

DTU Library

Relationship of baseline and maximum glucocorticoid concentrations to migration propensity – a field test with wild sub-adult brown trout (Salmo trutta)

Jain-Schlaepfer, Sofia; Midwood, Jonathan D.; Larsen, Martin Hage; Aarestrup, Kim; King, Greg; Suski, Cory; Cooke, Steven J.

Published in: Canadian Journal of Zoology

Link to article, DOI: 10.1139/cjz-2018-0044

Publication date: 2018

Document Version Peer reviewed version

Link back to DTU Orbit

Citation (APA):

Jain-Schlaepfer, S., Midwood, J. D., Larsen, M. H., Aarestrup, K., King, G., Suski, C., & Cooke, S. J. (2018). Relationship of baseline and maximum glucocorticoid concentrations to migration propensity – a field test with wild sub-adult brown trout (Salmo trutta). Canadian Journal of Zoology, 96(12), 1346-1352. DOI: 10.1139/cjz-2018-0044

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

• Users may download and print one copy of any publication from the public portal for the purpose of private study or research.

- · You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Relationship of baseline and maximum glucocorticoid concentrations to migration propensity -1 2 a field test with wild sub-adult brown trout (Salmo trutta) 3 S.M.R. Jain-Schlaepfer^{*a}, J.D. Midwood^b, M.H. Larsen^{cd}, K. Aarestrup^c, G.D. King^e, C.D. Suski^e and 4 S.J. Cooke^a 5 6 ^aEcology and Conservation Physiology Laboratory, Department of Biology and Institute of 7 Environmental Science, Carleton University, 1125 Colonel By Dr. Ottawa, ON K1S 5B6, Canada 8 b Great Lakes Laboratory for Fisheries and Aquatic Science. Fisheries and Oceans Canada, 9 Burlington, ON, L7S1A1, Canada 10 ^c National Institute of Aquatic Resources, Freshwater Fisheries, Technical University of 11 Denmark, Vejlsøvej 39, 8600 Silkeborg, Denmark 12 ^d Danish Centre for Wild Salmon, Brusgårdsvej 15, DK-8960 Randers SØ, Denmark. 13 ^eDepartment of Natural Resources and Environmental Sciences, University of Illinois, 1102 S. 14 Goodwin Ave, Urbana, IL 61801, USA. 15 16 *sofiaj@rogers.com, 0467694922, 32 McKinley Street, North Ward 4810, Australia 17

19 Relationship of baseline and maximum glucocorticoid concentrations to migration propensity –

20 a field test with wild sub-adult brown trout (Salmo trutta)

S.M.R. Jain-Schlaepfer, J.D. Midwood, M.H. Larsen, K. Aarestrup, G.D. King, C.D. Suski and S.J.

22 Cooke

21

23 Abstract

There is considerable variation in individuals' glucocorticoid (GC) baseline status and stress 24 25 responses, yet the cause and consequence of this variation remains ambiguous. Attempts to 26 relate GC levels to fitness and life-history tradeoffs have yielded variable results. In this study, 27 we evaluated whether baseline and post-stressor GC hormone concentrations predicted 28 migration strategy (i.e., resident or migrant) and successful seaward migration in a partially migrating population of juvenile brown trout (Salmo trutta (Linnaeus, 1758)). Baseline (N=99) 29 or post-stressor (N=102) plasma cortisol concentrations were obtained from trout and they 30 were tagged with passive integrated transponder (PIT) and released in a natural Danish stream. 31 32 Subsequently, fish were tracked with PIT reader systems and the stream was resampled for resident individuals. GC levels were not found to be associated with recapture of resident 33 individuals or migration propensity to our first tracking station (S1), but increased baseline (and 34 35 not post-stressor) GC levels were associated with increased passage from S1 to our second tracking station, which anecdotally was an area of high predation or challenge. Our study found 36 no evidence to suggest that cortisol regulates the migration life-history in juvenile brown trout, 37 but intermediate increases in baseline GC (and not post-stressor GC) levels may favor migration 38 39 performance.

41 Key Words

42 Cortisol, Fish, Glucocorticoid, Migration, Stress response, Brown trout, Salmo trutta

43

44 Abbreviations

- 45 GC, glucocorticoid; PIT, passive integrated transponder; S1, tracking station 1; S2, tracking
- 46 station 2; HPA, hypothalamic–pituitary–adrenal axis; HPI, hypothalamic–pituitary–interrenal
- 47 axis; K, Fulton's condition factor.

49 The study of the physiological basis of stress is useful for understanding physical, chemical, 50 social and perceived challenges faced by organisms, and the functional responses employed by organisms to deal with those stressors (Denver 2009). In vertebrates, real or perceived 51 52 stressors stimulate the hypothalamic-pituitary-adrenal axis (HPA; or the hypothalamicpituitary-interrenal (HPI) axis in fish) resulting in the release of glucocorticoid (GC) hormones 53 into the blood stream (Sapolsky et al. 2000; Barton 2002). Thus, circulating concentrations of 54 55 GC hormones are often measured to understand an organism's stress response (Dantzer et al. 2014). Within a species, individuals often show repeatable baseline (prior to a stressor) GC 56 levels and a repeatable GC response following a stressor (Pottinger and Carrick 2001; 57 58 Ouedraogo et al. 2004; Wada et al. 2008; Rensel and Schoech 2011; Cook et al. 2012; Narayan et al. 2013; Love et al. 2015) resulting in different reactive scope profiles (difference between 59 60 GC response and baseline) between individuals (Romero et al. 2009). The biological 61 functionality, as well as the individual and evolutionary consequences of such variability among individua, are still debated (Breuner et al. 2008; Bonier et al. 2009). 62

