

Research Article

Sparfloxacin-Metal Complexes as Urease Inhibitors: Their Synthesis, Characterization, Antimicrobial, and Antienzymatic Evaluation

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Four new metal complexes (S12–S15) of SPFX (third-generation quinolones) via heavy metals have been synthesized in good yield and characterized by physicochemical and spectroscopic methods including TLC, IR, NMR, and elemental analyses. Sparfloxacinato ligand binds with metals through pyridone and oxygen atom of carboxylic group. The biological actives of complexes have been tested against four Gram-positive and seven Gram-negative bacteria and six different fungi. Statistical analysis of antimicrobial data was done by one-way ANOVA, Dunnett's test; it was observed that S13, S14, and S15 were found to be most active complexes. Antifungal data confirm that all four synthesized complexes are most active and show significant activity against *F. solani* with respect to parent drug and none of complexes show activity against *A. parasiticus*, *A. effuris*, and *S. cervicis*. To study inhibitory effects of newly formed complexes, enzyme inhibition studies have been conducted against urease, α -chymotrypsin, and carbonic anhydrase. Enzymatic activity results of these complexes indicated them to be good inhibitors of urease enzyme while all complexes show mild activities against carbonic anhydrase enzyme. Further research may prove the promising role of these synthesized complexes as urease inhibitors.

1. Introduction

For infectious diseases, multiple therapies are usually required and so the possibility of drug-drug interactions increased. Careful consideration of concomitant drug therapy is needed. Literature survey reveals that fluoroquinolones showed several important interactions with many drugs [1]. Usually fluoroquinolones are prescribed for many diseases including respiratory and urinary tract infections. Sparfloxacin (SPFX) is an orally active synthetically broad-spectrum third-generation quinolones use for upper respiratory tract infection. Metals are considered essential to a human body in performing physiologically important and vital functions, in the body [2]. The action of many drugs is dependent on coordination with metal ions or/and

the inhibition on the formation of metalloenzymes [3]. The proposed mechanism of the interaction is chelation between the 4-oxo and adjacent carboxyl group of quinolone and metal cations [4–8]. Literature survey reveals that concurrent administration of magnesium and aluminium containing antacid with ciprofloxacin resulted in a nearly complete loss of activity of the drug [9] and patients who orally administrated fluoroquinolones should avoid mixtures containing multivalent cations, because quinolones binds with these metals through chelation, in consequence formed metal complex in the gastric system [10]. Ma et al. [11] published norfloxacin interaction with aluminium, magnesium and calcium and Alkaysi et al. [12] compiled interaction of 16 metals with eight quinolones. Absorption of fluoroquinolones is manifestly reduced by antacids, calcium

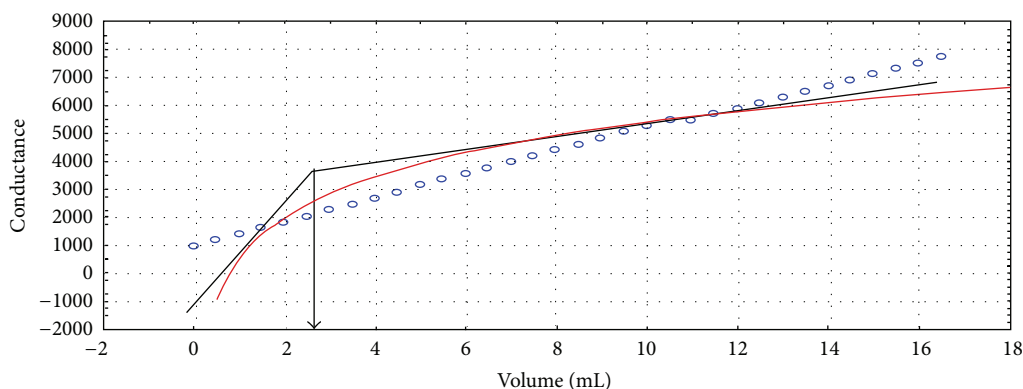


FIGURE 1: Representation of SPFX metal complexes ratio via conductance.

carbonate, ferrous sulphate, and sucralfate. Despite the fact that quantitative differences between fluoroquinolones exist, these combinations should be avoided whenever possible [7]. A reasonable recommendation may be to avoid using sucralfate and norfloxacin concurrently or avoid administration of norfloxacin and ciprofloxacin within two hours of sucralfate administration. Magnesium- and aluminum-containing antacids may also interfere with quinolones absorption. Survey assembled a number of different complexation of quinolones. Mononuclear dioxomolybdenum(VI) complexes with enrofloxacin and sparfloxacin were discovered by Efthimiadou and co-workers [13]. They also [14] discovered ciprofloxacin, cinoxacin, norfloxacin and nalidixic acid complexation with VO^{2+} , Mn^{2+} , Fe^{3+} , Co^{2+} , Ni^{2+} , Zn^{2+} , MoO^2 , Cd^{2+} , and UO^{2+} , vanadyl complex with enrofloxacin [15], and copper complex with sparfloxacin [16] Skyrianou et al. [17] reported nickel complex with sparfloxacin. Ciprofloxacin interaction with Mn^{2+} , Fe^{3+} , Co^{2+} , Ni^{2+} , and MoO^2 was presented by Psomas [18]. Alkaysi et al. [19] published norfloxacin interaction with aluminum, magnesium, and calcium and Turel [20] compiled interaction of 16 metals with eight quinolones. He also published ciprofloxacin complex with Cu(II) [21] and Ionic complexes of protonated norfloxacin with Zn(II) and Cu(II) [22]. Wallis and co-workers have reported complexes of ciprofloxacin with V(IV) O^{2+} , Fe(III) [23], and copper(II) [24]. Complexes of norfloxacin with Zn(II) and Cu(II) were prepared by Chen et al. [25] and complexes of ofloxacin with Cu(II) was discovered by Macías et al. [26], while Wang et al. [27] reported norfloxacin complex with Mn in 2002.

Interaction studies of SPFX metal complexes urged an idea of their synthesis [28]. Now, here we present synthesis of these complexes to aid in proving interaction studies. My research group has worked on this clinically important field of metal interaction and complexation for the last few years [29, 30]. We have already published metal complexes of SPFX as antifungal agents [9].

In this section, spectroscopic characterization of these novel neutral mononuclear metal complexes has been conducted with spectroscopic techniques such as IR, $^1\text{H-NMR}$, and elemental analyses (CHN). Prior to synthesis, M : L ratios were determined by conductance. The antimicrobial activity

TABLE 1: Physicochemical parameters of SPFX and SPFX-metal complexes.

