



Subscriber access provided by UQ Library

# Ecotoxicology and Human Environmental Health

# Experimental investigation and modelling of the transformation of illicit drugs in a pilot-scale sewer system

Jiaying Li, Jianfa Gao, Phong K. Thai, Adam Shypanski, Ludwika Nieradzik, Jochen F. Mueller, Zhiguo Yuan, and Guangming Jiang

Environ. Sci. Technol., Just Accepted Manuscript • DOI: 10.1021/acs.est.8b06169 • Publication Date (Web): 11 Mar 2019 Downloaded from http://pubs.acs.org on March 13, 2019

## Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.



Table of Contents

- <sup>1</sup> Experimental investigation and modelling of the
- <sup>2</sup> transformation of illicit drugs in a pilot-scale sewer
- 3 system
- 4 Jiaying Li<sup>a</sup>, Jianfa Gao<sup>b</sup>, Phong K. Thai<sup>b</sup>, Adam Shypanski<sup>a</sup>, Ludwika Nieradzik<sup>a</sup>, Jochen F.
- 5 Mueller<sup>b</sup>, Zhiguo Yuan<sup>a</sup>, Guangming Jiang<sup>a,c,\*</sup>
- <sup>6</sup> <sup>a</sup>Advanced Water Management Centre, The University of Queensland, St Lucia, QLD
- 7 4072, Australia
- 8 <sup>b</sup>Queensland Alliance for Environmental Health Sciences, The University of
- 9 Queensland, Brisbane, QLD 4102, Australia
- <sup>10</sup> <sup>c</sup>School of Civil, Mining and Environmental Engineering, University of Wollongong,
- 11 Wollongong, NSW 2522, Australia
- 12
- 13 KEYWORDS
- 14 Wastewater-based epidemiology, illicit drugs, biotransformation, pilot sewer system,
- 15 model validation
- 16 ABSTRACT

In-sewer stability of illicit drug biomarkers has been evaluated by several reactor-17 based studies but less has been done in sewer pipes. Experiments conducted in sewer 18 pipes have advantages over lab-scale reactors in providing more realistic biomarker 19 stability due to the flow and biological dynamics. This study assessed the 20 transportation and transformation of seven illicit drug biomarker compounds in a 21 22 pilot-scale rising main and a gravity sewer pipe. Biomarkers presented diverse stability patterns in the pilot sewers, based on which a drug transformation model was 23 calibrated. This model was subsequently validated using transformation datasets from 24 literature, aiming to demonstrate the predictability of the pilot-based transformation 25 coefficients under varying sewer conditions. Furthermore, transformation coefficients for 26 five investigated biomarkers were generated from four studies and their prediction 27 capabilities under the pilot sewer conditions were jointly assessed using performance 28 29 statistics. The transformation model was successful in simulating the in-sewer stability for most illicit drugs. However, further study is required to delineate the sources and 30 pathways for those compounds with potential formations to be simulated in the 31 transformation model. Overall, the transformation model calibrated using the pilot-32 sewer data is a credible tool for the application of wastewater-based epidemiology. 33

#### 34 INTRODUCTION

Wastewater-based epidemiology (WBE) has been developed rapidly over the last decade in a bid to achieve objective and timely assessment of community health and consumption behaviours via analysing trace levels of substances (termed as

Page 4 of 39

biomarkers) in wastewater, including illicit drugs, pharmaceuticals and new 38 psychoactive substances.<sup>1</sup> Back-estimating the catchment-wide usage of illicit drugs 39 is an emerging area of WBE, which is demonstrated to be a useful complementary 40 tool to conventional drug monitoring approaches.<sup>2-5</sup> In order to improve the 41 accuracy of back-estimation, researches have been widely conducted to address the 42 uncertainties associated with sampling method and chemical analysis,<sup>5-9</sup> while a 43 comprehensive understanding of biomarkers stability in real sewers is still ongoing.<sup>10-</sup> 44 <sup>17</sup> Biomarkers are subjected to physiochemical and biological processes during their 45 transport in rising main and gravity sewer pipes, where the hydraulic retention time 46 (HRT) may last for hours.<sup>18</sup> Neglecting the biomarkers transformation (e.g., the 47 degradation or formation in sewers) will lead to an under- or over-estimation of drug 48 consumption in a catchment.<sup>4, 10, 12, 13, 15, 19</sup> This uncertainty varies depending on the 49 stability of biomarkers and the characteristics of sewer systems such as HRT 50 distributions, which was suggested to be negligible for the stable biomarkers<sup>8</sup> but 51 significantly increased for the unstable biomarkers with >40% median mass losses in 52 the catchments.<sup>15</sup> 53

Depending on the different experimental scales and conditions utilized, research on biomarker stability can be divided into four categories: 1) *in-water study* using clean/sterile bottles or other containers where abiotic processes such as chemical hydrolysis occur in water;<sup>14, 20, 21</sup> 2) *in-wastewater study* that is conducted in raw wastewater where suspended biomass and certain microbial activities contribute to the transformation process;<sup>12, 14, 21-25</sup> 3) *sewer reactor study* employing lab reactors
with intact/suspended biofilms or activated sludge to mimic the biologically active
sewer environments;<sup>10-12, 14, 16, 19</sup> 4) *real sewer pipe study* using the sewer pipes with
the same (or similar) size and operational conditions as the real sewer networks.<sup>10, 15, 17, 26, 27</sup>

Among the *real sewer pipe studies*, two of them assessed the change of biomarkers 64 using 24-h composite samples, however, with limited understanding of the 65 concurrent in-pipe hydraulics or biological activities.<sup>15, 26</sup> Two other studies spiked 66 biomarkers in a real rising main pipe and evaluated their variations from the pipe 67 upstream to a downstream sampling point, coupled with the measurements of flow 68 dynamics and biological activities in the pipe. However, due to the poor accessibility 69 of real sewer pipes and the narrow HRT windows, the obtained data points were 70 insufficient for kinetics evaluation.<sup>10, 27</sup> In comparison to the inherent limitations of 71 static lab-reactors and complex real sewers<sup>28</sup>, a pilot-scale sewer can be more 72 73 beneficial for the study of biomarker stability in sewer pipes by providing multiple sampling points, online monitoring, controllable flow, and other environmental 74 factors. This has been demonstrated in a recent *real sewer pipe study* investigating 75 the fate of pharmaceutical biomarkers.<sup>17</sup> 76

The temporal transformation data obtained in pilot-scale sewers can be very valuable
for the modelling of biomarker stability. Transformation modelling is a useful
approach with which to utilize information on biomarker stability in the application

of WBE in reality. So far, three sewer reactor studies investigated the transformation 80 modelling of illicit drugs,<sup>10, 14, 16</sup> but only one study validated the estimated 81 transformation coefficients in a real rising main pipe.<sup>10</sup> It is thus imperative to 82 calibrate the illicit drug transformation model using dynamic data from sewer pipes 83 (e.g. pilot-scale sewers) instead of lab reactors. More importantly, the transformation 84 coefficients derived from different studies need to be systematically evaluated for 85 their transferability across diverse sewer conditions. 86 The first objective of this study is to measure and model the stability of illicit drug 87 biomarkers in the pilot-scale sewer pipes. Experiments were conducted in a pilot-88 scale sewer system, including a rising main pipe and a gravity sewer pipe with online 89 monitoring and control system. A drug transformation model was calibrated with the 90 pilot-sewer data and was subsequently validated using literature data under varying 91 92 sewer conditions. The second objective is to systematically evaluate the transformation coefficients generated by previous studies, through the comparison 93 94 of the prediction capabilities using performance statistics. Collectively speaking, this work intends to advance WBE through not only providing valuable data on illicit drug 95 transformation in sewer pipes, but also enhancing the generalizability and 96 applicability of the transformation model to the application of WBE. 97

