

Research Article

The Neuroprotective Effects of *Ratanasampil* on Oxidative Stress-Mediated Neuronal Damage in Human Neuronal SH-SY5Y Cells

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Received 29 December 2014; Revised 13 April 2015; Accepted 16 April 2015

Academic Editor: Vincent Pialoux

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We previously found that *Ratanasampil* (RNSP), a traditional Tibetan medicine, improves the cognitive function of mild-to-moderate AD patients living at high altitude, as well as learning and memory in an AD mouse model (Tg2576); however, mechanism underlying the effects of RNSP is unknown. In the present study, we investigated the effects and molecular mechanisms of RNSP on oxidative stress-induced neuronal toxicity using human neuroblastoma SH-SY5Y cells. Pretreatment with RNSP significantly ameliorated the hydrogen peroxide- (H_2O_2 -) induced cytotoxicity of SH-SY5Y cells in a dose-dependent manner (up to 60 $\mu\text{g/mL}$). Furthermore, RNSP significantly reduced the H_2O_2 -induced upregulation of 8-oxo-2'-deoxyguanosine (8-oxo-dG, the oxidative DNA damage marker) but significantly reversed the expression of repressor element-1 silencing transcription factor (REST) from H_2O_2 associated (100 μM) downregulation. Moreover, RNSP significantly attenuated the H_2O_2 -induced phosphorylation of p38 mitogen-activated protein kinase (MAPK) and extracellular signal-regulated kinase 1/2 (ERK 1/2) in SH-SY5Y cells. These observations strongly suggest that RNSP may protect the oxidative stress-induced neuronal damage that occurs through the properties of various antioxidants and inhibit the activation of MAPKs. We thus provide the principle molecular mechanisms of the effects of RNSP and indicate its role in the prevention and clinical management of AD.

1. Introduction

By the year 2030, roughly 20% of the world's population will be over the age of 65 years [1]. As the mean life expectancy continues to increase, the cognitive impairments associated with Alzheimer's disease (AD) will be one of the major issues in aging societies around the world. Oxidative stress is a major component of the harmful cascades activated in the development of aging-related neurodegenerative disorders, including AD [2–4], because oxidative stress through the overproduction of reactive oxygen species (ROS) causes cell damage through the promotion of lipid peroxidation, DNA damage, and the regulation of death proteins [5]. Numerous

reports have provided direct morphological and biochemical evidence indicating the connection between oxidative stress and cell death in the brain of AD patients [6–8]. Antioxidant therapy is considered to be one of the promising approaches to the prevention and clinical management of AD [9].

Mitogen-activated protein kinase (MAPK) cascades are sensitive to oxidative stress. The overproduction of ROS during oxidative stress plays a role as a second messenger in signal transduction cascades leading to the activation of MAPKs, because the intracellular redox state of the cells regulates the cellular signaling pathways [10, 11]. The members of the major MAPK subfamily, c-Jun N-terminal kinase (JNK), p38 MAPK, and extracellular signal-regulated kinase

1/2 (ERK 1/2), are known cell death factors that increase in response to oxidative stress [12, 13]. Indeed, increased levels of phosphorylated MAPKs have been found in the postmortem examinations of the brains of AD patients [14–16].

Ratanasampil (RNSP), the Tibetan language name which translates to “seventy-taste pearl-balls,” has been traditionally used for the treatment of stroke and cerebrovascular diseases. Our previous studies have found that RNSP increases learning and memory and reduces β -amyloid ($A\beta$) protein levels in mouse AD models (Tg2576) [17, 18]. Furthermore, we have recently found that RNSP improves the cognitive functions in mild-to-moderate AD patients living at high altitude and decreases serum $A\beta_{42}$ concentration as well as proinflammatory factors, including Tumor Necrosis Factor (TNF- α) and interleukins IL-1 β and IL-6. These findings indicate the therapeutic effects of RNSP in the prevention and clinical management of AD [19]. However, the mechanism underlying these effects has remained unknown.

RNSP is composed of more than seventy components, including saffron and *Glycyrrhiza uralensis* [20, 21]. The beneficial effects of RNSP components have been received. Previous studies have shown that saffron effectively inhibited the TNF α -induced cell death of PC12 cells [22] and protected neurons from the neurotoxic activity of 6-hydroxydopamine hydrobromide [23], via its antioxidant properties [24–26]. More recent studies have reported that *Glycyrrhiza uralensis* exerts anti-inflammatory effects by inhibiting the activation of nuclear factor-kappa B (NF κ B) as well as the NOD-like receptor family, a pyrin domain containing 3 (NLRP3) inflammasome [27, 28]. In the present study, we focused on the effects and molecular mechanisms of RNSP in hydrogen peroxide- (H_2O_2 -) induced neuronal death using cultured SH-SY5Y cells, because H_2O_2 is one of the most important ROS generated during oxidative stress [29, 30] and because SH-SY5Y cells are widely used for the study of neuronal cell death induced by oxidative stress [31, 32]. Pretreatment with RNSP was found to significantly ameliorate the H_2O_2 -induced cytotoxicity of SH-SY5Y cells in a dose-dependent manner (from 10 μ g/mL to 60 μ g/mL). Furthermore, RNSP was found to significantly reduce the H_2O_2 -upregulated expression of 8-oxo-deoxyguanosine (8-oxo-G, major product of DNA oxidation that is widely used as an oxidative DNA damage marker) but significantly reserved repressor element-1 silencing transcription factor (REST) from H_2O_2 -related downregulation. Moreover, RNSP significantly attenuated the H_2O_2 -induced phosphorylation of p38 and ERK1/2 in SH-SY5Y cells. These observations strongly suggest that RNSP may provide a protective effect against oxidative stress-induced neuronal death via its antioxidant properties and through the inhibition of MAPK activation. The findings of this study may thus provide the principle molecular mechanisms supporting the clinical usage of RNSP in the prevention and clinical management of AD.

