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Published in:
Algal Research

Link to article, DOI:
10.1016/j.algal.2018.08.019

Publication date:
2018

Document Version
Peer reviewed version

Light attenuation in photobioreactors and algal pigmentation under different growth conditions – model identification and complexity assessment

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Abstract

Microalgae are photosynthetic organisms, and thus one of the most important factors affecting their growth is light. Effective design and operation of cultivation systems requires mathematically consistent simulation models that can accurately predict light availability and its impact on microalgae growth in photobioreactors (PBR). Three cylindrical column reactors, mimicking typical open pond reactors, with different diameters were used to conduct experiments where the light distribution was monitored inside the reactor. A batch experiment was conducted where the effect of nutrients and light availability on the pigmentation of the microalgae was monitored together with the light distribution. The effect of reactor size and cultivation conditions on the light distribution in PBRs was evaluated. Moreover, we assessed the effect of using different simulation model structures on the model prediction accuracy and uncertainty propagation. Results obtained show that light scattering can have a significant effect on light distribution in reactors with narrow diameter (typical to panel-type PBRs) and under cultivation conditions that promote low pigmentation. The light attenuation coefficient was estimated using the Lambert-Beer equation and it was compared to Schuster's law. The light attenuation was found to be dependent on biomass

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concentration and microalgae pigmentation. Using a discretized layer model to describe the light distribution in PBRs resulted in the most accurate prediction of microalgal growth and lowest uncertainty on model predictions.

Keywords

Green microalgae; Pigments; Light attenuation; Photobioreactor operation; Model identification

1. Introduction

Optimizing microalgal cultivation is critical for effective reactor operation. One of the most important factors affecting microalgal growth is light availability [1]. Light is essential for microalgae to conduct photosynthesis and photoautotrophic cultivation is not viable without sufficient light in the reactor [2]. During photosynthesis microalgae convert carbon dioxide and water into carbohydrates and oxygen using light as an energy source [3]. In the light reactions, the light harvesting antenna collects the incoming light (i.e., photons) that is transported to the reaction centres (PSI and PSII) where this energy is converted into chemical energy in the form of $NADPH_2$ and ATP [4]. In the dark reaction or Calvin cycle the produced chemical energy is used to reduce carbon dioxide to phosphoglycerate, which can be further converted to, e.g., carbohydrates [4]. In closed photobioreactors (PBR), the light is more efficiently distributed as a result of optimal reactor designs, e.g., flat-panel [5]. However, in open pond cultivation systems, 90% of the incoming light intensity is absorbed in the first few centimetres of the culture, resulting in an inefficient distribution of photons [6]. Consequently, effective mixing is required to ensure that microalgal cells are regularly exposed to light [7]. Therefore for proper design of algal cultivation systems, the application of process models that accurately describe light distribution dynamics is essential [8]. Another factor affecting microalgae cultivation in open pond cultivation is the potential contamination by bacteria or protozoa [6]. Open cultivation of microalgae is used especially in used

water resource recovery systems, where the potential for bacterial contamination is high [9]. The presence of bacteria can further affect the light distribution in PBRs.

There are two major groups of photosynthetic pigments in green algae: chlorophylls (green pigment), absorbing in two spectrum bands (blue (450-475 nm) and red (630-675 nm)), and carotenoids (yellow pigment), absorbing at 400-550 nm. Chlorophylls are the main photon-harvesting pigments, whilst carotenoids can serve as protective pigments against high irradiance and reactive oxygen species and improve the light absorbance and the light utilization [10,11]. Depending on the culture conditions – mainly nitrogen and light availability – chlorophylls and carotenoids are expressed in different quantities [12–15]. Pigments are also important high value products that can be used as, e.g., food and feed ingredients or cosmetics [11,16–18].

Typically, there are three distinct light regimes prevailing through algal growth. Under light limited conditions, photosynthesis shows linear dependency on light intensity. The maximum photosynthetic rate is reached at saturation light intensity, from where the photosynthetic rate is limited by the dark reactions [2]. Light intensity that is higher than the saturation level causes photoinhibition, whereby the photosynthetic rate declines due to non-photochemical quenching to dissipate the excess energy as heat [19]. Algae exposed to inhibiting light intensities for more than 1 min will be affected by photoinhibition [19]. Due to light dynamics, microalgae have developed acclimation mechanisms to cope with light intensity changes. Regulation occurs in the reaction centres, mainly in PSII, by altering their photon-harvesting capacity or the number of reaction centres [20]. Under light limiting conditions microalgae increase the amount of chlorophyll, i.e. their photon-harvesting capacity. Under high light intensity, chlorophyll levels are reduced to avoid excess energy harvesting [19].

Light attenuation in the PBR is affected by the absorption capacity of photosynthetic pigments, the shading effect by cells and light scattering caused by reactor wall and cells [10]. The Lambert-Beer expression accounts for the light absorption in the reactor by the biomass concentration [21] or by the combination of biomass and pigments concentration [22], but does not account for scattering. Schuster's law can be used in cases where the light scattering is considered [21]. When the pigment concentration impact on light distribution is considered, it is necessary to include pigments concentration in the biological model as a state-variable. There are several approaches to model pigment concentration: i) relating the intracellular chlorophyll content to the internal nitrogen quota [22] or to the nitrogen assimilation [23], ii) considering photo-acclimation as the driving force of chlorophyll accumulation [20], or iii) relating the chlorophyll synthesis to inorganic carbon uptake [24]. The dependence of microalgal growth on light intensity can be modelled by following three complexity levels [19]. Type 1 consists of biokinetic models that employ incident or average light intensity, i.e., algal cells are assumed to be exposed to the same light intensity through the entire reactor volume and have the same photosynthetic rate, thus neglecting the effect of photo-acclimation and light attenuation (see, e.g.,[25]). Type II models account for light distribution in the culture by applying, e.g., the Lambert-Beer expression (e.g.,[21,26]) to predict the light intensity at a given reactor depth. Finally, type III models account for culture history in terms of light exposure as cells move around in the system (e.g.,[27]). Light intensity is commonly measured and expressed in the photosynthetically active radiation (PAR) range (400-700 nm) (e.g. [5,28,29]).

A microalgal biokinetic process model developed in the framework of activated sludge modelling (ASM-A) was proposed earlier [25], including photoautotrophic and heterotrophic microalgal growth, nitrogen and phosphorus uptake and storage and biomass decay processes. The effect of light intensity on photoautotrophic growth was experimentally assessed and found to be best described by the Steele equation. An average light intensity is used to account for light intensity

inside the reactor (i.e., Type I model). Moreover, in the paper, the effect of light intensity on heterotrophic growth was assessed. The goal of the ASM-A model is to move towards a consensus based process model for green microalgae. As discussed above, light intensity within PBRs can be accounted for in different ways, which was not evaluated in the original ASM-A biokinetic process model. Thus, to further develop a comprehensive process modelling framework for green microalgae, in this paper, different approaches to predict the effects of light intensity on microalgal growth are assessed.

Hence, the objectives of this study are: (i) to assess the distribution of light intensity in column reactors used for microalgae cultivation with different dimensions, biomass concentrations and pigmentation, receiving light from the top; (ii) to assess the effect of cultivation conditions on the light distribution and the pigment synthesis during batch cultivation; (iii) to identify a process model structure that can describe pigments accumulation and degradation as a function of substrate availability; (iv) to compare different simulation model complexity levels used to predict light intensity in PBRs.

2. Materials and methods

2.1. Microalgae and culture media

A mixed green microalgal consortium consisting mainly of *Chlorella sorokiniana* and *Scenedesmus sp.* was used in this study [25]. The mixed culture was cultivated using the MWC+Se synthetic medium [30] by adjusting the nitrogen and phosphorus concentrations as later specified. The consortium was also grown in effluent water from a laboratory-scale enhanced biological phosphorus removal (EBPR) system [31] operated at 16 days of solids retention time (SRT) fed with pre-clarified used water from Lundtofte WWTP (Kgs. Lyngby, Denmark).

2.2. Microalgal cultivation in batch reactors

Batch experiments were carried out in an 8-L batch reactor (made out of clear acrylic material, see Fig. S1, Supporting Information (SI)), to assess the effect of nutrients and light availability on the pigments concentration of the microalgae. The cylindrical reactor had a diameter of 140 mm, height of 0.6 m and working volume of 8-L. Constant aeration with CO_2 enriched air (5 % CO_2) at a flow rate of 20 L/h was used to mix the biomass and to provide CO_2. Light was supplied from the top of the reactor with a custom-built lamp, providing 1500 ± 150 µmol photons m^{-2} s^{-1}, with a metal-halide light bulb (OSRAM©, Germany). The reactor wall was covered with a black cloth from the outside to reduce the effect of ambient light on the monitoring of the incoming light intensity. The light sensor (described in section 2.3) was only placed inside the reactor for the course of the light intensity measurements (otherwise it was kept outside of the reactor to not interfere with the light penetration). The inoculum for the batch cultivation was taken from a reactor where the culture was cultivated under light limited conditions due to high biomass concentration (data not reported). Moreover, the inoculum was grown in a modified MWC+Se medium, and kept under nutrients in excess conditions for the inoculation period (data not shown). The MWC+Se medium was modified to reach 7.55 mg NH_4^+-N/L, 12.7 mg NO_3^--N /L and 3.5 mg PO_4-P/L. The reactor was kept at room temperature (23-24 °C). The pH of the algal culture varied in the range of 6.8 - 7.9 during the experiments. After 15 days of starvation, when nutrients were depleted in the cultivation medium, nitrogen and phosphorus were spiked again reaching 1.8 mg NH_4^+-N/L, 6.6 mg NO_3^--N /L and 0.6 mg PO_4-P/L. Algae biomass was diluted by replacing 20% of the culture with fresh cultivation medium, thereby supplying other micronutrients that were likely depleted.

Moreover, three reactors (made out of clear acrylic material, Fig. S2, SI) of different diameters were used in the experiments where the effect of reactor size, nutrient availability and cultivation media on light attenuation were assessed. Reactor 1 had a diameter of 240 mm, height of 0.6 m and

working volume of 22.5-L. Reactor 2 had a diameter of 140 mm, height of 0.6 m and working volume of 8-L. Finally, reactor 3 had a diameter of 110 mm, height of 1.2 m and working volume of 10.5-L. Light was supplied from the top of the reactor from 30 W fluorescent lamps (Philips, The Netherlands) in case of the tests with synthetic medium. Custom made light source was used during the tests done with used water resources. In order to eliminate ambient light, the reactor walls were covered with a black cloth during the measurements. The incident light intensity measured in each experiment is reported in Table S1, SI. Light intensity distribution in the algae suspension cultivated in synthetic medium was measured for three different concentrations of algal biomass in each reactor. Two tests were conducted using synthetic medium. In the first case microalgae were cultivated under nutrient limited conditions. The light attenuation in the culture was measured on day 1, day 2 and on day 4 of the nutrient limited cultivation. Thus three different concentrations were achieved (Table S1). In the second case microalgae were cultivated in nutrients in excess medium. The culture was grown to reach the highest biomass concentration (158 mg/L) and the light attenuation was measured. The culture was diluted two times with synthetic medium, to conduct the light attenuation measurements at the lower concentrations as well (at 79 mg/L and 39.5 mg/L). More details on the experimental design are reported in the SI, SI-1.

2.3. Analytical methods

LI-193 SA Spherical Quantum Sensor (LI-COR, USA) was used to measure the light intensity inside the reactors, connected to a LI-1400 data logger (Fig. S1, SI). The sensor measures within the PAR range. The sensor has a uniform sensitivity to light wavelengths between 400 and 700 nm, which corresponds to light used by algae for photosynthesis. The light intensity sensor was placed in a circular fitting, to ensure that it stayed vertical during the measurement (Fig. S3, SI). It was submerged at the centre of each reactor through the top opening of the reactor and the cable was fitted through a 20 mm hole in the bottom (Fig. S3, SI). The sensor could be moved up and down

the reactor column. Light intensity was measured every 2-2.5 cm over the operational depth of each reactor.

pH was monitored with a pH-electrode Sentix 940 sensor, connected to a MultiLine multi-meter 3430 (WTW, Germany), and dissolved oxygen was monitored using a FDO 925 optical oxygen sensor (WTW, Germany), connected to the same multi-meter.

Total suspended solids (TSS) measurement was carried out using glass fibre filters (Advantec©, USA) with a pore size of 0.6 µm based on standard methods [32]. Total nitrogen and phosphorus measurements in the suspension were done using commercial test kits (Hach-Lange©, USA). Ammonium, nitrate, nitrite and phosphate concentrations were measured after sample filtration through 0.2 µm syringe filters (Sartorius, Germany) using test kits supplied by Merck© (USA). The internal cell quota of nitrogen was obtained based on the difference of total nitrogen measured in the algal suspension (algae + medium) and total soluble nitrogen in the filtrate (soluble organic N + ammonium + nitrite + nitrate). The internal cell quota of phosphorus was calculated by taking the difference of total phosphorus measured in the algal suspension and soluble phosphate measured in the filtrate.

Pigments extraction method was adapted from literature [11,33] and the detailed protocol is reported in the SI, SI-2. The pigments were analysed using ultra high performance liquid chromatography (UHPLC) based on [33]. We targeted chlorophyll *a* and *b* as well as some carotenoids (lutein, β-carotene, violaxanthin) as these were the most common pigments found in *Chlorella sp.* according to literature [11,16,33,34].

2.4. Calculations and statistical analysis

Principle component analysis (PCA) was carried out to assess the relevant correlations between factors that could affect the light attenuation using Matlab (The MathWorks, USA). The variables

that were considered in the PCA were chosen to be the internal nitrogen and phosphorus quota, the chlorophyll a and b content, the violaxanthin, β-carotein and lutein concentrations and the biomass concentration. The variables were standardized based on their mean and standard deviation in order to be able to represent them on the same scale.

SigmaPlot® was used to fit regression on the experimental data obtained in the three reactors presented in section 2.2. The standard error of the estimate parameter values obtained through the fitting is shown as error bars in the figures in the results section.

The Lambert-Beer expression and the Schuster's law (see, e.g., [21]) were fitted on light distribution curves measured inside the reactor in SigmaPlot® (CA, USA). The two equations were chosen to compare the fitting including light scattering (Schuster´s law) and without light scattering (Lambert-Beer equation).

The Schuster's law is expressed as [35]:

$$I = I_0 * \frac{4\alpha}{(1+\alpha)^2 * e^{\delta * X_{Alg} * z} - (1-\alpha)^2 * e^{-\delta * X_{Alg} * z}} \qquad \text{Eq. 1}$$

where

$$\alpha = \sqrt{\frac{E_a}{E_a + E_s}} \quad \text{and} \quad \delta = \sqrt{E_a * (E_a + E_s)}$$

where I (µmol $m^{-2}s^{-1}$) is the light intensity measured at depth z (m), I_0 (µmol $m^{-2}s^{-1}$) is the incident light intensity, X_{Alg} (g m^{-3}) is the biomass concentration, E_a is the light absorption coefficient and E_s is the light scattering coefficient.

Another approach proposed in this study accounts for light scattering by increasing the measured light path length (depth of the reactor) with a correction factor. An optical path length multiplication

(PLM) factor was determined from the curve fit by fitting the Lambert-Beer equation, modified with the PLM:

$$I = I_0 * e^{-k_a * X_{Alg} * z * PLM} \quad \text{Eq. 2}$$

where I (µmol $m^{-2}s^{-1}$) is the light intensity measured at depth z (m), I_0 (µmol $m^{-2}s^{-1}$) is the incident light intensity, k_a (m^2 g^{-1}) is the attenuation coefficient, X_{Alg} (g m^{-3}) is the biomass concentration and PLM (-) is the path length multiplication factor. In this way, the true optical path length caused by scattering was predicted.

2.5. ASM-A model complexity analysis and model extension

As discussed earlier, different model complexities are used to account for light intensity in the PBR. We tested three different assumptions to account for light intensity during model simulations, all of them based on the Lambert-Beer law – see section 3.3. Complexity level 1 (CL-1) assumes that there is a constant average light intensity available in the reactor throughout the simulation. The average light intensity (I_{av}), which was set constant over time, was calculated by integration of the Lambert-Beer law as presented in Wágner et al.[25]. CL-2 includes the dynamic calculation of the average light intensity (by integration of the Lambert-Beer law, as in CL-1) for each time-step of the simulation. In this way, light intensity can be updated over time taking into account the impact of biomass concentration. Finally, in CL-3, the culture volume was discretized into *n* equal layers parallel to each other and orthogonal to the light source, which entered from the top of the reactor. The layer model structure is similar to the model reported by Huesemann et al.[29]. The light intensity is calculated in the middle of each layer using the Lambert-Beer equation. The ASM-A biokinetic model is then solved in each layer for one time-step, whereby different growth rates are expected due to the gradient in light intensity within the PBR. The reactor is modelled as a continuously stirred tank reactor (CSTR) operated as a batch. Therefore, the state-variables

228 calculated in each layer are then numerically averaged for the entire volume at the end of each time-
229 step and average values were used as initial conditions for the next time-step. The optimal time-step
230 and the number of layers were estimated to be 0.1 d and 10 layers, respectively, by comparing the
231 root mean square normalized error (RMSNE) of the simulations (Fig. S4, SI). The RMSNE was
232 calculated by comparing the simulation to the experimental data in Batch 1. The attenuation
233 coefficient present in the Lambert-Beer equation was estimated first based on the TSS concentration
234 (Eq. 3) and then based on the total chlorophyll concentration (Eq. 4) resulting in six assessments in
235 total.