# 98 MATERIALS AND METHODS

## 99 **Compounds for Investigation**

100	This study investigated the parent compounds of major illicit drug biomarkers by
101	spiking them into the pilot-scale sewer pipes, including cocaine (COC), ketamine
102	(KET), 3,4-methylenedioxymethamphetamine (MDMA), morphine (MOR), and
103	methadone (MTD). According to the analysis results of wastewater samples at the
104	experiment site, the spiking concentrations (2.5-8 ppb) for most biomarkers were
105	higher than their native residues in raw wastewater (<0.5 ppb). The native
106	methamphetamine (METH) was investigated without being spiked because of its
107	relatively high background concentrations (around 1 ppb). The native
108	benzoylecgonine (BE) was evaluated as a specific metabolite of COC without being
109	spiked. Other metabolites of the parent compounds were not spiked in separate
110	tests. The flow tracer rhodamine was added into the spiking mixture solution.
111	Rhodamine signal was measured by a portable Cyclops <sup>®</sup> -7 Submersible Rhodamine
112	Sensor coupled with a Cyclops <sup>®</sup> Explorer.

# 113 **Tests in the pilot-scale sewer system**

Experiments were conducted in a pilot sewer system located at the Luggage Point Wastewater Treatment Plant, Queensland, Australia. This study employed two 300-m long sewer pipes, one rising main and one gravity pipe, on a controllable platform equipped with programmable logic controller, pumps, meters, sensors, etc.<sup>17, 29</sup> The experimental procedures in this study were the same as reported previously.<sup>17</sup> The layout and parameters of the sewer system are illustrated in Supplementary Information S1.1.

The **Rising main pipe** is completely filled with wastewater and anaerobic biofilm 121 were cultivates on the pipe's inner surface with a thickness of  $1 \sim 2 \text{ mm.}^{29}$  The internal 122 diameter is 100 mm, leading to a biofilm-area-to-wastewater-volume (A/V) ratio of 123 40 m<sup>-1</sup>. This rising main pipe is constructed to spiral up from the ground layer (inlet) 124 to the top layer (outlet) and wastewater is driven by pumping events in a plug-flow 125 126 regime. As shown in Figure S1.1, eight sampling ports are distributed along the sewer pipe, i.e. at 0, 15, 45, 75, 105, 135, 195, and 240 m from the inlet for port #1 to 127 #8, respectively. 128

Triplicate batch tests were carried out over 3 consecutive days (Day 1, 2 and 3) in the 129 rising main. To achieve a typical hydraulic condition, the main pump was turned on 130 for 1 min every hour with a flow rate of 236 L min<sup>-1</sup>, producing a wastewater slug of 131 30 m and an intermittent shear stress of 0.6 N m<sup>-2</sup> in the pipe. With the first pumping 132 event of each test, a mixture solution of biomarkers and rhodamine was spiked into 133 influent using an external peristaltic pump, resulting in the first and the only spiked 134 135 wastewater slug at time 0 ( $t_0$ ). This spiked wastewater slug was pushed 30 m downstream by the subsequent non-spiked wastewater slugs at every following 136 pumping event and arrived at the final sampling port after 7 h of HRT in the pipe. 137 Through matching the length of every wastewater slug with the distance between 138 two sampling ports (i.e. 30 m), the central area of this spiked slug could be captured 139 at #1 to #8 sampling ports in sequence after each hourly pumping event. Meanwhile, 140 during every 1-h pump-off period, samples of the spiked wastewater were collected 141

at 15 min intervals through the sampling port where the spiked slug was located.
Concurrently with the transportation of the spiked wastewater slug, a rhodamine
sensor was connected to #1 to #8 sampling ports for the monitoring of the
rhodamine concentration.

The **Gravity sewer pipe** comprises both water and air phases, between which gas 146 transfer leads to  $1 \sim 4 \text{ mg } L^{-1}$  dissolved oxygen in the bulk liquid phase. A removable 147 section of pipe showed the prevailing existence of sediments at the bottom. This 148 gravity sewer pipe has an internal diameter of 225 mm and is constructed to spiral 149 down from the top layer (inlet) to the ground layer (outlet) with a slope of 0.56%, 150 where wastewater flow is driven by gravitation. The traveling time of wastewater 151 from the pipe inlet to the outlet ranged around 8~10 min. Moreover, this pilot 152 gravity sewer pipe has the unique capability of allowing recirculation. Under the 153 recirculation mode, wastewater effluent at the pipe outlet is collected in a 154 recirculation tank and re-directed to the feeding tank by a recirculation pump. The 155 156 recirculation mode enables wastewater to flow in the gravity sewer pipe as long as required. The average flow velocity was 0.38 m s<sup>-1</sup>; the in-pipe water depth was 157 158 around 5 cm; and the average A/V ratio was approximately 27.5 m<sup>-1</sup> (S1.2 and Figure S1.2). The average shear stress under this flow condition was 0.5 N m<sup>-2</sup>. A sampling 159 160 port is installed near the outlet, allowing access to wastewater in the pipe (Figure S1.1). 161

Triplicate tests were conducted over 3 consecutive days (Day 4, 5 and 6) in the 162 gravity sewer pipe under recirculation mode. At the beginning of each test, a mixture 163 solution of biomarkers and rhodamine was directly spiked into the pipe and 164 continuously mixed with the flowing wastewater. Meanwhile, a rhodamine sensor 165 was connected to the sampling port for online readings during experimental periods. 166 As shown in preliminary tests, the spiked rhodamine presented 2~3 signal peaks over 167 the first few cycles of recirculation and then a sufficiently mixed stage appeared at 168 1.5 h after the spiking event as indicated by the consistent signal intensity. Therefore, 169 for experiments in the recirculating gravity sewer,  $t_0$  of the non-spiked biomarker 170 commenced at the beginning of each test. However,  $t_0$  of the spiked biomarkers was 171 defined when a homogeneously mixed status was reached, in order to minimize the 172 uncertainty of mixing on the evaluation of the spiked biomarkers stability. The HRTs 173 of the recirculating wastewater in the gravity sewer pipe were 3~4 h, during which 174 wastewater samples were collected at 15 min intervals. 175

176 The collected samples were pretreated on site for the analyses of biomarkers and

177 wastewater parameters, including sulfur species, dissolved methane, volatile fatty

acids (VFAs), soluble chemical oxygen demand (SCOD), total and volatile suspended

- solids (TSS and VSS) (for sample pre-treatment and analytical methods see S2).
- 180 Temperature and pH of samples were measured on site using a portable
- 181 pH/temperature meter (TPS Aqua-pH pH/Temp meter).

