

Effect of Previous Exposure to Malaria on Blood Pressure in Kilifi, Kenya: A Mendelian Randomization Study

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Background—Malaria exposure in childhood may contribute to high blood pressure (BP) in adults. We used sickle cell trait (SCT) and α^+ thalassemia, genetic variants conferring partial protection against malaria, as tools to test this hypothesis.

Methods and Results—Study sites were Kilifi, Kenya, which has malaria transmission, and Nairobi, Kenya, and Jackson, Mississippi, where there is no malaria transmission. The primary outcome was 24-hour systolic BP. Prevalent hypertension, diagnosed using European Society of Hypertension thresholds was a secondary outcome. We performed regression analyses adjusting for age, sex, and estimated glomerular filtration rate. We studied 1 127 participants in Kilifi, 516 in Nairobi, and 651 in Jackson. SCT frequency was 21% in Kilifi, 16% in Nairobi, and 9% in Jackson. SCT was associated with -2.4 (95% CI, -4.7 to -0.2) mm Hg lower 24-hour systolic BP in Kilifi but had no effect in Nairobi/Jackson. The effect of SCT in Kilifi was limited to 30- to 59-year-old participants, among whom it was associated with -6.1 mm Hg (CI, -10.5 to -1.8) lower 24-hour systolic BP. In pooled analysis allowing interaction by site, the effect of SCT on 24-hour systolic BP in Kilifi was -3.5 mm Hg (CI, -6.9 to -0.1), increasing to -5.2 mm Hg (CI, -9.5 to -0.9) when replacing estimated glomerular filtration rate with urine albumin to creatinine ratio as a covariate. In Kilifi, the prevalence ratio for hypertension was 0.86 (CI, 0.76–0.98) for SCT and 0.89 (CI, 0.80–0.99) for α^+ thalassemia.

Conclusions—Lifelong malaria protection is associated with lower BP in Kilifi. Confirmation of this finding at other sites and elucidating the mechanisms involved may yield new preventive and therapeutic targets. (*J Am Heart Assoc.* 2019;8:e011771. DOI: 10.1161/JAHA.118.011771.)

Key Words: ambulatory blood pressure monitoring • high blood pressure • hypertension • malaria • Mendelian randomization • sickle cell disease • sickle cell trait • thalassemia

High blood pressure (BP) is a major cause of morbidity and mortality worldwide, and its impact is particularly substantial in sub-Saharan Africa.¹ Examining factors unique

to, or more prevalent in, sub-Saharan Africa might reveal pathophysiological mechanisms underlying hypertension that could be exploited to reduce the burden of hypertension. More than half of Africa's population lives in areas with moderate to high malaria transmission.² *Falciparum* malaria is associated with low birth weight, childhood malnutrition, and chronic inflammation, each of which has been associated with the development of hypertension.³ In addition, children of women who experienced malaria in pregnancy have higher BP at 1 year of age than those whose mothers did not have malaria during pregnancy.⁴

Mendelian randomization studies, in which genetic polymorphisms are used to represent environmental exposures, are increasingly being applied in determining causality.⁵ These natural experiments, based on genes acquired at conception, overcome the limitations of observational studies while avoiding the expense and ethical concerns that prevent the conduct of randomized clinical trials.

Sickle cell trait (SCT) and α^+ thalassemia are common genetic polymorphisms among African populations that

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Accompanying Data S1, Tables S1 through S15, and Figures S1 through S4 are available at <https://www.ahajournals.org/doi/suppl/10.1161/JAHA.118.011771>

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Clinical Perspective

What Is New?

- We tested the hypothesis that previous exposure to malaria is associated with increased blood pressure (BP).
- We compared BP in individuals with and without sickle cell trait, which protects against malaria, at 3 sites, 1 where there is ongoing malaria transmission and 2 where there is no malaria transmission.
- We found that individuals with sickle cell trait had lower BP than those without sickle cell trait, but this was only in the area with malaria transmission, indicating that malaria is associated with high BP.

What Are the Clinical Implications?

- Confirmation of the existence of a link between previous exposure to malaria and high BP at other sites and elucidation of the mechanisms involved may help in finding new ways to prevent and treat high BP.

provide protection against malaria. Examining differences in BP among adults with and without these polymorphisms can allow inferences to be made as to whether malaria influences BP.³ In the present study, we tested whether SCT and α^+ thalassemia are associated with lower BP and a lower prevalence of hypertension in Kilifi, Kenya, where there is malaria transmission. For comparison, we investigated the same association in Nairobi, Kenya, and Jackson, Mississippi, 2 areas where there is no malaria transmission.

Methods

Data and Materials Availability

The authors confirm that, for approved reasons, some access restrictions apply to the data underlying the findings. The authors are unable to make the data more freely available because of the terms for data sharing included in the consent forms for this study. Data are available through the Data Governance Committee of the KEMRI Wellcome Trust Research programme where uses are compatible with the consent obtained from participants for data collection in this study. Requests can be sent to the coordinator of the Data Governance Committee on dgc@kemri-wellcome.org and the Jackson Heart Study on jhspub@umc.edu.

This Mendelian randomization study was performed among residents of 3 sites (Figure S1) with different levels of malaria transmission; Kilifi, Kenya, where there has been moderate malaria transmission,^{6–8} and 2 sites (Nairobi, Kenya, and Jackson, Mississippi) where there is no malaria transmission.^{9–11} We utilized SCT, which offers 50% to 90% protection

against malaria episodes,^{12,13} predominantly in childhood,¹⁴ to represent malaria exposure (Figure S2). In a secondary analysis among Kenyan participants, we used α^+ thalassemia, which provides a lower level of protection against malaria than SCT. To be eligible for the study in Kenya, individuals had to be 10 years or older at investigation and to be lifelong residents of Kilifi or Nairobi. In Kilifi, we recruited randomly selected residents of Chasimba and Junju locations within the Kilifi Health and Demographic Surveillance System.¹⁵ Kilifi participants were predominantly of the Chonyi subtribe of the Mijikenda ethnic group. In Nairobi, we recruited randomly selected residents of the Nairobi Urban Health and Demographic Surveillance System.¹⁶ Study participants in Nairobi were randomly selected from among those who had self-identified as belonging to ethnic groups known to have a high frequency of malaria-protective polymorphisms (Luhya, Luo, Teso, Mijikenda).^{17,18} Data were collected in Kenya between December 2015 and June 2017. Participants in the United States were blacks aged 21 years and older recruited into the Jackson Heart Study between 2000 and 2004.¹⁹

We used appropriately sized cuffs on the nondominant arm to undertake ambulatory BP monitoring (ABPM). We used the Arteriograph24 (TensioMed Ltd.) device in Kenya and the Spacelabs 90207 (Spacelabs) device in the United States. For the primary analyses, we defined completeness of ABPM recordings using European Society of Hypertension (ESH) criteria.²⁰ These criteria require ≥ 20 daytime (9 AM–9 PM) and ≥ 7 nighttime (1 AM–6 AM) readings.²⁰ The respective time periods were used to determine mean daytime and nighttime BP. The mean 24-hour BP was calculated using all available readings. Prevalent hypertension was diagnosed among participants 16 years and older who met any of the ESH-defined thresholds for ABPM hypertension regardless of whether they were taking antihypertensive medication. The thresholds used were 24-hour systolic BP (SBP) ≥ 130 mm Hg or 24-hour diastolic BP ≥ 80 mm Hg, daytime SBP ≥ 135 mm Hg or daytime diastolic BP ≥ 85 mm Hg, and/or nighttime SBP ≥ 120 mm Hg or nighttime diastolic BP ≥ 70 mm Hg BP.^{21,22} Detailed study procedures are described in the supplement.

Statistical Analysis

The primary outcome was the difference in 24-hour SBP by SCT status. We determined that a sample of 1115 participants in Kilifi and 1270 participants in Nairobi and Jackson combined would provide at least 80% statistical power to detect a 4-mm Hg difference in 24-hour SBP between participants with and without SCT at each of these sites, assuming an SCT frequency of 15% in Kilifi and 10% in Nairobi/Jackson and an SD for 24-hour SBP of 15 mm Hg. We defined secondary outcomes as the genotype-specific

differences in systolic and diastolic components of daytime and nighttime BP, and hypertension as defined above. A 2-sided α level of ≤ 0.05 was taken to indicate statistical significance.

We used χ^2 test and Student *t* test to compare categorical and continuous variables at each site by genotype. Hardy–Weinberg equilibrium was evaluated using a χ^2 test. Nonnormally distributed variables were log-transformed before analysis.

Two types of analyses were conducted to test the hypothesis. First, we compared BP among participants with and without SCT at each of the 3 sites, while adjusting for confounders as described below. Second, we pooled data from the 3 sites and analyzed whether there was an interaction in the effect of SCT on BP by site.

In the first analyses, which were site-specific, we performed linear regression to determine whether SCT status was associated with 24-hour SBP, adjusting for age, sex, and estimated glomerular filtration rate (eGFR)²³ (Figure S3), which were specified a priori as potential confounders. These covariates were also used in Poisson regression models with robust variance to assess whether SCT was associated with prevalent hypertension. As α^+ thalassemia modifies the protective effect of SCT against malaria,²⁴ we tested for statistical interaction in their effect under both dominant and additive conditions among Kilifi participants.

The second, pooled analyses were conducted as follows. Initially, we tested for heterogeneity in the effect of SCT on BP in the 2 sites with no malaria transmission, Nairobi and Jackson, by conducting a linear regression allowing interaction by site. As there was no evidence of heterogeneity in this analysis, we combined data from these 2 sites and then tested whether the effect of SCT on BP differed across sites with different levels of malaria transmission (Kilifi versus Nairobi/Jackson). This was conducted in a regression model that tested for the main effects of age, sex, eGFR, malaria site (Kilifi versus Nairobi/Jackson) and SCT, and an interaction term for SCT and malaria site.

Prespecified subgroup analyses included stratification by age category (10–29, 30–59, and ≥ 60 years) and sex, and after exclusion of participants taking antihypertensive medication. These subgroup analyses were performed for Kilifi separately and for Nairobi and Jackson combined. We also examined the effect of replacing eGFR with log-transformed urine albumin to creatinine ratio as the covariate representing renal function in the pooled regression models.

In sensitivity analyses, we repeated the analyses using an expanded data set that included participants whose ABPM data met the less stringent IDACO (International Database of Ambulatory Blood Pressure in relation to Cardiovascular Outcome) study criteria for completeness. These criteria

require a minimum of 10 daytime (10 AM–8 PM) readings and 5 nighttime (12 AM–6 AM hours) readings.

All analyses were conducted using Stata version 15 software (StataCorp).

The ethical review committees/institutional review boards of the Kenya Medical Research Institute, London School of Hygiene and Tropical Medicine, Jackson State University, Tougaloo College, and the University of Mississippi Medical Center approved the study. Analysis of Jackson Heart Study data was approved by the University of Alabama at Birmingham's institutional review board. Written informed consent, and assent for participants younger than 18 years in Kenya, was obtained from study participants or parents.

Results

We identified 9543 lifelong residents in the Kilifi population register and, using a random number sequence, we invited 2790 to participate in the study (Figure 1). Characteristics of participants by consent to undergo ABPM and by completeness of ABPM recordings are summarized in Tables S1 and S2, respectively. Individuals with homozygous sickle cell disease ($n=5$) were excluded from all analyses. Complete data were available for 1127 participants. None of the participants were previously aware of their genetic status. Overall, 238 (21%) participants had SCT and 768 (67%) were either heterozygous ($-\alpha/\alpha\alpha$) or homozygous ($-\alpha/-\alpha$) for α^+ thalassemia, equally distributed by SCT status (69% for those with SCT versus 67% for those without SCT, $P=0.5$). There was no departure from Hardy–Weinberg equilibrium for either SCT ($P=0.5$) or α^+ thalassemia ($P=0.8$). Mean 24-hour SBP was 127 ± 18 mm Hg.

Characteristics of participants by study site and SCT status are presented in Table 1. A higher proportion of participants without SCT versus participants with SCT in Jackson were taking antihypertensive medication. In Kilifi, eGFR was lower among participants with SCT versus participants without SCT. Urine albumin to creatinine ratio was higher among participants with SCT versus participants without SCT in Kilifi and Jackson.

Mean 24-hour SBP in Kilifi was 126 ± 18 mm Hg in individuals with SCT and 127 ± 18 mm Hg in individuals without SCT. In Nairobi/Jackson, mean 24-hour SBP was 123 ± 13 mm Hg in individuals with SCT and 123 ± 14 mm Hg in individuals without SCT. After adjusting for age, sex, and eGFR in regression analyses, SCT was associated with a -2.4 (95% CI, -4.7 to -0.2) mm Hg ($P=0.037$) lower 24-hour SBP in Kilifi (Figure 2). In Nairobi and Jackson, there was no association between SCT and 24-hour SBP. As there was no heterogeneity between Nairobi and Jackson in the effect of SCT on any BP measure (test for interaction, $P=0.409$ – 0.944 ; Table S3) data from these sites

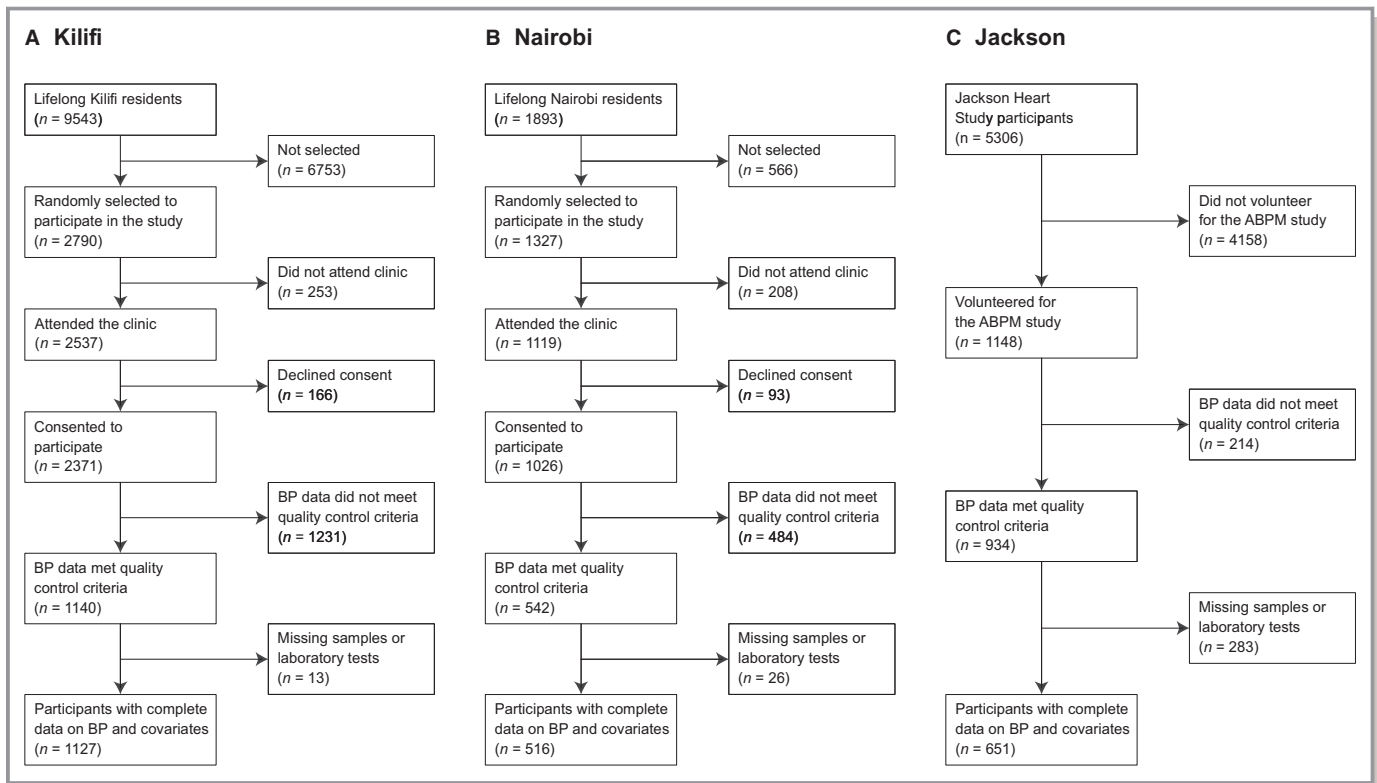


Figure 1. Study flow chart. ABPM indicates ambulatory blood pressure monitoring; BP, blood pressure.

were pooled in subsequent analyses. The proportion of variation in BP explained by the regression model as determined by the adjusted r^2 statistic was 25% in Kilifi, 16% in Nairobi, and 7% in Jackson.