236 The ASM-A model was extended with the prediction of the chlorophyll content of the microalgae.
237 As previously reported in the literature (e.g.[22]), the chlorophyll content is set proportional to the
238 internal nitrogen quota ($X_{Alg,N}$). Chlorophyll is reported to be an easily accessible nitrogen source
239 from the internal nitrogen pool that can be degraded under nitrogen limitation [36]. Thus, it is
240 hypothesized that the chlorophyll that is degraded provides nitrogen to be used inside the cells. We
241 introduced an independent decay term for the chlorophyll content (R7, Table 1), assuming that it is
242 degraded faster than the internal nitrogen content.

243 **<Table 1>**

244 **2.6. Model implementation, calibration and evaluation**

245 The different model structures were implemented in Matlab (The MathWorks, USA) as extensions
246 of the ASM-A simulation model by Wágner et al.[25]. Parameter estimation and model
247 identifiability analysis were carried out based on the Latin-Hypercube-Sampling-based priors for
248 Simplex (LHSS) method [25]. Parameter identifiability is assessed by analysing the posteriori
249 parameter distribution, i.e., parameter 95% confidence interval and covariance based on 500

Simplex runs. Values for parameters not estimated in this study were taken from the original ASM-A calibration.

The model complexity was compared based on four criteria: (1) model accuracy assessment based on the root mean square normalised error (RMSNE) and Akaike's information criterion (AIC) [37]; (2) parameter uncertainty based on the comparison of mean value and 95% confidence interval; (3) parameter correlation based on [38]; (4) model prediction uncertainty, assessed based on the 95% confidence bands using average relative interval length (ARIL, based on Dotto et al.[39]) together with the coverage, expressed as ARILC by Ramin et al.[40]. For further details on calculating the above criteria, the reader is referred to the SI, SI-3.

3. Results and discussion

3.1. Estimation of light attenuation under different growth conditions – preliminary evaluation in short term batch experiments

As light penetrates in a PBR containing a microalgal suspension, there is a decrease in the light intensity with increasing depth (see an example in Fig. 1a). This is due to the light absorption and shading effect by the culture [7]. The Lambert-Beer equation was fitted to light distribution data (see examples in Fig. 1a and 1b) experimentally obtained to estimate the light attenuation coefficient in three PBRs, having different reactor diameters and using three different biomass concentrations. The light attenuation coefficient (k_a) (reported in Table S2, SI) was found to vary as a function of biomass concentrations (Fig. 2a and 2b, Table S3, SI). There was no significant difference in the dependence of k_a on biomass concentration between 240 and 140 mm diameter reactors (Fig. 2a and 2b), whilst the narrowest reactor (110 mm diameter) showed a different relationship with the biomass concentration. We note that the sensor used to measure the light intensity inside the reactor has a diameter comparable to that of R3 (6.1 cm and 11 cm,

273 respectively), which could potentially affect our observations (e.g., increase of light scattering).
274 This factor is assumed to be negligible in influencing measured light intensity data in our study.
275 Additionally, the nutrient availability was found to have significant impact on the predicted light
276 attenuation coefficient (Fig. 2a and 2b). The bubble size did not significantly (based on standard
277 deviation and student t-test) affect the light attenuation in the PBR (Fig. S5, SI). However, a more
278 dedicated analysis of using different diffusers, mixing-conditions and air-flows should be done to
279 thoroughly evaluate the effect of bubble size on, e.g. light scattering.

280 **<Figure 1>**

281 As light penetrates through the culture it can be affected by the scattering from the reactor walls and
282 back-scattering from the microalgal cells [5]. Scattering from the reactor walls can enhance the light
283 intensity as light penetrates through the reactor (see Fig. 1c). Scattering from the reactor walls
284 changes the direction of the light beam as it propagates through the reactor, thereby changing and
285 potentially increasing the true optical path length of light within the algal culture. Therefore, light
286 scattering was quantified by increasing the measured path length with a correction factor (i.e. the
287 optical path length multiplier, PLM), thereby predicting the true optical path length. The Lambert-
288 Beer equation was fitted on the curves, using k_a as estimated in Table S3 (SI) for the nutrient
289 limited and nutrients in excess scenarios. Values of PLM were estimated using the curve fit (Eq. 2,
290 Table S4, SI). The best fit was obtained based on R^2. In the case of the narrowest reactor for all
291 tested biomass concentrations, the model predictions can be improved by using the PLM. In case of
292 the wider reactors PLM was only needed for the nutrient limited scenario (Table S4, SI). Under
293 nutrient limited condition the pigment composition changes in the culture, which can result in
294 decreased light absorption by the biomass (Fig. 2a and 2b) compared to nutrients in excess
295 cultivation. Due to the lower light absorption, light scattering can be enhanced by the reactor wall,
296 and thus the use of PLM can improve model prediction. Additionally, Pandey et al.[41] found that

297 the wall reflection of light in the reactor has higher impact at lower biomass concentration than in
298 high concentration. Our results (Fig. 2c and 2d) suggest higher attenuation coefficients at low
299 biomass concentrations. Observations made using the PLM were confirmed by fitting Schuster's
300 law on the light distribution curves (Table S5, SI). In case scattering becomes insignificant, the
301 parameter E_s in Schuster's law approaches 0, and thus the expression becomes identical to the
302 Lambert-Beer law. Comparing the attenuation coefficient (k_a) estimated using the Lambert-Beer
303 law (Table S2, SI) and E_a estimated using the Schuster's law (Table S5, SI), indicates that when E_s
304 is 0, E_a and k_a are equal or not significantly different. This was found to be the case for R1 and 2
305 under in-excess nutrients concentrations, whilst under nutrient limited conditions the Schuster's law
306 gave better fit. In case of R3 (narrowest diameter) a better fit was obtained by applying Schuster's
307 law compared to that of Lambert-Beer under both nutrient limited and in-excess conditions. Thus,
308 modelling the effect of scattering by implementing Schuster's law [21] or the PLM approach is
309 needed to accurately predict light attenuation in reactors with narrow diameter, e.g. flat-plate PBR.
310 However, the prediction by the Lambert-Beer equation, i.e. without accounting for scattering, is
311 sufficient in reactors intended to be used at high biomass concentrations, typically the case in PBRs.

312 Based on the correlation between the attenuation coefficient and the TSS concentration an
313 exponential relation was obtained, and used to approximate data points (Fig. 2c):

314 $$k_a = a * e^{-b*X_{Alg}} \qquad \text{Eq. 3}$$

315 where a ($m^2\ g^{-1}$) and b ($m^3\ g^{-1}$) are the correlation parameters estimated and X_{Alg} ($g\ m^{-3}$) is the
316 biomass concentration.

317 The light distribution in a PBR also depends on the cultivation conditions, i.e. nutrient availability
318 and culture medium, which can affect microalgal physiology (e.g., pigments content and
319 composition). Under nutrient limited cultivation the estimated attenuation coefficient values are

significantly higher than for parameters estimated under nutrients in-excess cultivation (Fig. 2c), suggesting that algae absorb less light when algae are cultivated under nutrient limited conditions. As a result of different cultivation conditions, algae change their pigmentation (see Fig. S6, SI). Under nitrogen limitation, chlorophylls are considered to be the first nitrogen pools inside the algae accessed by the cells [14], and thus the chlorophyll content of the algae is expected to decrease together with nutrient availability. Moreover, in more diluted cultures, the light intensity that the algae is exposed to is comparably high, thereby promoting the production of carotenoids serving as photo-protective pigments by capturing energy on characteristic wavelengths [42]. This effect can alter the light absorption of the microalgal cells and thus the light attenuation in the reactor.

<Figure2>

The composition of cultivation medium can also affect the light distribution in PBR, e.g., treated wastewater contains chromophores and particulate matter that can interfere with light attenuation in PBRs. We assessed the effect of such chromophores on light attenuation in PBR using treated water derived from a laboratory scale EBPR (Fig. 1c and Fig. 2c). We found that using treated used water as cultivation medium resulted in comparably high absorbance (i.e., lower k_a) than using clear and colourless synthetic medium (Fig. 2c). Moreover, we assessed the effect of increasing bacterial biomass concentration on light attenuation. Increasing bacterial biomass concentration can further increase light absorption in the reactor (Fig. 2d). Thus, experiments designed for the estimation of the attenuation coefficient should be carried out using the cultivation medium relevant for the system. This effect is crucial to model combined bacterial-algal cultivation systems where the bacterial biomass concentration can vary (e.g.[43,44]).

3.2. Effect of cultivation condition on pigments synthesis and light attenuation – evaluation under dynamic conditions

A 16-day batch experiment was run (Batch 1), where after 3 days the nutrients were depleted from the medium, whilst the biomass concentration kept increasing until day 6 (Fig. 3a and 3b). The chlorophyll *a* and *b* concentration inside the biomass decreased from the beginning of the experiment, reaching a plateau after 4 days (Fig. 3c). A slight increase in the chlorophyll *a* (the primary chlorophyll type) content of the microalgae can also be observed by the end of the cultivation (Fig. 3c). Among the measured carotenoids, lutein was present in the highest concentration (Fig. 3d).

<Figure3>

As opposed to the observed trends in chlorophyll depletion, carotenoids were accumulated in the first 2 days and then depleted until the end of the cultivation period, possibly due to the increase of biomass concentration, which results in reduced light intensity inside the reactor (Fig. 4a). When microalgae are exposed to high light intensities, the chlorophyll production is suppressed and carotenoids are synthesized due to photo-acclimation processes against high light intensity [11,15,20,21,42,45,46]. In the beginning of the cultivation the sudden increase of light intensity (average light intensity was 215 µmol m^{-2} s^{-1} in the start of the cultivation after the inoculum was acclimated to low light intensity) could potentially result in photo-inhibition as suggested by, e.g., García-Camacho et al.[20] or Vaquero et al.[42]. Moreover, Adesanya et al.[24] report the decrease of chlorophyll in *Chlorella vulgaris* instantaneously after the start of batch cultivation due to nitrogen limitation in the culture with initial nitrogen concentration similar to our case. Ferreira et al.[36] report that microalgae increase their chlorophyll content under low light intensity to harvest light more efficiently, which can be observed in our experiment at the end of the cultivation period. Furthermore, photo-protective pigments such as carotenoids can be used by microalgae to reduce

365 the negative effects of elevated light intensity. Carotenoids can dissipate excess light through non-
366 photochemical processes, and as antioxidants they can reduce the effect of reactive oxygen species
367 [13,42]. Thus the increase in carotenoids content in the beginning of the process is likely due to the
368 elevated light intensities. Moreover, the increase in the carotenoid concentration in the end of the
369 experiment (when chlorophyll content increases as well) can be related to the widening of the light
370 absorption spectrum, whereby carotenoids enhance the light harvesting capacity to enhance the
371 photosynthetic activity [36,42].

372 **<Figure4>**

373 In the beginning of the second batch experiment (Batch 2), to assess the change in pigments
374 concentration under changing nutrient availability, nutrients were spiked to the starved culture.
375 There is an increase in chlorophyll *a* and *b* concentration during the first 2 days (Fig. 3e). This is
376 possibly due to the available nitrogen in the medium that promotes the synthesis of chlorophyll to
377 enhance photosynthesis [21,36]. 2 days after the bulk nitrogen source is depleted, there is a decrease
378 of the chlorophyll *a* and *b* concentration. As previously stated, chlorophyll is reported to be an
379 easily degradable nitrogen source for microalgae [36] and under nitrogen starvation chlorophyll is
380 degraded to support growth [14]. Lutein concentration increases slightly in Batch 2 (Fig. 3f). In this
381 case it is unlikely that lutein serves as a photo-protective pigment, as the average light intensity is
382 similar to the one estimated in the end of the cultivation in Batch 1 (Fig. 4a). Likely, lutein serves to
383 widen the light absorption spectrum, to promote effective photosynthesis [36]. Results are subject to
384 the pigment extraction protocol which were demonstrated to be inefficient for lutein extraction [47].

385 **3.3. Modelling of the effect of chlorophyll on light attenuation**

386 The total chlorophyll concentration was expressed as nitrogen based on the nitrogen content of
387 chlorophyll in the molecular formula (chlorophyll *a*: $C_{55}H_{72}O_5N_4Mg$ and chlorophyll *b*:

388 $C_{55}H_{70}O_6N_4Mg$; [48]). We found a linear correlation with the nitrogen quota of the microalgae and
389 the total chlorophyll content (Fig. 5), as also suggested by Bernard[22]. Moreover, Ikaran et al.[49]
390 found similar trends between the stored protein and chlorophyll content of microalgae during batch
391 cultivation, where protein is suggested to be part of the nitrogen quota [14]. However, the maximum
392 nitrogen content present as chlorophyll in the total nitrogen quota was about 2% in our study and
393 thus it forms an insignificant fraction of nitrogen storage. This is in agreement with Geider and La
394 Roche [48], who reported that 0.2-3% of the intracellular nitrogen is associated with chlorophyll.

395 **<Figure 5>**

396 The k_a was estimated inside the reactor during the course of the 8-L batch experiments using the
397 Lambert-Beer expression. Similar to the previous results presented in section 3.1, values of k_a
398 change as function of the TSS concentration (Fig. S7, SI), which can be described using an
399 exponential relation (Table S6, SI). Thus, to effectively predict the light distribution in the PBR, the
400 value of k_a cannot be expressed as a constant value, but as a variable updated during the cultivation
401 period (Fig. 4b). We calculated the effective attenuation coefficient that is the product of the
402 attenuation coefficient (k_a) and the biomass concentration (X_{Alg}), to decouple the effect of biomass
403 concentration on the light attenuation. This value increases (Fig. 4c) during the cultivation period.
404 As the biomass concentration increases and thus the light intensity inside the reactor decreases the
405 effective attenuation coefficient increases, as can be seen in Eq. 2.

406 Results obtained in a PCA analysis (Fig. 6a) - whereby the smaller the angles between vectors the
407 stronger correlation is [50] – suggest k_a to be the most dependent on the chlorophyll *a* and *b* content
408 and the internal nitrogen quota and not dependent on the carotenoids, whereas it is negatively
409 correlated with the biomass concentration. Consequently, the k_a expressed as a function of total
410 chlorophyll concentration is proposed. We found different trends between the attenuation and the

total chlorophyll concentration than in the case of TSS (Fig. 6b). The correlation between the cellular pigment content-specific attenuation coefficient ($k_{a,p}$) and the chlorophyll concentration is assessed based on analysing different algebraic expressions (in SigmaPlot®) and it is identified (based on R^2) as:

$$k_{a,p} = \frac{d}{X_{Chl}} - c \qquad \text{Eq. 4}$$

where $k_{a,p}$ (m^2 g^{-1} Chl) is the attenuation coefficient specific for pigments, c (m^2 g^{-1} Chl) and d (m^{-1}) are the regression parameters estimated and X_{Chl} (g Chl m^{-3}) is the total chlorophyll concentration that cannot equal zero. Above approximately 4 mg Chl/L, $k_{a,p}$ becomes independent of the chlorophyll content (Fig. 6b). The correlation between $k_{a,p}$ and the pigments concentration (based on Eq. 4) is shown in Table S6, SI.

<Figure 6>

The cellular chlorophyll content can be modelled as a function of the internal nitrogen quota and by introducing a specific chlorophyll decay process rate (R7, Table 1). The specific chlorophyll decay rate coefficient (b_{XChl}) was estimated using measured data obtained in Batch 1. A value of b_{XChl}=0.45±0.043 d^{-1} was estimated using the LHSS method. The fraction of chlorophyll-nitrogen (fXN_{Chl}) to the total cellular nitrogen quota was estimated from the slope of Fig. 5, i.e. fXN_{Chl} = 0.026 gN-Chl/gN. The chlorophyll concentration can effectively be predicted using the extended ASM-A simulation model (Fig. 7). The variability of b_{XChl} was assessed using the Janus coefficient (J) by comparing RMSNE values obtained with Batch 1 (used for model calibration) and Batch 2 (used for model validation). J~1, thus b_{XChl} estimate derived from Batch 1 can be used to achieve accurate model prediction in Batch 2 (Fig. S8, SI). In the following section (3.3) we evaluate the difference of calculating k_a as a function of TSS and chlorophyll content on the model simulations (in all complexity levels CL1-CL3).