2. Materials and Methods

2.1. Reagents. *Ratanasampil* (RNSP, Zhunzi Z63020062) was purchased from Qinghai Jinke Tibetan Medicine Pharmaceutical Co. Ltd. (Xining, China). A suitable concentration

of methanol was titrated for cell culture in order to prevent interference from the methanol solvent. H_2O_2 (30%) was purchased from Sigma-Aldrich (St. Louis, MO, USA). Mouse monoclonal anti-8-oxo-deoxyguanosine (8-oxo-dG) was purchased from NOF Corporation (Kyoto, Japan); rabbit anti-phospho-p38, rabbit anti-p38, rabbit anti-phospho-ERK1/2, rabbit anti-phospho-ERK1/2, rabbit anti-phospho-pJNK (1:1000), and rabbit anti-pJNK (1:1000) were purchased from Cell Signaling Technology (Danvers, MA, USA).

2.2. SH-SY5Y Cell Culture. Cells of the SH-SY5Y human neuroblastoma cell line, which were purchased from American Type Culture Collection (Manassas, VA, USA), were cultured in DMEM/F-12 mixture supplemented with 10% fetal bovine serum (FBS, ICN Biomedicals, Eschwege, Germany), 2 mM L-glutamine, and 1% antibiotic and antimycotic solution (Sigma, St. Louis, MO, USA) in a humid atmosphere of 5% CO_2 and 95% air at 37°C. To determine the toxicity of the reagents, the cells were treated with freshly prepared H_2O_2 (from 30% stock) at concentrations ranging from 5 to 500 μ M (diluted with sterile purified water), and RNSP was tested at concentrations ranging from 0.01 to 100 μ g/mL (diluted with methanol or sterile purified water) for different treatment times.

2.3. Cell Survival Assay. An MTT assay was performed as described previously [33]. Based on the preliminary observations, cells in the exponential phase of growth were seeded in 96-well plates (1×10^5 cells/well), allowed to adhere to plates (for 24 h), and treated with various concentrations of RNSP (diluted with methanol or sterile purified water), H_2O_2 (diluted with sterile purified water), and vehicle (methanol or sterile purified water). The relative cell viability was measured using CellQuanti-MTT Cell Viability Assay Kits (Bio Assay Systems, Hayward, USA). The assay was performed according to the manufacturer's protocol. The absorbency at 570 nm was evaluated using a microplate reader.

2.4. Observations of Morphological Changes. The cells were seeded in 24-well plates (2×10^5 cells/well) for 24 h and then treated with H_2O_2 at a concentration of 100 μ M for 24 h with or without RNSP (methanol extraction) at different concentrations. The cellular morphology was observed and photographed using a bright-field microscope (Nikon, ECLIPSE Ti-S, Japan).

2.5. Detection of Mitochondrial ROS. Mitochondrial ROS were measured using MitoSOX Red (Invitrogen, USA), which is a live-cell permeant that rapidly and selectively targets mitochondria [33]. Once in the mitochondria, MitoSOX Red reagent is oxidized by superoxide and exhibits red fluorescence (with excitation at 510 nm and emission at 580 nm). The cultured SH-SY5Y cells were seeded in 24-well plates (2×10^5 cells/well) and incubated with or without RNSP for 2 h (methanol extraction, 60 μ g/mL). The cells were then further exposed to H_2O_2 (100 μ M) for 1 h. After incubation in Hank's balanced salt solution (HBSS) containing 5 mM MitoSOX Red for 10 min at 37°C, the cells were washed twice with PBS and then mounted in a warm buffer for imaging. Images

were collected using a $\times 20$ objective lens (NA = 0.50, 200x magnification, yielding a frame of 0.575 mm^2). The procedure resulted in arbitrary optical density values on a scale of 0 (background staining) to 255.

2.6. Real-Time Quantitative RT-PCR Analysis. SH-SY5Y cells were seeded in a 10 cm^2 dish (1×10^7 /dish), which was pretreated with RNSP (methanol extraction, $60 \mu\text{g/mL}$) for 2 h. The cells were then treated with H_2O_2 ($100 \mu\text{M}$) for 1 or 2 h. The mRNAs isolated from the RNSP-pretreated or nontreated cells were subjected to a real-time quantitative RT-PCR. The total RNA was extracted with a PureLink RNA micro kit (Invitrogen, Japan), which was used in accordance with the manufacturer's instructions. A total of 800 ng of extracted RNA were reverse-transcribed to cDNA using High Capacity RNA-to-cDNA Master Mix (Applied Biosystems, Foster City, CA). Thermal cycling was conducted at 50°C for 2 min and then at 95°C for 10 min, followed by 40 cycles of 95°C for 15 s and 60°C for 1 min. The cDNA was amplified in duplicate using TaqMan Universal PCR Master Mix (Applied Biosystems, Foster City, CA) with an Applied Biosystems 7500/7500 Fast Real-Time PCR System. The data were evaluated using the 7500 software program (version 2.0, Applied Biosystems). The following primer sequences were used: repressor element-1 silencing transcription factor (REST): 5'-TGG TGG GTG CCC AAA TTG TA-3' and 5'-ACC TGC ATG GGA GCA GAT TC-3'. For data normalization, an endogenous control (actin) was assessed to control for the cDNA input, and the relative units were calculated using a comparative Ct method. All of the real-time RT-PCR experiments were repeated three times, and the results are presented as the means of the ratios \pm SEM.

2.7. Immunofluorescence Imaging. Immunofluorescence imaging was performed as described previously [34]. Briefly, SH-SY5Y cells were seeded in 24-well plates (2×10^5 cells/well) for 24 h, then treated with H_2O_2 ($100 \mu\text{M}$) for 4 h, with or without pretreatment with RNSP (methanol extraction, $60 \mu\text{g/mL}$) for 2 h, and fixed with 4% paraformaldehyde. After washing the cells with PBS twice, they were incubated with mouse anti-8-oxo-dG (1:500) overnight at 4°C and then incubated with anti-mouse Alexa 488 (1:2000, Jackson ImmunoResearch Lab. Inc.) at 4°C for 2 h. The cells were mounted in the antifading medium, VECTASHIELD, and the fluorescence images were taken using a confocal laser scanning microscope (CLSM, C2si, Nikon, Japan).