S. no.	Complexes	Colour	M.P $^{\circ}\text{C}$	%Yield
1	SPFX	Bright yellow	260	—
2	S12	Yellow	170	45.43
3	S13	Light yellow	242	42
4	S14	Yellow	238–241	46
5	S15	Dark yellow	220	59.2

of these complexes has been evaluated against four Gram-positive and seven Gram-negative bacteria while antifungal activity against six different fungi has been determined also. Statistical analysis of antimicrobial data was done by one-way ANOVA, Dunnett's test. Enzyme inhibition studies have been conducted against urease, carbonic anhydrase, and α -chymotrypsin enzymes. Also physicochemical parameters have been recorded carefully.

2. Experimental

2.1. Materials and Reagents. Sparfloxacin was a kind gift by Abbott Pharmaceuticals (Karachi) while solvents and chemicals of analytical grade were purchased from the market. Metal salts ($\text{Al}(\text{OH})_3$, As_2O_3 , AgCl , and PbCO_3) were of pious grade from E. Merck. All solutions were prepared fresh before work.

2.2. Instruments. The melting points were taken on an electrothermal melting point apparatus (Gallenkamp) in open capillary tubes and are uncorrected. TLC spots were detected by UV lamp. Infrared spectra were recorded as KBr pellets on Shimadzu 470 instrument. ^1H NMR spectra were obtained by using Bruker/XWIN NMR spectrometer with TMS as internal standard. Complexes were dissolved in CDCl_3 , D_2O , or MeOD for NMR. An elemental analysis is done by Carlo Erba Strumentazione Elemental analyzer-MOD 1106 instrument.

2.3. Stoichiometric Study. Conductometric titration was performed to inspect the stoichiometric ratio of the ligand and

TABLE 2: FTIR absorption data of SPFX and its metal complexes (4000–400 cm⁻¹).

S. no.	Complexes	CH-F	Pri.NH	O-H stretching	C=O _{as} ,	C=O _s ,	Δ ^a
1	SPFX	1029	1441	3346–3468 ^c	1720 ^b	—	—
2	S12	1028	1442	3336	1636	1387	249
3	S13	1027	1440	3340	1641	1387	254
4	S14	1029	1442	3347	1636	1373	263
5	S15	1027	1437	3094	1639	1368	271

^aΔ = ν(CO₂)_{asym} - ν(CO₂)_{sym}, ^bν(COOH), ^cNH (str).

TABLE 3: ¹H-NMR data of SPFX and its metal complexes.

S. no.	Complexes	H-NMRδ: ppm
1	SPFX	0.53–0.28 (3H-cyclopropyl), 3.56–3.31 (m, 4H, piperazinyl ring protons), 4.0 (NH ₂), 7.96 (1H-phenyl), 8.51 (1H-quinolone), 11 (1H-OH, carbonyl).
2	S12	1.17–1.53 (5H-cyclopropyl), 3.04–3.31 (singlet piperazinyl ring), 3.91 (NH ₂), 7.24 (1H-phenyl), 8.62 (1H-quinolone).
3	S13	1.04–1.20 (5H-cyclopropyl), 3.29–3.31 (singlet piperazinyl ring), 3.92 (NH ₂), 6.44 (1H-phenyl), 8.62 (1H-quinolone).
4	S14	1.03–1.06 (3H-cyclopropyl), 3.20–3.31 (piperazinyl), 3.98 (NH ₂), 6.450–7.20 (phenyl), 8.46 (1H-CH ₂).
5	S15	1.17–1.21 (4H-cyclopropyl), 3.29–3.31 (singlet piperazinyl ring), 3.90 (NH ₂), 6.44 (1H-phenyl), 8.61 (1H-quinolone).

TABLE 4: Elemental analyses of the complexes.

Metal complexes	Found (calcd.) %	Found (calcd.) %	Found (calcd.) %
	C	H	N
SPFX	58.16 (57.09)	5.65 (5.53)	14.28 (14.06)
[Al(SPFX) ₂].3H ₂ O	53.24 (53.76)	5.96 (5.78)	12.74 (12.52)
[As(SPFX) ₂].2H ₂ O	53.55 (53.63)	5.76 (5.48)	12.81 (12.75)
[Ag(SPFX) ₂ (H ₂ O) ₂].Cl ₂ .2H ₂ O	49.72 (49.66)	4.92 (5.23)	11.89 (11.82)
[Pb(SPFX) ₂ .(H ₂ O) ₂].CO ₃	45.07 (45.28)	4.35 (4.42)	10.51 (10.88)

metal ions. For this purpose, 1 mM alcoholic solutions of drug (SPFX) and metal salts were prepared individually. In 20 mL of drug (SPFX) solution, 2 mL of metal solution was added each time; after every 2 min the conductance value was carefully noted. All the values of conductance were noted until state of chemical equilibrium is achieved. Graph was plotted between corrected conductivity and the volume of titrant added and the end point was determined. Results show that all complexes have stoichiometries of 2 : 1 (drug : metal). Figure 1 represents conductometric ratio.

2.4. Synthesis of Complexes. A warm methanolic, unimolar solution of metal salts was mixed with a bimolar solution of SPFX in methanol (1 : 2) in round bottomed flask and was refluxed for 4 h, above 80 °C on a water bath with constant stirring. The solution was filtered and the product left for slow evaporation and then crystallized at room temperature. After a few days, crystals deposited were collected, washed with methanol, and dried. % yield, color, melting points, and solubility of all the complexes were carried out in different solvents as water, methanol, chloroform, and dimethyl sulfoxide.