To define the biological consequences of individual variation in GC levels during the stress
response, researchers have attempted to relate inter-individual differences in baseline and
post-stressor GC concentrations to performance (e.g., migration, parental care, territorial
behavior) and fitness (e.g. Comendant et al. 2003; Cabezas et al. 2007; Breuner et al. 2008).
From a theoretical point of view, it is often hypothesized that individuals with greater poststressor GC levels should have increased long term fitness (Breuner et al. 2008), because an

individual's stress response increases chances of surviving the stressor (e.g. predation attempt) 70 71 and mediates trade-offs between reproduction and survival. Indeed, experimental studies show variability in the relationship between the magnitude of the stress response and performance 72 73 or fitness measures (reviewed by Breuner et al. 2008). This relationship is increasingly 74 understood as context dependent, because an elevated stress response in an emergency situation shifts resources from reproduction to survival (Wingfield et al. 1998); however, an 75 76 elevated stress response when the perception of stressors is exaggerated can lead to the 77 misallocation of resources, thereby limiting investment in growth and reproduction (Hawlena and Schmitz 2010). There is a paucity of studies directly examining inter-individual differences 78 79 in stress response while concurrently measuring performance or fitness (Romero and Wikelski 2001; Blas et al. 2007; Cook et al. 2011). 80

82 In contrast to the theorized positive association between elevated post-stressor GC levels and performance or fitness, higher baseline GC levels are often inferred to indicate reduced 83 condition, because it suggests an organism is experiencing greater environmental stressors 84 (Cort-Fitness hypothesis; reviewed by Bonier et al. 2009). Higher GC levels are in circulation 85 when exposed to a stressor and are associated with the reallocation of resources from fitness-86 87 related factors, such as growth and reproduction, towards the restoration of physiological homeostasis. Despite the potential negative consequences associated with higher baseline GC 88 levels, some studies have found that moderate long term increases in GCs can actually increase 89 90 individual performance (Comendant et al. 2003; Pravosudov 2003; Cabezas et al. 2007). Further

Page 6 of 29

91 investigation is therefore warranted to understand the relationship between baseline GC
92 hormones and performance.

Both baseline and post-stressor GC concentrations have a heritable component in some species 94 95 (Evans et al. 2006; Jenkins et al. 2014) and are associated with a suite of other traits that can lead to different life history and coping strategies (Koolhaas et al. 1999; Pottinger and Carrick 96 97 1999; Blas et al. 2007; Øverli et al. 2007; Wada et al. 2008). Selection on heritable variation in 98 GC levels and response is thought to drive the evolution of life-history traits (Bolnick et al. 2003; Williams 2008), such as migration behavior. Migration in particular, requires large amounts of 99 energy and often exposes individuals to many stressors including predation and changes in 100 environmental condition (Dingle 2014). Thus, GC levels have the potential to be an important 101 102 factor determining successful migration (Holberton 1999; Piersma et al. 2000; Angelier et al. 103 2009), which could affect fitness and potentially drive the evolution of migration strategies. Migration success, meaning successfully reaching the migration destination, has been linked to 104 105 GC levels in a single species of fish, adult sockeye salmon (Oncorhynchus nerka (Walbaum, 106 1792)), where upriver migration success was found to increase with stress responsiveness but 107 not baseline stress levels (Cook et al. 2014). GC levels have also been related to the 108 smoltification process in anadromous salmonids. Smolting fish generally have higher baseline and peak GC, which appears to induce the physiological changes required to cope with the 109 transition from freshwater to saltwater (e.g., increased osmoregularity capacity; Barton et al. 110 111 1985; Redding et al. 1991; Björnsson et al. 2011). Stressors that cause a GC response may cause a reallocation of energy towards immediate survival resulting in a change in life history and 112

134

coping strategy (Landys et al. 2006; Shepard et al. 2008). Thus, GC variation resulting from past
or current conditionsmay influence the "decision" to migrate. Alternative migration strategies
have the potential to be heritable through heritable GC regulation (Zera and Harshman 2001;
Williams 2008).

For our study species, brown trout (Salmo trutta (Linnaeus, 1758)), migration success provides a 118 119 measure of performance, and also acts as an indicator of alternate life history strategies. More specifically, European populations of brown trout show a continuum of migration strategies, 120 where subpopulations are capable of anadromous activity, smaller scale migrations within 121 122 freshwater systems (potamodromous), and residency in natal streams (Cucherousset et al. 2005). This phenomenon is termed partial migration and reflects the fact that only a 123 124 component of the population engages in migration, a common trait among some salmonid 125 species (Jonsson and Jonsson 1993). Previous studies on the propensity of migration in brown trout have found that individuals with high metabolisms and high growth rates tend to migrate, 126 127 likely due to higher resource requirements (onsson and Jonsson 1993; Forseth et al. 1999). 128

The goal of this study is to define relationships between GC levels (baseline and post-stressor), and migration propensity in juvenile brown trout. Given the role of GCs in smoltification, we would expect to see higher post-stress and baseline GC levels in migrating individuals. The effects of individual body condition and size (length, mass) in relation to migration propensity will also be quantified.

Page 8 of 29

135 Methods

136 Experimental Design

137 Brown trout were captured using backpack electroshocking (Scubla ELT 60 II GI; run at 300 138 volts) in four regions of the Gudsø Stream, Jutland, Denmark (Figure 1) between 11 March and 139 18 March 2013. This time period was selected as it fell prior to the spring migration, which typically occurs in mid-April (Midwood et al. 2014). Individuals between 120 and 182 mm in 140 141 total length (TL) were used to minimize the effects of allometry on results. To evaluate an individual's resting plasma cortisol levels (baseline), blood was collected (~0.2-0.3 ml) from a 142 143 subset of fish within 3 min of capture via caudal puncture of the haemal arch using a 1 ml 144 syringe and 25-gauge needle. A separate subset of fish (Table 1) were exposed to a 145 standardized 3 minute air exposure and then held in a 20 L container with 5 L of freshwater for 146 30 min prior to the collection of a post-stressor blood sample. Blood was sampled in the same 147 manner as above at 30 ± 3 minutes post-air exposure corresponding with the approximate peak in GCs in brown trout (Pickering and Pottinger 1989). Blood samples were held in a water-ice 148 149 slurry for no more than 2 hrs until centrifuged for 5 min (10,000 rpm) to isolate plasma. Plasma 150 isolate was decanted, flash frozen and stored (-80°C) for later analysis of plasma cortisol 151 concentrations. Total length and wet mass of each individual were then measured. Next, a 152 uniquely coded 23 mm PIT tag (RI-TRP-RRHP, half duplex, 134 kHz, diameter 3.85 mm and a mass of 0.6 g in air; Texas Instruments, Plano, Texas, USA) was inserted into the body cavity 153 through a small incision made with a scalpel. Previous tag retention studies on salmonids have 154 155 found negligible effects on survival and growth (Larsen et al. 2013) and generally low PIT tag 156 rejection rates (< 4% after seven months; Ombredane et al. 1998). Following tagging fish were

then placed in a 60 L container of fresh river water and monitored for a minimum of 30 minutes to ensure recovery before they were ultimately released back into the stream near their point of capture. All fish recovered. Length and mass were used to calculate Fulton's condition factor (*K*) for each sampled fish where *K* = mass (g) × 100 / length³ (cm) (Ricker 1975).

