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The cost of toxin production in phytoplankton: the case of PST producing dinoflagellates

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1	Title of the paper: The cost of toxin production in phytoplankton: the case of
2	PST producing dinoflagellates
3	Running title: Cost of toxin production
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19 Abstract

20 Many species of phytoplankton produce toxins that may provide protection from 21 grazing. In that case one would expect toxin production to be costly; else all species 22 would evolve toxicity. However, experiments have consistently failed to show any 23 costs. Here, we show that costs of toxin production are environment dependent but 24 can be high. We develop a fitness optimization model to estimate rate, costs, and 25 benefits of toxin production, using PST (paralytic shellfish toxin) producing 26 dinoflagellates as an example. Costs include energy and material (nitrogen) costs 27 estimated from well-established biochemistry of PSTs, and benefits are estimated 28 from relationship between toxin content and grazing mortality. The model reproduces 29 all known features of PST production: inducibility in the presence of grazer cues, low 30 toxicity of nitrogen-starved cells, but high toxicity of P- and light-limited cells. The 31 model predicts negligible reduction in cell division rate in nitrogen replete cells, 32 consistent with observations, but >20% reduction when nitrogen is limiting and 33 abundance of grazers high. Such situation is characteristic of coastal and oceanic 34 waters during summer when blooms of toxic algae typically develop. The investment 35 in defense is warranted, since the net growth rate is always higher in defended than in 36 undefended cells.

37 Key words

38 Defense; Trade-off; Environment-dependent cost

39

40

41

42 Introduction

43 Many phytoplankton species produce substances that are toxic to humans, hence we 44 consider such phytoplankton 'toxic algae', but the evolution and functional role of 45 these secondary metabolites remain unclear. They may be released into the 46 environment and have allelochemical effects to combat competitors [1, 2] or grazers 47 [3], they may be mainly intracellular and have toxic and/or deterrent effects on 48 grazers [4, 5], or in mixotrophic species they may have offensive roles, functioning as 49 a venom towards prey [6–8]. Some toxic algae may also form dense blooms, 50 potentially promoted by their toxicity and consequent grazer deterrent effects, and a 51 defensive role of toxin production is thus often assumed [4, 9]. This interpretation is 52 supported by the observations that algal toxins often have a deterrent rather than a 53 toxic effect on grazers [10–15], and that toxin production may be upregulated in the presence of grazers or their cues, as demonstrated in dinoflagellates, Alexandrium 54 55 spp. [13, 16, 17], and in diatoms, *Pseudo-nitzschia* spp. [18]. The latter observation 56 also suggests that toxin production comes at a cost – why else regulate the production 57 in response to the need? Optimal defense theory, well founded in terrestrial plant 58 ecology, predicts that inducibility of defense should evolve only when the defense is 59 costly and variable in time [19]. However, experiments have generally been unable to 60 demonstrate such costs in toxic phytoplankton, and the growth rates of grazer-induced 61 and un-induced cells, or of toxic and nontoxic strains of the same phytoplankton 62 species appear to be similar [3, 13, 16, 18], and model exercises have similarly 63 suggested costs to be trivial [20]. However, if defense via toxin production is both 64 effective and costless, one would expect most phytoplankton to be toxic, which is far from being the case [19]. Also, the promotion of phytoplankton diversity in the ocean 65 due to grazing and consequent evolution of defense mechanisms, as demonstrated in 66

both theoretical models and considerations [21–23] and whole-community

68 experiments [24], functions only if the defense comes at a cost.

69 One of the most studied groups of toxic phytoplankton is the dinoflagellate 70 Alexandrium spp. since the toxins it produces, Paralytic Shellfish Toxins (PSTs), have 71 serious effects on humans. The main potential grazers of dinoflagellates are copepods. 72 While the toxin production may be induced by grazer (copepod) cues [13], the 73 production also depends on the nutrient status of the algae. Specifically, when 74 nitrogen or carbon (light) is limiting the PST production is reduced, while nutrient 75 balanced or P-limited cells may produce PSTs at a high rate (growth of P-limited cells 76 decreases while the production of toxins continues) [25, 26]. PSTs contain large 77 amounts of nitrogen, and so this dependency suggests that the costs of PST 78 production only become obvious when N or light is limiting, and may become 79 manifest either as a reduction in growth rate because part of the assimilated N goes 80 towards PST production; or as a cessation of the production of PSTs in favor of a 81 higher growth rate with a consequent loss of the defense; or a combination of the two. 82 Here, we explore by means of a simple resource allocation model the costs of 83 chemical defense in phytoplankton, using PST producing dinoflagellate Alexandrium 84 spp. as an example. We consider both energy and material costs of PST production, 85 their dependency on environmental conditions, and the reduction in mortality that is achieved by the defense investments. Through fitness optimization we demonstrate 86 87 that costs can be substantial, leading to > 20% reduction in cell division rate, when 88 grazer abundance is high and nitrogen availability is low. We also examine the 89 conditions under which the production of toxins provides maximum benefit to the 90 toxin producing species.