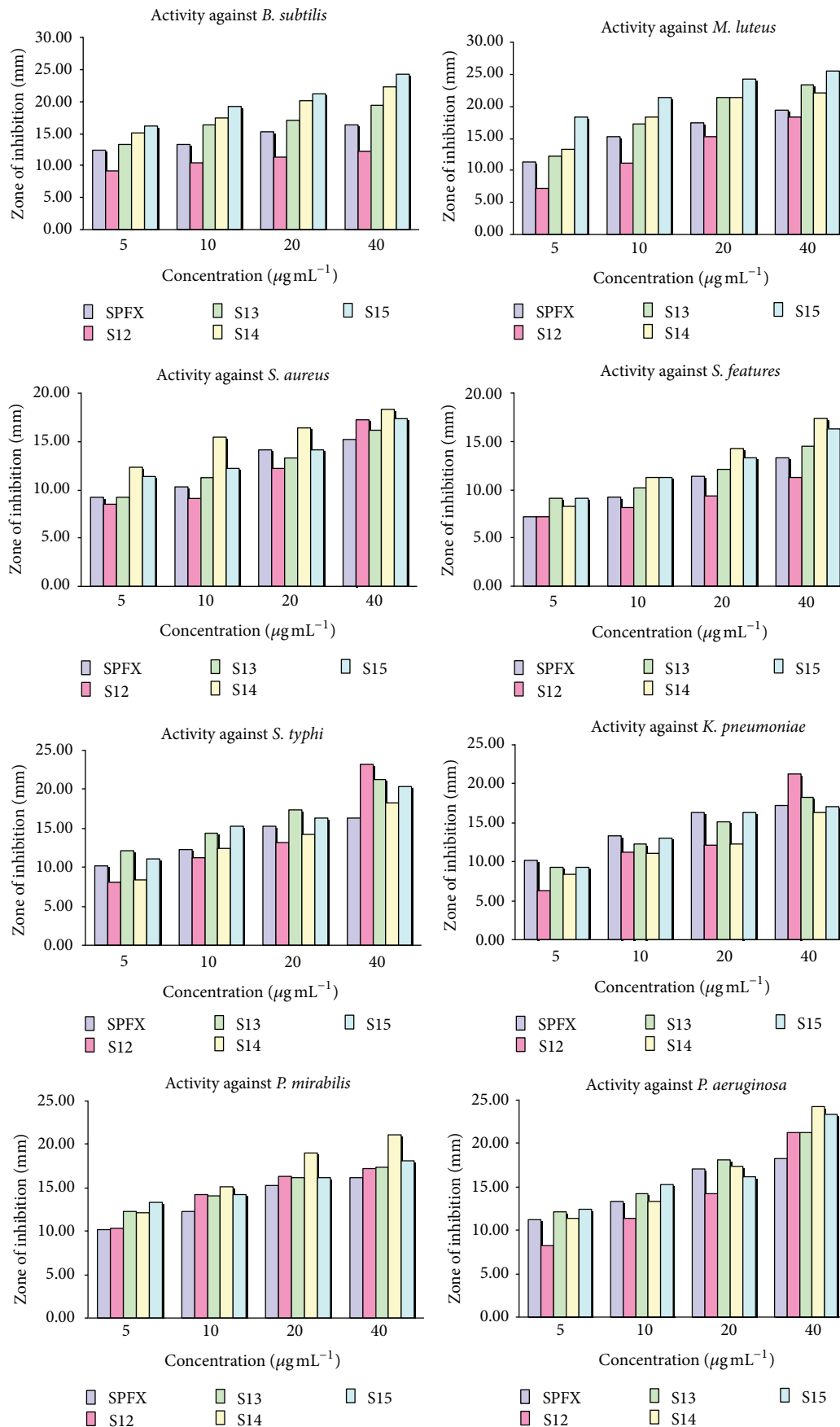
2.5. Antimicrobial Activity. For antibacterial and antifungal studies, disk susceptibility technique was used. The diffusion technique developed by Bauer et al. [31] recommended by

the FDA [32] has been adopted which has most extensively been used in the clinical laboratories [33].

2.6. Preparation of Dried Paper Disk. The stock solutions of standard drug (SPFX) and SPFX-metal complexes were prepared in water to get the concentration of 100 μg mL⁻¹ and diluted in four concentrations of 40, 20, 10, and 5 μg mL⁻¹. Three mm filter paper discs were impregnated with 20 mL of each of the different dilutions.

Discs were allowed to remain at room temperature till complete diluents evaporation and kept under refrigeration (ready to be used).

2.7. Procedure for Antimicrobial Activity. Organisms studied were taken from the slant with the help of wire loop and were immersed in the tube containing nutrient broth which was incubated at 37 °C for 4–6 hrs until the turbidity exceeded that of 0.5 MacFarland standards. Nutrient agar was prepared, autoclaved at 121 °C for 15, minutes then poured in dry, sterile Petri dishes, cooled, and set. The bacterial inoculum was uniformly spread using sterile cotton swab on a sterile Petri dish with agar. Discs soaked with metal complexes and derivatives were placed onto the surface of the agar with bacterial inoculum and sparfloxacin disk was used for control. These were then incubated at 36 °C ± 1 °C, for 24 h,



(a)

FIGURE 2: Continued.

