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On the need to consider wood formation processes in global vegetation models and a suggested approach

Key message: Dynamic global vegetation models are key tools for interpreting and forecasting the responses of terrestrial ecosystems to climatic variation and other drivers. They estimate plant growth as the outcome of the supply of carbon through photosynthesis. However, growth is itself under direct control, and not simply controlled by the amount of available carbon. Therefore predictions by current photosynthesis-driven models of large increases in future vegetation biomass due to increasing concentrations of atmospheric CO₂ may be significant over-estimations. We describe how current understanding of wood formation can be used to reformulate global vegetation models, with potentially major implications for their behaviour.

Keywords: dynamic-global-vegetation-model; xylogenesis; carbon; source; sink

Global vegetation models: from a source dominated to a balanced source-sink approach

The last couple of decades have seen the emergence of so-called earth system models (ESMs) for forecasting global climate responses to emissions of anthropogenic greenhouse gases (Hajima et al., 2014). ESMs are built around a general circulation model of the atmosphere, coupled to representations of ocean and land components, including the exchanges of carbon between these different reservoirs. Land surface carbon exchange with the atmosphere and structural dynamics are simulated using a sub-model of vegetation and soil processes, usually described as a “Dynamic Global Vegetation Model” (DGVM). These sub-models are dynamic in the sense that they simulate changes in vegetation distribution and structure over time from underlying physiological and ecological principles (e.g., Friend and White, 2000; Sitch et al., 2003), as opposed to a static Global Vegetation Model in which vegetation distribution (and sometimes structure) is prescribed, for example from remote sensing of the land surface (e.g., Sellers et al., 1996; Wang et al., 2010).

DGVMs have been used extensively for the simulation of historical and future land-atmosphere carbon fluxes in order to attribute (e.g., Keenan and Williams, 2018; Le Quéré et al., 2018) and predict (e.g., Cramer et al., 2001; Friend et al., 2014) terrestrial biosphere responses to, primarily, climate, CO₂, and land use changes. A common feature of these simulations is that while historical dynamics are largely consistent with atmospheric CO₂ constraints (e.g., Le Quéré et al., 2018), future predictions diverge quite markedly (e.g., Arora et al., 2013; Jones et al., 2013; Friedlingstein et al., 2014), with differences in the responses of vegetation carbon fluxes playing a major role. Moreover, this uncertainty has remained “stubbornly consistent” (Quetin and Swann, 2018), despite major efforts to expand the biological and physical process representations in these models (Lovenduski and Bonan, 2017). DGVMs largely attribute the historical net carbon sink on land to the CO₂ fertilization of plant growth through enhanced photosynthesis, and most predict this effect to become stronger in coming decades, resulting in a large terrestrial carbon sink, especially in forests, but with substantial variation between models (e.g., Friend et al., 2014).

55 **Figure 1. (A)** Schematic representation of carbon flows and controls in a generic DGVM. Photosynthesis
56 is under strong environmental control, resulting in gross uptake of carbon (GPP: gross primary productivity),
57 from which respiration is subtracted to give net primary productivity (NPP). This NPP is then partitioned
58 to various sinks, with relative proportions determined by allometric coefficients (e.g. fixed or based on goal-
59 seeking/optimisation assumptions), or based on passive filling in the case of a reserve pool. Here we indicate
60 that the prime purpose of the reserve pool is to replenish the foliage following complete leaf loss such as
61 during winter in a cold deciduous tree, as for example in the ORCHIDEE model (Krinner et al. (2005)).
62 Turnover of structural sinks is incorporated into soil organic matter, which decays back to atmospheric CO₂.
63 The positive feedback from the leaf sink to photosynthesis is due to the dependency of radiation interception
64 on leaf area. **(B)** Schematic representation of a proposed growth- and source/sink feedback-enabled DGVM.
65 A labile carbon pool of sugars receives carbon from photosynthesis and, potentially, storage reserves, and
66 loses it to respiration and flows to various sinks. The sink strengths are explicitly modelled, and therefore the
67 flows to them (and their growth) are the outcomes of their activities, rather than the rate of photosynthesis.
68 The activities of the sinks are under their own environmental and internal controls, including signalling effects
69 from the size of the labile pool itself (orange arrows). The labile pool also affects photosynthetic capacity
70 through negative feedback. The dynamics of the labile pool thereby ensure coordination between growth and
71 photosynthesis.

