| 1 | Life history parameters in acellular |
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| 2 | extrinsic fiber cementum microstructure |
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27 Abstract

Life-history parameters such as pregnancies, skeletal trauma, and renal disease have previously 28 been identified from hypomineralized growth layers (incremental lines) of acellular extrinsic 29 30 fiber cementum (AEFC). The precise periodicity of these growth layers remains vaguely approximated, so causal life-history explanations using tooth cementum cannot yet be 31 rigorously calculated or tested. On the other hand, we show how life history parameters in 32 AEFC can be identified by two contrasting elemental detection methods. Based on our results 33 we reject the possibility of accurate estimation of pregnancies and other life history parameters 34 from cementum using scanning electron microscopy alone. Here, we propose a new 35 methodological approach for cementum research, Time-of-Flight Secondary Ion Mass 36 Spectrometry (ToF-SIMS), to measure degree and distribution of mineralization of cementum 37 growth layers. Our results show that Tof-SIMS can significantly increase our knowledge of 38 cementum composition and is therefore a powerful new tool for life history researchers. 39

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41 Introduction

Acellular extrinsic fiber cementum (AEFC) is deposited in a regular annual rhythm in the form 42 of incremental lines around the roots of human teeth, with varying degrees of mineralization 43 [1, 2]. Life-history parameters (LHP), such as pregnancies, skeletal trauma, and renal disease, 44 can be identified and precisely datable from incremental lines of AEFC in human teeth by 45 observing their visual effects [3]. These life history parameters appear to change calcium 46 47 metabolism [4, 5], and lack of available calcium at the mineralization front of the cementum causes formation of a such visually different incremental AEFC line [3]. In a study on humans 48 [3], as well as in great apes [6] "suspicious" AEFC lines were successfully detected as being 49

visibly broader and translucent in tooth ground sections (70 - 80µm thick) under optical 50 magnification with transmuted polarized light. These studies also showed that some of the 51 LHPs affecting mineralization of AEFC are precisely datable from the AEFC cross-sections. 52 On the other hand, in a controlled study undertaken on goats, Lieberman [2] showed that AEFC 53 bands corresponding to a control diet low in minerals including calcium and phosphorus 54 appeared to be opaquer and relatively narrower, as observed from x-ray microradiographs of 55 56 thin ground sections (50µm thick). Lieberman described these bands as hypermineralized (denser) due to reduced cementogeniesis. In contrast, a study undertaken by Cool and 57 58 colleagues [7] reported that cementum growth layers are not the result of changes in mineral density at all, as they failed to detect cementum growth layers using scanning electron 59 microscope (SEM) equipped with backscattered electrons detector (BSE). The most recent 60 61 study on composition and structure of AEFC, using Raman imaging analysis [8] argues that darker AEFC lines correspond to higher mineral/organic ratio when compared to brighter lines. 62

However, due to the relatively regular annual rhythm in their layering, AEFC incremental lines 63 are more frequently used as a chronological age estimation aid. This optical detection technique 64 65 has been in relatively frequent use as an individual age estimation aid [9 - 21], although many results are carefully qualified or subsequently disputed [22, 23]. Continued caution is required, 66 67 as cementum is the least known of all the mineralized tissues [1], and rigorously controlled human clinical studies have rarely been used to support these findings. As such, life history 68 researchers cannot entirely rely on the results of these pioneering studies yet. Furthermore, 69 70 cementum research is considerably hampered by an over-emphasis on optical microscopy as summarised by Nadji and colleagues [24]. We do not fully understand the optical appearance 71 of cementum incremental lines yet, let alone the underlying complex mineralisation 72 process(es). 73

Here we investigate if chemical composition and the degree of mineralization of AEFC can detect one important LHP, namely full-term pregnancies, from human teeth. To do so, we employ a comparative approach towards the study of AEFC incremental lines. Firstly, we compared direct measurements of degree and distribution of mineralization of AEFC from a patient with a known life history of six full term pregnancies, using two different microscopic methods, Scanning Electron Microscopy (SEM) with energy Dispersive X-Ray Analysis (EDS) and Time-of-Flight Secondary Ion Mass Spectroscopy (ToF-SIMS).