<Figure 7>

3.4. Simulation model complexity evaluation

Three different model structures to predict the impact of light on algal growth were compared, together with different expressions for the light attenuation coefficient (k_a or $k_{a,p}$), using four selection criteria (see section 2.6). Model accuracy assessment was based on RMSNE and AIC calculations (Table S7, SI). Based on these two criteria, the accuracy of the predicted biomass concentration (X_{Alg}) improved by using a model structure with higher complexity, i.e. the layer-model, regardless of the constitutive equation used to calculate the attenuation coefficient. The sum of RMSNE calculations suggest, as opposed to the AIC results, that there is a worse overall fit with using the layer model. This discrepancy is due to that normalized objective functions, e.g. RMSNE used in this study, result larger values when experimental data are low (e.g., values below 1) [51]. In case of the AIC calculation there is no normalization included (Eq. S2, SI). We hypothesise that, using an average and constant light intensity value might result in the inaccurate prediction of the measurement data in both cases. Under high biomass concentrations the simulation model tends to over-predict the experimental data (Fig. S9 and S10, SI). Implementing the time-variable average light intensity function reduces this over-prediction (Fig. S11 and S12, SI). Finally, using a one-dimensional model structure improves the goodness of fit predominantly for the prediction of the biomass concentration (Fig. S13 and S14, SI and Table S7, SI), as a result of the more realistic prediction of light availability for algal growth in the PBR.

The parameter uncertainty was assessed based on the comparison of the mean value and 95% confidence interval of the parameter subset estimated using the different simulation model complexity levels and attenuation coefficients (Fig. 8, Table S8, SI,). The different model structures do not significantly influence the parameter estimates across the scenarios. The mean estimate for

457 the maximum specific growth rate using the layer model structure is similar to the maximum
458 specific growth rate estimated for *Chlorella sorokiniana* [52], which was estimated in a flat-plate
459 PBR, where no light limitation occurs.

460 **<Figure 8>**

461 The parameter correlation was compared using the LHSS method. The *posteriori* parameter
462 distributions were presented as histograms (Table S9-S14, SI). The histograms are narrow and the
463 95% confidence interval is low (below 40%) in case of CL-2 and CL-3. However, in case of CL-1,
464 the 95% confidence interval is higher than 40% in case of $\mu_{A,max}$ and $k_{NO,Alg}$, due to the simplifying
465 assumption of using average light intensity. The covariance matrices show that most of the
466 parameters are identifiable (covariance is below 0.5) in case of CL-2 and CL-3, thus the reduction
467 of uncertainty with more complex model structures might improve parameter identifiability.
468 Interestingly, $\mu_{A,max}$ and $k_{NO,Alg}$ show correlation in all cases that can be due to the challenges of
469 calibrating $k_{NO,Alg}$, as discussed in Wágner et al.[25].

470 The model prediction uncertainty was assessed based on the 95% confidence bands using ARIL
471 divided by the percentage coverage, expressed as ARILC (based on Ramin et al.[40]). The
472 simulation model performance is improved with increasing model structural complexity. The width
473 of the uncertainty bands is reduced as model complexity increases (Fig. 9; Table S7, SI; Fig. S15-
474 S19, SI). This is due to the reduced parameter uncertainty, based on 95% confidence interval (Fig.
475 8) when using a more complex model to predict light impact on algal growth. However, in the case
476 of the internal nitrogen cell quota there is a significant number of data points outside of the
477 prediction band for both variable light intensity and layer model cases, mainly due to the decrease
478 of the wideness of the prediction.

Using the average light intensity to account for light (CL-1) gave the least accurate predictions. This scenario is furthest from reality as we assume that the light intensity is the same throughout the cultivation, which is not true, because among others, the biomass concentration increases and thus light intensity decreases. Using the variable average light intensity (CL-2) as a measure of modelling light inside the reactor gives comparably more accurate model predictions. This scenario is also closer to reality, as we account with the effect of the change in biomass on light intensity in the reactor. Using a model with discretized layers (CL-3) to predict the light distribution in PBRs resulted in the most accurate prediction of the microalgal growth as well as the reduction of the uncertainty of the overall model output. However, the computational time significantly increases (although the optimal layer number and time-step was optimised in Fig. S4.) in case of using the layer model (up to 100 fold increase compared to CL-1) which can considerably increase the time and computational power needed. CL-2 and CL-3 performed similarly apart from the prediction of biomass concentration (a critical variable in microalgae cultivation). Thus, we conclude that using CL-3 can improve the prediction accuracy especially in case of biomass concentration. Therefore, the modeler should choose between CL-3 and CL-2 depending on the system to be modeled and the accuracy required to predict, e.g., biomass productivity.

<Figure 9>

4. Conclusions

In this study, we developed a consistent simulation model extension to the ASM-A framework to accurately predict light attenuation and distribution in PBRs using cylindrical PBRs with different diameters and under different cultivation conditions.

Three different simulation model structures were compared to predict light intensity inside the PBR. Light scattering had an effect on light distribution in reactors with narrow diameter or under cultivation conditions that promote low biomass concentrations and decreased pigmentation. This is important e.g. when biomass is grown for lipid accumulation in PBR under nutrient limitation and one must be careful to account with possible scattering.

Nitrogen limited conditions resulted in the decrease of chlorophyll content, whilst elevated light intensity promoted the synthesis of carotenoids. In the new model, the light attenuation coefficient is predicted as a function of the pigmentation – calculated as total chlorophyll content of microalgae, thus defining it as a dynamic variable. Algal chlorophyll content is predicted by the model as a function of the internal nitrogen quota and the pigment decay process rate.

We propose a consistent simulation model structure using a one-dimensional discretization (layers) to predict the light distribution in PBRs. As a result, more accurate prediction of the microalgal growth as well as the reduction of the uncertainty of the overall model output is obtained. This comes at a cost of increased computational time.

The ASM-A simulation model shows high predictive accuracy with the dynamic laboratory-scale systems. High variability of nutrient loading is typically the case in used water resource recovery systems. Under such conditions, it is also important to consider the effect of the cultivation medium, which is now also accounted for by the developed simulation model.

The significant outcomes of the paper help to better understand and predict the effects of cultivation conditions on light attenuation in PBRs. For practitioners, investigating other cultures, the implementation of the simulation model developed - using rigorous experimental, statistical and computational approaches (used in our previous study and this study) - is straightforward.

Acknowledgements

Dorottya S. Wágner thanks the European Commission (E4WATER Project, FP7-NMP-2011.3.4-1 grant agreement 280756) for the financial support. Borja Valverde-Pérez thanks the Integrated Water Technology (InWaTech) project (http://www.inwatech.org) for the financial support. The authors thank Mariann Sæbø and Michael Steidl for conducting some of the experiments and Dr. Arnaud Dechesne for the discussion on the PCA analysis.

Author contribution declaration

DSW, BGP and BVP contributed to the design of the experiments. Experiments were carried out by DSW, supported by BVP. The analysis and interpretation of the data was carried out by DSW, supported by BGP and BVP. DSW drafted the manuscript; all authors contributed to its revision and completion, and approved the final submission.

Conflict of interest statement

The authors declare that there are no known conflicts of interest associated with this publication.

Statement of informed consent, human/animal rights

No conflicts, informed consent, human or animal rights applicable.

Declaration of authors agreement to authorship

The work described has not been published previously and it is not under consideration for publication elsewhere. The publication and submission of the manuscript for peer review is approved by all authors.

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Figures

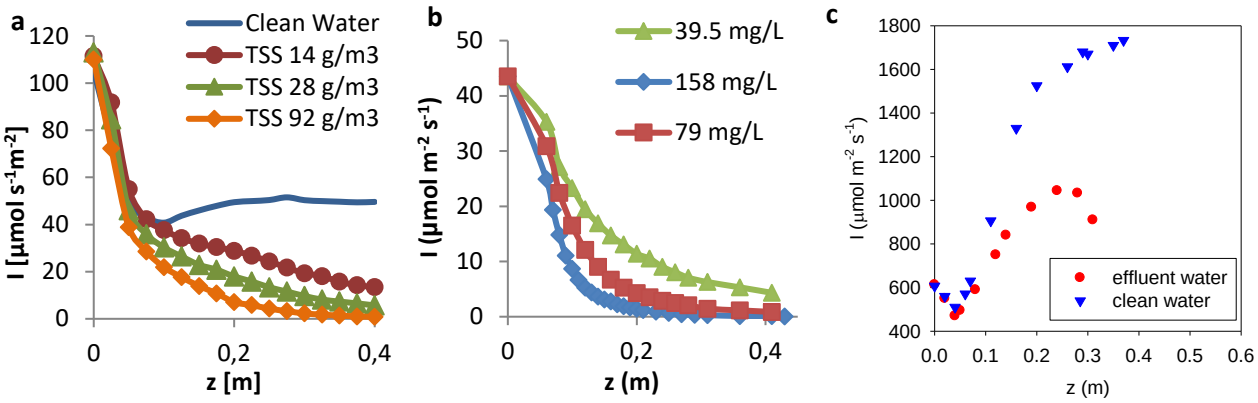
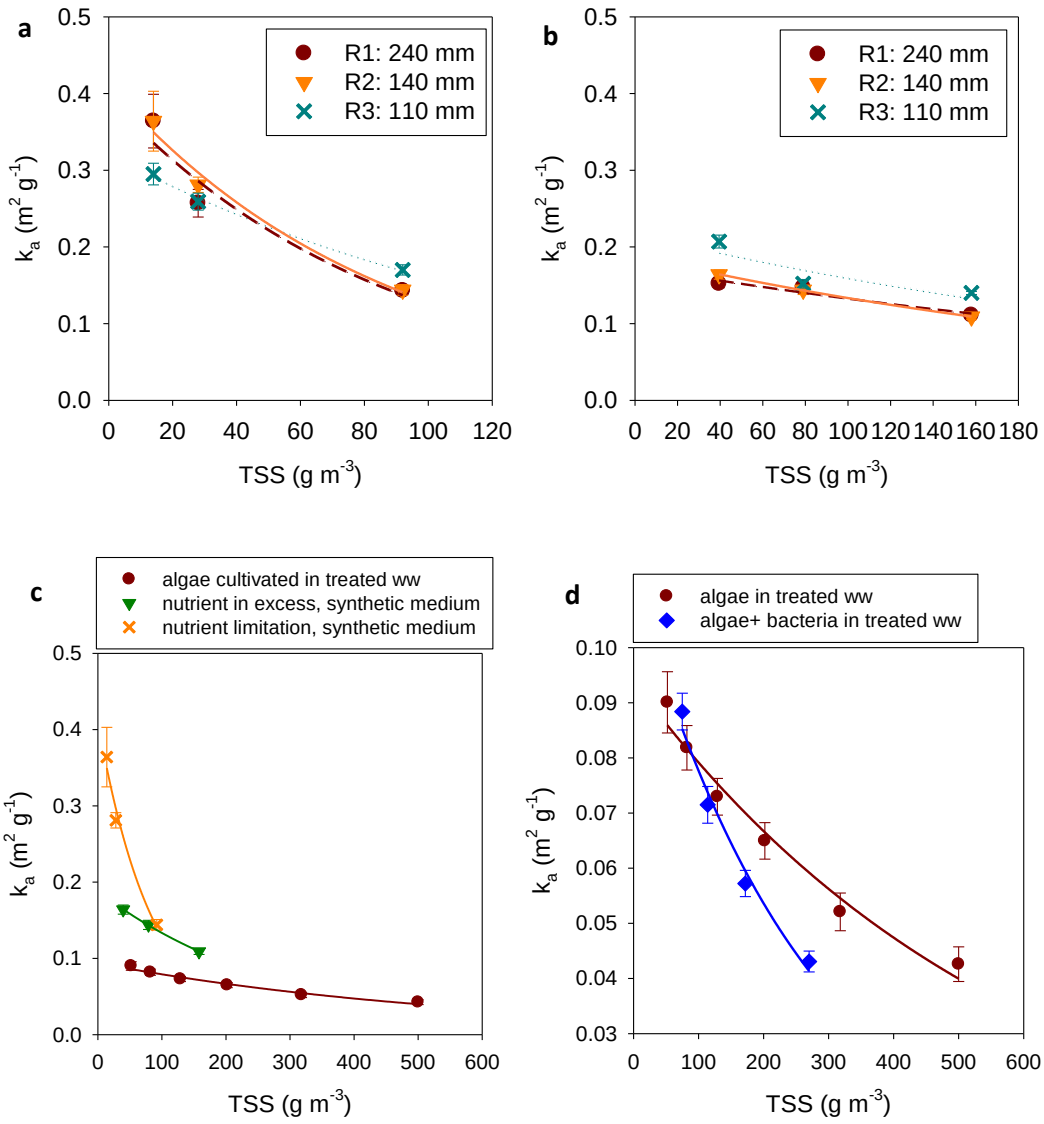


Figure 1: Light attenuation and scattering in photobioreactors, PBR (reactor diameter: 0.14 m) - effects of scattering on light distribution in PBRs. Due to the scattering on the reactor walls, the light intensity increases towards the bottom of the reactor – the bottom and the sides of the reactor were both covered with black cloth, thus light only entered from the top of the reactor. (**a**) Light attenuation inside the PBRs at different biomass concentrations, with nutrient-limited cultivation, and in clean water; (**b**) Light attenuation inside the PBR at different biomass concentrations with nutrients in excess cultivation; (**c**) Light attenuation inside the PBR with clean water and effluent used water.



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693 **Figure 2**: Estimation of the attenuation coefficient (k_a) values in PBRs with three different diameters and at
694 three different biomass concentrations. The estimation of k_a values was done both at (**a**) nutrient limited
695 conditions and at (**b**) nutrients-in-excess conditions (see Fig. S6, SI where the different pigmentations are
696 shown). The lines show exponential regression functions fitted on the measured data sets (values of
697 regression coefficients shown in Table S4). The dashed red line shows the fitting for R1, the solid orange
698 line shows the fitting for R2 and the dotted blue line shows the fitting for R3. (**c**) Values of k_a obtained at
699 different biomass concentrations with algae cultivated in synthetic medium and EBPR process effluent water
700 (denoted as ww in the legend). The observations were made in Reactor 2 (140 mm diameter). (**d**) The effect

701 of increased bacterial biomass concentration on the light attenuation in the PBR. TSS in this figure represents
702 the total TSS of algal + bacterial biomass where the amount of bacteria was increased whilst algal biomass
703 was kept constant (at 75 mg/L). The observations were made in Reactor 2 (140 mm diameter). The error bars
704 present the standard error of the estimate parameter value obtained through regression in SigmaPlot®.

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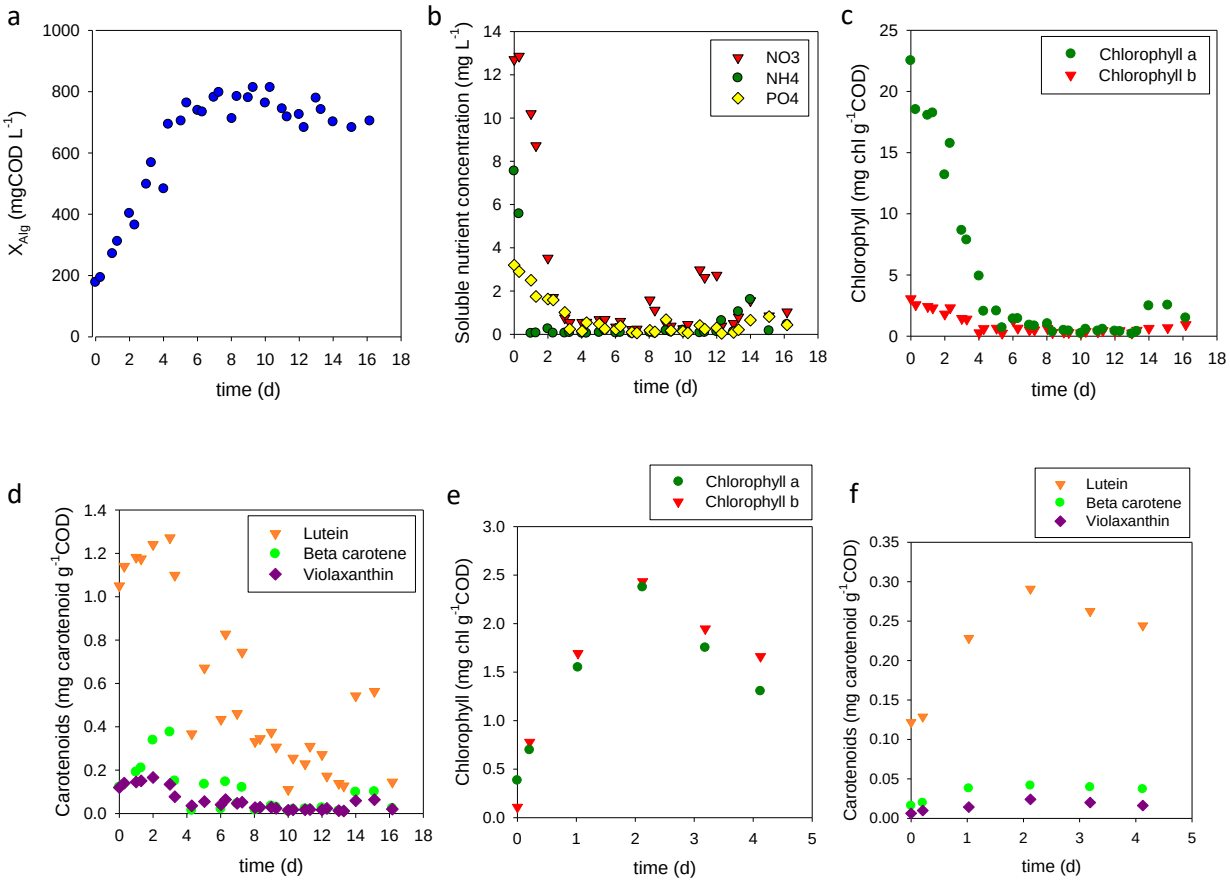

