



The Effect of High Dose Isoflavone Supplementation on Serum Reverse T₃ in Euthyroid Men With Type 2 Diabetes and Post-menopausal Women

Thozhukat Sathyapalan¹, Josef Köhrle², Eddy Rijntjes², Alan S. Rigby¹,
Soha R. Dargham³, Eric S. Kilpatrick⁴ and Stephen L. Atkin^{3*}

¹ Academic Diabetes, Endocrinology and Metabolism, Hull York Medical School, University of Hull, Hull, United Kingdom, ² Institut für Experimentelle Endokrinologie, Charité-Universitätsmedizin Berlin, Berlin Institute of Health, CVK, Humboldt-Universität zu Berlin, Berlin, Germany, ³ Weill Cornell Medical College Qatar, Doha, Qatar, ⁴ Department of Clinical Chemistry, Sidra Medical and Research Center, Doha, Qatar

OPEN ACCESS

Edited by:

Alessandro Antonelli,
Università degli Studi di Pisa, Italy

Reviewed by:

Cheng Han,
Albert Einstein College of Medicine,
United States
Trevor Edmund Angell,
University of Southern California,
United States

*Correspondence:

Stephen L. Atkin
sla2002@qatar-med.cornell.edu

Specialty section:

This article was submitted to
Thyroid Endocrinology,
a section of the journal
Frontiers in Endocrinology

Received: 08 September 2018

Accepted: 06 November 2018

Published: 22 November 2018

Citation:

Sathyapalan T, Köhrle J, Rijntjes E,
Rigby AS, Dargham SR, Kilpatrick ES
and Atkin SL (2018) The Effect of High
Dose Isoflavone Supplementation on
Serum Reverse T₃ in Euthyroid Men
With Type 2 Diabetes and
Post-menopausal Women.
Front. Endocrinol. 9:698.
doi: 10.3389/fendo.2018.00698

Background: The health benefits of soy are widely reported but there are queries on the effect of soy isoflavones on thyroid function and the underlying mechanism of action.

Materials and Methods: We examined the effect of soy isoflavones on reverse tri-iodothyronine (or 3,3',5'-tri-iodothyronine; rT₃) in two studies comprising 400 patients: 200 men (study 1; 3 months) and 200 post-menopausal women (study 2; 6 months) who were randomized to consume 15 g soy protein with 66 mg of isoflavones (SPI) daily, or 15 g soy protein alone without isoflavones (SP) daily.

Results: SPI supplementation increased rT₃ serum concentration in both men 0.41 (0.12) vs. 0.45 (0.14) nmol/L and women 0.33 (0.12) vs. 0.37 (0.09) nmol/L at 3 months compared to SP that was not seen at 6 months. Thyroid stimulating hormone (TSH) serum concentrations increased while free thyroxine (fT₄) concentrations decreased with 3 months of SPI compared to SP supplementation for both men and women. rT₃ correlated with TSH in both studies ($p = 0.03$) but not with either fT₃ or fT₄. fT₃ levels did not differ between the SPI and SP preparations.

Conclusion: Soy isoflavones transiently increased rT₃ levels within 3 months though reverted to baseline at 6 months. The mechanism for this would be either rT₃ degrading deiodinase 1 and/or deiodinase 2 activities are transiently inhibited at 3 months, or inhibition of deiodinase 3, which generates rT₃ from T₄ is induced at 6 months. These changes were mirrored in the TSH concentrations, suggesting that short-term high dose isoflavone transiently impairs thyroid function in the first 3 months and may impact on general health during this period.

ISRCTN Registry: ISRCTN 90604927; ISRCTN34051237.