81

82 Material and methods

A mandibular canine was extracted from a white Caucasian woman undergoing necessary 83 dental intervention at the University Hospital Kosovska Mitrovica (University of Priština). The 84 informed consent to use the tooth for this research was obtained from the patient as well as her 85 anamnestic data (Table 1). The patient was born and raised in the region of Kosovska 86 Mitrovica. At the time of the extraction she was 66 years old with no previous history of renal 87 disease, endocrinal problems, skeletal fractures or trauma. The patient was considered an 88 excellent candidate for a fertility related analysis, as she reported six pregnancies that carried 89 to full-term, starting at the age of 19 with the last one at age of 31 (Table 1). After the extraction, 90 the tooth was placed in a labelled vial containing physiological saline (solution of 0.90% w/v 91 of NaCl). The tooth was free from obvious signs of pathology. 92

93

94 Table 1. The patient's anamnestic data

| Tooth extracted: 43 (left mandibular canine) | | | | | | |
|--|----------------|----|----|----|----|----|
| Sex: Female | | | | | | |
| Age of extraction: 66 years | | | | | | |
| Tooth eruption | n age: 9.6 yea | rs | | | | |
| Age at | 19 | 21 | 25 | 27 | 30 | 31 |
| pregnancies: | | | | | | |

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97 Scanning Electron Microscopy (SEM) with Energy Dispersive X-

98 Ray Analysis (EDS) for measurement of elements

The resin block with exposed mid-root surface represented the sample to be analysed by SEM-99 EDS was coated with carbon (Quorum K975x carbon coater; Quorum Technologies, UK). The 100 prepared sample was examined at 20 keV by scanning electron microscopy using a Philips 101 XL30 E-SEM (Hillsboro, OR, USA) equipped with an Oxford instruments energy dispersive 102 x-ray analysis detector using Oxford Instruments INCA software. The EDS analysis was 103 104 employed to determine whether there are mineral component compositional changes between cementum growth layers. EDS analysis was performed on the same sections employed for 105 106 SIMS imaging. Beam positioning was achieved by viewing the BSE image at 500× magnification. 107

Line scans of cementum growth layers was subject to an acquisition time of 100 sec (working
distance 10 mm, take-off angle 35°) to obtain X-ray spectra. The X-ray spectra were used to
determine which minerals were present and the Ca:P atomic percent ratio.

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112 Identification of Ca and HAp by Time of Flight–Secondary Ion

113 Mass Spectrometry Imaging

The sample preparation for ToF-SIMS imaging comprised several steps and it was performed 114 115 at University College London, Institute of Archaeology. The procedure was tailor made for this research, namely the identification of Ca and HAp form AEFC growth layers using ToF-SIMS. 116 After the cross section was cut out from the mid-root of the patient's tooth, the exposed root 117 surface mounted in the resin represented the sample to be analysed using ToF-SIMS. The 118 following preparation step was polishing, undertaken using a rotating wheel and polishing 119 media. This step is required to remove the surface damage that occurred during sectioning and 120 to provide a flat surface. The polishing procedure included the use of a series of progressively 121 finer polishing pads and diamond compounds, from $2 - \frac{1}{4} \mu$ (Kemet Diamond paste). The final 122 step was ultrasonic cleaning (1min at xx kHz) using deionized water. This step was performed 123 in order to thoroughly remove all traces of contamination tightly adhering or embedded onto 124 the sample surface. 125

Secondary ion mass spectrometry and secondary ion mapping was performed using a 126 TOF.SIMS5 mass spectrometer (ION-TOF, Münster, Germany) at Imperial College London, 127 128 Micrometre resolution was used for secondary ion mapping within m/z 0-880. The system is comprised of a bismuth primary ion beam, operating at 25 keV and tuned to use the Bi₃ ⁺ cluster 129 130 for greater secondary ion yield, and a low energy electron flood gun for charge compensation. Cluster ion sources, such as Bi₃, are used to identify larger HAp fragment ions at for example 131 m/z 485, 541, 597, and 653, identified as $Ca_5P_3O_{12}^+$, $Ca_6P_3O_{13}^+$, $Ca_7P_3O_{14}^+$, and $Ca_8P_3O_{15}^+$, 132 respectively. Ionic species sputtered from the surface under the bismuth bombardment are 133 steered into a reflectron time-of-flight mass analyzer. Before mass spectrometry was 134 performed, an Ar_n^+ cluster ion beam was used to remove any surface organic contaminants. 135

Identified peaks strongly localized to cementum growth layers were mapped on single ion
maps. Positive-ion spectra were acquired from two different 100×100 μm regions of tooth
encompassing the entire cementum width from mesio-buccal and disto-buccal side of the tooth,
respectively to localize of HAp and identification of different CaP phases within cementum
layers.