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708 **Figure 3:** Batch algal cultivation. (**a**) Microalgal biomass growth during the batch cultivation where
709 nutrients were added to a dilute culture (185 mg COD/L initial algal biomass) at day 0 and were depleted by
710 day 3 (Batch 1). (**b**) Nitrogen and phosphorus concentration during cultivation in Batch 1. (**c**) Chlorophyll *a*
711 and *b* and (**d**) carotenoids concentrations obtained in Batch 1. (**e**) Chlorophyll *a* and *b* and (**f**) carotenoids
712 concentration obtained in batch cultivation in Batch 2 where nutrients were added to a dense (400 mg
713 COD/L initial biomass concentration) and highly nutrient limited culture at day 0 and were depleted by day 2

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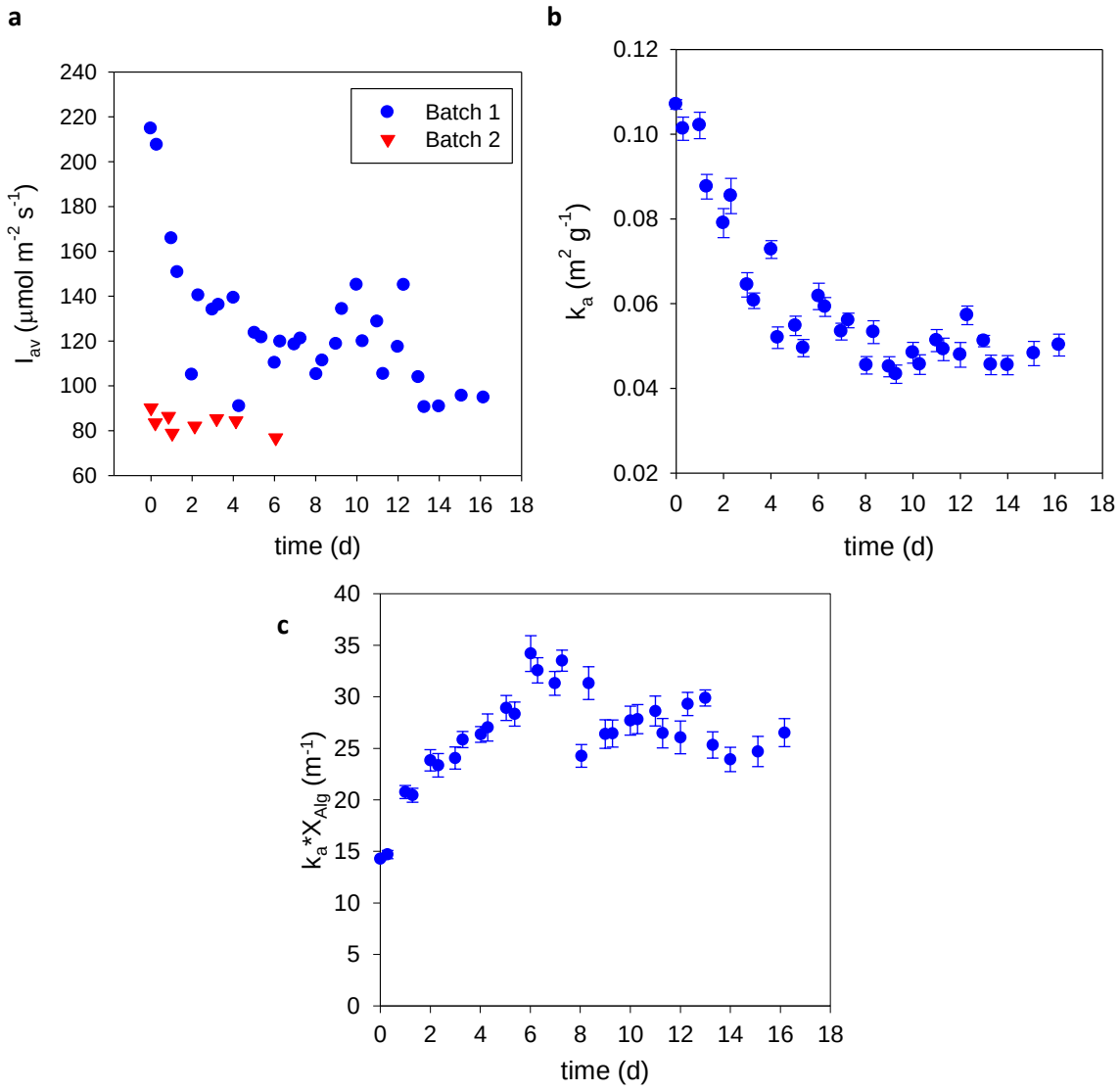

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717 **Figure 4:** Light intensity and attenuation in Batch 1 and 2. (**a**) Average light intensity in the reactor during
718 Batch 1 and Batch 2 cultivation. The average light intensity was calculated by integrating the Lambert-Beer
719 equation at each time step. (**b**) Variation of the light attenuation coefficient (k_a) over time during the batch
720 cultivation (in Batch 1). Values of k_a were estimated by measuring the light intensity at different depths of
721 the reactor and fitting the Lambert-Beer equation. (**c**) Variation of the effective attenuation coefficient,
722 calculated by the product of k_a and the biomass concentration (X_{Alg}). The error bars present the standard error
723 of the estimate parameter value obtained through regression in SigmaPlot®.

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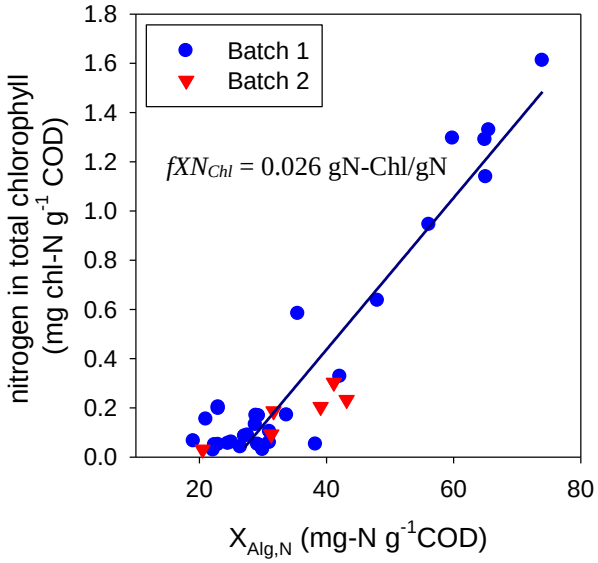
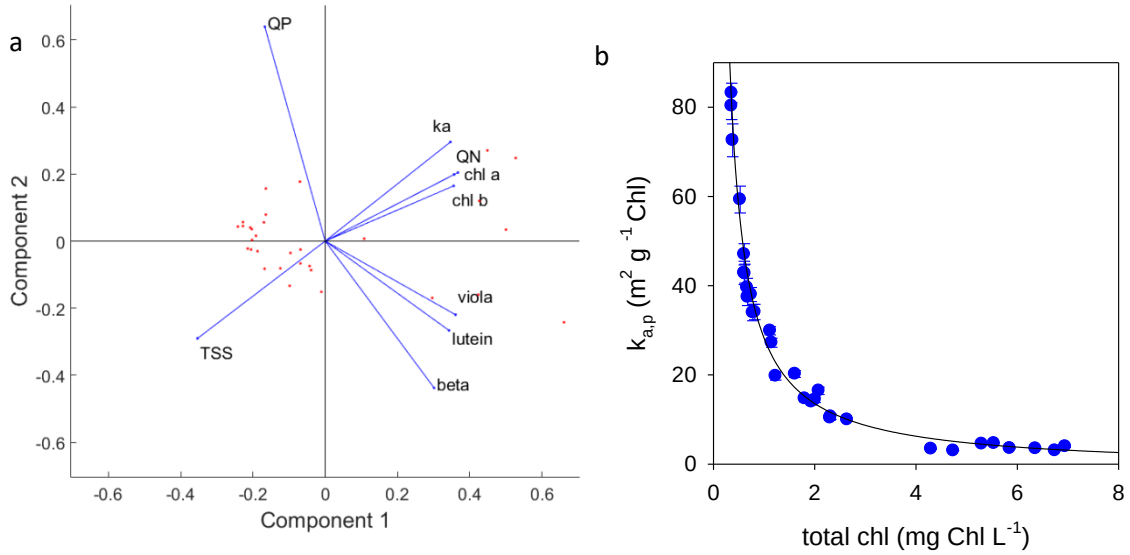


Figure 5: The nitrogen content of total chlorophyll expressed as Chl-N plotted against the internal nitrogen quota. The fraction of chlorophyll-nitrogen (fXN_{Chl}) to the total cellular nitrogen quota was estimated from the slope.



731

732 **Figure 6**: Light attenuation and prediction of k_a in PBR. (**a**) PCA analysis showing the factors that can
733 affect the light attenuation. (**b**) Estimation of the attenuation coefficient specific for the chlorophyll
734 content in Batch 1. The error bars present the standard error of the estimate parameter value obtained through
735 regression in SigmaPlot®.

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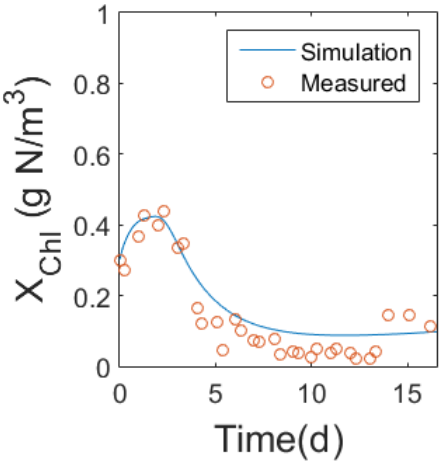

737

738 **Figure 7**: Simulation of batch experimental data using the extended ASM-A model. Prediction of

739 the chlorophyll content of the microalgae in Batch 1.

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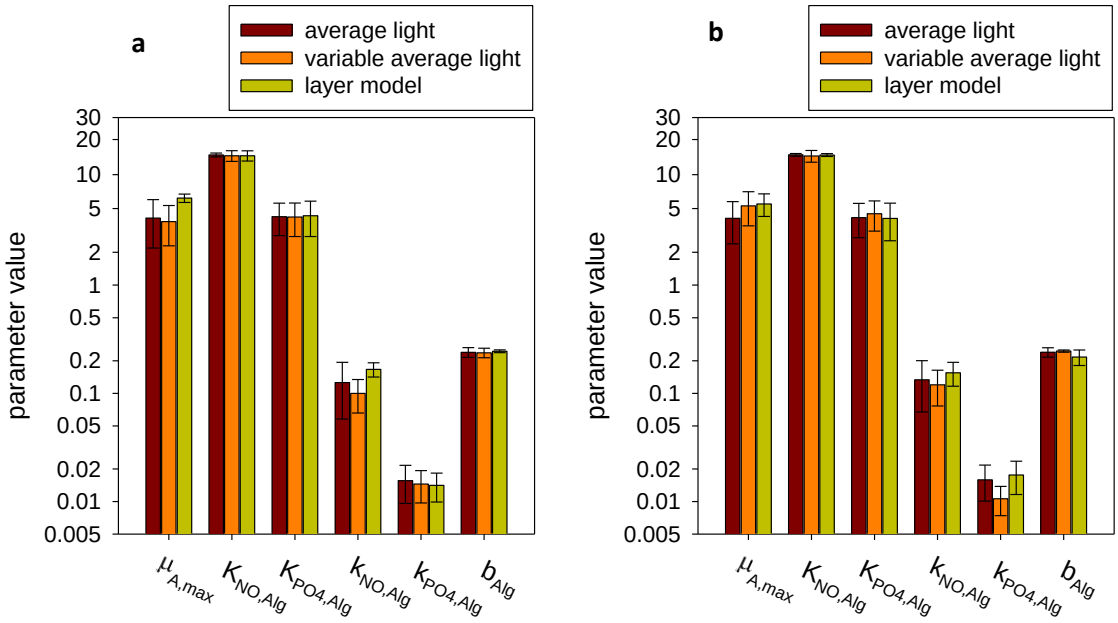
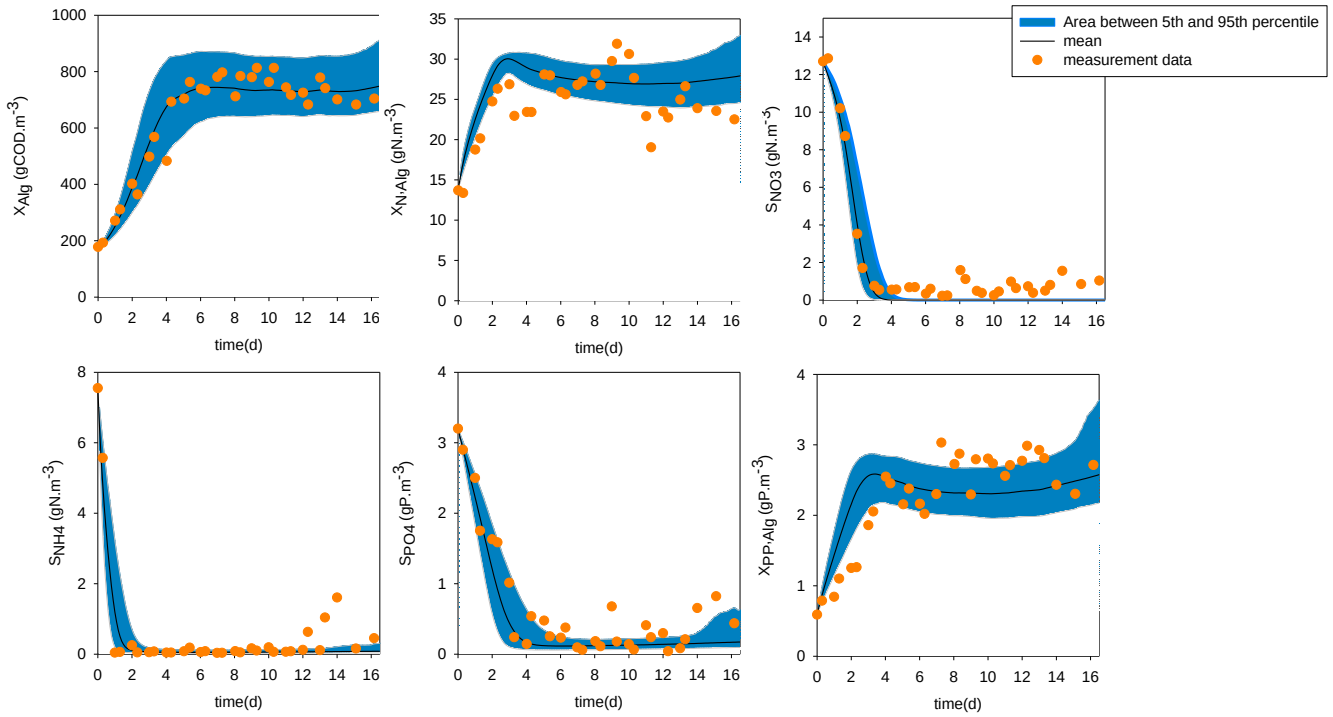


741

742 **Figure 8:** Comparison of the estimated parameter (mean value and 95% confidence interval) values
743 using different model complexity levels (CL1 - CL3). On the y-axis Weibull type scaling is used to
744 allow comparison of parameter values at different scales. (**a**) The TSS is used to calculate the
745 attenuation coefficient (k_a). (**b**) The simulation model extended to predict the algal chlorophyll
746 content is used to estimate the pigment specific attenuation coefficient ($k_{a,p}$).

747

748



749

750 **Figure 9:** Simulation of batch experimental data (Batch 1) using the extended ASM-A implemented
751 as CL- 3 (one-dimensional layer model) with the mean values of the parameters estimated. The
752 uncertainty bands are shown in blue. The chlorophyll content is used to calculate the pigment
753 specific attenuation coefficient ($k_{a,p}$) that is used in the simulations to predict the light intensity.

754

Table 1: The Gujer matrix of ASM-A model including the state-variables, the stoichiometric coefficients and the process rate equations identified in [25]. The grey highlighted columns and rows include the model extension presented in this paper to estimate the chlorophyll content.