91 Model description:

92

93 et al. [27]. The division rate of phytoplankton depends on the acquisition of three 94 resources, viz. carbon (via photosynthesis), nitrogen (as nitrate), and phosphorous (as 95 phosphate), as well as on metabolic expenses. The cells may invest some of their 96 assimilated nitrogen into toxin production and combust some of their fixed organic 97 carbon to cover costs of toxin synthesis, and in return experience reduced grazing 98 mortality. We search for the investment that maximizes the fitness of the cells, 99 defined as the difference between cell division rate and mortality rate (= net growth 100 rate). We use the model to explore the dependency of division rate, net growth rate, 101 and toxicity on the environmental resource availability and predation risk. 102 Uptake of carbon, nitrogen and phosphorous to the cell is described by the symbol J_i 103 (mass flows *i* being carbon via light-dependent photosynthesis (C), nitrogen (N) or 104 phosphorous (P) in units of μg per day; see Table 1 for central symbols and 105 parameters), and are combined to synthesize new biomass (Fig. 1). Respiratory costs, R_{C} (units of µg C per day), include costs of biomass synthesis (incl. transport) and 106 maintenance of the structure. Toxin is produced at a rate, T_r , and implies an additional 107 respiratory cost, R_T . Biomass synthesis rate, J_{tot} , is constrained by the stoichiometric 108 109 balance between carbon, nitrogen and phosphorous. Finally, we assume that toxins 110 and structure have constant but different stoichiometry.

The model is based on a resource allocation optimization model modified from Berge

111 Uptake of carbon, nitrogen, and phosphorous

112 The potential uptake J_i of resource i (C, N, P) is governed by a standard saturating
113 functional response:

$$J_i = M_i \frac{A_i Y_i}{A_i Y_i + M_i}, (1)$$

114 where A_i is the affinity for resource *i*, Y_i resource concentration (μ g L⁻¹), and M_i is 115 the maximum uptake rate.

116 Costs

117 Respiratory costs include costs of both uptake and mobilization of resources for

118 synthesis through each pathway, and the maintenance of the structure. This metabolic

119 cost is assumed to be 30% of the total carbon budget [28] plus a constant basal

120 respiration (R_0) independent of J_c , i.e.,

$$R_C = 0.3 J_C + R_0.$$
 (2)

121 Rate and cost of toxin production

122 Let θ be the fraction of nitrogen uptake that a toxic phytoplankton cell devotes to

123 toxin production. Then the potential rate of toxin production is (units of μ g N d⁻¹):

124
$$T_{pot}(\theta) = \theta J_N.$$
(3)

As the toxin production needs carbon both for building the toxin molecules and to
fuel the respiratory costs of toxin production, the actual toxin production rate may be
limited by the available carbon to:

128
$$T_r(\theta) = \min[T_{pot}(\theta), (J_C - R_C)/(n_T + r_T)], (4)$$

129 where n_T is the mass of carbon need per mass of nitrogen in the toxin, and r_T is the 130 respiratory cost per nitrogen synthesized into toxins (units of g C (g N)⁻¹). 131 The total cost of toxin production in terms of carbon then becomes (μ g C d⁻¹):

132
$$R_T(\theta) = (n_T + r_T)T_r(\theta). \quad (5)$$

133 Synthesis and growth rate

134 The assimilated carbon, nitrogen and phosphorous are combined to synthesize new structure. We assume constant C:N mass ratio, Q_{CN} (units of $\mu g C (\mu g N)^{-1}$) and C:P 135 mass ratio, Q_{CP} (units of $\mu g C (\mu g P)^{-1}$) of the cell. The total available carbon for 136 growth is then $J_C - R_C - R_T$ where J_C represents the total uptake of carbon through 137 photosynthesis, and R_C and R_T represent the costs of maintenance and biomass 138 139 synthesis, and costs of toxin production, respectively. The carbon required to 140 synthesize biomass from nutrients is $Q_{CN}(J_N - T_r)$ and $Q_{CP}J_P$ for nitrate and 141 phosphate, respectively. The growth rate is constrained by the limiting resource 142 (Liebig's law of the minimum) such that the total flux of carbon (and nutrients) 143 available for growth J_{tot} is:

$$J_{\text{tot}}(\theta) = \min[J_C - R_C - R_T(\theta), \ Q_{CN}(J_N - T_r(\theta)), \ Q_{CP}J_P].$$
(6)