The association between SCT and 24-hour SBP observed in Kilifi was unaffected by excluding participants taking antihypertensive medication (Table S4).

In Kilifi, SCT was associated with lower 24-hour SBP (-6.1 mm Hg; 95% CI, -10.5 to -1.8 [$P=0.006$]) among participants aged 30 to 59 years but not in participants in the other age categories (Table 2). In Nairobi and Jackson combined, there was no association between SCT and any BP measure in any age group except for 24-hour and daytime diastolic BP in those 60 years and older, where BP was higher among participants with SCT. In Kilifi, there was a numerically stronger association between SCT and lower 24-hour SBP in women compared with men but there was no statistically significant interaction by sex ($P=0.218$, Table S5).

In the pooled analyses, SCT was associated with -3.5 mm Hg (CI, -6.9 to -0.1) lower 24-hour SBP ($P=0.041$) in Kilifi when compared with Nairobi and Jackson (Figure 3). This interaction model was associated with an adjusted r^2 statistic of 20%. The magnitude of the difference was larger (-5.2 mm Hg; CI, -9.5 to -0.9 [$P=0.019$]) when we adjusted for log-transformed urine albumin to creatinine

ratio instead of eGFR (Table S6). Stratified by sex, the effect of SCT on 24-hour SBP in Kilifi compared with Nairobi/Jackson was not stronger in women compared with men ($P=0.419$, Table S7).

The prevalence of hypertension among study participants was 56% in Kilifi, 28% in Nairobi, and 62% in Jackson. In Kilifi, the adjusted prevalence ratio (PR) for hypertension among participants with SCT versus those without SCT was 0.86 (CI, 0.76–0.98, $P=0.027$). In Nairobi and Jackson, the corresponding adjusted PR was 0.96 (CI, 0.80–1.16; $P=0.706$). When using clinic BP to define hypertension (SBP ≥ 140 and/or diastolic BP ≥ 90 mm Hg), the adjusted PR for hypertension among participants with SCT versus those without SCT in Kilifi was 0.82 (CI, 0.64–1.05; $P=0.113$). In Nairobi and Jackson, the corresponding adjusted PR when using clinic BP was 1.45 (CI, 0.62–3.42; $P=0.396$).

α^+ Thalassemia deletions were not associated with the primary outcome in either Kilifi and Nairobi (Table S8), but a significant association with prevalent hypertension was present in Kilifi. In Kilifi, the presence of ≥ 1 thalassemia deletion was associated with an adjusted PR for hypertension of 0.89 (CI, 0.80–0.99; $P=0.036$). There was no effect of α^+ thalassemia on hypertension in Nairobi (PR, 1.43; CI, 0.95–2.15 [$P=0.085$]).

There was no interaction between α^+ thalassemia and SCT on 24-hour SBP in Kilifi (test for interaction, $P=0.865$).

Table 1. Characteristics of Study Participants With and Without SCT by Study Site

Characteristic	Kilifi (N=1127)		Nairobi (N=516)		Jackson (N=651)	
	SCT (n=238)	Non-SCT (n=889)	SCT (n=82)	Non-SCT (n=434)	SCT (n=58)	Non-SCT (n=593)
	No. (%)	No. (%)	No. (%)	No. (%)	No. (%)	No. (%)
Women	126 (53)	528 (59)	40 (49)	235 (54)	38 (66)	392 (66)
Smoker	17 (7)	78 (9)	3 (4)	7 (2)	5 (9)	74 (12)
Previously diagnosed with hypertension*	37 (16)	127 (14)	7 (9)	53 (12)	31 (53)	367 (62)
Taking antihypertensive medication	9 (4)	26 (3)	0 (0)	8 (2)	26 (46)	338 (61)
	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)
Age, y	41 (22)	39 (22)	20 (14)	23 (18)	61 (12)	60 (11)
BMI, kg/m ²	20.6 (3.6)	20.6 (3.8)	20.2 (3.9)	20.5 (4.3)	31.1 (7.0)	30.9 (6.3)
HbA _{1c} , %	5.2 (0.6)	5.1 (0.8)	5.2 (1.1)	5.2 (1.0)	6.0 (1.2)	6.1 (1.3)
Hemoglobin, g/dL	12.6 (2.0)	12.6 (1.6)	13.4 (1.7)	13.2 (1.7)	12.8 (1.4)	13.0 (1.4)
WBC count ×10 ⁹ /L	5.7 (1.5)	5.7 (1.4)	5.4 (1.4)	5.4 (1.6)	4.9 (1.1)	5.3 (1.5)
Platelet count ×10 ⁹ /L	267 (97)	262 (83)	283 (99)	287 (100)	230 (58)	243 (61)
Plasma osmolality, mOsm/kg	290 (6.6)	290 (5.8)	291 (10)	291 (11)	... (...)	... (...)
eGFR, mL/min per 1.73 m ²	108 (35)	114 (42)	118 (27)	115 (24)	85 (26)	87 (26)
Log UACR, mg/g	1.3 (0.5)	1.2 (0.6)	1.3 (0.7)	1.5 (0.7)	1.06 (0.6)	0.90 (0.5)

Plasma osmolality measurements were not available for Jackson participants. BMI indicates body mass index; eGFR, estimated glomerular filtration rate; HbA_{1c}, glycosylated hemoglobin; SCT, sickle cell trait; UACR, urine albumin to creatinine ratio; WBC, white blood cell.

*Answered "yes" to the question: Has a doctor or healthcare worker previously told you that you have high blood pressure?

Results Based on IDACO ABPM Criteria

The results of sensitivity analyses are presented in Tables S9 through S15. The number of participants who satisfied IDACO criteria for completeness of ABPM data was 2048 (86%). As expected, the associations between SCT and α^+ thalassemia and BP were weaker than in the primary analyses, but the associations with hypertension remained significant. In Kilifi, SCT was associated with an adjusted PR for hypertension of 0.89 (CI, 0.80–0.99; $P=0.025$). In Nairobi and Jackson, the corresponding adjusted PR was 0.96 (CI, 0.81–1.14; $P=0.659$). The presence of any α^+ thalassemia deletion among individuals in Kilifi was associated with an adjusted PR of hypertension of 0.92 (CI, 0.84–0.99; $P=0.037$). In Nairobi, the corresponding adjusted PR for α^+ thalassemia was 1.32 (CI, 0.95–1.86 [$P=0.096$]).

Discussion

In this study, SCT, a genetic polymorphism associated with partial protection against malaria, was associated with a 2.4-mm Hg lower mean 24-hour SBP and a 14% lower prevalence of hypertension in Kilifi, an area with malaria transmission, but not in Nairobi and Jackson, areas with no malaria transmission. α -Thalassemia, which provides a lower level of protection against malaria was associated with an 11% reduction in the prevalence of hypertension in Kilifi but not in Nairobi. This

suggests that increased risk of malaria is associated with higher adult BP. In the absence of malaria, in Nairobi and Jackson, mean BP estimates were marginally higher among participants with SCT than those without SCT. Incorporating this baseline observation in a pooled analysis that compared malaria with nonmalaria sites, we estimate that malaria is responsible for a mean increase in 24-hour SBP of 3.5 mm Hg.

This difference in BP is roughly similar to those attributed to reduction of salt intake by ≈ 4 g/d²⁵ or a dose of 10 mg/d of ramipril in the HOPE (Heart Outcomes Prevention Evaluation) trial.²⁶ At the population level, a reduction in SBP of 3 mm Hg may avert a substantial number of cardiovascular events including an $\approx 15\%$ reduction in the incidence of stroke.²⁷ However, several factors suggest that the actual effect of malaria on BP might be greater. First, SCT is only associated with a 50% reduction in incidence of nonsevere malaria¹³ and 90% reduction against severe malaria episodes,²⁸ and this relative protection wanes with age.¹⁴ Second, the protection offered by SCT against malaria is reduced in individuals with concurrent α^+ thalassemia,²⁴ who comprised 67% of participants in the current study who had SCT. The current study was not powered to analyze the effect of this interaction. In addition, because Kilifi has low to moderate malaria transmission,⁶ the effect of malaria on BP in other parts of Africa where malaria is endemic could be considerably higher.

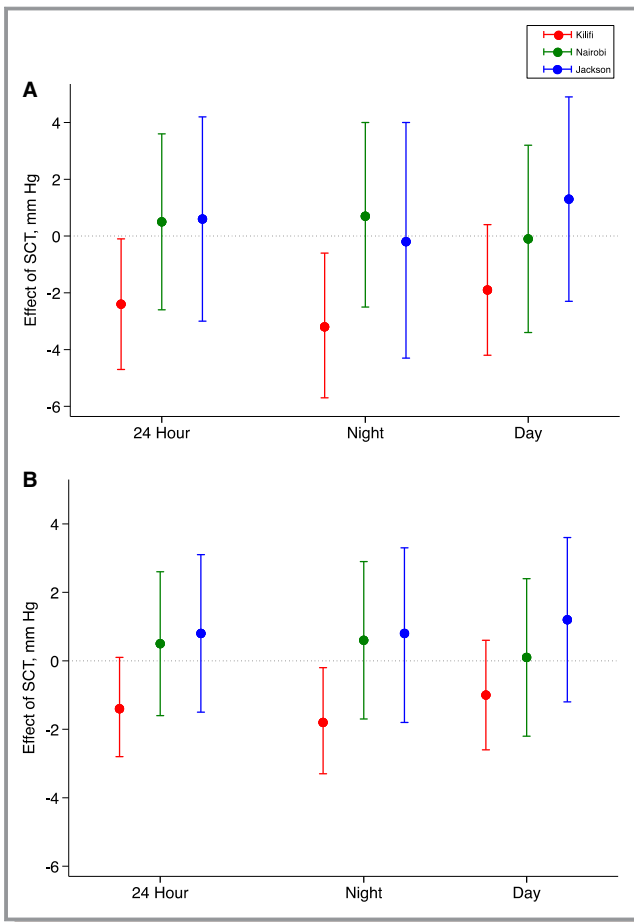


Figure 2. Effect of sickle cell trait (SCT) on blood pressure by study site. **A**, Systolic blood pressure and **B**) diastolic blood pressure. Linear regression models with adjustment for age, sex, and estimated glomerular filtration rate.

In this epidemiological study, we were not able to study the physiological mechanisms by which malaria results in higher BP. However, there are a number of plausible hypotheses. For example, hypertension may be the consequence of chronic inflammation in childhood induced by malaria³ and inflammation, itself, has been associated with the development of hypertension.²⁹ Malaria also causes stunting and malnutrition, which could influence BP,³ although anthropometric indices such as BMI were similar in the groups we studied. The numerically stronger association that we observed between SCT and BP in women could be explained by SCT-mediated protection against malaria in pregnancy.³⁰ Malaria in pregnancy has been associated with gestational hypertensive disorders that place women at risk for chronic hypertension. CD4⁺ and CD8⁺ T-cells, which play a role in responses to malaria,³¹ as well as partially explain sex differences in hypertension,³² could possibly explain the sex differences that we observed. Theoretically, BP could be modulated by exposure to antimalarial treatment, but this seems an unlikely explanation because no commonly used antimalarial drug is

Table 2. Age-Specific Effects of SCT on BP by Study Site

Age, y	No.	24-h BP			Nighttime BP			Daytime BP					
		SBP	95% CI	DBP	SBP	95% CI	DBP	SBP	95% CI	DBP			
Kilifi													
10 to 29	494	-0.1	-3.2 to 1.3	-0.05	-1.6 to 1.5	-0.8	-3.2 to 1.7	0.05	-1.7 to 1.8	-0.1	-4.0 to 0.9	-0.1	-1.6 to 2.0
30 to 59	384	-6.1	-10.5 to -1.8	-4.5	-7.4 to -1.6	-7.6	-12 to -2.9	-4.9	-7.9 to -2.0	-4.7	-9.1 to -0.3	-3.9	-7.0 to -0.8
≥60	249	0.2	-6.1 to 6.5	1.0	-2.8 to 4.8	-1.3	-8.7 to 6.1	-0.3	-4.4 to 3.9	1.6	-4.6 to 7.8	1.9	-2.1 to 5.8
Nairobi and Jackson pooled together*													
10 to 29	399	0.6	-2.3 to 3.5	0.6	-1.2 to 2.5	1.1	-1.8 to 4.0	0.9	-1.2 to 2.8	-0.4	-3.6 to 2.8	-0.04	-2.2 to 2.1
30 to 59	389	-1.3	-6.1 to 3.4	-1.3	-4.4 to 1.8	-1.5	-6.8 to 3.9	-0.5	-3.8 to 3.2	-1.0	-5.7 to 3.7	-1.2	-4.5 to 2.1
≥60	378	3.8	-1.4 to 8.9	4.1	0.8-7.3	2.1	-3.8 to 8.1	3.5	-0.2 to 7.0	4.8	-0.5 to 10	4.8	1.4-8.2

Results of linear regression models adjusted for age, sex, and estimated glomerular filtration rate. BP indicates blood pressure; DBP, diastolic blood pressure; SBP, systolic blood pressure; SCT, sickle cell trait.