162 Cortisol Analysis

Plasma cortisol (ng mL⁻¹) was quantified using a commercially available enzyme linked
 immunosorbant assay (ELISA) (Enzo Life Sciences, Cortisol EIA Kit [901-071], BioAssay Systems)
 previously validated for use in fishes (Sink et al. 2008). This assay has a detection limit of 0.0567
 ng mL⁻¹, therefore, the 5 samples with undetectable cortisol concentrations were assigned
 values equal to this detection limit (Haddy and Pankhurst 1999; Liss et al. 2013).

168

161

169 Tracking

To track movements of individuals from their point of release to the sea, two tracking stations 170 were placed approximately 1 km upstream of Kolding Fjord on Gudsø Stream, which was the 171 closest feasible place to the ocean. The first station (S1) was placed upstream of a millpond and 172 the second station (S2) was placed downstream of the millpond, approximately 150 m from S1. 173 174 Each station was comprised of a paired set of PIT gates spaced approximately 10-m apart. This allowed for an assessment of the direction of movement for individuals at each station. The 175 same array was used by Midwood et al. (2015) and found to have a detection efficiency at S1 of 176 96.3%; detection efficiency could not be evaluated at S2, but missed-detections were assumed 177

Page 10 of 29

to affect all trout equally and therefore should not bias conclusions related to baseline GC or
 post-stressor GC correlations with relative migration success to S1 and S2.

180

181 To estimate the proportion of individuals that stayed resident and did not migrate to the ocean, 182 the stream was re-sampled using back-pack electrofishing on 18 and 19 June, 2013, at which 183 point migratory individuals should have left the stream. Surveys started near the antenna and 184 proceeded upstream through all areas where brown trout were initially captured (Figure 2). It 185 was not possible to resample the entire system as it is greater than 16 km in length; however, given the shallow and narrow nature of the stream and the relatively high efficiency of capture 186 187 for salmonid species using electrofishing (Kennedy and Strange 1981; Haynes and Baird 1994), 188 efforts were likely sufficient to provide a relative estimate of residency.

189

190 Statistical Analyses

Multiple linear regression was used to quantify relationships between plasma cortisol 191 192 concentrations (for both post-stressor and baseline groups) and timing of migration (time from 193 sampling to S1 passage), while controlling for K, length or mass. Assumption of homogeneity of 194 variance and normal distribution for the models were ensured by visual inspection of residual 195 plots. If violated, the response variable was log or square root transformed. Multiple binomial logistic regression was used to test for the effect of plasma cortisol concentration (baseline and 196 post-stressor) on the probability of successful migration to S1 (used as a proxy of migration 197 strategy) and on the probability of successful migration to S2 given successful passage of S1 198 199 (used as a measure of migration success through the millpond for migrating individuals). Again,

separate tests were conducted using mass, length or *K* as a covariate in the model. Length,
mass and *K* were also tested separately for effects on all migration response variables with the
baseline and post-stressor fish subsets pooled. Two outliers were removed before analyses as
they were outside the standard deviation of the mean: a baseline cortisol value of 44.5ng/mL
and a peak cortisol value of 3.43ng/mL. All significant results found without the outliers were
also found when the outliers were included in the analyses.

206

207 Variation in association with recorded mean values is given as standard error (\pm SE) throughout. 208 Statistical significance for all analyses was set at α = 0.05 and all statistical analyses were 209 conducted in R (v 3.2.2; R Core Team, 2016).

210

211 Results

212 Of the 198 fish tagged, 19 were recaptured in mid-June, 83 were detected passing S1 and 45 passing S2 (Table 2). All fish detected at S2 were also detected at S1, suggesting a detection 213 efficiency of 100% at S1. Mean baseline cortisol was 5.6 ± 6.7 ng/mL and post-stressor cortisol 214 was 69.2 \pm 40.1 ng/mL (Table 1). Post-stressor (Multiple logistic regression: $Z_{2,96}$ = 1.44, p = 215 0.15) and baseline (Multiple logistic regression: $Z_{2,96} = 0.15$, p = 0.89) plasma cortisol 216 217 concentrations did not have a significant effect on the probability of successful passage at S1 when controlling for K, mass, or length (p>0.1; Figure 2b and 2e). For fish that passed S1, the 218 219 probability of successful passage to S2 increased with increasing individual baseline cortisol when controlling for mass (Multiple logistic regression: $Z_{2,27} = 1.96$, p = 0.05) and length 220 (Multiple logistic regression: $Z_{2,27}$ = 2.01, p = 0.045), but not for K (p>0.05). For fish that passed 221

S1, the probability of successful passage to S2 showed no relationship with post stressor
cortisol (All: *p*>0.1)

224

Mean migration time from blood sampling to S1 passage was 33.4±19.1 days and mean time from S1 to S2 was 3.2±7.6 days. Multiple regression showed no significant relationship between migration duration or date of passage, and post-stressor or baseline cortisol when controlling for mass, length or *K* (All: *p*>0.1). Migration time decreased with increasing fish length (Linear regression: $F_{1,79}$ = 6.82, p = 0.01, R^2 = 0.8) and fish mass (Linear regression: $F_{1,79}$ = 4.4, p = 0.04, R^2 = 0.05), but no relationship was found with *K* (Linear regression: $F_{1,79}$ = 3.14, p = 0.08, R^2 = 0.04).

232

Multiple logistic regression revealed no effect of post-stressor and baseline cortisol on the probability of recapture in resident streams when controlling for K, length or mass (Figure 2; All: p>0.1). Length, mass and K also were found to have no effect on the probability of recapture in resident streams (All: Logistic regression: p>0.1). No relationship was found between K, mass or length and baseline or post-stressor cortisol (All: Linear regression: p>0.1). The probability of S1 passage and the probability of passage between S1 and S2 had no relationship with K, fish length and fish mass (All: p>0.08).