141 Study Approval

Use of human tissues and human sensitive data for this study was approved by the Ethics
Committee of Faculty of Medicine, University of Pristina at Kosovska Mitrovica, Ministry of
Health, Republic of Serbia, as well as by the Ethical Board of Research Executive Agency,
European Commission, Brussels.

146 **Results**

147 SEM-EDS

No visual evidence of cementum growth layering was found SEM micrograph using Philips 148 XL30 FEG-SEM (Hillsboro, OR, USA) equipped with an Oxford instruments energy 149 dispersive x-ray analysis detector (Fig 1a). The Ca:P ratio (by atomic percent) ranged from 150 1.47 to 1.73, with 1.59 as average value. Line scan for Ca showed no significant change in its 151 relative amounts across the width of the cementum (Fig 1b), except in the case of line spectrum 152 (8) which exhibits the lowest relative amount of Ca as of 0.13(atomic %), as well as the lowest 153 Ca:Pa ratio as of 1.29 by atomic % (Fig.1a). However, these low readings for the line spectrum 154 (8) are due to an intruding artefact deposited in cementum, which can be clearly observed on 155 electron photomicrograph of a transverse section of midroot cementum (Fig 1a), and should 156 not be taken into account when interpreting mineral distribution across the AEFC width of our 157 sample. 158

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Fig 1. Results of EDS line analysis of the cross section of the patient's tooth. The SEM photomicrograph of the cross section (a) is showing the location across mid-root acellular extrinsic fiber cementum (AEFC) and dentine (D) where the line spectrum was taken. The line charts (b) represent results of EDS line scan analyses for Ca (upper chart) and Ca/P ratio (lower chart) across the width of AEFC.

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166 **ToF-SIMS**

Layering of AEFC is somewhat visible from the ION-TOF.SIMS5 camera view (Fig 2a), but 167 not in a form of clearly defined incremental lines. Furthermore, AEFC incremental lines in our 168 sample were not clearly discernible even when observed under transmitted polarized light 169 microscope, using 400× magnification, and flowing the established Tooth Cementum 170 171 Annulation protocol [25] (see S1 Appendix and S1 Fig). However, we were able to estimate approximal width of AEFC incremental lines in our sample by using the available 172 measurements, and knowledge on teeth eruption AEFC annulation. Having the total width of 173 174 the intact cementum layer measured from the ToF-SIMS micrograph (Fig 2a) of our sample, assuming that it grows in a regular annual rhythm [1 - 3], we were able to calculate the 175 approximate width of incremental lines. As shown on Figure 2a, the analysed AEFC is flanked 176 by cemento-dentinal junction on one side and outer edge of the tooth on the other side. More 177 precisely, the total width of AEFC in our sample equals 73µm, as it spreads across the area 178 between 4 µm - 77 µm on the linescans (Fig 2b). Given the age of extraction for this tooth (66 179 years), as well as the average sex specific year of eruption of the tooth, which is 9.6 for 180 mandibular canines in females [26, 27], the estimated approximal width of AEFC incremental 181 182 lines for this individual is c. 1.3 µm.

Elemental and molecular maps, as well as line scans of Ca+ and HAp+ (Ca/P ratio) are obtained from the AEFC surface. We have detected a variation in the intensity of Ca+ across the analyzed AEFC surface (Fig 2b). A depletion in relative Ca+ intensity can be observed from $12 - 54\mu$ m respectively, but no depletion in intensity of HAp+ in the same line scan (Fig 2b, Fig 3). This corresponds to the patient's 2nd -5th decade of life (Fig 3). The lowest point in Ca+ intensity depletion is recorded at $32^{nd} \mu$ m (Fig 3) which corresponds to start of the patent's 4th decade of life (around age her age of 30).

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Fig 2. Distribution of the molecular ions identified by SIMS. (a) A ToF-SIMS micrograph
of the area of interest. "D" represents dentine; "AEFC" represents acellular extrinsic fiber
cementum; and red square is demarking the area which has been analysed by ToF-SIMS. (b)
Molecular ion mapping and line scan ion images of calcium (Ca+), and (C) hydroxyapatite
(HAp+).

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Fig. 3. Secondary ion yield (counts) of calcium (Ca+) and hydroxyapatite (HAp+) across
the width of the cementum. Right side of the plots represents outside of the tooth.