Keywords: soy, isoflavones, phytoestrogens, reverse tri-iodothyronine, tri-iodothyronine, thyroxine, TSH

INTRODUCTION

The consumption of soy food products have increased due to the reported potential health benefits that have been suggested to be due to the isoflavone components, leading to the development of isoflavone supplements and the fortification of foods with isoflavones (1, 2). It has been suggested that the soy isoflavones might provide protection against breast and prostate cancer (3–5), osteoporosis (6), cardiovascular diseases (7, 8) and alleviate hot flashes (9). However, there are concerns in susceptible individuals that soy may adversely affect thyroid function (10–14). The mechanism by which soy isoflavones may interfere with thyroid function is unclear, but it is critical to understand given the wide spread use of soy products. Animal studies have suggested that soy isoflavones interfere with thyroid function via thyroid peroxidase inhibition, as well as, with tissue deiodinase enzyme activities, which may affect extrathyroidal thyroid hormone metabolism, including rT₃ concentrations (15). Isoflavones might also displace thyroid hormones from their distributor proteins in the blood (16).

We have shown that high dose isoflavone intake, in comparison with isoflavone free soy, impair thyroid function in patients with type 2 diabetes (T2DM) (Study 1) (17) and also in post-menopausal healthy women (Study 2) (18). Study 1 was a randomized double-blind parallel study investigating the effect of soy isoflavones on testosterone serum concentrations in men with T2DM; 3 months of high dose isoflavone intake resulted in a significant increase in serum concentrations of thyrotropin (TSH) and a reduction of free thyroxine (fT₄) with no changes in serum free tri-iodothyronine (fT₃) concentration. Similarly, in study 2, a double-blind randomized parallel study investigating the effect of high dose isoflavone intake on bone turnover markers in women within 2 years of onset of menopause, 6 months of high dose isoflavone resulted in a significant increase in TSH and reduction of fT₄ with no changes in fT₃. As rT₃ is a major endogenous T₄ metabolite, probably devoid of major biological action in adults, we analyzed potential rT₃ concentration changes in serum, which might be harbingers of altered thyroid hormone distribution and metabolism (19). We conducted this *post-hoc* analysis to understand the impact of high dose isoflavones on rT₃ concentrations and their correlation with other thyroid measurement parameters tests including fT₃, fT₄ and TSH.

RESEARCH DESIGN AND METHODS

Study 1 involved 200 men aged between 45 and 75 years with T2DM, low early morning total testosterone concentrations (total testosterone less than the lower level of the reference range of 12 nmol/L) and normal gonadotropins who participated in a randomized double blind parallel study investigating the effect of soy isoflavones on serum testosterone concentrations (17). They were randomized either to intake of 15 g soy protein with 66 mg of isoflavones (SPI) per day or 15 g soy protein alone without any isoflavones (SP) per day for 3 months in the form of snack bars (**Figure 1A**). Study 2 involved 200 healthy women within 2 years of the onset of their menopause who were recruited

