## **Treatments targeting inotropy**

## A position paper of the Committees on Translational Research and Acute Heart Failure of the Heart Failure Association (HFA) of the European Society of Cardiology (ESC)

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## Abstract

Acute heart failure and in particular, cardiogenic shock are associated with high morbidity and mortality. A therapeutic dilemma is that the use of positive inotropic agents, such as catecholamines or phosphodiesterase-inhibitors, is associated with increased mortality. Newer drugs, such as levosimendan or omecamtiv mecarbil, target sarcomeres to improve systolic function putatively without elevating intracellular Ca<sup>2+</sup>. Although meta-analyses of smaller trials suggested that levosimendan is associated with a better outcome than dobutamine, larger comparative trials failed to confirm this observation. For omecamtiv mecarbil, phase II clinical trials suggest a favorable hemodynamic profile in patients with acute and chronic heart failure, and a phase III morbidity/mortality trial in patients with chronic heart failure has recently begun. Here, we review the pathophysiological basis of systolic dysfunction in patients with heart failure and the mechanisms through which different inotropic agents improve cardiac function. Since adenosine triphosphate and reactive oxygen species production in mitochondria are intimately linked to the processes of excitationcontraction coupling, we also discuss the impact of inotropic agents on mitochondrial bioenergetics and redox regulation. Therefore, this position paper should help identify novel targets for treatments that could not only safely improve systolic and diastolic function acutely, but potentially also myocardial structure and function over a longer term.

#### **Key words**

Heart failure, acute decompensated heart failure, inotropes, cardiogenic shock, excitationcontraction coupling, calcium, sarcomeres, mitochondria, energetics, adrenergic receptors, contractility, levosimendan, omecamtiv mecarbil, nitroxyl.

## Introduction

Heart failure (HF) is a leading cause for hospital admissions in developed countries worldwide,<sup>1</sup> and its incidence is further increasing as average life expectancy rises. While the prognosis of patients with *chronic* HF has improved over the last decades, the prognosis of *acute* HF is still poor. Although only a small percentage of patients with acute HF present with cardiogenic shock (~3%) and/or signs of hypoperfusion (~15%), their inhospital mortality is much higher than of patients with acute HF without these severe conditions.<sup>2</sup> An important treatment option in these patients is inotropic agents to acutely increase cardiac output.<sup>3</sup> However, since the use of most inotropic agents (in particular, catecholamines and PDE-inhibitors) is complicated by adverse short- and long-term effects, their use is not recommended in the absence of hypotension or hypoperfusion.<sup>3</sup> On the other hand, the oldest inotropic agent (digitalis) can be safely applied to patients with chronic HF and improves morbidity.<sup>4</sup> [...] Here, we discuss the detailed mechanisms of inotropic agents to estimate which mechanisms of action may provide benefit in either acute or chronic HF.

In patients with HF with reduced ejection fraction (HFrEF), the inability of the heart to eject sufficient blood for the needs of peripheral tissues is caused by defects of excitation-contraction (EC) coupling in cardiac myocytes (**Figure 1**).<sup>5</sup> Traditionally, an acute increase in cardiac output is achieved with drugs that increase intracellular cyclic adenosine monophosphate (cAMP), such as catecholamines or phosphodiesterase (PDE) inhibitors (**Figure 2**). However, these agents increase myocardial oxygen (O<sub>2</sub>) consumption, predispose to life-threatening arrhythmias<sup>6</sup> and activate signaling pathways of hypertrophy and cell death,<sup>7, 8</sup> which may explain why they are associated with adverse outcome.<sup>9</sup> Consequently, recent developments aimed at increasing contractility *without* increasing cAMP or Ca<sup>2+</sup> through Ca<sup>2+</sup>-sensitizer levosimendan suggest an overall benefit,<sup>10, 11</sup> larger comparative trials failed to show a survival benefit despite hemodynamic improvements.<sup>12-15</sup> The myosin activator omecamtiv mecarbil has passed phase II trials,<sup>16-20</sup> and a phase III trial was recently launched in patients with chronic HF (GALACTIC-HF; NCT0292932).

Considering the poor outcome of patients in cardiogenic shock and/or signs of hypoperfusion<sup>2</sup> and the mostly disappointing results of inotropic agents in patients with acute HF, novel strategies are urgently needed. To this end, the Committees on Translational Research and on Acute Heart Failure of the Heart Failure Association (HFA) of the European Society of Cardiology (ESC) developed this position paper that addresses the following **three key issues**:

- 1) Why have classical inotropic agents failed?
- 2) Is direct targeting of sarcomere function therapeutically advantageous?
- 3) Which novel concepts are promising?

To understand the mechanisms of traditional, current and future medicines, it is essential to understand the physiology of EC coupling and its pathological alterations in HF. We will also discuss the bioenergetic consequences of inotropic interventions, since these may impact the long-term prognosis of HF patients. Finally, we will give recommendations for basic and clinical research directed at developing novel strategies for inotropic interventions in acute HF.

## Physiology of excitation-contraction coupling

During each action potential, Ca<sup>2+</sup> enters cardiac myocytes via L-type Ca<sup>2+</sup> channels (LTCC), triggering even greater Ca<sup>2+</sup> release from the Ca<sup>2+</sup> stores of the cell, i.e., the sarcoplasmic reticulum (SR; **Figure 1**). This Ca<sup>2+</sup> binds to troponin C, facilitating actinmyosin interaction that induces the contraction of the heart muscle. During diastole, Ca<sup>2+</sup> diffuses away from troponin C, initiating relaxation. The Ca<sup>2+</sup> that was released from the SR is taken back up by the SR Ca<sup>2+</sup> ATPase (SERCA), whereas the amount of Ca<sup>2+</sup> that entered the cell via LTCCs is exported by the Na<sup>+</sup>/Ca<sup>2+</sup> exchanger (NCX).<sup>4</sup>

Cardiac contractility is increased by three principle mechanisms:

- 1)  $\beta$ -Adrenergic stimulation,
- 2) the Frank-Starling mechanism and
- 3) the positive force-frequency relation (also known as "Bowditch-Treppe").

Noradrenaline (NA) is released from sympathetic nerve endings in the myocardium and stimulates  $\beta_1$ -adrenergic receptors ( $\beta_1$ -ARs), which couple to the stimulatory G-protein (G<sub>s</sub>) and activate the adenylyl cyclase (AC) to produce cAMP (Figures 1 and 2). Cyclic AMP activates protein kinase A (PKA) which (through phosphorylation) activates LTCCs, accelerates SERCA, increases SR Ca<sup>2+</sup> release via ryanodine receptors (RyRs) and decreases myofilament Ca<sup>2+</sup> affinity. Furthermore, cAMP activates the *exchange protein directly activated by cAMP* (Epac), further activating Ca<sup>2+</sup>/Calmodulin-dependent protein kinase II (CaMKII) which phosphorylates various Na<sup>+</sup>- and Ca<sup>2+</sup>-transporting proteins, mostly synergistic with PKA-mediated actions (Figure 2).<sup>21</sup> The net result is an increase and acceleration of force generation and relaxation (positive *inotropic* and *lusitropic* effects),

maintaining the refilling of the ventricles at elevated heart rates (positive *chronotropic* effect). In the human heart, the breakdown of cAMP is governed primarily by PDE3 and to a lesser extent by PDE4.<sup>22</sup>

Besides  $\beta$ -AR stimulation, the "Bowditch Treppe" increases cardiac contractility at elevated heart rates. This is explained by slower Ca<sup>2+</sup>-efflux (via the NCX) than -influx kinetics (via LTCCs), which results in intracellular accumulation of Ca<sup>2+</sup> at higher heart rates that is sequestered into the SR by SERCA, from where it is released in greater amounts on the ensuing beat. The Frank Starling mechanism is caused by a length-dependent increase of the Ca<sup>2+</sup> affinity of the myofilaments, resulting in stronger contraction at unchanged cytosolic Ca<sup>2+</sup> concentrations ([Ca<sup>2+</sup>]<sub>c</sub>).

# Pathophysiological changes of excitation-contraction coupling in heart failure

The central deficit of EC coupling in myocytes from failing hearts is a decreased Ca<sup>2+</sup> load of the SR. This is primarily the result of reduced SERCA expression and activity and a Ca<sup>2+</sup> leak from the SR via RyRs, reducing systolic SR Ca<sup>2+</sup> release and thereby the activator Ca<sup>2+</sup> at the myofilaments (Figure 1).<sup>5</sup> The reduced rate of SERCA-mediated Ca<sup>2+</sup> re-uptake into the SR also slows relaxation. Furthermore, the cytosolic  $Na^+$  concentration ( $[Na^+]_i$ ) is elevated in failing cardiac myocytes through changes in the "late Na<sup>+</sup> current" (late  $I_{Na}$ ). Na<sup>+</sup>/H<sup>+</sup> exchanger (NHE) and Na<sup>+</sup>/K<sup>+</sup>-ATPase (NKA) activities.<sup>23</sup> While this facilitates Ca<sup>2+</sup>influx via the reverse-mode of the NCX during the action potential,<sup>5</sup> partly compensating for decreased systolic SR Ca<sup>2+</sup> release,<sup>24</sup> relaxation is further slowed by hampering diastolic Ca<sup>2+</sup> extrusion via the forward-mode NCX. This is particularly problematic at higher heart rates, when diastole progressively shortens. Consequently, the normally positive forcefrequency relationship is blunted or even negative in failing human hearts, mediated by a variable combination of elevated diastolic [Ca<sup>2+</sup>]<sub>c</sub> and tension as well as decreased SR Ca<sup>2+</sup> load and release.<sup>25, 26</sup> This decreases left ventricular ejection fraction (LVEF) and cardiac output at higher heart rates in vivo.<sup>27</sup> Finally, elevated diastolic [Ca<sup>2+</sup>]<sub>c</sub> and increased open probability of RyRs increases the probability of spontaneous SR Ca<sup>2+</sup> release events, which (by subsequent Ca2+ extrusion via the electrogenic NCX) can induce delayed afterdepolarizations, a well-defined trigger of ventricular arrhythmias.

In addition to the defects in ion handling and the ensuing inversion of the force-frequency relationship, continuous stimulation of cardiac  $\beta_1$ -ARs through activation of the sympathetic

nervous system desensitizes and downregulates  $\beta_1$ -ARs (Figure 2),<sup>7, 8</sup> blunting their response to endogenous or exogenous catecholamines. Consequently, phosphorylation of several Ca<sup>2+</sup> handling proteins is reduced. In failing hearts, the Ca<sup>2+</sup> affinity of the myofilaments is increased as a result of decreased PKA-mediated phosphorylation of troponin I,<sup>28, 29</sup> although this issue is not fully settled yet.<sup>30</sup>

## **Energetic aspects**

EC coupling requires high amounts of energy in the form of ATP, which is replenished by oxidative phosphorylation in mitochondria. During *β*-adrenergic stimulation, mitochondria take up Ca<sup>2+</sup> to stimulate the Krebs cycle, which produces NADH as the main electron donor for ATP production at the respiratory chain (Figure 3).<sup>31</sup> Mitochondrial function is impaired in HF, resulting in energetic deficit and oxidative stress (Figure 3).<sup>31-33</sup> Mitochondrial dysfunction is linked to defects in EC coupling, since the Krebs cycle requires stimulation of its key enzymes by Ca<sup>2+</sup>, and decreased SR Ca<sup>2+</sup> release hampers mitochondrial Ca<sup>2+</sup> uptake via the uniporter (MCU). In addition, elevated [Na<sup>+</sup>], accelerates mitochondrial  $Ca^{2+}$  efflux via the mitochondrial  $Na^+/Ca^{2+}$  exchanger (NCLX; Figure 3). Impaired Krebs cycle activity limits NADH-dependent ATP production at the respiratory chain and provokes excess emission of reactive oxygen species (ROS) through depletion of the NADPH-dependent anti-oxidative capacity, causing oxidative stress.<sup>31</sup> Reduced ATP production can limit the contractile reserve of the LV,<sup>34</sup> and most Na<sup>+</sup> and Ca<sup>2+</sup> transporting mechanisms are sensitive to redox-dependent modifications.<sup>35</sup> Thus, the tight interplay between EC coupling and mitochondrial energetics (Figure 3) can set in motion a vicious cycle of deteriorated ion handling, energetic deficit and oxidative stress to aggravate systolic and diastolic dysfunction in HF.