144 Synthesis is not explicitly limited by a maximum synthesis capacity; limitation of 145 synthesis is taken care of by the limitation of uptake of carbon, nitrogen and 146 phosphorous in the functional responses (Eq. 1). The division rate μ of the cells (d⁻¹) 147 is the total flux of carbon divided by the carbon mass of the cell (w_X):

148
$$\mu(\theta) = J_{\text{tot}}(\theta)/w_X.$$
 (7)

149 Further subtracting the predation mortality (m_p) yields the net growth rate (r):

150
$$r(\theta) = \mu(\theta) - m_p(\theta). \quad (8)$$

We assume that predation mortality increases linearly with zooplankton biomass (Z),
and due to the toxin production, zooplankton reduces its grazing pressure on toxic
cells exponentially as:

154
$$m_p(\theta) = m_{p,0} Z e^{-\beta T(\theta)}, \quad (9)$$

where $m_{p,0}$ is a mortality constant, *T* is the cellular toxin content (μ g N cell⁻¹)

estimated as the toxin production rate divided by the cell division rate (= T_r/μ), and

157 β represents the strength of toxic effect.

158 The resulting population growth rate, $r(\theta)$, is a measure of the fitness of the

159 phytoplankton and we assume that the cell has the ability to optimize its growth rate

160 by regulating its resource allocation to toxin production such that it maximizes its

161 fitness. The optimal proportion of assimilated N devoted to toxin production then

162 becomes:

163
$$\theta^* = \arg\max_{\theta} \{r(\theta)\}.$$
(10)

164 Model parameterization

- 165 Calibration of parameters is based on laboratory measurements on the dinoflagellates
- 166 Alexandrium minutum and A. tamarense as the toxic species, and the copepods
- 167 *Acartia clausi* and *A. tonsa* as the grazer zooplankton. To calibrate the basic grazing
- 168 parameters, we use data for non-toxic strains of *A. minutum*.

169 *Parameters related to phytoplankton division rate* (μ) *:*

170 We use experimental observations reported in the literature for cell division rate (μ)

171 of A. minutum as a function of light intensity (L) and nutrient concentrations (N and

172 P) to estimate parameter values for maximum uptake rates (M_i) and affinities (A_i)

173 (Fig. 2a-c). While calibrating these parameters a non-toxic strain of A. minutum was

174 considered, and as a result, no cost is deducted. For calibration, we adjust the

175 parameters manually to fit the curves with data and keep them close to the existing

176 values of the parameters from other studies (when available). Due to Liebig's

177 minimum law for synthesis (eq. 6), the synthesis is limited by one of the resources

178 (either C or N) and further growth cannot materialize in spite of the availability of

179 other non-limiting resource. As a result, growth cannot increase any further.

180 We use a constant value $1.07 \times 10^{-4} \mu \text{g C d}^{-1}$ for the basal respiration rate (R_0) taken 181 from the range reported in Frangoulis *et al.* [29].

182 *Parameters related to the cost of toxin production:*

183 To estimate the two parameters related to the cost of toxin production (n_T, r_T) , we

184 consider the stoichiometry of PSTs and the biochemistry of PST synthesis. PSTs

185 produced by *Alexandrium* spp. (and other organisms) consist of saxitoxin and

186 multiple derivatives; they are cyclic nitrogenous compounds that are synthesized from

187 amino acid precursors. Here we consider the synthesis of saxitoxin, one of the

188 dominating toxins, from the amino acid glutamate via arginine to estimate the

approximate costs of toxin production. The costs are two-fold; i.e. the metabolic cost

190 of biosynthesis, r_T , and the cost in terms of material invested in the toxin, n_T .

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191 Material investment (n_T): The molecular formula of saxitoxin in C<sub>10</sub>H<sub>17</sub>N<sub>7</sub>O<sub>4</sub>, that is
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192 10 moles of carbon per 7 moles of nitrogen, or $n_T = (10x12)/(7x14) = 1.23 \ \mu g C \ (\mu g$

