 cases from the literature. *Medicine (Baltimore)* 85:295–330 (LoE ⊗⊗⊗)
88. Jensen RT, Cadiot G, Brandi ML, de Herder WW, Kaltsas G, Komminoth P, Scoazec JY, Salazar R, Sauvanet A, Kianmanesh R, Barcelona Consensus Conference participants (2012) ENETS Consensus Guidelines for the management of patients with digestive neuroendocrine neoplasms: functional pancreatic endocrine tumor syndromes. *Neuroendocrinology* 95:98–119
89. Service F (1999) Diagnostic approach to adults with hypoglycaemic disorders. *Endocrinol Metab Clin North Am* 28:519–532
90. Vezzosi D, Bennet A, Fauvel J, Boulanger C, Tazi O, Louvet JP, Caron P (2003) Insulin levels measured with an insulin-specific assay in patients with fasting hypoglycaemia related to endogenous hyperinsulinism. *Eur J Endocrinol* 149:413–419 (LoE ⊗⊗⊗)
91. Wermers RA, Fatourehchi V, Kvols LK (1996) Clinical spectrum of hyperglucagonemia associated with malignant neuroendocrine tumors. *Mayo Clin Proc* 71:1030–1038
92. Krejs GJ (1987) Vipoma syndrome. *Am J Med* 82:37–48
93. Grier JF (1995) WDHA (watery diarrhea, hypokalemia, achlorhydria) syndrome: clinical features, diagnosis, and treatment. *South Med J* 88:22–24
94. Sahani DV, Bonaffini PA, Fernandez-Del Castillo C, Blake MA (2013) Gastroenteropancreatic neuroendocrine tumors: role of imaging in diagnosis and management. *Radiology* 266:38–61
95. Sundin A, Vullierme MP, Kaltsas G, Plöckinger U, Mallorca Consensus Conference participants, European Neuroendocrine Tumor Society (2009) ENETS consensus guidelines for the standards of care in neuroendocrine tumors: radiological examinations. *Neuroendocrinology* 90:167–183
96. Falconi M, Bartsch DK, Eriksson B, Klöppel G, Lopes JM, O'Connor JM, Salazar R, Taal BG, Vullierme MP, O'Toole D, Barcelona Consensus Conference participants (2012) ENETS consensus guidelines for the management of patients with digestive neuroendocrine neoplasms of the digestive system: well-differentiated pancreatic non-functioning tumors. *Neuroendocrinology* 95:120–134
97. Malagò R, D'Onofrio M, Zamboni GA, Faccioli N, Falconi M, Boninsegna L, Mucelli RP (2009) Contrast-enhanced sonography of non-functioning pancreatic neuroendocrine tumors. *AJR Am J Roentgenol* 192:424–430 (LoE ⊗⊗⊗)
98. Quايا E, Stacul F, Gaiani S, Ricci P, Passariello R, Curzio D, Pozzi Mucelli R (2004) Comparison of diagnostic performance of unenhanced vs. SonoVue-enhanced ultrasonography in focal liver lesions characterization. The experience of three Italian centers. *Radiol Med* 108:71–81 (LoE ⊗⊗⊗)
99. Mork H, Ignee A, Schuessler G, Ott M, Dietrich CF (2007) Analysis of neuroendocrine tumour metastases in the liver using contrast-enhanced ultrasonography. *Scand J Gastroenterol* 42:652–662 (LoE ⊗⊗⊗)
100. Hoeffel C, Job L, Ladam-Marcus V, Vitry F, Cadiot G, Marcus C (2009) Detection of hepatic metastases from carcinoid tumor: prospective evaluation of contrast-enhanced ultrasonography. *Dig Dis Sci* 54:2040–2046 (LoE ⊗⊗⊗)
101. Wu W, Chen MH, Yin SS, Yan K, Fan ZH, Yang W, Dai Y, Huo L, Li JY (2006) The role of contrast enhanced sonography of focal liver lesions before percutaneous biopsy. *AJR Am J Roentgenol* 187:752–761 (LoE ⊗⊗⊗)
102. Chatzipantelis P, Salla C, Konstantinou P, Karoumpalis I, Sakellariou S, Doumani I (2008) Endoscopic ultrasound-guided fine-needle aspiration cytology of pancreatic neuroendocrine tumors: a study of 48 cases. *Cancer* 114:255–262 (LoE ⊗⊗⊗)
103. Jani N, Khalid A, Kaushik N, Brody D, Bauer K, Schoedel K, Ohori NP, Moser AJ, Lee K, McGrath K (2008) EUS guided FNA diagnosis of pancreatic endocrine tumors: new trends identified. *Gastrointest Endosc* 67:44–50 (LoE ⊗⊗⊗)
104. Fidler JL, Fletcher JG, Reading CC, Andrews JC, Thompson GB, Grant CS, Service FJ (2003) Preoperative detection of pancreatic insulinomas on multiphasic helical CT. *AJR Am J Roentgenol* 181:775–780 (LoE ⊗⊗⊗)
105. Cwikla JB, Buscombe JR, Caplin ME, Watkinson AF, Walecki J, Gorczyca-Wiśniewska E, Hilson AJ (2004) Diagnostic imaging of carcinoid metastases to the abdomen and pelvis. *Med Sci Monit* 10(suppl 3):9–16 (LoE ⊗⊗⊗)
106. Kumbasar B, Kamel IR, Tekes A, Eng J, Fishman EK, Wahl RL (2004) Imaging of neuroendocrine tumors: accuracy of helical CT versus SRS. *Abdom Imaging* 29:696–702 (LoE ⊗⊗⊗)
107. Johanssen S, Boivin M, Lochs H, Voderholzer W (2006) The yield of wireless capsule endoscopy in the detection of NET in comparison with CT enteroclysis. *Gastrointest Endosc* 63:660–665 (LoE ⊗⊗⊗)
108. Pilleul F, Penigaud M, Milot L, Saurin JC, Chayvialle JA, Vallette PJ (2006) Possible small bowel neoplasms: contrast-enhanced and water-enhanced multidetector CT enteroclysis. *Radiology* 241:796–801 (LoE ⊗⊗⊗)
109. Kamaoui I, De Luca V, Ficarella S, Mennesson N, Lombard-Bohas C, Pilleul F (2010) Value of CT enteroclysis in suspected small-bowel carcinoid tumors. *Am J Roentgenol* 194:629–633 (LoE ⊗⊗⊗)
110. Waldmann J, Fendrich V, Habbe N, Bartsch DK, Slater EP, Kann PH, Rothmund M, Langer P (2009) Screening of patients with multiple endocrine neoplasia type 1 (MEN-1): a critical analysis of its value. *World J Surg* 33:1208–1218 (LoE ⊗⊗⊗)