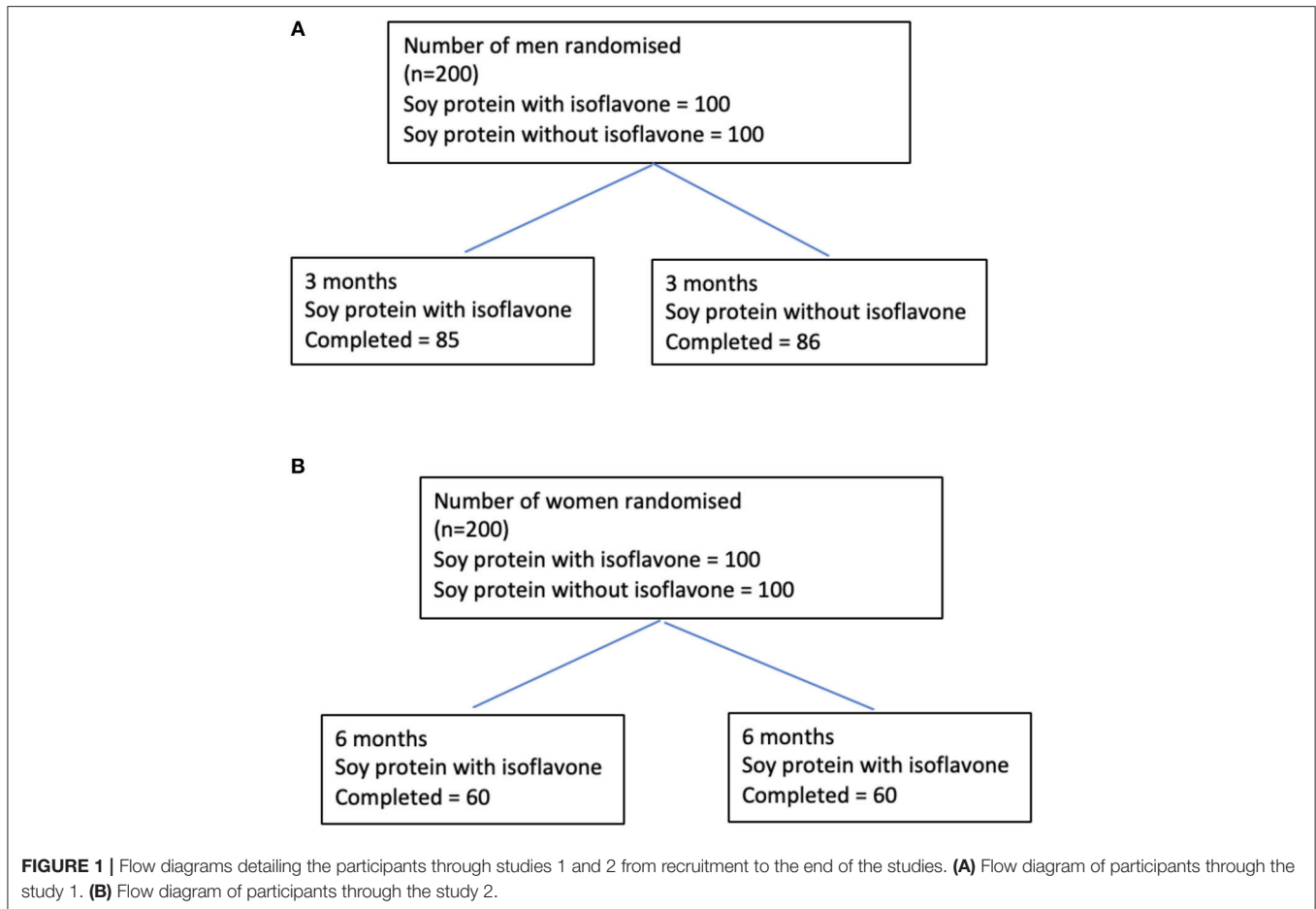
(20) to assess the impact of high dose soy isoflavones on bone turnover markers. They were randomized into either the SPI group (15 g soy protein with 66 mg of isoflavones) or SP group (15 g soy protein alone, isoflavone free) daily intake for a period of 6 months in the form of snack bars (**Figure 1B**). Thyroid function tests including fT₃, fT₄, and TSH were secondary outcomes for both studies.

Participants were required to avoid food products containing soy, alcohol, mineral, or vitamin supplementation. All patients were euthyroid and none of the patients were on thyroxine supplementation or on medications, which are known to affect thyroid function. The snack bars were consumed twice daily between meals. Dietary reinforcement together with measurement of plasma isoflavone concentrations was undertaken at each visit to ensure adherence. All subjects gave their written informed consent. Both studies received ethical approval by the Research Ethics Committee (East Yorkshire & North Lincolnshire Research Ethics Committee, ref: 09/H1304/45 and 09/1304/45). Computer-generated block randomization list was performed by Essential Nutrition Ltd., Brough, UK, who held the randomization codes for both the studies.

TSH, fT₃, and fT₄ assays were performed on an Abbott Architect i4000 immunoassay analyser (Abbott Diagnostics Division, UK) as batched samples. Analytical performance in terms of imprecision at discreet levels (% CV, Mean) for fT₄ was 4%, 13.7 pmol/L, for fT₃ was 11.2%, 0.33 nmol/L, and that of TSH was 4.2%, 1.32 mU/L. The rT₃ concentrations were measured in duplicate by competitive radioimmunoassay (total rT₃ RIA, lot R-EW-125-1614A; Radim Deutschland GmbH, Freiburg, Germany) according to the manufacturer's instructions as a batched analysis. The assay range was 0.04–3.10 nmol/L and inter-assay %CV was <11.2%. The isoflavones were extracted and analyzed from serum by LGC, Fordham, Cambridgeshire, UK using isotope-dilution LC-MS/MS (21). LC-MS/MS was conducted using a Sciex4000Qtrap with separation achieved using a C18 column and mobile phases of acetonitrile and water, both containing acetic acid (17).