193 N)⁻¹.

194 *Metabolic expenses* (r_T) : Saxitoxin is synthesized from arginine [30], which in turn is typically synthesized from glutamate that in phytoplankton has to be synthesized *de* 195 196 *novo*. The metabolic cost of converting glutamate to arginine is 12 mol ATP per mol 197 of arginine, and the synthesis of arginine precursors (i.e. glutamate and carbamoyl-P) requires another 32 mol of ATP, i.e. a total of 44 mol ATP per mol of arginine [31]. 198 199 We have no estimate of the cost of synthesizing saxitoxin from arginine (e.g., 200 necessary transcripts and translation to enzymes) as well as costs due to actual or potential autotoxicity (hence we ignore it), but it takes 3 mol of arginine to synthesize 201 202 one mol of saxitoxin. Therefore, it requires at least 132 mol of ATP to synthesize 1 203 mol of saxitoxin. The respiratory equivalent of ATP synthesis is about 0.235 mol ATP 204 synthesized per liter of oxygen consumed; or about 2 g of organic carbon combusted 205 per mol of ATP synthesized [32]. Thus, 2x132 g of organic carbon is respired per mol of toxin synthesized. With 7x14 g N per mol toxin yields an estimate of $r_T = 2.7 \ \mu g C$ 206 $(\mu g N)^{-1}$. 207

208 Parameters related to zooplankton feeding:

209 The parameters related to reduction in feeding on toxic cell (β) was calibrated from

210 Teegarden & Cembella [12] who quantified the feeding rate on a toxic strain of *A*.

211 *minutum* by *A. tonsa* as a function of cellular toxin content (μ g N cell⁻¹). We fit Eq. 9

to the experimental data to estimate β (Fig. 2d). We use $8.95 \times 10^{-4} \mu g C$ as the

cellular carbon content of *A. minutum* [28], and chose the value of the mortality

214 constant $(m_{p,0})$ as 0.008 L $(\mu g C)^{-1} d^{-1}$ based on the clearance rate of A. tonsa [34].

215

216 **Results**

217 *Optimal allocation strategy*

218 The optimal allocation of nitrogen to toxin production is the one that yields the 219 highest population growth rate (Fig. 3). The optimal allocation of N to toxin 220 production increases with decreasing environmental nitrogen availability and 221 increasing concentration of zooplankton (Fig. 3). As a result, the investment in 222 defense - toxin production - in toxic dinoflagellates varies with both N-availability 223 and grazing pressure, with implications to cell division rate, grazing mortality rate, 224 and population growth rate (Fig. 4, Fig. 5, Fig. 6 and Fig. 7). 225 Toxin production, grazing mortality, and cost of defense 226 At high N concentrations, the cells produce toxin whenever zooplankton is present but 227 production ceases in the absence of grazers (Fig. 4a). Note that for simplicity we 228 assume that toxin production rate becomes zero when there is no grazer. In contrast, 229 at low N, phytoplankton produce toxins only when zooplankton biomass exceeds a 230 threshold concentration, and the cellular toxin content increases with the biomass of

the zooplankton (Fig. 4a, 6b).

232 Defended cells experience lower grazing mortality than undefended cells especially

when nitrogen availability is high and zooplankton concentration is low (Fig. 4d) as

cellular toxin content remains high (Fig. 4a). Taken together the grazing mortality

increases with zooplankton density and decreases with availability of nitrogen.

The cost of the defense can be quantified as a reduction in the cell division rate of

237 defended relative to undefended cells. This cost is significant only at high

238 zooplankton biomass and/or low nitrogen availability (Fig. 4b, Fig. 5a) and increases

with increasing zooplankton biomass and decreasing nitrogen concentration (Fig. 6c,

240 d). At realistically high zooplankton biomass and realistically low nitrogen 241 availability, cell division rate may be reduced by more than 20%. At high nitrogen 242 concentrations, the cells produce toxins from the excess nitrogen (not used for 243 growth) and therefore the costs are, of course, unmeasurably low (Fig. 6d). The net outcome of the defense investment is that defended cells have similar or higher 244 245 population growth rates (fitness) than undefended cells under all nutrient conditions 246 (Fig. 4c). The absolute enhancement is largest at high N (Fig. 6e) whereas the relative 247 advantage is most pronounced at low N and high zooplankton biomass (Fig. 6f).

248 Effects of P and light limitation

249 If phosphate or light rather than nitrogen limit cell growth, N is in excess and the 250 excess N can be allocated to toxin production and the cells consequently become well 251 defended. Figs. 7a and b display the effects of light limitation on toxin production and 252 cellular toxin contents, respectively. There are two regions in this parameter space: 253 the left region where light is limiting and toxin production increases with light 254 intensity while toxin content decreases with light intensity; and the right region, 255 where nitrogen is limiting and both toxin production and cellular toxin content are 256 independent of light intensity. Toxin production and cellular contents similarly vary 257 in the nitrogen-phosphorous parameter space, between phosphorous limitation at low 258 P and nitrogen limitation at high P (Fig. 7c, d). Under all conditions, cellular toxin 259 contents increase with decreasing ambient P.