In patients with HF, iron deficiency (ID) predicts adverse outcome,<sup>36</sup> while iron supplementation improves functional capacity and quality of life.<sup>37</sup> In failing hearts, myocardial iron content is reduced and associated with reduced activity of Krebs cycle dehydorgenases and expression of antioxidative enzymes.<sup>38</sup> In preclinical models, severe cardiac or skeletal muscle ID perturbs mitochondrial function and induces systemic metabolic derangements and cardiomyopathy.<sup>39, 40</sup> Accordingly, ID may further aggravate energy supply and demand mismatch and oxidative stress in HF. However, since plasma ID does not directly correlate with myocardial ID, these issues require further investigation (for more in-depth discussion see Ref.<sup>41</sup>).

## "Classical" inotropic agents

## Digitalis

Digitalis-derived cardiotropic glycosides (CTG) are the oldest inotropic drugs and increase [Na<sup>+</sup>]<sub>i</sub> in cardiomyocytes by inhibiting Na<sup>+</sup> export via the NKA (**Figure 4**). [Na<sup>+</sup>]<sub>i</sub> accumulation hampers diastolic Ca<sup>2+</sup> extrusion via the NCX and supports Ca<sup>2+</sup> influx via the reverse mode NCX during systole, thereby increasing diastolic [Ca<sup>2+</sup>], Ca<sup>2+</sup> transient amplitudes and consequently, inotropy (**Figure 4**). On the other hand, elevated [Na<sup>+</sup>]<sub>i</sub> accelerates mitochondrial Ca<sup>2+</sup> efflux via the NCLX, reducing Ca<sup>2+</sup>-activation of the Krebs cycle and its regeneration of NADH and NADPH. Since NADPH is required for anti-oxidative enzymes to detoxify ROS (**Figure 3**), CTG-induced NADPH oxidation increases mitochondrial ROS emission and thereby arrhythmias (**Figure 4**).<sup>42</sup> The pro-arrhythmic actions of CTG narrow its therapeutic range.<sup>43</sup>

In patients with HFrEF, digoxin may be considered in symptomatic patients in sinus rhythm despite treatment with an ACE-inhibitor (or angiotensin receptor blocker), a β-blocker and a mineralocorticoid antagonist to reduce the risk of hospitalizations (class IIb, B<sup>3</sup>). However, the Digitalis Investigation Group (DIG) Trial<sup>4</sup> was conducted before β-blockers became a mainstay of HF therapy, and digoxin did not improve all-cause mortality, but reduced hospitalization due to HF worsening. At the lower dosages used in current practice, digitalis may be preferentially a modulator of autonomic tone and less inotropic stimulator.<sup>44</sup> The ongoing DIGIT-HF study prospectively investigates the role of digitoxin in patients with HFrEF already on current guideline-adherent therapy (http://digit-hf.de). In atrial fibrillation, clinical evidence is limited to small and observational studies, and there is an ongoing debate on potentially increased mortality.<sup>45</sup> However, prescription bias in the retrospective analyses likely mimics the perceived digoxin driven mortality.<sup>46</sup> In the current HF Guidelines, an intravenous bolus of digoxin should be considered in digoxin-naïve-patients with New York Heart Association (NYHA) class IV patients and rapid ventricular rate to slow heart rate (Class IIa, B).<sup>3</sup>

### Catecholamines

The three endogenous catecholamines dopamine, adrenaline and noradrenaline stimulate  $\beta_1$ -ARs, and to variable degrees also  $\alpha_1$ - and  $\beta_2$ -ARs and dopaminergic D<sub>1</sub>- and D<sub>2</sub>-receptors (**Table 1**).<sup>6</sup> Thus, their hemodynamic actions are not limited to increasing cardiac contractility via  $\beta_1$ -ARs (**Figure 2**). In fact,  $\alpha_1$ -AR-mediated vasoconstriction by adrenaline and noradrenaline (**Table 1**) renders them useful in patients with septic shock where vasodilation underlies hypotension. The hemodynamic profile of dopamine, the immediate

precursor to noradrenaline in the synthetic pathway, is dominated by vasodilation at low concentrations at which binding to dopaminergic  $D_1$ - and  $D_2$ -receptors increases blood flow to the heart, brain, kidney and various other organs. Its traditional use to increase renal blood flow, however, was discouraged after neutral effects in trials on patients with acute decompensated heart failure, with or without renal dysfunction.<sup>47-49</sup> At higher doses, dopamine increases blood pressure by vasoconstriction via  $\alpha_1$ -ARs and positive inotropic and chronotropic effects via  $\beta$ -ARs, respectively.

Since in patients with cardiogenic shock, neuroendocrine activation induces vasoconstriction and tachycardia, the ideal drug should be positive inotropic without further increasing systemic vascular resistance (SVR). Dobutamine fulfils these requirements.<sup>50</sup> It is a full agonist at  $\beta_1$ -ARs, inducing a positive inotropic effect with similar efficacy as isoproterenol, a synthetic  $\beta_1$ - and  $\beta_2$ -AR agonist without any  $\alpha$ -AR agonism (Table 1). The affinity of dobutamine for  $\beta_2$ -AR is ~10-fold lower than for  $\beta_1$ -ARs and in particular, its agonist efficacy at  $\beta_2$ -ARs and  $\alpha_1$ -ARs much weaker than at  $\beta_1$ -ARs.<sup>51</sup> Through its dominating inotropic effect and mutually offsetting vascular effects of  $\alpha_1$ - and  $\beta_2$ -AR agonism, the decrease of SVR at intermediate and higher doses is mediated by reflex withdrawal of the endogenous sympathetic tone.<sup>52</sup> For any given increase in cardiac contractility, the increase of heart rate and blood pressure is lower with dobutamine than with dopamine or noradrenaline, further reflecting dobutamine's selectivity for  $\beta_1$ - over  $\beta_2$ - and  $\alpha_1$ -ARs.<sup>50</sup> However, this favorable hemodynamic profile comes at the cost of elevated myocardial oxygen (O<sub>2</sub>) consumption<sup>50, 52</sup> and arrhythmias. Noradrenaline may be considered in patients who have cardiogenic shock, despite treatment with another inotrope to increase blood pressure and vital organ perfusion.<sup>3</sup> The combination of adrenaline with dobutamine, however, appears to portend a particular risk for adverse outcome.53

In patients with HF, elevated plasma noradrenaline levels predict adverse outcome.<sup>54</sup> Chronic  $\beta$ -AR stimulation desensitizes and downregulates  $\beta$ -ARs via PKA, GRK2 (also known as  $\beta$ -ARK1) and  $\beta$ -arrestin.<sup>7, 8, 55</sup> Furthermore,  $\beta$ -arrestin activates CaMKII which sustains contractility despite desensitization of  $\beta$ -ARs from PKA-mediated inotropy (**Figure** 2).<sup>21</sup> CaMKII activation, however, is a major driver of cardiac arrhythmias.<sup>56</sup> The net functional consequence of all these processes is a decreased responsiveness of the human failing heart to  $\beta$ -AR stimulation. Therefore, despite its favourable short-term hemodynamic profile that improves symptoms,<sup>9</sup> dobutamine treatment is associated with tolerance,<sup>57</sup> arrhythmias and mortality, respectively.<sup>58, 59</sup>

#### **PDE Inhibitors**

To overcome desensitization and downregulation of cardiac  $\beta$ -ARs as well as their blockade through  $\beta$ -blockers, PDE-inhibitors were developed. In human failing myocardium, inhibition of PDE3, but not PDE4 potentiates  $\beta$ -AR mediated positive inotropic effects.<sup>60</sup> Conversely, in human atrial myocardium, inhibition of PDE4 potentiates arrhythmias induced by both  $\beta_1$ - and  $\beta_2$ -AR stimulation, while PDE3 inhibition only potentiates  $\beta_1$ -AR-induced arrhythmias.<sup>61</sup> PDE-inhibitors also decrease SVR through cAMP-mediated vasodilation, which is beneficial in patients with acute HF and high SVR, but limits its application in patients with cardiogenic shock due to reductions in blood pressure. In patients with HF treated with  $\beta$ -blockers, the efficacy and potency of PDE-inhibitors is maintained, while the effects of dobutamine are blunted.<sup>62</sup>

#### Why have classical inotropes failed?

Currently, dobutamine, dopamine and PDE-inhibitors are recommended in patients with hypotension (SBP <90 mmHg) and/or signs/symptoms of hypoperfusion despite adequate filling status to increase cardiac output and blood pressure and to improve peripheral perfusion and maintain end-organ function (IIb, C).<sup>3</sup> However, while in analyses that evaluated cAMP-dependent inotropes in general, their short term use yielded neutral effects, longer term use was associated with adverse outcome in patients hospitalized for acute HF despite improved quality of life.<sup>58, 63, 64</sup> Therefore, inotropic agents are explicitly restricted to patients that fulfil the above mentioned criteria and are not recommended for any other patients (class IIIA).<sup>3</sup> The adverse long-term effects may be related to PKA- and CaMKII-induced maladaptive cardiac remodeling through inducing hypertrophy, apoptosis and fibrosis (Figure 2).<sup>7,8</sup> In particular, activation of  $\beta_1$ -ARs induces apoptosis.<sup>65</sup> which is an important mechanism for LV remodelling and dysfunction in HF.<sup>66, 67</sup> Furthermore, βadrenergic activation alters myocardial substrate utilization and thereby, may trigger energetic deficit and oxidative stress.<sup>59</sup> These data indicate that despite their favourable acute hemodynamic profile, the use of adrenergic agonists and PDE-inhibitors leads to adverse outcome by triggering maladaptive cardiac remodelling and arrhythmias, while vice versa, antagonizing  $\beta_1$ -ARs can reverse remodelling, improve LV function and prolong survival.<sup>7</sup> Therefore, a new generation of inotropes had to be developed to avoid activation of adrenergic pathways and increase contractility without raising Ca<sup>2+</sup>, which is proarrhythmic and causes higher energy consumption through activation of Ca<sup>2+</sup> transporting systems.

## **Treatments targeting sarcomeres**

## Ca<sup>2+</sup> sensitizers

 $Ca^{2+}$  sensitizers shift the relationship between  $[Ca^{2+}]_c$  and force development of sarcomeres (i.e., the pCa-force relationship) to the left, increasing force at any given  $[Ca^{2+}]_c$ . They were developed in the early 1980s as cardiotonic agents with a number of theoretical advantages over catecholamines and PDE-inhibitors:

- (i) Ca<sup>2+</sup> sensitizers should neither increase trans-sarcolemmal influx of Ca<sup>2+</sup> nor alter SR Ca<sup>2+</sup> fluxes during systole or diastole. This should be less proarrhythmic and less energy consuming.
- (ii) Ca<sup>2+</sup> sensitizers should not affect heart rate or blood pressure if a compound was selective for cardiac myofilaments, which may have positive effects on energetics.
- (iii)  $Ca^{2+}$  sensitizers should be independent of the desensitized  $\beta$ -AR/cAMP system in HF and should themselves not induce tolerance.