199

200 Discussion and conclusions

This study compared two different elemental analyses methods in order to establish which one best estimates the degree and distribution of mineralisation of AEFC. The results of the two methods were compared with the recorded life history parameters of a subject who had six fullterm pregnancies. We demonstrate that degree and distribution of mineralization of human

AEFC varies across the width of the resultant cementum sample. Furthermore, we show how life history parameter detection in AEFC can vary between two elemental detection methods.

207 Scanning electron microscope with electro-dispersive probe did not detect any significant variation in Ca relative amounts across the AEFC width, nor in Ca:P ratio respectively (Fig 208 1b). The range for the Ca/P ratio (by atomic percent) varied between 1.47 to 1.73, where the 209 210 majority of the values (Fig 1b) fell below 1.65, and the value 1.73 was read only once. This implies that our results for Ca/P atomic percent ratio are significantly lower than the Ca/P 211 atomic ratio bioapatite standard (1.69 - 1.71). These results suggest that the AEFC analysed 212 here is relatively hypomineralized overall. The aim of this study was to detect variation in 213 degree and distribution of mineralization across the AEFC cross-section, therefore, it is not of 214 any use to discuss the average Ca/P ratio we have obtained. In terms of previous research that 215 found no obvious variation in the concentration profile for calcium and phosphorus, our results 216 are in general agreement [7]. The authors of that study reported that calcium and phosphorus 217 218 are present in cementum at a ratio (1.70) similar to the bioapatite standard, which disagrees with our results. They concluded that "cementum growth involves a constant rate of both 219 mineralization and matrix production, rather than variations in the rate of matrix production 220 with mineralization continuing at a uniform rate" [7], which is opposite of what was reported 221 in Lieberamn's work [28] (Lieberman, 1994). Apart from stating that the cross-section was 222 taken through areas shown to contain growth layers, the authors [7] had not specified precisely 223 which type of cementum analyzed in their study. This implies that they were observing 224 cementum as a single uniform type of tissue. A similar approach was taken in a few previous 225 studies when analyzing cementum with electron probes [29, 30]. Importantly, there are various 226 types of cementum which can be distinctly classified based on presence or absence of cells, 227 nature and origin of organic matrix, or a combination of these factors [1, 31, 32]. Different 228 types of cementum are formed at different rates, but only AEFC is considered annually 229

deposited. Most importantly, incremental lines are found in different types of cementum (e.g.
in acellular and cellular cementum). Furthermore, EDS analysis can only be used to obtain the
relative amounts of concretions even when certified standards are used; as EDS operating
software does not provide us with continuous elemental values. This might be the reason we
were not able to detect any obvious variation in Ca and P profiles. With all this in mind, we
argue that the SEM-EDS technique is inadequate for measuring the degree and distribution of
AEFC mineralization.

ToF-SIMS micro-image revealed clearly defined AEFC in our cross-section. Although some 237 appearance of layering within AEFC can be also observed (Fig 2a), the incremental lines of 238 AEFC were not clearly discernible from the micrograph. On the other hand, we were able to 239 estimate the approximate width of AEFC incremental lines using the micrograph itself. The 240 linescans showed obvious variation in Ca+ intensity across the AEFC width, but not for HAp+ 241 (Fig 2b, Fig 3). Ca+ intensity variation has been detected in a form of a depletion which might 242 correlate with the with patient's pregnancies, as the depletion of Ca+ intensity corresponds to 243 the patient's $2^{nd} - 5^{th}$ life decade (Fig 3). The lowest point in Ca+ intensity depletion is recorded 244 at 32nd µm which corresponds to the beginning of the patent's 4th life decade. The initial Ca+ 245 intensity depletion was detected four years prior to the patient's age at the time of the first 246 pregnancy. The lowest value of Ca intensity is measured around the age corresponding to the 247 patient's last pregnancy. From that point on, the intensity values for Ca+ are seen to rise or 248 perhaps normalise, after final pregnancy. 249

In conclusion, our ToF-SIMS results imply that life history parameters, such as pregnancies, are more likely to influence AEFC in terms of relatively reduced mineralization, which is in accordance with work done by Kagerer and Grupe [3]. We also demonstrated how use of SEM-EDS technique is inadequate when measuring the degree and distribution of AEFC

mineralization. Our results point towards an alternative direction for life history research using 254 tooth cementum data. To detect life history parameters, which have a marked impact on Ca 255 metabolism, such as pregnancies, we propose a new methodological approach, namely Time-256 of-Flight Secondary Ion Mass Spectrometry. As a clear relationship between degree and 257 distribution of AEFC mineralization and reported pregnancies is observed by our study, this 258 technique appears to have great potential for investigating various biological events in the 259 historical development of humans and animals. Clearly, more clinical tests are required. In the 260 meantime, far more caution is required by cementum-oriented life history researchers. 261

262

263 **Supporting Information**

264 S1 Appendix. Transmitted Polarized Light Microscopy Analysis

S1 Table. Results of the reported counts of incremental lines from the tooth cross section.