*Participants in Jackson were 21 years and older.

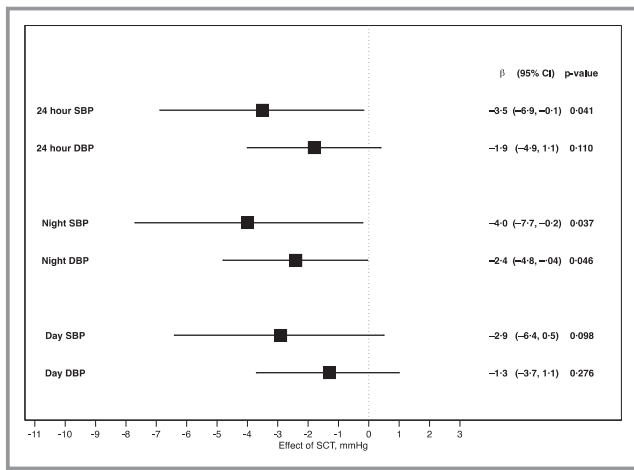


Figure 3. Effect of sickle cell trait on blood pressure (BP) in Kilifi vs Nairobi and Jackson pooled together. Points (and bars) represent the difference in BP (and 95% CI), measured in mm Hg, associated with sickle cell trait (SCT) in Kilifi (a malaria site) vs Nairobi/Jackson (nonmalaria sites). Estimates were derived as the interaction term (malaria vs nonmalaria sites) in a linear regression of BP by SCT status after adjusting for age, sex, and estimated glomerular filtration rate. DBP indicates diastolic blood pressure; SBP, systolic blood pressure.

known to elevate BP over the long term. Elucidating mechanisms by which malaria leads to higher BP could drive the design of novel therapies for the prevention or treatment of hypertension.

We studied populations in Nairobi and Jackson that had not been exposed to malaria to exclude the possibility that the lower BP associated with SCT in Kilifi could have been caused by factors other than malaria (illustrated in Figure S4). Although the study was not powered to detect small changes in BP at these sites, there was no statistically significant difference in BP by SCT status at these 2 sites and there was no evidence of heterogeneity by site. Other studies also suggest that SCT does not influence BP independently. In a study of 15 975 black patients, where SCT was associated with an increased risk of kidney disease, there was no difference in baseline clinic BP, based on SCT.²³ There was no difference in incident hypertension among participants with or without SCT in an analysis of 1995 black patients followed for 25 years.³³ A preliminary analysis from the present data set, restricted to 11- to 17-year-old Nairobi residents, showed that SCT and α^+ thalassemia do not directly influence BP.^{34,35} In addition, large genome-wide association studies have not reported a statistically significant association between SCT and BP.^{36–38}

To invalidate the interpretation of malaria as the cause of elevated BP in this study it would be necessary for SCT and α^+ thalassemia, or loci in linkage disequilibrium with them, to be associated with a large reduction in BP. This is highly unlikely because: (1) none of the loci in the Bantu/Central African Republic haplotype that is predominant among individuals with

SCT in Kilifi has been associated with BP traits; (2) studies in the United States show that SCT does not reduce the risk of hypertension-related outcomes such as stroke, heart failure, and chronic kidney disease,^{23,39,40} as would be expected if it lowered BP; and (3) most genetic polymorphisms influencing BP tend to have small effects.^{36,37}

Study Strengths

This study had several strengths. First was the use of ABPM, considered the reference standard for measuring BP.⁴¹ By performing multiple repeated measurements, ABPM gives more accurate estimates of BP.²⁰ The largest differences were observed in the primary analyses that utilized more stringent quality criteria, which provided for better accuracy in measurement without introducing a selection bias. In addition, in both the primary and sensitivity analyses, the largest differences were observed when comparing nighttime measurements, which are less susceptible to interference by daytime activities. Nocturnal BP indices are also more predictive of cardiovascular outcomes than daytime or 24-hour values.^{20,42,43} Second, the Mendelian randomization approach that we used is a robust design for elucidating causal relationships.⁵ We used genetic variants that are strongly associated with malaria and showed associations with the outcomes that were consistent with the different levels of protection against malaria afforded by each variant. Third, study participants in Kilifi were of the same ethnicity, minimizing the possibility that population stratification could explain the differences observed. Fourth, we used prospectively collected health and demographic surveillance system records^{15,16} to ascertain residence in malaria/nonmalaria sites.

Study Limitations

Although Mendelian randomization is a well-established method for inferring causality, there are some residual limitations. As we did not have medical record data for the participants in Kilifi, we could not determine the timing, number, or severity of malaria episodes required to elevate BP in adult life. These questions could be investigated using sequential birth cohort studies that take into account the fact that there has been a marked reduction in malaria transmission in Kilifi from the year 2000.⁷ In addition, replication studies in other areas with malaria transmission are needed to confirm the observations we made in Kilifi.

Conclusions

SCT was associated with lower BP and reduced prevalence of hypertension in Kilifi but not in Nairobi, Kenya, or Jackson, Mississippi, an observation compatible with a causal association between malaria and higher BP. One implication is that

malaria elimination would lead to health benefits well beyond those currently described. A second implication is that elucidating the mechanisms by which malaria leads to an elevation in BP could yield new preventive strategies for hypertension and consequent cardiovascular disease.

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Disclosures

None.

References

- Ibrahim MM, Damasceno A. Hypertension in developing countries. *Lancet*. 2012;380:611–619.
- Snow RW, Sartorius B, Kyalo D, Maina J, Amratia P, Mundia CW, Bejon P, Noor AM. The prevalence of *Plasmodium falciparum* in sub-Saharan Africa since 1900. *Nature*. 2017;550:515–518.
- Etyang AO, Smeeth L, Cruickshank JK, Scott JA. The malaria-high blood pressure hypothesis. *Circ Res*. 2016;119:36–40.
- Ayoola OO, Omotade OO, Gemmell I, Clayton PE, Cruickshank JK. The impact of malaria in pregnancy on changes in blood pressure in children during their first year of life. *Hypertension*. 2014;63:167–172.
- Davey Smith G, Ebrahim S. 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? *Int J Epidemiol*. 2003;32:1–22.
- Mbogho CN, Snow RW, Kabiru EW, Ouma JH, Githure JI, Marsh K, Beier JC. Low-level *Plasmodium falciparum* transmission and the incidence of severe malaria infections on the Kenyan coast. *Am J Trop Med Hyg*. 1993;49:245–253.
- O'Meara WP, Bejon P, Mwangi TW, Okiro EA, Peshu N, Snow RW, Newton CR, Marsh K. Effect of a fall in malaria transmission on morbidity and mortality in Kilifi, Kenya. *Lancet*. 2008;372:1555–1562.
- Snow RW, Kibuchi E, Karuri SW, Sang G, Gitonga CW, Mwandawiro C, Bejon P, Noor AM. Changing malaria prevalence on the Kenyan coast since 1974: climate, drugs and vector control. *PLoS One*. 2015;10:e0128792.
- Mudhune SA, Okiro EA, Noor AM, Zurovac D, Juma E, Ochola SA, Snow RW. The clinical burden of malaria in Nairobi: a historical review and contemporary audit. *Malar J*. 2011;10:138.
- Noor AM, Gething PW, Alegana VA, Patil AP, Hay SI, Muchiri E, Juma E, Snow RW. The risks of malaria infection in Kenya in 2009. *BMC Infect Dis*. 2009;9:180.
- Williams LL Jr. Malaria eradication in the United States. *Am J Public Health Nations Health*. 1963;53:17–21.
- Malaria Genomic Epidemiology Network. Reappraisal of known malaria resistance loci in a large multicenter study. *Nat Genet*. 2014;46:1197–1204.
- Williams TN, Mwangi TW, Wambua S, Alexander ND, Kortok M, Snow RW, Marsh K. Sick cell trait and the risk of *Plasmodium falciparum* malaria and other childhood diseases. *J Infect Dis*. 2005;192:178–186.
- Williams TN, Mwangi TW, Roberts DJ, Alexander ND, Weatherall DJ, Wambua S, Kortok M, Snow RW, Marsh K. An immune basis for malaria protection by the sickle cell trait. *PLoS Med*. 2005;2:e128.
- Scott JA, Bauni E, Moisi JC, Ojal J, Gatakaa H, Nyundo C, Molyneux CS, Kombe F, Tsoka B, Marsh K, Peshu N, Williams TN. Profile: the Kilifi Health and Demographic Surveillance System (KHDSS). *Int J Epidemiol*. 2012;41:650–657.
- Beguy D, Elung'ata P, Mberu B, Oduor C, Wamukoya M, Nganyi B, Ezeh A. Health & demographic surveillance system profile: the Nairobi Urban Health and Demographic Surveillance System (NUHDSS). *Int J Epidemiol*. 2015;44:462–471.
- Flint J, Hill AV, Bowden DK, Oppenheimer SJ, Sill PR, Serjeantson SW, Bana-Koiri J, Bhatia K, Alpers MP, Boyce AJ, Weatherall DJ, Clegg JB. High frequencies of alpha-thalassaemia are the result of natural selection by malaria. *Nature*. 1986;321:744–750.
- Foy H, Kondi A, Timms GL, Brass W, Bushra F. The variability of sickle cell rates in the tribes of Kenya and the Southern Sudan. *Br Med J*. 1954;1:294–297.
- Taylor HA Jr. The Jackson Heart Study: an overview. *Ethn Dis*. 2005;15:S6–13.
- O'Brien E, Parati G, Stergiou G, Asmar R, Beilin L, Bilo G, Clement D, de la Sierra A, de Leeuw P, Dolan E, Fagard R, Graves J, Head GA, Imai Y, Kario K, Lurbe E, Mallion JM, Mancia G, Mengden T, Myers M, Ogedegbe G, Ohkubo T, Omboni S, Palatini P, Redon J, Ruilope LM, Shennan A, Staessen JA, vanMontfrans G, Verdecchia P, Waeber B, Wang J, Zanchetti A, Zhang Y; European Society of Hypertension Working Group on Blood Pressure Monitoring. European Society of Hypertension position paper on ambulatory blood pressure monitoring. *J Hypertens*. 2013;31:1731–1768.
- Lurbe E, Agabiti-Rosei E, Cruickshank JK, Dominiczak A, Erdine S, Hirth A, Invitti C, Litwin M, Mancia G, Pall D, Rascher W, Redon J, Schaefer F, Seeman T, Sinha M, Stabouli S, Webb NJ, Wuhl E, Zanchetti A. 2016 European Society of Hypertension guidelines for the management of high blood pressure in children and adolescents. *J Hypertens*. 2016;34:1887–1920.
- Williams B, Mancia G, Spiering W, Agabiti Rosei E, Azizi M, Burnier M, Clement DL, Coca A, de Simone G, Dominiczak A, Kahan T, Mahfoud F, Redon J, Ruilope L, Zanchetti A, Kerins M, Kjeldsen SE, Kreutz R, Laurent S, Lip GYH, McManus R, Narkiewicz K, Ruschitzka F, Schmieder RE, Shlyakhto E, Tsioufias C, Aboyans V, Desormais I; ESC Scientific Document Group. 2018 ESC/ESH guidelines for the management of arterial hypertension. *Eur Heart J*. 2018;39:3021–3104.
- Naik RP, Derebail VK, Grams ME, Franceschini N, Auer PL, Peloso GM, Young BA, Lettre G, Peralta CA, Katz R, Hyacinth HI, Quarells RC, Grove ML, Bick AG, Fontanillas P, Rich SS, Smith JD, Boerwinkle E, Rosamond WD, Ito K, Lanzkrone S, Coresh J, Correa A, Sarto GE, Key NS, Jacobs DR, Kathiresan S, Bibbins-Domingo K, Kshirsagar AV, Wilson JG, Reiner AP. Association of sickle cell trait with chronic kidney disease and albuminuria in African Americans. *JAMA*. 2014;312:2115–2125.
- Williams TN, Mwangi TW, Wambua S, Peto TE, Weatherall DJ, Gupta S, Recker M, Penman BS, Uyoga S, Macharia A, Mwacharo JK, Snow RW, Marsh K. Negative epistasis between the malaria-protective effects of alpha-thalassaemia and the sickle cell trait. *Nat Genet*. 2005;37:1253–1257.
- He FJ, Li J, Macgregor GA. Effect of longer term modest salt reduction on blood pressure: cochrane systematic review and meta-analysis of randomised trials. *BMJ*. 2013;346:f1325.
- Heart Outcomes Prevention Evaluation Study I, Yusuf S, Sleight P, Pogue J, Bosch J, Davies R, Dagenais G. Effects of an angiotensin-converting-enzyme inhibitor, ramipril, on cardiovascular events in high-risk patients. *N Engl J Med*. 2000;342:145–153.