Study Product

“The intervention consisted of a snack bar containing 7.5 g isolated soy protein powder (Solcon F, Solbar Industries, Israel) with 33 mg of isoflavones (SPI) (Solgen 40, Solbar Industries, Israel) given twice daily between meals (15 g soy protein and 66 mg of isoflavones per day), or 7.5 g of the isolated soy protein given twice daily (15 g soy protein per day without isoflavones per day) as control (SP). The latter had an isoflavone concentration of <300 parts per billion following serial alcohol extraction by Dishman Ltd., India (20); and product isoflavones assayed by FERA, Sand Hutton, UK (20). Analysis showed the composition of the dose materials to be 12% glycitein, 35% daidzein, and 54% genistein as aglycones. 90% of isoflavones were in the primary glucoside form, with the remaining 10% as aglycones as malonyl and acetyl glucosides. The snack bars were consumed twice daily between meals for 6 months. The study bars were prepared and packaged by Halo foods, Swindon, UK.” Soy bars were identical and



had similar macronutrient content and a tasting panel had determined that there was no discernible difference between the 2 products.

Statistical Analysis

This is a *post-hoc* exploratory analysis of two studies to understand the impact of high dose isoflavones on rT₃ concentrations and their correlation with other serum thyroid hormone measurement parameters tests including fT₃, fT₄, and TSH hence apriori power calculation was not done for changes in rT₃.

Baseline continuously distributed data is presented as median (25/75th centiles); categorical data by *n* (%). Within-group differences (difference between 12 weeks/24 weeks values and baseline values) are shown for each treatment group separately by a mean and a standard deviation (SD). Between-group comparisons were performed using the independent sample *t*-test. The *t*-test assumes equal variance between groups. Changes of rT₃ concentrations (Δ rT₃) with each supplementation was correlated with changes in fT₃ (Δ fT₃), changes in fT₄ (Δ fT₄) and changes in TSH (Δ TSH) by Spearman correlation coefficient method. For all statistical analyses, a two-tailed *P* < 0.05 was considered to indicate statistical significance. Statistical analysis was performed using the STATA statistical computer

package (StataCorp 2013. *Stata Statistical Software*: StataCorp LP, USA).

RESULTS

In Study 1, there was a significant increase in rT₃ with SPI supplementation [0.41 (0.12) vs. 0.45 (0.14) nmol/L] at 3 months compared to SP supplementation [0.43 (0.10) vs. 0.40 (0.15) *p*-value < 0.001] (Tables 1, 2) in men with type 2 diabetes. SPI supplementation increased TSH within 3 months [Mean (SD)] [1.81 (0.92) vs. 3.23 (1.03) mU/L] compared to SP supplementation [1.82 (0.93) vs. 1.96 (1.11) mU/L]. Conversely, SPI supplementation decreased free T₄ within 3 months [12.68 (1.90) vs. 11.09 (2.00) pmol/L] compared to SP supplementation [13.06 (1.74) vs. 12.74 (1.62) pmol/L]. There were no changes in fT₃ with 3 months of either SPI or SP supplementation. There was a significant correlation between changes in rT₃ with TSH (*r* = 0.52; *p* = 0.03) but no correlation with changes in fT₃ (*r* = 0.18; *p* = 0.81) and fT₄ (*r* = 0.13; *p* = 0.62).

In Study 2 in healthy women within 2 years of menopause (Table 3), SPI supplementation increased rT₃ [0.33 (0.12) vs. 0.37 (0.09) nmol/L] compared to SP supplementation (*p* < 0.001) within 3 months. The rT₃ decreased after 6 months 0.31 (0.13)

TABLE 1 | Changes in reverse T₃ at 3 months for study 1 and at 3 and 6 months for study 2.