Component	NH_4	NO_3	Internal quota N	PO_4	Internal quota P	Inorganic carbon	Acetate	O_2	Algal Biomass	Inert Particulates	Slowly biodegradable Particulate	Chlorophyll content	Process rate equations
Symbol	S_{NH4}	S_{NO}	$X_{Alg,N}$	S_{PO4}	$X_{Alg,PP}$	S_{Alk}	S_A	S_{O2}	X_{Alg}	X_I	X_S	X_{Chl}	
Unit	gN/m^3	gN/m^3	gN/m^3	gP/m^3	gP/m^3	gC/m^3	$gCOD/m^3$	$gCOD/m^3$	$gCOD/m^3$	$gCOD/m^3$	$gCOD/m^3$	gN/m^3	
Process	Stoichiometric Matrix												
Uptake and storage of nitrogen from NH4	-1		$1-fXN_{Chl}$									fXN_{Chl}	R1
Uptake and storage of nitrogen from NO3		-1	$1-fXN_{Chl}$									fXN_{Chl}	R2
Uptake and Storage of PO4				-1	1								R3
Autotrophic growth			$-iN_{Xalg}$		$-iP_{Xalg}$	$-1/Y_{Xalg,SAlk}$		$2.67/Y_{Xalg,SAlk}$	1				R4
Heterotrophic growth			$-iN_{Xalg}$		$-iP_{Xalg}$	$0.4/Y_{Ac}$	$-1/Y_{Ac}$	$-(1/Y_{Ac}-1)$	1				R5
Decay	$iN_{Xalg}-fX_I\cdot iN_{XalgI}-(1-fX_I)\cdot iN_{XalgS}$			$iP_{Xalg}-fX_I\cdot iP_{XalgI}-(1-fX_I)\cdot iP_{XalgS}$				$-(1-fX_I)$	-1	fX_I	$1-fX_I$		R6
Decay of X_{Chl}			1									-1	R7
	Process rate equations												
R1 [g N m^{-3} d^{-1}]	$k_{NH4,Alg}\cdot\frac{S_{NH4}}{S_{NH4}+K_{NH4,Alg}}\cdot\frac{X_{Alg,Nmax}\cdot X_{Alg}-X_{Alg,N}}{X_{Alg,Nmax}\cdot X_{Alg}}\cdot X_{Alg}$												
R2 [g N m^{-3} d^{-1}]	$k_{NO,Alg}\cdot\frac{S_{NO}}{S_{NO}+K_{NO,Alg}}\cdot\frac{K_{NH4,Alg}}{K_{NH4,Alg}+S_{NH4}}\cdot\frac{X_{Alg,Nmax}\cdot X_{Alg}-X_{Alg,N}}{X_{Alg,Nmax}\cdot X_{Alg}}\cdot X_{Alg}$												
R3 [g P m^{-3} d^{-1}]	$k_{PO4,Alg}\cdot\frac{S_{PO4}}{S_{PO4}+K_{PO4,Alg}}\cdot\frac{X_{Alg,PPmax}\cdot X_{Alg}-X_{Alg,PP}}{X_{Alg,PPmax}\cdot X_{Alg}}\cdot X_{Alg}$												
R4 [g COD m^{-3} d^{-1}]	$\mu_{A,max}\cdot(1-\frac{X_{Alg,Nmin}X_{Alg}}{X_{Alg,N}})\cdot(1-\frac{X_{Alg,PPmin}X_{Alg}}{X_{Alg,PP}})\cdot\frac{S_{Alk}}{S_{Alk}+K_{Alk}}\cdot\frac{I_{Av}}{I_S}\cdot e^{1-\frac{I_{Av}}{I_S}}\cdot X_{Alg}$												
R5 [g COD m^{-3} d^{-1}]	$\mu_{H,max}\cdot(1-\frac{X_{Alg,Nmin}X_{Alg}}{X_{Alg,N}})\cdot(1-\frac{X_{Alg,PPmin}X_{Alg}}{X_{Alg,PP}})\cdot\frac{S_A}{S_A+K_A}\cdot\frac{S_{O2}}{S_{O2}+K_{O2}}\cdot\frac{K_I}{K_I+I_{Av}}\cdot X_{Alg}$												
R6 [g COD m^{-3} d^{-1}]	$b_{Xalg}\cdot X_{Alg}$												
R7 [g N m^{-3} d^{-1}]	$b_{XChl}\cdot X_{Chl}$												

Supporting Information

"Light attenuation in photobioreactors and algal pigmentation under different growth conditions – model identification and complexity assessment"

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The Supporting Information consists of 29 pages. It contains 14 tables (pages 6-16) and 19 figures (pages 17-28).

SI-1 Measurement of light distribution in three reactors with different diameters

First, a blank test was carried out, where light intensity was measured over depth in reactors filled with clean tap water. The effect of aeration with different bubble size was assessed. Three different diffusers were tested during the experiments. Bubble size was measured manually based on pictures taken during the experiments, by relating the bubble size to the size of the diffuser.

Microalgae were cultivated in the effluent water of a laboratory-scale EBPR system (as described in section 2.1), to assess the effect of effluent water on the light attenuation in the reactor. A blank test was carried out to assess the light attenuation in the reactor in effluent water, without the addition of algae. Moreover, bacterial biomass was taken from the EBPR system and was spiked in the reactor containing microalgae cultivated in the effluent water to assess its impact on the light attenuation. Biomass concentrations used in each experiment are reported in Table S1.

SI-2 Pigment extraction protocol

1 ml of microalgae sample was collected in 1.5 ml Eppendorf tube and centrifuged for 10 min at 10000 rpm. The pellet was kept at -20 ºC until the extraction. The pigment extraction was done in darkness using green light to minimize the degradation of extracted pigments and when possible keeping them in ice. 1 ml 99.9 % HPLC grade methanol (Sigma-Aldrich, Germany) was added to the pellets and mixed with vortex. Ultrasonic bath (Retsch U1, Germany) was used to break the microalgal cells. During the sonication the samples were cooled with ice. Following the 60 min sonication the samples were kept on ice for 30 min to enhance extraction of pigments. The samples were then centrifuged for 5 min at 10000 rpm. The supernatant was filtered through 0.2 µm syringe filters (Agilent Technologies, USA) and 200 µl filtered sample was mixed with 600 µl 28mM Tetrabutylammoniumacetate buffer solution in amber glass vials. The samples were placed in the UHPLC and were cooled at 8 ºC until analysis.

SI-3 Model evaluation criteria

The RMSNE was calculated as:

$$RMSNE = \sqrt{\frac{1}{n}\sum_{i=1}^{n}\left(\frac{y_m - y}{y_m}\right)^2} \qquad \text{Eq. S1}$$

where n is the number of measurement points, y_m is the measured value and y is the predicted value. The AIC criterion is estimated by Akaike (1973):

$$AIC = N * \ln\left(\frac{SS}{N}\right) + 2 * K \qquad \text{Eq. S2}$$

where N is the number of data points, SS is the sum of squares of the difference between the measured data and model prediction, K is the number of parameters estimated plus one. This criteria indicates the goodness of fit of the model predictions, where a lower AIC suggests better fit. Mean and 95% confidence interval of the estimated parameter subsets were compared in the second criterion, and the parameter correlation in the third criterion, thereby assessing the impact of model structure on parameter identifiability based on the LHSS output. Finally, in the fourth criterion, the model prediction uncertainties were compared. Monte Carlo simulations were performed to obtain a confidence interval of model predictions (Sin et al., 2009). The uncertainty classes were assigned to each parameter based on Wágner et al. (2016). The probability range of the estimated parameters was calculated by the mean and the 95% confidence interval. 1000 MC simulations were run as specified by Wágner et al. (2016).

ARIL is calculated based on (Dotto et al., 2012):

$$ARIL = \frac{1}{N}\sum_{i=1}^{N}\frac{Limit_{upper,i} - Limit_{lower,i}}{X_{obs,i}} \qquad \text{Eq. S3}$$

where $Limit_{upper,i}$ and $Limit_{lower,i}$ are the upper and lower bounds, based on the 95% confidence interval obtained in the Monte Carlo simulations, $X_{obs,i}$ is the measured value, N is the number of measurement points. ARIL is used in combination with the coverage, which is the percentage of the observations that are within the prediction bands. Lower ARIL and a higher coverage suggest better model performance. Ramin et al. (2016) expressed the combination of the two evaluation criteria:

$$ARILC = \frac{ARIL}{coverage} \qquad \text{Eq. S4}$$

where a smaller ARILC indicates better model prediction.

Tables

Table S1: Initial conditions of the experiments used to assess the light distribution in three different reactors (R1, R2 and R3) at different biomass concentrations.

		R1 (240 mm)		R2 (140 mm)		R3 (110 mm)	
	X_{Alg} (mg/L)	I_0 (µmol/m^2/s)	$X_{bacteria}$ (mg/L)	I_0 (µmol/m^2/s)	$X_{bacteria}$ (mg/L)	I_0 (µmol/m^2/s)	$X_{bacteria}$ (mg/L)
nutrient limited cultivation	14	104		112		229	
	28	112		113		234	
	92	112		110		266	
nutrients in excess cultivation	39.5	42		44		353	
	79	42		44		353	
	158	38		44		353	
cultivation in used water resources	52			1032			
	82			1021			
	129			1054			
	202			1087			
	318			975			
	500			1170			
addition of bacteria	75			1099	0		
	75			1059	39		
	75			1068	97		
	75			1281	195		

Table S2: The estimated attenuation coefficients (based on the Lambert-Beer equation) for the nutrient limited and nutrients in excess cultivation in three different reactor diameters and six biomass concentrations. The blue shading refers to scenarios where the estimated attenuation coefficient is equal (or not significantly different from) to the E_a estimated by the Schuster's law.

	TSS (mg/l)	Diameter (mm)		
		240	140	110
		k_a (m^2/g)	k_a (m^2/g)	k_a (m^2/g)
nutrient limited	14	0.364	0.364	0.295
	28	0.257	0.281	0.259
	92	0.14	0.144	0.17
nutrients in excess	39.5	0.15	0.16	0.207
	79	0.15	0.14	0.152
	158	0.11	0.11	0.14

Table S3: Light parameters *a* and *b* estimated (average ± standard deviation) for the nutrient limited, nutrients in-excess cultivation in synthetic medium and cultivation in treated water. Constitutive relation for light attenuation: $k_a = a * e^{-b*X_{Alg}}$.

	Nutrient limited	Nutrient in-excess	Treated water
a (m^2/g)	0.374±0.029	0.194±0.0079	0.094±0.003
b (g/m^3)	0.01±0.0017	0.0031±0.0004	0.0017±0.0001

Table S4: In-reactor light path length multiplier (PLM) calculated for the nutrient limited and nutrients in excess scenarios. Curve fitting was done by using the estimated k_a in the two scenarios and the Lambert-Beer equation. PLM was used to improve fit (based on R^2). The blue shading refers to scenarios where PLM is not needed.

	TSS (mg/l)	diameter (mm)			diameter (mm)			diameter (mm)		
		240	140	110	240	140	110	240	140	110
		PLM (-)			R^2 of fit without PLM			R^2 of fit with PLM		
nutrient limited	14	1.4	1.6	1.4	0.51	0.04	0.5	0.82	0.7	0.74
	28	1.1	1.3	1.3	0.86	0.69	0.79	0.88	0.83	0.87
	92	1	1.1	1.3	0.97	0.92	0.87	0.97	0.96	0.95
nutrients in excess	39.5	1	1	1.6	0.93	0.97	0.33	0.93	0.97	0.86
	79	1	1	1.1	0.93	0.94	0.9	0.93	0.94	0.96
	158	1	1	1.2	0.93	0.95	0.91	0.93	0.95	0.99

Table S5: The Lambert-Beer equation is fitted on the light curves and their fit is compared to the fitting with Schuster's law. The comparison is based on R^2 of the fit. The blue shading refers to scenarios where scattering is not relevant thus Lambert-Beer equation and Shuster's law give the same fitting.

	TSS (mg/l)	Diameter (mm)					
		240		140		110	
		E_a (m^2/g)	E_s (m^2/g)	E_a (m^2/g)	E_s (m^2/g)	E_a (m^2/g)	E_s (m^2/g)
nutrient limited	14	0.003	1.86	0.01	2.4	0.017	2.35
	28	0.019	1.26	0.016	1.63	0.023	1.64
	92	0.14	0	0.025	0.53	0.026	0.64
nutrients in excess	39.5	0.15	0	0.16	0	0.013	1.2
	79	0.14	0	0.14	0	0.021	0.64
	158	0.11	0	0.1	0	0.14	0
		R^2 of the fit with Lambert-Beer equation					
nutrient limited	14	0.82		0.7		0.74	
	28	0.88		0.83		0.87	
	92	0.98		0.96		0.95	
nutrients in excess	39.5	0.98		0.98		0.86	
	79	0.93		0.97		0.96	
	158	0.94		0.96		0.99	
		R^2 of the fit with Schuster's law					
nutrient limited	14	0.96		0.97		0.98	
	28	0.99		0.98		0.99	
	92	0.98		0.99		0.98	
nutrients in excess	39.5	0.98		0.98		0.99	
	79	0.93		0.97		0.99	
	158	0.94		0.96		0.99	

Table S6: Light parameters *a* and *b* estimated for Batch 1 defining the attenuation coefficient based on the TSS ($k_a = a * e^{-b*X_{Alg}}$). Light parameters *c* and *d* estimated for Batch 1 defining the attenuation coefficient based on the chlorophyll content ($k_{a,p} = \frac{d}{X_{Chl}} - c$).

	a (m^2/g TSS)	b (m^3/g TSS)	c (m^2/g Chl)	d (m^3/g Chl)
Batch 1	0.135±0.009	0.0018±0.0003	1.06±0.8	29.3±0.65

Table S7: RMSNE, AIC and ARILC values obtained with simulations using three different light modelling complexities, first using TSS to calculate the attenuation coefficient (k_a) and second using the chlorophyll content to calculate the pigment specific attenuation coefficient ($k_{a,p}$).

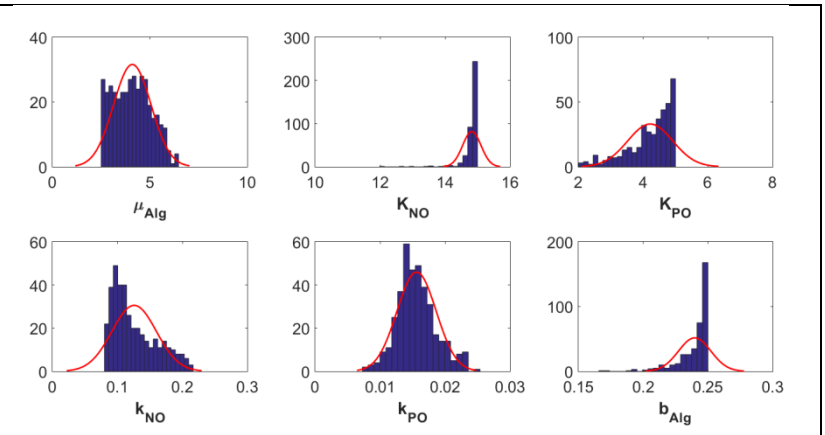
	calculated with k_a			calculated with $k_{a,p}$		
	Average light	Variable average light	Layer model	Average light	Variable average light	Layer model
RMSNE (-)						
X_{Alg}	0.167	0.157	0.092	0.149	0.159	0.094
X_{AlgN}	0.124	0.142	0.171	0.125	0.149	0.164
S_{NO3}	0.88	0.87	0.889	0.879	0.881	0.879
X_{AlgP}	0.281	0.284	0.229	0.277	0.24	0.293
S_{PO4}	0.803	0.885	1.036	0.775	1.379	0.692
sum	2.255	2.338	2.417	2.204	2.808	2.121
AIC (-)						
X_{Alg}	-97	-101	-134	-104	-100	-133
X_{AlgN}	-115	-107	-96	-115	-104	-98
S_{NO3}	6	5	7	6	6	6
X_{AlgP}	-65	-64	-77	-66	-75	-62
S_{PO4}	0.4	6	16	-2	33.9	-9
sum	-271	-260	-284	-281	-239	-296
ARILC (-)						
X_{Alg}	0.0043	0.0041	0.0011	0.0041	0.0039	0.0036
X_{AlgN}	0.003	0.0031	0.0007	0.0033	0.0033	0.0048
S_{NO3}	0.033	0.022	0.0074	0.026	0.02	0.018
X_{AlgP}	0.0081	0.0077	0.0032	0.0091	0.0071	0.0084
S_{PO4}	0.023	0.021	0.02	0.022	0.034	0.019
sum	0.071	0.057	0.033	0.065	0.068	0.054

Table S8: Comparison of the estimated parameter subsets using the three complexity levels. The values are presented as mean ± 95% confidence interval.

	calculated with k_a			calculated with $k_{a,p}$		
	CL-1 Average light	CL-2 Variable average light	CL-3 Layer model	CL-1 Average light	CL-2 Variable average light	CL-3 Layer model
$\mu_{A,max}$ (d^{-1})	4.1±1.91	3.81±1.51	6.2±0.53	4.08±1.69	5.28±1.8	5.5±1.27
$K_{NO,Alg}$ ($gN\ m^{-3}$)	14.83±0.56	14.59±1.53	14.61±1.46	14.86±0.4	14.52±1.68	14.82±0.42
$K_{PO4,Alg}$ ($gP\ m^{-3}$)	4.22±1.38	4.19±1.41	4.31±1.52	4.14±1.42	4.49±1.37	4.07±1.52
$k_{NO,Alg}$ ($gN\ g^{-1}COD\ d^{-1}$)	0.13±0.068	0.1±0.034	0.17±0.025	0.13±0.067	0.12±0.044	0.16±0.04
$k_{PO4,Alg}$ ($gN\ g^{-1}COD\ d^{-1}$)	0.016±0.006	0.015±0.005	0.014±0.004	0.016±0.006	0.011±0.003	0.018±0.006
b_{Alg} (d^{-1})	0.24±0.025	0.24±0.024	0.25±0.008	0.24±0.024	0.25±0.006	0.22±0.036

Table S9: Model calibration and identifiability analysis using the average constant light intensity-CL1. TSS was used to calculate the attenuation coefficient (k_a). Histograms obtained for the posterior parameter distribution and correlation matrix.