267 S1 Fig. Ground cross-sections of the patient's tooth under the transmitted polarized light

268 microscope (a and b \times 400). The red arrows indicate a pronounced eruption line. The white 269 arrows indicate possible "crisis" lines which appear as broad and translucent layers in acellular

extrinsic fiber cementum (AEFC) of the patient. (D) represents dentine, and (CDJ) is cemento-

271 dentinal junction.

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273 **References**

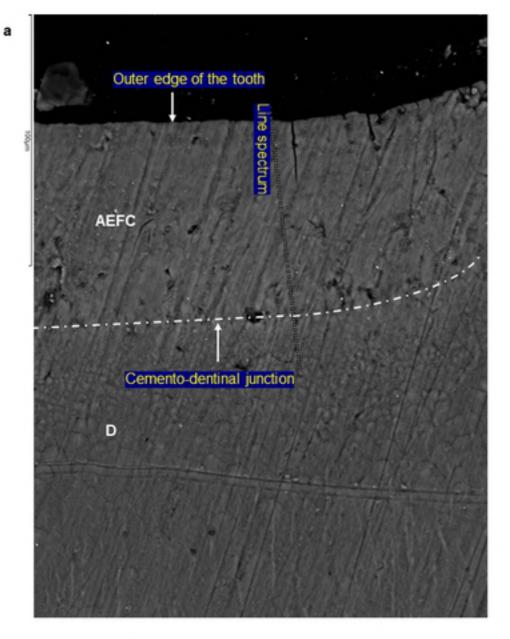
| 274 | 1. | Berkovitz BG, Holland G, Moxham B. Oral Anatomy, Histology and Embryology. 4th |
|-----|----|---|
| 275 | | ed. Elsevier Limited; 2009. |
| 276 | 2. | Lieberman DE. Life history variables preserved in dental cementum microstructure. |
| 277 | | Science. 1993; 261: 1162-1164. DOI: 10.1126/science.8356448. |
| 278 | 3. | Kagerer P, Grupe G. Age-at-death diagnosis and determination of life-history |
| 279 | | parameters by incremental lines in human dental cementum as an identification aid. |
| 280 | | Forensic Sci Int. 2001; 118: 75-82. |
| 281 | 4. | Kovacs CS, Kroneneberg HM. Maternal-fetal calcium and bone metabolism during |
| 282 | | pregnancy, puerperium, and lactation. Endocrine Reviews. 1997; 18(6): 832-872. |
| 283 | 5. | Harada S, Rodan GA, Control of osteoblast function and regulation of bone mass. |
| 284 | | Nature. 2003; 423: 349-355. |
| 285 | 6. | Cipriano A. Cold stress in captive great apes recorded in incremental lines of dental |
| 286 | | cementum. Folia Primatol. 2002; 73: 21-31. |
| 287 | 7. | Cool SM, Forwood MR, Campbell P, Bennett MB. Comparisons between bone and |
| 288 | | cementum compositions and the possible basis for their layered appearances. Bone. |
| 289 | | 2002; 30(2): 386-392. |
| 290 | 8. | Colard T, Falgayrac G, Bertrand B, Naji S, Devos O, Balsack C, et al. New Insights |
| 291 | | on the Composition and the Structure of the Acellular Extrinsic Fiber Cementum by |
| 292 | | Raman Analysis. PLoS ONE. 2016; 11(12): e0167316. |
| 293 | | doi:10.1371/journal.pone.0167316 |
| 294 | 9. | Laws RM. A new method of age determination for mammals. Nature. 1952: 972-973. |
| 295 | 10 | . Stott GG, Sis RF, Levy BM. Cemental annulation as an age criterion in forensic |
| 296 | | dentistry. J. Dent. Res. 1982; 61:814-7. PMID: 6953121 |
| 297 | 11 | . Lieberman DE, Meadow RH. The biology of cementum increments (with an |
| 298 | | archaeological application). Mammal Rev. 1992; 22(2): 57-77. |