111. van der Lely AJ, de Herder WW, Krenning EP, Kwekkeboom DJ (2003) Octreoscan radioreceptor imaging. *Endocrine* 20:307–311
112. Kwekkeboom DJ, Krenning EP, Scheidhauer K, Lewington V, Lebtahi R, Grossman A, Vitek P, Sundin A, Plöckinger U, Mallorca Consensus Conference participants, European Neuroendocrine Tumor Society (2009) ENETS consensus guidelines for the standards of care in neuroendocrine tumors: somatostatin receptor imaging with ¹¹¹In-pentetreotide. *Neuroendocrinology* 90:184–189
113. Perri M, Erba P, Volterrani D, Lazzeri E, Boni G, Grosso M, Mariani G (2008) Octreo-SPECT/CT imaging for accurate detection and localization of suspected neuroendocrine tumors. *Q J Nucl Med Mol Imaging* 52:323–333 (LoE ⊗⊗⊗)
114. Gibril F, Reynolds JC, Doppmann JL, Chen CC, Venzon DJ, Termanini B, Weber HC, Stewart CA, Jensen RT (1996) Somatostatin receptor scintigraphy: its sensitivity compared with that of other imaging methods in detecting primary and metastatic gastrinomas: a prospective study. *Ann Int Med* 125:26–34 (LoE ⊗⊗⊗)
115. Shi W, Johnston CF, Buchanan KD, Ferguson WR, Laird JD, Crothers JG, McIlrath EM (1998) Localization of neuroendocrine tumours with [¹¹¹In]-DTPA-octreotide scintigraphy (Octreoscan): a comparative study with CT and MRI imaging. *QJM* 91:295–301 (LoE ⊗⊗⊗)
116. Kaltsas G, Mukherjee JJ, Grossman AB (2001) The value of radiolabeled MIBG and octreotide in the diagnosis and management of neuroendocrine tumours. *Ann Oncol* 12(Suppl 2):S47–S50

117. Kaltsas G, Rockall A, Papadogias D, Reznick R, Grossman AB (2004) Recent advances in radiological and radionuclide imaging and therapy of neuroendocrine tumours. *Eur J Endocrinol* 151:15–27
118. de Herder WW, Kwekkeboom DJ, Feelders RA, van Aken MO, Lamberts SW, van der Lely AJ, Krenning EP (2006) Somatostatin receptor imaging for neuroendocrine tumors. *Pituitary* 9:243–248
119. Krenning EP, Kwekkeboom DJ, Bakker WH, Breeman WA, Kooij PP, Oei HY, van Hagen M, Postema PT, de Jong M, Reubi JC et al (1993) Somatostatin receptor scintigraphy with [¹¹¹In-DTPA-D-Phe¹]- and [¹²³I-Tyr³]- octreotide: the Rotterdam experience with more than 1,000 patients. *Eur J Nucl Med* 20:716–731
120. Reubi JC, Schär JC, Waser B, Wenger S, Heppeler A, Schmitt JS, Mäcke HR (2000) Affinity profiles for human somatostatin receptor subtypes SST1–SST5 of somatostatin radiotracers selected for scintigraphic and radiotherapeutic use. *Eur J Nucl Med Mol Imaging* 27:273–282
121. Hofmann M, Maecke H, Borner R, Weckesser E, Schoffski P, Oei L, Schumacher J, Henze M, Heppeler A, Meyer J, Knapp H (2001) Biokinetics and imaging with the somatostatin receptor PET radioligand ⁶⁸Ga-DOTATOC: preliminary data. *Eur J Nucl Med Mol Imaging* 28:1751–1757 (LoE ⊗○○○)
122. Kowalski J, Henze M, Schumacher J, Maecke HR, Hofmann H, Haberkorn U (2003) Evaluation of positron emission tomography imaging using ⁶⁸Ga-DOTA-Phe¹-Tyr³-octreotide in comparison to ¹¹¹In-DTPAOC SPECT. First results in patients with neuroendocrine tumors. *Mol Imag Biol* 5:42–48 (LoE ⊗○○○)
123. Buchmann I, Henze M, Engelbrecht S, Eisenhut S, Runz A, Schäfer M, Schilling T, Haufe S, Herrmann T, Haberkorn U (2007) Comparison of ⁶⁸Ga-DOTATOC PET and ¹¹¹In-DTPAOC (Octreoscan) SPECT in patients with neuroendocrine tumours. *Eur J Nucl Med Mol Imaging* 34:1617–1626 (LoE ⊗⊗⊗○)
124. Putzer D, Gabriel M, Henninger B, Kendler D, Uprimny C, Dobrozemsky G, Decristoforo C, Bale RJ, Jaschke W, Virgolini IJ (2009) Bone metastases in patients with neuroendocrine tumor: ⁶⁸Ga-DOTA-Tyr³-octreotide PET in comparison to CT and bone scintigraphy. *J Nucl Med* 50:1214–1221 (LoE ⊗⊗⊗○)
125. Kayani I, Bomanji JB, Groves A, Conway G, Gacinovic S, Win T, Dickson J, Caplin M, Ell PJ (2008) Functional imaging of neuroendocrine tumors with combined PET/CT using ⁶⁸Ga-DOTATATE (DOTA-DPhe¹, Tyr³-octreotate) and ¹⁸F-FDG. *Cancer* 112:2447–2455 (LoE ⊗⊗○○)
126. Prasad V, Ambrosini V, Hommann M, Hoersch D, Fanti S, Baum RP (2010) Detection of unknown primary neuroendocrine tumours (CUP-NET) using ⁶⁸Ga-DOTANOC receptor PET/CT. *Eur J Nucl Med Mol Imaging* 37:67–77 (LoE ⊗⊗○○)
127. Srirajakanthan R, Kayani I, Quigley AM, Soh J, Caplin ME, Bomanji J (2010) The role of ⁶⁸Ga-DOTATATE PET in patients with neuroendocrine tumors and negative or equivocal findings on ¹¹¹In-DTPA-octreotide scintigraphy. *J Nucl Med* 51:875–878 (LoE ⊗⊗○○)
128. Poeppel TD, Binse I, Petersenn S, Lahner H, Schott M, Antoch G, Brandau W, Bockisch A, Boy C (2011) ⁶⁸Ga-DOTATOC versus ⁶⁸Ga-DOTATATE PET/CT in functional imaging of neuroendocrine tumors. *J Nucl Med* 52:1864–1870 (LoE ⊗⊗⊗○)
129. Kabasakal L, Demirci E, Ocak M, Decristoforo C, Araman A, Ozsoy Y, Uslu I, Kanmaz B (2012) Comparison of ⁶⁸Ga-DOTATATE and ⁶⁸Ga-DOTANOC PET/CT imaging in the same patient group with neuroendocrine tumours. *Eur J Nucl Med Mol Imaging* 39:1271–1277 (LoE ⊗⊗⊗○)