72
73 An overview of how carbon fluxes are represented in current DGVMs is shown in Figure 1(A). It is ev-
74 ident from the relationships in this structure that variability in the input of carbon (“Atmospheric CO₂”)
75 through photosynthesis (“GPP”) has a dominant influence on overall dynamics, especially of the plant and
76 soil carbon reservoirs. Furthermore, plant growth (“NPP”) is determined directly from the difference between
77 photosynthesis and plant respiration, with no explicit representation of growth processes themselves. This
78 implicit approach to growth has not been perceived as a problem for a variety of reasons, the main one being
79 the belief that plant productivity is only, or mainly, limited by the input of carbon through photosynthesis,
80 i.e. growth is C source-limited (e.g. eqn 3 of Lloyd and Farquhar (1996)). However, evidence that growth
81 processes have greater environmental sensitivities than photosynthesis, and even control photosynthesis un-
82 der many conditions through internal feedback, has led to calls for a re-evaluation of this C-source driven
83 production paradigm, and for it to be replaced with one in which the demand (i.e. “sink”) for carbon plays
84 at least as important a role as its supply (e.g., Millard et al., 2007; Fatichi et al., 2014; Körner, 2015; Fatichi
85 et al., 2018; Zuidema et al., 2018). These arguments are supported by the failure of the large stimulation
86 of photosynthesis by elevated CO₂ in experimental manipulations to be translated into equivalent growth
87 responses (e.g., Kirschbaum, 2011; Woodward, 2002; Dawes et al., 2015; Ellsworth et al., 2017), and evidence
88 that direct environmental constraints on growth, such as drought, low temperatures, may be stronger than
89 those on photosynthesis (e.g., Hsiao, 1973; Parent et al., 2010; Muller et al., 2011). Indeed, observations
90 of non-structural carbohydrate (NSC) accumulations under stress (Hoch, 2015; Hartmann et al., 2018), and
91 feedback inhibition of photosynthesis when growth is limited (e.g., Paul and Foyer, 2001), indicate the po-
92 tential for sink strength to limit overall carbon assimilation into durable biomass. We therefore suggest that
93 the arguments for an important role for sink processes are persuasive enough that it would be worthwhile to
94 incorporate them into a DGVM framework in order to explore their implications for ecosystem carbon fluxes
95 and community dynamics.

96
97 Attempts to incorporate sink-limited growth into DGVMs have so far been limited (Fatichi et al., 2018),
98 probably because of the lack of a perceived need and/or no clear approach to how it might be achieved,
99 particularly within current model structures. Nevertheless, some efforts have been undertaken, such as mod-
100 ifications to existing allocation routines (e.g., Guillemot et al., 2017), or highly empirical approaches (e.g.,
101 Leuzinger et al., 2013). In the latter, for example, an empirical model of the temperature impact on annual
102 NPP was implemented based on growing-degree days within the Lund-Potsdam-Jena (LPJ) DGVM
103 (Sitch et al., 2003). It was found that the addition of this constraint greatly reduced predicted biomass
104 under low-temperature limited situations, especially at high latitudes and altitudes, compared to when only
105 photosynthesis limits growth. Other relevant developments include the incorporation of nutrient cycling in
106 DGVMs, with the potential to constrain sink strength. For example, the HYBRID4.1 (Friend and White,
107 2000) and O-CN models (Zaehle and Friend, 2010) simulate the dynamics of a labile plant N pool, which
108 then constrains tissue growth through stoichiometric limits (Friend et al., 1997). Therefore available N has

109 the potential to limit growth, with major impacts on predicted responses to increasing atmospheric CO₂
110 (Cramer et al., 2001; Zaehle et al., 2010). However, these developments do not explicitly consider growth
111 processes, with growth still the outcome of carbon balance.

112

113 While not DGVMs, Functional-Structural Plant Models treat sink activity explicitly. The L-PEACH
114 model of Allen et al. (2005), for example, includes a number of features directly relevant to modelling
115 source-sink dynamics. Photosynthesis is inhibited by the accumulation of carbohydrate in the leaf due to
116 inadequate sink strength, the flow of carbohydrates around the plant is based on concentration gradients
117 across resistances (Thornley, 1972), storage compartments are included as sinks and sources, and the uptake
118 of carbohydrates by sinks is a function of the local sugar concentration in the phloem and the degree of water
119 stress. However, these models focus on the simulation of plant form (e.g. branching structure), and have
120 not been used to address the significance of sink-limited growth *per se*, as far as we know, and their detailed
121 consideration of small stem segments makes them unsuitable for global modelling. We therefore conclude
122 that there is a need for a new process-based methodology simulating plant growth within DGVMs, with a
123 core element being the explicit treatment of sink processes and their controls.