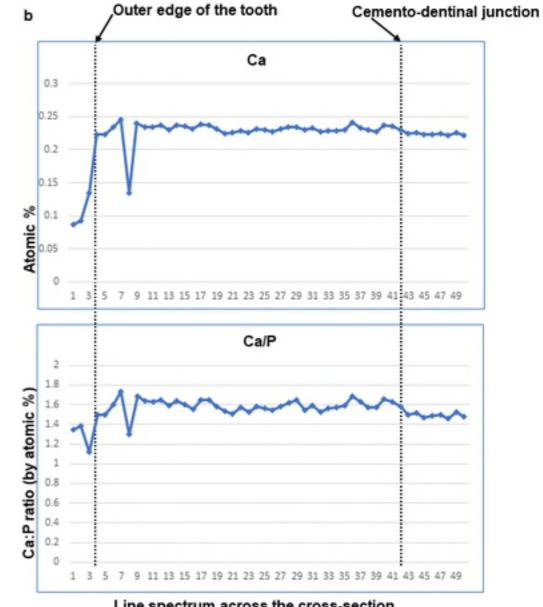
| 299 | . Wittwer-Backofen U, Buba H. Age estimation by tooth cementum annulations. In: | |
|-----|---|----|
| 300 | Hoppa RD, Vaupel JW, editors. Paleodemography: Age Distributions from Skeleta | .1 |
| 301 | Samples. Cambridge: Cambridge University Press; 2002. pp. 107-128. | |
| 302 | . Bojarun R, Garmus A, Jankauskas R. Microstructure of dental cementum and | |
| 303 | individual biological age estimation. Medicina (Kaunas). 2003; 39(10): 960-4. | |
| 304 | . Wittwer-Backofen U, Gampe J, Vaupel JW. Tooth cementum annulation for age | |
| 305 | estimation: Results from a large known-age validation study. American Journal of | |
| 306 | Physical Anthropology. 2004; 123(2): 119-129. | |
| 307 | Lippitsch A, Grupe G. Variability of the apposition of the acellular, extrinsic fiber | |
| 308 | cementum and its influence on the tooth cementum annulation technique in human | 5: |
| 309 | the influence of physical demands and functional morphology. In: Grupe G, Peters | J, |
| 310 | editors. Skeletal Series and Their Socio-Economic Context. Rahden/Westf. : M. | |
| 311 | Leidorf; 2007. pp. 87-112. | |
| 312 | . Dias PE, Beaini TL, Melani RF. Age estimation from dental cementum incrementa | 1 |
| 313 | lines and periodontal disease. J. Forensic Odontostomatol. 2010; 28: 13-21. | |
| 314 | . Kasetty S, Rammanohar M, Raju Ragavendra T. Dental cementum in age estimatic | n: |
| 315 | a polarized light and stereomicroscopic study. J. Forensic Sci. 2010; 55: 779-783. | |
| 316 | 8. Radovic M. Ageing in the Danube Gorges population (9500 – 5500 BC) – Tooth | |
| 317 | Cementum Annulation method. Starinar. 2012; 42: 9-18. doi: | |
| 318 | 10.2298/STA1262009R. | |
| 319 | 9. Schug GR, Brandt ET, Lukacs JR. Cementum annulations, age estimation, and | |
| 320 | demographic dynamics in Mid-Holocene foragers of North India. HOMO-Journal | of |
| 321 | Comparative Human Biology. 2012; 63(2): 94-109. | |