Examples of this class of drugs are EMD-57033, CGP-48506, pimobendan and levosimendan. The mechanisms of Ca<sup>2+</sup> sensitization differ between Ca<sup>2+</sup> sensitizers. While levosimendan and pimobendan increase the affinity of troponin C to bind Ca<sup>2+</sup>,<sup>68-70</sup> CGP-48506 acts downstream of troponin C, and EMD 57033 affects the actin-myosin interaction by direct binding to the myosin motor domain.<sup>71, 72</sup> Independent of the mechanism, the shift of the pCa<sup>2+</sup>-force curve to the left increases systolic force generation for any given [Ca<sup>2+</sup>]<sub>c</sub>, but on the other hand impedes relaxation following the decrease in [Ca<sup>2+</sup>]<sub>c</sub>. The slowing of diastolic relaxation is an inherent property of pure Ca<sup>2+</sup> sensitizers and may be the reason why most pharmaceutical companies stopped their development. In this context, mutations in sarcomeric proteins that cause hypertrophic cardiomyopathy (HCM) commonly increase myofilament Ca<sup>2+</sup> sensitivity as a unifying disease mechanism.<sup>73, 74</sup> Moreover, both HCM mutations and drugs that increase Ca<sup>2+</sup> sensitivity are arrhythmogenic, presumably by providing a sink for Ca<sup>2+</sup> that is released during diastole, causing depolarisations via the electrogenic NCX.<sup>75, 76</sup>

### Levosimendan

In contrast to CGP-48506, levosimendan does not prolong relaxation time or compromise diastolic relaxation. Thus, it does not have the same profile as pure  $Ca^{2+}$  sensitizers. The most likely reason is that levosimendan is not only a  $Ca^{2+}$  sensitizer binding to troponin C,<sup>69</sup> but also a potent and selective PDE3-inhibitor with an IC<sub>50</sub> in the nanomolar range, but

~1000-fold lower affinity for PDE4.<sup>77, 78</sup> Accordingly, levosimendan increases cAMP with similar potency as it increases force.<sup>79</sup> Furthermore, its positive inotropic effect is abolished by the muscarinic receptor agonist carbachol, which acts through inhibiting cAMP generation. Also in human myocardium, the inotropic effects of levosimendan require  $\beta$ -adrenergic pre-stimulation and/or elevations of  $[Ca^{2+}]_c$  and can be prevented by PDE3-, but not PDE4-inihibition.<sup>80-82</sup>

Levosimendan's clinical activity during long-term treatment is mainly governed by its active metabolite OR-1896, which has a much longer half-life (81 vs. 1 h<sup>83</sup>). OR-1896 stimulated contractile force with a roughly similar potency as levosimendan, had a 4.5-fold lower potency as a Ca<sup>2+</sup> sensitizer and a 38-fold lower potency as a PDE3 inhibitor.<sup>78</sup> Yet, even the inotropic effect of OR-1896 is sensitive to carbachol and therefore likely mediated by PDE3 inhibition.<sup>84</sup> Therefore, also for OR-1896, a combination of PDE3-inhibition with Ca<sup>2+</sup> sensitization is the mechanism that is responsible for positive inotropy.

A common alternative explanation why levosimendan does not prolong relaxation (other than through PDE3-inhibition) is that levosimendan's binding to troponin C is Ca<sup>2+</sup>-dependent. However, it is currently unclear – if not rather unlikely – whether levosimendan can bind and unbind troponin C on a beat-to-beat basis in a millisecond and micromolar range, as would be required to explain the lack of relaxation prolongation by this mechanism (see the **Online Supplement** for a detailed discussion of this issue). Besides its effects on EC coupling, levosimendan also activates glibenclamide-sensitive *sarcolemmal* ATP-dependent K<sup>+</sup>-currents ( $I_{KATP}$ ), which may add to its vasodilating activity and potentially provide cardioprotective effects through activation of *mitochondrial*  $I_{KATP}$  (see **Online Supplement** for a more detailed discussion).

These data indicate that for levosimendan, PDE3 inhibition synergizes with Ca<sup>2+</sup> sensitization for its inotropic action (**Figure 5**), which may be particularly relevant to human failing myocardium in which PDE3 plays the dominant role for controlling intracellular cAMP.<sup>60</sup> From this it can be predicted that the more  $\beta$ -ARs are pre-activated by endogenous or exogenous catecholamines, the more pronounced is the inotropic effect of levosimendan, and the more this effect is mediated by PDE3-inibition rather than Ca<sup>2+</sup> sensitization. Conversely, at low  $\beta$ -AR pre-activation (such as during pharmacological  $\beta$ -blockade), the Ca<sup>2+</sup> sensitization effect of levosimendan may become more important for inotropy. In this context, it is interesting to observe that in the SURVIVE trial, patients with (but not without)  $\beta$ -blocker pre-treatment had improved short-term survival with levosimendan compared to dobutamine.<sup>85, 86</sup> Conversely, in patients with septic shock who

were all co-treated with catecholamines, levosimendan was associated with higher rates of supraventricular tachycardia and a numerical, but non-significant increase in mortality (hazard ratio 1.24, p=0.17).<sup>87</sup> Therefore, although the clinical evidence for these considerations is limited, levosimendan's PDE3-inhibitory effect may be potentiated through pre-activation of  $\beta$ -ARs by endogenous or exogenous catecholamines also under *in vivo* conditions, and such potentiated adrenergic signaling may potentially contribute to arrhythmias and adverse consequences for cardiomyocyte biology (Figure 2).

#### **Clinical trials**

Several clinical trials tested the effects of levosimendan in patients with HF, comparing it to either placebo or dobutamine. In the LIDO trial, levosimendan improved hemodynamics more effectively than dobutamine and was associated with lower mortality than dobutamine after 180 days.<sup>88</sup> In the SURVIVE trial, however, mortality after 180 days (the primary endpoint) was not different between dobutamine and levosimendan despite a more favourable hemodynamic profile (BNP reduction) in the first 5 days after randomization.<sup>12</sup> As mentioned above, pre-treatment with a  $\beta$ -blocker was associated with improved short-term outcome at day 5 in a posthoc analysis.<sup>85</sup>

In the REVIVE trial,<sup>13</sup> levosimendan (compared to placebo) was associated with more frequent hypotension and cardiac arrhythmias during the infusion period, and a numerical (but insignificant) risk of death despite improved symptoms and reduced plasma BNP levels. In a meta-analysis on 5,480 patients in 45 randomized clinical trials, however, levosimendan was associated with a 20% relative risk reduction of mortality, and this reduction was confirmed in studies with placebo (-18%; p<0.05) or dobutamine as comparator (-32%; p<0.005).<sup>10</sup> These trends were confirmed by another meta-analysis.<sup>11</sup> However, in these meta-analyses, trials on patients with acute cardiac events were combined with trials on patients undergoing elective cardiac surgery. Furthermore, in the recent CHEETAH<sup>14</sup> and LEVO-CTS<sup>15</sup> trials, levosimendan did not improve outcome of patients with systolic HF undergoing cardiac surgery, although the use of inotropes 24 hours after surgery was reduced with levosimendan in LEVO-CTS.<sup>15</sup>

Several smaller and mostly underpowered trials evaluated the usefulness of repeated doses of levosimendan for patients with terminal HF in outpatient settings. Since posthocand meta-analyses of these smaller trials suggest that levosimendan may have favourable effects on hemodynamics, symptoms, rehospitalisation and biomarkers, the initiation of an adequately powered trial is warranted.<sup>89</sup>

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Taken together, the principle of Ca<sup>2+</sup> sensitization alone was no breakthrough in the treatment of the common forms of HF, because it is associated with worsening of diastolic relaxation, which is already compromised in HF in the first place. The ancillary PDE-inhibitory effect of levosimendan improves its hemodynamic profile compared to other, more pure Ca<sup>2+</sup> sensitizers, although this may come at the cost of cAMP-related side effects (e.g., arrhythmias). No single trial on its own, however, could so far show superiority of levosimendan compared to placebo or a comparator drug.

Based on the clinical profile of levosimendan, the current HF Guidelines make the following recommendations for the use of levosimendan:

- Short-term intravenous infusion of levosimendan may be considered in patients with hypotension (SBP <90 mmHg) and/or signs/symptoms of hypoperfusion despite adequate filling status to increase cardiac output and improve peripheral perfusion and maintain end-organ function (similar to dobutamine, dopamine and PDEinhibtors; IIb, C).<sup>3</sup>
- When mean arterial pressure needs pharmacological support, a vasopressor (preferably noradrenaline) may be used in combination with levosimendan.<sup>3</sup>
- An intravenous infusion of levosimendan (or a PDE inhibitor) may be considered to reverse the effect of β-blockade if β-blockade is thought to be contributing to hypotension with subsequent hypoperfusion (IIb, C).<sup>3</sup>
- Levosimendan is not recommended unless the patient is symptomatically hypotensive or hypoperfused because of safety concerns (IIIA).<sup>3</sup>

## **Omecamtiv mecarbil**

Omecamtiv mecarbil (OM) is a small-molecule, selective cardiac myosin activator whose therapeutic rationale and discovery were described elsewhere.<sup>90, 91</sup> OM binds to the catalytic domain of cardiac myosin, stabilizing the pre-powerstroke state,<sup>92</sup> thus increasing the transition rate of myosin into the strongly actin-bound force-generating state (**Figure 6A**)<sup>93</sup> and thus increasing cardiac contractility. The pharmacodynamic signature of OM is an increase in the systolic ejection time (SET). This is a consequence of the increase in the number of myosin heads interacting with actin filaments, facilitating a longer duration of systole, even as  $[Ca^{2+}]_c$  already decays. OM prolongs the time and increases the amplitude, but not the rate of cell shortening, and does not interfere with  $[Ca^{2+}]_c$  transients (**Figure 6B**).<sup>93</sup>

A number of dose-finding studies with an intravenous formulation of OM were performed in a dog model of HF,<sup>94</sup> healthy men<sup>17</sup> and patients with acute<sup>19</sup> and chronic HF,<sup>16</sup> respectively. In the dog HF model, OM prolonged SET and increased stroke volume (SV; **Table 2**).<sup>93, 94</sup> As a consequence, cardiac output increased, thereby decreasing SVR and heart rate, presumably through reducing endogenous sympathetic activation.<sup>93, 94</sup> Although OM might increase ATP turnover at the level of the sarcomere,<sup>93</sup> this increase appears to be counterbalanced by the reduction in heart rate, SVR (reducing cardiac afterload) and end diastolic volume (reducing myocardial wall stress), resulting in no significant change of cardiac O<sub>2</sub> consumption despite the increase of cardiac output.<sup>93, 94</sup> Consequently, cardiac efficiency should improve.

In healthy men and patients with stable HF, at comparable increases in SET, however, the net increases in SV, cardiac output and the ensuing decreases in heart rate were overall smaller than in the preclinical studies, perhaps reflecting the broader range of baseline conditions found in human studies (**Table 2**). In these early studies, OM was studied over a broad range of plasma concentrations, in some cases exceeding 1200 ng/ml. Increases in SET are noted at plasma concentrations as low as 100-200 ng/ml, while the effect on SV appeared to plateau at 400-500 ng/ml. In some individuals, myocardial ischemia developed with chest pain, ECG changes and/or troponin rises at plasma concentrations beyond 1200 ng/ml.<sup>16, 17</sup> This may be explained by an excessive increase in SET, prolonging cardiac contraction, and progressively shortening diastole (during which coronary perfusion takes place).<sup>16, 17</sup> In a trial of patients with ischemic cardiomyopathy and angina in daily life, however, OM at target plasma concentrations of 295 ng/ml and 550 ng/ml, respectively, did not affect symptom-limited exercise capacity in treadmill tests or plasma troponin I levels.<sup>18</sup> Subsequent trials focused on dose regimens that constrain exposure to less than 1000 ng/ml.

In the ATOMIC-AHF study on patients with AHF and an LVEF  $\leq$ 40%, the primary endpoint of dyspnea relief was not reached by 3 ascending doses of intravenous infusion of OM versus placebo.<sup>19</sup> However, in the highest dose group (n=202), more patients responded with dyspnea relief to OM (51%) than to placebo (37%; p=0.034). In an echocardiographic substudy, OM prolonged SET and decreased LV end-systolic dimension, although LV stroke volume was not increased.<sup>19</sup> Additionally, slight decreases in heart rate (-2 bpm) and increases in systolic blood pressure were noted (Table 2).

In the COSMIC-HF trial, *oral* OM at either a fixed dose (25 mg twice daily) or dosing based on a pharmacokinetic titration protocol was tested against placebo in patients with *stable* 

(not acute) systolic HF receiving standard of care therapy.<sup>20</sup> After 20 weeks, moderate increases in SET and SV and a slight reduction in heart rate were noted in the pharmacokinetic titration group (**Table 2**). The latter effect may reflect slightly reduced endogenous sympathetic activity.<sup>95</sup> Furthermore, the LV end-diastolic volume decreased by 11 ml and NT-proBNP levels dropped by 970 pg/ml compared to placebo, respectively. As in ATOMIC-AHF,<sup>19</sup> there was a small increase in cardiac troponin I that did not correlate with OM plasma concentrations.<sup>16, 17</sup> The frequency of deaths, arrhythmias, hospital admissions or adverse events was not different between groups, suggesting safety.