2.8. Electrophoresis and Immunoblotting. SH-SY5Y cells were cultured at a density of 1×10^7 cells in 10 cm^2 dishes. The cytosolic samples of the cells were collected at 10, 30, and 60 min after H_2O_2 ($100 \mu\text{M}$) treatment, with or without RNSP (methanol extraction, $60 \mu\text{g/mL}$). The lysed samples were electrophoresed in 12% SDS-polyacrylamide gels, and the proteins on the SDS gels were electrophoretically transferred to nitrocellulose membranes. Following blocking, the membranes were incubated at 4°C overnight under gentle agitation with each primary antibody: rabbit anti-phospho-p38 (1:1000), rabbit anti-p38 (1:1000),

rabbit anti-phospho-pERK1/2 (1:1000), rabbit anti-phospho-ERK1/2 (1:1000), rabbit anti-phospho-pJNK (1:1000), rabbit anti-pJNK (1:1000), and rabbit anti-phospho-pERK1/2 (1:1000) antibodies. After washing, the membranes were incubated with horseradish peroxidase- (HRP-) labeled anti-rabbit (1:2000, GE Healthcare, UK) antibody for 2 h at 24°C . The protein bands were then detected using an enhanced chemiluminescence detection system (ECK kit, Amersham Pharmacia Biotech) with an image analyzer (LAS-4000, Fuji Photo Film, Tokyo, Japan).

2.9. Statistical Analysis. The data are represented as the means \pm SEM. Statistical analyses were performed using one-way or two-way analysis of variance (ANOVA) with post hoc Tukey's test using the GraphPad Prism software package. A *p* value of <0.05 was considered to indicate statistical significance (GraphPad Software Inc., San Diego, CA, USA).

3. Results

3.1. Effect of RNSP on Cell Viability in SH-SY5Y Cells. We first examined the effects of RNSP on the cell viability of SH-SY5Y cells using an MTT assay. The mean cell viability was not significantly changed after treatment with the methanol extracts of RNSP at final concentrations between 0.3 and $60 \mu\text{g/mL}$ for 5 days (Figure 1(a)). However, the mean cell viability was significantly reduced after pretreatment with RNSP at a final concentration of over $100 \mu\text{g/mL}$ (82% of viable cells). It is known that the concentration of H_2O_2 in healthy individuals is normally quite low, and the elevation of H_2O_2 concentration upon cerebral ischemic and reperfusion injury could act as a significant signal [35, 36]. We next examined the toxic effect of H_2O_2 at different concentrations ($5\text{--}500 \mu\text{M}$) in SH-SY5Y cells, because H_2O_2 has frequently been used as an oxidative stimulus to identify redox-sensitive processes. The viability of SH-SY5Y cells was decreased after exposure to H_2O_2 for 24 h in a dose-dependent manner. In comparison to nonexposed cells, 56.8% of the viable cells were detected at an H_2O_2 concentration of $100 \mu\text{M}$, while less than 50% of the viable cells were detected at an H_2O_2 concentration of over $125 \mu\text{M}$ (Figure 1(b)). We therefore used methanol extracts of RNSP at concentrations of up to $60 \mu\text{g/mL}$ and H_2O_2 at $100 \mu\text{M}$ concentration to investigate the effects of RNSP on the H_2O_2 -induced cytotoxicity of SH-SY5Y cells in the subsequent experiments.

3.2. The Effects of RNSP on H_2O_2 -Induced Toxicity in SH-SY5Y Cells. Pretreatment with methanol extracts of RNSP for 5 days prevented H_2O_2 -induced cytotoxicity in SH-SY5Y cells in a dose-dependent manner (Figure 1(c)). In comparison to the H_2O_2 -treated cells, the cell viabilities were significantly restored from treatment with RNSP at a concentration of $10 \mu\text{g/mL}$, and 92.8% of viable cells were detected in RNSP with a concentration of $60 \mu\text{g/mL}$. The protective effect of RNSP was further confirmed by the microscopic evaluation of the cells. Morphological changes that were observed included the degeneration of H_2O_2 -treated SH-SY5Y cells, which exhibited the disappearance of the neuritis and shrinkage (Figure 1(d)). The percentage of viable cells was