| 322 | 20. Gauthier J, Schutkowski H. Assessing the application of tooth cementum annulation |
|-----|--|
| 323 | relative to macroscopic aging techniques in an archeological sample. Homo. 2013; |
| 324 | 64:42-57. doi: 10.1016/j.jchb.2012.11.001 PMID: 23218650 |
| 325 | 21. Bertrand B, Robbins Schug G, Polet C, Naji S, Colard T. Age-at-death estimation of |
| 326 | pathological individuals: a complementary approach using teeth cementum |
| 327 | annulations Int J Paleopathol. 2016; 15:120-127. doi: 10.1016/j.ijpp.2014.04.001. |
| 328 | 22. Lipsinic FE, Paunovich E, Houston GD, Robison SF.Correlation of age and |
| 329 | incremental lines in the cementum of human teeth. J Forensic Sci. 1986; 31(3):982-9. |
| 330 | 23. Renz H, Radlanski RJ. Incremental Lines in Root Cementum of Human Teeth – A |
| 331 | Reliable Age Marker? HOMO - Journal of Comparative Human Biology. 2006; 57(1): |
| 332 | 29-50. DOI:10.1016/j.jchb.2005.09.002 |
| 333 | 24. Naji S, Colard T, Blondiaux J, Bertrand B, d'Incau E, Bocquet-Appel J. |
| 334 | Cementochronology, to cut or not to cut? International Journal of Paleopathology. |
| 335 | 2016; 15: 113-119. PMID: 29539545 DOI: 10.1016/j.ijpp.2014.05.003 |
| 336 | 25. Wittwer-Backofen U. Age Estimation Using Tooth Cementum Annulation. In: Bell |
| 337 | LS, editor. Forensic Microscopy for Skeletal Tissues: Methods and Protocols. |
| 338 | Totowa, NJ: Humana Press; 2012. p. 129-43. |
| 339 | 26. Helm S. Seidler B. Timing of permanent tooth emergence in Danish children. |
| 340 | Community Dentistry and Oral Epidemiology. 1974; 2: 122-129. doi:10.1111/j.1600- |
| 341 | 0528.1974.tb01669.x |
| 342 | 27. Nelson SJ, Ash MM. Wheeler's Dental Anatomy, Physiology, and Occlusion. 9th ed. |
| 343 | St. Louis, Mo. : Saunders Elsevier; 2009. |
| 344 | 28. Lieberman DE. The biological basis of seasonal increments in dental cementum and |
| 345 | their application to archaeological research. J Archaeol Sci. 1994; 21:525-539. |

| 346 | 29. Hals E, Selvig KA. Correlated electron probe microanalysis and microradiography of |
|-----|--|
| 347 | carious and normal dental cementum. Caries Res. 1997; 11:62-75. |
| 348 | 30. Selvig KA, Hals E. Periodontally diseased cementum studied by correlated |
| 349 | microradiography, electron probe analysis and electron microscopy. J Periodont Res. |
| 350 | 1977; 12:419-429. PMID: 145479 |
| 351 | 31. Yamamoto H, Niimi T, Yokota-Ohta R, Suzuki K, Sakae T, Kozawa Y. Diversity of |
| 352 | Acellular and Cellular Cementum Distribution in Human Permanent Teeth. Journal of |
| 353 | Hard Tissue Biology. 2009; 18(1): 40-44. |
| 354 | 32. Ho SP, Marshall SJ, Ryder MI, Marshall GW. The tooth attachment mechanism |
| 355 | defined by structure, chemical composition and mechanical properties of collagen |
| 356 | fibers in the periodontium. Biomaterials, 2007; 28(35):5238-5245. doi: |