# 182 Calibration of Drug Transformation Model

#### **Environmental Science & Technology**

As widely applied in previous studies,<sup>10, 14, 30-32</sup> a first-order kinetics is adopted for
illicit drug transformation model in this work (eq 1):

$$C_t = C_0 \cdot e^{-\left(k_{ww} + k'_{bio} \cdot \frac{A}{V}\right) \cdot t}$$
(1)

 $C_t$  is biomarker concentration (µg L<sup>-1</sup>) at time t (h) and  $C_0$  is the initial concentration 185 ( $\mu$ g L<sup>-1</sup>).  $k_{ww}$  (h<sup>-1</sup>) represents the processes in the bulk liquid wastewater, mainly 186 chemical hydrolysis assuming that the sorption to suspended solids or biofilm is 187 limited for investigated biomarkers<sup>5, 14, 16, 33</sup> and the biological activity of suspended 188 solids is negligible compared to the sewer biofilms or sediments.<sup>34-36</sup> It is further 189 postulated that  $k_{ww}$  remains the same under aerobic and anaerobic conditions, 190 according to the findings of McCall et al.<sup>14</sup> and Ramin et al.<sup>16</sup> (about the minor 191 impact of redox condition on abiotic transformation rates).  $k'_{bio}$  (m h<sup>-1</sup>) includes  $k'_{bioa}$ 192 and  $k'_{bioan}$ , representing the biofilm effect under aerobic and anaerobic condition 193 with the normalization of A/V ratio, respectively. The effects of mass transfer 194 limitation on k'bio is considered to be negligible for illicit drug compounds under the 195 experimental conditions as discussed in S1.2. In conclusion, the overall in-sewer 196 transformation rate (h<sup>-1</sup>) depends on  $k_{ww}$  and the  $k'_{bio}$  coupled with a specific A/V 197 ratio. Consequently, the percentage contributions from wastewater and biofilm 198 processes to the overall transformation of a biomarker are quantified by the ratio of 199  $k_{ww}$  against  $k'_{bio} \cdot \frac{A}{V}$ . This study assumed that the mature sewer biofilms in the pilot 200

201 sewers were under steady state with negligible biomass growth over the course of202 experiments.

203	To calibrate the transformation model, experimental datasets obtained from the pilot
204	rising main pipe (Day 1 and 2) and the pilot gravity sewer (Day 4 and 5) pipe were
205	used to estimate $k'_{bioan}$ and $k'_{bioa}$ , respectively, through a Bayesian procedure
206	described in Li <i>et al</i> . <sup>10</sup> The datasets of Day 3 (rising main) and Day 6 (gravity sewer)
207	were reserved for the subsequent model validation. As shown in a previous field-
208	scale study, <sup>10</sup> rhodamine signal was used to normalize the biomarker concentrations
209	which effectively minimized the potential hydraulic uncertainty during wastewater
210	transport. Therefore, for the spiked biomarkers in this pilot study, $C_t$ is normalized by
211	the ratio of the initial rhodamine signal against the signal at $t$ . $k_{ww}$ is estimated using
212	the data on biomarkers transformation in the bulk wastewater (Figure S3.1), which is
213	collected from the experiments using the control sewer reactor without biofilms and
214	from previous <i>in-wastewater studies</i> . <sup>10, 12</sup> The estimation of $k'_{bio}$ is carried out in R
215	(Version 3.2) which executes the Bayesian method in OpenBUGS
216	(http://www.openbugs.net).

# 217 Validation of the pilot-based model

In order to evaluate the validity of the transformation coefficients (*k* values)

219 estimated by the abovementioned pilot-based model across diverse sewer

220 environments, experiment datasets obtained from the pilot rising main at Day 3 and

the pilot gravity sewer at Day 6 together with the literature data collected from three

independent sewer reactor studies<sup>10, 14, 16</sup> are used as the observation inputs 222 representing different testing sewer conditions (datasets in Table S3.2). For each 223 investigated biomarker, the pilot-based k values are used to generate predictions 224 (mean with 95% confidence bounds) with the specific A/V ratios of the four studies 225 (for descriptions of prediction scenarios see Table S3.2). Under each scenario, 226 rejection probability is determined by counting the percentage of experimental 227 observations (i.e. the data on biomarker transformation) located outside the 228 corresponding predictive region. The lower rejection probability reflects the higher 229 validity of the pilot-based k values. 230

## 231 **Performance comparisons of different** *k*-value sets

232 To date, several lab-scale studies have assessed the in-sewer stability of illicit drug biomarkers and estimated the transformation coefficients under their specific testing 233 conditions.<sup>10, 14, 16</sup> Although these k-value sets demonstrated adequate prediction 234 performance in the corresponding studies, their predictive abilities have neither been 235 validated beyond their original testing conditions nor jointly compared under the 236 237 realistic sewer conditions. Importantly, measurements of this pilot study are valuable to model validation, which were obtained from an enlarged pilot-scale system with 238 typical operations and dynamic hydraulics of real sewer systems. 239

For this reason, multiple *k*-value sets ( $k_{ww}^{M_i}$  and  $k_{bio}^{\prime M_i}$ ) are obtained from four different stability studies, i.e., this pilot study and three *sewer reactor studies* (defined as M1~M4 in Table S3.2), and their prediction capabilities are jointly compared under

243	the conditions of the pilot-scale sewer system. Multiple prediction scenarios (mean
244	with 95% confidence bounds) under the conditions of the pilot rising main and the
245	gravity sewer are generated for each investigated biomarker (Table S3.2).
246	Furthermore, in order to identify the $k$ -value set with the highest agreement of
247	prediction to the observations, performance statistics are computed for the
248	predictive scenarios of M1~M4 under each condition, including a stochastic
249	validation metric (the Bayes factor) and two accuracy measures (Pearson correlation
250	coefficient <i>r</i> and variance explained by predictive models based on cross-validation
251	(VEcv)) calculated using RStudio (Version 1.0.143).
252	1) The Bayes Factor
253	The Bayes Factor (BF) is a typical model validation metric, which quantitatively
254	measures the agreement between predictions and measurements together with their
255	internal uncertainties based on stochastic characteristics. <sup>37</sup> As explained by eq 2, the
256	Bayes Factor quantifies the ratio of the probabilities of observations under null
257	hypothesis $H_0$ and alternative hypothesis $H_1$ , respectively, at each validation site (for
258	visual illustrations see Figure S3.2):
	$BF = \frac{P(\text{data} \mid H_0)}{P(\text{data} \mid H_1)} $ (2)

where  $H_0$  is the null hypothesis representing the better match between the observations in pilot sewers and the predictions of a calibrated model, e.g. M1;  $H_1$  is the alternative hypothesis representing the better match between observations and

262	the predictions of other competing models, e.g., M2~M4. For each biomarker under
263	the pilot gravity sewer or the rising main condition, up to three log(BFs) can be
264	computed depending on the available k-value sets in literature, i.e., log(BF12),
265	log(BF13), and log(BF14). At a validation site, log(BF)>0 means $H_0$ is true, indicating
266	that predictions by the $k$ -value set of M1 is favoured by the observed data. A larger
267	absolute value of log(BF) suggests the higher preference to the pilot-based M1 when
268	log(BF)>0, or to the reactor-based M2, M3, or M4 when log(BF)< 0.
269	2) Accuracy of model predictions