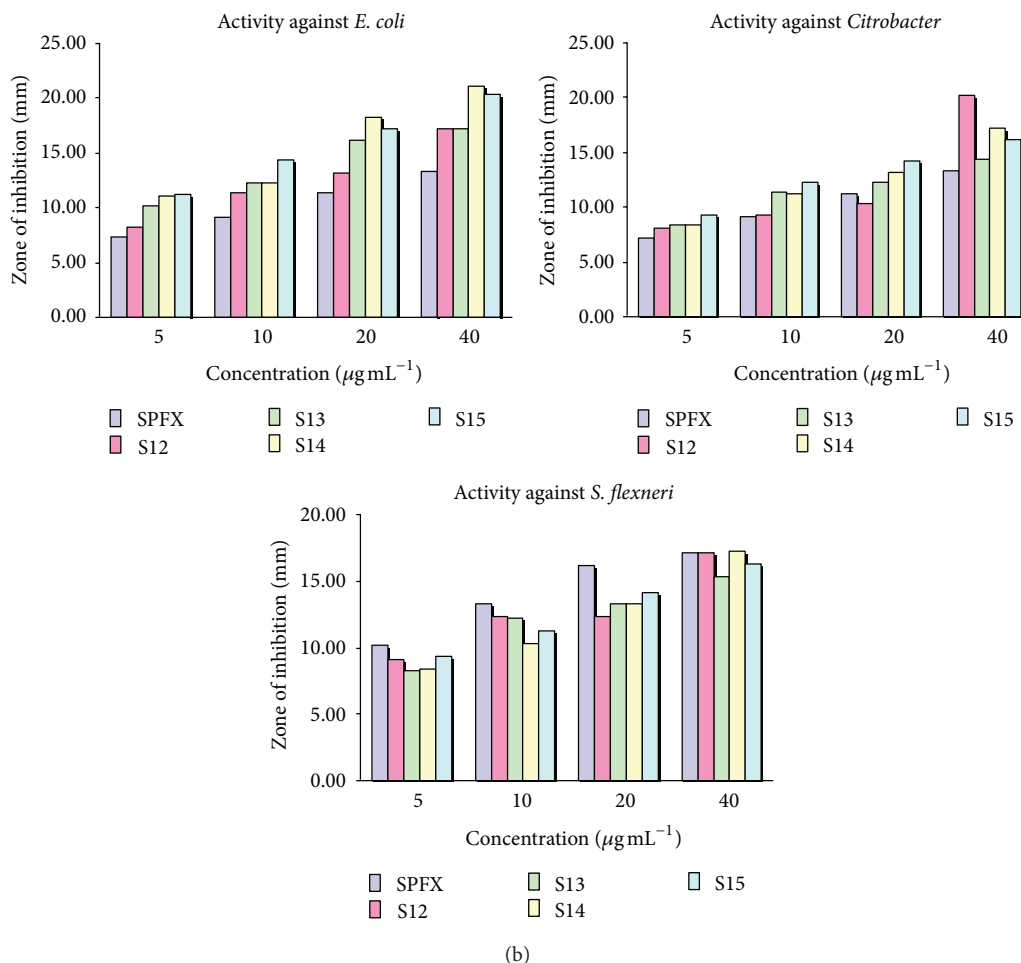


FIGURE 2: Graphical representation of inhibition zone against Gram-positive and Gram-negative bacteria.

while the water paper discs were used as a positive control. Three replicate trials were conducted against each organism for each concentration. Statistical analysis was used for data interpretation included calculation of the mean values, standard deviation, and investigation of significant differences in results. Similar procedures were adopted for antifungal activities. Derivative discs (5, 10, 20, and 40 $\mu\text{g mL}^{-1}$) were placed on SDS medium plates previously seeded with fungal culture and incubated for seven days at $36^\circ\text{C} \pm 1^\circ\text{C}$, for 48 hours. Zones of inhibition were carefully measured using Vernier caliper.

2.8. Statistical Study. Statistical analysis of antimicrobial data was done by one-way ANOVA, Dunnett's test through SPSS software version 10.0 (Carry, NC, USA).

3. Results and Discussion

3.1. Synthesis of SPFX-Metal Complexes with Heavy Metals. Four metal complexes were synthesized by refluxing metal salt solutions of $\text{Al}(\text{OH})_3$, As_2O_3 , AgCl , and Pb_2CO_3 in methanol with SPFX in the ratio of 1:2 [M:L] (determined

by conductance), for 4 hours, and the volume was reduced by evaporation. Moreover, their melting points and solubility were noted. Solubility facts of these complexes show that Al^{3+} and As^{3+} were soluble in CdCl_2 , Ag^+ was soluble in MeOH, and Pb^{2+} was soluble in both MeOH and CdCl_2 . Physico-chemical parameters of SPFX and SPFX-metal complexes are given in Table 1. The antimicrobial activity of these complexes has been evaluated against mentioned bacteria and fungi and analysis of data was done by one-way ANOVA. Enzyme inhibition studies have been conducted against the above-mentioned enzymes.

3.2. Proposed Structure of SPFX Metal Complexes. The coordination chemistry of some quinolones (including sparfloxacin) antibiotics with transition and d10 metal ions has been reported [34–37]. In this case, the SPFX has several potential donor sites but, due to steric hindrances, the ligand can provide a maximum of two donor atoms to any one metal centre. The spectroscopic changes suggested that the SPFX acts as a bidentate ligand and its coordination occurs through the metal via the pyridone and one carboxylate oxygen atom and forms slightly distorted octahedral geometry.