130. Haug AR, Cindea-Drinus R, Auernhammer CJ, Reincke M, Wängler B, Uebleis C, Schmidt GP, Göke B, Bartenstein P, Hacker M (2012) The role of ⁶⁸Ga-DOTATATE PET/CT in suspected neuroendocrine tumors. *J Nucl Med* 53:1686–1692 (LoE ⊗⊗○○)
131. Gabriel M, Oberauer A, Dobrozemsky G, Decristoforo C, Putzer D, Kendler D, Uprimny C, Kovacs P, Bale R, Virgolini IJ (2009) ⁶⁸Ga-DOTA-Tyr³-octreotide PET for assessing response to somatostatin-receptor-mediated radionuclide therapy. *J Nucl Med* 50:1427–1434 (LoE ⊗⊗⊗○)
132. Frilling A, Sotiropoulos GC, Radtke A, Malago M, Bockisch A, Kuehl H, Li J, Broelsch CE (2010) The impact of ⁶⁸Ga-DOTATOC positron emission tomography/computed tomography on the multimodal management of patients with neuroendocrine tumors. *Ann Surg* 252:850–856 (LoE ⊗⊗○○)
133. Ruf J, Heuck F, Schiefer J, Denecke T, Elgeti F, Pascher A, Pavel M, Stelter L, Kropf S, Wiedenmann B, Amthauer H (2010) Impact of multiphase ⁶⁸Ga-DOTATOC-PET/CT on therapy management in patients with neuroendocrine tumors. *Neuroendocrinology* 91:101–109 (LoE ⊗⊗○○)
134. Kumar R, Sharma P, Garg P, Karunanithi S, Naswa N, Sharma R, Thulkar S, Lata S, Malhotra A (2011) Role of ⁶⁸Ga-DOTATOC PET-CT in the diagnosis and staging of pancreatic neuroendocrine tumours. *Eur Radiol* 21:2408–2416 (LoE ⊗⊗○○)
135. Bombardieri E, Ambrosini V, Aktolun C, Baum RP, Bishof-Delaloye A, Del Vecchio S, Maffioli L, Mortelmans L, Oyen W, Pepe G, Chiti A, Oncology Committee of the EANM (2010) ¹¹¹In-pentetreotide scintigraphy: procedure guidelines for tumour imaging. *Eur J Nucl Med Mol Imaging* 37:1441–1448
136. Adams S, Baum R, Rink T, Schumm-Dräger PM, Usadel KH, Hör G (1998) Limited value of fluorine-18 fluorodeoxyglucose positron emission tomography for the imaging of neuroendocrine tumours. *Eur J Nucl Med* 25:79–83 (LoE ⊗⊗⊗○)
137. Pasquali C, Rubello D, Sperti C, Gasparoni P, Liessi G, Chierichetti F, Ferlin G, Pedrazzoli S (1998) Neuroendocrine tumor imaging: can ¹⁸F-fluorodeoxyglucose positron emission tomography detect tumors with poor prognosis and aggressive behavior? *World J Surg* 22:588–592 (LoE ⊗○○○)
138. Giammarile F, Billotey C, Lombard-Bohas C, Le Bars D, Bournaud C, Masson S, Walter T, Houzard C, Scoazec JY, Hervieu V, Vuillez JP, Cornu C, Janier M, Borson-Chazot F (2011) ¹⁸F-FLT and ¹⁸F-FDG positron emission tomography for the imaging of advanced well-differentiated gastro-entero-pancreatic endocrine tumours. *Nucl Med Commun* 32:91–97 (LoE ⊗⊗⊗○)
139. Binderup T, Knigge U, Loft A, Mortensen J, Pfeifer A, Federspiel B, Hansen CP, Højgaard L, Kjaer A (2010) Functional imaging of neuroendocrine tumors: a head-to-head comparison of somatostatin receptor scintigraphy, ¹²³I-MIBG scintigraphy, and ¹⁸F-FDG PET. *J Nucl Med* 51:704–712 (LoE ⊗⊗⊗○)
140. Abgral R, Leboulleux S, Déandreis D, Aupérin A, Lumbroso J, Dromain C, Duvillard P, Elias D, de Baere T, Guigay J, Ducreux M, Schlumberger M, Baudin E (2011) Performance of ¹⁸Fluorodeoxyglucose positron emission tomography and somatostatin receptor scintigraphy for high Ki-67 (≥10%) well-differentiated endocrine carcinoma staging. *J Clin Endocrinol Metab* 96:665–671 (LoE ⊗⊗⊗○)
141. Garin E, Le Jeune F, Devillers A, Cuggia M, de Lajarte-Thirouard AS, Bourriel C, Boucher E, Raoul JL (2009) Predictive value of ¹⁸F-FDG PET and somatostatin receptor scintigraphy in patients with metastatic endocrine tumors. *J Nucl Med* 50:858–864 (LoE ⊗⊗⊗○)
142. Fuccio C, Musto A, Cambioli S, Castellucci P, Pantaleo MA, Nanni C, Montini GC, Campana D, Tomassetti P, Bellan E, Massaro A, Grassetto G, Rubello D, Fanti S (2011) When should ¹⁸F-18 FDG PET/CT be used instead of ⁶⁸Ga-DOTA-peptides to investigate metastatic neuroendocrine tumors? *Clin Nucl Med* 36:1109–1111 (LoE ⊗○○○)

143. Binderup T, Knigge U, Loft A, Federspiel B, Kjaer A (2010) ^{18}F -fluorodeoxyglucose positron emission tomography predicts survival of patients with neuroendocrine tumors. *Clin Cancer Res* 16:978–985 (LoE $\otimes\otimes\otimes\circ$)
144. Becherer A, Szabó M, Karanikas G, Wunderbaldinger P, Angelberger P, Raderer M, Kurtaran A, Dudczak R, Kletter K (2004) Imaging of advanced neuroendocrine tumors with ^{18}F -FDOPA PET. *J Nucl Med* 45:1161–1167 (LoE $\otimes\otimes\otimes\circ$)
145. Haug A, Auernhammer CJ, Wangler B, Tiling R, Schmidt G, Göke B, Bartenstein P, Pöppel G (2009) Intraindividual comparison of ^{68}Ga -DOTA-TATE and ^{18}F -DOPA PET in patients with well-differentiated metastatic neuroendocrine tumours. *Eur J Nucl Med Mol Imaging* 36:765–770 (LoE $\otimes\otimes\otimes\circ$)
146. Koopmans KP, Neels OC, Kema IP, Elsinga PH, Sluiter WJ, Vanghillewe K, Brouwers AH, Jager PL, de Vries EG (2008) Improved staging of patients with carcinoid and islet cell tumors with ^{18}F -dihydroxy-phenyl-alanine and ^{11}C -5-hydroxy-tryptophan positron emission tomography. *J Clin Oncol* 26:1489–1495 (LoE $\otimes\otimes\otimes\circ$)
147. Jager PL, Chirakal R, Marriott CJ, Brouwers AH, Koopmans KP, Gulenchyn KY (2008) 6-L- ^{18}F -fluorodihydroxyphenylalanine PET in neuroendocrine tumors: basic aspects and emerging clinical applications. *J Nucl Med* 49:573–586