The Pearson correlation coefficient r is widely used to assess model predictions 270 against observed data via quantifying the strength and direction of a linear 271 relationship between the two variables (eq 3). When observations x are perfectly 272 linearly related to model predictions y (i.e.,  $y = \beta_0 + \beta_1 x$ ,  $\beta_0$  and  $\beta_1$  are coefficients), r 273 is a suitable indicator of predictive accuracy:<sup>38</sup> 274

$$r = \frac{\sum_{1}^{n} (x_i - \overline{x})(y_i - \overline{y})}{\sqrt{\sum_{1}^{n} (x_i - \overline{x})^2 (y_i - \overline{y})^2}}$$
(3)

where *n* is the number of observations;  $x_i$  is the observed value *i*;  $\overline{x}$  is the mean of 275 the observed values;  $y_i$  is the predicted value *i*;  $\overline{y}$  is the mean of the predicted values. 276 However, when x and y are not well correlated, i.e., when noticeable noise  $\varepsilon$  appears 277 in  $y = \beta_0 + \beta_1 x + \varepsilon$ , r becomes potentially biased and VEcv (eq 4) is recommended as 278 the correct measure of predictive accuracy instead:<sup>38</sup> 279

$$VEcv = (1 - \frac{\sum_{1}^{n} (x_{i} - y_{i})^{2}}{\sum_{1}^{n} (x_{i} - \overline{x})^{2}}) \times 100(\%)$$
(4)

#### 280 RESULTS AND DISCUSSION

#### 281 Wastewater Compositions and Biological Activities

The observations of wastewater parameters in this study were the same as reported 282 in Gao et al.,<sup>17</sup> which conducted experiments in the same setup. The sewer 283 characteristics in the pilot sewer system, in terms of wastewater parameters and 284 biological activities, are also compared to the literature data. It is found that the 285 variations of wastewater parameters in the pilot sewers (Figure S1.3) are similar to 286 those in other sewer studies (Table S1.2).<sup>10, 39, 40</sup> For instance, wastewater pH kept 287 relatively stable over 8 h in the pilot rising main (6.99±0.11), while pH in the pilot 288 gravity sewer increased by ~0.25 units during the first 2 h, likely due to the CO<sub>2</sub> and 289 H<sub>2</sub>S stripping. Wastewater temperature remained consistently stable in the pilot 290 sewers (22.9±0.6 °C). TSS was higher in the pilot gravity sewer (500-800 mg L<sup>-1</sup>) than 291 the pilot rising main (200-410 mg L<sup>-1</sup>) due to resuspension of sediments. The interday 292 deviations of TSS were attributed to the daily variation of real wastewater, while the 293 intraday changes of TSS were relatively insignificant over the experimental period. 294 Based on previous studies, sulfate reducing bacteria and methanogenic archaea are 295 the primary microorganisms responsible for not only the carbon/sulfur 296 transformation, but also the biomarker stability in biofilms.<sup>14, 18, 41, 42</sup> In this work, 297

298	sulfide and methane production rates are used as the major indicators of biological
299	activities in the pilot sewers, which are comparable to the literature data (Table S1.2).
300	The simultaneously increasing pattern of sulfide and methane in the pilot rising main
301	(Figure S1.3) was also similar to that in real sewers. <sup>43-46</sup> In contrast, sulfide or
302	methane production was detected to be negligible or even negative in the pilot
303	gravity sewer, which could be explained by the faster sulfide oxidation than the
304	concurrent sulfate reduction and/or the emission of sulfide and methane gas into the
305	air phase.
306	VFAs and SCOD are the major substrates for heterotrophic bacteria and
307	methanogens, which present varying consumption rates in different sewer
308	environments (Table S1.2). Primarily being produced by fermentation process under
309	anaerobic condition, <sup>39, 47</sup> the consumption rate of VFAs in the pilot rising main was
310	lower than that in the gravity sewer (Figure S1.3). The consumption rate of SCOD was
311	higher than VFAs in the pilot rising main, while in the pilot gravity sewer, the
312	concurrent consumption of SCOD and VFAs was generally similar. Overall, this
313	unique pilot sewer system has an environmental condition representative of real
314	sewers.

# **Biomarkers Stability in the Pilot Sewer System**

The online monitoring of rhodamine showed the dynamic flow patterns in the pilot sewer pipes during experiment periods (Figure 1). The rhodamine profiles shared the same results of Gao *et al.*<sup>17</sup> In the pilot rising main, the staged profiles of rhodamine revealed the plug-flow regime as a response to the intermittent pumping events. In the pilot gravity sewer under recirculation mode, rhodamine profiles reflected the continuous mixing and indicated the commencing of a sufficiently mixed stage at 1.5 h after the spiking event.

Various patterns were found for the investigated biomarkers in the pilot sewer 323 system (Figure 1). Over the consecutive tests in the pilot gravity sewer or the rising 324 main, the interday divergence of the transformations for each biomarker was 325 relatively limited as indicated by the low standard deviations. COC, MDMA, and MTD 326 exhibited decreasing trends during the testing periods in both the pilot rising main 327 and the gravity sewer, where MTD was observed to have the most rapid degradation 328 with ~25% loss in 3 h, followed by ~20% loss of COC and MDMA. During the longer 329 HRT periods in the pilot rising main, these biomarkers still presented similar 330 degradation trends and overall 35~40% losses were observed after 8 h. By contrast, 331 KET and METH showed relatively good stability with <20% losses in both pipes. In 332 333 addition, formations of BE and MOR were observed over the experimental periods, which were the combined results of multiple concurrent in-sewer processes, 334 335 including the transformation of biomarker itself, the back-transformation of parent compounds and/or the potential deconjugation of glucuronides in the raw 336 wastewater.<sup>11, 12</sup> Data on the transformations of MOR and MTD in the pilot sewers 337 was also reported previously.<sup>17</sup> Similarly, BE and MOR formations were also found in 338

other testing sewer conditions, e.g., in lab reactors<sup>11, 12, 14, 16, 23, 31, 48</sup> and real sewer
pipe.<sup>10</sup>

#### 341 **Drug Transformation Models**

## 342 Calibration and validation of the transformation model

Calibrations of the transformation model based on the measured data of this pilot 343 study are presented in Figure S3.3 and the values of transformation coefficients are 344 reported as mean with 95% credible intervals (CI) in Table S3.1. Among the estimated 345  $k'_{bio}$  values, the relatively high  $k'_{bioa}$  and/or  $k'_{bioan}$  for COC, MDMA, and MTD suggest 346 the important effect of biofilm on their stability in sewers. Moreover, for most 347 biomarkers, their  $k'_{bioa}$  values are found to be higher than the corresponding  $k'_{bioan}$ 348 values, indicating the higher biofilm-specific impact in aerobic condition compared 349 to anaerobic condition. Effect of mass transfer resistance on biodegradation process 350  $(k'_{bio})$  is considered to be limited for most investigated biomarkers (see discussions in 351 S1.2). Meanwhile, the relatively high  $k_{ww}$  for COC, KET, and MTD indicate their 352 comparatively evident decreases in the bulk liquid wastewater due to the processes 353 such as chemical hydrolysis.<sup>10, 12</sup> However, k value was not estimated for BE or MOR, 354 which showed significant formations in the pilot sewers (Figure 1). This is because the 355 knowledge on BE or MOR formation was limited since 1) it is difficult to accurately 356 quantify the various contributing sources, such as the parent compounds and/or 357 conjugated forms in the raw wastewater and 2) this study did not spike labelled BE or 358 MOR to exclude the contribution from other precursors and hence to identify their 359