124

125 **A way forward: xylogenesis**

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127 The need to better understand climate-growth relationships in dendroecology has led to the development
128 of xylogenesis models. As these explicitly consider wood growth, they are promising frameworks for incor-
129 porating sink activity in DGVMs. Perhaps the best known is the Vaganov-Shashkin (VS) simulation model
130 (Vaganov et al., 2006), which aims to go beyond previous statistical frameworks (e.g., Fritts et al., 1971) in
131 providing a mechanistic tool for addressing questions such as details of climatic controls on the formation of
132 conifer tree rings in dry, cold, and temperate regions. The dynamics of cambial cell production, and subse-
133 quent cell enlargement, wall thickening, and eventual death of xylem cells are treated, with dependencies on
134 temperature and soil water content. Xylogenesis models such as these point the way to how sink activity in
135 trees can be modelled as they explicitly consider the dynamics of volume and mass increases at the cellular
136 level, and how they are driven by environmental factors. However, the VS model is not mechanistically
137 tied to the whole tree, and so does not provide a complete framework for inclusion of growth in a DGVM.
138 Moreover, these models have not yet been validated at the scale of the processes they are representing, for
139 example using wood formation monitoring data.

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141 Fatichi et al. (2014) recommended a series of processes that should be implemented in DGVMs in order
142 for them to realistically simulate growth, or at least “C allocation”. However, their list includes components
143 that would be very challenging to implement efficiently in a global model. Moreover, they do not explicitly
144 discuss wood growth, whereas this is the process whereby the majority of carbon is sequestered into long-lived
145 plant material, and so needs to be central to any DGVM representation of growth processes.

146

147 Here we outline a possible approach for building a balanced source-sink treatment of growth in DGVMs,
148 based on a representation of xylogenesis, scaled to the whole tree in terms of total carbon balance, with links
149 to tree size and shape. While we focus on the process of wood formation as this represents the dominant
150 carbon sink on land, much of the approach is relevant to all higher plant life forms.

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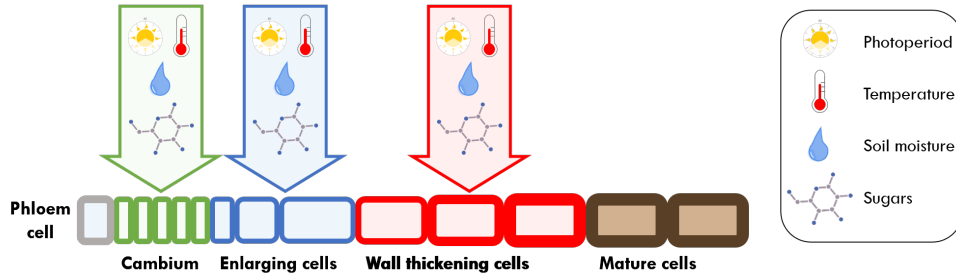
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163 **Figure 2.** A radial file of developing tracheids, including dividing cells in the cambium, and enlarging,
 164 wall thickening, and mature cells in the forming xylem. By including each developmental phase, the model
 165 can be compared in detail with observations. External and internal factors are assumed to act directly on
 166 the differentiating cells, influencing the rates and/or the duration of cell differentiation.

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 168 We do not advocate a completely sink- or source-driven approach, but one in which these two controls
 169 are coupled. Plants grow as integrated wholes, and must achieve a balance between carbon uptake, storage,
 170 and growth (Smith and Stitt, 2007). A model of coupled source- and sink-controls on tree growth therefore
 171 needs to include both an explicit treatment of wood formation and a link between growth and photosynthesis.
 172 Storage pools are required to buffer supply and demand on different timescales, and growth processes need to
 173 be simulated on a sufficiently short time step to allow for the influence of climatic variability. The whole-plant
 174 carbon balance also requires treatments of respiration and non-wood sinks such as reproduction. A proposal
 175 for such a scheme is presented in Figure 1(B). Whether this scheme is applied at the individual or stand scale
 176 will depend on the overall structure of the model, but an individual-scale implementation allows resolution of
 177 those features that are size- (e.g., Bennett et al., 2015), age- (e.g., Hayat et al., 2017), and shape-dependent.
 178 As a tree grows, the relative proportions of its different components can change, affecting the mass, volume,
 179 and area ratios between source and sink tissues, with important consequences for controls on wood formation
 180 and growth (Stephenson et al., 2014; Hayat et al., 2017; Hartmann et al., 2018), making an individual-based
 181 approach desirable. Such an approach also allows for the treatment of successional dynamics and competition
 182 (e.g., Friend et al., 1997).