240

241 Discussion

As previous studies have found cortisol to be elevated during smoltification (Barton et al. 1985), we expected individual brown trout with higher baseline and post-stressor cortisol to have a higher probability of migrating downstream to S1 and S2, coupled with a lower probability of
recapture as resident. However, no such relationships were found.

246

247 We found no evidence suggesting that migration is correlated with endocrine regulation in 248 juvenile brown trout, despite several studies having documented that cortisol levels rise during 249 smoltification in andromous fish (Barton et al. 1985; Redding et al. 1991; Björnsson et al. 2011), 250 and also that GC regulation has the potential to play a role in determining alternative coping 251 strategies (Zera and Harshman 2001; Landys et al. 2006; Williams 2008). Mass, length and body 252 condition were also not found to have a relationship with plasma cortisol (baseline or post-253 stressor). Fish of greater mass and length (but not condition) were found to have shorter migration times similar to previous findings (Bohlin et al. 1996). Body condition, length and 254 255 mass were not found to affect migration propensity to S1. the lack of correlation between our 256 condition metrics and migration propensity is counter to previous findings of reduced condition leading to increased migration propensity (Peiman et al. 2017); however, it is possible that the 257 258 comparatively limited recapture rate in the present study (N=19) prevented the detection of a 259 size-based effect.

260

Baseline, but not post-stressor, cortisol concentration was found to be positively associated with passage through the millpond (i.e., from S1 to S2). Previous studies have found brown trout baseline cortisol without chronic stressors to be 0-5 ng/mL, while those exposed to chronic stressors such as confinement or crowding have shown baseline cortisol elevated to 10 ng/mL (Pickering and Pottinger 1989). The average baseline cortisol found in individuals

recaptured in resident streams, individuals that passed S1 and individuals that passed S2 was
5.6 ng/mL, 5.8 ng/mL and 8.4 ng/mL, respectively (Table 2). Thus, individuals that passed
through the millpond had baseline cortisol levels of individuals approaching chronically stressed
levels on average, while other fish had cortisol levels closer to what would be expected for fish
not showing chronic stress.

272 The distance between S1 and S2 is only 150 m, thus it is unlikely that S1 versus S2 passage 273 would be indicative of migration strategy. The millpond may therefore be an area of increased 274 mortality, suggesting S1 passage is more indicative of migration strategy (resident and 275 potamodromous versus anadromous) while passage between S1 and S2 is more indicative of migration success of migrant individuals. Forty five percent of all brown trout that passed S1 276 277 were not detected passing S2, which is only 150 meters downstream, and those that did 278 migrate through the millpond, took much longer than when migrating through the stream, taking an average of 3.2 days to move 150 meters. Similar patterns are found in other studies of 279 migrating fish at Gudso (Midwood et al. 2014, 2015). This indicates the millpond was 280 281 challenging for migrating trout. We suggest that many of these individuals likely suffered 282 mortality, as it is unlikely for such a large proportion of migrants to take up residency in such a 283 small area. Furthermore, there is anecdotal evidence of high concentration of foraging grey heron (Ardea cinerea Linnaeus, 1758) and other birds at the outflow of millpond. Previous 284 285 studies on salmonid smolts have documented large losses resulting from predation in lakes (Jepsen et al. 1998; Koed et al. 2006) and at weirs and related millponds (Aarestrup and Koed 286 287 2003). For example, Jepsen et al. (1998) found 90% of smolts died while passing through Lake

Tange, Denmark, with 56 % of the mortality cause by pike (Esox lucius (Linnaeus, 1758)) and 288 289 31% by birds. Another possibility for the reduced passage and long migration time through the 290 millpond is that there is only one small exit and trout may have had a hard time getting out. While it is possible that overall lower passage at S2 is driven by reduced detection efficiency at 291 292 this station, these missed-detections would affect all trout equally and thus would not affect 293 the relationship found between baseline cortisol and passage between S1 and S2 other than by 294 reducing statistical power. Our results, therefore, do not support the link between GC levels 295 and migration strategy in brown trout, but they do suggest that baseline GC (and not post-296 stressor GC) is positively associated with migration success.

297

298 Generally it is hypothesized that increased baseline GC are associated with decreased 299 survivorship because increased baseline GC are associated with increased chronic stressors 300 (Cort-Fitness hypothesis: Bonier et al. 2009). Our findings appear to support the opposite trend as migratory fish with lower baseline cortisol appear to have lower successful passage through 301 302 the millpond (Figure 2). Several other studies have also found higher baseline cortisol to be 303 correlated with increased survivorship: Cabezas et al. (2007) found moderately elevated corticosterones were associated with decreased body condition but increased probability of 304 305 survival in European wild rabbits (Oryctolagus cuniculus (Linnaeus, 1758)); Comendant et al. (2003) found higher corticosterone in female lizards (Uta stansburiana (Baird and Girard, 1852)) 306 307 to be associated with increased survival to second ovulation; and Cote et al. (2006) found increased corticosterone to modify behaviour (locomotor and social activity), increasing survival 308 309 in male adult common lizards (Lacerta vivipara (Jacquin, 1787)).

310 Another possible explanation for why our results are not consistent with the Cort-Fitness 311 hypothesis is that it is difficult to discern whether higher baseline cortisol indicates exposure of the individual to greater chronic stress (which is assumed to be the case by the Cort-Fitness 312 313 hypothesis; Bonier et al. 2009), or indicates a more competent ability to cope with chronic 314 stressors. The first would likely lead to decreased survivorship with increasing baseline GC level, while the later would likely result in the opposite trend. In our study, all individuals may be 315 316 subjected to similar chronic stressors as they were all reared in the same stream reach. Thus, 317 baseline stress may be a better indicator of better ability to cope with chronic stressors than 318 exposure to greater environmental stressors.