Overall, the hemodynamic profile of OM appears promising within its therapeutic range. The increase in cardiac contractility and subsequent prolongation of SET increases LV stroke volume in patients with chronic heart failure and consequently, blood pressure should rise initially which then may reduce endogenous sympathetic activation. This is indicated by the slight though consistent lowering of heart rate in human and animal studies. As a result, cardiac output in humans appears largely unchanged despite the modest decrease in heart rate, suggesting improved cardiac efficiency. Furthermore, the decrease in LV filling pressures, as indicated by the decrease in NT-proBNP in COSMIC-HF or the decrease of LV end-diastolic pressures in acute studies in the dog indicate LV unloading that may facilitate reverse remodeling of the LV. The now initiated GALACTIC-HF trial (NCT02929329), which aims to include 8000 patients with chronic HF will eventually clarify the long-term outcome by OM. Meanwhile, some uncertainties remain regarding the mechanisms of action of OM and their implications for cardiac function and long-term outcome.

#### Diastolic dysfunction

In cardiac myocytes, the increase in systolic function by OM comes at the cost of increased diastolic tension (**Figure 6C**), indicated by shortening of diastolic cell length. At low [OM] of 200-400 nM, systolic improvement outweighs the diastolic deficit in rat cardiac myocytes, while at 800 nM, this relation reverses. In an *in vivo* pig model of myocardial stunning after ischemia/reperfusion, OM reduced both end-diastolic and end-systolic volumes to similar extents (**Figure 6E**) at concentrations that prolonged SET by 20% (**table 2**). Thereby LVEF pseudo-increased, while SV did not.<sup>96</sup> Furthermore, the OM-induced increase in cardiac output was smaller in humans with or without HF compared to the dogs with HF (**table 3**). In light of the results on post-ischemic pigs,<sup>96</sup> it needs to be considered whether an improvement of SET (and therefore, SV) by OM may have been (partly) offset by decreases in end-diastolic volume or filling. The improvement of NT-proBNP by long-term OM in

COSMIC-HF,<sup>20</sup> however, rather argues against a meaningful deterioration of diastolic function by these doses of OM.

#### **Bioenergetic aspects**

In dogs with HF, OM did not increase O<sub>2</sub> consumption<sup>94</sup> although in the post-ischemic pig model, O<sub>2</sub> consumption tended to increase.<sup>96</sup> In isolated mouse hearts, OM impaired myocardial efficacy by increasing O<sub>2</sub> consumption in working hearts and during basal (resting) metabolism, which was abolished by a myosin-ATPase inhibitor.<sup>96</sup> These data suggest that OM increases (tonic) myosin ATPase activity<sup>97</sup> and thereby O<sub>2</sub> consumption, which however contrasts with the effect of OM to inhibit the basal ATPase of myosin *in vitro*.<sup>96</sup> In skinned rat cardiac myocytes, OM shifted the pCa/force relationship to the left, indicative of sensitizing myofilaments to Ca<sup>2+</sup> (Figure 6F).<sup>99</sup> In human myocardium, OM increased the myosin duty ratio which resulted in enhanced Ca<sup>2+</sup> sensitivity, but slower force development.<sup>100</sup> In a mouse model of dilated cardiomyopathy with *decreased* myofilament Ca<sup>2+</sup> sensitivity, OM resensitized myofilaments towards control levels.<sup>101</sup> However, in the majority of patients with HF, the Ca<sup>2+</sup> affinity of the myofilaments is *increased* rather than *decreased*,<sup>28-30, 102, 103</sup> and in LV myocardium of patients with terminal HF, increased *diastolic* tension consumes as much ATP and O<sub>2</sub> as *systolic* tension, and elevated diastolic tension is a substantial energetic burden in failing hearts especially at higher heart rates.<sup>104</sup>

In conclusion, whether OM has a neutral or even net energy-sparing effect on myocardial bioenergetics, or whether the drug's net effect on myosin could increase O<sub>2</sub>-consumption is a question future research should continue to address. Furthermore, the development of small molecules targeting sarcomeric motor proteins is an emerging field that is discussed in more detail in the **Online Supplement**.

## Alternative treatments targeting EC coupling

### Nitroxyl

Nitroxyl (HNO) is produced by NO synthase under conditions of oxidative or nitrosative stress. HNO donated by Angeli's salt (AS) improves cardiac function in normal and failing dogs, independently of β-AR signaling, with no change in cGMP levels.<sup>105, 106</sup> In cardiac myocytes, HNO increases fractional shortening (FS) and Ca<sup>2+</sup> transients with no involvement of cAMP/PKA or cGMP/protein kinase G signaling.<sup>107</sup> Instead, HNO modifies cysteine residues to enhance Ca<sup>2+</sup> handling and increase myofilament Ca<sup>2+</sup> sensitivity. In particular, HNO alters the inhibitory interaction between phospholamban and SERCA2a in a redox-dependent manner, improving SR Ca<sup>2+</sup> uptake and release in isolated myocytes/hearts (**Figure 7**).<sup>108, 109</sup> In addition, HNO modifies the actin-tropomyosin and myosin heavy chain-myosin light chain 1 interactions, increasing Ca<sup>2+</sup> sensitivity and force generation in intact and skinned muscles (**Figure 7**).<sup>110</sup>

Whereas beneficial effects of NO signaling to cGMP may be lost in conditions of cardiac oxidative stress (by the high reactivity of NO with ROS), this does not occur with HNO, whose efficacy is preserved in conditions with altered redox balance.<sup>111, 112</sup> Novel HNO donors (e.g. CXL-1020) are long-lasting and more specific. CXL-1020 has HNO-dependent positive inotropic and lusitropic effects in isolated cardiac myocytes *in vitro* and in whole animal studies *in vivo*, in both normal and failing conditions. Human phase I-IIa clinical trials were recently completed (NCT01092325, NCT01096043). In patients with systolic HF, CXL-1020 reduced both left and right heart filling pressures and SVR, while increasing SV. Heart rate was unchanged, and arterial pressure declined modestly.<sup>113</sup> Currently, an improved second-generation HNO donor, CXL-1427, is tested in Phase I and II trials on healthy volunteers and patients with HF (NCT02157506, NCT02819271).

### Other compounds and interventions

The mechanisms and, where appropriate, clinical results of Istaroxime, SERCA2a gene therapy and EF hand proteins are discussed in the **Online Supplement**.

## **Energetic considerations**

Taken together, inotropic agents have three principal modes of action:

- 1) Activation of the adrenergic system,
- 2) sensitization of myofilaments to Ca<sup>2+</sup> and
- 3) reconstitution of cytosolic  $Ca^{2+}$  handling independent of adrenergic activation.

The modes of action of inotropic drugs have important energetic consequences. Most cellular ATP is consumed by SERCA, NKA and myosin ATPase.<sup>114</sup> A central mechanism to match ATP supply to demand is "parallel activation" by Ca<sup>2+,31</sup> where Ca<sup>2+</sup> activates both ATP consumption and -regeneration (Figures 3 and 8): Increases in [Ca<sup>2+</sup>]<sub>c</sub> accelerate ATP consumption by EC coupling, hastening respiration via ADP which oxidizes NADH and FADH<sub>2</sub> at the respiratory chain. On the other hand, Ca<sup>2+</sup> enters mitochondria to activate Krebs cycle dehydrogenases, accelerating NADH and FADH<sub>2</sub> regeneration (Figure 8).<sup>31</sup> Consequently, hormones or drugs that increase Ca<sup>2+</sup> handling (e.g., catecholamines) in the short term induce this "parallel activation" of respiration, maintaining the redox state of NADH and FADH<sub>2</sub> in *normal* hearts (Figure 8).<sup>31</sup> In the failing heart, however, mitochondrial Ca<sup>2+</sup> uptake is impaired, resulting in NADH oxidation during  $\beta$ -adrenergic stimulation.<sup>31, 33</sup> Since NADH is coupled to the NADPH pool, and NADPH required for ROS detoxification (Figures 3 and 8), a mismatch between cardiac work and mitochondrial Ca<sup>2+</sup> uptake induces oxidative stress.<sup>31, 33</sup> This may contribute to arrhythmias, systolic dysfunction and maladaptive remodeling through necrosis and other redox-sensitive signaling pathways.<sup>33</sup> Therefore, improving SR Ca<sup>2+</sup> content and release, as has been observed with AAV1/SERCA2a<sup>115</sup> or HNO<sup>107</sup> in animal models of HF, may improve the efficiency of mitochondrial Ca<sup>2+</sup> uptake and make the failing heart less sensitive towards  $\beta$ -AR-mediated oxidative stress and damage. In patients with improved LVEF in response to β-blockers, SERCA gene expression was strongly upregulated,<sup>116</sup> suggesting that also β-blockers may improve mitochondrial redox regulation through restoring defective EC coupling in the long term.

Conversely, when sensitizing myofilaments to  $Ca^{2+}$ , the increase in ATP consumption at the myofilaments may not be adequately matched by  $Ca^{2+}$ -induced Krebs cycle activation in mitochondria (**Figure 8**). In fact, pre-stretching cardiac trabeculae increases force production independent of an increase in  $Ca^{2+}$  via the Frank Starling mechanism, which is related to myofilament  $Ca^{2+}$  sensitization, and this oxidizes NADH.<sup>117</sup> Whether such energetic mismatch and possibly further downstream consequences such as energetic deficit and oxidative stress are also the result of  $Ca^{2+}$  sensitization through inotropic drugs

(i.e., EMD-57033, levosimendan, OM, nitroxyl etc.), mutations occurring in patients with HCM<sup>73, 74</sup> and/or post-translational modifications occurring in patients with systolic HF<sup>103</sup> needs to be clarified by future research. In the context of inotropic drugs, however, a profile where Ca<sup>2+</sup> sensitization is coupled to restoration of Ca<sup>2+</sup> handling (i.e. nitroxyl) or increases of Ca<sup>2+</sup> (i.e., levosimendan; **table 3**) may ameliorate energetic/redox mismatch occurring through Ca<sup>2+</sup> sensitization *per se*.

Finally, it should be considered whether targeting mitochondria may be an alternative indirect inotropic intervention, since in dogs with HF, elamipretide – which accumulates in mitochondria and improves mitochondrial function – *acutely* increased cardiac output by a similar extent (+25%)<sup>118</sup> as OM (+22 and +29%, respectively; **Table 3**).<sup>93, 94</sup> Furthermore, trimetazidine and perhexilline target substrate metabolism of mitochondria and improve LVEF, hemodynamics, cardiac energetics and symptoms in patients with HF (for more details on these compounds see **Online Supplement**).<sup>119-123</sup> Finally, cardiac myocyte-specific iron deficiency impaired the response to dobutamine in preclinical models of HF which could be restored by iron supplementation,<sup>124</sup> suggesting that pharmacological restoration of mitochondrial function may also regenerate the heart's response to inotropic stimulation.

## Summary

Catecholamines and PDE-inhibitors are associated with excess mortality presumably related to the induction of arrhythmias in the short term and the activation of signaling pathways that aggravate maladaptive remodelling of the failing heart in the long term. Although levosimendan has so far been viewed as a Ca<sup>2+</sup> sensitizer, its inotropic effect relies on PDE3-inhibition as well. As a myosin activator, OM improves systolic function without activating adrenergic signaling or increasing cytosolic Ca<sup>2+</sup>, but its therapeutic range is limited by diastolic dysfunction at higher doses. Nitroxyl restores cytosolic Ca<sup>2+</sup> handling in failing hearts without activating cAMP-dependent signaling pathways and shows a promising hemodynamic profile, but its clinical usefulness awaits further clinical testing. Besides the impact of adrenergic signaling, bioenergetics aspects need to be considered to estimate the comprehensive profile and long-term consequences of any agent that affects inotropy.