260 Sensitivity analysis of β and r_T

261 Since the parameter β was calibrated based on only four available data points (Figure 262 2d), we perform a sensitivity analysis by varying β , which represents the reduction in

263 zooplankton grazing due to cellular toxin (see supplementary material fig. A1). 264 Overall, the qualitative patterns described above are robust to changes in β . When 265 nitrogen concentrations are high, there is no observable change in cellular toxin 266 concentration with varying β as organisms produce toxins at their maximum rate and 267 consequently division rates remain same. However, due to the increase in predation 268 pressure with decreasing β , population growth rate decreases. On the other hand, with 269 decrease in nitrogen concentrations, organisms produce more toxin with decrease in 270 β , leading to reduction in division rates as well as growth rates.

271 Similarly, varying the parameter r_T , representing the metabolic cost of synthesizing 272 toxin, does not lead to observable changes in the system dynamics (see supplementary 273 material fig. A2).

274 Discussion

Experiments have demonstrated that PST producing dinoflagellates become less toxic
when N-starved, that toxin content is high in exponentially growing cells in N:P
balanced environments, and that the cells become most toxic when P-limited [20, 26,
35–37]. Light-limited cells also accumulate more toxins [38] and toxin production is
enhanced in the presence of grazer cues [13]. Our model qualitatively reproduces all
these observations.

The model further conforms with the experimental observation that the cost of toxin production, quantified as a reduction in cell division rate of toxin-producing cells compared to cells or strains that produce less or no toxins, is negligible when resources, mainly N and light, are plentiful [3, 13]. However, when light or nitrogen are limiting cell growth, and when grazers are abundant, we predict that the cost of

286 investing in toxin production may be substantial and lead to > 20% reduction in cell 287 division rate. There is evidence for other defense mechanisms in phytoplankton where 288 the cost of the defense only materializes when resources are limiting [39–41]. 289 However, the prediction of costs of toxin production still remains to be tested experimentally. The costs of the investment are two-fold: (i) the cells need nitrogen to 290 291 build toxin molecules, and this requirement compete with the nitrogen investment in 292 cell structure and growth. While the nitrogen requirement for toxin production is 293 small in absolute terms, leading John and Flynn [20] to consider it trivial, it becomes 294 significant when nitrogen is limiting, and may eventually lead to a total shut down of 295 toxin production. (ii) The cells further need energy for the synthesis of toxins, and this 296 energy eventually comes from photosynthesis. This is why toxin production rate 297 increases with light intensity when light is the limiting resource. In this situation, cell 298 division rate increases faster than toxin production rate with increasing light, and 299 therefore cellular toxin content decreases with increasing light, an effect and a 300 mechanism in agreement with experimental observations [38].

301 The environmental conditions that promote cell toxicity, i.e., high grazer biomass, 302 actually coincide with the time of the year when nitrogen is the most limiting resource 303 in temperate shelf regions. Thus, zooplankton (copepod) biomass is at its seasonal maximum during summer and may easily exceed 10-100 μ g C L⁻¹ [42], which will 304 305 impose a high predation mortality and thus induce high cell toxicity. At this time of 306 the year, concentrations of inorganic nitrogen in surface waters in temperate shelf regions are low, typically below 1-10 μ g N L⁻¹ [43, 44], and under these conditions 307 the model predicts that the cost of toxin production is substantial, leading to > 20%308 309 decrease in cell division rate. However, the investment in defense pays off since defended cells experience lower grazing mortality, and may consequently have net 310

growth rates up to twice or more of that of undefended cells (e.g. Fig. 6f). If the toxins
have deterrent rather than toxic effects, i.e., preventing the toxic cells from being
consumed rather than killing grazers that do consume the cells [10, 11, 45], then the
toxic cells may have a competitive advantage and a monospecific bloom may
develop. Indeed, blooms of toxic algae typically occur during summer [46], which is
consistent with these considerations.

Furthermore, the toxic species can also gain growth advantage under high nitrogen and high grazer biomass, as growth rates can be doubled due to the benefit provided by toxin production at a negligible production cost (Fig. 6d, f). Such situations are comparable with many coastal areas where N:P ratios remain considerably high due to erratic input of nitrogen from human activities, and can consequently result in toxic blooms [47]. Thus, potentially toxic species can also become toxic when exposed to high N regime caused by eutrophication.