27. Lewington S, Clarke R, Qizilbash N, Peto R, Collins R; Prospective Studies C. Age-specific relevance of usual blood pressure to vascular mortality: a meta-analysis of individual data for one million adults in 61 prospective studies. *Lancet*. 2002;360:1903–1913.
28. Hill AV, Allsopp CE, Kwiatkowski D, Anstey NM, Twumasi P, Rowe PA, Bennett S, Brewster D, McMichael AJ, Greenwood BM. Common West African HLA antigens are associated with protection from severe malaria. *Nature*. 1991;352:595–600.
29. Rodriguez-Iturbe B, Pons H, Johnson RJ. Role of the immune system in hypertension. *Physiol Rev*. 2017;97:1127–1164.
30. Adeyemi AB, Adediran IA, Kuti O, Owolabi AT, Durosimi MA. Outcome of pregnancy in a population of Nigerian women with sickle cell trait. *J Obstet Gynaecol*. 2006;26:133–137.
31. Bediako Y, Ngoi JM, Nyangweso G, Wambua J, Opiyo M, Nduati EW, Bejon P, Marsh K, Ndungu FM. The effect of declining exposure on T cell-mediated immunity to *Plasmodium falciparum*—an epidemiological “natural experiment”. *BMC Med*. 2016;14:143.
32. Gillis EE, Sullivan JC. Sex differences in hypertension: recent advances. *Hypertension*. 2016;68:1322–1327.
33. Liem RI, Chan C, Vu TT, Fornage M, Thompson AA, Liu K, Carnethon MR. Association among sickle cell trait, fitness, and cardiovascular risk factors in CARDIA. *Blood*. 2017;129:723–729.
34. Etyang AO, Khayeka-Wandabwa C, Kapesa S, Muthumbi E, Odipo E, Wamukoya M, Ngomi N, Haregu T, Kyobutungi C, Tendwa M, Makale J, Macharia A, Cruickshank JK, Smeeth L, Scott JA, Williams TN. Blood pressure and arterial stiffness in Kenyan adolescents with alpha-thalassemia. *J Am Heart Assoc*. 2017;6:e005613. DOI: 10.1161/JAHA.117.005613.
35. Etyang AO, Wandabwa CK, Kapesa S, Muthumbi E, Odipo E, Wamukoya M, Ngomi N, Haregu T, Kyobutungi C, Williams TN, Makale J, Macharia A, Cruickshank JK, Smeeth L, Scott JA. Blood pressure and arterial stiffness in Kenyan adolescents with the sickle cell trait. *Am J Epidemiol*. 2018;187:199–205.
36. Warren HR, Evangelou E, Cabrera CP, Gao H, Ren M, Mifsud B, Ntalla I, Surendran P, Liu C, Cook JP, Kraja AT, Drenos F, Loh M, Verweij N, Marten J, Karaman I, Lepe MP, O'Reilly PF, Knight J, Snieder H, Kato N, He J, Tai ES, Said MA, Porteous D, Alver M, Poulter N, Farrall M, Gansevoort RT, Padmanabhan S, Magi R, Stanton A, Connell J, Bakker SJ, Metspalu A, Shields DC, Thom S, Brown M, Sever P, Esko T, Hayward C, van der Harst P, Saleheen D, Chowdhury R, Chambers JC, Chasman DI, Chakravarti A, Newton-Cheh C, Lindgren CM, Levy D, Kooner JS, Keavney B, Tomaszewski M, Samani NJ, Howson JM, Tobin MD, Munroe PB, Ehret GB, Wain LV; International Consortium of Blood Pressure (ICBP) 1000G Analyses, BIOS Consortium, Lifelines Cohort Study, Understanding Society Scientific group, CHD Exome + Consortium, Exome BP Consortium, T2D-GENES Consortium, Go T2DGenes Consortium, Cohorts for Heart and Ageing Research in Genome Epidemiology (CHARGE) BPEC BP Exome Consortium, International Genomics of Blood Pressure (iGEN-BP) Consortium, UK Biobank CardioMetabolic Consortium BP working group. Genome-wide association analysis identifies novel blood pressure loci and offers biological insights into cardiovascular risk. *Nat Genet*. 2017;49:403–415.
37. Padmanabhan S, Caulfield M, Dominiczak AF. Genetic and molecular aspects of hypertension. *Circ Res*. 2015;116:937–959.
38. Staley JR, Blackshaw J, Kamat MA, Ellis S, Surendran P, Sun BB, Paul DS, Freitag D, Burgess S, Danesh J, Young R, Butterworth AS. PhenoScanner: a database of human genotype-phenotype associations. *Bioinformatics*. 2016;32:3207–3209.
39. Caughey MC, Loehr LR, Key NS, Derebail VK, Gottesman RF, Kshirsagar AV, Grove ML, Heiss G. Sickle cell trait and incident ischemic stroke in the Atherosclerosis Risk in Communities study. *Stroke*. 2014;45:2863–2867.
40. Bello NA, Hyacinth HI, Roetker NS, Seals SR, Naik RP, Derebail VK, Kshirsagar AV, Key NS, Wilson JG, Correa A, Adams RJ, Egede LD, Longstreth WT Jr, Choudhary G, Gee BE, Hughes AL, Shah AM, Manson JE, Allison M, Burke GL, Folsom AR, Carty CL, Reiner AP, Solomon SD, Konety SH. Sickle cell trait is not associated with an increased risk of heart failure or abnormalities of cardiac structure and function. *Blood*. 2017;129:799–801.
41. Flynn JT, Daniels SR, Hayman LL, Maahs DM, McCrindle BW, Mitsnefes M, Zachariah JP, Urbina EM; American Heart Association Atherosclerosis Hypertension and Obesity in Youth Committee of the Council on Cardiovascular Disease in the Young. Update: ambulatory blood pressure monitoring in children and adolescents: a scientific statement from the American Heart Association. *Hypertension*. 2014;63:1116–1135.
42. Hansen TW, Li Y, Boggia J, Thijs L, Richart T, Staessen JA. Predictive role of the nighttime blood pressure. *Hypertension*. 2011;57:3–10.
43. Roush GC, Fagard RH, Salles GF, Pierdomenico SD, Reboldi G, Verdecchia P, Eguchi K, Kario K, Hoshida S, Polonia J, de la Sierra A, Hermida RC, Dolan E, Fapohunda J; ABC-H Investigators. Prognostic impact of sex-ambulatory blood pressure interactions in 10 cohorts of 17 312 patients diagnosed with hypertension: systematic review and meta-analysis. *J Hypertens*. 2015;33:212–220.

Supplemental Material

Data S1.

Study Procedures

a) Study Procedures in Kilifi and Nairobi, Kenya

Participants were recruited from December 2015 to June 2017. We recruited participants aged ≥ 10 years that were lifelong residents of the Nairobi Urban Health and Demographic Surveillance System¹ and the Kilifi Health and Demographic Surveillance System² respectively (locations shown in Figure S1). Lifelong residency was confirmed using prospectively collected residency data from enumeration rounds that are conducted every 3-4 months within the demographic surveillance systems. Continuous residency was required in order to minimize misclassification of exposure to malaria as Nairobi and Kilifi have markedly contrasting malaria transmission patterns. Study participants in Nairobi were randomly selected from those who had self-identified as belonging to ethnic groups known to have a high frequency of malaria protective polymorphisms (Luhya, Luo, Teso, Mijikenda) as a result of hailing from parts of Kenya that are known to be endemic for malaria.^{3, 4} Study participants in Kilifi were predominantly from the Chonyi subtribe of the Mijikenda community. The prevalence of hypertension within the Kilifi Health and Demographic Surveillance System which covers an area of 900km² is ~17%.⁵ However there are significant differences in the incidence to death due to stroke within the study area. Chasimba where >75% of study participants came from has an incidence of death due to stroke that is three times that of Kilifi township, suggesting that there are local geographical differences in the prevalence of hypertension which is the main risk factor for stroke.

In both Kilifi and Nairobi, trained study staff visited all individuals who had been selected to participate in the study at their homes and requested them to come to the

study clinic to undergo study procedures. Those who failed to come to the clinic within 3 months of being requested to do so were considered to have declined our invitation to participate in the study.

At the clinic participants first underwent an interview where they answered questions about their past medical history and their socioeconomic status based on the multi-dimensional poverty (MDP) index.⁶ Weight and height were measured using a validated SECA 874™ weighing machine and a portable stadiometer (Seca 213™), respectively. Body mass index was calculated as the weight in kilograms divided by height in meters squared (kg/m²). We did not classify BMI by age-category in the adolescents that we studied. Mid-upper arm circumference (MUAC) was measured in a standardized manner using TALC™ MUAC tapes. All participants were subsequently fitted with a validated Arteriograph24™ (TensioMed Ltd., Budapest, Hungary) device for 24-hour ABPM measurement.⁷ The devices were attached on the non-dominant arm and were programmed to take measurements every 20 minutes during daytime hours (0600-2200 hrs) and every 40 minutes at night (2200-0600 hrs). At the end of the 24-hour period, participants returned to the study clinic where the Arteriograph was removed and data downloaded onto computers that would later (within 12 hours) synchronize their data onto an MySQL database hosted on servers located at the KEMRI-Wellcome Trust Research Programme offices in Kilifi, Kenya.

We collected 10ml of blood from participants for full blood count, determination of sickle hemoglobin status and serum electrolytes. After performing automated full blood counts using an ACT 5™ machine, whole blood samples were frozen at -80°C and then transported to the KEMRI-Wellcome Trust Research Programme laboratories in Kilifi, Kenya for determination of sickle hemoglobin status. DNA was

extracted retrospectively from the frozen samples by use of Qiagen™ DNA blood mini-kits (Qiagen, Crawley, United Kingdom) and typed for sickle hemoglobin and α^+ thalassemia using polymerase chain reaction. Glycosylated hemoglobin levels were determined using the Biorad™ D-10 machine (Bio-rad Laboratories Inc, Hercules, California).

Serum and urine samples collected from participants were frozen at -80°C within 4 hours of collection and later transported to the laboratories in Kilifi for analysis. We determined urea and creatinine levels in these samples using ion electrophoresis and the jaffe method, respectively.⁸ Creatinine measurements were performed using Isotope dilution mass spectrometry traceable methods. In addition, we determined albumin levels in the urine samples by immunoturbidometry using a Quantex™ microalbumin kit. Estimated glomerular filtration rate (eGFR) was calculated using the Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) equation in adults and the Schwartz equation in those aged ≤ 16 years.^{9, 10}

b) Study Procedures in Jackson, Mississippi, USA

The Jackson Heart Study (JHS)¹¹ is a population-based prospective cohort study, which was designed to evaluate cardiovascular disease risk among blacks. The JHS enrolled 5306 noninstitutionalized blacks, aged ≥ 21 years, between 2000 and 2004. The participants were recruited from the Atherosclerosis Risk in the Community site in Jackson, MS, and a representative sample of urban and rural Jackson, MS, metropolitan tricounty (Hinds, Madison, and Rankin counties) residents, volunteers, randomly selected individuals, and secondary family members.¹² The current analysis was restricted to JHS participants who completed ABPM soon after the baseline study visit (visit 1).

During in-home interviews, trained African American interviewers administered standardized questionnaires to collect self-reported information on socio-demographics (e.g. age, sex, education, marital status and socioeconomic status), previously diagnosed co-morbid conditions and selected health-related behaviors (e.g. current smoking). Weight and height were measured during a clinic visit. At the clinic visit, blood samples were collected for full blood count, genetic studies and determination of serum sodium, potassium and creatinine concentrations. 24-hour urine samples were collected for determination of creatinine and albumin concentrations. Full blood counts were performed using the Coulter GenS machine (BeckmanCoulter, Hialeah, Florida, USA). DNA was extracted from whole blood samples using Puregene reagents (Gentra System, Minneapolis, USA) and genetic studies were performed as previously described.¹² Biochemical tests were performed using a Vitros 950 or 250 Ortho-Clinical Diagnostics analyzer (Raritan, new Jersey, USA). Urine albumin was measured on a Dade-Behring BN 11 nephelometer (dade-Behring, Newark, Delaware, USA). All tests were performed at the University of Minnesota laboratory with the exception of hematology tests, which were done at the University of Mississippi Medical Center.¹³ Creatinine measurements were performed using Isotope dilution mass spectrometry traceable methods. Participants were fitted with an ABPM device (Spacelabs 90207, Spacelabs, Redmond, WA) on their non-dominant arm. Ambulatory BP was recorded every 20 minutes. After 24 hours, the device was removed, and data were downloaded onto a computer and processed with Medifacts International's Medicom software (Rockville, MD)

Statistical Methods and Considerations

a) Reporting format

While there are no specific guidelines for reporting Mendelian randomization (MR) studies, the principles outlined in the Strengthening the Reporting of Observational studies (STROBE)¹⁴ guidelines as well as the STROBE Extension for Genetic Association studies (STREGA)¹⁵ were used. Reporting was also guided by the review by Boef *et al.* of the quality of reporting of MR studies.¹⁶

b) Sample size estimation

The sample size calculation for Kilifi was based on a two-sample t-test comparing mean 24-hour systolic blood pressure in those with and without the sickle cell trait (SCT). The following assumptions were made:

- That the prevalence of SCT would be $\geq 15\%$ ¹⁷
- That the standard deviation of 24-hour systolic BP would be ≤ 15 mm Hg^{5, 18}

Based on these assumptions we calculated that, for Kilifi, we would need a minimum of 1115 participants with complete data in order to detect a statistically significant 4 mm Hg difference in 24-hour systolic BP with at least 80% statistical power.

For participants in Nairobi/Jackson we assumed that the combined SCT prevalence for the two sites would be $\geq 10\%$.¹⁹ Other assumptions were similar to those for Kilifi.

Based on these assumptions we calculated that for Nairobi/Jackson, we would need minimum of 1270 participants with complete data in order to detect a statistically significant 4 mm Hg difference in 24-hour systolic BP with at least 80% statistical power.

We assumed that with these numbers, we would achieve enough power for the primary outcome measure, a linear regression to determine the effect of SCT on 24-hour BP measures, while adjusting for age, sex and estimated glomerular filtration

rate (eGFR). The literature suggests that the major consideration for sample size calculations in linear regression models is to ensure that there are at least 2-50 individuals per variable in the model²⁰, a requirement that would almost certainly be achieved if most of the assumptions stated above held true.

c) Quality control criteria for ABPM data

There are 2 internationally recognized quality control criteria used for ABPM data, which are based on completeness of observations. The International Database of Ambulatory blood pressure in relation to Cardiovascular Outcomes (IDACO) study²¹ defined ABPM data as acceptable if they include ≥ 10 daytime and ≥ 5 nighttime readings, where daytime is defined as 1000-2000 hrs and nighttime as 0000-0600 hrs.²¹ The guidelines from the European Society of Hypertension (ESH) are more stringent; they require ≥ 20 daytime and ≥ 7 nighttime readings where daytime is defined as 0900 to 2100 hrs and nighttime as 0100 to 0600 hours.²² It is important to note that these criteria were arbitrarily set by experts and were not based on outcome studies. As the ESH criteria are more stringent they are likely to lead to a greater loss of data and subsequent loss of power and precision. However, in order to reduce measurement bias and obtain as accurate an effect size as possible, an *a priori* decision was made to restrict our primary analysis to data that met the ESH criteria.

d) Primary and secondary outcome measures

The primary outcome measure was estimated using a linear regression model to determine the effect of SCT on 24-hour systolic blood pressure, after adjusting for age, sex and estimated glomerular filtration rate (eGFR). Blood pressures were obtained by ambulatory blood pressure monitoring using the Arteriograph24™

device.⁷ Numerous studies have shown that the more accurate measurements resulting from repeated inflations and more standardized procedures in ABPM make it a much better predictor of cardiovascular events than other BP measurement methods.²² The justification for adjusting for age, sex and eGFR is given in the section below on confounders and model building.

Secondary outcome measures were defined as follows:

- a) effect of α^+ thalassaemia on 24-hour, daytime and nighttime systolic blood pressures, after adjusting for age, sex and estimated glomerular filtration rate
- b) effect of SCT on 24-hour, daytime and nighttime diastolic blood pressures, after adjusting for age, sex and estimated glomerular filtration rate
- c) effect of α^+ thalassaemia on 24-hour, daytime and nighttime diastolic blood pressures, after adjusting for age, sex and estimated glomerular filtration rate
- d) prevalence ratio for hypertension in those with and without SCT using log-binomial regression, adjusting for age, sex and estimated glomerular filtration rate

Hypertension was diagnosed by any one of the following criteria in individuals aged ≥ 16 years: ^{22, 23}

- i) 24-hours systolic BP ≥ 130 mmHg and/or 24-hour diastolic BP ≥ 90 mm Hg
- ii) Daytime systolic BP ≥ 135 mm Hg and/or daytime diastolic BP ≥ 85 mm Hg
- iii) Nighttime systolic BP ≥ 120 mm Hg and/or nighttime diastolic BP ≥ 70 mm Hg

Adjustment for multiple testing was not considered necessary in this scenario of a limited number of clinically relevant pre-specified tests (e.g. compared to GWAS studies)²⁴

e) Adjusting for confounders and model building

The theoretical basis for the malaria-high blood pressure hypothesis has been published previously.²⁵ Briefly, the primary hypothesis is that individuals in Kilifi who were exposed to more malaria disease in childhood (represented by those having haemoglobin AA) would have higher 24-hour systolic blood pressure than those who were exposed to less malaria disease (represented by those having haemoglobin AS [SCT]). The proposed causal diagram drawn purely for purposes of informing the analytical plan can be found in Figure S2.