## **Statements and recommendations**

Based on these preclinical and clinical data as well as the bioenergetic considerations, the Committees on Translational Research and on Acute Heart Failure of the HFA of the ESC make the following statements and recommendations:

- **Currently available** drugs primarily targeting inotropy are cardiotropic glycosides, catecholamines (in particular, dobutamine), PDE-inhibitors and levosimendan.
- According to the current HF Guidelines,<sup>3</sup> the use of catecholamines, PDE-inhibitors and levosimendan should be limited to patients with hypotension (SBP <90 mmHg) and/or signs/symptoms of hypoperfusion despite adequate filling status to increase cardiac output and improve peripheral perfusion to maintain end-organ function (IIb, C).</li>
- PDE-inhibitors and levosimendan can **cause hypotension** due to vasodilatory actions.
- Vasopressors (preferably norepinephrine) may be considered to increase blood pressure and vital organ perfusion in patients with cardiogenic shock despite the use of inotropes (IIb, B).<sup>3</sup>
- So far, all inotropic drugs recommended for the use in patients with *acute* HF (including levosimendan) activate adrenergic signaling at least to some extent.
- Longer-term use of drugs that **exclusively target adrenergic signaling** (catecholamines, PDE-inhibitors) are associated with adverse outcome.
- Levosimendan, with its hybrid Ca<sup>2+</sup> sensitization and adrenergic action (PDE3 inhibition), could be useful in selected patient populations, which may include patients treated with β-blockers.
- Since **mitochondrial function** is intimately linked to cellular Ca<sup>2+</sup> handling,<sup>31</sup> the bioenergetic consequences of treatments targeting inotropy need to be considered to understand their short- and long-term consequences.
- On theoretical grounds, treatments that restore the defects of cytosolic Ca<sup>2+</sup> handling in the failing heart without activating adrenergic signaling may be a promising avenue since they avoid diastolic dysfunction and potentially bioenergetic mismatch of pure Ca<sup>2+</sup> sensitization, but also adverse long-term consequences of adrenergic activation.
- Future research should be directed towards deepening our understanding of the close interplay between EC coupling and mitochondrial energetics, since only the integration of these aspects will resolve the net biological effects of drugs targeting inotropy in the short- and long-term.

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## References:

1. Ambrosy AP, Fonarow GC, Butler J, Chioncel O, Greene SJ, Vaduganathan M, Nodari S, Lam CSP, Sato N, Shah AN, Gheorghiade M. The Global Health and Economic Burden of Hospitalizations for Heart Failure: Lessons Learned From Hospitalized Heart Failure Registries. Journal of the American College of Cardiology 2014;**63**(12):1123-1133.

2. Chioncel O, Mebazaa A, Harjola V-P, Coats AJ, Piepoli MF, Crespo-Leiro MG, Laroche C, Seferovic PM, Anker SD, Ferrari R, Ruschitzka F, Lopez-Fernandez S, Miani D, Filippatos G, Maggioni AP, on behalf of the ESCHFL-TRI. Clinical phenotypes and outcome of patients hospitalized for acute heart failure: the ESC Heart Failure Long-Term Registry. Eur J Heart Fail 2017;**19**(10):1242-1254.

3. Ponikowski P, Voors AA, Anker SD, Bueno H, Cleland JG, Coats AJ, Falk V, Gonzalez-Juanatey JR, Harjola VP, Jankowska EA, Jessup M, Linde C, Nihoyannopoulos P, Parissis JT, Pieske B, Riley JP, Rosano GM, Ruilope LM, Ruschitzka F, Rutten FH, van der Meer P, Authors/Task Force M, Document R. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC). Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. Eur J Heart Fail 2016;**18**(8):891-975.

4. The Effect of Digoxin on Mortality and Morbidity in Patients with Heart Failure. New England Journal of Medicine 1997;**336**(8):525-533.

5. Bers DM. Altered Cardiac Myocyte Ca Regulation In Heart Failure. Physiology (Bethesda) 2006;**21**(6):380-387.

6. Overgaard CB, Dzavik V. Inotropes and vasopressors: review of physiology and clinical use in cardiovascular disease. Circulation 2008;**118**(10):1047-56.

7. Bristow MR. Treatment of chronic heart failure with beta-adrenergic receptor antagonists: a convergence of receptor pharmacology and clinical cardiology. Circ Res 2011;**109**(10):1176-94.

8. Lohse MJ, Engelhardt S, Eschenhagen T. What is the role of beta-adrenergic signaling in heart failure? Circ Res 2003;**93**(10):896-906.

9. Francis GS, Bartos JA, Adatya S. Inotropes. Journal of the American College of Cardiology 2014;**63**(20):2069-78.

10. Landoni G, Biondi-Zoccai G, Greco M, Greco T, Bignami E, Morelli A, Guarracino F, Zangrillo A. Effects of levosimendan on mortality and hospitalization. A meta-analysis of randomized controlled studies. Crit Care Med 2012;**40**:634-46.

11. Pollesello P, Parissis J, Kivikko M, Harjola VP. Levosimendan meta-analyses: Is there a pattern in the effect on mortality? International journal of cardiology 2016;**209**:77-83.

12. Mebazaa A, Nieminen MS, Packer M, Cohen-Solal A, Kleber FX, Pocock SJ, Thakkar R, Padley RJ, Poder P, Kivikko M, Investigators S. Levosimendan vs dobutamine for patients with acute decompensated heart failure: the SURVIVE Randomized Trial. JAMA : the journal of the American Medical Association 2007;**297**(17):1883-91.

13. Packer M, Colucci W, Fisher L, Massie BM, Teerlink JR, Young J, Padley RJ, Thakkar R, Delgado-Herrera L, Salon J, Garratt C, Huang B, Sarapohja T, Group RHFS. Effect of levosimendan on the short-term clinical course of patients with acutely decompensated heart failure. JACC Heart failure 2013;**1**(2):103-11.

14. Landoni G, Lomivorotov VV, Alvaro G, Lobreglio R, Pisano A, Guarracino F, Calabrò MG, Grigoryev EV, Likhvantsev VV, Salgado-Filho MF, Bianchi A, Pasyuga VV, Baiocchi M, Pappalardo F, Monaco F, Boboshko VA, Abubakirov MN, Amantea B, Lembo R, Brazzi L, Verniero L, Bertini P, Scandroglio AM, Bove T, Belletti A, Michienzi MG, Shukevich DL, Zabelina TS, Bellomo R, Zangrillo A. Levosimendan for Hemodynamic Support after Cardiac Surgery. N Engl J Med 2017;**376**(21):2021-2031.

15. Mehta RH, Leimberger JD, van Diepen S, Meza J, Wang A, Jankowich R, Harrison RW, Hay D, Fremes S, Duncan A, Soltesz EG, Luber J, Park S, Argenziano M, Murphy E, Marcel R, Kalavrouziotis D, Nagpal D, Bozinovski J, Toller W, Heringlake M, Goodman SG, Levy JH, Harrington RA, Anstrom KJ, Alexander JH. Levosimendan in Patients with Left

Ventricular Dysfunction Undergoing Cardiac Surgery. N Engl J Med 2017;**376**(21):2032-2042.

16. Cleland JG, Teerlink JR, Senior R, Nifontov EM, Mc Murray JJ, Lang CC, Tsyrlin VA, Greenberg BH, Mayet J, Francis DP, Shaburishvili T, Monaghan M, Saltzberg M, Neyses L, Wasserman SM, Lee JH, Saikali KG, Clarke CP, Goldman JH, Wolff AA, Malik FI. The effects of the cardiac myosin activator, omecamtiv mecarbil, on cardiac function in systolic heart failure: a double-blind, placebo-controlled, crossover, dose-ranging phase 2 trial. Lancet 2011;**378**(9792):676-83.

17. Teerlink JR, Clarke CP, Saikali KG, Lee JH, Chen MM, Escandon RD, Elliott L, Bee R, Habibzadeh MR, Goldman JH, Schiller NB, Malik FI, Wolff AA. Dose-dependent augmentation of cardiac systolic function with the selective cardiac myosin activator, omecamtiv mecarbil: a first-in-man study. Lancet 2011;**378**(9792):667-75.

18. Greenberg BH, Chou W, Saikali KG, Escandon R, Lee JH, Chen MM, Treshkur T, Megreladze I, Wasserman SM, Eisenberg P, Malik FI, Wolff AA, Shaburishvili T. Safety and tolerability of omecamtiv mecarbil during exercise in patients with ischemic cardiomyopathy and angina. JACC Heart failure 2015;**3**(1):22-9.

19. Teerlink JR, Felker GM, McMurray JJ, Ponikowski P, Metra M, Filippatos GS, Ezekowitz JA, Dickstein K, Cleland JG, Kim JB, Lei L, Knusel B, Wolff AA, Malik FI, Wasserman SM, Investigators A-A. Acute Treatment With Omecamtiv Mecarbil to Increase Contractility in Acute Heart Failure: The ATOMIC-AHF Study. Journal of the American College of Cardiology 2016;**67**(12):1444-55.

20. Teerlink JR, Felker GM, McMurray JJV, Solomon SD, Adams Jr KF, Cleland JGF, Ezekowitz JA, Goudev A, Macdonald P, Metra M, Mitrovic V, Ponikowski P, Serpytis P, Spinar J, Tomcsányi J, Vandekerckhove HJ, Voors AA, Monsalvo ML, Johnston J, Malik FI, Honarpour N. Chronic Oral Study of Myosin Activation to Increase Contractility in Heart Failure (COSMIC-HF): a phase 2, pharmacokinetic, randomised, placebo-controlled trial. The Lancet 2016;**388**(10062):2895-2903.

21. Tilley DG. G protein-dependent and G protein-independent signaling pathways and their impact on cardiac function. Circ Res 2011;**109**(2):217-30.

22. Bobin P, Belacel-Ouari M, Bedioune I, Zhang L, Leroy J, Leblais V, Fischmeister R, Vandecasteele G. Cyclic nucleotide phosphodiesterases in heart and vessels: A therapeutic perspective. Archives of cardiovascular diseases 2016;**109**(6-7):431-43.

23. Clancy CE, Chen-Izu Y, Bers DM, Belardinelli L, Boyden PA, Csernoch L, Despa S, Fermini B, Hool LC, Izu L, Kass RS, Lederer WJ, Louch WE, Maack C, Matiazzi A, Qu Z, Rajamani S, Rippinger CM, Sejersted OM, O'Rourke B, Weiss JN, Varro A, Zaza A. Deranged sodium to sudden death. J Physiol 2015;**593**(6):1331-45.

24. Weisser-Thomas J, Piacentino V, 3rd, Gaughan JP, Margulies K, Houser SR. Calcium entry via Na/Ca exchange during the action potential directly contributes to contraction of failing human ventricular myocytes. Cardiovasc Res 2003;**57**(4):974-85.

25. Mulieri LA, Hasenfuss G, Leavitt B, Allen PD, Alpert NR. Altered myocardial forcefrequency relation in human heart failure. Circulation 1992;**85**(5):1743-50.

26. Hasenfuss G, Schillinger W, Lehnart SE, Preuss M, Pieske B, Maier LS, Prestle J, Minami K, Just H. Relationship between Na+-Ca2+-exchanger protein levels and diastolic function of failing human myocardium. Circulation 1999;**99**(5):641-8.

27. Hasenfuss G, Holubarsch C, Hermann HP, Astheimer K, Pieske B, Just H. Influence of the force-frequency relationship on haemodynamics and left ventricular function in patients with non-failing hearts and in patients with dilated cardiomyopathy. European heart journal 1994;**15**(2):164-70.

28. van der Velden J, Klein LJ, Zaremba R, Boontje NM, Huybregts MAJM, Stooker W, Eijsman L, de Jong JW, Visser CA, Visser FC, Stienen GJM. Effects of Calcium, Inorganic Phosphate, and pH on Isometric Force in Single Skinned Cardiomyocytes From Donor and Failing Human Hearts. Circulation 2001;**104**(10):1140-1146.

29. Wolff MR, Buck SH, Stoker SW, Greaser ML, Mentzer RM. Myofibrillar calcium sensitivity of isometric tension is increased in human dilated cardiomyopathies: role of altered beta-adrenergically mediated protein phosphorylation. J Clin Invest 1996;**98**(1):167-176.

30. Marston SB, de Tombe PP. Troponin phosphorylation and myofilament Ca2+sensitivity in heart failure: increased or decreased? Journal of molecular and cellular cardiology 2008;**45**(5):603-7.

31. Bertero E, Maack C. Calcium Signaling and Reactive Oxygen Species in Mitochondria. Circ Res 2018;**122**(10):1460-1478.

32. Neubauer S. The failing heart--an engine out of fuel. The New England journal of medicine 2007;**356**(11):1140-51.

33. Nickel A, Kohlhaas M, Maack C. Mitochondrial reactive oxygen species production and elimination. Journal of molecular and cellular cardiology 2014;**73C**:26-33.

34. Tian R, Halow JM, Meyer M, Dillmann WH, Figueredo VM, Ingwall JS, Camacho SA. Thermodynamic limitation for Ca2+ handling contributes to decreased contractile reserve in rat hearts. Am J Physiol 1998;**275**(6 Pt 2):H2064-71.