324 The success of toxic bloom formation also depends on the evolutionary history of 325 zooplankton grazer and toxic species [40]. Evidence from both fresh- and marine 326 waters shows that grazers can evolve full or partial resistance against the toxic algae 327 [48, 49]. Our results suggest that cellular toxin content will increase in the presence of 328 toxin-resistant grazers as will the costs (see supplementary material fig. A1). 329 However, if the benefits in terms of reduced grazing mortality vanish due to grazer 330 resistance, the possibility of success of the toxic species in terms of forming bloom will be reduced or disappear. 331

332 Many phytoplankton species have evolved what is supposed to be defense

333 mechanisms to avoid or reduce predation, ranging from hard shells and spines, to

evasive behaviors and toxin production, and such defenses may have significant

335 implications on predator-prey interactions, population dynamics, and diversity of 336 phytoplankton communities [50]. However, the trade-offs are rarely quantified and 337 often not even documented. While there is increasing evidence that toxin-production 338 in many cases provides partial protection from grazing in dinoflagellates, the costs 339 have hitherto not been properly quantified. The currencies of benefits and costs are 340 growth and mortality. Here, we have shown that the costs are not only strongly 341 dependent on the concentration of grazers but also on the resource availability, and 342 that the realized costs in nature are typically highest during summer when the defense 343 is most needed. The delicate balance between costs, benefits, and resource availability 344 not only explains why the defense is inducible but also has implications for the timing 345 of toxic algal blooms. Further studies on costs and benefits of toxin production are 346 needed to experimentally test our model predictions (e.g., toxin production and growth under sufficient and deficient resources), and for deeper understanding of the 347 348 mechanisms and evolution of inducible toxin production. At present, very little is 349 known about the consequences of inducible toxin production on the community level 350 in complex communities. Future work should be devoted towards investigating the 351 complex integrated ecological issues of inducible toxin production, species diversity, 352 and food web structure.

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357 Conflict of interest

358 The authors declare that they have no conflict of interest.

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- **Table 1.** Central symbols and general parameters. Index *i* refers to carbon (C) via
- 531 light-dependent photosynthesis where light intensity is measured in units of
- 532 $\mu E m^{-2}s^{-1}$, nitrogen (*N*) in units of $\mu g N L^{-1}$ or phosphorous (P) in units of $\mu g P L^{-1}$.
- 533 Calibration of parameters is based on data from laboratory measurements and
- 534 provided in the 'Model parameterization' section.

Symbol	Description	Value	Unit	Reference

L	Light flux in the environment	-	$\mu E m^{-2} s^{-1}$	-
Ν	Concentration of nitrogen in			
	the environment	-	μ g N L^{-1}	-
Р	Concentration of phosphorous			
	in the environment	-	μ g P L^{-1}	-
W_X	Cellular mass of toxic algae	8.95×10^{-4}	μ g C	[28]
Z	Biomass of zooplankton	-	μ g C L^{-1}	-
	Functional responses			
J _i	Flux of assimilated substance		μ g C d ⁻¹ , μ g N d ⁻¹ , or μ g P d ⁻¹	Eq. (1)
A_L	Affinity for light	3.1×10^{-5}	μ g C (μ E m ⁻² s ⁻¹) ⁻¹ d ⁻¹	Calibrated
A_N	Affinity for nitrogen	3×10^{-6}	$L d^{-1}$	Calibrated
A_P	Affinity for phosphorous	3×10^{-7}	$L d^{-1}$	Calibrated
M_L	Max. uptake rate of C through			
	photosyntehsis	9.5×10^{-4}	μ g C d ⁻¹	Calibrated
M_N	Max. uptake rate of N	1.1×10^{-4}	μ g N d ⁻¹	Calibrated
M_P	Max. uptake rate of P	1.3×10^{-5}	μ g P d ⁻¹	Calibrated
	Costs and toxin production			
R_{C}	Total metabolic cost	-	μ g C d ⁻¹	Eq. (2)
R_0	Basal respiration rate	1.07×10^{-4}	μ g C d ⁻¹	[29]
θ	Fraction of N devoted to toxin	-	-	Eq. (10)
T_{pot}	Potential toxin production rate	-	μ g N d ⁻¹	Eq. (3)
T_r	Actual toxin production rate	-	μ g N d ⁻¹	Eq. (4)
Т	Cellular toxin content	-	μ g N cell ⁻¹	-
R_T	Cost of toxin production	-	μ g C d ⁻¹	Eq. (5)
n_T	Material cost of toxin			
	production	1.23	$\mu g C (\mu g N)^{-1}$	Calibrated
r_T	Metabolic cost of synthesizing			
	toxin	2.7	$\mu g C (\mu g N)^{-1}$	Calibrated
	Predation			
m_p	Predation mortality	-	d ⁻¹	Eq. (9)