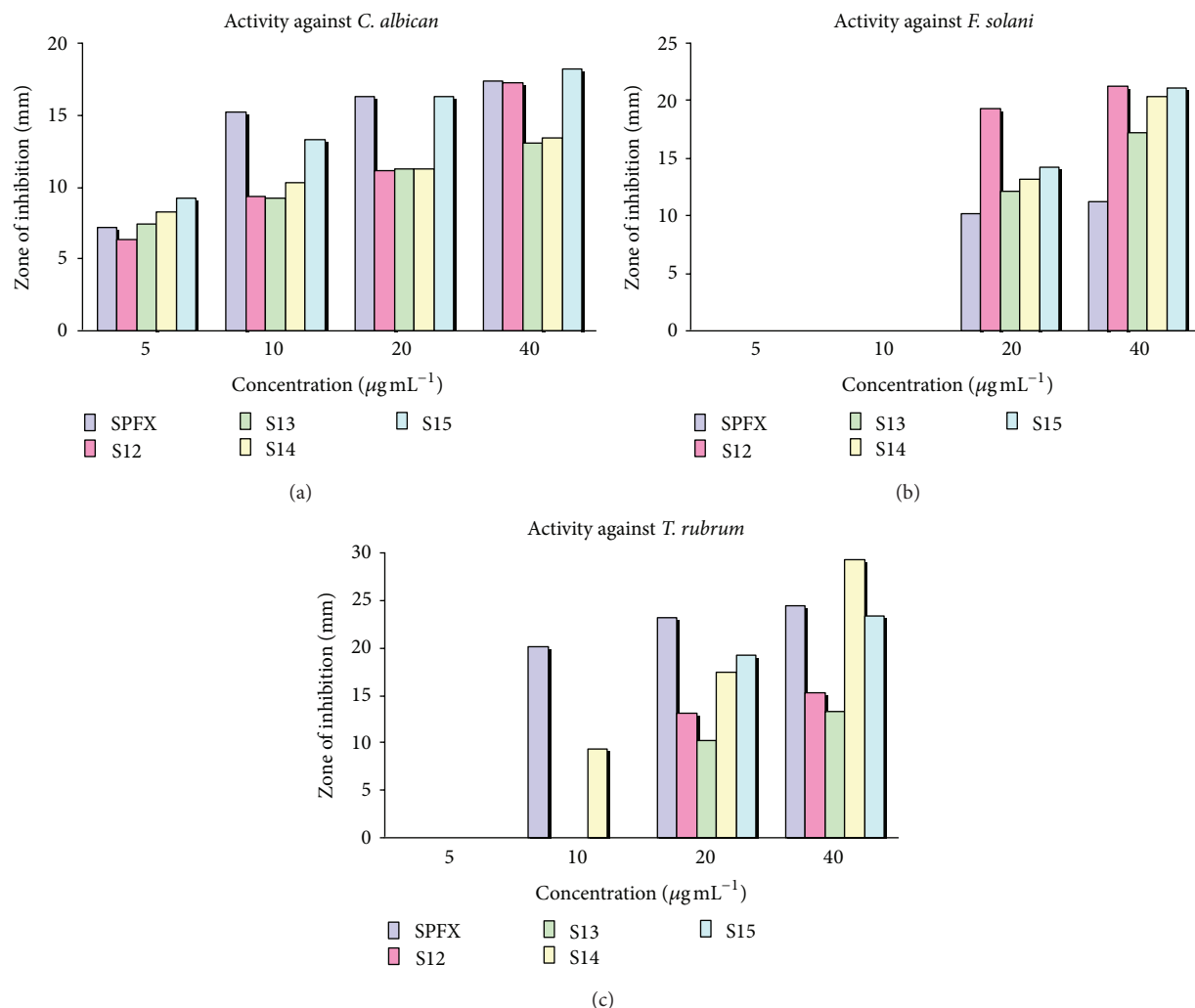


FIGURE 3: Graphical representation of inhibition zone against fungi.

3.3. Spectroscopic Studies

3.3.1. Infrared Analysis. IR spectra of SPFX_M.complex (S12–S15) revealed that the absorption at 1720 cm^{-1} observed in the spectrum of sparfloxacin, attributed to the $\nu(\text{C}=\text{O})_{\text{carb}}$, has been replaced with two very strong characteristic bands in the range of $1636\text{--}1641\text{ cm}^{-1}$ and $1368\text{--}1387\text{ cm}^{-1}$ assigned as $\nu(\text{O}-\text{C}-\text{O})$ asymmetric and symmetric stretching vibrations, respectively (Table 2) [31]. The $\Delta[\nu_{\text{asym}}(\text{CO}_2) - \nu_{\text{sym}}(\text{CO}_2)]$ values fall in the range of $249\text{--}271\text{ cm}^{-1}$ indicating a monodentate coordination mode of the carboxylato group via the pyridone and one carboxylato oxygen atom [7]. The vibration $\nu(\text{C}=\text{O})_{\text{p}}$, pyridone stretch is slightly shifted from 1641 to $1636\text{--}1641\text{ cm}^{-1}$ upon bonding [35]. Broad split band at $3094\text{--}3347\text{ cm}^{-1}$ can be assigned to the O–H stretching vibrations of water molecules and also includes the N–H stretching vibration of the piperazinyl moiety [38, 39]. New bands around $470\text{--}490\text{ cm}^{-1}$ seemed in the spectra of complexes can be assigned to $\nu(\text{M}-\text{O})$ [40].

3.3.2. ¹H NMR Analysis. The proton NMR spectrum of complexes showed a set of signals which were almost identical to those of SPFX, while changes occurred particularly at carboxylic protons as well as protons of aromatic C–NH. Singlet at δ 3.90–3.98 ppm was assigned to C–NH₂ group. The spectra showed multiplet at δ 1.03–1.53 ppm for cyclopropyl protons, multiplet at δ 3.04–3.31 ppm for iperazinyl protons, and singlet at δ 8.46–8.62 ppm for –CH₂ protons (Table 3). No broad weak band for acidic proton at δ : 11 ppm seen in spectra of complexes indicating that this targeted moiety took part in complexation of metals with SPFX and SPFX acts as bidentate deprotonated ligands bound to the metal through the pyridone oxygen and one carboxylate oxygen [9].

3.3.3. Elemental Analysis. The results obtained from elemental analysis CHN point toward that all of the complexes are formed from the reaction of the metal salt with drug in 1:2 molar ratio (Table 4).

TABLE 5: (a) Inhibition zones (mm) against *Bacillus subtilis*. (b) Inhibition zones (mm) against *Micrococcus luteus*. (c) Inhibition zones (mm) against *Staphylococcus aureus*. (d) Inhibition zones (mm) against *Streptococcus* features. (e) Inhibition zones (mm) against *Salmonella typhi*. (f) Inhibition zones (mm) against *Klebsiella pneumoniae*. (g) Inhibition zones (mm) against *Proteus mirabilis*. (h) Inhibition zones (mm) against *Pseudomonas aeruginosa*. (i) Inhibition zones (mm) against *Escherichia coli*. (j) Inhibition zones (mm) against *Citrobacter* species. (k) Inhibition zones (mm) against *Shigella flexneri*.

(a)

Complexes	Concentrations (μgmL^{-1})			
	5 (μgmL^{-1})	10 (μgmL^{-1})	20 (μgmL^{-1})	40 (μgmL^{-1})
SPFX	7.21 \pm 0.2	15.3 \pm 0.21	16.31 \pm 0.04	17.28 \pm 0.11
S12	8.09 \pm 0.36** (-12.21)	9.29 \pm 0.1** (39.28)	11.1 \pm 0.08** (31.94)	14.05 \pm 0.25** (18.69)
S13	8.84 \pm 0.16** (-22.61)	11.01 \pm 0.27** (28.04)	15.08 \pm 0.18** (7.54)	18.32 \pm 0.14** (-6.02)
S14	0 \pm 0** (100)	0 \pm 0** (100)	9.31 \pm 0.25** (42.92)	10.28 \pm 0.23** (40.51)
S15	6.97 \pm 0.07** (3.33)	10.16 \pm 0.2** (33.59)	12.21 \pm 0.12** (25.14)	14.37 \pm 0.06** (16.84)
ANOVA (df = 10, 22)	F-334.993 P < 0.001	F-44.427 P < 0.001	F-132.338 P < 0.001	F-123.338 P < 0.001

Mean \pm S.D (percent zone of inhibition), ** P < 0.005, * P < 0.05.

(b)

Complexes	Concentrations (μgmL^{-1})			
	5 (μgmL^{-1})	10 (μgmL^{-1})	20 (μgmL^{-1})	40 (μgmL^{-1})
SPFX	7.21 \pm 0.2	15.3 \pm 0.21	16.31 \pm 0.04	17.28 \pm 0.11
S12	8.09 \pm 0.36** (-12.21)	9.29 \pm 0.1** (39.28)	11.1 \pm 0.08** (31.94)	14.05 \pm 0.25** (18.69)
S13	8.84 \pm 0.16** (-22.61)	11.01 \pm 0.27** (28.04)	15.08 \pm 0.18** (7.54)	18.32 \pm 0.14** (-6.02)
S14	0 \pm 0** (100)	0 \pm 0** (100)	9.31 \pm 0.25** (42.92)	10.28 \pm 0.23** (40.51)
S15	6.97 \pm 0.07** (3.33)	10.16 \pm 0.2** (33.59)	12.21 \pm 0.12** (25.14)	14.37 \pm 0.06** (16.84)
ANOVA (df = 10, 22)	F-742.153 P < 0.001	F-192.257 P < 0.001	F-760.801 P < 0.001	F-467.658 P < 0.001

Mean \pm S.D (percent zone of inhibition), ** P < 0.005, * P < 0.05.