For purposes of this analysis it is important to note that because malnutrition and stunting are on the causal pathway from malaria to the outcome, adjustment for body mass index (BMI) and other anthropometric indices (e.g. mid upper arm circumference) would be inappropriate.

i) Confounders

The principle of Mendelian randomization holds that because comparisons are based on genetic traits acquired at conception, any relationships between the genetic trait and the outcome are unlikely to be confounded by other exposures as these will be randomly distributed between carriers and non-carriers of the trait.²⁶ However age, sex, and BMI are known to have a very strong influence on BP and other cardiovascular diseases²⁷, and are usually adjusted for as 'fixed covariates' in MR/Genome wide association studies²⁸⁻³¹. We have outlined above why it would be inappropriate to adjust for BMI.

Sickle cell trait has been associated with impaired kidney function as measured by decline in estimated glomerular filtration rate (eGFR) and albuminuria.¹⁹ This association is independent of blood pressure elevation. Impaired kidney function is associated with elevations in blood pressure as a result of sodium retention³², increased activity of the renin-angiotensin system³³, increased sympathetic activity³⁴, secondary hyperparathyroidism³⁵, impaired nitric oxide synthesis³⁶ and increased prevalence of nocturnal non-dipping BP pattern.³⁷ It is also possible that kidney disease could arise from hypertension.³⁸ The direction of the relationship between blood pressure and kidney function, has been the subject of debate.³⁹ However, evidence from genetic studies suggests that the association between renal function and blood pressure is likely to be explained by decreased renal function giving rise to high blood pressure. In a large (n>200,000) genome wide association study (GWAS), loci that were associated with blood pressure elevation and cardiovascular disease showed no association with kidney disease or kidney function.²⁹ If SCT compromises renal function and this in turn leads to elevated blood pressure, this would result in a bias toward a null result when using SCT as a proxy for testing the malaria-high blood pressure hypothesis. As can be seen in Figure S3, impaired kidney function (as measured by eGFR) is associated with both the exposure and the outcome, but is not on the causal pathway from malaria to the outcome. Kidney function is therefore considered a confounder and we adjusted for eGFR in all regression analyses. We also examined the effect of using urine albumin to creatinine ratio in place of eGFR in the regression models.

If, however, renal function lies on the causal pathway between malaria and high blood pressure it would be inappropriate to include it within the regression models. Severe malaria does occasionally present with acute renal failure and repeated episodes of malaria could result in chronic pyelonephritis and elevated BP. However, acute renal failure is a very rare complication of malaria in Kilifi, for example, it occurred in 2 out of 1844 children admitted with malaria.⁴⁰ This suggests that renal failure is an unlikely mediator of the potential association between malaria and elevated BP.

We confirmed that each of the *a priori* specified covariates (age, sex and eGFR) significantly improved the regression models using the likelihood ratio test.

Confounding due to pleiotropy

A special type of confounding can also occur if the genetic trait influences the outcome through a pathway that is independent of the exposure (pleiotropy)⁴¹ as illustrated in Figure S4.

In contrast with renal function, which is a known confounder and can be measured and adjusted for in regression analyses, confounding due to other (often unknown) causes can only be detected by examining the relationship between sickle cell trait and blood pressure in individuals who have not been exposed to malaria. The existence of pleiotropy can invalidate the use of the genetic trait as a marker for the infectious disease exposure. In order to exclude pleiotropy as a potential explanation for the association between SCT and BP, we studied lifelong residents of Nairobi, Kenya and Jackson, Mississippi, two sites where there is no malaria transmission. In addition, we conducted a pooled analysis incorporating data from the three study sites and conducted a linear regression with the previously specified covariates plus SCT and study site and their interaction. This increased the power to detect any

independent effect of SCT on BP while simultaneously checking for differential effect of SCT according to study site.

ii) Effect modifier: α^+ thalassemia

α^+ thalassemia, in which there is reduction in the amount of alpha hemoglobin, is common in regions where malaria transmission occurs. Williams et al⁴² have demonstrated a negative epistatic effect when α^+ thalassemia is coinherited with SCT. The effect of coinheritance of the mutations is to reduce the malaria protective effect of SCT to about 27% (from 50%) for uncomplicated malaria and to 44% (from 80%) for severe malaria.⁴² Put simply, the presence of α^+ thalassemia reduces the protective effect of SCT against both uncomplicated and severe malaria by about half. We therefore included α^+ thalassemia as an interaction term (interacting with SCT) in the main regression model and examined whether its inclusion changed the effect estimate for SCT in predicting blood pressure.

In a related analysis, we ran a linear regression model examining the effect of α^+ thalassemia on blood pressure with the same covariates used in the main analysis for SCT. Because α^+ thalassemia confers less protection against malaria than SCT, we expected that the effect estimate in this model would be lower than that of SCT.

f) Testing for cohort effects

Malaria incidence in Kilifi has been changing over time and we considered that this could influence results obtained. Data on the changing levels of transmission go back to 1990 and they show that a significant drop in transmission in Kilifi began around 1999-2000.⁴³ In addition, because blood pressure rises with age, it is possible that the effects of malaria on outcome measures may be more apparent later in life. While it is not possible to determine the individual contributions of

changing malaria exposure and aging to any differences observed in outcome measures, we attempted to display these differences by performing comparisons of the outcomes by sickle trait in 3 age strata.

Table S1. Characteristics of those who consented to undergo ABPM versus those who declined.

Characteristic	Kilifi N=2537					Nairobi N=1119					Jackson N=5306				
	Consented n=2371		Did not consent n=166		p-value	Consented n=1026		Did not consent n=93		p-value	Consented n=1148		Did not consent n=4158		p-value
n	(%)	n	(%)	n		(%)	n	(%)	n		(%)	n	(%)	n	
Female	1361	(54)	84	(51)	0.408	480	(47)	45	(48)	0.443	780	(68)	2591	(62)	<0.001
Mean Age, years	mean	(SD)	mean	(SD)	p-value	mean	(SD)	mean	(SD)	p-value	mean	(SD)	mean	(SD)	p-value
	39	(22)	48	(22)	<0.001	22	(16)	25	(17)	0.073	59	(11)	54	(13)	<0.001

Table S2. Characteristics of participants with and without good quality ABPM data.

Characteristic	Kilifi N=2371					Nairobi N=1026					Jackson N=1148				
	Met ESH criteria n=1140		Did not meet ESH criteria n=1231		p-value	Met ESH criteria n=542		Did not meet ESH criteria n=484		p-value	Met ESH criteria n=934		Did not meet ESH criteria n=214		p-value
	n	(%)	n	(%)		n	(%)	n	(%)		n	(%)	n	(%)	
Female	660	(58)	520	(48)	<0.001	290	(54)	237	(49)	0.300	630	(67)	150	(73)	0.134
Have sickle cell trait	240	(21)	205	(19)	0.240	83	(15)	50	(14)	0.637	58	(8.9)	10	(7.5)	0.588
Smoker	96	(8)	87	(8)	0.761	11	(2)	12	(3)	0.198	103	(11)	40	(19)	0.001
Previously diagnosed hypertensive [§]	165	(14)	133	(12)	0.140	62	(11)	23	(7)	0.017	560	(60)	122	(59)	0.779
On anti-hypertensive medication	36	(3)	16	(1.5)	0.009	9	(2)	3	(1)	0.315	519	(59)	116	(59)	0.930
	mean	(SD)	mean	(SD)	p-value	mean	(SD)	mean	(SD)	p-value	mean	(SD)	mean	(SD)	p-value
Age, years	40	(22)	36	(21)	<0.001	23	(17)	19	(15)		59	(11)	58	(12)	0.360
BMI kg/m ²	20.6	(3.8)	20.6	(4.0)	0.689	20.4	(4.1)	19.7	(4.0)	0.010	30.9	(6.5)	33.1	(5.8)	<0.001
HbA1c, %	5.1	(0.7)	5.1	(0.6)	0.134	5.3	(0.98)	5.4	(1.2)	0.043	6.0	(1.3)	6.1	(1.4)	0.776
Hemoglobin, g/dl	12.6	(1.7)	12.9	(1.6)	<0.001	13.3	(1.7)	13.4	(1.7)	0.421	13.0	(1.4)	13.0	(1.5)	0.661
WBC count X10 ⁹ /L	5.7	(1.4)	5.7	(1.6)	0.718	5.4	(1.6)	5.5	(1.4)	0.187	5.3	(1.6)	5.8	(1.9)	0.002
Platelet count X10 ⁹ /L	264	(86)	257	(81)	0.064	289	(100)	302	(88)	0.046	243	(62)	250	(69)	0.078
eGFR, ml/min/1.73m ²	113	(41)	119	(40)	<0.001	116	(25)	119	(24)	0.090	86.6	(25)	87.8	(25)	0.540
Log UACR, mg/g	1.2	(0.62)	1.2	(0.61)	0.674	1.3	(0.71)	1.40	(0.60)	0.345	0.90	(0.50)	0.90	(0.56)	0.860

ABPM: Ambulatory blood pressure monitoring; eGFR: Estimated glomerular filtration rate; ESH: European society of hypertension. UACR: urine albumin to creatinine ratio
ESH criteria require a minimum of 20 valid readings taken between 9 a.m. and 9 p.m. and a minimum of 7 valid readings taken between 1a.m. and 6 a.m. in order for ABPM readings to be considered as complete.

[§]Answered "yes" to the question: Has a doctor or healthcare worker previously told you that you have high blood pressure?

Table S3. Effect of sickle cell trait on blood pressure in Nairobi and Jackson.

ABPM measure	Nairobi (N=516)			Jackson (N=651)			p-value for interaction
	β (mm Hg)	(95% CI)	p value	β (mm Hg)	(95% CI)	p value	
24 hour SBP	0.6	(-2.5, 3.7)	0.722	0.6	(-3.0, 4.2)	0.732	0.944
24 hour DBP	0.5	(-1.6, 2.6)	0.652	0.8	(-1.5, 3.1)	0.489	0.695
Nighttime SBP	0.7	(-2.5, 4.0)	0.669	-0.2	(-4.3, 4.0)	0.938	0.766
Nighttime DBP	0.6	(-1.7, 2.9)	0.610	0.8	(-1.8, 3.3)	0.558	0.779
Daytime SBP	-0.1	(-3.4, 3.2)	0.963	1.3	(-2.3, 4.9)	0.486	0.567
Daytime DBP	0.1	(-2.3, 2.4)	0.951	1.2	(-1.2, 3.6)	0.310	0.409

Linear regression models adjusted for age, sex and estimated glomerular filtration rate

SBP: Systolic blood pressure

DBP: Diastolic blood pressure

p-value for interaction represents result of regression models testing for difference in effect of sickle cell trait on BP in Nairobi versus Jackson

Table S4. Effects of sickle cell trait on blood pressure: effect of excluding participants taking anti-hypertensive medication.

All participants

ABPM measure	Kilifi (N=1127)			Nairobi and Jackson (N=1166)		
	β (mm Hg)	(95% CI)	p value	β (mm Hg)	(95% CI)	p value
24 hour SBP	-2.4	(-4.7, -0.2)	0.037	0.7	(-1.6, 3.1)	0.542
24 hour DBP	-1.4	(-2.8, 0.1)	0.068	0.1	(-1.5, 1.8)	0.860
Nighttime SBP	-3.2	(-5.7, -0.6)	0.015	0.5	(-2.2, 3.1)	0.727
Nighttime DBP	-1.8	(-3.3, -0.2)	0.026	0.3	(-1.5, 2.0)	0.773
Daytime SBP	-1.9	(-4.2, 0.4)	0.113	0.7	(-1.7, 3.1)	0.566
Daytime DBP	-1.0	(-2.6, 0.6)	0.223	0.1	(-1.6, 1.8)	0.889

Excluding participants on medication

ABPM measure	Kilifi (N=1092)			Nairobi and Jackson (N=755)		
	β (mm Hg)	(95% CI)	p value	β (mm Hg)	(95% CI)	p value
24 hour SBP	-2.5	(-4.8, -0.2)	0.034	0.2	(-2.5, 2.8)	0.896
24 hour DBP	-1.4	(-2.8, 0.1)	0.067	-0.2	(-2.0, 1.5)	0.787
Nighttime SBP	-3.1	(-5.7, -0.6)	0.017	0.8	(-2.1, 3.6)	0.597
Nighttime DBP	-1.8	(-3.3, -0.2)	0.028	0.1	(-1.8, 2.1)	0.902
Daytime SBP	-2.0	(-4.3, 0.4)	0.096	-0.4	(-3.1, 2.4)	0.802
Daytime DBP	-1.0	(-2.6, 0.6)	0.205	-0.5	(-2.4, 1.5)	0.630

Linear regression models adjusted for age, sex and estimated glomerular filtration rate
 SBP: Systolic blood pressure
 DBP: Diastolic blood pressure

Table S5. Effect of Sickle cell trait on blood pressure by sex and study site.