148. Kauhanen S, Seppänen M, Ovaska J, Minn H, Bergman J, Korsoff P, Salmela P, Saltevo J, Sane T, Välimäki M, Nuutila P (2009) The clinical value of [^{18}F]fluoro-dihydroxy-phenylalanine positron emission tomography in primary diagnosis, staging, and restaging of neuroendocrine tumors. *Endocr Relat Cancer* 16:255–265 (LoE $\otimes\otimes\otimes\circ$)
149. Orlefors H, Sundin A, Garske U, Juhlin C, Oberg K, Skogseid B, Langstrom B, Bergstrom M, Eriksson B (2005) Whole-body ^{11}C -5-hydroxytryptophan positron emission tomography as a universal imaging technique for neuroendocrine tumors: comparison with somatostatin receptor scintigraphy and computed tomography. *J Clin Endocrinol Metab* 90:3392–3400 (LoE $\otimes\otimes\otimes\circ$)
150. Sundin A, Garske U, Orlefors H (2007) Nuclear imaging of neuroendocrine tumours. *Best Pract Res Clin Endocrinol Metab* 21:69–85
151. Christ E, Wild D, Forrer F, Brändle M, Sahli R, Clerici T, Gloor B, Martius F, Maecke H, Reubi JC (2009) Glucagon-Like Peptide-1 receptor imaging for localization of insulinomas. *J Clin Endocrinol Metab* 94:4398–4405 (LoE $\otimes\otimes\otimes\circ$)
152. Vannella L, Lahner E, Annibale B (2012) Risk for gastric neoplasias in patients with chronic atrophic gastritis: a critical reappraisal. *World J Gastroenterol* 18:1279–1285
153. Delle Fave G, Kwekkeboom DJ, Van Cutsem E, Rindi G, Kos-Kudla B, Knigge U, Sasano H, Tomassetti P, Salazar R, Ruszniewski P (2012) ENETS consensus guidelines for the management of patients with gastroduodenal neoplasms. *Neuroendocrinology* 95:74–87
154. Merola E, Sbrozzi-Vanni A, Panzuto F, D'Ambrà G, Di Giulio E, Pillozzi E, Capurso G, Lahner E, Bordi C, Annibale B, Delle Fave G (2012) Type I gastric carcinoids: a prospective study on endoscopic management and recurrence rate. *Neuroendocrinology* 95:207–213 (LoE $\otimes\otimes\otimes\circ$)
155. Ruszniewski P, Amouyal P, Amouyal G, Grange JD, Mignon M, Bouch O, Bernades P (1995) Localization of gastrinomas by endoscopic ultrasonography in patients with Zollinger–Ellison syndrome. *Surgery* 117:629–635 (LoE $\otimes\otimes\otimes\circ$)
156. Yoshikane H, Suzuki T, Yoshioka N, Ogawa Y, Hamajima E, Hasegawa N, Hasegawa C (1995) Duodenal carcinoid tumor: endosonographic imaging and endoscopic resection. *Am J Gastroenterol* 90:642–644 (LoE $\otimes\otimes\otimes\circ$)
157. Ramage JK, Goretzki PE, Manfredi R, Komminoth P, Ferone D, Hyrdel R, Kaltsas G, Kelestimir F, Kvols L, Scoazec JY, Garcia MI, Caplin ME, Frascati Consensus Conference participants (2008) Consensus guidelines for the management of patients with digestive neuroendocrine tumors: well differentiated colon and rectum tumor/carcinoma. *Neuroendocrinology* 87:31–39
158. Scherubl H (2009) Rectal carcinoids are on the rise: early detection by screening endoscopy. *Endoscopy* 41:162–165
159. Hara AK, Leighton JA, Sharma VK, Heigh RI, Fleischer DE (2005) Imaging of small bowel disease: comparison of capsule endoscopy, standard endoscopy, barium examination, and CT. *Radiographics* 25:697–711
160. Bellutti M, Fry L, Malfertheiner P, Monkemuller K (2009) Detection of NET of the small bowel by double balloon enteroscopy. *Dig Dis Sci* 54:1050–1058 (LoE $\otimes\otimes\otimes\circ$)
161. Pais SA, Al-Haddad M, Mohamadnejad M, Leblanc JK, Sherman S, McHenry L, DeWitt JM (2010) EUS for pancreatic neuroendocrine tumors: a single-center, 11-year experience. *Gastrointest Endosc* 71:1185–1193 (LoE $\otimes\otimes\otimes\circ$)
162. Atiq M, Bhutani MS, Bektas M, Lee JE, Gong Y, Tamm EP, Shah CP, Ross WA (2012) EUS-FNA for pancreatic neuroendocrine tumors: a tertiary cancer center experience. *Dig Dis Sci* 57:791–800 (LoE $\otimes\otimes\otimes\circ$)
163. Puli SR, Bechtold ML, Buxbaum JL, Eloubeidi MA (2013) How good is endoscopic ultrasound-guided fine-needle aspiration in diagnosing the correct etiology for a solid pancreatic mass? A meta-analysis and systematic review. *Pancreas* 42:20–26 (LoE $\otimes\otimes\otimes\circ$)
164. Puli SR, Kalva N, Bechtold ML, Pamulaparthi SR, Cashman MD, Estes NC, Pearl RH, Volmar FH, Dillon S, Shekleton MF, Forcione D (2013) Diagnostic accuracy of endoscopic ultrasound in pancreatic neuroendocrine tumors: a systematic review and metaanalysis. *World J Gastroenterol* 19:3678–3684 (LoE $\otimes\otimes\otimes\circ$)
165. Shumacher B, Lubke HJ, Frieling T, Strohmeier G, Starke AAR (1996) Prospective study on the detection of insulinomas by endoscopic ultrasonography. *Endoscopy* 28:273–276 (LoE $\otimes\otimes\otimes\circ$)
166. Gouya H, Vignaux O, Augui J, Dousset B, Palazzo L, Louvel A, Chaussade S, Legmann P (2003) CT, endoscopic sonography, and a combined protocol for preoperative evaluation of pancreatic insulinomas. *AJR Am J Roentgenol* 181:987–992 (LoE $\otimes\otimes\otimes\circ$)
167. Bansal R, Tierney W, Carpenter S, Thompson N, Scheiman JM (1999) Cost effectiveness of EUS for preoperative localization of pancreatic endocrine tumors. *Gastrointest Endosc* 49:19–25 (LoE $\otimes\otimes\otimes\circ$)