(c)

Complexes	Concentrations (μgmL^{-1})			
	5	10	20	40
SPFX	7.21 \pm 0.2	15.3 \pm 0.21	16.31 \pm 0.04	17.28 \pm 0.11
S12	8.09 \pm 0.36** (-12.21)	9.29 \pm 0.1** (39.28)	11.1 \pm 0.08** (31.94)	14.05 \pm 0.25** (18.69)
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S14	0 \pm 0** (100)	0 \pm 0** (100)	9.31 \pm 0.25** (42.92)	10.28 \pm 0.23** (40.51)
S15	6.97 \pm 0.07** (3.33)	10.16 \pm 0.2** (33.59)	12.21 \pm 0.12** (25.14)	14.37 \pm 0.06** (16.84)
ANOVA (df = 10, 22)	F-434.618 P < 0.001	F-200.157 P < 0.001	F-680.164 P < 0.001	F-326.531 P < 0.001

Mean \pm S.D (percent zone of inhibition), ** P < 0.005, * P < 0.05.

(d)

Complexes	Concentrations (μgmL^{-1})			
	5	10	20	40
SPFX	7.21 \pm 0.2	15.3 \pm 0.21	16.31 \pm 0.04	17.28 \pm 0.11
S12	8.09 \pm 0.36** (-12.21)	9.29 \pm 0.1** (39.28)	11.1 \pm 0.08** (31.94)	14.05 \pm 0.25** (18.69)
S13	8.84 \pm 0.16** (-22.61)	11.01 \pm 0.27** (28.04)	15.08 \pm 0.18** (7.54)	18.32 \pm 0.14** (-6.02)
S14	0 \pm 0** (100)	0 \pm 0** (100)	9.31 \pm 0.25** (42.92)	10.28 \pm 0.23** (40.51)
S15	6.97 \pm 0.07** (3.33)	10.16 \pm 0.2** (33.59)	12.21 \pm 0.12** (25.14)	14.37 \pm 0.06** (16.84)
ANOVA (df = 10, 22)	F-1304.177 P < 0.001	F-1543.733 P < 0.001	F-622.545 P < 0.001	F-420.443 P < 0.001

Mean \pm S.D (percent zone of inhibition), ** P < 0.005, * P < 0.05.

(e)

Complexes	Concentrations (μgmL^{-1})			
	5	10	20	40
SPFX	7.21 \pm 0.2	15.3 \pm 0.21	16.31 \pm 0.04	17.28 \pm 0.11

(e) Continued.

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
S12	$8.09 \pm 0.36^{**}$ (-12.21)	$9.29 \pm 0.1^{**}$ (39.28)	$11.1 \pm 0.08^{**}$ (31.94)	$14.05 \pm 0.25^{**}$ (18.69)
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Mean \pm S.D (percent zone of inhibition), $^{**}P < 0.005$, $^{*}P < 0.05$.

(f)

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
SPFX	7.21 ± 0.2	15.3 ± 0.21	16.31 ± 0.04	17.28 ± 0.11
S12	$8.09 \pm 0.36^{**}$ (-12.21)	$9.29 \pm 0.1^{**}$ (39.28)	$11.1 \pm 0.08^{**}$ (31.94)	$14.05 \pm 0.25^{**}$ (18.69)
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Mean \pm S.D (percent zone of inhibition), $^{**}P < 0.005$, $^{*}P < 0.05$.

(g)

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
SPFX	7.21 ± 0.2	15.3 ± 0.21	16.31 ± 0.04	17.28 ± 0.11
S12	$8.09 \pm 0.36^{**}$ (-12.21)	$9.29 \pm 0.1^{**}$ (39.28)	$11.1 \pm 0.08^{**}$ (31.94)	$14.05 \pm 0.25^{**}$ (18.69)
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S15	$6.97 \pm 0.07^{**}$ (3.33)	$10.16 \pm 0.2^{**}$ (33.59)	$12.21 \pm 0.12^{**}$ (25.14)	$14.37 \pm 0.06^{**}$ (16.84)
ANOVA (df = 10, 22)	F-1304.177 $P < 0.001$	F-1543.733 $P < 0.001$	F-622.545 $P < 0.001$	F-420.443 $P < 0.001$

Mean \pm S.D (percent zone of inhibition), $^{**}P < 0.005$, $^{*}P < 0.05$.

(h)

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
SPFX	7.21 ± 0.2	15.3 ± 0.21	16.31 ± 0.04	17.28 ± 0.11
S12	$8.09 \pm 0.36^{**}$ (-12.21)	$9.29 \pm 0.1^{**}$ (39.28)	$11.1 \pm 0.08^{**}$ (31.94)	$14.05 \pm 0.25^{**}$ (18.69)
S13	$8.84 \pm 0.16^{**}$ (-22.61)	$11.01 \pm 0.27^{**}$ (28.04)	$15.08 \pm 0.18^{**}$ (7.54)	$18.32 \pm 0.14^{**}$ (-6.02)
S14	$0 \pm 0^{**}$ (100)	$0 \pm 0^{**}$ (100)	$9.31 \pm 0.25^{**}$ (42.92)	$10.28 \pm 0.23^{**}$ (40.51)
S15	$6.97 \pm 0.07^{**}$ (3.33)	$10.16 \pm 0.2^{**}$ (33.59)	$12.21 \pm 0.12^{**}$ (25.14)	$14.37 \pm 0.06^{**}$ (16.84)
ANOVA (df = 10, 22)	F-1304.177 $P < 0.001$	F-1543.733 $P < 0.001$	F-622.545 $P < 0.001$	F-420.443 $P < 0.001$

Mean \pm S.D (percent zone of inhibition), $^{**}P < 0.005$, $^{*}P < 0.05$.