183
 184 Two key features of this scheme will require innovations that go beyond current global dynamic vegeta-
 185 tion models: (1) explicit wood formation; and (2) associated dependencies and feedback between sink- and
 186 source-activities in order to achieve coordinated uptake and growth. Below we outline a possible approach to
 187 achieve this, discuss how such changes might alter model behaviour, and suggest how future research might
 188 fill critical remaining knowledge gaps.

189 **Developing an explicit wood growth model for incorporation into DGVMs**

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 192 A mechanistic approach to the incorporation of explicit internal carbon sink processes into a DGVM
 193 will need to treat the influences of environmental factors, such as temperature and the supply of water and
 194 nutrients, on xylem formation. Carbon supply through photosynthesis and/or from reserves will also affect
 195 xylem formation, either directly as a substrate for growth and associated energetic requirements, and/or, as
 196 evidence suggests, indirectly as a signal metabolite (Smith and Stitt, 2007).

197
 198 As mentioned, wood formation consists of the differentiation of secondary xylem and associated cell de-
 199 velopmental phases, often described as “xylogenesis”, during which xylem precursors divide, enlarge, undergo
 200 secondary wall thickening, and finally succumb to programmed cell death (see Figure 2). Modelling this pro-
 201 cess mechanistically is challenging due to complex interactions between environmental conditions and internal
 202 signalling pathways linked to physiological and developmental factors. Moreover, periods of volume growth
 203 and mass increment occur at different times and in different cell developmental phases (Cuny et al., 2015).
 204 Despite this complexity, we believe that there is now sufficient knowledge concerning the basic processes of
 205 xylem differentiation and how they respond to environmental and developmental factors, at least in conifers

206 (Rathgeber et al., 2016), to build a model suitable for testing hypotheses and as a basis for a treatment of
207 whole-tree growth for insertion into a DGVM.

208
209 During the growing season, the vascular cambium produces, at most, one new xylem cell per radial file
210 per day, which then takes 1-2 months to mature (Rathgeber et al., 2016). This suggests that a suitable
211 approach would be to simulate division and differentiation of the cells in a radial file on a daily timestep.
212 Each cell would be classed as either cambial (i.e. a mother xylem cell still capable of division), enlarging
213 (i.e. a primary cell-walled differentiating xylem cell no longer capable of division), wall thickening (i.e. a
214 secondary-cell-walled maturing xylem cell), or mature (i.e. a dead but fully functional xylem cell). Transi-
215 tions between these cell developmental phases can be computed on a daily basis, as can be the amount of
216 enlargement and cell wall deposition for the appropriate cell developmental phases. Xylogenesis consumes
217 carbon through respiration and the deposition of wall materials, thus providing a major sink for internal
218 carbohydrates. Xylogenesis not only determines the amount of carbon sequestered, but also shapes the mor-
219 phology (i.e. lumen size and wall thickness) of the xylem cells. This morphology in turn determines the
220 tree’s water transport capacity (which limits foliage area and stomatal conductance) and safety (which can
221 determine mortality), and also its mechanical properties (which affect height growth and branching pattern).

222
223 While the sequence of secondary xylem differentiation in conifers is well known, details of the controls
224 on the rates and timings of division, enlargement, cell wall thickening, and eventual death are not yet fully
225 understood (Hartmann et al., 2017). Factors believed to be important for rates include cambial tempera-
226 ture, xylem cell turgor, and internal signals (e.g. auxins, peptide ligands, sucrose). Durations within cell
227 development phases are related to cell age, cell size, and internal signals (both positional and related to
228 season and environmental conditions). External signals such as wind speed can also affect tree growth; while
229 the mechanisms are not well understood and have not been incorporated into growth models of the type
230 discussed here, recent work suggests an approach for doing so in relation to both primary and secondary
231 growth (Mouliat et al., 2015).