168. Hooper K, Mukhtar F, Li S, Eltoun IA (2013) Diagnostic error assessment and associated harm of endoscopic ultrasound-guided fine-needle aspiration of neuroendocrine neoplasms of the pancreas. *Cancer Cytopathol* 121:653–660 (LoE $\otimes\otimes\otimes\circ$)
169. Chang F, Chandra A, Culora G, Mahadeva U, Meenan J, Herbert A (2006) Cytologic diagnosis of pancreatic endocrine tumors by endoscopic ultrasound-guided fine-needle aspiration: a review. *Diagn Cytopathol* 34:649–658
170. Alsohaibani F, Bigam D, Kneteman N, Shapiro AMJ, Sandha GS (2008) The impact of preoperative endoscopic ultrasound on the surgical management of pancreatic neuroendocrine tumours. *Can J Gastroenterol* 22:817–820 (LoE $\otimes\otimes\otimes\circ$)
171. Haynes AB, Deshpande V, Ingkakul T, Vagefi PA, Szymonifka J, Thayer SP, Ferrone CR, Wargo JA, Warshaw AL, Fernandez-del Castillo C (2011) Implications of incidentally discovered, non-functioning pancreatic endocrine tumors. Short-term and long-term patient outcomes. *Arch Surg* 146:534–538 (LoE $\otimes\otimes\otimes\circ$)
172. Cheslyn-Curtis S, Sitaram V, Williamson RC (1993) Management of non-functioning neuro-endocrine tumours of the pancreas. *Br J Surg* 80:625–627
173. Caplin M, Sundin A, Nillson O, Baum RP, Klose KJ, Kelestimir F, Plöckinger U, Papotti M, Salazar R, Pascher A (2012) ENETS

- consensus guidelines for the management of patients with digestive neuroendocrine neoplasms: colorectal neuroendocrine neoplasms. *Neuroendocrinology* 95:88–97
174. Modlin IM, Kidd M, Latich I, Zikusoka MN, Shapiro MD (2005) Current status of gastrointestinal carcinoids. *Gastroenterology* 128:1717–1751
 175. Van der Lely AJ, de Herder WW (2005) Carcinoid syndrome: diagnosis and medical management. *Arq Bras Endocrinol Metab* 49:850–860
 176. Vinik AI, McLeod MK, Fig LM, Shapiro B, Lloyd RV, Cho K (1989) Clinical features, diagnosis, and localization of carcinoid tumors and their management. *Gastroenterol Clin North Am* 18:865–896
 177. Kulke MH, Mayer RJ (1999) Carcinoid tumors. *N Engl J Med* 340:859–868
 178. Janson ET, Holmberg L, Stridsberg M, Eriksson B, Theodorsson E, Wilander E, Öberg K (1997) Carcinoid tumors: analysis of prognostic factors and survival in 301 patients from a referral center. *Ann Oncol* 8:685–690 (LoE ⊗⊗⊗○)
 179. Soga J, Yakuwa Y, Osaka M (1999) Carcinoid syndrome: a statistical evaluation of 748 reported cases. *J Exp Clin Cancer Res* 18:133–141 (LoE ⊗⊗○)
 180. Tomassetti P, Migliori M, Lalli S, Campana D, Tomassetti V, Corinaldesi R (2001) Epidemiology, clinical features and diagnosis of gastroenteropancreatic endocrine tumours. *Ann Oncol* 12(Suppl 2):S95–S99
 181. Kaltsas G, Besser GM, Grossman AB (2004) The diagnosis and medical management of advanced neuroendocrine tumors. *Endocr Rev* 25:458–511
 182. Creutzfeldt W (1996) Carcinoid tumours: development of our knowledge. *World J Surg* 20:126–131
 183. Davì MV, Bodei L, Francia G, Bartolomei M, Oliani C, Scilanga L, Reghellin D, Falconi M, Paganelli G, Lo Cascio V, Ferdeghini M (2006) Carcinoid crisis induced by receptor radioligand therapy with 90Y-DOTATOC in a case of liver metastases from bronchial neuroendocrine tumor (atypical carcinoid). *J Endocrinol Invest* 29:563–567 (LoE ⊗○○○)
 184. Vinik AI, Gonzales MRC (2011) New and emerging syndromes due to neuroendocrine tumors. *Endocrinol Metab Clin North Am* 40:19–63
 185. McCormick D (2002) Carcinoid tumors and syndrome. *Gastroenterol Nurs* 25:105–111
 186. Gustafsson BI, Hauso O, Drosdo I, Kidd M, Modlin IM (2008) Carcinoid heart disease. *Int J Cardiol* 129:318–324
 187. Izikson L, English JC 3rd, Zirwas MJ (2006) The flushing patient: differential diagnosis, workup, and treatment. *J Am Acad Dermatol* 55:193–208
 188. Feldman J, Lee E (1985) Serotonin content of foods: effect on urinary excretion of 5-hydroxyindoleacetic acid. *Am J Clin Nutr* 42:639–643
 189. Richter G, Stockmann F, Conlon JM, Creutzfeldt W (1986) Serotonin release into blood after food and pentagastrin. Studies in healthy subjects and in patients with metastatic carcinoid tumors. *Gastroenterology* 91:612–618 (LoE ⊗○○○)
 190. Mashige F, Matsushima Y, Kanazawa H, Sakuma I, Takai N, Bessho F, Ohkubo A (1996) Acidic catecholamine metabolites and 5-hydroxyindoleacetic acid in urine: the influence of diet. *Ann Clin Biochem* 33:43–49
 191. Feldman JM (1987) Carcinoid tumors and syndrome. *Semin Oncol* 14:237–246
 192. Feldman JM, O'Dorisio TM (1986) Role of neuropeptides and serotonin in the diagnosis of carcinoid tumors. *Am J Med* 81:41–48
 193. Eriksson B, Öberg K, Stridsberg M (2000) Tumor markers in neuroendocrine tumors. *Digestion* 62(Suppl 1):33–38
 194. Öberg K (2011) Circulating biomarkers in gastroenteropancreatic neuroendocrine tumours. *Endocr Relat Cancer* 18(Suppl 1):S17–S25
 195. Tomassetti P, Migliori M, Simoni P, Casadei R, De Iasio R, Corinaldesi R, Gullo L (2001) Diagnostic value of plasma chromogranin A in neuroendocrine tumours. *Eur J Gastroenterol Hepatol* 13:55–58