			0.000	• (• • • • • • • • • • • • • • • • • •	[2.4]
	$m_{p,0}$	Mortality constant	0.008	$L (\mu g C)^{-1} d^{-1}$	[34]
	β	Reduction in grazing due to			
		toxin	1.147×10^{5}	cells (μ g N) ⁻¹	Calibrated
		Synthesis and growth			
	J _{tot}	Total available C flux	-	μ g C d ⁻¹	Eq. (6)
	μ	Division rate of algae	-	d ⁻¹	Eq. (7)
	r	Growth rate of algae	-	d ⁻¹	Eq. (8)
	Q_{CN}	C:N mass ratio	5.68	$\mu g C (\mu g N)^{-1}$	[51]
	Q_{CP}	C:P mass ratio	41	$\mu g C (\mu g P)^{-1}$	[51]
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Figure 1. Schematic representation of the model showing how fluxes of nitrogen 544 545 (dotted), carbon (solid) and phosphorous (dash-dot) are lost through respiration (gray 546 explosion) and toxin production (gray rectangles), and combined (gray ellipse) to 547 determine growth rate. White triangle symbols represent the functional responses for the uptake mechanisms. $R_{\rm C}$ represents the respiratory cost that includes the costs of 548 549 both uptake and mobilization of resources for synthesis and the maintenance of 550 structure. The rate at which toxin is synthesized from C and N is $n_T T_r$, and the 551 respiratory cost of toxin production is $r_T T_r$ where T_r is the toxin production rate. The 552 ellipse represents synthesis of biomass from the available C, N and P following Liebig's law of the minimum and constrained by the Redfield ratio ($\mu g C (\mu g N)^{-1}$ 553 ¹=5.68, μ g C (μ g P)⁻¹=41) [51]. In our steady state consideration, the excess amounts 554 of assimilated C, N or P are assumed lost as excess resources. µ represents the 555

division rate, and r is the population growth rate after subtracting predation mortality

557 from μ.





Figure 2. Comparison of division rates (μ) between available data and model outcome with the calibrated parameters by varying (a) nitrogen ($P = 200 \ \mu g P L^{-1}$) (at three different light intensities $L = 100 \ \mu E m^{-2} s^{-1}$, 50 $\mu E m^{-2} s^{-1}$, 25 $\mu E m^{-2} s^{-1}$) [52], (b) phosphorous (L =45 $\mu E m^{-2} s^{-1}$, $N = 200 \ \mu g N L^{-1}$) [53], and (c) light ($N = 6000 \ \mu g N L^{-1}$, P =

- 564 $400 \ \mu g P L^{-1}$ [52, 54]. Grazing rate at different toxin concentrations (d). Data for
- 565 grazing on toxic *A. tamarense* strain CCMP 115 by *A. tonsa* were used [12].



Figure 3. Population growth rate as a function of the fraction of assimilated nitrogen that is allocated to toxin production (a) at low and high environmental concentration of N (N=80 μ g N L^{-1} and 10 μ g N L^{-1} with concentration of grazer $Z = 10 \ \mu$ g C L^{-1}), and (b) at high and low concentration of zooplankton ($Z = 20 \ \mu$ g C L^{-1} and 1 μ g C L^{-1} with $N = 75 \ \mu$ g N L^{-1}). The maximum of the curves shows the optimal

572 allocation strategy (θ^*) (marked by arrows). Thin lines represent growth rates in the

by absence of toxin production. Other resources are phosphate (P) = 120 μ g P L^{-1} , light

574 intensity (L) = 150 μ E m⁻²s⁻¹, and phytoplankton biomass (X) = 90 μ g C L⁻¹.



Figure 4. Optimal cellular toxin content (a), cell division rate (b), population growth rate (c), and predation mortality (d) as a function zooplankton biomass at low (N=20 μ g N L^{-1}) and high (N=150 μ g N L^{-1}) N concentrations of defended (toxin producing) and undefended cells (non-toxic strain). Curves of division rates for defended and undefended cells under high N concentration lie on top of each other. Light intensity $L = 150 \ \mu$ E m⁻²s⁻¹ and phosphorous concentration $P = 120 \ \mu$ g P L⁻¹ in all plots.





Figure 5. Relative reduction in cell division rate as a function of nitrogen

584 concentration (L=150 μ E m⁻²s⁻¹, P=120 μ g P L⁻¹) (a), phosphorous concentration

585 (L=150 μ E m⁻²s⁻¹, N=40 μ g N L⁻¹) (b), and light intensities (N=120 μ g N L⁻¹,

586 P=120 μ g P L⁻¹) (c) at high (Z=20 μ g C L⁻¹) and low (Z=1 μ g C L⁻¹) zooplankton

- 587 biomasses. Cellular toxin content as a function of N at high and low zooplankton
- 588 biomasses (d).