Page 20 of 39

360	specific transformation in sewers. In consequence, the transformation model was not
361	calibrated for the prediction of the in-sewer formations of BE or MOR.
362	Validity of the estimated pilot-based k values in diverse sewer environments are
363	subsequently examined. For each investigated biomarker, the measurement datasets
364	obtained from different stability studies and the predictive regions generated by the
365	pilot-based k values under the corresponding scenario (e.g., A/V ratio and redox
366	condition) are jointly plotted in Figure 2. Through showing whether the literature
367	data points are encompassed or rejected by the prediction regions, Figure 2 visually
368	reflects the transferability and applicability of the pilot-based $k$ values to various
369	sewer conditions. The rejection probability for each scenario is reported in Table S3.3.
370	The prediction capability of the pilot-based k values is successfully validated for KET
371	and METH under all sewer conditions. As shown in Figure 2, the predictive regions
372	almost completely encompass the observations from the four stability studies under
373	aerobic and anaerobic conditions, except for a few data points over long HRTs.
374	Validity of the pilot-based $k$ values is successfully demonstrated for COC, MDMA,
375	and MTD in the pilot sewers but is less successful against the testing conditions of
376	the three sewer reactor studies. For these three biomarkers, relatively high rejection
377	probabilities of the pilot-based $k$ values are determined for different reasons: 1) the
378	overestimation of COC decreases at the very high A/V ratio condition (175 $m^{-1}$ ) of
379	Ramin <i>et al.</i> <sup>21</sup> ; 2) the overestimation of MDMA transformation compared to its
380	observed insignificant changes in lab reactors, <sup>10, 14</sup> suggesting that MDMA could be

381	partially degraded by certain microbes that existed in the realistic pilot sewers but
382	not in lab-scale reactors. Similarly, McCall et al. <sup>14</sup> also found the biofilm specific
383	transformation for MDMA and attributed it to the divergent biofilm growing
384	conditions and the different transformation potentials; 3) the underestimation of
385	MTD losses compared to the observed drastic decreases in the anaerobic reactor of
386	Li <i>et al.</i> <sup>10</sup> The diffusion limitation could affect $k'_{bio}$ in the pilot rising main (with
387	intermittent pumping events) due to the high biodegradability of MTD (full
388	discussion in S1.2).

# 389 Model performance evaluation I: the different k-value sets

Multiple *k*-value sets ( $k_{ww}^{M_i}$  and  $k_{bio}^{'M_i}$ ) are generated from four stability studies as explained in Table S3.2 and illustrated in Figure S3.3. Figure 3 shows the joint distributions of  $k_{ww}^{M_i}$  and  $k_{bio}^{'M_i}$  which visually reflect the consistency or discrepancy among the *k*-value sets estimated by different studies.

For most investigated biomarkers, the close distributions of  $k_{ww}^{M_i}$  indicate the general congruence of  $k_{ww}^{M_i}$  derived from different stability studies. Such consistency suggests that the processes in the bulk wastewater (mainly the abiotic physicochemical process) of different sewer environments could lead to similar transformation of biomarkers. However, certain deviations are found for COC, KET, and MTD under aerobic and/or anaerobic conditions, where the centroids of  $k_{ww}^{M_1}$  and/or  $k_{ww}^{M_2}$  show relative distances with those of  $k_{ww}^{M_3}$  and/or  $k_{ww}^{M_4}$ . The deviations may be attributed to

401	the assumptions about the independent relationship of $k_{ww}$ with wastewater
402	compositions and properties such as the redox potential or suspended solids
403	concentration. Specific experiments are required to evaluate the effects of
404	wastewater composition, redox potential, and pH on $k_{ww}$ and the associated $k_{bio}$ .
405	By contrast, distributions of $k_{bio}^{M_i}$ are more deviated for most investigated biomarkers.
406	Since $k_{bio}^{M_i}$ represents the contribution of biofilm to biomarker stability in sewers, such
407	deviations reveal that the biofilm-specific effects in different studies are strongly
408	related to the microbial communities, functions, and activities in biofilms. Generally
409	speaking, $k'_{bio}^{M_1}$ and/or $k'_{bio}^{M_2}$ are usually found to be higher than $k'_{bio}^{M_3}$ and/or $k'_{bio}^{M_4}$ ,
410	except for KET with closer distributions of $k_{bio}^{\prime M_i}$ at minor levels. This could be
411	explained by the stronger biological activities of biofilms in the pilot sewer pipes and
412	the sewer reactors of Li et al. <sup>10</sup> compared to biofilms in the reactors of McCall et al. <sup>14</sup>
413	and/or Ramin <i>et al.</i> <sup>16</sup> Moreover, $k_{bio}^{M_1}$ and/or $k_{bio}^{M_2}$ appear to be generally equivalent
414	for COC, KET, and METH, while present certain dissimilarity for MDMA and MTD
415	under anaerobic and/or aerobic conditions. Such discrepancies could be due to the
416	different microbial communities in biofilms grown in pilot-scale sewer pipes and lab-
417	scale sewer reactors. Also, the continuous mixing condition in the lab reactors <sup>10, 42</sup>
418	could accelerate the transformation of certain biomarkers in biofilms.
419	The embedded histograms in Figure 3 show the contributions from wastewater and
420	biofilm processes to the overall transformation of biomarkers in the pilot sewer
421	system. For most biomarkers, the k-value sets of M1 and M2 usually suggest the

422	dominant role of biofilms because of the relatively high $k_{bio}^{\prime M_1}$ and $k_{bio}^{\prime M_2}$ values, except
423	for MDMA and METH under anaerobic condition and for KET under both aerobic and
424	anaerobic conditions. The k-value sets of M3 and M4 also recognize the governing
425	effect of biofilms on MTD under anaerobic condition and on KET, MDMA and METH
426	under both aerobic and anaerobic conditions. The overall KET variation predicted by
427	M3 is actually negligible and akin to the observed high stability of KET under
428	multiple experimental conditions of McCall <i>et al.</i> <sup>14</sup> On the contrary, the $k$ -value sets
429	of M3 and M4 indicate higher impact of wastewater processes ( $k_{ww}$ ) on MTD under
430	aerobic condition and on COC under both aerobic and anaerobic conditions. These
431	are consistent with the findings of McCall <i>et al</i> . <sup>14</sup> and Ramin <i>et al.</i> , <sup>16</sup> suggesting that
432	processes in the bulk wastewater dominated the transformation of MTD (under
433	aerobic condition) and COC, while sewer biofilms did not significantly enhance their
434	transformation.

# 435 Model performance evaluation II: the Bayes Factor, r, and VEcv values

436 Figure 4 shows the Bayes Factor, *r*, and *VEcv* across the predictive scenarios

437 generated by the *k*-value sets of M1~M4 under the pilot sewer conditions (for detail

numbers see Table S3.4 to S3.6). Predictions of M1~M4 versus observations in the

pilot sewer pipes (Day 3 and Day 6) are jointly plotted for each biomarker in FigureS3.4.