319

320 Post-stressor cortisol, a proxy for stress responsiveness, was not found to be a significant 321 predictor of migration timing, survival or success. Potentially, the effect of stress 322 responsiveness was masked, because we were not able to control for individual variability in baseline as fish were too small to obtain both a baseline and a post-stressor cortisol sample. 323 324 However post-stressor cortisol was more variable (SD=40.7 ng/mL) than baseline cortisol 325 (SD=7.7 ng/mL). Cockrem (2013) reviewed studies on baseline and stress-induced GCs and 326 determined that there tended to be more variation in baseline GCs among individuals than in 327 stress-induced levels. Although our findings are not consistent with that generalization, Cockrem (2013) did note that it was not ubiquitous with a number of exceptions. It is unclear 328 why that pattern was observed here but reasons, among others, could include inter-individual 329 differences in how the stressor was perceived or different stress histories. Despite the widely 330 331 accepted belief that acute stress responses are adaptive, similar to our study several other

studies have failed to find a relationship between stress responsiveness and performance 332 333 (Romero and Wikelski 2001; Blas et al. 2007), or may even reduce performance. These results may be an indication that different reactive scopes profiles may allow individuals to adequately 334 cope with stressors. Additionally, the benefits of stress responsiveness are increasingly viewed 335 336 as context dependent, as traits induced by a stress response can have variable consequences to 337 an organisms' performance depending on the nature of the stressor (Breuner et al. 2008). 338 339 Similar to our findings in brown trout, the only other study we are aware of that examines 340 individual variation in endocrine regulation in relation to migration in fish (anadromous 341 semelparous sockeye salmon) found no correlation between GC levels and migration timing or rate. They also found individuals with higher stress responsiveness to have greater migration 342 343 success, but that baseline cortisol not to be a significant predictor of migration success (Cook et 344 al. 2014). Differences in the relationship between GC secretion and migration success found by our study may be attributed to differences in the stressors experienced. For example, salmon 345 may have more chronic stressors such that their baseline levels are at the turning point of the 346 baseline cortisol-survivorship curve hypothesized by Cabezas et al. (2007), where differences in 347 baseline levels have negligible effects on survivorship. The inconsistency between findings may 348 also be reflective of differences in physiology between species. 349

351

350

352 Conclusions

In summary, we found no evidence suggesting that cortisol regulates the migration life history trade off in juvenile brown trout. But, slightly higher levels of baseline GC (and not post-stressor GC) may favor migration performance. Further study is warranted to help determine the relationship between migration success and GC, with special focus on stress responsiveness within an individual.

360 Acknowledgements

This project was funded through an NSERC Discovery Grant awarded to S.J.C. as well as grants to the Technical University of Denmark from the Danish National Fishing License Funds, and the strategic project SMOLTPRO, financed by the Swedish Research Council Formas. S.J.C. is further supported by the Canada Research Chairs Program. The authors thank Michael Holm, Hans-Jørn Christensen, Jørgen Skole Mikkelsen, and Morten Carøe for assistance in the field, Henrik Baktoft for technical support, as well as Tanya Prystay and Michael Lawrence for assistance with editing the manuscript.

368

358

359

369 References

Aarestrup, K., and Koed, A. 2003. Survival of migrating sea trout (Salmo trutta) and Atlantic
 salmon (Salmo salar) smolts negotiating weirs in small Danish rivers. Ecol. Freshw. Fish, 12:
 169–176.

Angelier, F., Holberton, R.L., and Marra, P.P. 2009. Does stress response predict return rate in a
 migratory bird species? A study of American redstarts and their non-breeding habitat.
 Proc. Biol. Sci. 276(1672): 3545–3551. doi:10.1098/rspb.2009.0868.

	376 377	Barton, B.A. 2002. Stress in fishes: a diversity of responses with particular reference to changes in circulating corticosteroids. Integr. Comp. Biol. 42 (3): 517–525. doi:10.1093/icb/42.3.517.
	378 379 380	Barton, B.A., Schreck, C.B., Ewing, R.D., Hemmingsen, A.R., and Patiño, R. 1985. Changes in plasma cortisol during stress and smoltification in Coho Salmon, Oncorhynchus kisutch. Gen. Comp. Endocrinol. 59 (3): 468–471. doi:10.1016/0016-6480(85)90406-X.
	381 382 383	Björnsson, B.T., Stefansson, S.O., and McCormick, S.D. 2011. Environmental endocrinology of salmon smoltification. Gen. Comp. Endocrinol. 170 (2): 290–298. doi:10.1016/j.ygcen.2010.07.003.
•	384 385 386	Blas, J., Bortolotti, G.R., Tella, J.L., Baos, R., and Marchant, T. a. 2007. Stress response during development predicts fitness in a wild, long lived vertebrate. Proc. Natl. Acad. Sci. U. S. A. 104 (21): 8880–8884. doi:10.1073/pnas.0700232104.
)	387 388	Bohlin, T., Dellefors, C., and Faremo, U. 1996. Date of smolt migration depends on body size but not age in wild sea-run brown trout. J. Fish Biol. 49 (1): 157–164.
)	389 390 391	Bolnick, D.I., Svanbäck, R., Fordyce, J. A, Yang, L.H., Davis, J.M., Hulsey, C.D., and Forister, M.L. 2003. The ecology of individuals: incidence and implications of individual specialization. Am. Nat. 161 (1): 1–28. doi:10.1086/343878.
•	392 393	Bonier, F., Martin, P.R., Moore, I.T., and Wingfield, J.C. 2009. Do baseline glucocorticoids predict fitness? Trends Ecol. Evol. 24 (11): 634–642. doi:10.1016/j.tree.2009.04.013.
1	394 395 396	Breuner, C.W., Patterson, S.H., and Hahn, T.P. 2008. In search of relationships between the acute adrenocortical response and fitness. Gen. Comp. Endocrinol. 157 (3): 288–295. doi:10.1016/j.ygcen.2008.05.017.
4	397 398 399	Cabezas, S., Blas, J., Marchant, T. A., and Moreno, S. 2007. Physiological stress levels predict survival probabilities in wild rabbits. Horm. Behav. 51 (3): 313–320. doi:10.1016/j.yhbeh.2006.11.004.
	400 401	Cockrem, J.F. 2013. Individual variation in glucocorticoid stress responses in animals. Gen. Comp. Endocrinol. 181 (1): 45–58. doi:10.1016/j.ygcen.2012.11.025.
	402 403 404	Comendant, T., Sinervo, B., Svensson, E.I., and Wingfield, J. 2003. Social competition, corticosterone and survival in female lizard morphs. J. Evol. Biol. 16 (5): 948–955. doi:10.1046/j.1420-9101.2003.00598.x.