CL1-Average light



	$\mu_{A,max}$ (d^{-1})	$K_{NO,Alg}$ ($gN{\cdot}m^{-3}$)	$K_{PO4,Alg}$ ($gP{\cdot}m^{-3}$)	$k_{NO,Alg}$ ($gN{\cdot}g^{1}COD{\cdot}d^{-1}$)	$k_{PO4,Alg}$ ($gP{\cdot}g^{-1}COD{\cdot}d^{-1}$)	b_{Alg} (d^{-1})
$\mu_{A,max}$ (d^{-1})	1					
$K_{NO,Alg}$ ($gN{\cdot}m^{-3}$)	0.023	1				
$K_{PO4,Alg}$ ($gP{\cdot}m^{-3}$)	0.104	-0.018	1			
$k_{NO,Alg}$ ($gN{\cdot}g^{1}COD{\cdot}d^{-1}$)	-0.936	0.076	-0.152	1		
$k_{PO4,Alg}$ ($gP{\cdot}g^{-1}COD{\cdot}d^{-1}$)	-0.665	-0.038	0.61	0.663	1	
b_{Alg} (d^{-1})	0.237	-0.045	0.367	-0.319	0.099	1

Table S10: Model calibration and identifiability analysis using the average constant light intensity - CL1. The chlorophyll content was used to calculate the pigment specific attenuation coefficient ($k_{a,p}$). Histograms obtained for the posterior parameter distribution and correlation matrix.

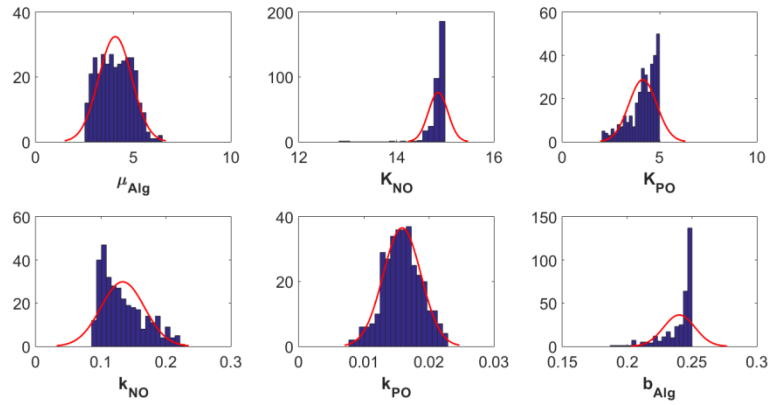
CL1- Average light						
	$\mu_{A,max}$ (d^{-1})	$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	$k_{NO,Alg}$ ($gN \cdot g^{1}COD \cdot d^{-1}$)	$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	b_{Alg} (d^{-1})
$\mu_{A,max}$ (d^{-1})	1					
$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	0.0798	1				
$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	0.091	0.035	1			
$k_{NO,Alg}$ ($gN \cdot g^{1}COD \cdot d^{-1}$)	-0.947	-0.019	-0.207	1		
$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	-0.639	-0.028	0.659	0.571	1	
b_{Alg} (d^{-1})	0.31	0.046	0.411	-0.385	0.148	1

Table S11: Model calibration and identifiability analysis using the variable average light intensity – CL2. TSS was used to calculate the attenuation coefficient (k_a). Histograms obtained for the posterior parameter distribution and correlation matrix.

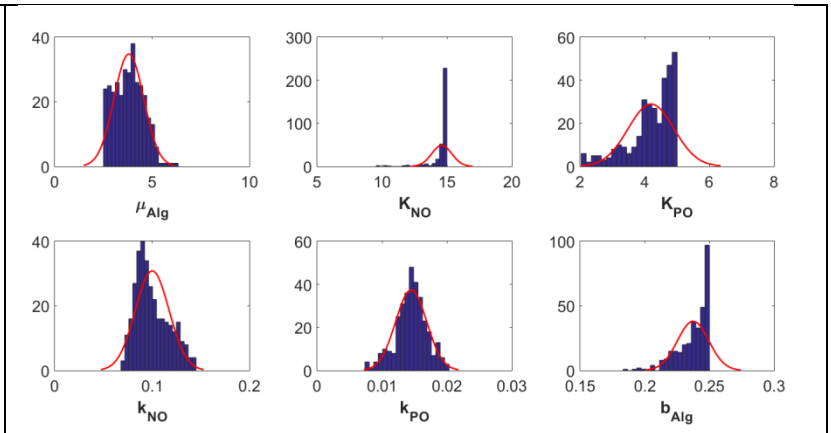
CL2- Variable average light						
	$\mu_{A,max}$ (d^{-1})	$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	$k_{NO,Alg}$ ($gN \cdot g^{1}COD \cdot d^{-1}$)	$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	b_{Alg} (d^{-1})
$\mu_{A,max}$ (d^{-1})	1					
$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	-0.081	1				
$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	-0.038	-0.022	1			
$k_{NO,Alg}$ ($gN \cdot g^{1}COD \cdot d^{-1}$)	-0.912	0.347	-0.1	1		
$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	-0.571	0.026	0.804	0.446	1	
b_{Alg} (d^{-1})	0.271	-0.030	0.278	-0.351	0.146	1

Table S12: Model calibration and identifiability analysis using the variable average light intensity – CL2. The chlorophyll content was used to calculate the pigment specific attenuation coefficient ($k_{a,p}$). Histograms obtained for the posterior parameter distribution and correlation matrix.

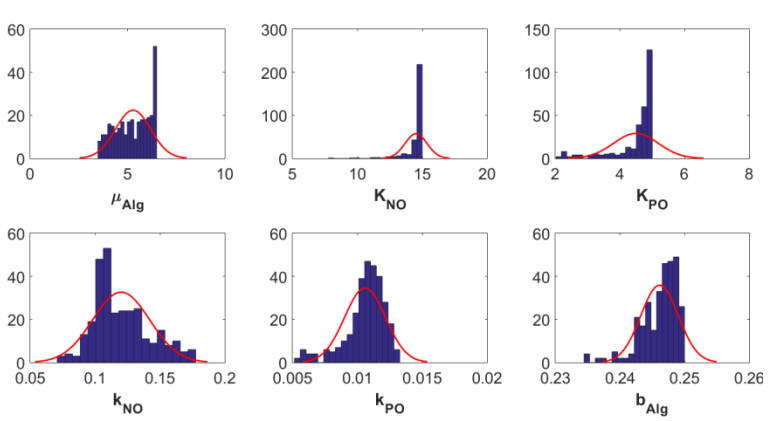
CL2- Variable average light						
	$\mu_{A,max}$ (d^{-1})	$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	$k_{NO,Alg}$ ($gN \cdot g^{1}COD \cdot d^{-1}$)	$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	b_{Alg} (d^{-1})
$\mu_{A,max}$ (d^{-1})	1					
$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	-0.172	1				
$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	-0.021	-0.096	1			
$k_{NO,Alg}$ ($gN \cdot g^{1}COD \cdot d^{-1}$)	-0.882	0.376	-0.233	1		
$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	-0.491	-0.057	0.807	0.195	1	
b_{Alg} (d^{-1})	0.206	0.25	0.131	-0.229	0.008	1

Table S13: Model calibration and identifiability analysis using the layer model – CL3. TSS was used to calculate the attenuation coefficient (k_a). Histograms obtained for the posterior parameter distribution and correlation matrix.

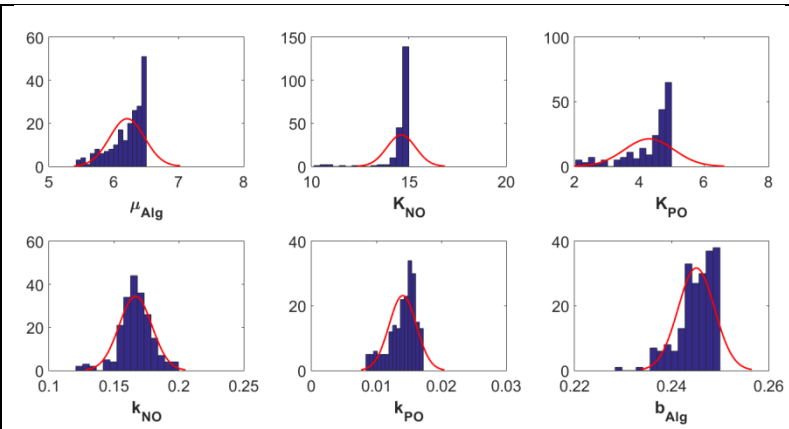
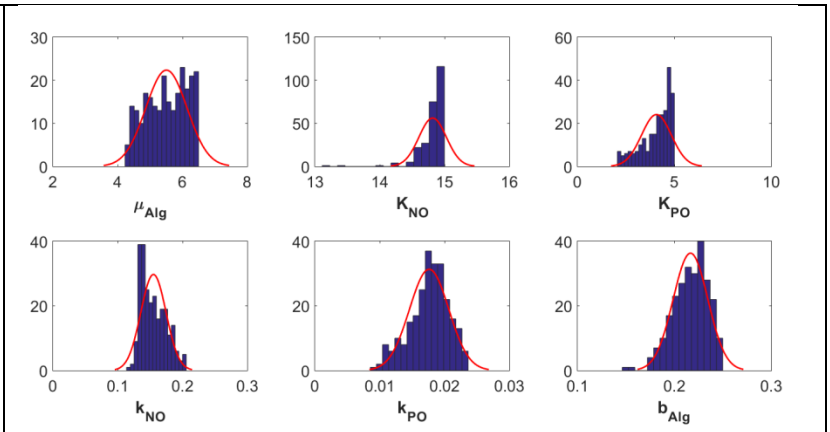
CL3- Layer model						
	$\mu_{A,max}$ (d^{-1})	$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	$k_{NO,Alg}$ ($gN \cdot g^{-1}COD \cdot d^{-1}$)	$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	b_{Alg} (d^{-1})
$\mu_{A,max}$ (d^{-1})	1					
$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	0.005	1				
$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	-0.116	-0.003	1			
$k_{NO,Alg}$ ($gN \cdot g^{-1}COD \cdot d^{-1}$)	-0.213	0.596	-0.423	1		
$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	-0.293	-0.026	0.931	-0.518	1	
b_{Alg} (d^{-1})	0.186	0.195	-0.006	0.034	0.041	1

Table S14: Model calibration and identifiability analysis using the layer model – CL3. The chlorophyll content was used to calculate the pigment specific attenuation coefficient ($k_{a,p}$). Histograms obtained for the posterior parameter distribution and correlation matrix.

CL3- Layer model						
	$\mu_{A,max}$ (d^{-1})	$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	$k_{NO,Alg}$ ($gN \cdot g^{1}COD \cdot d^{-1}$)	$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	b_{Alg} (d^{-1})
$\mu_{A,max}$ (d^{-1})	1					
$K_{NO,Alg}$ ($gN \cdot m^{-3}$)	0.115	1				
$K_{PO4,Alg}$ ($gP \cdot m^{-3}$)	0.035	-0.017	1			
$k_{NO,Alg}$ ($gN \cdot g^{1}COD \cdot d^{-1}$)	-0.961	-0.008	-0.167	1		
$k_{PO4,Alg}$ ($gP \cdot g^{-1}COD \cdot d^{-1}$)	-0.401	-0.099	0.867	0.28	1	
b_{Alg} (d^{-1})	0.459	0.042	0.324	-0.418	0.246	1

Figures

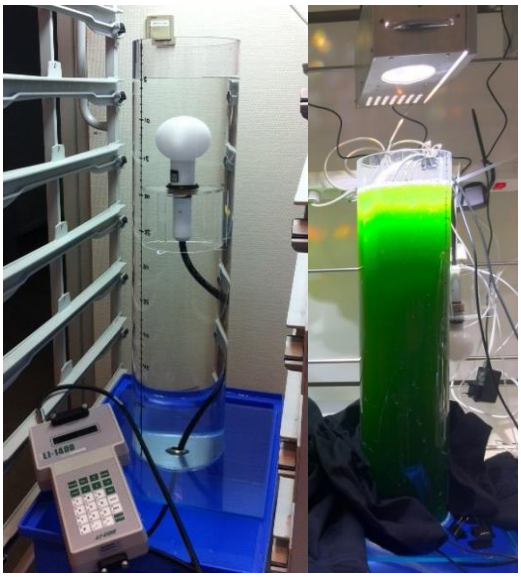


Figure S1: The cylindrical shaped clear walled plastic reactor used for the experiments, with the light sensor inside, connected to a data logger (picture on the left). The 8-L reactor used for the batch experiments with the custom built lamp mounted above (picture on the right). The black cloth on the bottom was used to cover the reactor wall from the side.

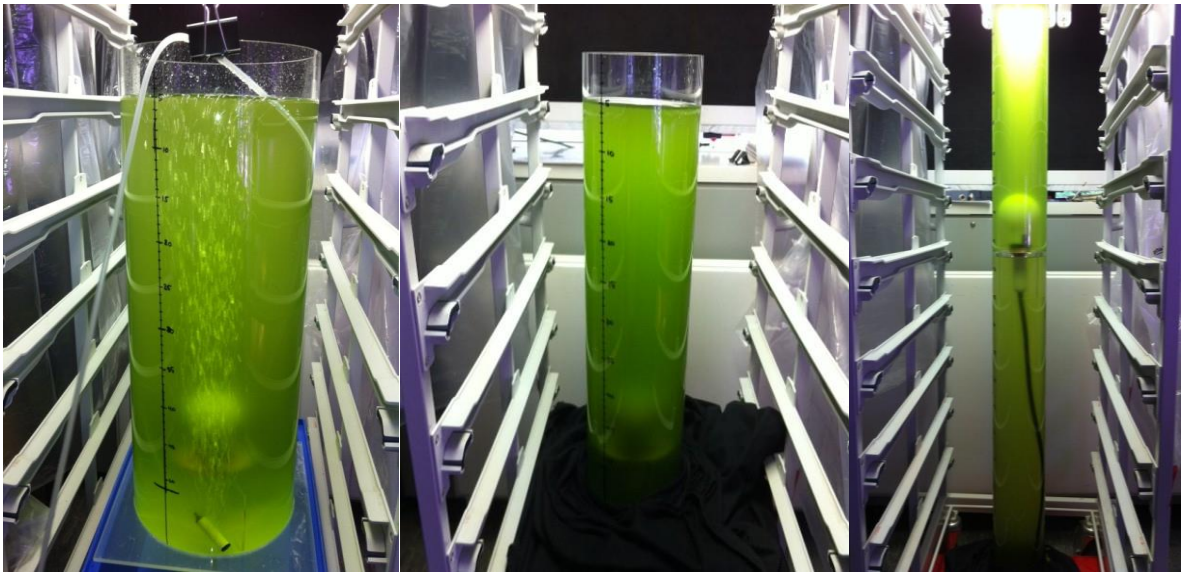


Figure S2: The cylindrical shaped clear walled plastic reactors with three different diameters used for the experiments. Reactor 1 with a diameter of 240 mm (picture on the left). Reactor 2 with a diameter of 140 mm (picture in the middle). Reactor 3 with a diameter of 110 mm (picture on the right).

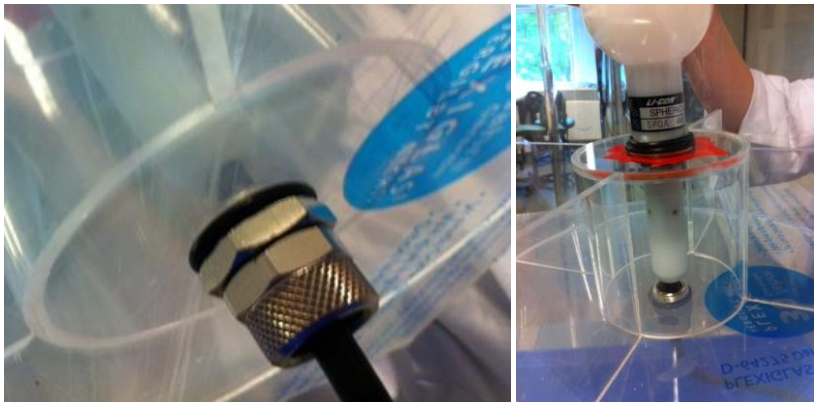


Figure S3: The fitting on the bottom of the reactor, used to mount the cable of the light sensor (picture on the left). The plastic fitting, used to keep the light sensor in a vertical upward position inside the reactor (picture on the right).