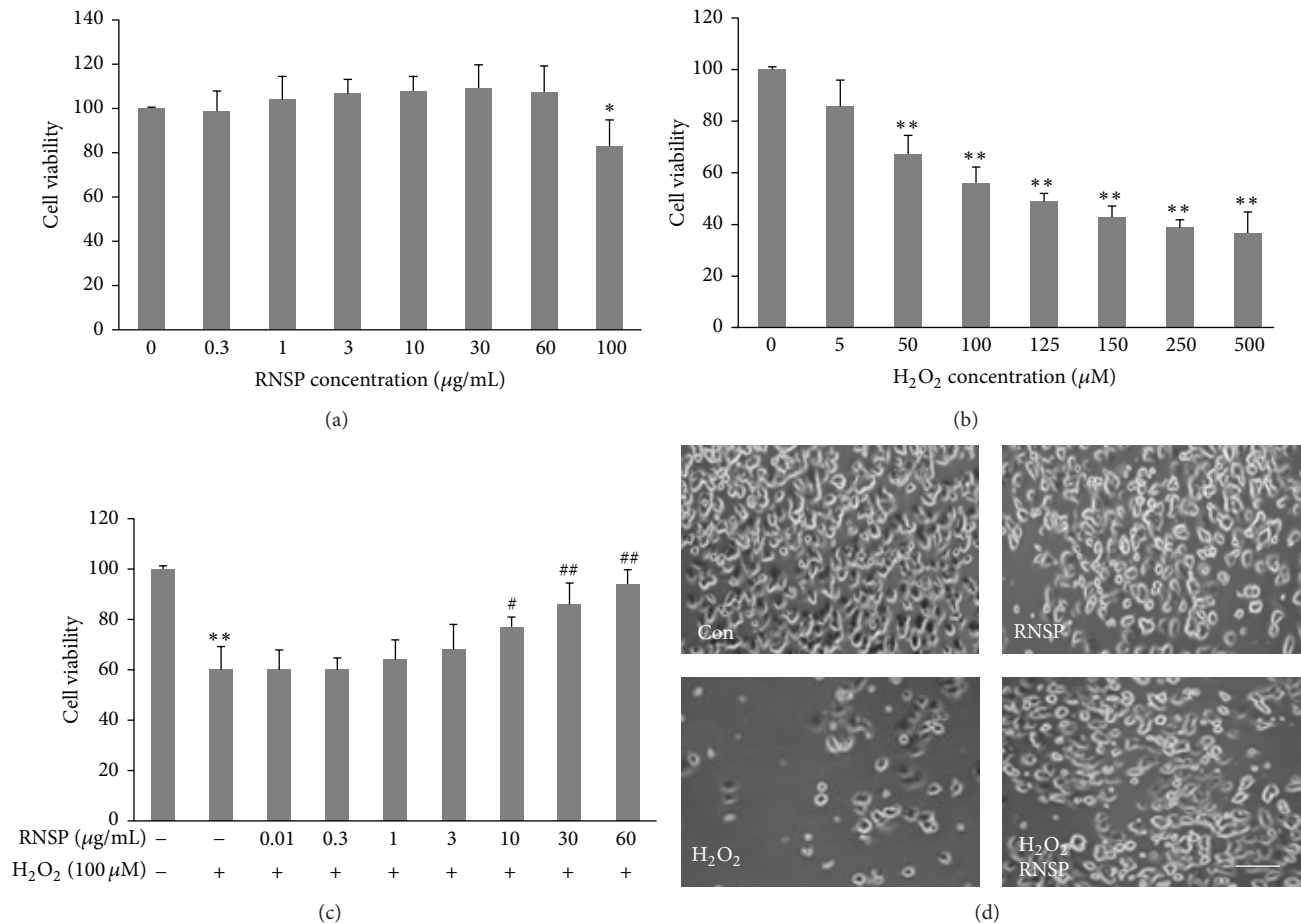


FIGURE 1: The effects of RNSP on H₂O₂-induced toxicity in SH-SY5Y cells. (a) The cell viability of SH-SY5Y cells after treatment with RNSP for 5 days. (b) The cell viability of SH-SY5Y cells after 24 h of exposure to H₂O₂. (c) The effect of RNSP on H₂O₂-induced toxicity in SH-SY5Y cells. Pretreatment was performed with RNSP (5 days) before exposure to H₂O₂ (100 µM, 24 h). Each column and bar represent the mean ± SEM ($n = 4$ each). Asterisks indicate a statistically significant difference from the value in untreated cells (* $p < 0.05$, ** $p < 0.01$). Pound signs indicate a statistically significant difference from the value in H₂O₂-exposed cells without pretreatment with RNSP (# $p < 0.05$, ## $p < 0.01$). (d) The morphological changes of SH-SY5Y cells with or without pretreatment with RNSP (60 µg/mL, 5 days) after exposure to H₂O₂ for 24 h. Scale bar = 20 µm.

also reduced by H₂O₂ in a dose-dependent manner (data not shown). It is noted that the neuritis and shrinkage of cells were attenuated by pretreatment with RNSP (Figure 1(d)). To determine the effects of the soluble components of RNSP, we further examined the effects of water extracts of RNSP on H₂O₂-induced toxicity in SH-SY5Y cells in comparison to the methanol extracts of RNSP. As shown in Figures 2(a) and 2(b), pretreatment with the RNSP water extracts for 5 days reduced the H₂O₂-induced toxicity in SH-SY5Y cells in a dose-dependent manner. No significant difference was detected between the water and methanol extracts (Figures 2(a) and 2(b)). To further address the protective effects of RNSP over short time periods, pretreatment with methanol extracts of RNSP for 2 h as well as twice for 2 h with a 2 h interval (2-2 h) was performed in SH-SY5Y cells before exposure to H₂O₂ (24 h). As shown in Figures 2(c) and 2(d), pretreatment with RNSP for both 2 h and 2-2 h significantly inhibited the H₂O₂-induced cytotoxicity in SH-SY5Y cells in a dose-dependent manner, and no significant difference

was detected between the two treatment schedules in the effects on the H₂O₂-induced toxicity in cells. Taken together, these observations strongly demonstrate that pretreatment with RNSP protects SH-SY5Y cells from H₂O₂-induced cytotoxicity. Pretreatment with methanol extracts of RNSP (60 µg/mL) for 2 h was set up in the subsequent experiments.

3.3. The Effects of RNSP on H₂O₂-Induced DNA Damage in SH-SY5Y Cells. Oxidative stress is an important inducer of neurotoxicity in AD patients [37], causing damage to cardinal cellular components, including the DNA, and initiating subsequent cell death [38]. Following our previous experiments, we used two approaches to address the effects of RNSP on H₂O₂-induced DNA damage in SH-SY5Y cells: one approach was the use of a MitoSOX Red probe, as a marker for mitochondria-derived ROS generation [33], and the other was immunofluorescence imaging for a biomarker of oxidation-damaged DNA marker, 8-oxo-dG [39]. In comparison to the untreated cells, the expression of MitoSOX