Figure 6. Surface plots of toxin production rate (a), and cellular toxin content (b), as well as absolute and relative changes in cell division rate $(\mu(\theta^*) - \mu(\theta = 0))$ and $(\mu(\theta^*) - \mu(\theta = 0)) \times 100/\mu(\theta = 0))$ (c, d), and population growth rates $(g(\theta^*) - g(\theta = 0)) = 0$ and $(g(\theta^*) - g(\theta = 0)) \times 100/g(\theta = 0))$ (e, f) of defended relative to undefended cells as a function of N-availability and zooplankton biomass. Light intensity $L = 150 \ \mu \text{E} \text{ m}^{-2} \text{s}^{-1}$ and phosphorous concentration $P = 120 \ \mu \text{g} \text{ P} \text{ L}^{-1}$ in all plots.



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Figure 7. Surface plots of toxin production rate (a), and cellular toxin content (b) as a function of N-availability and light intensity. Zooplankton biomasses $Z=10 \ \mu g \ C \ L^{-1}$, and phosphorous concentration P=120 $\mu g \ P \ L^{-1}$ in both plots. Surface plots of toxin production rate (c), and cellular toxin content (d) as a function of N and P-availability. Zooplankton biomasses Z=10 $\mu g \ C \ L^{-1}$, and light intensity L=150 $\mu E \ m^{-2} \ s^{-1}$ in both plots.

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611 Supplementary material

612 Appendix A. Sensitivity analysis

Fig. A1 shows the color plot with continuous variations in both N concentration and

- 614 the benefit from toxin production (β). The toxin production rate is high when the
- benefit from toxin production remains within certain range. However, it decreases at

616 very high benefit range as organisms receive large benefit by producing small amount

- 617 of toxin (Fig. A1b). As a result, benefits of toxin production in terms of population
- 618 growth rate remains high when benefits are high and also N concentration is high.
- Fig. A2 shows the color plot with continuous variations in both N concentration and

620 the metabolic cost of synthesizing toxin (r_T) . When the cost is relatively low, an

621 increase in N concentration increases toxin production (Fig A2b), as organisms get

622 more benefit from toxin production rather than increasing division rate. However,

623 when the toxin production is costly and N is sufficient in the system, organisms invest

624 their energy in increasing division rate rather than production of costly toxin.

625 Although, since low N does not support high growth, organisms produce toxin to

626 increase their population growth rate in spite of their high cost.

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635 Figure legends

- **Figure A1.** Surface plots of optimal allocation of N to toxin production (a), toxin
- 637 production rate (b), and cellular toxin content (c), as well as absolute and relative
- 638 changes in cell division rate (($\mu(\theta^*) \mu(\theta = 0)$) and ($\mu(\theta^*) \mu(\theta = 0)$)×100/ $\mu(\theta = 0)$)
- 639 0))) (d, e), and population growth rates (($g(\theta^*) g(\theta = 0)$) and ($g(\theta^*) g(\theta = 0)$)))
- 640 0))×100/g(θ = 0))) (f, g) of defended relative to undefended cells as a function of N-
- 641 availability and the strength of toxicity (β). The position of arrow indicates the value
- 642 of β used for other figures. The phytoplankton biomasses X=90 μ g C L^{-1} ,
- 643 zooplankton biomasses Z=10 μ g C L^{-1} , light intensity L=150 μ E m⁻²s⁻¹, and
- 644 phosphorous concentration P=120 μ g P L⁻¹ in all plots.
- **Figure A2.** Surface plots of optimal allocation of N to toxin production (a), toxin
- 646 production rate (b), and cellular toxin content (c) as well as absolute and relative
- 647 changes in cell division rate (($\mu(\theta^*) \mu(\theta = 0)$) and ($\mu(\theta^*) \mu(\theta = 0)$)×100/ $\mu(\theta = 0)$)
- 648 ())) (d, e) and population growth rates (($g(\theta^*) g(\theta = 0)$) and ($g(\theta^*) g(\theta = 0)$)
- 649 0))×100/g($\theta = 0$))) (f, g) of defended relative to undefended cells as a function of N-
- availability and the cost of toxin production (r_T) . The position of arrow indicates the
- value of r_T used for other figures. The phytoplankton biomasses X=90 μ g C L^{-1} ,
- 652 zooplankton biomasses Z=10 μ g C L^{-1} , light intensity L=150 μ E m⁻²s⁻¹, and
- 653 phosphorous concentration $P=120 \mu g P L^{-1}$ in all plots.





655 Figure A1