As indicated by the results of performance statistics, the *k*-value set of M1 provides
high prediction accuracy for COC under the pilot rising main and for MDMA and

443	MTD under both pilot gravity sewer and rising main conditions. In these cases, the
444	likelihood of log(BFs) locating in the positive area is dominant, suggesting the closer
445	agreements of pilot-sewer measurements to the outputs of M1 than those of
446	M2~M4 (Table S3.4). Moreover, experimental observations and the outputs of M1
447	are more closely matched (i.e., close to $y = x$ ) compared to the correlations for
448	M2~M4. Consistently, higher <i>r</i> and <i>VEcv</i> values are quantified for M1 (Table S3.5 to
449	S3.6), corroborating the higher prediction accuracy of M1 for the discussed cases. It
450	should be noted that the datasets of Day 3 and Day 6 from the pilot sewers may
451	naturally favour M1, which is calibrated under similar conditions.
452	Meanwhile, all the $k$ -value sets of M1~M4 provide similarly high prediction accuracy
453	for COC under the pilot gravity sewer condition. The performances of M1~M2 are
454	also comparable for KET under the pilot rising main condition, where a strong linear
455	relationship between observations and the predictions of M1 or M2 is found. In these
456	cases, log(BFs) mainly spread out within the area close to zero, implying the
457	comparable prediction outputs of M1~M4 (Table S3.4). Moreover, the $r$ and VEcv
458	values of M1~M4 are similar and high for the discussed cases, which further verify
459	the comparable and reasonable prediction capabilities of the k-value sets derived
460	from different stability studies (Table S3.5 to S3.6).
461	For KET under the pilot gravity sewer and METH under both pilot gravity sewer and
462	rising main conditions, the predicted overall transformations by all k-value sets are
463	limited, i.e. $\leq$ 20% as shown in Figure S3.4. The prediction accuracy of M1~M4 is

476 On the contrary, validity of the *k*-value sets derived from different stability studies cannot be effectively assessed for BE or MOR because their actual stability in the 477 pilot sewers might be masked by formations from other sources and/or 478 deconjugation processes. Such in-sewer formations bring uncertainty to the back-479 estimation of the drugs that could be metabolized/transformed to BE (i.e., the 480 consumption of COC) or MOR (e.g., the consumptions of codeine, heroin, and 481 morphine itself in glucuronide form). Information on the sources contributing to BE 482 and MOR formation, such as the concentrations and transformation pathways of 483

their parent compounds and/or conjugated forms, should be determined before the
formation processes are incorporated into the transformation model.

The comprehensive evaluations of different calibrated transformation models help to 486 choose the k values to give the best predictive performance in the application of 487 WBE. Collectively speaking, for the biomarkers presenting evident losses in sewers 488 (e.g., COC and MTD), both lab-scale and pilot-scale studies estimate relatively high k489 values to predict such decreases over time. On the other hand, most stability studies 490 suggest the overall insignificant transformation of KET and METH in sewers. Their 491 predicted transformations are also limited over time based on all k-value sets, 492 although with some random variations. Importantly, due to the complex microbial 493 processes in real sewers, the pilot-based k values exhibit advantage in predicting the 494 fate of the biomarkers (e.g., MDMA) which are stable in lab-scale reactors but show 495 partial degradation in the pilot sewer pipes, especially over long HRT. Unfortunately, 496 none of the available k-value sets are capable of simulating the increasing patterns 497 498 of biomarkers (e.g., BE and MOR) in pilot sewers. In short, the transformation model calibrated by the measurements in the pilot-scale sewer pipes can be applied as a 499 500 credible tool for the future WBE study.

501 ASSOCIATED CONTENT

## 502 Supporting Information

Additional information about the layout and parameters of the pilot sewer system, experimental observations of this pilot study, analytical methods for wastewater samples, and results of model validation and comparison are provided. Supporting information is available free of charge via the Internet.

#### 507 AUTHOR INFORMATION

## 508 Corresponding Author

509 \*Guangming Jiang: E-mail: gjiang@uow.edu.au

#### 510 ACKNOWLEDGEMENT

- 511 This research was supported by the ARC Discovery project (DP150100645). Jiaying Li
- receives the support from China Scholarship Council. Jianfa Gao receives an ARC
- scholarship (DP150100645). Phong K. Thai was funded by the QUT VC Fellowship during
- 514 part of this study. Jochen Mueller acknowledges the UQ Research Fellowship. Guangming
- Jiang is the recipient of an Australian Research Council DECRA Fellowship
- 516 (DE170100694). We specially acknowledge the collaboration with Queensland Urban
- 517 Utilities for the measurement campaign at Innovation Centre, Luggage Point Wastewater
- 518 Treatment Plant. We thank for the assistance of Natasha Rossi and Tobias Hesse during
- the onsite experiments. We thank Chris Paxman for English proof reading.

#### 520 REFERENCE

- 521 1. Daughton, C. G. Pharmaceuticals and Personal Care Products in the
- 522 Environment: Overarching Issues and Overview. **2001**, *791*, 2-38.

2. Daughton, C. G., Illicit drugs in municipal sewage: Proposed new nonintrusive 523 tool to heighten public awareness of societal use of illicit-abused drugs and their 524 potential for ecological consequences. In *Pharmaceuticals and Personal Care Products* 525 in the Environment: Scientific and Regulatory Issues, American Chemical Society: 526 Washington DC, 2001; Vol. 791, pp 348-364. 527 528 3. Zuccato E; Chiabrando C; Castiglioni S; Calamari D; Bagnati R; Schiarea S; R., F. Cocaine in surface waters- a new evidence-based tool to monitor community drug 529 abuse. Environ. Health 2005, 4 (14), 7. 530 van Nuijs, A. L. N.; Castiglioni, S.; Tarcomnicu, I.; Postigo, C.; de Alda, M. L.; 4. 531 Neels, H.; Zuccato, E.; Barcelo, D.; Covaci, A. Illicit drug consumption estimations 532 derived from wastewater analysis: A critical review. Sci. Total Environ. 2011, 409 (19), 533 3564-3577. 534 European Monitoring Centre for Drugs and Drug Assessing illicit drugs in 535 5. wastewater Advances in wastewater-based drug epidemiology; Luxembourg, 2016. 536 6. Ort, C.; Lawrence, M. G.; Rieckermann, J.; Joss, A. Sampling for Pharmaceuticals 537 and Personal Care Products (PPCPs) and Illicit Drugs in Wastewater Systems: Are 538 Your Conclusions Valid? A Critical Review. Environ. Sci. Technol. 2010, 44 (16), 6024-539 6035. 540 Lai, F. Y.; Ort, C.; Gartner, C.; Carter, S.; Prichard, J.; Kirkbride, P.; Bruno, R.; Hall, 7. 541 W.; Eaglesham, G.; Mueller, J. F. Refining the estimation of illicit drug consumptions 542 from wastewater analysis: co-analysis of prescription pharmaceuticals and 543

544 uncertainty assessment. *Water Res* **2011**, *45* (15), 4437-48.