Changes in reverse T ₃ (rT ₃)						
	Baseline	3 months	p-value (baseline vs. 3 months between groups)	6 months	p-value (baseline vs. 6 months between groups)	p-value (3 vs. 6 months between groups)
STUDY 1						
SPI	0.41 (0.12)	0.45 (0.14)	<0.001			
SP	0.43 (0.10)	0.40 (0.15)				
STUDY 2						
SPI	0.33 (0.12)	0.37 (0.09)	<0.001	0.31 (0.13)	0.81	0.001
SP	0.33 (0.11)	0.33 (0.12)		0.30 (0.12)		

SPI, 15 g soy protein with 66mg of isoflavones; SP, 15 g soy protein alone isoflavone free. Results in Mean (SD).

TABLE 2 | Baseline characteristics of study 1 participants.

Parameters	Soy protein without isoflavone	Soy protein with isoflavone
Body mass index (kg/m ²)	31.6 (29.2, 35.0)	31.8 (28.8, 34.7)
Age (years)	52.0 (50.0, 55.0)	52.0 (50.0, 55.0)
HbA1c (mmol/mol)	58 (53, 64)	56 (52, 60)
Duration of diabetes (years)	7.9 (4.4, 9.1)	7.3 (4.2, 8.8)
HbA1c (mmol/mol)	58 (53, 64)	56 (52, 60)
fT ₄ (pmol/L)	13.0 (12.0, 14.0)	12.0 (12.0, 14.0)
fT ₃ (pmol/L)	4.6 (4.2, 4.9)	4.6 (4.2, 5.0)
TSH (mU/L)	1.6 (1.2, 2.5)	1.6 (1.2, 2.4)

Data are provided as medians (25/75th centiles).

fT₄, free thyroxine; fT₃, free tri-iodo thyronine; TSH, thyroid stimulating hormone.

TABLE 3 | Baseline characteristics of study 2 participants.

Parameters	Soy protein without isoflavone	Soy protein with isoflavone
Body mass index (kg/m ²)	24.6 (22.7, 28.4)	26.3 (24.3, 30.7)
Age (years)	52 (50, 55)	52 (49, 56)
fT ₄ (pmol/L)	13 (12, 14)	13 (13, 15)
fT ₃ (pmol/L)	4.7 (4.3, 4.9)	4.6 (4.3, 5.1)
TSH (mU/L)	1.6 (0.9, 2.3)	1.5 (0.9, 2.2)

Data are provided as medians (25/75th centiles). fT₄, free thyroxine; fT₃, free tri-iodo thyronine; TSH, thyroid stimulating hormone.

of SPI supplementation and was comparable to baseline (p -value = 0.81). Mean TSH increased significantly [mean (SD) 1.58 (0.93) vs. 2.61 (1.24) mU/L, $p < 0.01$] with a corresponding reduction in mean fT₄ [13.5 (2.2) vs. 11.2 (1.8) pmol/L, $p < 0.01$] from baseline to 3 months. There was a significant correlation between changes in rT₃ with TSH ($r = 0.612$; $p = 0.02$) but no correlation with changes in fT₃ ($r = 0.22$; p -value = 0.42) or fT₄ ($r = 0.18$; p -value = 0.38) at 3 months. There was no correlation between changes in rT₃ with changes in TSH, fT₄ and fT₃ at 6 months.

There were no changes in TSH and fT₄ between 3 and 6 months. There were no differences in the fT₃ between both preparations.

DISCUSSION

There was a significant increase in rT₃ in both studies after 3 months of high dose isoflavone supplementation, whereas the rT₃ values did not differ in isoflavone-free soy, suggesting that it is the isoflavone component that is responsible for the rT₃ changes seen. However, in the study 2 involving post-menopausal women the rT₃ decreased to baseline values after 6 months of SPI supplementation, suggesting that the isoflavone induced changes are transient and return to normal pretreatment thyroid homeostasis over a 6-months period.