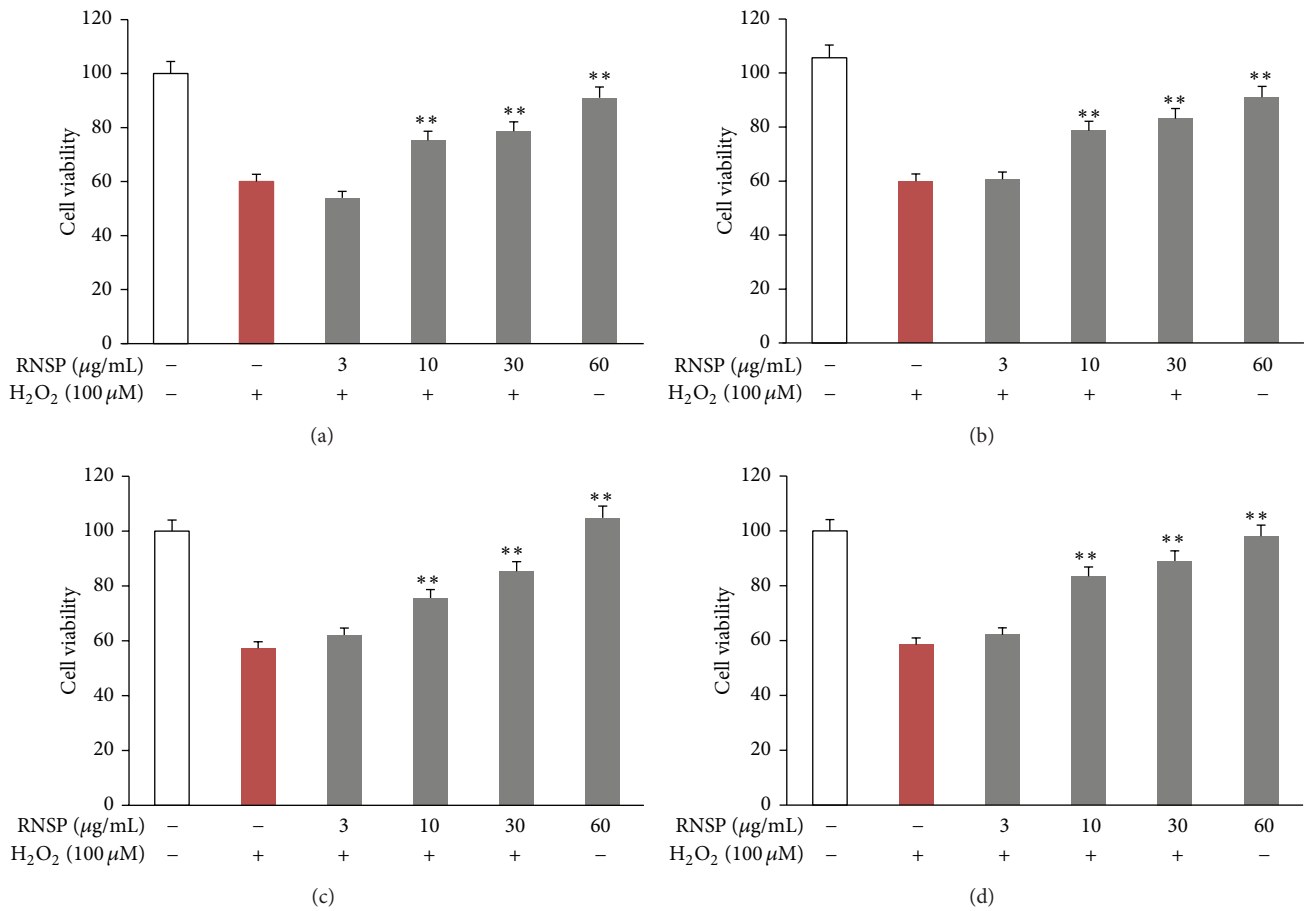


FIGURE 2: The effects of RNSP in different solvents and times on H₂O₂-induced toxicity in SH-SY5Y cells. Pretreatment with water extracts (a) and methanol extracts (b) of RNSP for 5 days. Each column and bar represent the mean \pm SEM ($n = 4$ each). Asterisks indicate a statistically significant difference from the value in the H₂O₂-exposed cells without RNSP (** $p < 0.01$). The effect of pretreatment with methanol extracts of RNSP for 2 h (c) and 2 h two times in a 2 h interval (2-2 h) (d) on H₂O₂-exposed SH-SY5Y cells. Each column and bar represent the mean \pm SEM ($n = 4$ each). Asterisks indicate a statistically significant difference from the value in the H₂O₂-incubated cells without RNSP (** $p < 0.01$).

Red signals was significantly increased in SH-SY5Y cells after exposure to H₂O₂ for 1 h, suggesting that the mitochondria are the early origin of ROS generation during oxidative stress. Pretreatment with RNSP significantly inhibited the H₂O₂-induced mitochondria-derived ROS generation in SH-SY5Y cells (Figure 3(a)), thus confirming the antioxidant properties of RNSP. Immunofluorescence imaging showed a significant inverse relationship between Hoechst and 8-oxo-dG after exposure of SH-SY5Y cells to H₂O₂ for 4 h (Figure 3(c)), and the mean fluorescent intensity of 8-oxo-dG was found to significantly increase in comparison to that in the cells that were not exposed to H₂O₂ (7.22 versus 2.75, *** $p < 0.001$). It is noted that pretreatment with the RNSP methanol extracts for 2 h significantly reduced the immunofluorescence intensity of 8-oxo-dG in the H₂O₂-exposed SH-SY5Y cells (3.60 versus 7.22, ### $p < 0.001$, Figures 3(c) and 3(d)). These observations demonstrate that RNSP could attenuate oxidative stress-induced DNA damage in neuronal cells.

3.4. The Effects of RNSP on the H₂O₂-Induced Downregulation of REST in SH-SY5Y Cells. Recently, the neuroprotective role of repressor element-1 silencing transcription factor (REST) has been shown in the repression of ROS production and oxidative stress [40]. We thus examined the effects of RNSP on H₂O₂-induced REST expression in SH-SY5Y cells. A real-time quantitative RT-PCR analysis showed that the REST expression was significantly reduced after the exposure of SH-SY5Y cells to 100 µM of H₂O₂ (reduced to 85% at 1 h and 68% at 2 h in comparison to the nonexposed cells, Figure 3(b)), indicating that the downregulation of REST expression is preceded to H₂O₂-induced oxidative DNA damage. To our surprise, pretreatment with methanol extracts of RNSP for 2 h significantly reversed the H₂O₂-reduced the expression of REST from 1 h (reversed 87.8% at 1 h and 87.8% at 2 h in comparison with H₂O₂-exposed cells, Figure 3(b)). This observation suggests that the reversion of RNSP in H₂O₂-reduced REST expression may be involved in preventing oxidative DNA damage in SH-SY5Y cells.