232
233 Despite our lack of knowledge concerning many of these controls, understanding has recently greatly ben-
234 efit from measurements of the seasonal dynamics of wood formation using microcores, extracted at weekly
235 intervals during the growing season (e.g., Cuny et al., 2014; Balducci et al., 2016; Cuny and Rathgeber, 2016;
236 Cuny et al., 2018). To date these measurements have mainly been made on conifers with their relatively
237 simple wood anatomy. Our modelling approach is therefore initially focused on these species. However, we
238 aim to model angiosperm species as well, and indeed extend it to non-woody plants.

239
240 A dynamic model of cellular differentiation along a single radial file can be separated into two conceptual
241 components. The first is a spatially-explicit representation of a linear radial file of cells with particular iden-
242 tities, which determine their potentials for division, enlargement, thickening, and death, and the sequential
243 dynamic transitions between those identities. This component is an unchangeable computational framework
244 as it mirrors how xylogenesis actually occurs (although increased complexity will be necessary to treat woody
245 angiosperm anatomies). The second component is a representation of the controls on these rates and transi-
246 tions (including onset and cessation), which are in many cases uncertain and must therefore be included as
247 hypotheses to be tested through consistency with observations.

248 **Key data sources**

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251 Two important types of observational data are available with which to test hypotheses and determine
252 parameter values, kinetic and anatomical. The recent development of a tracheid differentiation kinetics ap-
253 proach is providing data and results that can be directly used to test mechanistic wood formation model
254 hypotheses and calibrate parameter values (Cuny et al., 2013). These kinetic data quantify the temporal
255 course of cellular dynamics, such as the number of cells per developmental zone (e.g. the cell enlargement
256 zone), the characteristics of those cells (e.g. size and cell wall thicknesses), and the timings of their transi-
257 tions (e.g. the beginning and end dates for cells in a given phase during the growing season). This approach
258 is based on the statistical analysis of wood formation monitoring data, and is associated with quantitative
259 wood anatomy data. Wood formation monitoring data consist of weekly counts of cell number in the four

260 differentiation zones (i.e., cambium, enlarging, wall thickening, and mature). From these data, the duration
261 spent by each forming cell in each differentiation zone can be estimated (Wodzicki, 1971; Rossi et al., 2006;
262 Cuny et al., 2013).

263

264 In contrast, quantitative anatomical data concern the cells within the tree ring at the end of the growing
265 season, and provide measurements of tracheid dimensions (e.g. cell wall thicknesses and lumen diameters).
266 Knowing the time a cell spent in each differentiation stage from kinetic data, and the result in terms of
267 morphology from end-of-season anatomy, it is possible to compute the rate of change for the different pro-
268 cesses (see Figure S2 of Cuny et al. (2018) for a graphical explanation). For example, an earlywood cell that
269 spent 15 days in enlargement for a final cell lumen diameter of 30 μm , enlarged at a rate of 2 $\mu\text{m}/\text{day}$. The
270 rates of the cell differentiation processes, and the cell differentiation phase durations, will both be represented
271 in the mechanistic wood formation model and so can be used for its parameterisation as well as its validation.

272

273 We also suggest that once a basic model has been developed using tracheid differentiation kinetics data,
274 the model could be tested and further developed at larger temporal and spatial scales using new data sets
275 from quantitative wood anatomy which are being produced (e.g., Castagneri et al., 2017; Ziaco et al., 2016).
276 Tree-ring microdensitometric profiles are an additional source of data for the relationship between wood
277 anatomy and wood density (Cuny et al., 2014).

278

279 Finally, while wood formation studies to date span a relatively limited amount of time (e.g. up to 13 yr in
280 Rossi et al. (2016); up to 7 yr in Cuny et al. (2018)), anatomical data can be obtained after wood formation
281 has ceased and so can be extended over many years, or even centuries, into the past. This increases the
282 range of responses to environmental conditions that can be analysed, including to climatic variability, CO_2
283 concentration, and successional stage. For example, an 87 yr cellular anatomy dataset was used to study
284 climatic controls on tracheid development in *Picea abies* along an altitudinal gradient (Castagneri et al.,
285 2017), and Fonti et al. (2013) investigated the relationship between temperature and tracheid anatomy using
286 a 312 yr tree-ring chronology from a southern Siberian larch forest. These types data are potentially very
287 useful additional sources of information for the xylogenesis model envisaged here, and, moreover, the model
288 could be used to interpret these data types mechanistically.