35. Wagner S, Rokita AG, Anderson ME, Maier LS. Redox regulation of sodium and calcium handling. Antioxidants & redox signaling 2013;**18**(9):1063-77.

36. Jankowska EA, Rozentryt P, Witkowska A, Nowak J, Hartmann O, Ponikowska B, Borodulin-Nadzieja L, Banasiak W, Polonski L, Filippatos G, McMurray JJV, Anker SD, Ponikowski P. Iron deficiency: an ominous sign in patients with systolic chronic heart failure. European heart journal 2010;**31**(15):ehq158-1880.

37. Jankowska EA, Tkaczyszyn M, Suchocki T, Drozd M, von Haehling S, Doehner W, Banasiak W, Filippatos G, Anker SD, Ponikowski P. Effects of intravenous iron therapy in iron-deficient patients with systolic heart failure: a meta-analysis of randomized controlled trials. European Journal of Heart Failure 2016;**18**(7):786-795.

38. Melenovsky V, Petrak J, Mracek T, Benes J, Borlaug BA, Nuskova H, Pluhacek T, Spatenka J, Kovalcikova J, Drahota Z, Kautzner J, Pirk J, Houstek J. Myocardial iron content and mitochondrial function in human heart failure: a direct tissue analysis. European journal of heart failure 2016.

39. Xu W, Barrientos T, Mao L, Rockman HA, Sauve AA, Andrews NC. Lethal Cardiomyopathy in Mice Lacking Transferrin Receptor in the Heart. Cell Rep 2015;**13**(3):533-545.

40. Barrientos T, Laothamatas I, Koves TR, Soderblom EJ, Bryan M, Moseley MA, Muoio DM, Andrews NC. Metabolic Catastrophe in Mice Lacking Transferrin Receptor in Muscle. EBioMedicine 2015;**2**(11):1705-17.

41. von Hardenberg A, Maack C. Mitochondrial Therapies in Heart Failure. Handb Exp Pharmacol 2017;**243**:491-514.

42. Liu T, Brown DA, O'Rourke B. Role of mitochondrial dysfunction in cardiac glycoside toxicity. Journal of molecular and cellular cardiology 2010;**49**(5):728-736.

43. Gonano LA, Petroff MV. Subcellular Mechanisms Underlying Digitalis-Induced Arrhythmias: Role of Calcium/Calmodulin-Dependent Kinase II (CaMKII) in the Transition from an Inotropic to an Arrhythmogenic Effect. Heart Lung Circ 2014;**23**(12):1118-1124.

44. van Bilsen M, Patel HC, Bauersachs J, Bohm M, Borggrefe M, Brutsaert D, Coats AJS, de Boer RA, de Keulenaer GW, Filippatos GS, Floras J, Grassi G, Jankowska EA, Kornet L, Lunde IG, Maack C, Mahfoud F, Pollesello P, Ponikowski P, Ruschitzka F, Sabbah HN, Schultz HD, Seferovic P, Slart R, Taggart P, Tocchetti CG, Van Laake LW, Zannad F, Heymans S, Lyon AR. The autonomic nervous system as a therapeutic target in heart failure: a scientific position statement from the Translational Research Committee of the Heart Failure Association of the European Society of Cardiology. Eur J Heart Fail 2017;epub ahead of print 2017 Sep 26.

45. Vamos M, Erath JW, Hohnloser SH. Digoxin-associated mortality: a systematic review and meta-analysis of the literature. European heart journal 2015;**36**(28):1831-1838.

46. Bavendiek U, Aguirre Davila L, Koch A, Bauersachs J. Assumption versus evidence: the case of digoxin in atrial fibrillation and heart failure. European heart journal 2017;**38**(27):2095-2099.

47. Chen HH, Anstrom KJ, Givertz MM, Stevenson LW, Semigran MJ, Goldsmith SR, Bart BA, Bull DA, Stehlik J, LeWinter MM, Konstam MA, Huggins GS, Rouleau JL, O'Meara E, Tang WH, Starling RC, Butler J, Deswal A, Felker GM, O'Connor CM, Bonita RE, Margulies KB, Cappola TP, Ofili EO, Mann DL, Davila-Roman VG, McNulty SE, Borlaug BA, Velazquez EJ, Lee KL, Shah MR, Hernandez AF, Braunwald E, Redfield MM, Network NHFCR. Low-dose dopamine or low-dose nesiritide in acute heart failure with renal dysfunction: the ROSE acute heart failure randomized trial. JAMA : the journal of the American Medical Association 2013;**310**(23):2533-43.

48. Giamouzis G, Butler J, Starling RC, Karayannis G, Nastas J, Parisis C, Rovithis D, Economou D, Savvatis K, Kirlidis T, Tsaknakis T, Skoularigis J, Westermann D, Tschöpe C, Triposkiadis F. Impact of Dopamine Infusion on Renal Function in Hospitalized Heart Failure Patients: Results of the Dopamine in Acute Decompensated Heart Failure (DAD-HF) Trial. Journal of cardiac failure 2010;**16**(12):922-930.

Triposkiadis FK, Butler J, Karayannis G, Starling RC, Filippatos G, Wolski K, Parissis J, Parisis C, Rovithis D, Koutrakis K, Skoularigis J, Antoniou C-K, Chrysohoou C, Pitsavos C, Stefanadis C, Nastas J, Tsaknakis T, Mantziari L, Giannakoulas G, Karvounis H, Kalogeropoulos AP, Giamouzis G. Efficacy and safety of high dose versus low dose furosemide with or without dopamine infusion: The Dopamine in Acute Decompensated Heart Failure II (DAD-HF II) Trial. International journal of cardiology 2014;**172**(1):115-121.
 Tuttle RR, Mills J. Dobutamine: development of a new catecholamine to selectively increase cardiac contractility. Circ Res 1975;**36**(1):185-96.

51. Williams RS, Bishop T. Selectivity of dobutamine for adrenergic receptor subtypes: in vitro analysis by radioligand binding. J Clin Invest 1981;**67**(6):1703-11.

52. Ruffolo RR, Jr. The pharmacology of dobutamine. The American journal of the medical sciences 1987;**294**(4):244-8.

53. Tarvasmäki T, Lassus J, Varpula M, Sionis A, Sund R, Køber L, Spinar J, Parissis J, Banaszewski M, Silva Cardoso J, Carubelli V, Di Somma S, Mebazaa A, Harjola V-P. Current real-life use of vasopressors and inotropes in cardiogenic shock - adrenaline use is associated with excess organ injury and mortality. Critical Care 2016;**20**(1):208.

54. Cohn JN, Levine TB, Olivari MT, Garberg V, Lura D, Francis GS, Simon AB, Rector T. Plasma norepinephrine as a guide to prognosis in patients with chronic congestive heart failure. The New England journal of medicine 1984;**311**(13):819-23.

55. Rockman HA, Koch WJ, Lefkowitz RJ. Seven-transmembrane-spanning receptors and heart function. Nature 2002;**415**(6868):206-12.

56. Wagner S, Maier LS, Bers DM. Role of Sodium and Calcium Dysregulation in Tachyarrhythmias in Sudden Cardiac Death. Circ Res 2015;**116**(12):1956-1970.

57. Unverferth DA, Blanford M, Kates RE, Leier CV. Tolerance to dobutamine after a 72 hour continuous infusion. The American journal of medicine 1980;**69**(2):262-6.

58. Tacon CL, McCaffrey J, Delaney A. Dobutamine for patients with severe heart failure: a systematic review and meta-analysis of randomised controlled trials. Intensive care medicine 2012;**38**(3):359-67.

59. Stapel B, Kohlhaas M, Ricke-Hoch M, Haghikia A, Erschow S, Knuuti J, Silvola JMU, Roivainen A, Saraste A, Nickel AG, Saar JA, Sieve I, Pietzsch S, Müller M, Bogeski I, Kappl R, Jauhiainen M, Thackeray JT, Scherr M, Bengel FM, Hagl C, Tudorache I, Bauersachs J, Maack C, Hilfiker-Kleiner D. Low STAT3 expression sensitizes to toxic effects of  $\beta$ -adrenergic receptor stimulation in peripartum cardiomyopathy. European heart journal 2017;**38**(5):349-361.

60. Molenaar P, Christ T, Hussain RI, Engel A, Berk E, Gillette KT, Chen L, Galindo-Tovar A, Krobert KA, Ravens U, Levy FO, Kaumann AJ. PDE3, but not PDE4, reduces beta(1) - and beta(2)-adrenoceptor-mediated inotropic and lusitropic effects in failing ventricle from metoprolol-treated patients. British journal of pharmacology 2013;**169**(3):528-38.

61. Molina CE, Leroy J, Richter W, Xie M, Scheitrum C, Lee IO, Maack C, Rucker-Martin C, Donzeau-Gouge P, Verde I, Llach A, Hove-Madsen L, Conti M, Vandecasteele G, Fischmeister R. Cyclic adenosine monophosphate phosphodiesterase type 4 protects against atrial arrhythmias. Journal of the American College of Cardiology 2012;**59**(24):2182-90.

62. Metra M, Nodari S, D'Aloia A, Muneretto C, Robertson AD, Bristow MR, Dei Cas L. Beta-blocker therapy influences the hemodynamic response to inotropic agents in patients with heart failure: a randomized comparison of dobutamine and enoximone before and after chronic treatment with metoprolol or carvedilol. Journal of the American College of Cardiology 2002;**40**(7):1248-58.

63. Mebazaa A, Parissis J, Porcher R, Gayat E, Nikolaou M, Boas FV, Delgado JF, Follath F. Short-term survival by treatment among patients hospitalized with acute heart failure: the global ALARM-HF registry using propensity scoring methods. Intensive care medicine 2011;**37**(2):290-301.

64. Packer M, Carver JR, Rodeheffer RJ, Ivanhoe RJ, DiBianco R, Zeldis SM, Hendrix GH, Bommer WJ, Elkayam U, Kukin ML, et al. Effect of oral milrinone on mortality in severe chronic heart failure. The PROMISE Study Research Group. The New England journal of medicine 1991;**325**(21):1468-75.

65. Communal C, Singh K, Pimentel DR, Colucci WS. Norepinephrine stimulates apoptosis in adult rat ventricular myocytes by activation of the beta-adrenergic pathway. Circulation 1998;**98**(13):1329-34.

66. Narula J, Haider N, Virmani R, DiSalvo TG, Kolodgie FD, Hajjar RJ, Schmidt U, Semigran MJ, Dec GW, Khaw BA. Apoptosis in myocytes in end-stage heart failure. The New England journal of medicine 1996;**335**(16):1182-9.

67. Wencker D, Chandra M, Nguyen K, Miao W, Garantziotis S, Factor SM, Shirani J, Armstrong RC, Kitsis RN. A mechanistic role for cardiac myocyte apoptosis in heart failure. J Clin Invest 2003;**111**(10):1497-504.

68. Fujino K, Sperelakis N, Solaro RJ. Sensitization of dog and guinea pig heart myofilaments to Ca2+ activation and the inotropic effect of pimobendan: comparison with milrinone. Circ Res 1988;**63**(5):911-922.

69. Pollesello P, Ovaska M, Kaivola J, Tilgmann C, Lundström K, Kalkkinen N, Ulmanen I, Nissinen E, Taskinen J. Binding of a new Ca2+ sensitizer, levosimendan, to recombinant human cardiac troponin C. A molecular modelling, fluorescence probe, and proton nuclear magnetic resonance study. J Biol Chem 1994;**269**(46):28584-28590.

70. Schlecht W, Li K-L, Hu D, Dong W. Fluorescence Based Characterization of Calcium Sensitizer Action on the Troponin Complex. Chem Biol Drug Design 2016;**87**(2):171-181.

71. Radke MB, Taft MH, Stapel B, Hilfiker-Kleiner D, Preller M, Manstein DJ. Small molecule-mediated Refolding and Activation of Myosin Motor Function. eLife 2014:DOI: 10.7554/eLife.01603.

72. Solaro RJ, Gambassi G, Warshaw DM, Keller MR, Spurgeon HA, Beier N, Lakatta EG. Stereoselective actions of thiadiazinones on canine cardiac myocytes and myofilaments. Circ Res 1993;**73**(6):981-990.

73. Moore JR, Leinwand L, Warshaw DM. Understanding cardiomyopathy phenotypes based on the functional impact of mutations in the myosin motor. Circ Res 2012;**111**(3):375-85.