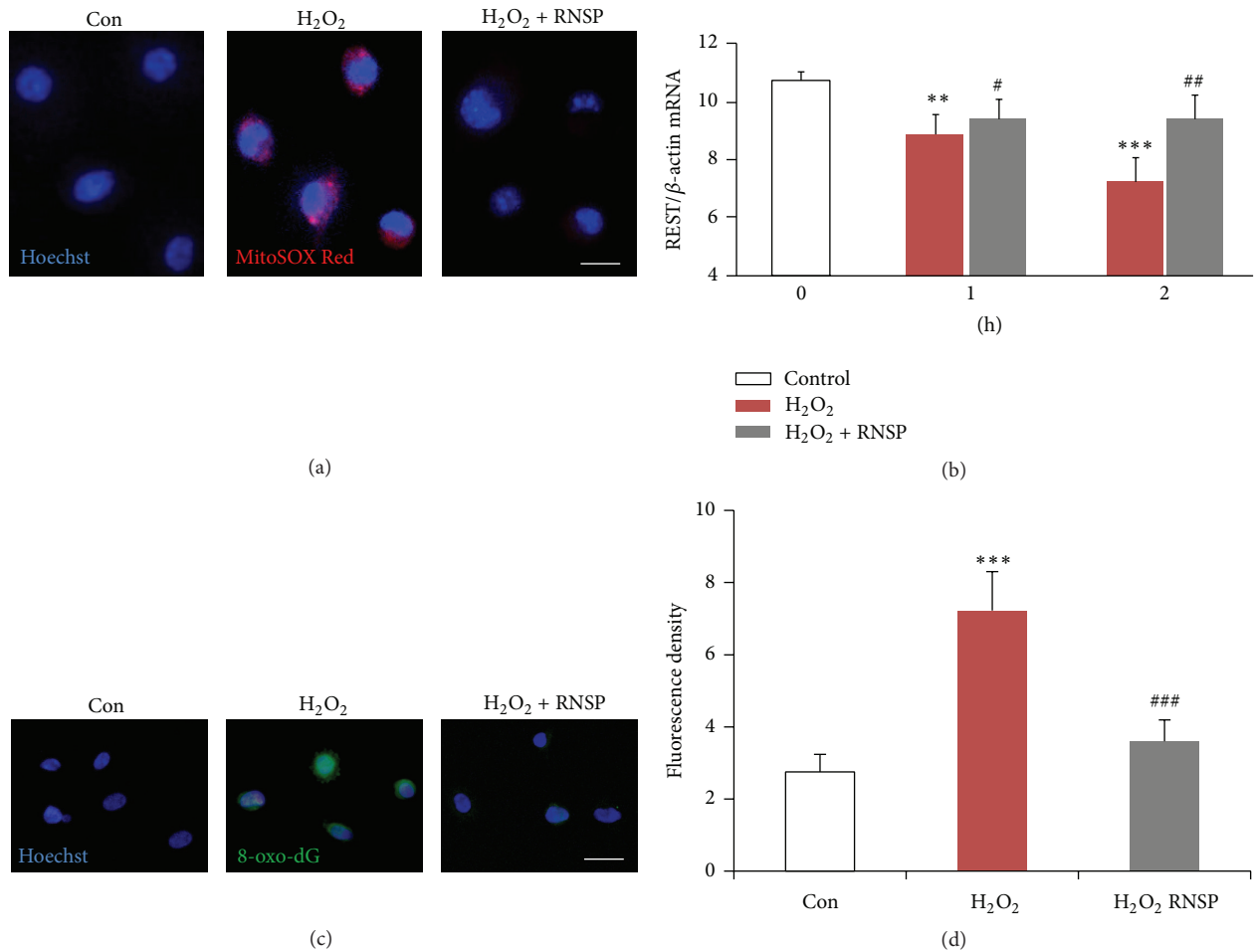


FIGURE 3: The effects of RNSP on H₂O₂-induced DNA damage in SH-SY5Y cells. (a) Fluorescent images of MitoSOX Red signals in the H₂O₂-incubated cells (100 μM, 1 h) with or without RNSP (60 μg/mL, 2 h). Scale bar = 10 μm. (b) REST expression, as analyzed by real-time quantitative RT-PCR, in the H₂O₂-exposed cells (100 μM, 1 h, 2 h), with or without RNSP (60 μg/mL, 2 h). Each column and bar represent the mean ± SEM ($n = 4$ each). Asterisks indicate a statistically significant difference from the value in the untreated cells (** $p < 0.01$, *** $p < 0.001$). Pound signs indicate a statistically significant difference from the value in the H₂O₂-incubated cells without RNSP (# $p < 0.05$, ## $p < 0.01$). (c) Immunofluorescent CLMS images of 8-oxo-dG (green) with Hoechst-stained nuclei (blue) in the H₂O₂-exposed cells (100 μM, 4 h), with or without RNSP (60 μg/mL, 2 h). Scale bar = 10 μm. (d) The quantitative analyses of 8-oxo-dG immunofluorescence signal intensity in (c). Each column and bar represent the mean ± SEM ($n = 4$ each). Asterisks indicate a statistically significant difference from the value in the untreated cells (** $p < 0.01$, *** $p < 0.001$). Pound signs indicate a statistically significant difference from the value in the H₂O₂-exposed cells without RNSP (## $p < 0.01$, ### $p < 0.001$).