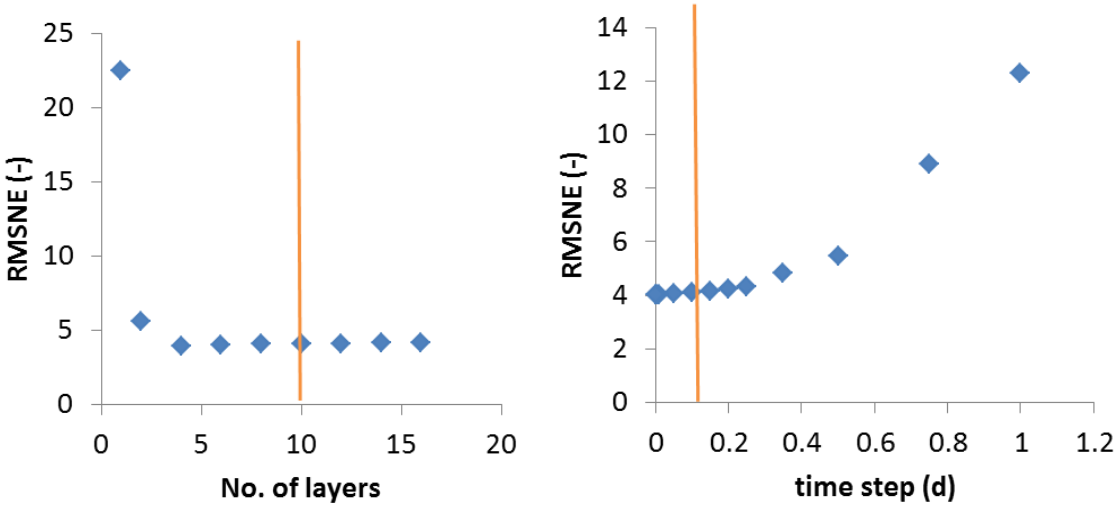


Figure S4: Evaluation of optimal number of layers and optimal time-step of the layer model based on the RMSNE of the simulation. The RMSNE was calculated by comparing the simulation to the experimental data in Batch 1.

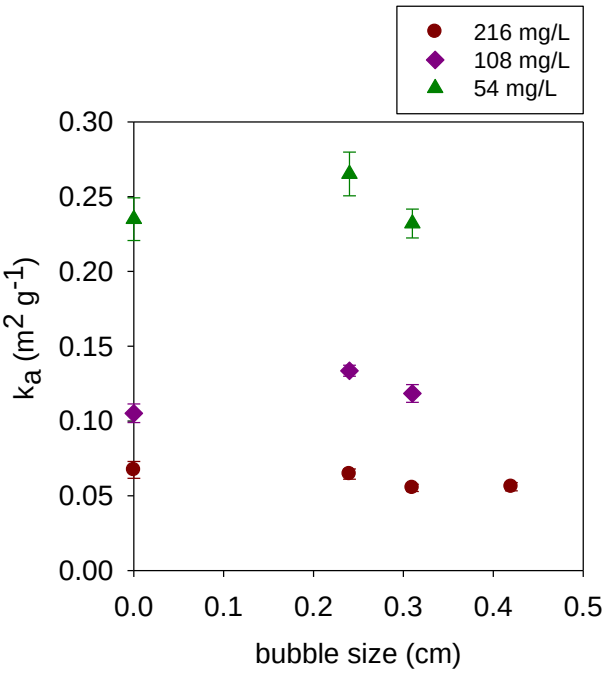


Figure S5: Light attenuation inside the PBR with 140 mm diameter assessing the effect of bubble size.

Figure S6: Colour change due to nutrient limited conditions (see Fig. 2a). The left metal plate contains a glass-fibre filter that has a deep green colour, due to high chlorophyll content at nutrients in excess conditions (see Fig. 2b). The metal plate on the right contains a filter that has yellowish colour due to the increase in carotenoid level under nutrient limited conditions.

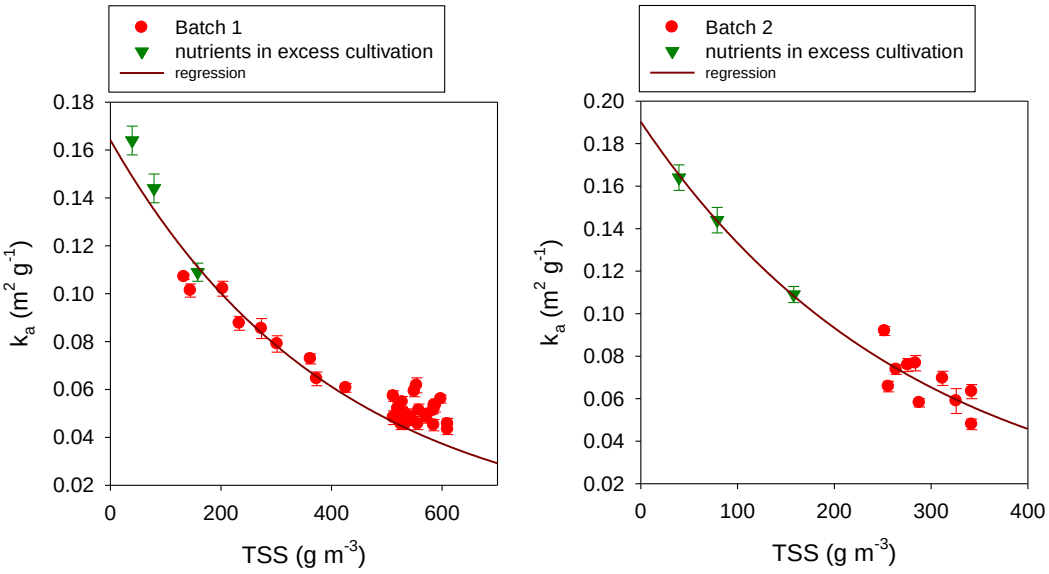


Figure S7: Attenuation coefficient during the batch cultivation as a function of biomass concentration. The figures include the data for nutrients in excess cultivation, for comparison with the batches.

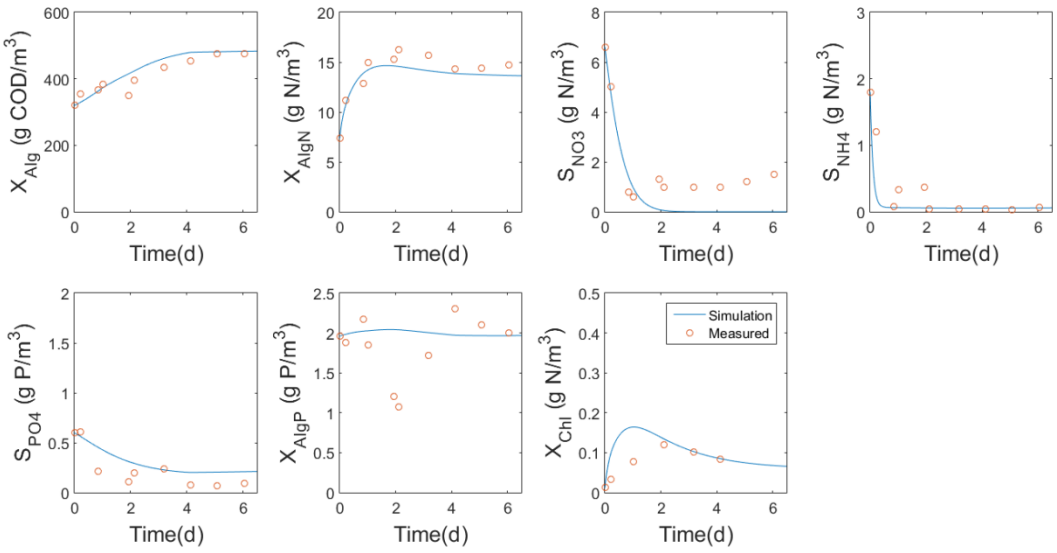


Figure S8: Simulations of Batch 2 using the parameter set estimated in Batch 1.

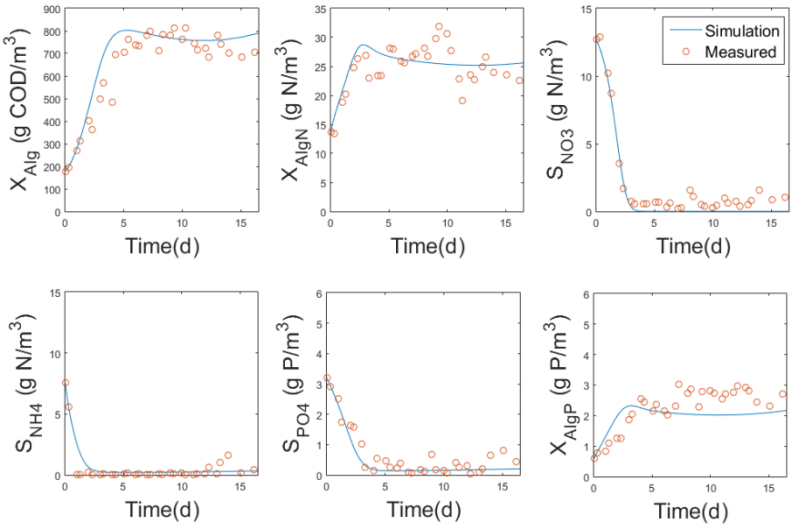


Figure S9: Simulation using model CL-1 (average constant light intensity, 127 µmol $m^{-2}s^{-1}$) with the mean values of the paremeters estimated. TSS was used to calculate the attenuation coefficient (k_a).

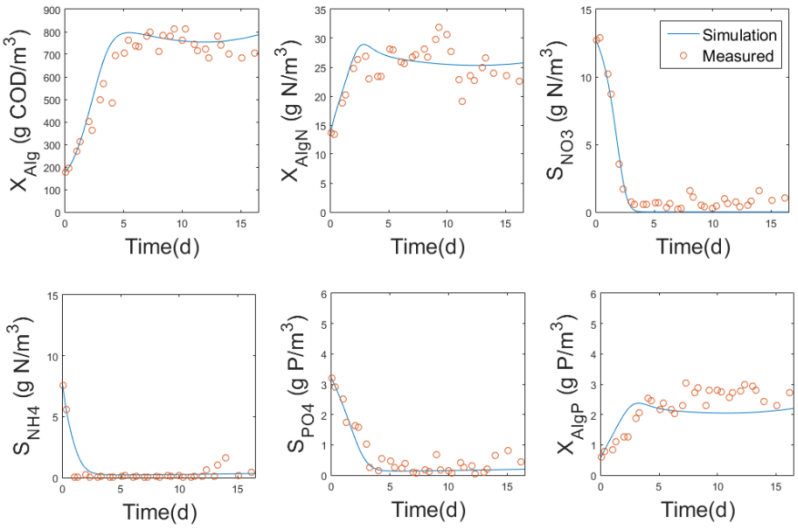


Figure S10: Simulation using model CL-1 (average constant light intensity, 118 µmol $m^{-2}s^{-1}$) with the mean values of the paremeters estimated. The chlorophyll content was used to calculate the pigment specific attenuation coefficient ($k_{a,p}$).

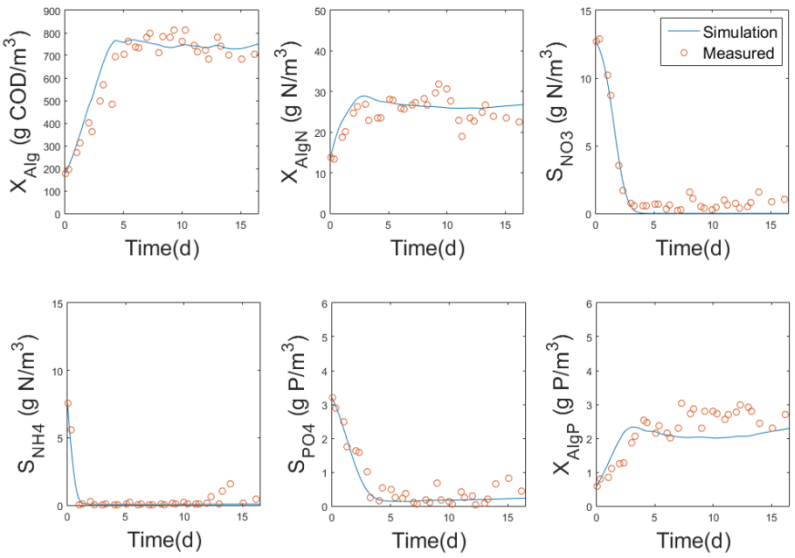


Figure S11: Simulation using model CL-2 (time-variable light intensity) with the mean values of the paremeters estimated. TSS was used to calculate the attenuation coefficient (k_a).

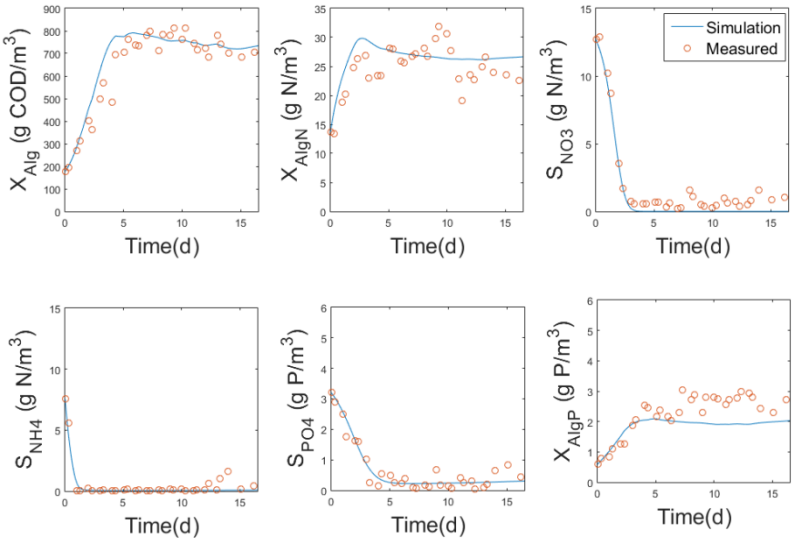


Figure S12: Simulation using model CL-2 (time-variable light intensity) with the mean values of the paremeters estimated. The chlorophyll content was used to calculate the pigment specific attenuation coefficient ($k_{a,p}$).

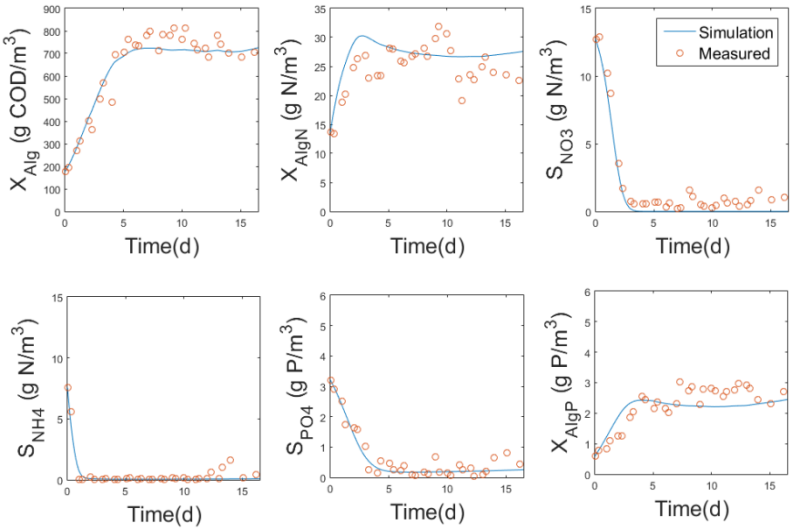


Figure S13: Simulation using model CL-3 (discretized layer model) with the mean values of the paremeters estimated. TSS was used to calculate the attenuation coefficient (k_a).

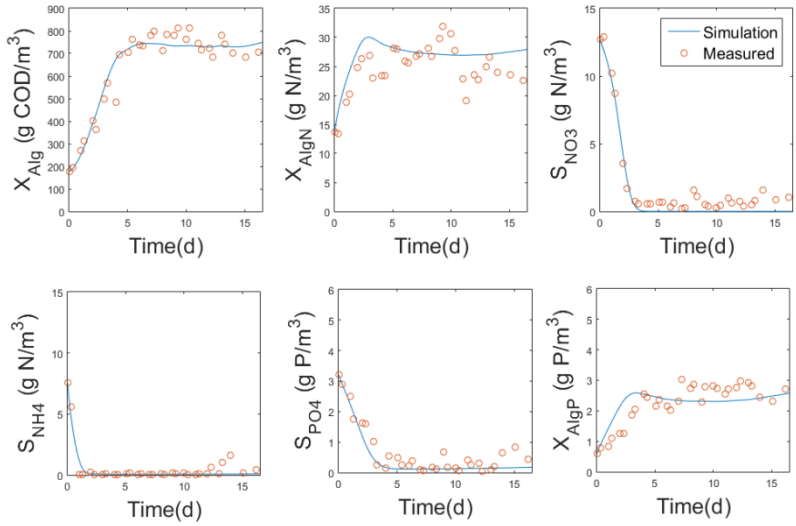


Figure S14: Simulation using model CL-3 (discretized layer model) with the mean values of the paremeters estimated. The chlorophyll content was used to calculate the pigment specific attenuation coefficient ($k_{a,p}$).