74. Watkins H, Ashrafian H, Redwood C. Inherited cardiomyopathies. The New England journal of medicine 2011;**364**(17):1643-56.

75. Schober T, Huke S, Venkataraman R, Gryshchenko O, Kryshtal D, Hwang HS, Baudenbacher FJ, Knollmann BC. Myofilament Ca sensitization increases cytosolic Ca binding affinity, alters intracellular Ca homeostasis, and causes pause-dependent Ca-triggered arrhythmia. Circ Res 2012;**111**(2):170-9.

76. Baudenbacher F, Schober T, Pinto JR, Sidorov VY, Hilliard F, Solaro RJ, Potter JD, Knollmann BC. Myofilament Ca2+ sensitization causes susceptibility to cardiac arrhythmia in mice. J Clin Invest 2008;**118**(12):3893-903.

77. Raasmaja A, Talo A, Haikala H, Nissinen E, Linden IB, Pohto P. Biochemical properties of OR-1259--a positive inotropic and vasodilatory compound with an antiarrhythmic effect. Advances in experimental medicine and biology 1992;**311**:423.

78. Szilagyi S, Pollesello P, Levijoki J, Kaheinen P, Haikala H, Edes I, Papp Z. The effects of levosimendan and OR-1896 on isolated hearts, myocyte-sized preparations and phosphodiesterase enzymes of the guinea pig. European journal of pharmacology 2004;**486**(1):67-74.

79. Boknik P, Neumann J, Kaspareit G, Schmitz W, Scholz H, Vahlensieck U, Zimmermann N. Mechanisms of the contractile effects of levosimendan in the mammalian heart. The Journal of pharmacology and experimental therapeutics 1997;**280**(1):277-83.

80. Hasenfuss G, Pieske B, Castell M, Kretschmann B, Maier LS, Just H. Influence of the novel inotropic agent levosimendan on isometric tension and calcium cycling in failing human myocardium. Circulation 1998;**98**(20):2141-7.

81. Orstavik O, Ata SH, Riise J, Dahl CP, Andersen GO, Levy FO, Skomedal T, Osnes JB, Qvigstad E. Inhibition of phosphodiesterase-3 by levosimendan is sufficient to account for its inotropic effect in failing human heart. British journal of pharmacology 2014;**171**(23):5169-81.

82. Brixius K, Reicke S, Schwinger RH. Beneficial effects of the Ca(2+) sensitizer levosimendan in human myocardium. Am J Physiol Heart Circ Physiol 2002;**282**(1):H131-7.

83. Antila S, Kivikko M, Lehtonen L, Eha J, Heikkila A, Pohjanjousi P, Pentikainen PJ. Pharmacokinetics of levosimendan and its circulating metabolites in patients with heart failure after an extended continuous infusion of levosimendan. British journal of clinical pharmacology 2004;**57**(4):412-5.

84. Orstavik O, Manfra O, Andressen KW, Andersen GO, Skomedal T, Osnes JB, Levy FO, Krobert KA. The Inotropic Effect of the Active Metabolite of Levosimendan, OR-1896, Is Mediated through Inhibition of PDE3 in Rat Ventricular Myocardium. PloS one 2015;**10**(3):e0115547.

85. Mebazaa A, Nieminen MS, Filippatos GS, Cleland JG, Salon JE, Thakkar R, Padley RJ, Huang B, Cohen-Solal A. Levosimendan vs. dobutamine: outcomes for acute heart failure patients on  $\beta$ -blockers in SURVIVE<sup>†</sup>. Eur J Heart Fail 2009;**11**(3):304-311.

86. Kivikko M, Pollesello P, Tarvasmäki T, Sarapohja T, Nieminen MS, Harjola V-P. Effect of baseline characteristics on mortality in the SURVIVE trial on the effect of levosimendan vs dobutamine in acute heart failure: Sub-analysis of the Finnish patients. International journal of cardiology 2016;**215**:26-31.

87. Gordon AC, Perkins GD, Singer M, McAuley DF, Orme RML, Santhakumaran S, Mason AJ, Cross M, Al-Beidh F, Best-Lane J, Brealey D, Nutt CL, McNamee JJ, Reschreiter H, Breen A, Liu KD, Ashby D. Levosimendan for the Prevention of Acute Organ Dysfunction in Sepsis. The New England journal of medicine 2016;**375**(17):1638-1648.

88. Follath F, Cleland JG, Just H, Papp JG, Scholz H, Peuhkurinen K, Harjola VP, Mitrovic V, Abdalla M, Sandell EP, Lehtonen L, Steering C, Investigators of the Levosimendan Infusion versus Dobutamine S. Efficacy and safety of intravenous levosimendan compared with dobutamine in severe low-output heart failure (the LIDO study): a randomised double-blind trial. Lancet 2002;**360**(9328):196-202.

89. Pölzl G, Altenberger J, Baholli L, Beltrán P, Borbély A, Comin-Colet J, Delgado JF, Fedele F, Fontana A, Fruhwald F, Giamouzis G, Giannakoulas G, Garcia-González MJ, Gustafsson F, Kaikkonen K, Kivikko M, Kubica J, von Lewinski D, Löfman I, Malfatto G, Manito N, Martínez-Sellés M, Masip J, Merkely B, Morandi F, Mølgaard H, Oliva F, Pantev E, Papp Z, Perna GP, Pfister R, Piazza V, Bover R, Rangel-Sousa D, Recio-Mayoral A, Reinecke A, Rieth A, Sarapohja T, Schmidt G, Seidel M, Störk S, Vrtovec B, Wikström G, Yerly P, Pollesello P. Repetitive use of levosimendan in advanced heart failure: need for stronger evidence in a field in dire need of a useful therapy. International journal of cardiology 2017;**243**:389-395.

90. Malik FI, Morgan BP. Cardiac myosin activation part 1: from concept to clinic. J Mol Cell Cardiol;**51**(4):454-461.

91. Morgan BP, Muci A, Lu PP, Qian X. Discovery of omecamtiv mecarbil the first, selective, small molecule activator of cardiac myosin. ACS Med Chem Lett.

92. Planelles-Herrero VJ, Hartman JJ, Robert-Paganin J, Malik FI, Houdusse A. Mechanistic and structural basis for activation of cardiac myosin force production by omecamtiv mecarbil. Nat Commun 2017;**8**(1):190.

93. Malik FI, Hartman JJ, Elias KA, Morgan BP, Rodriguez H, Brejc K, Anderson RL, Sueoka SH, Lee KH, Finer JT, Sakowicz R, Baliga R, Cox DR, Garard M, Godinez G, Kawas R, Kraynack E, Lenzi D, Lu PP, Muci A, Niu C, Qian X, Pierce DW, Pokrovskii M, Suehiro I, Sylvester S, Tochimoto T, Valdez C, Wang W, Katori T, Kass DA, Shen YT, Vatner SF, Morgans DJ. Cardiac myosin activation: a potential therapeutic approach for systolic heart failure. Science 2011;**331**(6023):1439-43. 94. Shen YT, Malik FI, Zhao X, Depre C, Dhar SK, Abarzua P, Morgans DJ, Vatner SF. Improvement of cardiac function by a cardiac Myosin activator in conscious dogs with systolic heart failure. Circulation Heart failure 2010;**3**(4):522-7.

95. Mann DL. Searching for the perfect agent to improve cardiac contractility. Lancet 2016;**388**(10062):2845-2847.

96. Bakkehaug JP, Kildal AB, Engstad ET, Boardman N, Naesheim T, Ronning L, Aasum E, Larsen TS, Myrmel T, How OJ. Myosin Activator Omecamtiv Mecarbil Increases Myocardial Oxygen Consumption and Impairs Cardiac Efficiency Mediated by Resting Myosin ATPase Activity. Circulation Heart failure 2015;**8**(4):766-75.

97. Bakkehaug JP, Kildal AB, Engstad ET, Boardman N, Næsheim T, Rønning L, Aasum E, Larsen TS, Myrmel T, How O-J. Response to Letter Regarding Article, "Myosin Activator Omecamtiv Mecarbil Increases Myocardial Oxygen Consumption and Impairs Cardiac Efficiency Mediated by Resting Myosin ATPase Activity". Circulation Heart failure 2015;**8**(6):1142.

98. Liu Y, White HD, Belknap B, Winkelmann DA, Forgacs E. Omecamtiv Mecarbil Modulates the Kinetic and Motile Properties of Porcine  $\beta$ -Cardiac Myosin. Biochemistry 2015;**54**(10):1963-1975.

99. Nagy L, Kovacs A, Bodi B, Pasztor ET, Fulop GA, Toth A, Edes I, Papp Z. The novel cardiac myosin activator omecamtiv mecarbil increases the calcium sensitivity of force production in isolated cardiomyocytes and skeletal muscle fibres of the rat. British journal of pharmacology 2015;**172**:4506–4518.

100. Swenson AM, Tang W, Blair CA, Fetrow CM, Unrath WC, Previs MJ, Campbell KS, Yengo CM. Omecamtiv Mecarbil Enhances the Duty Ratio of Human Beta Cardiac Myosin Resulting in Increased Calcium Sensitivity and Slowed Force Development in Cardiac Muscle. J Biol Chem 2017;**292**(9):3768-3778.

101. Utter MS, Ryba DM, Li BH, Wolska BM, Solaro RJ. Omecamtiv Mecarbil, a Cardiac Myosin Activator, Increases Ca2+ Sensitivity in Myofilaments With a Dilated Cardiomyopathy Mutant Tropomyosin E54K. Journal of cardiovascular pharmacology 2015;**66**(4):347-353. 102. Messer AE, Jacques AM, Marston SB. Troponin phosphorylation and regulatory function in human heart muscle: dephosphorylation of Ser23/24 on troponin I could account for the contractile defect in end-stage heart failure. Journal of molecular and cellular cardiology 2007;**42**(1):247-59.

103. van der Velden J, Papp Z, Zaremba R, Boontje NM, de Jong JW, Owen VJ, Burton PBJ, Goldmann P, Jaquet K, Stienen GJM. Increased Ca2+-sensitivity of the contractile apparatus in end-stage human heart failure results from altered phosphorylation of contractile proteins. Cardiovasc Res 2003;**57**(1):37-47.

104. Meyer M, Keweloh B, Guth K, Holmes JW, Pieske B, Lehnart SE, Just H, Hasenfuss G. Frequency-dependence of myocardial energetics in failing human myocardium as quantified by a new method for the measurement of oxygen consumption in muscle strip preparations. Journal of molecular and cellular cardiology 1998;**30**(8):1459-70.

105. Paolocci N, Katori T, Champion HC, St John ME, Miranda KM, Fukuto JM, Wink DA, Kass DA. Positive inotropic and lusitropic effects of HNO/NO- in failing hearts: independence from beta-adrenergic signaling. Proc Natl Acad Sci U S A 2003;**100**(9):5537-42.

106. Paolocci N, Saavedra WF, Miranda KM, Martignani C, Isoda T, Hare JM, Espey MG, Fukuto JM, Feelisch M, Wink DA, Kass DA. Nitroxyl anion exerts redox-sensitive positive cardiac inotropy in vivo by calcitonin gene-related peptide signaling. Proc Natl Acad Sci U S A 2001;**98**(18):10463-8.

107. Tocchetti CG, Wang W, Froehlich JP, Huke S, Aon MA, Wilson GM, Di Benedetto G, O'Rourke B, Gao WD, Wink DA, Toscano JP, Zaccolo M, Bers DM, Valdivia HH, Cheng H, Kass DA, Paolocci N. Nitroxyl improves cellular heart function by directly enhancing cardiac sarcoplasmic reticulum Ca2+ cycling. Circ Res 2007;**100**(1):96-104.

108. Froehlich JP, Mahaney JE, Keceli G, Pavlos CM, Goldstein R, Redwood AJ, Sumbilla C, Lee DI, Tocchetti CG, Kass DA, Paolocci N, Toscano JP. Phospholamban thiols play a central role in activation of the cardiac muscle sarcoplasmic reticulum calcium pump by nitroxyl. Biochemistry 2008;**47**(50):13150-2.

109. Sivakumaran V, Stanley BA, Tocchetti CG, Ballin JD, Caceres V, Zhou L, Keceli G, Rainer PP, Lee DI, Huke S, Ziolo MT, Kranias EG, Toscano JP, Wilson GM, O'Rourke B, Kass DA, Mahaney JE, Paolocci N. HNO enhances SERCA2a activity and cardiomyocyte function by promoting redox-dependent phospholamban oligomerization. Antioxidants & redox signaling 2013;**19**(11):1185-97.