 196. Rockall AG, Reznick RH (2007) Imaging of neuroendocrine tumours (CT/MR/US). *Best Pract Res Clin Endocrinol Metab* 21:43–68
 197. Dromain C, de Baere T, Lumbroso J, Caillet H, Laplanche A, Boige V, Ducreux M, Duvillard P, Elias D, Schlumberger M, Sigal R, Baudin E (2005) Detection of liver metastases from endocrine tumors: a prospective comparison of somatostatin receptor scintigraphy, computed tomography, and magnetic resonance imaging. *J Clin Oncol* 23:70–78 (LoE ⊗⊗⊗○)
 198. Dromain C, de Baere T, Baudin E, Galline J, Ducreux M, Boige V, Duvillard P, Laplanche A, Caillet H, Lasser P, Schlumberger M, Sigal R (2003) MR imaging of hepatic metastases caused by neuroendocrine tumors: comparing four techniques. *AJR Am J Roentgenol* 180:121–128 (LoE ⊗⊗○○)
 199. Boudreaux JP, Klimstra DS, Hassan MM, Woltering EA, Jensen RT, Goldsmith SJ, Nutting C, Bushnell DL, Caplin ME, Yao JC, North American Neuroendocrine Tumor Society (NANETS) (2010) The NANETS consensus guideline for the diagnosis and management of neuroendocrine tumors: well-differentiated neuroendocrine tumors of the Jejunum, Ileum, Appendix, and Cecum. *Pancreas* 39:753–766
 200. Modlin IM, Latich I, Zikusoka M, Kidd M, Eick G, Chan AK (2006) Gastrointestinal carcinoids: the evolution of diagnostic strategies. *J Clin Gastroenterol* 40:572–582
 201. Bhattacharyya S, Toumpanakis C, Caplin ME, Davar J (2008) Analysis of 150 patients with carcinoid syndrome seen in a single year at one institution in the first decade of the twenty-first century. *Am J Cardiol* 101:378–381 (LoE ⊗⊗⊗○)
 202. Roy P, Venzon DJ, Shojamanesh H, Abou-Saif A, Peghini P, Doppman JL, Gibril F, Jensen RT (2000) Zollinger-Ellison syndrome: clinical presentation in 261 patients. *Medicine (Baltimore)* 79:379–411 (LoE ⊗⊗○○)
 203. Roy P, Venzon DJ, Feigenbaum KM, Koviack PD, Bashir S, Ojeaburu JV, Gibril F, Jensen RT (2001) Gastric secretion in Zollinger-Ellison syndrome. Correlation with clinical expression, tumor extent and role in diagnosis—a prospective NIH study of 235 patients and a review of 984 cases in the literature. *Medicine (Baltimore)* 80:189–222 (LoE ⊗⊗○○)
 204. Jensen RT, Niederle B, Mitry E, Ramage JK, Steinmuller T, Lewington V, Scarpa A, Sundin A, Perren A, Gross D, O'Connor JM, Pauwels S, Kloppel G, Frascati Consensus Conference; European Neuroendocrine Tumor Society (2006) Gastrinoma (duodenal and pancreatic). *Neuroendocrinology* 84:173–182
 205. Ellison EC, Johnson JA (2009) The Zollinger-Ellison syndrome: a comprehensive review of historical, scientific, and clinical considerations. *Curr Probl Surg* 46:13–106
 206. Corleto VD, Annibale B, Gibril F, Angeletti S, Serrano J, Venzon DJ, Delle Fave G, Jensen RT (2001) Does the widespread use of proton pump inhibitors mask, complicate and/or delay the diagnosis of Zollinger-Ellison syndrome? *Aliment Pharmacol Ther* 15:1555–1561
 207. Gibril F, Schumann M, Pace A, Jensen RT (2004) Multiple endocrine neoplasia type 1 and Zollinger-Ellison syndrome. A prospective study of 107 cases and comparison with 1,009 patients from the literature. *Medicine (Baltimore)* 83:43–83 (LoE ⊗⊗⊗○)
 208. Kulke MH, Anthony LB, Bushnell DL, de Herder WW, Goldsmith SJ, Klimstra DS, Marx SJ, Pasiaka JL, Pommier RF, Yao JC, Jensen RT, North American Neuroendocrine Tumor Society (NANETS) (2010) NANETS treatment guidelines: well-differentiated neuroendocrine tumors of the stomach and pancreas. *Pancreas* 39:735–752

209. Ramundo V, Milone F, Severino R, Savastano S, Di Somma C, Vuolo L, De Luca L, Lombardi G, Colao A, Faggiano A (2011) Clinical and prognostic implications of the genetic diagnosis of hereditary NET syndromes in asymptomatic patients. *Horm Metab Res* 43:794–800
210. Gibril F, Jensen RT (2005) Advances in evaluation and management of gastrinoma in patients with Zollinger-Ellison syndrome. *Curr Gastroenterol Rep* 7:114–121
211. Kloppel G, Anlauf M (2007) Gastrinoma—morphological aspects. *Wien Klin Wochenschr* 119:579–584
212. Stabile BE, Morrow DJ, Passaro E Jr (1984) The gastrinoma triangle: operative implications. *Am J Surg* 147:25–31 (LoE ⊗○○○)
213. Vinik AI, Woltering EA, Warner RR, Caplin M, O’Dorisio TM, Wiseman GA, Coppola D, Go VL (2010) NANETS consensus guidelines for the diagnosis of neuroendocrine tumor. *Pancreas* 39:713–734
214. Jensen RT (2001) Zollinger-Ellison syndrome. In: Doherty GM, Skogseid B (eds) *Surgical endocrinology: clinical syndromes*. Lippincott Williams & Wilkins, Philadelphia, pp 291–344
215. Lau JY, Sung J, Hill C, Henderson C, Howden CW, Metz DC (2011) Systematic review of the epidemiology of complicated peptic ulcer disease: incidence, recurrence, risk factors and mortality. *Digestion* 84:102–113
216. Jensen RT (2006) Consequences of long-term proton pump blockade: highlighting insights from studies of patients with gastrinomas. *Basic Clin Pharmacol Toxicol* 98:4–19