3.5. The Effects of RNSP on H₂O₂-Mediated MAPK Activation in SH-SY5Y Cells. We finally examined the effects of RNSP on H₂O₂-induced p38, ERK1/2, and JNK phosphorylation in SH-SY5Y cells, because they are known to be cell death factors which are mediated by oxidative stress [12, 13]. In comparison to the cells that were not exposed to H₂O₂, the mean level of phosphorylated p38 MAPK was significantly increased in SH-SY5Y cells from 30 min to 60 min after exposure to H₂O₂ (100 μM) (Figures 4(a) and 4(b)). Along the same lines, the phosphorylated ERK 1/2 MAPK was significantly increased from 60 min after exposure to H₂O₂ (Figures 4(c) and 4(d)). However, the mean level of phosphorylated JNK was not significantly increased in SH-SY5Y cells after exposure to H₂O₂ (data not shown). Interestingly, pretreatment with the methanol extracts of RNSP for 2 h significantly inhibited

the H₂O₂-induced phosphorylation of both p38 and ERK 1/2 MAPK from 60 min after incubation with H₂O₂ in SH-SY5Y cells (Figures 4(a)–4(d)). However, pretreatment with RNSP for 2 h did not significantly inhibit the phosphorylation of JNK in the SH-SY5Y cells. These observations demonstrate that RNSP could inhibit oxidative stress-induced MAPK activation in SH-SY5Y cells.

4. Discussion

The major findings of the present study are that RNSP protects SH-SY5Y cells from H₂O₂-induced neuronal cytotoxicity by reducing the oxidative stress-induced DNA damage and activation of MAPK (summarized in Figure 5). It

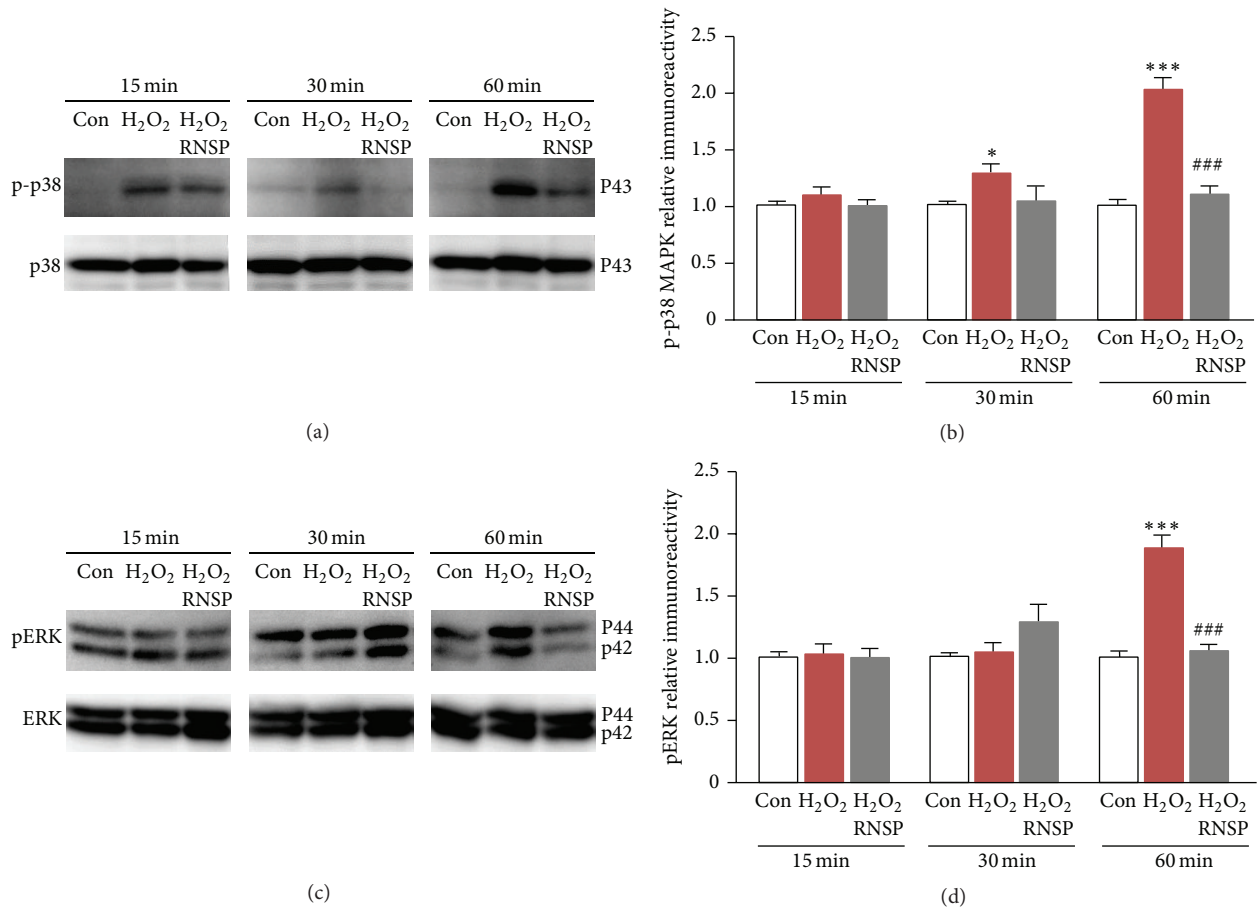


FIGURE 4: The effects of RNSP on H₂O₂-mediated MAPKs activation in SH-SY5Y cells. (a) The phosphorylation of p38 in H₂O₂-exposed cells with or without RNSP (60 μg/mL, 2 h). (b) The quantitative analyses of the immunoblotting from (a). Each column and bar represent the mean ± SEM (n = 4, each). An asterisk indicates a statistically significant difference from the value in the untreated cells (*p < 0.05, ***p < 0.001). Pound signs indicate a statistically significant difference from the value in H₂O₂-exposed cells without RNSP (###p < 0.001). (c) The phosphorylation of pERK1/2 in H₂O₂-exposed cells with or without RNSP (60 μg/mL, 2 h). (d) The quantitative analyses of the immunoblotting in (c). Each column and bar represent the mean ± SEM (n = 4 each). Asterisks indicate a statistically significant difference from the value in untreated cells (***p < 0.001). Pound signs indicate a statistically significant difference from the value in the H₂O₂-exposed cells without RNSP (###p < 0.001).