110. Gao WD, Murray CI, Tian Y, Zhong X, DuMond JF, Shen X, Stanley BA, Foster DB, Wink DA, King SB, Van Eyk JE, Paolocci N. Nitroxyl-mediated disulfide bond formation between cardiac myofilament cysteines enhances contractile function. Circ Res 2012;**111**(8):1002-11.

111. Paolocci N, Jackson MI, Lopez BE, Miranda K, Tocchetti CG, Wink DA, Hobbs AJ, Fukuto JM. The pharmacology of nitroxyl (HNO) and its therapeutic potential: not just the Janus face of NO. Pharmacology & therapeutics 2007;**113**(2):442-58.

112. Tocchetti CG, Stanley BA, Murray CI, Sivakumaran V, Donzelli S, Mancardi D, Pagliaro P, Gao WD, van Eyk J, Kass DA, Wink DA, Paolocci N. Playing with cardiac "redox switches": the "HNO way" to modulate cardiac function. Antioxidants & redox signaling 2011;**14**(9):1687-98.

113. Sabbah HN, Tocchetti CG, Wang M, Daya S, Gupta RC, Tunin RS, Mazhari R, Takimoto E, Paolocci N, Cowart D, Colucci WS, Kass DA. Nitroxyl (HNO): A novel approach for the acute treatment of heart failure. Circulation Heart failure 2013;**6**(6):1250-8.

114. Bers DM. *Excitation-contraction coupling and cardiac contractile force*. 2nd ed. Dordrecht, The Netherlands: Kluwer Academic Publisher; 2001.

115. Lyon AR, Bannister ML, Collins T, Pearce E, Sepehripour AH, Dubb SS, Garcia E, O'Gara P, Liang L, Kohlbrenner E, Hajjar RJ, Peters NS, Poole-Wilson PA, Macleod KT, Harding SE. SERCA2a gene transfer decreases sarcoplasmic reticulum calcium leak and reduces ventricular arrhythmias in a model of chronic heart failure. Circulation Arrhythmia and electrophysiology 2011;**4**(3):362-72.

116. Lowes BD, Gilbert EM, Abraham WT, Minobe WA, Larrabee P, Ferguson D, Wolfel EE, Lindenfeld J, Tsvetkova T, Robertson AD, Quaife RA, Bristow MR. Myocardial gene expression in dilated cardiomyopathy treated with beta-blocking agents. The New England journal of medicine 2002;**346**(18):1357-65.

117. Brandes R, Bers DM. Intracellular Ca2+ increases the mitochondrial NADH concentration during elevated work in intact cardiac muscle. Circ Res 1997;80(1):82-7.
118. Sabbah HN, Gupta RC, Kohli S, Wang M, Hachem S, Zhang K. Chronic Therapy With Elamipretide (MTP-131), a Novel Mitochondria-Targeting Peptide, Improves Left Ventricular and Mitochondrial Function in Dogs With Advanced Heart Failure. Circulation Heart failure 2016;9(2):e002206.

119. Fragasso G, Piatti MPM, Monti L, Palloshi A, Setola E, Puccetti P, Calori G, Lopaschuk GD, Margonato A. Short- and long-term beneficial effects of trimetazidine in patients with diabetes and ischemic cardiomyopathy. American heart journal 2003;**146**(5):854.

120. Fragasso G, Perseghin G, De Cobelli F, Esposito A, Palloshi A, Lattuada G, Scifo P, Calori G, Del Maschio A, Margonato A. Effects of metabolic modulation by trimetazidine on left ventricular function and phosphocreatine/adenosine triphosphate ratio in patients with heart failure. European heart journal 2006;**27**(8):942-948.

121. Zhang L, Lu Y, Jiang H, Zhang L, Sun A, Zou Y, Ge J. Additional Use of Trimetazidine in Patients With Chronic Heart Failure: A Meta-Analysis. Journal of the American College of Cardiology 2012;**59**(10):913-922.

122. Beadle RM, Williams LK, Kuehl M, Bowater S, Abozguia K, Leyva F, Yousef Z, Wagenmakers AJM, Thies F, Horowitz J, Frenneaux MP. Improvement in Cardiac Energetics by Perhexiline in Heart Failure Due to Dilated Cardiomyopathy. JACC: Heart Failure 2015;**3**(3):202-211.

123. Lee L, Campbell R, Scheuermann-Freestone M, Taylor R, Gunaruwan P, Williams L, Ashrafian H, Horowitz J, Fraser AG, Clarke K, Frenneaux M. Metabolic modulation with perhexiline in chronic heart failure: a randomized, controlled trial of short-term use of a novel treatment. Circulation 2005;**112**(21):3280-8.

124. Haddad S, Wang Y, Galy B, Korf-Klingebiel M, Hirsch V, Baru AM, Rostami F, Reboll MR, Heineke J, Flögel U, Groos S, Renner A, Toischer K, Zimmermann F, Engeli S, Jordan J, Bauersachs J, Hentze MW, Wollert KC, Kempf T. Iron-regulatory proteins secure iron availability in cardiomyocytes to prevent heart failure. European heart journal 2017;**38**(5):362-37.

## **Figure legends**

## Figure 1: The physiology of excitation-contraction coupling and how this is altered in systolic heart failure.

AR, adrenergic receptor; PDE, phosphodiesterase; cAMP, cyclic adenosine monophosphate; PKA, protein kinase A; SR, sarcoplasmic reticulum; SERCA, SR Ca<sup>2+</sup> ATPase; RyR, ryanodine receptor;  $I_{Ca}$  and  $I_{Na}$ , Ca<sup>2+</sup> and Na<sup>+</sup> currents; NCX; Na<sup>+</sup>/Ca<sup>2+</sup>- exchanger; NKA, Na<sup>+</sup>/K<sup>+</sup>-ATPase; ETC, electron transport chain; T-tubule, transversal tubule. Red arrows ( $\uparrow\downarrow$ ) indicate the direction of change in heart failure.

## Figure 2: Signal transduction of $\beta_1$ -adrenergic stimulation in cardiac myocytes and its impact on inotropy, but also arrhythmias, hypertrophy and apoptosis.

ADR, adrenaline; NA, noradrenaline; AR, adrenergic receptor; AC, adenylyl cyclase; cAMP, cyclic adenosine monophosphate; PDE, phosphodiesterase; PKA, protein kinase A; EPAC, exchange protein directly activated by cAMP; 5'-AMP, 5' adenosine monophosphate; CaMKII, Ca<sup>2+</sup>/calmodulin-dependent protein kinase II; GRK2, G-protein coupled receptor kinase 2;  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\alpha$ -,  $\beta$ - and  $\gamma$ -subunits of the stimulatory G-protein.

#### Figure 3: Interplay between EC coupling and mitochondrial energetics.

Krebs cycle activity is controlled by Ca<sup>2+</sup>, and mitochondrial Ca<sup>2+</sup> uptake is diminished in heart failure by changes in cytosolic Ca<sup>2+</sup> and Na<sup>+</sup> handling. This provokes an energetic deficit and oxidative stress, which further impairs EC coupling and aggravates systolic and diastolic function.

Nnt, nicotinamide nucleotide transhydrogenase; ETC, electron transport chain; ATPase,  $F_1F_0$ -ATP synthase; IDP<sub>m</sub>, isocitrate dehydrogenase; Mn-SOD, mitochondrial superoxide dismutase; CK, creatine kinase; Cr, creatine; PCr, phosphocreatine; SERCA, SR Ca<sup>2+</sup> ATPase; RyR, ryanodine receptor; late  $I_{Na}$ , late Na<sup>+</sup> current; NCLX, mitochondrial Na<sup>+</sup>/Ca<sup>2+</sup>- exchanger; MCU, mitochondrial Ca<sup>2+</sup> uniporter; CaMKII, Ca<sup>2+</sup>/calmodulin-dependent protein kinase II; AR, adrenergic receptor. Red arrows ( $\uparrow\downarrow$ ) indicate the direction of change in heart failure.

## Figure 4: Mechanisms of action of cardiotonic glycosides (CTG) and istaroxime (ISTA).

ROS, reactive oxygen species; SR, sarcoplasmic reticulum; SERCA, SR Ca<sup>2+</sup> ATPase; RyR, ryanodine receptor;  $I_{Ca}$  and  $I_{Na}$ , Ca<sup>2+</sup> and Na<sup>+</sup> currents; NCX; Na<sup>+</sup>/Ca<sup>2+</sup>-exchanger; NKA, Na<sup>+</sup>/K<sup>+</sup>-ATPase; NCLX, mitochondrial Na<sup>+</sup>/Ca<sup>2+</sup>-exchanger; TCA, tricarboxylic acid (Krebs) cycle. Red arrows ( $\uparrow\downarrow$ ) indicate the direction of change in response to CTG.

## Figure 5: Mode of action of levosimendan and its active metabolite OR-1896.

Both Ca<sup>2+</sup>-sensitization and PDE3-inhibition at nanomolar concentrations ([nM]) contribute to their inotropic and lusitropic effects. Activation of mitochondrial K<sub>ATP</sub> (mitoK<sub>ATP</sub>) channels at micromolar concentrations ([µM]) may provide protection against ischemia/reperfusion. AR, adrenergic receptor; PDE, phosphodiesterase; cAMP, cyclic adenosine monophosphate; PKA, protein kinase A; SR, sarcomplasmic reticulum; SERCA, SR Ca<sup>2+</sup> ATPase; RyR, ryanodine receptor;  $I_{Ca}$  and  $I_{Na}$ , Ca<sup>2+</sup> and Na<sup>+</sup> currents; NCX; Na<sup>+</sup>/Ca<sup>2+</sup>- exchanger; NKA, Na<sup>+</sup>/K<sup>+</sup>-ATPase; ETC, electron transport chain; T-tubule, transversal tubule. Red arrows ( $\uparrow\downarrow$ ) indicate the direction of change in heart failure, while green arrows ( $\uparrow\downarrow$ ) indicate the direction induced by levosimendan.

#### Figure 6: Mechanism of action and effects of omecamtiv mecarbil.

**A**, The mechanochemical cycle of myosin. Yellow indicates myosin weakly bound to actin, while red indicates the myosin strongly bound to actin. Omecamtiv mecarbil (OM) accelerates the transition rate of myosin into the strongly actin-bound force-generating state. **B**, Representative tracings showing that OM (200 nM) increases the time and amplitude of myocyte shortening without any effect on the cytosolic Ca<sup>2+</sup> transient. In contrast, the β-AR agonist isoproterenol increases myocyte shortening and diastolic cell length (**C**) as well as time to peak and maximal relaxation velocity (**D**) in isolated rat cardiac myocytes in response to escalating concentrations of OM. **A-D** are from Malik et al. <sup>93</sup> with permission. **E**, Impact of OM (20 min infusion at a dose that prolonged SET by 20%) on LV pressure-volume loops in a pig model of myocardial stunning (termed "post-ischemic" heart). The volumes indicate LV stroke volume and end-diastolic volume, of which EF is calculated. Taken from Bakkehaug et al. <sup>96</sup> with permission. **F**, The impact of OM at 0.1 or 1 µM on normalized isometric force in response to increasing Ca<sup>2+</sup> concentrations

(decreasing pCa) in skinned rat cardiac myocytes. Taken from Nagy et al. <sup>99</sup> with permission.

Figure 7: Mechanisms of action of nitroxyl (HNO) in HF. HNO affects redoxsensitive residues of various proteins involved in myocyte Ca<sup>2+</sup> handling. In particular, HNO increases SERCA activity and sensitizes myofilaments to Ca<sup>2+</sup>. In concert, these properties increase SR Ca<sup>2+</sup> load, systolic Ca<sup>2+</sup> transients and contraction. Red arrows ( $\uparrow\downarrow$ ) indicate the direction of change in heart failure, while green arrows ( $\uparrow\downarrow$ ) indicate the direction induced by HNO.

**Figure 8:** Known and hypothesized bioenergetic consequences of inotropic interventions that either increase cytosolic Ca<sup>2+</sup> or myofilament Ca<sup>2+</sup> sensitivity. ECC, excitation-contraction coupling; ETC, electron transport chain; MCU, mitochondrial Ca<sup>2+</sup> uptake.