ACS Paragon Plus Environment

545	8. Castiglioni, S.; Bijlsma, L.; Covaci, A.; Emke, E.; Hernandez, F.; Reid, M.; Ort, C.;
546	Thomas, K. V.; van Nuijs, A. L. N.; de Voogt, P.; Zuccato, E. Evaluation of Uncertainties
547	Associated with the Determination of Community Drug Use through the
548	Measurement of Sewage Drug Biomarkers. Environ. Sci. Technol. 2013, 47 (3), 1452-
549	1460.
550	9. van Nuijs, A. L. N.; Lai, F. Y.; Been, F.; Andres-Costa, M. J.; Barron, L.; Baz-
551	Lomba, J. A.; Berset, J. D.; Benaglia, L.; Bijlsma, L.; Burgard, D.; Castiglioni, S.;
552	Christophoridis, C.; Covaci, A.; de Voogt, P.; Emke, E.; Fatta-Kassinos, D.; Fick, J.;
553	Hernandez, F.; Gerber, C.; Gonzalez-Marino, I.; Grabic, R.; Gunnar, T.; Kannan, K.;
554	Karolak, S.; Kasprzyk-Hordern, B.; Kokot, Z.; Krizman-Matasic, I.; Li, A.; Li, X. Q.; Love,
555	A. S. C.; de Alda, M. L.; McCall, A. K.; Meyer, M. R.; Oberacher, H.; O'Brien, J.; Quintana,
556	J. B.; Reid, M.; Schneider, S.; Simoes, S. S.; Thomaidis, N. S.; Thomas, K.; Yargeau, V.;
557	Ort, C. Multi-year inter-laboratory exercises for the analysis of illicit drugs and
558	metabolites in wastewater: Development of a quality control system. Trac-Trend Anal
559	Chem <b>2018,</b> <i>103</i> , 34-43.
560	10. Li, J.; Gao, J.; Thai, P. K.; Sun, X.; Mueller, J. F.; Yuan, Z.; Jiang, G. Stability of Illicit
561	Drugs as Biomarkers in Sewers: From Lab to Reality. Environ. Sci. Technol. 2018, 52
562	(3), 1561-1570.
563	11. Gao, J.; Banks, A.; Li, J.; Jiang, G.; Lai, F. Y.; Mueller, J. F.; Thai, P. K. Evaluation of
564	in-sewer transformation of selected illicit drugs and pharmaceutical biomarkers. Sci.

565 *Total Environ*. **2017,** *609*, 1172-1181.

- 12. Thai, P. K.; Jiang, G.; Gernjak, W.; Yuan, Z.; Lai, F. Y.; Mueller, J. F. Effects of
- sewer conditions on the degradation of selected illicit drug residues in wastewater.
- 568 *Water Res.* **2014,** *48*, 538-47.
- 13. McCall, A. K.; Bade, R.; Kinyua, J.; Lai, F. Y.; Thai, P. K.; Covaci, A.; Bijlsma, L.; van
- 570 Nuijs, A. L.; Ort, C. Critical review on the stability of illicit drugs in sewers and
- 571 wastewater samples. *Water Res.* **2016**, *88*, 933-47.
- 14. McCall, A. K.; Scheidegger, A.; Madry, M. M.; Steuer, A. E.; Weissbrodt, D. G.;
- 573 Vanrolleghem, P. A.; Kraemer, T.; Morgenroth, E.; Ort, C. Influence of Different Sewer
- 574 Biofilms on Transformation Rates of Drugs. Environ. Sci. Technol. 2016, 50 (24),
- 575 13351-13360.
- 15. McCall, A. K.; Palmitessa, R.; Blumensaat, F.; Morgenroth, E.; Ort, C. Modeling
- 577 in-sewer transformations at catchment scale Implications on drug consumption
- estimates in wastewater-based epidemiology. *Water Res.* **2017**, *122*, 655-668.
- 16. Ramin, P.; Brock, A. L.; Causanilles, A.; Valverde Perez, B.; Emke, E.; de Voogt, P.;
- 580 Polesel, F.; Plosz, B. G. Transformation and sorption of illicit drug biomarkers in sewer
- 581 biofilms. *Environ. Sci. Technol.* **2017,** *51* (18), 10572–10584.
- 17. Gao, J.; Li, J.; Jiang, G.; Shypanski, A. H.; Nieradzik, L. M.; Yuan, Z.; Mueller, J. F.;
- 583 Ort, C.; Thai, P. K. Systematic evaluation of biomarker stability in pilot scale sewer
- 584 pipes. Water Res **2018**, 151, 447-455.
- 18. Hvitved-Jacobsen, T.; Vollertsen, J.; Nielsen, A. H. Sewer Processes: Microbial
- 586 and Chemical Process Engineering of Sewer Networks. Second ed.; CRC Press: Boca
- 587 Raton, 2013.

588	19.	Plosz, B. G.; Reid, M. J.; Borup, M.; Langford, K. H.; Thomas, K. V.	
589	Biotra	nsformation kinetics and sorption of cocaine and its metabolites and the	
590	factor	s influencing their estimation in wastewater. Water Res. 2013, 47 (7), 2129-	
591	2140.		
592	20.	Ostman, M.; Fick, J.; Nasstrom, E.; Lindberg, R. H. A snapshot of illicit drug use	
593	in Swe	eden acquired through sewage water analysis. Sci. Total Environ. 2014, 472,	
594	862-871.		
595	21.	Ramin, P.; Libonati Brock, A.; Polesel, F.; Causanilles, A.; Emke, E.; de Voogt, P.;	
596	Plosz,	B. G. Transformation and sorption of illicit drug biomarkers in sewer systems:	
597	under	standing the role of suspended solids in raw wastewater. Environ. Sci. Technol.	
598	2016,	50 (24), 13397-13408.	
599	22.	Castiglioni, S.; Zuccato, E.; Crisci, E.; Chiabrando, C.; Fanelli, R.; Bagnati, R.	
600	Identi	fication and measurement of illicit drugs and their metabolites in urban	
601	waste	water by liquid chromatography-tandem mass spectrometry. Anal Chem 2006,	
602	78 (24), 8421-8429.		
603	23.	Baker, D. R.; Kasprzyk-Hordern, B. Critical evaluation of methodology	
604	comm	only used in sample collection, storage and preparation for the analysis of	
605	pharm	naceuticals and illicit drugs in surface water and wastewater by solid phase	
606	extrac	tion and liquid chromatography-mass spectrometry. J Chromatogr A. 2011,	
607	1218	(44), 8036-59.	