217. Agréus L, Storskrubb T, Aro P, Ronkainen J, Talley NJ, Sipponen P (2009) Clinical use of proton-pump inhibitors but not H₂-blockers or antacid/alginates raises the serum levels of amidated gastrin-17, pepsinogen I and pepsinogen II in a random adult population. *Scand J Gastroenterol* 44:564–570 (LoE ⊗○○○)
218. Kim BW, Lee BI, Kim HK, Cho YS, Chae HS, Lee HK, Kim HJ, Han SW (2009) Influence of long-term gastric acid suppression therapy on the expression of serum gastrin, chromogranin A, and ghrelin. *Korean J Gastroenterol* 53:84–89 (LoE ⊗○○○)
219. Campana D, Piscitelli L, Mazzotta E, Bonora M, Serra C, Salomone L, Corinaldesi R, Tomassetti P (2005) Zollinger-Ellison syndrome. Diagnosis and therapy. *Minerva Med* 96:187–206
220. Poitras P, Gingras MH, Rehfeld JF (2012) The Zollinger-Ellison syndrome: dangers and consequences of interrupting antisecretory treatment. *Clin Gastroenterol Hepatol* 10:199–202 (LoE ⊗○○○)
221. Poitras P, Gingras MH, Rehfeld JF (2013) Secretin stimulation test for gastrin release in Zollinger-Ellison syndrome: to do or not to do? *Pancreas* 42:903–904
222. Desir B, Poltras P (2001) Oral pantoprazole for acid suppression in the treatment of patients with Zollinger-Ellison syndrome. *Can J Gastroenterol* 15:795–798 (LoE ⊗○○○)
223. Berna MJ, Hoffmann KM, Long SH, Serrano J, Gibril F, Jensen RT (2006) Serum gastrin in Zollinger-Ellison syndrome. II. Prospective study of gastrin provocative testing in 293 patients from the National Institutes of Health and comparison with 537 cases from the literature. Evaluation of diagnostic criteria, proposal of new criteria, and correlations with clinical and tumoral features. *Medicine (Baltimore)* 85:331–364 (LoE ⊗○○○)
224. Fraker DL, Norton JA, Alexander HR, Venzon DJ, Jensen RT (1994) Surgery in Zollinger-Ellison syndrome alters the natural history of gastrinoma. *Ann Surg* 220:320–330 (LoE ⊗○○○)
225. Norton JA, Fraker DL, Alexander HR, Gibril F, Liewehr DJ, Venzon DJ, Jensen RT (2006) Surgery increases survival in patients with gastrinoma. *Ann Surg* 244:410–419 (LoE ⊗○○○)
226. Morrow EH, Norton JA (2009) Surgical management of Zollinger-Ellison syndrome; state of the art. *Surg Clin North Am* 89:1091–1103
227. Senniappan S, Shanti B, James C, Hussain K (2012) Hyperinsulinaemic hypoglycaemia: genetic mechanisms, diagnosis and management. *J Inherit Metab Dis* 35:589–601
228. McAulay V, Deary IJ, Frier BM (2001) Symptoms of hypoglycaemia in people with diabetes. *Diabet Med* 18:690–705
229. Cox D, Gonder-Frederick L, McCall A, Kovatchev B, Clarke W (2002) The effects of glucose fluctuation on cognitive function and QOL: the functional costs of hypoglycaemia and hyperglycaemia among adults with type 1 or type 2 diabetes. *Int J Clin Pract* 56 (Suppl Jul 129):20–26
230. Okabayashi T, Shima Y, Sumiyoshi T, Kozuki A, Ito S, Ogawa Y, Kobayashi M, Hanazaki K (2013) Diagnosis and management of insulinoma. *World J Gastroenterol* 19:829–837
231. Cryer PE, Axelrod L, Grossman AB, Heller SR, Montori VM, Seaquist ER, Service FJ, Endocrine Society (2009) Evaluation and management of adult hypoglycemic disorders: an Endocrine Society Clinical Practice Guideline. *J Clin Endocrinol Metab* 94:709–728
232. Davda R, Seddon BM (2007) Mechanisms and management of non-islet cell tumour hypoglycaemia in gastrointestinal stromal tumour: case report and a review of published studies. *Clin Oncol (R Coll Radiol)* 19:265–268 (LoE ⊗○○○)
233. Waickus CM, de Bustros A, Shakil A (1999) Recognizing factitious hypoglycemia in the family practice setting. *J Am Board Fam Pract* 12:133–136 (LoE ⊗○○○)
234. Placzkowski KA, Vella A, Thompson GB, Grant CS, Reading CC, Charboneau JW, Andrews JC, Lloyd RV, Service FJ (2009) Secular trends in the presentation and management of functioning insulinoma at the Mayo Clinic, 1987–2007. *J Clin Endocrinol Metab* 94:1069–1073 (LoE ⊗○○○)
235. Toiari M, Davi MV, Dalle Carbonare L, Boninsegna L, Castellani C, Falconi M, Francia G (2013) Presentation, diagnostic features and glucose handling in a monocentric series of insulinoma. *J Endocrinol Invest* 36:753–758 (LoE ⊗○○○)
236. Phan GQ, Yeo CJ, Hruban RH, Lillemoie KD, Pitt HA, Cameron JL (1998) Surgical experience with pancreatic and peripancreatic neuroendocrine tumors: review of 125 patients. *J Gastrointest Surg* 2:473–482 (LoE ⊗○○○)
237. Mathur A, Gorden P, Libutti SK (2009) Insulinoma. *Surg Clin North Am* 89:1105–1121
238. Dizon AM, Kowalyk S, Hoogwerf BJ (1999) Neuroglycopenic and other symptoms in patients with insulinomas. *Am J Med* 106:307–310 (LoE ⊗○○○)
239. Starke A, Saddig C, Kirch B, Tschahargane C, Goretzki P (2006) Islet hyperplasia in adults: challenge to preoperatively diagnose non-insulinoma pancreatogenic hypoglycemia syndrome. *World J Surg* 30:670–679 (LoE ⊗○○○)
240. van Bon AC, Benhadi N, Endert E, Fliers E, Wiersinga WM (2009) Evaluation of endocrine tests. D: the prolonged fasting test for insulinoma. *Neth J Med* 67:274–278 (LoE ⊗○○○)