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290 **Scaling to the whole-tree**

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292 While it is relatively easy to code controls due to fundamental environmental factors such as temperature
293 and soil water (e.g. through the rate of cell cycling for temperature and rate of cell expansion for local turgor),
294 a representation of signalling pathways is harder to devise. A sensible approach is therefore to implement
295 these as simply as possible, such that they are adequate to explain first-order observed responses, for example
296 using simple spatial gradients in growth substances (e.g., Hartmann et al., 2017). A key dependency that
297 needs to be represented is the effect of the concentration of carbohydrate substrate: is this to be treated only
298 as a substrate pool for cell wall thickening (e.g. with Michaelis-Menten kinetics), or does it also, or mainly,
299 affect wood growth through a signalling pathway which controls sink activity (e.g. cambial division)?

300

301 Smith and Stitt (2007) provide evidence that growth is directly regulated by carbohydrate supply in order
302 to avoid carbon starvation. This is achieved through an effect of carbon availability on the synthesis of pro-
303 teins responsible for growth processes, controlling both cell proliferation (through controls on the cell cycle)
304 and cell wall synthesis. This suggests that a model of xylogenesis should include direct regulation of growth
305 activity by carbon supply (and potentially other growth regulators such as phytohormones) as a signalling
306 pathway, thus providing a mechanism for coordination between supply and demand, rather than regulation
307 only through a substrate-limited growth response. As Smith and Stitt (2007) suggest, this is consistent with
308 a regulatory framework in which resources are conserved when carbon availability is limited. They provide
309 evidence for rapid “acute” and acclimatory regulatory responses. Interestingly, genes involved in the cell cycle
310 have transcript levels that decrease during the night, suggesting that diurnal cycles in growth, and potentially
311 phenological responses on seasonal timescales, provide additional constraints that need to be considered. The
312 extent to which these processes occur in trees is yet to be determined, but studies such as Etchells et al.
313 (2015) indicate a strong regulatory control of wood growth.

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The regulation of source activity (i.e. photosynthetic rate) by sink demand in plants is well-known (Paul and Foyer, 2001), although the exact mechanism is unclear (Yan et al., 2013). There is good evidence for its occurrence in trees (e.g., Iglesias et al., 2002), and appears to be mediated by the accumulation of phloem sugars at the source sites in leaves (Franck et al., 2006; Ainsworth and Bush, 2011). Ainsworth and Bush (2011) describe how phloem loading plays a key role in balancing carbon source activity with sink utilisation in apoplastic loaders because loading can increase sugar concentrations against a gradient of 2 or 3 orders of magnitude. An increase in mesophyll sugar levels when sink strength is reduced must therefore be the result of down-regulation of phloem loaders in response to accumulation of phloem sugars, rather than as a purely physical effect. The resulting increases in mesophyll sugars can cause reductions in photosynthetic capacity (e.g., Krapp and Stitt, 1995; Paul and Foyer, 2001). A model approach can therefore be envisaged, applicable to both active and passive loaders, in which photosynthetic capacity is regulated in response to the concentrations of labile sugars, whether at the tissue or whole-plant level.

Taken together, there exists therefore a good theoretical basis for the development of a whole-plant regulatory framework linking sources and sinks mediated by the concentration of labile sugars, with sinks being driven by the production and differentiation of meristematic cells and sources by leaf photosynthesis.

As well as treating the activities of sink and source processes and the feedback between them, a model of whole-tree growth needs to scale the growth dynamics of the radial file, photosynthesis, and internal carbohydrate dynamics to the whole tree. This can be achieved by treating both the overall structure of the tree (canopy height, stem diameter, rooting depth, leaf area, crown area, etc.), as well as the internal dynamics of carbon, and potentially nutrients such as N and P as well as water. The coordinated nature of secondary cellular differentiation throughout the tree cambium means that the primary environmental influences on wood growth can be captured in a single radial file, making the problem of whole-tree growth dynamics eminently computable. Volume and mass growth of the stem can thus be treated as a function of the dynamics of a single radial file at some location on the stem, scaled to the whole tree using the summed stem, branches, and root surface areas to represent the entire secondary meristem. Apical meristems provide height and depth growth, and can use a similar approach as the secondary radial file to compute cellular differentiation. Other sinks, such as foliage and fine roots, can be included using approaches such as the pipe model. A parsimonious approach to scaling the radial file and an apical meristem to the whole tree structure with internal labile carbon dynamics was described and tested by Hayat et al. (2017), with the ratio of apical to secondary meristem activity controlled by leaf canopy shading. This approach used single undifferentiated volumes for the meristems, but could easily be extended to represent daily xylogenesis along a radial file.