(i)

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
SPFX	7.21 ± 0.2	15.3 ± 0.21	16.31 ± 0.04	17.28 ± 0.11
S12	$8.09 \pm 0.36^{**}$ (-12.21)	$9.29 \pm 0.1^{**}$ (39.28)	$11.1 \pm 0.08^{**}$ (31.94)	$14.05 \pm 0.25^{**}$ (18.69)
S13	$8.84 \pm 0.16^{**}$ (-22.61)	$11.01 \pm 0.27^{**}$ (28.04)	$15.08 \pm 0.18^{**}$ (7.54)	$18.32 \pm 0.14^{**}$ (-6.02)
S14	$0 \pm 0^{**}$ (100)	$0 \pm 0^{**}$ (100)	$9.31 \pm 0.25^{**}$ (42.92)	$10.28 \pm 0.23^{**}$ (40.51)
S15	$6.97 \pm 0.07^{**}$ (3.33)	$10.16 \pm 0.2^{**}$ (33.59)	$12.21 \pm 0.12^{**}$ (25.14)	$14.37 \pm 0.06^{**}$ (16.84)

(i) Continued.

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
ANOVA (df = 10, 22)	F-1304.177 $P < 0.001$	F-1543.733 $P < 0.001$	F-622.545 $P < 0.001$	F-420.443 $P < 0.001$

Mean \pm S.D (percent zone of inhibition), ** $P < 0.005$, * $P < 0.05$.

(j)

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
SPFX	7.21 \pm 0.2	15.3 \pm 0.21	16.31 \pm 0.04	17.28 \pm 0.11
S12	8.09 \pm 0.36** (-12.21)	9.29 \pm 0.1** (39.28)	11.1 \pm 0.08** (31.94)	14.05 \pm 0.25** (18.69)
S13	8.84 \pm 0.16** (-22.61)	11.01 \pm 0.27** (28.04)	15.08 \pm 0.18** (7.54)	18.32 \pm 0.14** (-6.02)
S14	0 \pm 0** (100)	0 \pm 0** (100)	9.31 \pm 0.25** (42.92)	10.28 \pm 0.23** (40.51)
S15	6.97 \pm 0.07** (3.33)	10.16 \pm 0.2** (33.59)	12.21 \pm 0.12** (25.14)	14.37 \pm 0.06** (16.84)
ANOVA (df = 10, 22)	F-1304.177 $P < 0.001$	F-1543.733 $P < 0.001$	F-622.545 $P < 0.001$	F-420.443 $P < 0.001$

Mean \pm S.D (percent zone of inhibition), ** $P < 0.005$, * $P < 0.05$.

(k)

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
SPFX	7.21 \pm 0.2	15.3 \pm 0.21	16.31 \pm 0.04	17.28 \pm 0.11
S12	8.09 \pm 0.36** (-12.21)	9.29 \pm 0.1** (39.28)	11.1 \pm 0.08** (31.94)	14.05 \pm 0.25** (18.69)
S13	8.84 \pm 0.16** (-22.61)	11.01 \pm 0.27** (28.04)	15.08 \pm 0.18** (7.54)	18.32 \pm 0.14** (-6.02)
S14	0 \pm 0** (100)	0 \pm 0** (100)	9.31 \pm 0.25** (42.92)	10.28 \pm 0.23** (40.51)
S15	6.97 \pm 0.07** (3.33)	10.16 \pm 0.2** (33.59)	12.21 \pm 0.12** (25.14)	14.37 \pm 0.06** (16.84)
ANOVA (df = 10, 22)	F-1304.177 $P < 0.001$	F-1543.733 $P < 0.001$	F-622.545 $P < 0.001$	F-420.443 $P < 0.001$

Mean \pm S.D (percent zone of inhibition), ** $P < 0.005$, * $P < 0.05$.

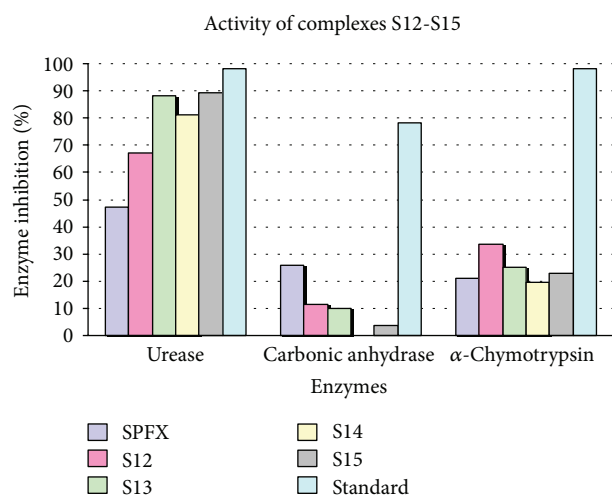


FIGURE 4: Graphical representation of enzymatic inhibition S12 to S15.

3.4. Antimicrobial Studies

3.4.1. Antibacterial Activity. Comparison of antibacterial activity data of novel SPFX metal complexes suggests that almost all complexes are active antimicrobial agents (Tables 5(a)–5(k), Figure 2) and most of them exhibit better activity

than parent drug. SPFX complexes including S13, S14, and S15 were found to be the most active complexes possessing higher antimicrobial activity against *B. subtilis* and *M. luteus* in all four tested concentrations. All synthesized complexes show moderate activity against *S. aureus* and *S. features* as compared to parent drug as well as other advance fluoroquinolones. Spectrum of Gram-negative activity indicated that all complexes show remarkable (excellent) activity against *P. aeruginosa*, *E. coli*, and *S. typh*e while S13, S14, and S15 exhibit good activity against *P. mirabilis* and citrobacter in comparison to SPFX and other standards, *M. luteus* and *S. typh*e. All complexes show almost same or less activity against *K. pneumoniae* and *S. flexneri*.

3.4.2. Antifungal Activity. These synthesized complexes were also evaluated for antifungal activity. The average results are shown in Tables 6(a)–6(c) (Figure 3). All of the complexes show excellent activity against *F. solani* as compared to the parent drug and other standards, while all synthesized complexes are less active against *T. rubrum* and more or less equally potent against *C. albican* in comparison to parentdrug. None of complexes show activity against *A. parasiticus*, *A. effuris*, and *S. cervicis*. In general, antifungal data and its statistical analysis confirm that all four synthesized complexes are most active and show significant activity against *F. solani* with respect to parent drug as well as other advance fluoroquinolones.

TABLE 6: (a) Inhibition zones (mm) against *C. albicans*. (b) Inhibition zones (mm) against *F. solani*. (c) Inhibition zones (mm) against *T. rubrum*.