Kilifi

ABPM measure	Women (N=659)			Men (N=473)			Interaction p-value
	β	(95% CI)	p value	β	(95% CI)	p value	
24 hour SBP	-3.7	(-7.1, -0.4)	0.028	-0.7	(-3.7, 2.3)	0.645	0.218
24 hour DBP	-1.9	(-2.8, 0.1)	0.079	-0.7	(-2.7, 1.3)	0.493	0.398
Nighttime SBP	-4.3	(-8.0, -0.6)	0.022	-1.6	(-5.1, 1.8)	0.356	0.335
Nighttime DBP	-2.0	(-4.2, 0.1)	0.067	-1.4	(-3.6, 0.8)	0.217	0.661
Daytime SBP	-3.1	(-6.4, 0.2)	0.068	-0.3	(-3.4, 2.9)	0.875	0.284
Daytime DBP	-1.6	(-3.9, 0.6)	0.154	-0.2	(-2.4, 2)	0.867	0.131

Nairobi and Jackson

ABPM measure	Women (N=705)			Men (N=461)			Interaction p-value
	β	(95% CI)	p value	β	(95% CI)	p value	
24 hour SBP	0.9	(-2.2, 4)	0.574	0.7	(-2.9, 4.4)	0.700	0.800
24 hour DBP	1.2	(-0.9, 3.3)	0.272	-0.4	(-2.9, 2.1)	0.737	0.106
Nighttime SBP	1.2	(-2.3, 4.7)	0.511	-0.2	(-4.2, 3.8)	0.915	0.512
Nighttime DBP	1.7	(-0.6, 3.9)	0.139	-0.8	(-3.5, 2.0)	0.586	0.046
Daytime SBP	0.7	(-2.5, 3.9)	0.679	1.0	(-2.8, 4.8)	0.617	0.998
Daytime DBP	0.8	(-1.4, 3.1)	0.464	-0.1	(-2.7, 2.5)	0.936	0.319

Linear regression models adjusted for age, sex and estimated glomerular filtration rate

SBP: Systolic blood pressure

DBP: Diastolic blood pressure

p-value for interaction represents result of regression models testing for difference in effect of sickle cell trait on BP in men versus women

Table S6. Results of interaction analysis comparing effect of sickle cell trait in Kilifi versus Nairobi/Jackson using log urine albumin to creatinine ratio as covariate instead of estimated Glomerular Filtration Rate.

	All (N=1583)			Women N=958			Men N=625		
	β	(95% CI)	p value	β	(95% CI)	p value	β	(95% CI)	p value
24 hour SBP	-5.2	(-9.5, -0.9)	0.019	-6.7	(-13, -0.6)	0.030	-3.5	(-9.5, 2.5)	0.249
24 hour DBP	-2.9	(-5.7, -0.1)	0.040	-4.6	(-8.5, -0.8)	0.019	-1.0	(-5.0, 3.1)	0.635
Nighttime SBP	-5.5	(-10, -0.7)	0.026	-6.3	(-13, 0.5)	0.067	-4.7	(-12, 2.1)	0.176
Nighttime DBP	-3.6	(-6.6, -0.6)	0.018	-5.1	(-9.1, -1.1)	0.013	-1.7	(-6.2, 2.8)	0.456
Daytime SBP	-4.8	(-9.2, -0.4)	0.032	-6.5	(-13, -0.4)	0.037	-3.2	(-9.5, 3.0)	0.308
Daytime DBP	-2.5	(-5.5, 0.5)	0.106	-4.1	(-8.3, 0.03)	0.052	-0.9	(-5.3, 3.4)	0.673

SCT: sickle cell trait

SBP: Systolic blood pressure

DBP: Diastolic blood pressure

Linear regression models tested for interaction in the effect of SCT by site. Other covariates were age, sex and log urine albumin to creatinine ratio

Table S7. Results of pooled analysis comparing effect of sickle cell trait in Kilifi versus Nairobi/Jackson stratified by sex.

ABPM measure	Women (N=1359)			Men (N=934)			Interaction p-value
	β	(95% CI)	p value	β	(95% CI)	p value	
24 hour SBP	-5.0	(-9.7, -0.2)	0.039	-1.9	(-6.6, 2.8)	0.431	0.419
24 hour DBP	-3.2	(-6.2, -0.1)	0.041	-0.3	(-3.5, 2.9)	0.871	0.110
Nighttime SBP	-5.8	(-11, -0.6)	0.030	-1.8	(-7.1, 3.5)	0.506	0.323
Nighttime DBP	-3.9	(-7.1, -0.7)	0.017	-0.5	(-4.0, 3.0)	0.782	0.086
Daytime SBP	-4.1	(-8.9, 0.7)	0.091	-1.8	(-6.7, 3.2)	0.489	0.549
Daytime DBP	-2.5	(-5.8, 0.7)	0.126	-0.2	(-3.6, 3.3)	0.925	0.207

Estimates were derived separately for each sex as the interaction term (malaria vs non-malaria sites) in a linear regression of blood pressure by sickle cell trait status after adjusting for age and estimated glomerular filtration rate. Interaction p-value is the result of 3-way interaction in regression models testing for difference in effect of sickle cell trait on BP in men versus women in Kilifi versus Nairobi and Jackson pooled together. DBP: diastolic blood pressure; SBP: systolic blood pressure.

Table S8. Effect of α^+ thalassemia on ambulatory blood pressure by study site**Kilifi (N=1125).**

ABPM measure	No. of -3.7kb α^+ thalassemia deletions					
	One deletion (Heterozygous)			Two deletions (Homozygous)		
	β mm Hg	(95% CI)	p value	β mm Hg	(95% CI)	p value
24 hour SBP	-0.1	(-2.2, 2.0)	0.921	-1.1	(-3.8, 1.6)	0.434
24 hour DBP	-0.3	(-1.7, 1.0)	0.651	-0.4	(-2.2, 1.3)	0.646
Nighttime SBP	0.2	(-2.2, 2.6)	0.879	-1.3	(-4.3, 1.8)	0.416
Nighttime DBP	-0.1	(-2.3, 1.4)	0.919	-0.5	(-2.4, 1.4)	0.597
Daytime SBP	-0.2	(-2.3, 2.0)	0.884	-1.0	(-3.7, 1.8)	0.500
Daytime DBP	-0.1	(-1.5, 1.4)	0.930	-0.3	(-2.2, 1.6)	0.765

Nairobi (N=514)

ABPM measure	No. of -3.7kb α^+ thalassemia deletions					
	One deletion (Heterozygous)			Two deletions (Homozygous)		
	β mm Hg	(95% CI)	p value	β mm Hg	(95% CI)	p value
24 hour SBP	1.1	(-1.3, 3.5)	0.373	3.9	(-0.5, 8.2)	0.083
24 hour DBP	0.6	(-1.0, 2.3)	0.440	2.6	(-0.4, 5.6)	0.086
Nighttime SBP	1.9	(-0.6, 4.4)	0.144	3.3	(-1.3, 7.9)	0.157
Nighttime DBP	1.8	(0.05, 3.6)	0.044	2.4	(-0.8, 5.6)	0.138
Daytime SBP	0.04	(-1.8, 2.6)	0.974	2.9	(-1.8, 7.5)	0.223
Daytime DBP	-0.3	(-2.1, 1.5)	0.748	2.2	(-1.0, 5.5)	0.181

SBP: Systolic blood pressure

DBP: Diastolic blood pressure

Linear regression models adjusted for age, sex and estimated glomerular filtration rate.

*No alpha thalassemia data were available for participants from Jackson

Results of analyses based on data meeting IDACO criteria for completeness

Table S9. Characteristics of study participants with and without sickle cell trait by study site (IDACO Criteria).

Characteristic	Kilifi N=2048				Nairobi N=835				Jackson N=724			
	SCT n=408		Non-SCT n=1640		SCT n=121		Non-SCT n=714		SCT n=63		Non-SCT n=661	
	n	(%)	n	(%)	n	(%)	n	(%)	n	(%)	n	(%)
Female	203	(50)	884	(54)	59	(49)	369	(52)	41	(65)	439	(66)
Smoker	34	(8)	134	(8)	5	(4)	17	(2)	9	(14)	83	(13)
Previously diagnosed hypertensive [§]	59	(15)	221	(14)	9	(7)	70	(10)	33	(52)	408	(62)
Taking antihypertensive medication	11	(3)	38	(2)	2	(2)	10	(1)	27	(44)	377	(61)
	mean	(SD)	mean	(SD)	mean	(SD)	mean	(SD)	mean	(SD)	mean	(SD)
Age, years	40	(22)	38	(22)	20	(14)	22	(16)	60	(12)	60	(11)
BMI kg/m ²	20.7	(3.8)	20.7	(3.9)	20.2	(4.0)	20.2	(4.1)	31.0	(6.9)	31.1	(6.4)
HbA1c, %	5.2	(0.6)	5.1	(0.7)	5.4	(1.3)	5.3	(1.0)	6.0	(1.2)	6.1	(1.3)
Hemoglobin, g/dl	12.7	(1.9)	12.7	(1.6)	13.4	(1.8)	13.3	(1.7)	12.8	(1.4)	13.0	(1.4)
WBC count X10 ⁹ /L	5.8	(1.5)	6.5	(22.5)	5.4	(1.3)	5.4	(1.5)	4.9	(1.2)	5.3	(1.5)
Platelet count X10 ⁹ /L	261	(89)	259	(81)	289	(96)	294	(97)	229	(57)	243	(62)
Plasma osmolality, mosm/Kg	290	(5.8)	290	(6.3)	291	(12)	291	(12)	-	-	-	-
eGFR, ml/min/1.73m ²	111	(39)	116	(40)	117	(27)	116	(24)	84	(27)	87	(26)
Log UACR, mg/g	1.3	(0.6)	1.2	(0.6)	1.5	(0.7)	1.3	(0.7)	1.1	(0.6)	0.9	(0.5)

BMI: Body mass index; eGFR: estimated glomerular filtration rate; HbA1c: glycosylated hemoglobin; SCT: Sickle cell trait; SD: standard deviation; UACR: urine albumin to creatinine ratio; WBC: white blood cell. Plasma osmolality measurements were not available for Jackson participants.

[§]Answered "yes" to the question: Has a doctor or healthcare worker previously told you that you have high blood pressure?

Table S10. Effect of sickle cell trait on blood pressure by malaria site (IDACO Criteria).

ABPM measure	Kilifi (N=1965)			Nairobi and Jackson (N=1500)			Nairobi/Jacks on Interaction
	β (mmHg)	(95% CI)	p value	β (mm Hg)	(95% CI)	p value	p-value for interaction
24 hour SBP	-1.4	(-3.1, 0.4)	0.128	0.4	(-1.6, 2.5)	0.697	0.557
24 hour DBP	-0.7	(-1.8, 0.4)	0.226	0.1	(-1.3, 1.5)	0.910	0.394
Nighttime SBP	-1.6	(-3.5, 0.4)	0.109	-0.4	(-2.6, 1.9)	0.744	0.622
Nighttime DBP	-0.9	(-2.1, 0.3)	0.146	-0.3	(-1.8, 1.2)	0.685	0.273
Daytime SBP	-0.9	(-2.7, 0.9)	0.327	0.6	(-1.6, 2.7)	0.602	0.264
Daytime DBP	-0.6	(-1.8, 0.7)	0.367	0.1	(-1.4, 1.6)	0.858	0.237

Linear regression models adjusted for age, sex and estimated glomerular filtration rate

SBP: Systolic blood pressure

DBP: Diastolic blood pressure

p-value for interaction represents result of regression models testing for difference in effect of sickle cell trait on BP in Nairobi versus Jackson

Table S11. Age specific effects of sickle cell trait on blood pressure by study site (IDACO Criteria).

Kilifi

Age, years	N	24 hour blood pressure				Night time blood pressure				Daytime blood pressure			
		SBP	(95% CI)	DBP	(95% CI)	SBP	(95% CI)	DBP	(95% CI)	SBP	(95% CI)	DBP	(95% CI)
10-29	917	-0.7	(-2.5, 1.1)	0.2	(-1.0, 1.4)	-0.2	(-2.2, 1.7)	0.4	(-0.9, 1.7)	-1.5	(-3.5, 0.6)	0	(-1.4, 1.4)
30-59	655	-3.0	(-6.3, 0.3)	-2.3	(-4.5, -0.1)	-3.8	(-7.5, -0.2)	-2.8	(-5.1, 0.5)	-1.5	(-4.8, 1.9)	-1.8	(-4.2, 0.5)
≥60	393	-0.2	(-5.3, 4.9)	-0.4	(-3.1, 3.0)	-0.8	(-6.6, 4.9)	-0.5	(-3.7, 2.7)	1.0	(-4.0, 6.1)	0.3	(-2.8, 3.6)

Nairobi and Jackson pooled together*

Age, years	N												
		SBP	(95% CI)	DBP	(95% CI)	SBP	(95% CI)	DBP	(95% CI)	SBP	(95% CI)	DBP	(95% CI)
10-29	620	-0.5	(-3.0, 1.9)	0	(-1.5, 1.5)	-1.1	(-3.6, 1.4)	-0.6	(-2.2, 1.1)	-0.9	(-3.6, 1.8)	-0.4	(-2.2, 1.5)
30-59	451	-0.4	(-4.6, 3.9)	-1.2	(-4.0, 1.6)	-0.7	(-5.5, 4.0)	-0.6	(-3.7, 2.5)	-0.2	(-4.5, 4.1)	-0.9	(-3.9, 2.1)
≥60	429	3.7	(-1.2, 8.6)	4.0	(0.9, 7.0)	2.0	(-3.5, 7.6)	3.1	(-0.2, 6.4)	5.1	(0.1, 10.0)	4.8	(1.5, 8.0)

SBP: Systolic blood pressure

DBP: Diastolic blood pressure

Results of linear regression models adjusted for age, sex and estimated glomerular filtration rate

*Participants in Jackson were aged 21 years and older

Table S12. Effect of Sick cell trait on blood pressure by sex and study site (IDACO Criteria).

Kilifi

ABPM measure	Women (N=1046)			Men (N=919)			Interaction p-value
	β	(95% CI)	p value	β	(95% CI)	p value	
24 hour SBP	-1.9	(-4.5, 0.6)	0.141	-0.8	(-3.2, 1.7)	0.537	0.533
24 hour DBP	-0.7	(-2.3, 0.9)	0.396	-0.7	(-2.3, 0.8)	0.362	0.910
Nighttime SBP	-2.0	(-4.9, 0.8)	0.160	-1.1	(-3.7, 1.6)	0.423	0.653
Nighttime DBP	-0.7	(-2.4, 1.0)	0.417	-1.1	(-2.8, 0.6)	0.200	0.825
Daytime SBP	-1.4	(-3.9, 1.2)	0.292	-0.4	(-3.0, 2.2)	0.759	0.612
Daytime DBP	-0.8	(-2.5, 0.9)	0.353	-0.4	(-2.1, 1.4)	0.679	0.620

Nairobi and Jackson

ABPM measure	Women (N=879)			Men (N=621)			Interaction p-value
	β	(95% CI)	p value	β	(95% CI)	p value	
24 hour SBP	0.1	(-2.7, 2.8)	0.962	1.0	(-2.1, 4.0)	0.525	0.272
24 hour DBP	0.7	(-1.1, 2.6)	0.429	-0.3	(-2.3, 1.8)	0.807	0.371
Nighttime SBP	-0.7	(-3.7, 2.4)	0.674	0.1	(-3.1, 3.4)	0.930	0.294
Nighttime DBP	0.5	(-1.4, 2.5)	0.604	-0.8	(-3.1, 1.4)	0.476	0.393
Daytime SBP	-0.3	(-3.1, 2.6)	0.856	1.8	(-1.4, 5.0)	0.275	0.096
Daytime DBP	0.2	(-1.8, 2.1)	0.881	0.6	(-1.7, 2.8)	0.612	0.110

Linear regression models adjusted for age, sex and estimated glomerular filtration rate

SBP: Systolic blood pressure

DBP: Diastolic blood pressure

p-value for interaction represents result of regression models testing for difference in effect of sickle cell trait on BP in men versus women

Table S13. Effects of sickle cell trait on blood pressure: effect of excluding participants taking anti-hypertensive medication (IDACO Criteria).