Assumptions regarding scaling of physiological feedback to the whole tree level can be equally parsimonious as a first step. As discussed, these need to treat the influence of both C source supply on xylem differentiation and the feedback of growth on photosynthesis. The simplest mechanism to achieve this, and one consistent with physiological understanding (e.g., Smith and Stitt, 2007), is to include one or more dynamic C reserve pools, which buffer supply and demand, and provide information on internal carbon status to source and sink processes. High levels of C reserves cause down-regulation of photosynthesis and, potentially, up-regulation of growth, depending on phenological and/or other controls. Key uncertainties are the precise relationships to use, the controls on reserve dynamics (Dietze et al. (2014); Hartmann et al. (2018)), and how feedback might interact with any internal dynamics of N and P and external factors such as soil water and incident light. However, as for the details of controls on xylogenetic processes, feedback processes can be represented as a set of first-order hypotheses and tested against a range of data sources (e.g., Würth et al., 2005; Furze et al., 2018; Smith et al., 2018).

There is clearly a need for more information on relationships between carbon supply and wood growth. However, elevated/reduced CO₂ experiments are difficult and expensive, and responses of seedlings in growth chambers may not reflect those in large mature trees. But there are other ways to manipulate C supply to radial meristems. These include girdling, phloem compression, and phloem chilling (Rademacher et al., submitted). In experiments on mature trees at Harvard Forest in which we are manipulating the supply of photoassimilates to stem sinks through phloem chilling, phloem compression, and girdling in order to inves-

368 tigate the response of xylogenesis (through weekly microcores) and NSC dynamics to variable carbon supply.
369 The outcomes of these manipulations will be extremely informative for developing a model of whole-tree
370 source-sink dynamics as envisaged here.

371 372 **Implications for vegetation models**

373
374 Inclusion of explicit wood growth and feedback on photosynthesis in trees is likely to have profound influ-
375 ences on the behaviour of DGVMs, especially their forecasts of future carbon uptake into durable biomass.
376 The actual degree of influence will depend on the extent to which limitations to growth are stronger than
377 those on photosynthesis, and over what timescales. At a minimum it is likely that such an approach, if
378 carefully implemented, will help reconcile model-data biases such as those due to lag effects (Keenan et al.,
379 2012), which invoke temporal separations between source and sink processes (Seftigen et al., 2018; Rocha
380 et al., 2006; Teets et al., 2018; Richardson et al., 2013). An additional important aspect is the distinction
381 between volume and mass growth. Inventory-type assessments of tree productivity use volume changes to
382 estimate mass changes, usually assuming fixed, species-specific values for wood density, carbon content, and
383 allometric relationships. Model predictions of (carbon-based) NPP dynamics are then compared with these
384 volume-derived observations. However, wood density varies significantly within and between growth rings,
385 and between individuals of the same species, and moreover there is very good evidence that significant reduc-
386 tions in wood density have occurred at large scales in recent decades due to environmental changes (Pretzsch
387 et al., 2018). Furthermore, wood carbon content as a percentage of total dry mass is known to significantly
388 vary from the widely used 50% of dry mass (e.g., Martin et al., 2018). A model which includes explicit
389 xylogenesis, of the type envisaged here, will be capable of reproducing divergences between volume and mass
390 changes, and may thus have major implications for the match between model and data more widely (Babst
391 et al., 2018). Moreover, density changes have implications for hydrological functioning (including leaf area,
392 stomatal conductance, and risk of cavitation), and mechanical support (Cannell and Dewar, 1994), which
393 can be built into our model approach dynamically (i.e. environmental factors can influence wood structure,
394 which affects hydrodynamics and risk of breakage). In addition, explicit consideration of the stoichiometric
395 requirements of secondary cell walls in the model would allow for the variation in carbon content to be taken
396 into account.