357 10.1016/j.biomaterials.2007.08.031 PMID: 17870156

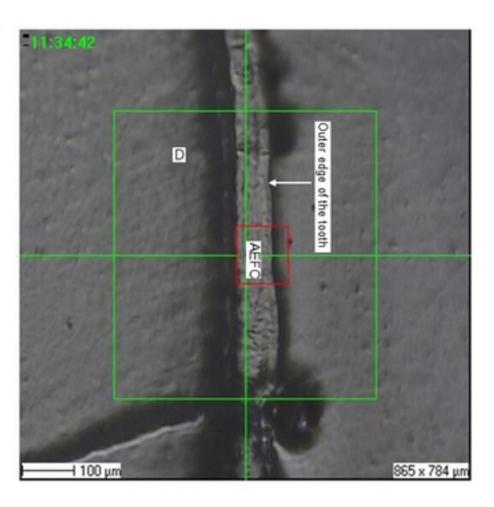




Line spectrum across the cross-section

Figure 1





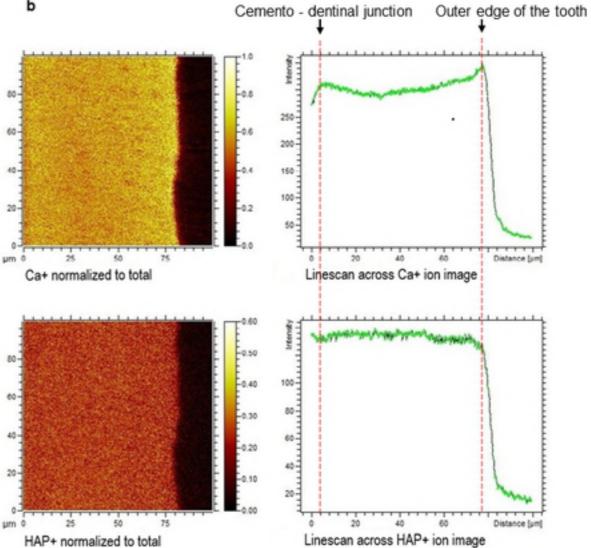


Figure 2

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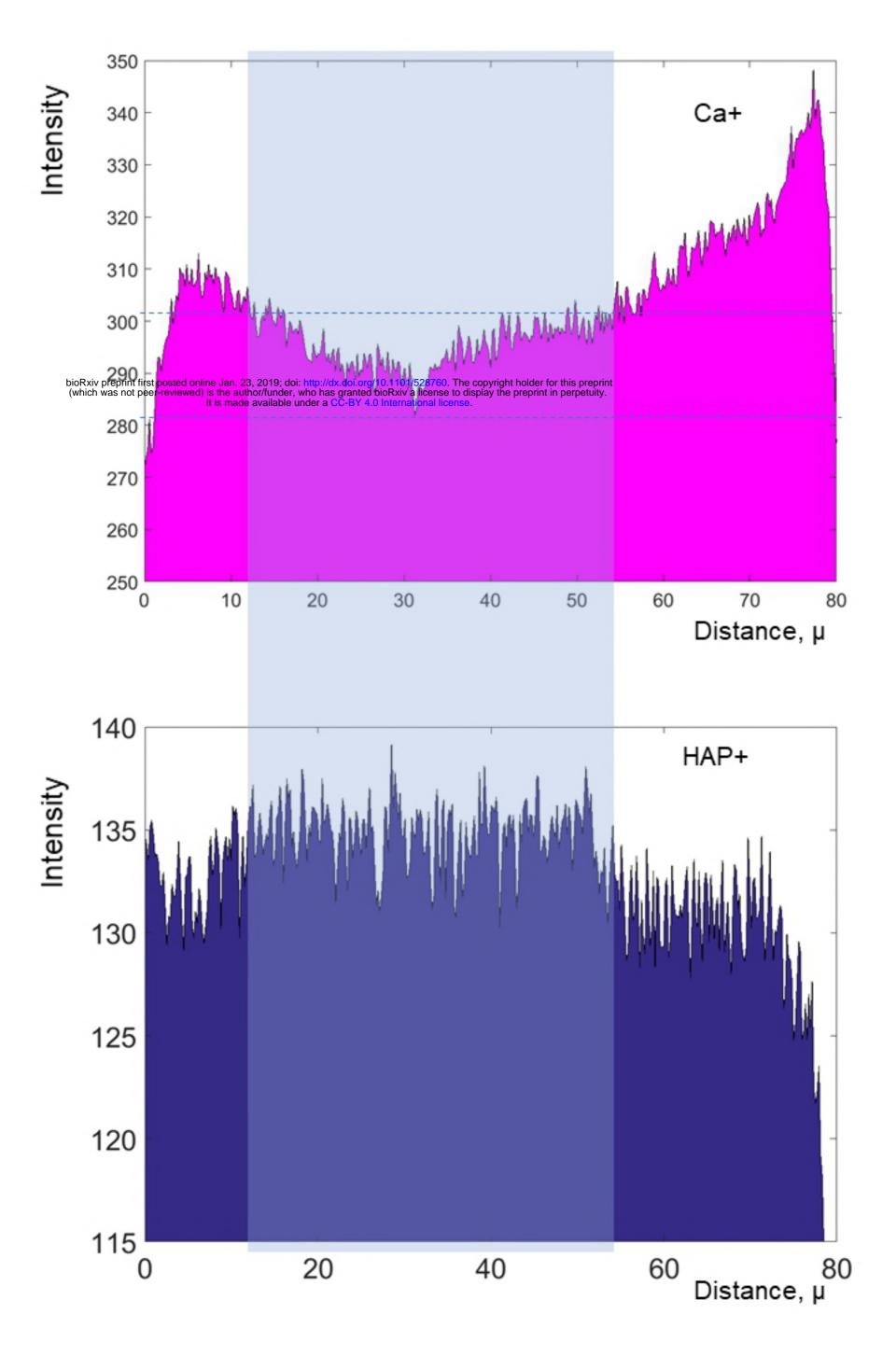


Figure 3