In both studies, there was a reduction in fT₄ and a corresponding rise in TSH, suggesting that the feedback response of the hypothalamo-pituitary-thyroid axis was intact. In situations where the thyroid cannot maintain thyroid hormone production, either due to an autoimmune process and/or lack of the essential trace element iodine, there would be a shift in thyroidal production from a 20:1 T₄:T₃ ratio to a ratio more in favor of T₃. A preferential production and secretion of T₃ compared to T₄ could potentially explain why there were no significant changes in fT₃ despite changes in TSH and fT₄. Dose and duration of SPI consumption might not yet be sufficient to also decrease serum fT₃ considering that the majority of T₃ is generated outside of the thyroid gland by Type 1 and Type II 5'-deiodinase activities. Furthermore, SPI isoflavones might enhance (hepatic and/or gastrointestinal) T₄ elimination by hepatic enzyme induction, as frequently observed for drugs (e.g., phenobarbital) and xenobiotics, while serum T₃ concentration are still maintained (22–24). However, given the uniform response of the increase in rT₃ in response to high dose isoflavones, this would suggest that this change is not idiosyncratic. Interestingly, changes in serum rT₃ in the SPI isoflavone consuming groups were similar in T2DM male patients and post-menopausal women, which in our opinion would exclude

effects solely restricted to T2DM patients or post-menopausal women.

Serum T₄ is metabolized either to the active thyroid hormone T₃ or to the inactive rT₃ in a reciprocal manner depending upon the relative actions of the tissue deiodinase enzymes types 1 to 3. The mechanism of the increase in rT₃ seen in both studies is unclear. One of the main sources of rT₃ is the peripheral conversion of thyroxine to rT₃. The enzyme responsible for this is deiodinase type 3 and it could be hypothesized that isoflavones may activate deiodinase type 3 (25), but no data has been reported on direct stimulation of expression of any of the deiodinases by isoflavones. Degradation of rT₃ is mainly accomplished by hepatic and renal deiodinase type 1, but also by deiodinase type 2 (25). Therefore, another plausible explanation would be that isoflavones (transiently) inhibit hepatic deiodinase 1 (and/or extrahepatic deiodinase 2) (25), which would lead to accumulation of rT₃ in blood. However, in rat-studies we have shown that deiodinase type 1 activity in the liver is increased after 16 weeks of isoflavone treatment (15), which might be due to higher hepatic uptake and exposure to thyroid hormone which may be displaced by flavonoids from its binding to transthyretin (16, 26). Administration of high concentrations of rT₃, administered in rodent experiments, did not affect serum TSH and thyroid hormones, albeit type 2 deiodinase activity was inhibited and hepatic gene expression was affected (27). The short half-life and rapid turnover of rT₃ might result in these transient changes and flavonoid effects on deiodinase isoenzymes and transthyretin might mainly manifest as variations in rT₃ serum concentrations which is only weakly bound to serum distribution proteins compared to T₄ and T₃. Whether the thyroid itself is also affected by isoflavones and/or responds to altered TSH concentration, which might induce activity and expression of type 1 and type 2 deiodinases and thus maintain T₃ concentration, requires detailed kinetic studies (28). There may be multiple mechanisms through which isoflavones act on thyroid hormone metabolism, distribution and/or transport systems with a complexity that needs further elucidation. It is also unknown if the isoflavone effects would be different between patients with and without autoimmune thyroid disease.

Reduced circulating levels of fT₃ are a sensitive marker of ill health especially in context of elevated rT₃ (29). In a population of elderly men, who were independently living, serum rT₃ concentrations increased with age and the presence of comorbidities (29). Higher serum rT₃ concentrations may result from decreased peripheral metabolism of TH due to the aging process itself and/or disease and may reflect a catabolic state (29). Indeed, fasting, cold exposure, and even minor infective or inflammatory disease are sufficient to reduce serum fT₃ and

elevate rT₃ concentrations in otherwise-healthy individuals (30, 31).