241. Baudin E, Caron P, Lombard-Bohas C, Tabarin A, Mitry E, Reznick Y, Taieb D, Pattou F, Goudet P, Vezzosi D, Scoazec JY, Cadiot G, Borson-Chazot F, Do Cao C, on behalf of the Société française d’endocrinologie and the Groupe d’étude des tumeurs endocrines (2013) Malignant insulinoma: recommendations for characterisation and treatment. *Ann Endocrinol* 74:523–533
242. McLean A (2004) Endoscopic ultrasound in the detection of pancreatic islet cell tumours. *Cancer Imaging* 4:84–91
243. Liu Y, Song Q, Jin HT, Lin XZ, Chen KM (2009) The value of multidetector-row CT in the preoperative detection of pancreatic insulinomas. *Radiol Med* 114:1232–1238 (LoE ⊗○○○)
244. Anaye A, Mathieu A, Closset J, Bali MA, Metens T, Matos C (2009) Successful preoperative localization of a small pancreatic insulinoma by diffusion-weighted MRI. *JOP* 10:528–531 (LoE ⊗○○○)

245. Minn H, Kauhanen S, Seppänen M, Nuutila P (2009) ^{18}F -FDOPA: a multiple-target molecule. *J Nucl Med* 50:1915–1918
246. Wild D, Christ E, Caplin ME, Kurzawinski TR, Forrer F, Brändle M, Seufert J, Weber WA, Bomanji J, Perren A, Ell PJ, Reubi JC (2011) Glucagon-like peptide-1 versus somatostatin receptor targeting reveals 2 distinct forms of malignant insulinomas. *J Nucl Med* 52:1073–1078 (LoE ⊗⊗○○)
247. Wiesli P, Uthoff H, Perren A, Pfammatter T, Zwimpfer C, Seiler H, Kindhauser R, Spinas GA, Schmid C (2011) Are biochemical markers of neuroendocrine tumors coreleased with insulin following local calcium stimulation in patients with insulinomas? *Pancreas* 40:995–999 (LoE ⊗○○○)
248. Guettier JM, Kam A, Chang R, Skarulis MC, Cochran C, Alexander HR, Libutti SK, Pingpank JF, Gorden P (2009) Localization of insulinomas to regions of the pancreas by intraarterial calcium stimulation: the NIH experience. *J Clin Endocrinol Metab* 94:1074–1080 (LoE ⊗⊗○○)
249. Proye C, Malvaux P, Pattou F, Filoche B, Godchaux JM, Maunoury V, Palazzo L, Huglo D, Lefebvre J, Paris JC (1998) Noninvasive imaging of insulinomas and gastrinomas with endoscopic ultrasonography and somatostatin receptor scintigraphy. *Surgery* 124:1134–1143 discussion 1143–1144 (LoE ⊗⊗⊗○)
250. Bellizzi AM (2013) Assigning site of origin in metastatic neuroendocrine neoplasms: a clinically significant application of diagnostic immunohistochemistry. *Adv Anath Pathol* 20:285–314
251. Kirshbom PM, Kherani AR, Onaitis MW, Feldman JM, Tyler DS (1998) Carcinoids of unknown origin: comparative analysis with foregut, midgut, and hindgut carcinoids. *Surgery* 124:1063–1070 (LoE ⊗⊗○○)
252. Pavel M, Baudin E, Couvelard A, Krenning A, Öberg K, Steinmüller T, Anlauf M, Wiedenmann B, Salazar R, all other Barcelona Consensus Conference participants (2012) ENETS consensus guidelines for the management of patients with liver and other distant metastases from neuroendocrine neoplasms of foregut, midgut, hindgut, and unknown primary. *Neuroendocrinology* 95:157–176
253. Catena L, Bichisao E, Milione M, Valente M, Platania M, Pusceddu S, Ducceschi M, Zilembo N, Formisano B, Bajetta E (2011) Neuroendocrine tumors of unknown primary site: gold dust or misdiagnosed neoplasms? *Tumori* 97:564–567
254. NCCN clinical practice guidelines in oncology. Neuroendocrine tumors. Version 2.2014. http://www.nccn.org/professionals/physician_gls/pdf/neuroendocrine.pdf. Accessed 21 Jan 2014
255. Srivastava A, Hornick JL (2009) Immunohistochemical staining for CDX-2, PDX-1, NESP-55, and TTF-1 can help distinguish gastrointestinal carcinoid tumors from pancreatic endocrine and pulmonary carcinoid tumors. *Am J Surg Pathol* 33:626–632 (LoE ⊗⊗○○)
256. Joseph S, Wang YZ, Boudreaux JP, Anthony LB, Campeau R, Raines D, O'Dorisio T, Go VL, Vinik AI, Cundiff J, Woltering EA (2011) Neuroendocrine tumors: current recommendations for diagnosis and surgical management. *Endocrinol Metab Clin N Am* 40:205–231
257. Wang SC, Parekh JR, Zuraek MB, Venook AP, Bergsland EK, Warren RS, Nakakura EK (2010) Identification of unknown primary tumors in patients with neuroendocrine liver metastases. *Arch Surg* 145:276–280
258. Gabriel M, Decristoforo C, Kendler D, Dobrozemsky G, Heute D, Uprimny C, Kovacs P, Von Guggenberg E, Bale R, Virgolini IJ (2007) ^{68}Ga -DOTA-Tyr³-octreotide PET in neuroendocrine tumors: comparison with somatostatin receptor scintigraphy and CT. *J Nucl Med* 48:508–518 (LoE ⊗⊗⊗○)
259. Wang SC, Fidelman N, Nakakura EK (2013) Management of well-differentiated gastrointestinal neuroendocrine tumors to the liver. *Semin Oncol* 40:69–74
260. Giesel FL, Kratochwil C, Mehndiratta A, Wulfert S, Moltz JH, Zechmann CM, Kauczor HU, Haberkorn U, Ley S (2012) Comparison of neuroendocrine tumor detection and characterization using DOTATOC-PET in correlation with contrast enhanced CT and delayed contrast enhanced MRI. *Eur J Radiol* 81:2820–2825 (LoE ⊗⊗○○)
261. Lindholm DP, Oberg K (2011) Biomarkers and molecular imaging in gastroenteropancreatic neuroendocrine tumors. *Horm Metab Res* 43:832–837
262. Ichikawa T, Peterson MS, Federle MP, Baron RL, Haradome H, Kawamori Y, Nawano S, Araki T (2000) Islet cell tumor of the pancreas: biphasic CT versus MR imaging in tumor detection. *Radiology* 216:163–171 (LoE ⊗⊗○○)