(a)

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
SPFX	7.21 \pm 0.2	15.3 \pm 0.21	16.31 \pm 0.04	17.28 \pm 0.11
S12	8.09 \pm 0.36** (-12.21)	9.29 \pm 0.1** (39.28)	11.1 \pm 0.08** (31.94)	14.05 \pm 0.25** (18.69)
S13	8.84 \pm 0.16** (-22.61)	11.01 \pm 0.27** (28.04)	15.08 \pm 0.18** (7.54)	18.32 \pm 0.14** (-6.02)
S14	0 \pm 0** (100)	0 \pm 0** (100)	9.31 \pm 0.25** (42.92)	10.28 \pm 0.23** (40.51)
S15	6.97 \pm 0.07** (3.33)	10.16 \pm 0.2** (33.59)	12.21 \pm 0.12** (25.14)	14.37 \pm 0.06** (16.84)
ANOVA (df = 10, 22)	F-270.407 P < 0.001	F-26.145 P < 0.001	F-78.461 P < 0.001	F-52.516 P < 0.001

Mean \pm S.D (percent zone of inhibition), ** P < 0.005, * P < 0.05.

(b)

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
SPFX	0 \pm 0	0 \pm 0	10.2 \pm 0.04	11.12 \pm 0.07
S12	20.23 \pm 0.24** (0)	22.2 \pm 0.22** (0)	23.29 \pm 0.06** (-128.33)	27.06 \pm 0.2** (-143.35)
S13	22.06 \pm 0.06** (0)	24.06 \pm 0.15** (0)	28.22 \pm 0.09** (-176.67)	30.04 \pm 0.02** (-170.14)
S14	12.09 \pm 0.22** (0)	15.98 \pm 0.04** (0)	21.17 \pm 0.35** (-107.55)	24.24 \pm 0.06** (-117.99)
S15	17.26 \pm 0.14** (0)	22.03 \pm 0.27** (0)	26.08 \pm 0.16** (-155.69)	30.13 \pm 0.2** (-170.95)
ANOVA (df = 10, 22)	F-67.16 P < 0.001	F-124.599 P < 0.001	F-246.703 P < 0.001	F-133.773 P < 0.001

Mean \pm S.D (percent zone of inhibition), ** P < 0.005, * P < 0.05.

(c)

Complexes	Concentrations ($\mu\text{g mL}^{-1}$)			
	5	10	20	40
SPFX	0 \pm 0	20.4 \pm 0.07	23.07 \pm 0.05	24.29 \pm 0.13
S12	22.22 \pm 0.14** (0)	22.07 \pm 0.4** (-8.19)	24.24 \pm 0.15** (-5.07)	27.06 \pm 0.09** (-11.4)
S13	18.95 \pm 0.37** (0)	25.31 \pm 0.07** (-24.07)	26.27 \pm 0.27** (-13.87)	27.08 \pm 0.31** (-11.49)
S14	12.22 \pm 0.09** (0)	17.02 \pm 0.17** (16.57)	19.11 \pm 0.08** (17.17)	22.18 \pm 0.07** (8.69)
S15	18.24 \pm 0.1** (0)	21.32 \pm 0.17** (-4.51)	24.1 \pm 0.03** (-4.46)	27.25 \pm 0.19** (-12.19)
ANOVA (df = 10, 22)	F-252.324 P < 0.001	F-2663.194 P < 0.001	F-476.634 P < 0.001	F-315.709 P < 0.001

Mean \pm S.D (percent zone of inhibition), ** P < 0.005, * P < 0.05.

TABLE 7: Enzymatic activities of SPFX metal complexes.

Enzymes Compounds	Urease		Carbonic anhydrase		α -Chymotrypsin	
	Inhibition (%)	IC ₅₀ \pm SEM (μm)	Inhibition (%)	IC ₅₀ \pm SEM (μm)	Inhibition (%)	IC ₅₀ \pm SEM (μm)
SPAR	47.3	—	26.00	—	21.1	—
S12	67.20	128.53 \pm 0.11	11.30	—	33.5	—
S13	88.30	316.2 \pm 0.60	10.00	—	25.1	—
S14	81.20	169.96 \pm 0.26	—	—	19.6	—
S15	89.20	145.13 \pm 0.09	3.70	—	22.9	—
Standard	98.2	21.00 \pm 0.11	78.1	0.30 \pm 0.0006	98.1	5.70 \pm 0.13

3.5. Enzymatic Activity. To study inhibitory effects of newly formed complexes, enzyme inhibition studies have been conducted against urease, α -Chymotrypsin, and carbonic anhydrase (Table 7, Figure 4). Results indicated that all complexes exhibit very good activities against urease as compared to standard (thiourea), while all complexes show no or little activity against carbonic anhydrase using acetazolamide as reference standard.

4. Conclusion


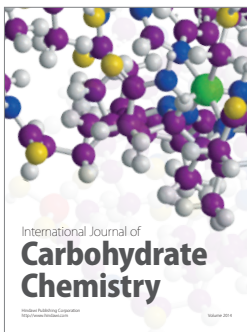
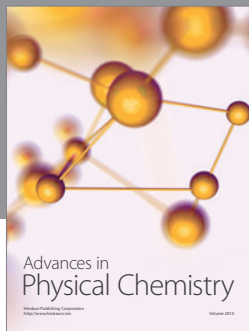
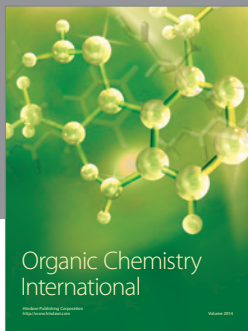
Metal complexes of SPFX via heavy metal have been synthesized in good yield and characterized by physicochemical and spectroscopic methods. Sparfloxacinato ligand binds with metals through pyridine and oxygen atom of carboxylic group. The biological activities of complexes have been tested against various bacteria and fungi; it was observed that S13,

S14, and S15 were found to be most active complexes and possess higher antimicrobial activity against *B. subtilis* and *M. luteus* in all four tested concentrations but less active than the parent drug, while all complexes show almost same or less activity against *K. pneumoniae* and *S. flexneri*. Antifungal data confirm that all four synthesized complexes are most active and show significant activity against *F. solani* with respect to parent drug as well as other advanced fluoroquinolones and none of complexes show activity against *A. parasiticus*, *A. effuris*, and *S. cervicis*. Enzymatic activity results of these complexes indicated them to be good inhibitors of urease enzyme while all complexes show mild activities against carbonic anhydrase enzyme. Further research may prove promising role of these synthesized complexes as urease inhibitors.

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