ABPM measure	Kilifi (N=1918)			Nairobi and Jackson (N=1041)		
	β (mm Hg)	(95% CI)	p value	β (mm Hg)	(95% CI)	p value
24 hour SBP	-1.3	(-3.1, 0.5)	0.157	-0.3	(-2.5, 1.9)	0.790
24 hour DBP	-0.7	(-1.8, 0.5)	0.254	-0.4	(-1.9, 1.1)	0.613
Nighttime SBP	-1.5	(-3.4, 0.5)	0.136	-0.6	(-2.9, 1.8)	0.639
Nighttime DBP	-0.9	(-2.0, 0.3)	0.162	-0.7	(-2.3, 0.8)	0.356
Daytime SBP	-0.9	(-2.7, 1.0)	0.348	-0.4	(-2.8, 2.0)	0.729
Daytime DBP	-0.5	(-1.8, 0.7)	0.382	-0.4	(-2.0, 1.3)	0.678

Table S14. Results of interaction analysis comparing effect of sickle cell trait in Kilifi versus Nairobi/Jackson (IDACO Criteria).

	Model 1 (N=3465)			Model 2 N=2379		
	β	(95% CI)	p value	β	(95% CI)	p value
24 hour SBP	-2.1	(-5.0, 0.7)	0.146	-3.3	(-7.0, 0.4)	0.078
24 hour DBP	-1.0	(-2.9, 0.8)	0.275	-2.3	(-4.7, 0.1)	0.057
Nighttime SBP	-1.5	(-4.6, 1.6)	0.339	-3.1	(-7.2, 0.9)	0.133
Nighttime DBP	-0.8	(-2.8, 1.1)	0.337	-2.7	(-5.3, -0.2)	0.032
Daytime SBP	-1.8	(-4.8, 1.1)	0.225	-3.1	(-6.8, 0.7)	0.112
Daytime DBP	-0.9	(-2.9, 1.1)	0.394	-2.0	(-4.6, 0.6)	0.126

Estimates were derived as the interaction term (malaria vs non-malaria sites) in a linear regression of blood pressure by sickle cell trait status after adjusting for age, sex and renal function. Renal function was represented by estimated glomerular filtration rate in model 1 and by log urine albumin to creatinine ratio in model 2. SBP: Systolic blood pressure. DBP: Diastolic blood pressure

Table S15. Effect of α^+ thalassemia on ambulatory blood pressure by study site (IDACO Criteria).

Kilifi (N=1961)

ABPM measure	No. of -3.7kb α^+ thalassemia deletions					
	One deletion (Heterozygous)			Two deletions (Homozygous)		
	β mm Hg	(95% CI)	p value	β mm Hg	(95% CI)	p value
24 hour SBP	-0.4	(-1.9, 1.2)	0.637	-1.9	(-3.9, 0.2)	0.073
24 hour DBP	-0.4	(-1.4, 0.7)	0.480	-0.7	(-2.0, 0.6)	0.286
Nighttime SBP	-0.1	(-1.9, 1.6)	0.879	-1.5	(-3.7, 0.8)	0.203
Nighttime DBP	-0.3	(-1.4, 0.8)	0.605	-0.6	(-2.0, 0.8)	0.397
Daytime SBP	-0.5	(-2.1, 1.2)	0.581	-2.1	(-4.1, 0.1)	0.059
Daytime DBP	-0.1	(-1.2, 1.0)	0.808	-0.7	(-2.1, 0.8)	0.361

Nairobi (N=771)

ABPM measure	No. of -3.7kb α^+ thalassemia deletions					
	One deletion (Heterozygous)			Two deletions (Homozygous)		
	β mm Hg	(95% CI)	p value	β mm Hg	(95% CI)	p value
24 hour SBP	0.7	(-1.3, 2.6)	0.494	1.6	(-2.0, 5.2)	0.390
24 hour DBP	0.4	(-0.6, 1.6)	0.572	1.4	(-0.9, 3.7)	0.238
Nighttime SBP	0.9	(-1.1, 3.0)	0.376	1.2	(-2.6, 4.9)	0.376
Nighttime DBP	1.0	(-0.4, 2.3)	0.164	1.0	(-1.4, 3.5)	0.420
Daytime SBP	0.1	(-3.7, 4.1)	0.941	0.2	(-3.7, 4.1)	0.923
Daytime DBP	-0.2	(-1.6, 1.3)	0.837	0.5	(-2.2, 3.1)	0.717

SBP: Systolic blood pressure

DBP: Diastolic blood pressure

Linear regression models adjusted for age, sex and estimated glomerular filtration rate.

*No alpha thalassemia data were available for participants from Jackson

Figure S1. Study locations.

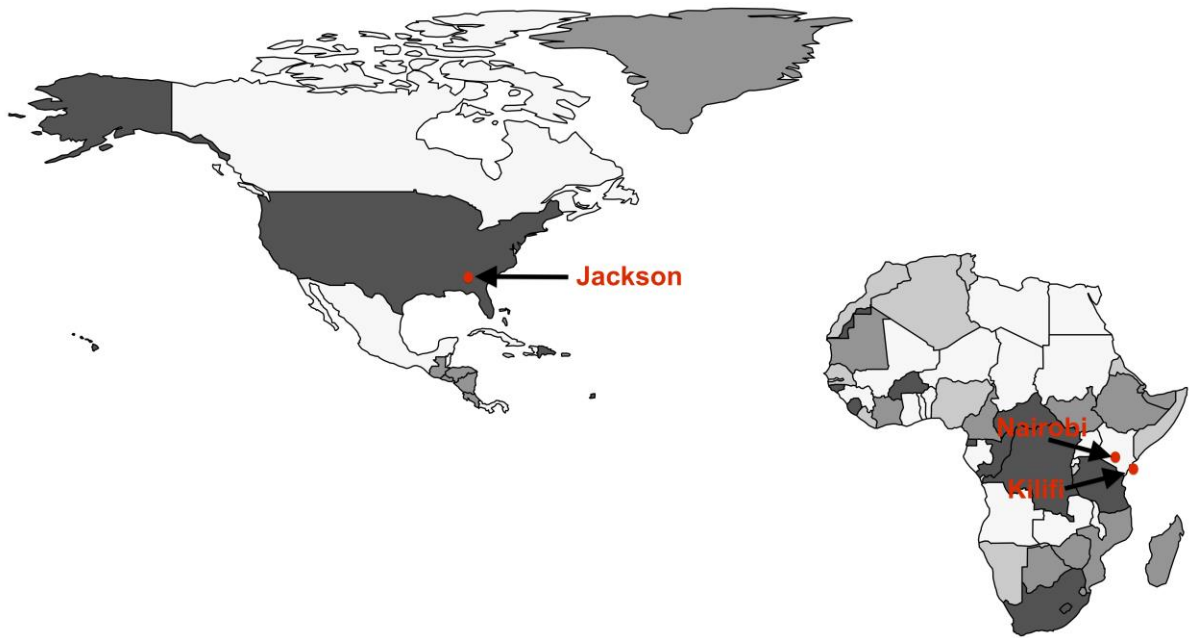


Figure S2. Causal diagram for the malaria high blood pressure hypothesis.

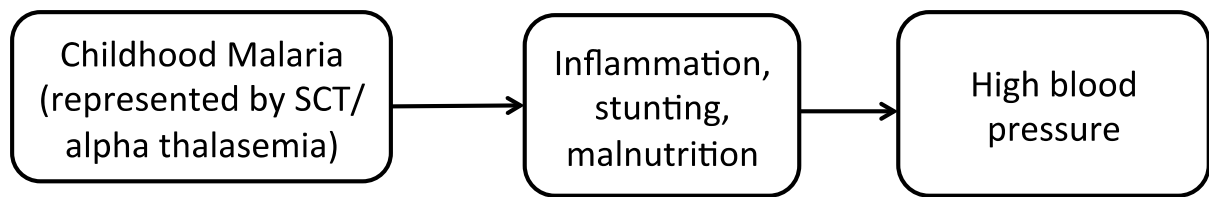


Figure S3. Illustrating confounding effect of kidney function (eGFR) in individuals with SCT.

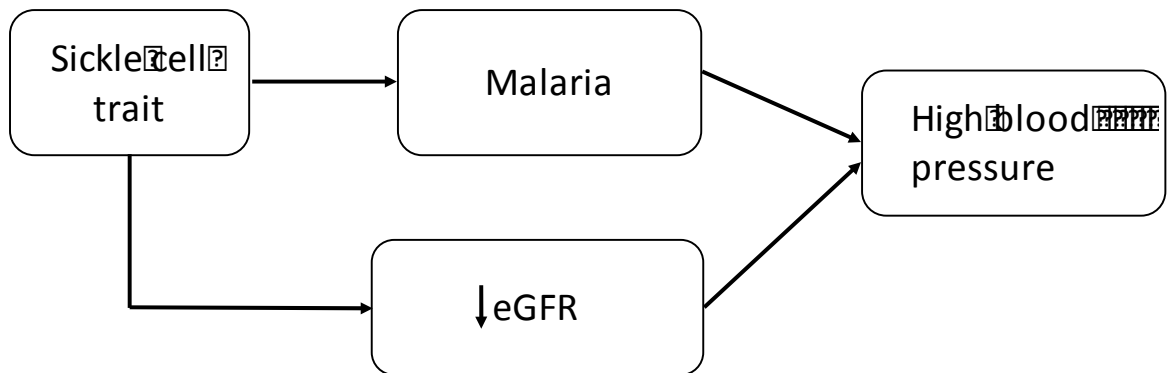
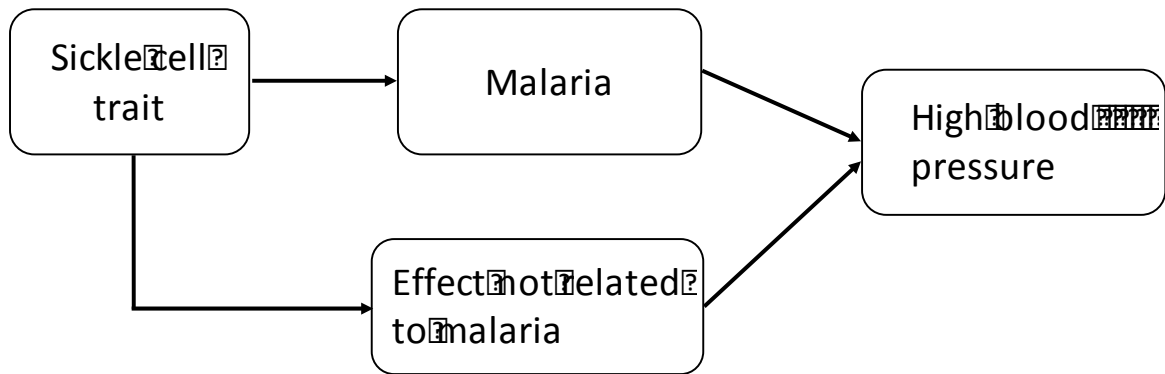


Figure S4. Illustrating confounding due to pleiotropy.



Supplemental References:

1. Beguy D, Elung'ata P, Mberu B, Oduor C, Wamukoya M, Nganyi B and Ezeh A. Health & Demographic Surveillance System Profile: The Nairobi Urban Health and Demographic Surveillance System (NUHDSS). *Int J Epidemiol*. 2015;44:462-71.
2. Scott JA, Bauni E, Moisi JC, Ojal J, Gatakaa H, Nyundo C, Molyneux CS, Kombe F, Tsofa B, Marsh K, Peshu N and Williams TN. Profile: The Kilifi Health and Demographic Surveillance System (KHDSS). *Int J Epidemiol*. 2012;41:650-7.
3. Flint J, Hill AV, Bowden DK, Oppenheimer SJ, Sill PR, Serjeantson SW, Bana-Koiri J, Bhatia K, Alpers MP, Boyce AJ and et al. High frequencies of alpha-thalassaemia are the result of natural selection by malaria. *Nature*. 1986;321:744-50.
4. Foy H, Ph D, Sc D, Kondi A, Bushra F, Hall F and Bari B-s. The variability of sickle cell rates in the tribes of Kenya and the Southern Sudan. *BMJ*. 1954;1:294-297.
5. Etyang AO, Warne B, Kapesa S, Munge K, Bauni E, Cruickshank JK, Smeeth L and Scott JA. Clinical and Epidemiological Implications of 24-Hour Ambulatory Blood Pressure Monitoring for the Diagnosis of Hypertension in Kenyan Adults: A Population-Based Study. *J Am Heart Assoc*. 2016;5:e004797-e004797.
6. Alkire S and Foster J. Understandings and misunderstandings of multidimensional poverty measurement. *The Journal of Economic Inequality*. 2011;9:289-314.
7. Nemeth Z, Moczar K and Deak G. Evaluation of the Tensioday ambulatory blood pressure monitor according to the protocols of the British Hypertension Society and the Association for the Advancement of Medical Instrumentation. *Blood Press Monit*. 2002;7:191-7.
8. Narayanan S and Appleton HD. Creatinine: a review. *Clin Chem*. 1980;26:1119-26.
9. Levey AS, Stevens LA, Schmid CH, Zhang YL, Castro AF, 3rd, Feldman HI, Kusek JW, Eggers P, Van Lente F, Greene T, Coresh J and Ckd EPI. A new equation to estimate glomerular filtration rate. *Ann Intern Med*. 2009;150:604-12.
10. Schwartz GJ, Munoz A, Schneider MF, Mak RH, Kaskel F, Warady BA and Furth SL. New equations to estimate GFR in children with CKD. *J Am Soc Nephrol*. 2009;20:629-37.
11. Taylor HA, Jr. The Jackson Heart Study: an overview. *Ethn Dis*. 2005;15:S6-1-3.
12. Wilson JG, Rotimi CN, Ekunwe L, Royal CDM, Crump ME, Wyatt SB, Steffes MW, Adeyemo A, Zhou J, Pharm D and Jr HAT. Study Design For Genetics In The Jackson Heart Study. *Ethn Dis*. 2005;15:S6-30 - S6-37.
13. Carpenter MA, Crow R, Steffes M, Rock W, Heilbraun J, Evans G, Skelton T, Jensen R and Sarpong D. Laboratory, reading center, and coordinating center data management methods in the Jackson Heart Study. *Am J Med Sci*. 2004;328:131-44.
14. Vandembroucke JP, von Elm E, Altman DG, Gøtzsche PC, Mulrow CD, Pocock SJ, Poole C, Schlesselman JJ and Egger M. Strengthening the Reporting of Observational Studies in Epidemiology (STROBE): explanation and elaboration. *PLoS Medicine*. 2007;4:e297-e297.