In a cohort of elderly people, baseline serum rT₃ concentration was associated with all-cause mortality during a 9-years period of follow up, suggesting that rT₃ may be a more sensitive marker for non-thyroidal illness than fT₃ (32). No significant mortality associations were found with serum fT₃, fT₄ or TSH concentrations. However, in men lower serum TSH, and in women higher rT₃ concentrations, predicted disability (32). These findings broadly confirm those of previous studied cohorts of older people (33, 34), and are in line with the known pathophysiological mechanisms whereby low fT₃ and higher rT₃ concentrations are non-specifically associated with increasing ill health and disease burden (31). In the elderly, high rT₃ is a stronger predictor of all-cause mortality than low fT₃ independent of disease burden (32).

Dietary intake of isoflavones in Asian soy diets has been estimated to be in the range of 30–50 mg per day of combined isoflavone aglycone equivalents (35, 36). In Western countries an average daily intake of ~2 mg isoflavones is seen though estimated to be 16 mg in vegetarians (37); therefore, the dose of 66 mg of isoflavones used in this study may be considered to be in the pharmacological range for this study purpose.

In conclusion, these studies show that the isoflavone component of soy protein transiently increases rT₃ concentrations when supplemented over a period of 3 months but afterwards rT₃ concentrations reverted to baseline at 6 months, perhaps due to deiodinase 3 inhibition. These changes were mirrored in the TSH values, suggesting that high dose isoflavones may transiently impair thyroid homeostasis, though it is not clear if this would impact clinically on general health.

AUTHOR CONTRIBUTIONS

TS, EK, and SA devised the study. JK and ER measured the rT₃ whilst. AR and SD performed the statistical analysis. All authors contributed to the writing and final review of the manuscript.

FUNDING

This study was supported by the Food Standards Agency, United Kingdom (T01057 and T01060). The sponsors did not influence the study design; the collection, analysis, and interpretation of data; writing of the report; and decision to submit the paper for publication. Any views or opinions expressed do not necessarily represent those of the FSA and are solely those of the authors. The publication of this article was funded by the Qatar National Library.

REFERENCES

- Nurmi T, Mazur W, Heinonen S, Kokkonen J, Adlercreutz H. Isoflavone content of the soy based supplements. *J Pharm Biomed Anal.* (2002) 28:1–11. doi: 10.1016/S0731-7085(01)00612-4
- Setchell KD. Soy isoflavones—Benefits and risks from nature's selective estrogen receptor modulators (SERMs). *J Am Coll Nutr.* (2001) 20 (Suppl. 5):354S–62S. discussion: 81S–83S. doi: 10.1080/07315724.2001.10719168
- Yamamoto S, Sobue T, Kobayashi M, Sasaki S, Tsugane S. Soy, isoflavones, and breast cancer risk in Japan. *J Nat Cancer Inst.* (2003) 95:906–13. doi: 10.1093/jnci/95.12.906
- Messina MJ. Emerging evidence on the role of soy in reducing prostate cancer risk. *Nutr Rev.* (2003) 61:117–31. doi: 10.1301/nr.2003.apr.117-131
- Lamartiniere CA, Cotroneo MS, Fritz WA, Wang J, Mentor-Marcel R, Elgavish A. Genistein chemoprevention: timing and mechanisms of