405	Cook, K. V., Crossin, G.T., Patterson, D. A., Hinch, S.G., Gilmour, K.M., and Cooke, S.J. 2014. The			
406	stress response predicts migration failure but not migration rate in a semelparous fish.			
407	Gen. Comp. Endocrinol. 202 : 44–49. doi:10.1016/j.ygcen.2014.04.008.			
408	Cook, K. V., O and Connor, C.M., McConnachie, S.H., Gilmour, K.M., and Cooke, S.J. 2012.			
409	Condition dependent intra-individual repeatability of stress-induced cortisol in a			
410	freshwater fish. Comp. Biochem. Physiol. A. 161 (3): 337–343.			
411	doi:10.1016/j.cbpa.2011.12.002.			
412	Cook, K. V, McConnachie, S.H., Gilmour, K.M., Hinch, S.G., and Cooke, S.J. 2011. Fitness and			
413	behavioral correlates of pre-stress and stress-induced plasma cortisol titers in pink salmon			
414	(Oncorhynchus gorbuscha) upon arrival at spawning grounds. Horm. Behav. 60 : 489–497.			
415	Cote, J., Clobert, J., Meylan, S., and Fitze, P.S. 2006. Experimental enhancement of			
416	corticosterone levels positively affects subsequent male survival. Horm. Behav. 49(3): 320-			
417	327. doi:10.1016/j.yhbeh.2005.08.004.			
418	Cucherousset, J., Ombredane, D., Charles, K., Marchand, F., and Bagliniere, J.L. 2005. A			
419	Continuum of Life History Tactics in a Brown Trout Salmo Trutta Population. Can. J. Fish.			
420	Aquat. Sci. 1610 : 1600–1610. doi:10.1139/f05-057.			
421	Dantzer, B., Fletcher, Q., Boonstra, R., and Sheriff, M. 2014. Measures of physiological stress: a			
422	transparent or opaque window into the status, management and conservation of species?			
423	Conserv. Physiol. 2(May): cou023. doi:10.1093/conphys/cou023.			
424	Denver, R.J. 2009. Structural and functional evolution of vertebrate neuroendocrine stress			
425	systems. Ann. N. Y. Acad. Sci. 1163 : 1–16. doi:10.1111/j.1749-6632.2009.04433.x.			
426	Dingle, H. 2014. Migration: the biology of life on the move. Oxford University Press, USA.			
427	Evans, M.R., Roberts, M.L., Buchanan, K.L., and Goldsmith, A.R. 2006. Heritability of			
428	corticosterone response and changes in life history traits during selection in the zebra			
429	finch. J. Evol. Biol. 19 (2): 343–352. doi:10.1111/j.1420-9101.2005.01034.x.			
430	Forseth, T., Næsje, T.F., Jonsson, B., and Hårsaker, K. 1999. Juvenile migration in brown trout: A			
431	consequence of energetic state. J. Anim. Ecol. 68 (4): 783–793. doi:10.1046/j.1365-			
432	2656.1999.00329.x.			
433	Haddy, J.A., and Pankhurst, N.W. 1999. Stress-induced changes in concentrations of plasma sex			
434	steroids in black bream. J. Fish Biol. 55 : 1304–1316.			

Page 21 of 29

435	Hawlena, D., and Schmitz, O.J. 2010. Physiological Stress as a Fundamental Mechanism Linking
436	Predation to Ecosystem Functioning. Am. Nat. 176 (5): 537–556. doi:10.1086/656495.
437	Haynes, J.W., and Baird, D.B. 1994. Estimating relative abundance of juvenile brown trout in
438	rivers by underwater census and electrofishing. N.Z. J. Mar. Freshw. Res. 28 (3): 243–253.
439	doi:10.1080/00288330.1994.9516612.
440	Holberton, R.L. 1999. Changes in patterns of corticosterone secretion concurrent with
441	migratory fattening in a neotropical migratory bird. Gen. Comp. Endocrinol. 116 (1): 49–58.
442	doi:10.1006/gcen.1999.7336.
443	Jenkins, B.R., Vitousek, M.N., Hubbard, J.K., and Safran, R.J. 2014. An experimental analysis of
444	the heritability of variation in glucocorticoid concentrations in a wild avian population.
445	Proc. R. Soc. Lond. B Biol. Sci. 281 (1790). Available from
446	http://rspb.royalsocietypublishing.org/content/281/1790/20141302.abstract.
447	Jepsen, N., Aarestrup, K., Økland, F., and Rasmussen, G. 1998. Survival of radio-tagged Atlantic
448	salmon (Salmo salar L .) and trout (Salmo trutta L .) smolts passing a reservoir during
449	seaward migration. Hydrobiologia, 371/372 (v-vii): 347–353.
450	Jonsson, B., and Jonsson, N. 1993. Partial migration: niche shift versus sexual maturation in
451	fishes. Rev. Fish Biol. Fish. 3 (4): 348–365. doi:10.1007/BF00043384.
452	Kennedy, G.J.A., and Strange, C.D. 1981. Efficiency of Electric Fishing for Salmonids in Relation
453	to River Width. Aquacult. Res. 12 (2): 55–60. doi:10.1111/j.1365-2109.1981.tb00010.x.
454	Koed, A., Baktoft, H., and Bak, B.D. 2006. Causes of mortality of Atlantic salmon (Salmo salar)
455	and brown trout (Salmo trutta) smolts in a restored river and its estuary. River Res.
456	Appl. 22 (1): 69–78. doi:10.1002/rra.894.
457	Koolhaas, J.M., Korte, S.M., De Boer, S.F., Van Der Vegt, B.J., Van Reenen, C.G., Hopster, H., De
458	Jong, I.C., Ruis, M. a W., and Blokhuis, H.J. 1999. Coping styles in animals: Current status in
459	behavior and stress- physiology. Neurosci. Biobehav. Rev. 23 (7): 925–935.
460	doi:10.1016/S0149-7634(99)00026-3.
161	Landys M.M. Ramonofsky M. and Wingfield J.C. 2006 Actions of alucocorticoids at a

Landys, M.M., Ramenofsky, M., and Wingfield, J.C. 2006. Actions of glucocorticoids at a
seasonal baseline as compared to stress-related levels in the regulation of periodic life
processes. Gen. Comp. Endocrinol. 148(2): 132–149. doi:10.1016/j.ygcen.2006.02.013.