15. Little J, Higgins JP, Ioannidis JP, Moher D, Gagnon F, von Elm E, Khoury MJ, Cohen B, Davey-Smith G, Grimshaw J, Scheet P, Gwinn M, Williamson RE, Zou GY, Hutchings K, Johnson CY, Tait V, Wiens M, Golding J, van Duijn C, McLaughlin J, Paterson A, Wells G, Fortier I, Freedman M, Zecevic M, King R, Infante-Rivard C, Stewart A, Birkett N and Studies STtRoGA. STrengthening the REporting of Genetic Association Studies (STREGA): an extension of the STROBE statement. *PLoS Med.* 2009;6:e22.
16. Boef AG, Dekkers OM and le Cessie S. Mendelian randomization studies: a review of the approaches used and the quality of reporting. *Int J Epidemiol.* 2015;44:496-511.
17. Williams TN, Mwangi TW, Wambua S, Alexander ND, Kortok M, Snow RW and Marsh K. Sick cell trait and the risk of Plasmodium falciparum malaria and other childhood diseases. *J Infect Dis.* 2005;192:178-86.
18. Abdalla M, Caughey MC, Tanner RM, Booth JN, 3rd, Diaz KM, Anstey DE, Sims M, Ravenell J, Muntner P, Viera AJ and Shimbo D. Associations of Blood Pressure Dipping Patterns With Left Ventricular Mass and Left Ventricular Hypertrophy in Blacks: The Jackson Heart Study. *J Am Heart Assoc.* 2017;6:e004847-e004847.
19. Naik RP, Derebail VK, Grams ME, Franceschini N, Auer PL, Peloso GM, Young BA, Lettre G, Peralta CA, Katz R, Hyacinth HI, Quarells RC, Grove ML, Bick AG, Fontanillas P, Rich SS, Smith JD, Boerwinkle E, Rosamond WD, Ito K, Lanzkron S, Coresh J, Correa A, Sarto GE, Key NS, Jacobs DR, Kathiresan S, Bibbins-Domingo K, Kshirsagar AV, Wilson JG and Reiner AP. Association of sickle cell trait with chronic kidney disease and albuminuria in African Americans. *JAMA.* 2014;312:2115-25.
20. Austin PC and Steyerberg EW. The number of subjects per variable required in linear regression analyses. *J Clin Epidemiol.* 2015;68:627-36.
21. Thijs L, Hansen TW, Kikuya M, Bjorklund-Bodegard K, Li Y, Dolan E, Tikhonoff V, Seidlerova J, Kuznetsova T, Stolarz K, Bianchi M, Richart T, Casiglia E, Malyutina S, Filipovsky J, Kawecka-Jaszcz K, Nikitin Y, Ohkubo T, Sandoya E, Wang J, Torp-Pedersen C, Lind L, Ibsen H, Imai Y, Staessen JA, O'Brien E and Investigators I. The International Database of Ambulatory Blood Pressure in relation to Cardiovascular Outcome (IDACO): protocol and research perspectives. *Blood Press Monit.* 2007;12:255-62.
22. O'Brien E, Parati G, Stergiou G, Asmar R, Beilin L, Bilo G, Clement D, de la Sierra A, de Leeuw P, Dolan E, Fagard R, Graves J, Head GA, Imai Y, Kario K, Lurbe E, Mallion JM, Mancia G, Mengden T, Myers M, Ogedegbe G, Ohkubo T, Omboni S, Palatini P, Redon J, Ruilope LM, Shennan A, Staessen JA, vanMontfrans G, Verdecchia P, Waeber B, Wang J, Zanchetti A, Zhang Y and European Society of Hypertension Working Group on Blood Pressure M. European Society of Hypertension position paper on ambulatory blood pressure monitoring. *J Hypertens.* 2013;31:1731-68.
23. Lurbe E, Agabiti-Rosei E, Cruickshank JK, Dominiczak A, Erdine S, Hirth A, Invitti C, Litwin M, Mancia G, Pall D, Rascher W, Redon J, Schaefer F, Seeman T, Sinha M, Stabouli S, Webb NJ, Wuhl E and Zanchetti A. 2016 European Society of Hypertension guidelines for the management of high blood pressure in children and adolescents. *J Hypertens.* 2016;34:1887-920.
24. Cook RJ and Farewell VT. Multiplicity Considerations in the Design and Analysis of Clinical Trials. *JR Statist Soc A.* 1996;159:93.
25. Etyang AO, Smeeth L, Cruickshank JK and Scott JA. The Malaria-High Blood Pressure Hypothesis. *Circ Res.* 2016;119:36-40.

26. Davey Smith G and Ebrahim S. 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease?*. *Int J Epidemiol.* 2003;32:1-22.
27. Stamler J, Stamler R, Rhombert P, Dyer A, Berkson DM, Reedus W and Wannamaker J. Multivariate analysis of the relationship of six variables to blood pressure: findings from Chicago community surveys, 1965--1971. *J Chronic Dis.* 1975;28:499-525.
28. Palmer TM, Nordestgaard BG, Benn M, Tybjaerg-Hansen A, Davey Smith G, Lawlor DA and Timpson NJ. Association of plasma uric acid with ischaemic heart disease and blood pressure: mendelian randomisation analysis of two large cohorts. *BMJ.* 2013;347:f4262.
29. International Consortium for Blood Pressure Genome-Wide Association S, Ehret GB, Munroe PB, Rice KM, Bochud M, Johnson AD, Chasman DI, Smith AV, Tobin MD, Verwoert GC, Hwang SJ, Pihur V, Vollenweider P, O'Reilly PF, Amin N, Bragg-Gresham JL, Teumer A, Glazer NL, Launer L, Zhao JH, Aulchenko Y, Heath S, Sober S, Parsa A, Luan J, Arora P, Dehghan A, Zhang F, Lucas G, Hicks AA, Jackson AU, Peden JF, Tanaka T, Wild SH, Rudan I, Igl W, Milaneschi Y, Parker AN, Fava C, Chambers JC, Fox ER, Kumari M, Go MJ, van der Harst P, Kao WH, Sjogren M, Vinay DG, Alexander M, Tabara Y, Shaw-Hawkins S, Whincup PH, Liu Y, Shi G, Kuusisto J, Tayo B, Seielstad M, Sim X, Nguyen KD, Lehtimaki T, Matullo G, Wu Y, Gaunt TR, Onland-Moret NC, Cooper MN, Platou CG, Org E, Hardy R, Dahgam S, Palmen J, Vitart V, Braund PS, Kuznetsova T, Uiterwaal CS, Adeyemo A, Palmas W, Campbell H, Ludwig B, Tomaszewski M, Tzoulaki I, Palmer ND, consortium CA, Consortium CK, KidneyGen C, EchoGen c, consortium C-H, Aspelund T, Garcia M, Chang YP, O'Connell JR, Steinle NI, Grobbee DE, Arking DE, Kardia SL, Morrison AC, Hernandez D, Najjar S, McArdle WL, Hadley D, Brown MJ, Connell JM, Hingorani AD, Day IN, Lawlor DA, Beilby JP, Lawrence RW, Clarke R, Hopewell JC, Ongen H, Dreisbach AW, Li Y, Young JH, Bis JC, Kahonen M, Viikari J, Adair LS, Lee NR, Chen MH, Olden M, Pattaro C, Bolton JA, Kottgen A, Bergmann S, Mooser V, Chaturvedi N, Frayling TM, Islam M, Jafar TH, Erdmann J, Kulkarni SR, Bornstein SR, Grassler J, Groop L, Voight BF, Kettunen J, Howard P, Taylor A, Guarrera S, Ricceri F, Emilsson V, Plump A, Barroso I, Khaw KT, Weder AB, Hunt SC, Sun YV, Bergman RN, Collins FS, Bonnycastle LL, Scott LJ, Stringham HM, Peltonen L, Perola M, Vartiainen E, Brand SM, Staessen JA, Wang TJ, Burton PR, Soler Artigas M, Dong Y, Snieder H, Wang X, Zhu H, Lohman KK, Rudock ME, Heckbert SR, Smith NL, Wiggins KL, Doumatey A, Shriner D, Veldre G, Viigimaa M, Kinra S, Prabhakaran D, Tripathy V, Langeveld CD, Rosengren A, Thelle DS, Corsi AM, Singleton A, Forrester T, Hilton G, McKenzie CA, Salako T, Iwai N, Kita Y, Ogiwara T, Ohkubo T, Okamura T, Ueshima H, Umemura S, Eyheramendy S, Meitinger T, Wichmann HE, Cho YS, Kim HL, Lee JY, Scott J, Sehmi JS, Zhang W, Hedblad B, Nilsson P, Smith GD, Wong A, Narisu N, Stancakova A, Raffel LJ, Yao J, Kathiresan S, O'Donnell CJ, Schwartz SM, Ikram MA, Longstreth WT, Jr., Mosley TH, Seshadri S, Shrine NR, Wain LV, Morken MA, Swift AJ, Laitinen J, Prokopenko I, Zitting P, Cooper JA, Humphries SE, Danesh J, Rasheed A, Goel A, Hamsten A, Watkins H, Bakker SJ, van Gilst WH, Janipalli CS, Mani KR, Yajnik CS, Hofman A, Mattace-Raso FU, Oostra BA, Demirkan A, Isaacs A, Rivadeneira F, Lakatta EG, Orru M, Scuteri A, Ala-Korpela M, Kangas AJ, Lyytikainen LP, Soininen P, Tukiainen T, Wurtz P, Ong RT, Dorr M, Kroemer HK, Volker U, Volzke H, Galan P, Hercberg S, Lathrop M, Zelenika D, Deloukas P, Mangino M, Spector TD, Zhai G, Meschia JF, Nalls MA, Sharma P, Terzic J, Kumar MV, Denniff M, Zukowska-Szczzechowska E, Wagenknecht LE, Fowkes FG, Charchar FJ, Schwarz PE, Hayward C, Guo X, Rotimi C, Bots ML, Brand E, Samani NJ, Polasek O, Talmud PJ, Nyberg F, Kuh D, Laan M, Hveem K, Palmer LJ, van der Schouw YT, Casas JP, Mohlke KL, Vineis P, Raitakari O, Ganesh SK, Wong TY, Tai ES, Cooper RS, Laakso M, Rao DC, Harris TB, Morris RW, Dominiczak AF, Kivimaki M, Marmot MG, Miki T, Saleheen D, Chandak GR, Coresh J, Navis G, Salomaa V, Han BG, Zhu X, Kooner JS, Melander O, Ridker PM, Bandinelli S, Gyllensten UB, Wright AF, Wilson JF, Ferrucci L, Farrall M, Tuomilehto J, Pramstaller PP, Elosua R, Soranzo N, Sijbrands EJ, Althuler D, Loos RJ, Shuldiner AR, Gieger C, Meneton P, Uitterlinden AG, Wareham NJ, Gudnason V, Rotter JI, Rettig R, Uda M, Strachan DP, Witteman JC, Hartikainen AL, Beckmann JS, Boerwinkle E, Vasani RS, Boehnke M, Larson MG, Jarvelin MR, Psaty BM, Abecasis GR, Chakravarti A, Elliott P, van Duijn CM, Newton-Cheh C, Levy D, Caulfield MJ and Johnson T. Genetic variants in novel pathways influence blood pressure and cardiovascular disease risk. *Nature.* 2011;478:103-9.
30. Cohen JC, Boerwinkle E, Mosley TH, Jr. and Hobbs HH. Sequence variations in PCSK9, low LDL, and protection against coronary heart disease. *N Engl J Med.* 2006;354:1264-72.

31. Ference BA, Robinson JG, Brook RD, Catapano AL, Chapman MJ, Neff DR, Voros S, Giugliano RP, Davey Smith G, Fazio S and Sabatine MS. Variation in PCSK9 and HMGCR and Risk of Cardiovascular Disease and Diabetes. *N Engl J Med*. 2016;375:2144-2153.
32. Khan S, Floris M, Pani A and Rosner MH. Sodium and Volume Disorders in Advanced Chronic Kidney Disease. *Adv Chronic Kidney Dis*. 2016;23:240-6.
33. de Borst MH, Vervloet MG, ter Wee PM and Navis G. Cross talk between the renin-angiotensin-aldosterone system and vitamin D-FGF-23-klotho in chronic kidney disease. *J Am Soc Nephrol*. 2011;22:1603-9.
34. Neumann J, Ligtenberg G, Klein, II, Koomans HA and Blankestijn PJ. Sympathetic hyperactivity in chronic kidney disease: pathogenesis, clinical relevance, and treatment. *Kidney Int*. 2004;65:1568-76.
35. Raine AE, Bedford L, Simpson AW, Ashley CC, Brown R, Woodhead JS and Ledingham JG. Hyperparathyroidism, platelet intracellular free calcium and hypertension in chronic renal failure. *Kidney Int*. 1993;43:700-5.
36. Passauer J, Pistrosch F, Bussemaker E, Lassig G, Herbrig K and Gross P. Reduced agonist-induced endothelium-dependent vasodilation in uremia is attributable to an impairment of vascular nitric oxide. *J Am Soc Nephrol*. 2005;16:959-65.
37. Portaluppi F, Montanari L, Massari M, Di Chiara V and Capanna M. Loss of nocturnal decline of blood pressure in hypertension due to chronic renal failure. *Am J Hypertens*. 1991;4:20-6.
38. Webster AC, Nagler EV, Morton RL and Masson P. Chronic Kidney Disease. *Lancet*. 2017;389:1238-1252.
39. Lewis JB. Blood pressure control in chronic kidney disease: is less really more? *J Am Soc Nephrol*. 2010;21:1086-92.
40. Marsh K, Forster D, Waruiru C, Mwangi I, Winstanley M, Marsh V, Newton C, Winstanley P, Warn P, Peshu N and et al. Indicators of life-threatening malaria in African children. *N Engl J Med*. 1995;332:1399-404.
41. Haycock PC, Burgess S, Wade KH, Bowden J, Relton C and Davey Smith G. Best (but oft-forgotten) practices: the design, analysis, and interpretation of Mendelian randomization studies. *Am J Clin Nutr*. 2016;103:965-78.
42. Williams TN, Mwangi TW, Wambua S, Peto TEa, Weatherall DJ, Gupta S, Recker M, Penman BS, Uyoga S, Macharia A, Mwacharo JK, Snow RW and Marsh K. Negative epistasis between the malaria-protective effects of alpha+-thalassemia and the sickle cell trait. *Nat Genet*. 2005;37:1253-7.
43. O'Meara WP, Bejon P, Mwangi TW, Okiro EA, Peshu N, Snow RW, Newton CR and Marsh K. Effect of a fall in malaria transmission on morbidity and mortality in Kilifi, Kenya. *Lancet*. 2008;372:1555-62.