is known that oxidative stress resulting from the overproduction of ROS causes damage to the cellular components, including DNA, resulting in subsequent cell death [5]. H₂O₂ is one of the most important ROS generated through oxidative stress [29, 30]. In the present study, the mean fluorescent intensity of 8-oxo-dG, a biomarker for oxidative stress-damaged DNA [41], was found to be significantly increased, mainly in the nuclei of SH-SY5Y cells from 4 h after exposure to H₂O₂. However, the expression of MitoSOX Red probe, a marker for mitochondria-derived ROS generation in the SH-SY5Y cells, increased from 1 h after exposure to H₂O₂, thus indicating mitochondrial damage to be the early origin of ROS generation before the nuclear DNA damage that is caused by oxidative stress. It is noted that both the long (5 days) and the short (2 h) pretreatments with RNSP could significantly inhibit the H₂O₂-induced mitochondrial ROS generation and nuclear DNA damage (Figures 3(a) and 3(b)) and the subsequent cell death in SH-SY5Y cells (Figures 1 and

2). This is the first evidence support the attenuation of stress-induced DNA damage in neuronal cells by RNSP.

Recent research has shown that REST is related to the reduction of oxidative damage. This was demonstrated by increased levels of ROS as well as oxidative DNA damage in response to REST knockdown. The overexpression of REST (up to 20-fold) can reduce H₂O₂-induced neuronal cell death [40]. In the present study, the expression of REST was significantly decreased as little as 1 h after cells were exposed to H₂O₂ (100 μM), and the time-dependent decrease of REST expression was positively correlated with subsequent DNA damage. These results agree with the hypothesis that REST knockdown increases oxidative DNA damage [40]. Surprisingly, pretreatment with RNSP for 2 h significantly reversed the H₂O₂-induced decrease of REST expression from 1 h (Figure 3(a)), inducing a parallel protective effect against subsequent oxidative DNA damage in SH-SY5Y cells (Figures 3(a) and 3(b)). Since REST is a protective factor

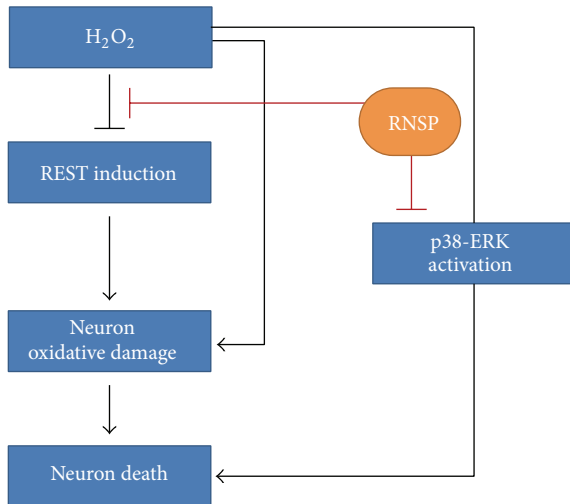


FIGURE 5: A schematic representation of the effects and the principle molecular mechanisms of RNSP on the protection from oxidative stress-induced neuronal death via the antioxidant properties and inhibition of MAPK activation.

in the ageing brain, which is in a state of decline from the early stages of AD [40], and because we have recently found that RNSP taken orally for 16 weeks can significantly improve cognitive function in the early stages of AD [18], the regulation of the H_2O_2 -mediated REST/oxidative stress may be one of the principle molecular mechanisms behind the protective effects of RNSP that are observed in neuronal cells.

Oxidative stress is known to be one of the major stimulators of the MAPK cascade. The overproduction of ROS during oxidative stress acts as a second messenger in signal transduction cascades leading to MAPK activation, because the intracellular redox state of the cells regulates the cellular signaling pathways [10, 11]. In the present study, p38 and ERK 1/2 MAPKs were significantly activated in SH-SY5Y cells after exposure to H_2O_2 (100 μM). This agrees with the hypothesis that ERK and p38 proteins are rapidly activated by H_2O_2 in PC12 cells [41, 42]. Pretreatment with RNSP for 2 h significantly reduced the H_2O_2 -induced phosphorylation of both p38 and ERK1/2, thus indicating that the reduction of the oxidative stress-mediated activation of p38 and ERK 1/2 may be another important molecular mechanism underlying the protective effects of RNSP on neuronal cells, because both p38 and ERK 1/2 MAPK are cell death factors which are mediated by oxidative stress [12, 13].

RNSP is composed of more than seventy components, including saffron and *Glycyrrhiza uralensis* [16, 17], and the beneficial effects of RNSP components have been paid attention [25]. Previous studies have shown that saffron, one of the components of RNSP, inhibits the $TNF\alpha$ -induced cell death of PC12 cells [22] and protects neurons from 6-hydroxydopamine hydrobromide-induced neurotoxic activity [23], through antioxidant properties of saffron [24–26]. Our previous studies have found improvements of the cognitive functions in mild-to-moderate AD patients living at high

altitude and increases in learning and memory in a mouse model of AD (Tg2576) [17–19]; the present observations have demonstrated the direct roles of RNSP in inhibiting oxidative stress-induced DNA damage and in the activation of MAPKs in neuronal cells, further confirming the benefits of RNSP in the prevention and clinical management of AD [7, 15]. Since oxidative stress is a potential contributor to the pathogenesis of AD [4, 7, 8] and since the levels of phosphorylated MAPKs are increased in the postmortem brains of AD patients [14–16], the antioxidant effects of RNSP can be expected to delay the development of AD.

5. Conclusion

RNSP was able to protect oxidative stress-induced neuronal damage via antioxidant properties and the inhibition of MAPK activation (schematic represented in Figure 5). These findings therefore provide the principle molecular mechanisms of the clinical effects of RNSP and support its use as therapeutic agent for the prevention and clinical management of AD.

Conflict of Interests

The authors declare they have no conflict of interests.

Authors' Contribution

Aiqin Zhu and Zhou Wu contributed equally to this work.

Acknowledgments

This work was supported partly by Key Grant for Returnees on Science and Technology from the Ministry of Personnel, China, to Aiqin Zhu (Ministry of Personnel 2014-414) and Grants-in-Aid for Scientific Research to Zhou Wu (24592802).

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