Larsen, M.H., Thorn, A.N., Skov, C., and Aarestrup, K. 2013. Effects of passive integrated
 transponder tags on survival and growth of juvenile Atlantic salmon Salmo salar Effects of
 passive integrated transponder tags on survival and growth of juvenile Atlantic salmon
 Salmo salar. Anim. Biotelemetry. 1: 1–7.

Liss, S. A., Sass, G.G., and Suski, C.D. 2013. Spatial and temporal influences on the physiological condition of invasive silver carp. Conserv. Physiol. **1**: 1–13. doi:10.1093/conphys/cot017.

Love, O.P., Shutt, L.J., Silfies, J.S., and Bird, D.M. 2015. Repeated restraint and sampling results
 in reduced corticosterone levels in developing and adult captive American kestrels (Falco sparverius). Physiol. Biochem. Zool. **76**(5): 753–761. doi:10.1086/376431.

Midwood, J.D., Larsen, M.H., Boel, M., Aarestrup, K., and Cooke, S.J. 2015. An experimental
 field evaluation of winter carryover effects in semi-anadromous brown trout (Salmo
 trutta). J. Exp. Zool. A. 323(9): 645–654. doi:10.1002/jez.1955.

Midwood, J.D., Larsen, M.H., Boel, M., Jepsen, N., Aarestrup, K., and Cooke, S.J. 2014. Does
cortisol manipulation influence outmigration behaviour, survival and growth of sea trout?
a field test of carryover effects in wild fish. Mar. Ecol. Prog. Ser. 496: 135–144.
doi:10.3354/meps10524.

Narayan, E.J., Cockrem, J.F., and Hero, J.M. 2013. Repeatability of baseline corticosterone and
 short-term corticosterone stress responses, and their correlation with testosterone and
 body condition in a terrestrial breeding anuran (Platymantis vitiana). Comp. Biochem.
 Physiol. A. 165(2): 304–312. Elsevier Inc. doi:10.1016/j.cbpa.2013.03.033.

Ombredane, D., Bagliniere, J.L., and Marchand, F. 1998. The effects of Passive Integrated
 Transponder tags on survival an growth of juvenile brown trout (Salmo trutta) and their
 use for studing movements in a small river. Hydrobiologia, **371**: 99–106.
 doi:10.1023/A:101702202

Ouedraogo, R.M., Goettel, M.S., and Brodeur, J. 2004. Behavioral thermoregulation in the
 migratory locust: a therapy to overcome fungal infection. Oecologia, **138**(2): 312–319.
 doi:10.1007/s00442-003-1431-0.

Øverli, Ø., Sørensen, C., Pulman, K.G.T., Pottinger, T.G., Korzan, W., Summers, C.H., and Nilsson,
 G.E. 2007. Evolutionary background for stress-coping styles: Relationships between
 physiological, behavioral, and cognitive traits in non-mammalian vertebrates. Neurosci.
 Biobehav. Rev. **31**(3): 396–412. doi:10.1016/j.neubiorev.2006.10.006.

495 Peiman, K.S., Birnie-Gauvin, K., Midwood, J.D., Larsen, M.H., Wilson, A.D.M., Aarestrup, K., and 496 Cooke, S.J. 2017. If and when: intrinsic differences and environmental stressors influence 497 migration in brown trout (Salmo trutta). Oecologia, 184(2): 375–384. doi:10.1007/s00442-498 017-3873-9. 499 Pickering, a. D., and Pottinger, T.G. 1989. Stress responses and disease resistance in salmonid 500 fish: Effects of chronic elevation of plasma cortisol. Fish Physiol. Biochem. 7(1–6): 253–258. 501 doi:10.1007/BF00004714. Piersma, T., Reneerkens, J., and Ramenofsky, M. 2000. Baseline corticosterone peaks in 502 503 shorebirds with maximal energy stores for migration: a general preparatory mechanism for 504 rapid behavioral and metabolic transitions? Gen. Comp. Endocrinol. 120(1): 118–126. 505 doi:10.1006/gcen.2000.7543. 506 Pottinger, T.G., and Carrick, T.R. 1999. A comparison of plasma glucose and plasma cortisol as 507 selection markers for high and low stress-responsiveness in female rainbow trout. 508 Aguaculture, 175(3-4): 351-363. doi:10.1016/S0044-8486(99)00107-6. 509 Pottinger, T.G., and Carrick, T.R. 2001. Stress responsiveness affects dominant-subordinate 510 relationships in rainbow trout. Horm. Behav. 40(3): 419-427. 511 doi:10.1006/hbeh.2001.1707. Pravosudov, V. V. 2003. Long-term moderate elevation of corticosterone facilitates avian food-512 caching behaviour and enhances spatial memory. Proc. Biol. Sci. 270(1533): 2599-2604. 513 514 doi:10.1098/rspb.2003.2551. R Core Team. 2016. R: A language and environment for statistical computing. R Foundation for 515 Statistical Computing, Vienna, Austria. Available from https://www.r-project.org/. 516 Redding, J.M., Patiño, R., and Schreck, C.B. 1991. Cortisol effects on plasma electrolytes and 517 thyroid hormones during smoltification in coho salmon Oncorhynchus kisutch. Gen. Comp. 518 519 Endocrinol. 81(3): 373-382. doi:10.1016/0016-6480(91)90164-2. 520 Rensel, M. A., and Schoech, S.J. 2011. Repeatability of baseline and stress-induced 521 corticosterone levels across early life stages in the Florida scrub-jay (Aphelocoma 522 coerulescens). Horm. Behav. 59(4): 497-502. doi:10.1016/j.yhbeh.2011.01.010. 523 Ricker, W.E. 1975. Computation and interpretation of biological statistics of fish

populations. In Bull. Fish. Res. Board Can. No. 191., Ottawa, Canada. p. 382.