608	24.	Chen, C.; Kostakis, C.; Irvine, R. J.; Felgate, P. D.; White, J. M. Evaluation of pre-
609	analys	is loss of dependent drugs in wastewater: stability and binding assessments.
610	Drug	Test Anal <b>2013,</b> 5 (8), 716-21.
611	25.	Senta, I.; Krizman, I.; Ahel, M.; Terzic, S. Assessment of stability of drug
612	bioma	rkers in municipal wastewater as a factor influencing the estimation of drug
613	consu	mption using sewage epidemiology. Sci Total Environ <b>2014,</b> 487, 659-65.
614	26.	Jelic, A.; Rodriguez-Mozaz, S.; Barcelo, D.; Gutierrez, O. Impact of in-sewer
615	transf	ormation on 43 pharmaceuticals in a pressurized sewer under anaerobic
616	condit	ions. <i>Water Res</i> . <b>2015,</b> 68, 98-108.
617	27.	Gao, J.; Li, J.; Jiang, G.; Yuan, Z.; Eaglesham, G.; Covaci, A.; Mueller, J. F.; Thai, P.
618	K. Stal	pility of alcohol and tobacco consumption biomarkers in a real rising main
619	sewer.	Water Res. <b>2018,</b> 138, 19-26.
620	28.	Ian A. Watson; Sascha E.Oswald; Steven A. Banwart; Roger S. Crouch;
621	Thorn	ton, S. F. Modeling the dynamics of fermentation and respiratory processes in a
622	groun	dwater plume of phenolic contam- inants interpreted from laboratory- to field-

- 623 scale. *Environ. Sci. Technol.* **2005,** *39* (22), 8829-8839.
- 624 29. Shypanski, A. H.; Yuan, Z.; Sharma, K. Influence of pressure main pumping
- 625 frequency on sulfide formation rates in sanitary sewers. *Environmental Science: Water*
- 626 *Research & Technology* **2018,** *4* (3), 403-410.
- 30. Ramin, P.; Polesel, F.; Brock, A. L.; Plósz, B. G. The impact of temperature on the
- transformation of illicit drug biomarkers in wastewater. Sci. Total Environ. 2018, 644,
- 629 **1612-1616**.

630	31.	Senta, I.; Krizman, I.; Ahel, M.; Terzic, S. Assessment of stability of drug
631	bioma	arkers in municipal wastewater, as a factor influencing the estimation of drug
632	consu	mption using sewage epidemiology. Sci. Total Environ. 2014, 487, 659-665.
633	32.	Devault, D. A.; Levi, Y.; Karolak, S. Applying sewage epidemiology approach to
634	estim	ate illicit drug consumption in a tropical context: Bias related to sewage
635	tempe	erature and pH. Sci. Total Environ. 2017, 584, 252-258.
636	33.	Baker, D. R.; Ocenaskova, V.; Kvicalova, M.; Kasprzyk-Hordern, B. Drugs of
637	abuse	in wastewater and suspended particulate matterfurther developments in
638	sewag	ge epidemiology. <i>Environ Int</i> <b>2012,</b> <i>4</i> 8, 28-38.
639	34.	Gutierrez, O.; Park, D.; Sharma, K. R.; Yuan, Z. Effects of long-term pH elevation
640	on the	e sulfate-reducing and methanogenic activities of anaerobic sewer biofilms.
641	Water	r Res. <b>2009,</b> <i>43</i> (9), 2549-2557.
642	35.	Guisasola, A.; Sharma, K. R.; Keller, J.; Yuan, Z. Development of a model for
643	assessing methane formation in rising main sewers. Water Res. 2009, 43 (11), 2874-	
644	2884.	
645	36.	Liu, Y. W.; Tugtas, A. E.; Sharma, K. R.; Ni, B. J.; Yuan, Z. G. Sulfide and methane
646	production in sewer sediments: Field survey and model evaluation. Water Res. 2016	
647	89, 14	2-150.
648	37.	Liu, Y.; Chen, W.; Arendt, P.; Huang, HZ. Toward a Better Understanding of
649	Mode	l Validation Metrics. J Mech Design 2011, 133 (7).
650	38.	Li, J. Assessing the accuracy of predictive models for numerical data: Not r nor
651	r2, wh	ny not? Then what? <i>Plos One <b>2017,</b> 12</i> (8), e0183250.

39. Æsøy, A.; Storfjell, M.; Mellgren, L.; Helness, H.; Thorvaldsen, G.; Ødegaard, H.;

Bentzen, G. A comparison of biofilm growth and water quality changes in sewers with

anoxic and anaerobic (septic) conditions. *Water Sci. Technol.* **1997**, *36* (1), 303-310.

40. Sharma, K.; Ganigue, R.; Yuan, Z. pH dynamics in sewers and its modeling.

- 656 *Water Res.* **2013,** *47* (16), 6086-6096.
- 41. Jiang, G.; Sharma, K. R.; Guisasola, A.; Keller, J.; Yuan, Z. Sulfur transformation

in rising main sewers receiving nitrate dosage. *Water Res.* **2009**, *43* (17), 4430-4440.

42. Thai, P. K.; Jiang, G. M.; Gernjak, W.; Yuan, Z. G.; Lai, F. Y.; Mueller, J. F. Effects of

sewer conditions on the degradation of selected illicit drug residues in wastewater.

- 661 *Water Res.* **2014,** *48*, 538-547.
- 43. Sharma, K. R.; Yuan, Z.; de Haas, D.; Hamilton, G.; Corrie, S.; Keller, J. Dynamics

and dynamic modelling of H(2)S production in sewer systems. *Water Res.* 2008, 42
(10), 2527-2538.

- 44. Foley, J.; Yuan, Z.; Keller, J.; Senante, E.; Chandran, K.; Willis, J.; Shah, A.; van
- Loosdrecht, M.; van Voorthuizen, E. N2O and CH4 emission from wastewater
- *collection and treatment systems: technical report;* Global Water Research Coalition:
- London, United Kingdom, 2011.

Guisasola, A.; de Haas, D.; Keller, J.; Yuan, Z. Methane formation in sewer
systems. *Water Res.* 2008, 42 (6), 1421-1430.

- 46. Liu, Y. W.; Sharma, K. R.; Fluggen, M.; O'Halloran, K.; Murthy, S.; Yuan, Z. G.
- 672 Online dissolved methane and total dissolved sulfide measurement in sewers. Water
- 673 *Res.* **2015,** *68*, 109-118.

- 47. Wang, K. J.; Zeeman, G.; Lettinga, G. Alteration in Sewage Characteristics Upon
- 675 Aging. Water Sci. Technol. **1995,** 31 (7), 191-200.
- 48. Ramin, P.; Valverde-Perez, B.; Polesel, F.; Locatelli, L.; Plosz, B. G. A systematic
- 677 model identification method for chemical transformation pathways the case of
- heroin biomarkers in wastewater. *Sci Rep* **2017**, *7* (1), 9390.

679



681 Table of Contents



**Figure 1.** Transportation of rhodamine as a flow tracer and stability of biomarkers in the pilot rising main pipe and the pilot gravity sewer pipe. (Transformations of MOR and MTD in the pilot sewer pipes were also reported recently<sup>17</sup>)



**Figure 2.** Validation of the pilot-based transformation coefficients through the predictive scenarios of the corresponding experimental observations in different stability studies.



**Figure 3**. Joint distributions of the transformation coefficients  $(k_{ww}^{M_i} \text{ and } k_{bio}^{'M_i})$  derived from the four stability studies (M1~M4) under aerobic and anaerobic conditions. The embedded histograms indicate the contributions from wastewater processes against biofilm effects to the overall biomarker transformation in the pilot sewer pipes.



**Figure 4**. Model performance comparison: results of the Bayes factor, *r*, and *VEcv* for comparing the predictive scenarios generated by the *k*-value sets of four stability studies (M1 to M4) under the conditions of the pilot sewer pipes.