397
398 However, we believe the implications of our approach will go beyond these issues, potentially challenging
399 the current interpretation of historical C sinks on land and having major impacts on future forecasts. For
400 example, the seeming inconsistencies between the top-down constraint of a large land C sink, and local obser-
401 vations of little, if any, increased plant growth, could be resolved by such an approach (Fatichi et al., 2018).
402 Moreover, if a mechanistic yet relatively simple model of tree growth can be shown to have a major effect
403 on DGVM behaviour, reconciling model-data biases and changing forecasts, then the role of photosynthesis
404 in these models is greatly reduced. DGVMs tend to treat photosynthesis and the canopy light environment
405 with great complexity, including vertical canopy gradients in physiology and radiation, and complex biochem-
406 ical functions with demanding computational numerics to balance the supply and demand of CO₂. This is
407 far more detail than envisaged in our approach to growth processes, and therefore a balanced treatment of
408 sources and sinks will make these models simpler as well as more realistic.

409 410 **Key remaining gaps in knowledge and future developments**

411
412 As mentioned, details of the relationship between the supply of photosynthate and the activity of differen-
413 tiating xylem is critical yet remains poorly understood. Even in *Arabidopsis*, while considerable knowledge
414 exists concerning the metabolic pathways involved in the conversion of sucrose to cell wall material, under-
415 standing the regulation of cell wall synthesis is still rudimentary (Verbančič et al., 2018). While synthesis
416 is inhibited at low rates of sugar supply, the range over which this occurs, and the controls when C supply
417 is saturating, remain unclear. More broadly, controls on cell wall synthesis include signalling pathways and
418 the regulation of genetic expression (Verbančič et al. (2018)), which are largely outside the scope of DGVMs.
419 However, observations in trees suggest influences from temperature and plant water content, which could
420 be implemented empirically to allow characterisation of C-sink strength at a cellular level (e.g., Antonova
421 and Stasova, 1997; Ziaco et al., 2014; Cuny and Rathgeber, 2016; Björklund et al., 2017). Both the rate of

422 synthesis and its duration are important, with the determination of the end of the maturation phase a key
423 factor. Interestingly, this usually occurs before environmental conditions deteriorate to levels that would be
424 expected to limit growth, implying strong phenological signalling. Implementing such controls mechanisti-
425 cally in a model of wood growth might be challenging, but is clearly of great importance, especially in the
426 context of decoupling growth from source activity. Other features of wood anatomy to consider, but not
427 discussed here, are the determination of cell types such as parenchyma and fibre cells, and the formation of
428 heartwood.

429
430 Xylogenetic studies have largely focused on conifers, with their relatively simple wood structure, and
431 therefore an important objective will be to extend this balanced source-sink model approach to other species.
432 Indeed, tropical trees contain the majority of plant carbon (Houghton et al., 2009), and so future work needs
433 to develop representations of wood development across a range of tree types and environmental conditions.
434 The responses of wood development to future conditions, particularly with respect to hydraulic properties,
435 will be a major determinant of the performance of individuals (cf., Pretzsch et al., 2018). Therefore, rep-
436 resenting differences in wood formation and resulting anatomical structures between tree types and species
437 (e.g., Spicer and Groover, 2010) will be key for predicting future community dynamics. The complexity of
438 angiosperm wood anatomy likely makes the level of detail possible in modelling conifer xylogenesis difficult
439 to achieve across all species. We are therefore exploring approaches that treat zones of cells types, rather
440 than individual cells, based on microcore observations of weekly growth dynamics in *Quercus rubra* and
441 *Acer rubrum* at Harvard Forest, in combination with phloem chilling and compression treatments.

442 443 **Conclusions**

444 We have identified a need to explore the implications of explicit representation of wood growth processes
445 in DGVMs and have outlined an approach for doing this. Our approach is to compute the daily growth
446 on an individual tree based on a treatment of xylogenesis along a radial file, with dependencies on external
447 and internal factors, and feedback on photosynthesis through labile carbon contents. Scaling to the whole
448 tree assumes this radial file represents all secondary growth, with tree size and shape determining the total
449 meristem mass through simple allometric relationships. A key uncertainty concerns the response of xylem
450 differentiation to carbon supply. Both wood formation dynamics and anatomical data from tree rings, es-
451 pecially where carbon supply is manipulated and labile concentrations measured, can be used to develop
452 and test hypotheses. We anticipate that when scaled up globally, a mechanistic representation of growth
453 processes has the potential to significantly alter our interpretation of the historical carbon sink on land and
454 greatly improve constraints on its likely future dynamics.

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