525 Romero, L.M., Dickens, M.J., and Cyr, N.E. 2009. The reactive scope model - A new model 526 integrating homeostasis, allostasis, and stress. Horm. Behav. 55(3): 375–389. 527 doi:10.1016/j.yhbeh.2008.12.009. 528 Romero, L.M., and Wikelski, M. 2001. Corticosterone levels predict survival probabilities of 529 Galapagos marine iguanas during El Nino events. Proc. Natl. Acad. Sci. U. S. A. 98(13): 530 7366-7370. doi:10.1073/pnas.131091498. Sapolsky, R.M., Romero, L.M., and Munck, a. U. 2000. How Do Glucocorticoids Influence Stress 531 Responses ? Preparative Actions *. Endocr. Rev. 21(April): 55-89. doi:10.1210/er.21.1.55. 532 533 Shepard, D.B., Kuhns, A.R., Dreslik, M.J., and Phillips, C.A. 2008. Roads as barriers to animal movement in fragmented landscapes. Anim. Conserv. 11(4): 288-296. doi:10.1111/j.1469-534 535 1795.2008.00183.x.

Sink, T.D., Lochmann, R.T., and Fecteau, K.A. 2008. Validation, use, and disadvantages of
enzyme-linked immunosorbent assay kits for detection of cortisol in channel catfish,
largemouth bass, red pacu, and golden shiners. Fish Physiol. Biochem. 34(1): 95–101.
doi:10.1007/s10695-007-9150-9.

Wada, H., Salvante, K.G., Stables, C., Wagner, E., Williams, T.D., and Breuner, C.W. 2008.
Adrenocortical responses in zebra finches (Taeniopygia guttata): Individual variation,
repeatability, and relationship to phenotypic quality. Horm. Behav. 53(3): 472–480.
doi:10.1016/j.yhbeh.2007.11.018.

Wingfield, J.C., Maney, D.L., Breuner, C.W., Jacobs, J.D., Lynn, S., Ramenofsky, M., and
Richardson, R.D. 1998. Ecological bases of hormone-behavior interactions: the "emergency
life history stage." Am. Zool. 38(1): 191–206. doi:10.1093/icb/38.1.191.

547 Zera, A.J. and Harshman, L.G., 2001. The physiology of life history trade-offs in animals. Annu.

Rev. Ecol. Evol. Syst. **32**: 95–126.

Table 1: The mean (with SD) and range of plasma cortisol, individual fish length, mass and
Fulton's Condition factor (*K*), for juvenile brown trout (*Salmo trutta*) collected from the Gudsø
Stream, Denmark. Collected fish were blood sampled either within three minutes of capture
(baseline) or 30 minutes following a standardized three minute air exposure (post-stressor).

Treatment	Baseline	Post-Stressor
Sample size	99	99
Cortisol (ng/mL)	5.6 ± 6.7	69.2 ± 40.1
	(0.1 to 25.5)	(7.2 to 197.6)
Length (mm)	146±126	147±13
	(120 to 182)	(122 to 171)
Mass (g)	28.2±7.6	29.2±8.1
	(15.8 to 48.5)	(17.1 to 45.8)
К	0.90±0.07	0.90±0.06
	(0.66 to 1.06)	(0.72 to 1.09)

554

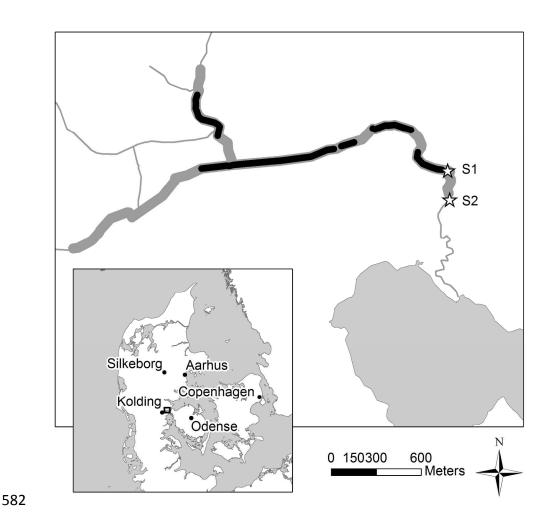
Table 2. Number of individuals recaptured, number of sampled individuals that were detected at S1 and the mean cortisol concentration (with SD) of these individuals for baseline (sampled within three minutes of capture) and post-stressor (sampled 30 min minutes following a standardized three minute air exposure) treatment groups of juvenile brown trout (*Salmo trutta*) collected from the Gudsø Stream, Denmark.

Treatment	Baseline	Post-Stressor
Number recaptured	13	6
Cortisol of recaptured (ng/mL)	5.6±5.0	69.9±40.1
Number S1 passage	37	46
Cortisol Passing S1(ng/mL)	5.8±7.7	76.2±42.3
Number S2 passage	19	26
Cortisol Passing S2 (ng/mL)	8.4±9.0	79.2±42.4

Figure 1. Location of Gudsø Stream in Denmark is shown as a black box in the bottom left panel (north-east of Kolding). The portions of the Stream where brown trout (*Salmo trutta*)were captured are shown as thick black lines and the areas re-sampled in June 2013 are shown as thick grey lines. The positions of the two PIT antennas, S1 and S2, are also shown in the larger map. Light gray areas represent water and the outflow location of Gudsø Stream into Kolding Fjord. Map data from the European Environment Agency.

569 Figure 2. The probability of recapture (a), successful S1 passage (b), and successful passage 570 between S1 and S2 (c) as a function of individual baseline cortisol plasma concentration 571 collected within 3 minutes of capture, for individual juvenile brown trout (Salmo trutta) on the Gudsø Stream, Denmark on the left, and on the right the probability of recapture (d), successful 572 573 S1 passage (e), and successful passage between S1 and S2 (f), as a function of individual cortisol 574 plasma concentration 30 minutes post 3 minute air exposure. Open circles represent actual values found for individuals and the line represents the values predicted using a binary logistic 575 regression with no covariates. Binary logistic regression found no significant relationships 576 577 except for (c): increased baseline cortisol was associated with greater probability of successful S2 passage for fish that had completed S1 passage, with marginal significance when mass 578 (p=0.05) or length (p=0.04) was included in the model as a covariate, but not when Fulton's 579 condition factor was included as a covariate. Significant relationships are indicated by three 580 581 stars.





583 Figure 1