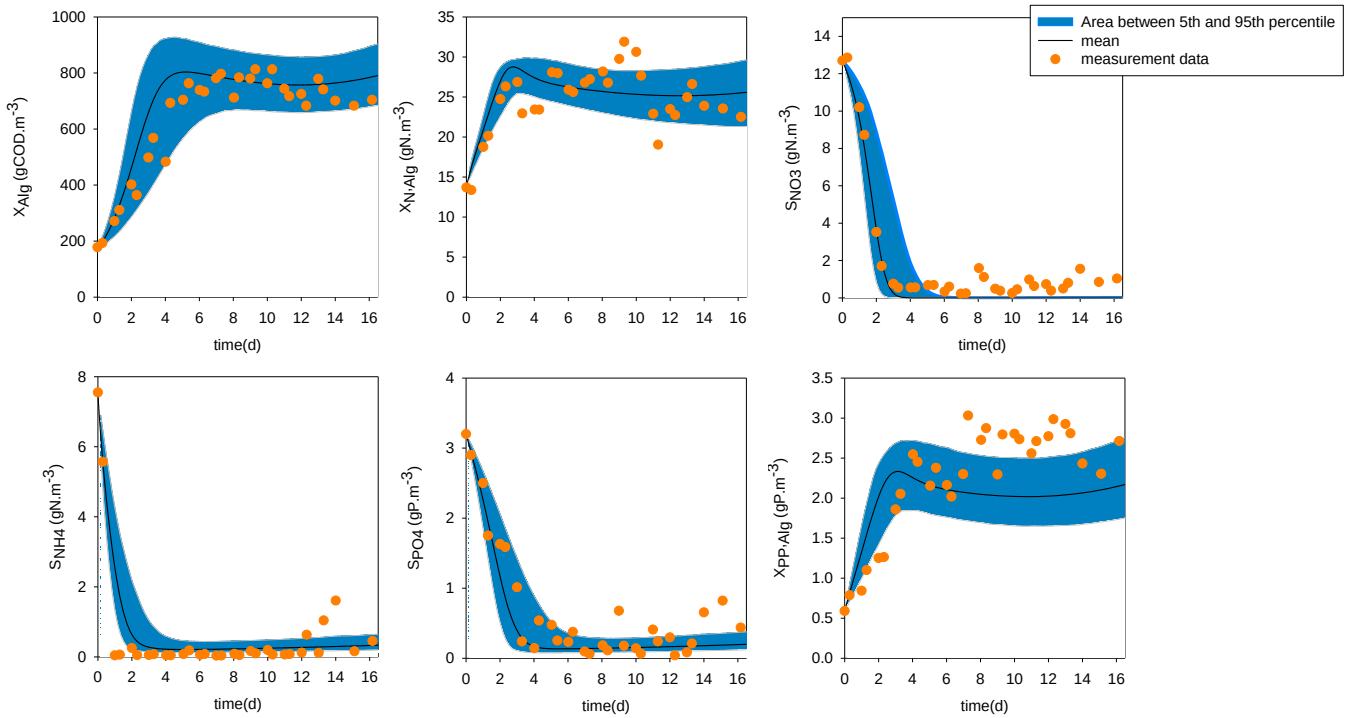


Figure S15: Simulation using model CL-1 (average constant light intensity, 127 µmol $m^{-2}s^{-1}$) with the mean values of the parameters estimated. The uncertainty bands are shown in blue. The TSS is used to calculate the attenuation coefficient (k_a).

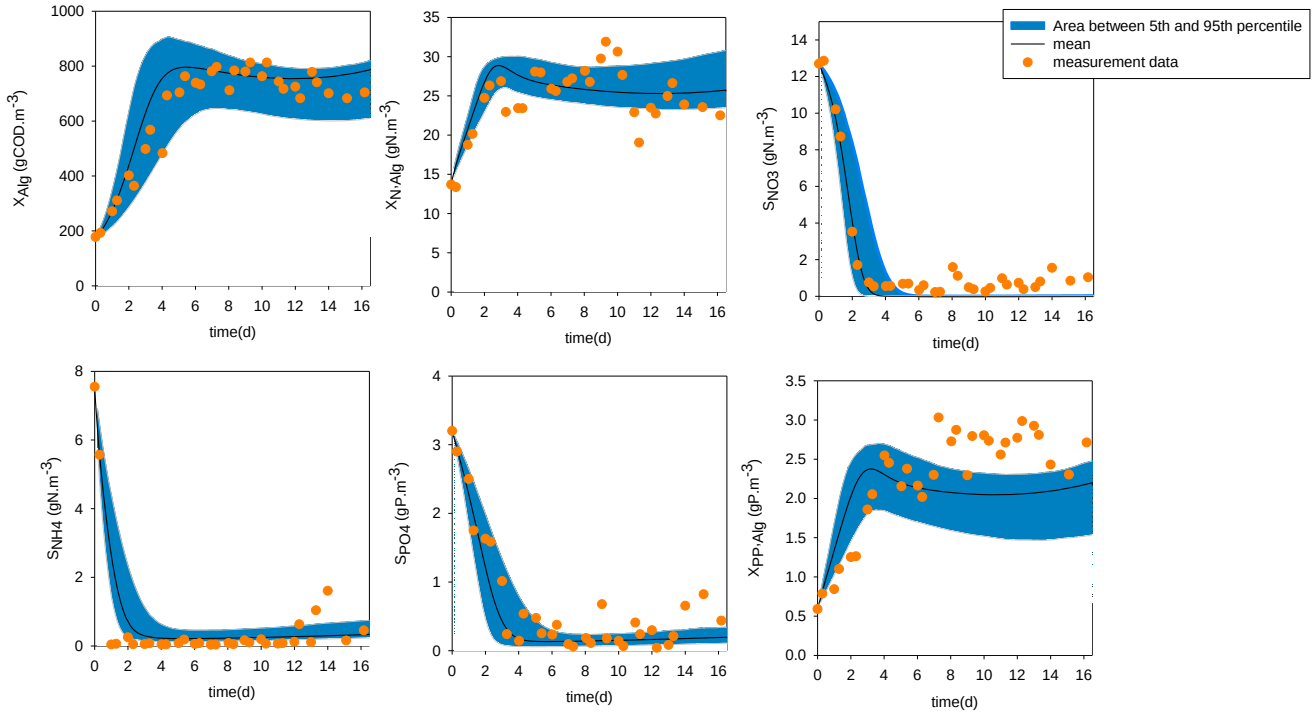


Figure S16: Simulation using model CL-1 (average constant light intensity, 118 µmol $m^{-2}s^{-1}$) with the mean values of the parameters estimated. The uncertainty bands are shown in blue. The chlorophyll content is used to calculate the pigment specific attenuation coefficient ($k_{a,p}$).

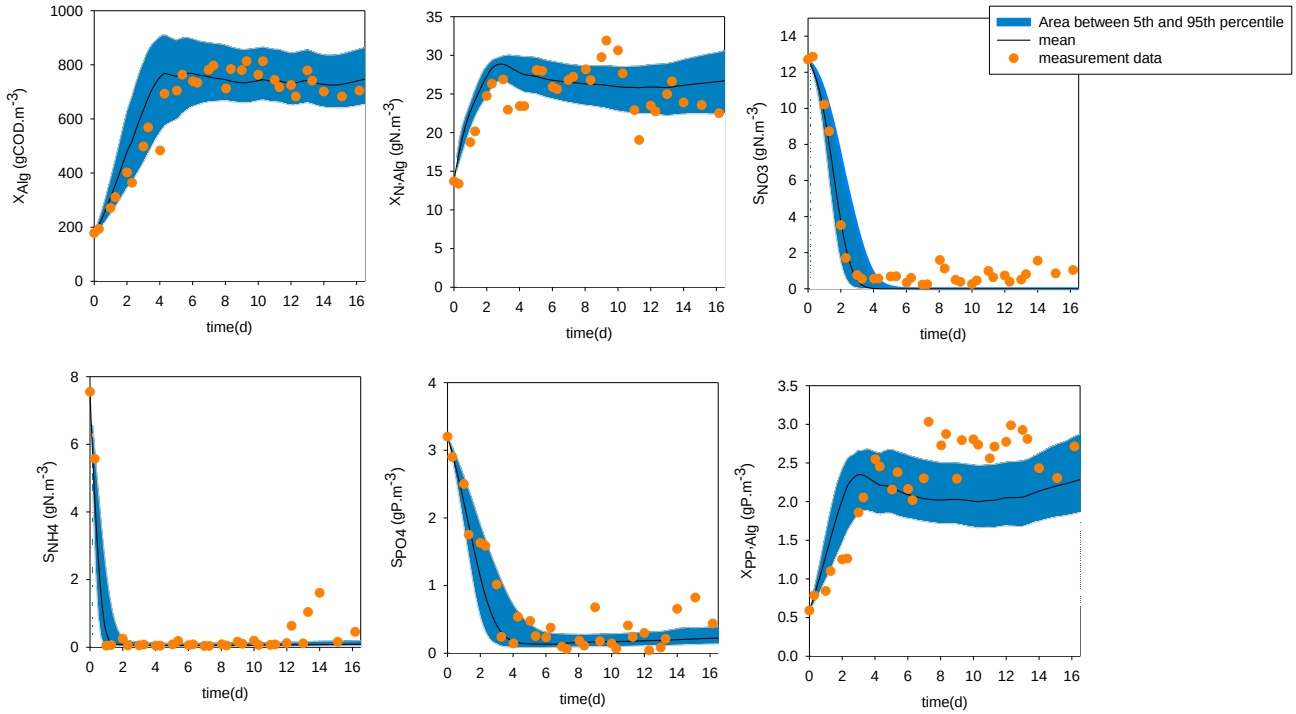


Figure S17: Simulation using model CL- 2 (time-variable light intensity) with the mean values of the parameters estimated. The uncertainty bands are shown in blue. The TSS is used to calculate the attenuation coefficient (k_a).

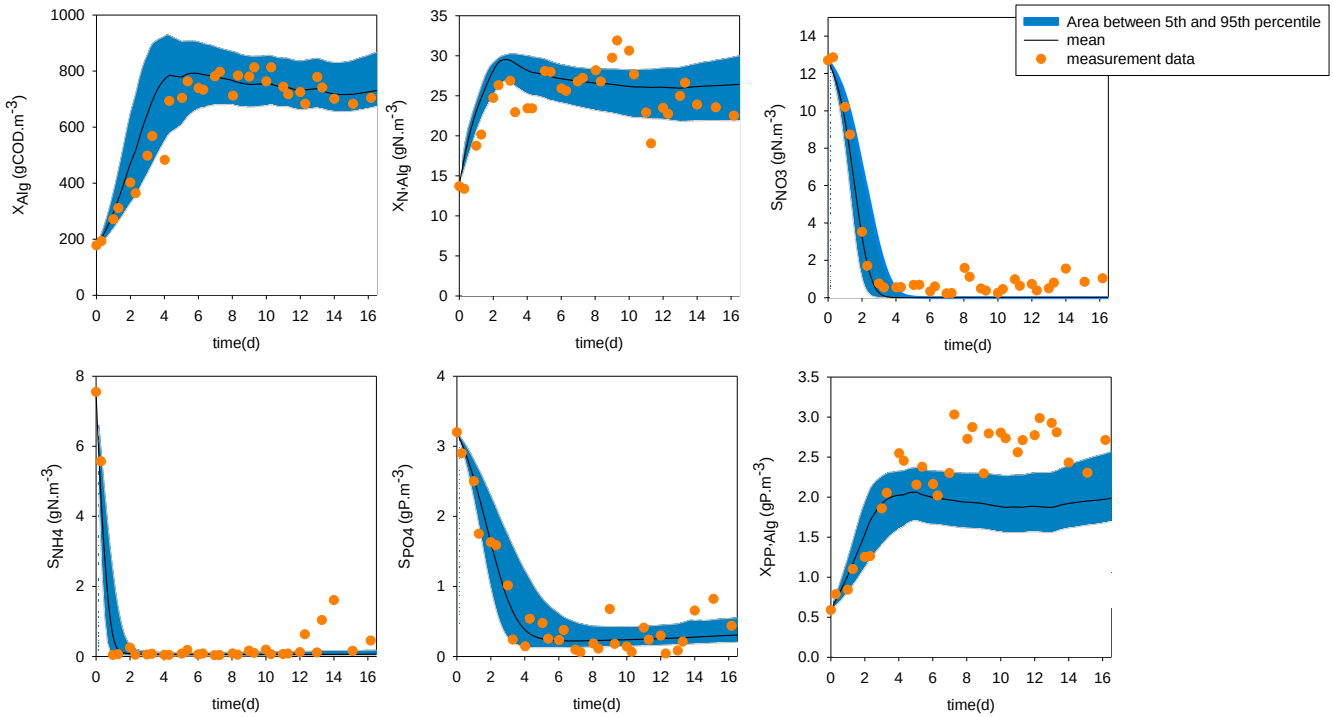


Figure S18: Simulation using model CL- 2 (time-variable light intensity) with the mean values of the parameters estimated. The uncertainty bands are shown in blue. The chlorophyll content is used to calculate the pigment specific attenuation coefficient ($k_{a,p}$).

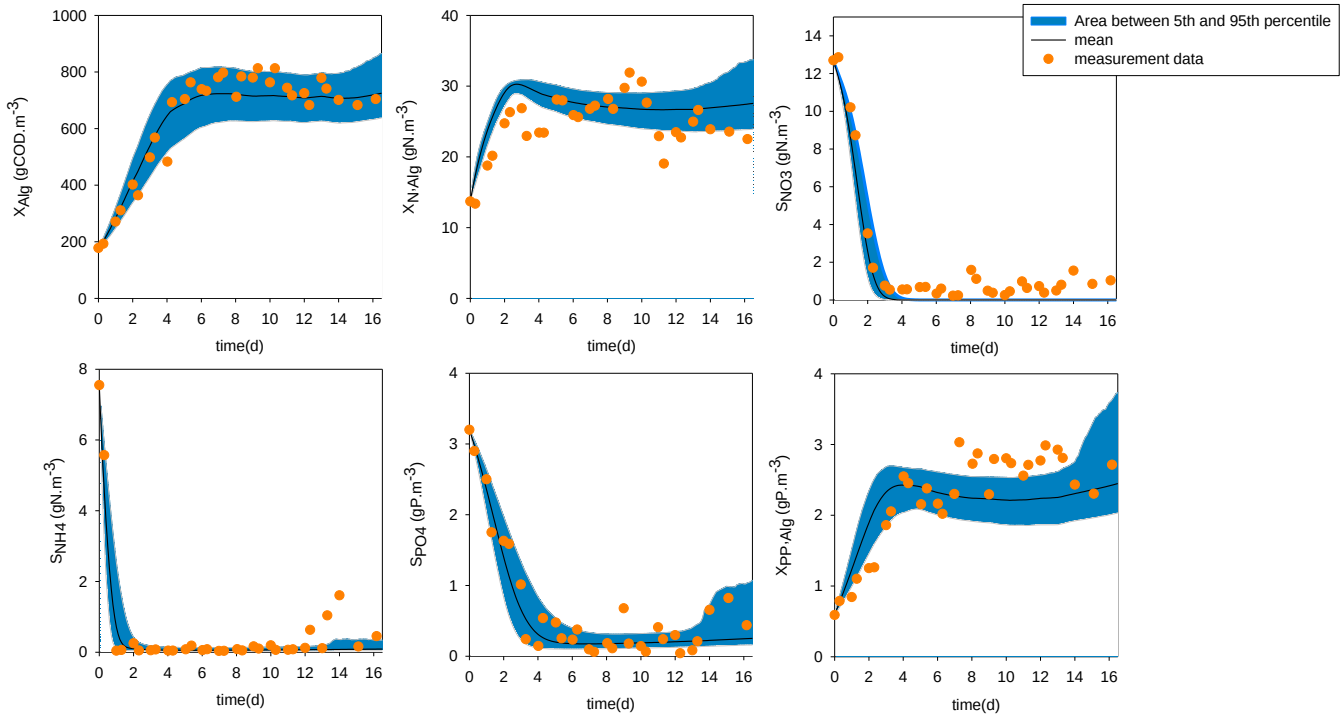


Figure S19: Simulation using model CL- 3 (one-dimensional layer model) with the mean values of the parameters estimated. The uncertainty bands are shown in blue. The TSS is used to calculate the attenuation coefficient (k_a).

References

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