- action in murine mammary and prostate. *J Nutr.* (2002) 132:552S–8S. doi: 10.1093/jn/132.3.552S
6. Messina M, Ho S, Alekel DL. Skeletal benefits of soy isoflavones: a review of the clinical trial and epidemiologic data. *Curr Opin Clin Nutr Metab Care* (2004) 7:649–58. doi: 10.1097/00075197-200411000-00010
 7. Weggemans RM, Trautwein EA. Relation between soy-associated isoflavones and LDL and HDL cholesterol concentrations in humans: a meta-analysis. *Eur J Clin Nutr.* (2003) 57:940–6. doi: 10.1038/sj.ejcn.1601628
 8. Nestel P. Isoflavones: their effects on cardiovascular risk and functions. *Curr Opin Lipidol.* (2003) 14:3–8. doi: 10.1097/00041433-200302000-00002
 9. Messina M, Hughes C. Efficacy of soyfoods and soybean isoflavone supplements for alleviating menopausal symptoms is positively related to initial hot flush frequency. *J Med Food* (2003) 6:1–11. doi: 10.1089/109662003765184697
 10. Fitzpatrick M. Soy formulas and the effects of isoflavones on the thyroid. *N Z Med J.* (2000) 113:24–6.
 11. Doerge DR, Sheehan DM. Goitrogenic and estrogenic activity of soy isoflavones. *Environ Health Perspect.* (2002) 110 (Suppl. 3):349–53. doi: 10.1289/ehp.02110s3349
 12. Sathyapalan T, Manuchehri AM, Thatcher NJ, Rigby AS, Chapman T, Kilpatrick ES, et al. The effect of soy phytoestrogen supplementation on thyroid status and cardiovascular risk markers in patients with subclinical hypothyroidism: a randomized, double-blind, crossover study. *J Clin Endocrinol Metab.* (2011) 96:1442–9. doi: 10.1210/jc.2010-2255
 13. Messina M, Redmond G. Effects of soy protein and soybean isoflavones on thyroid function in healthy adults and hypothyroid patients: a review of the relevant literature. *Thyroid* (2006) 16:249–58. doi: 10.1089/thy.2006.16.249
 14. Huser S, Guth S, Joost HG, Soukup ST, Kohrle J, Kreienbrock L, et al. Effects of isoflavones on breast tissue and the thyroid hormone system in humans: a comprehensive safety evaluation. *Arch Toxicol.* (2018) 92:2703–48. doi: 10.1007/s00204-018-2279-8
 15. Susic-Jurjevic B, Filipovic B, Wirth EK, Zivanovic J, Radulovic N, Jankovic S, et al. Soy isoflavones interfere with thyroid hormone homeostasis in orchidectomized middle-aged rats. *Toxicol Appl Pharmacol.* (2014) 278:124–34. doi: 10.1016/j.taap.2014.04.018
 16. Radovic B, Mentrup B, Kohrle J. Genistein and other soya isoflavones are potent ligands for transthyretin in serum and cerebrospinal fluid. *Br J Nutr.* (2006) 95:1171–6. doi: 10.1079/BJN20061779
 17. Sathyapalan T, Rigby AS, Bhasin S, Thatcher NJ, Kilpatrick ES, Atkin SL. Effect of soy in men with type 2 diabetes mellitus and subclinical hypogonadism: a randomized controlled study. *J Clin Endocrinol Metab.* (2017) 102:425–33. doi: 10.1210/jc.2016-2875
 18. Sathyapalan T, Aye M, Rigby AS, Fraser WD, Thatcher NJ, Kilpatrick ES, et al. Soy reduces bone turnover markers in women during early menopause: a randomized controlled trial. *J Bone Miner Res.* (2017) 32:157–64. doi: 10.1002/jbmr.2927
 19. Schmidt RL, LoPresti JS, McDermott MT, Zick SM, Straseski JA. Does reverse triiodothyronine testing have clinical utility? An analysis of practice variation based on order data from a National Reference Laboratory. *Thyroid* (2018) 28:842–8. doi: 10.1089/thy.2017.0645
 20. Sathyapalan T, Aye M, Rigby AS, Fraser WD, Thatcher NJ, Kilpatrick ES, et al. Soy reduces bone turnover markers in women during early menopause: a randomized controlled trial. *J Bone Miner Res.* (2016) 32:157–64. doi: 10.1002/jbmr.292
 21. Grace PB, Mistry NS, Carter MH, Leatham AJ, Teale P. High throughput quantification of phytoestrogens in human urine and serum using liquid chromatography/tandem mass spectrometry (LC-MS/MS). *J Chromatogr B Analyt Technol Biomed Life Sci.* (2007) 853:138–46. doi: 10.1016/j.jchromb.2007.03.011
 22. Zabka TS, Fielden MR, Garrido R, Tao J, Fretland AJ, Fretland JL, et al. Characterization of xenobiotic-induced hepatocellular enzyme induction in rats: anticipated thyroid effects and unique pituitary gland findings. *Toxicol Pathol.* (2011) 39:664–77. doi: 10.1177/0192623311406934
 23. Capen CC. Mechanisms of chemical injury of thyroid gland. *Prog Clin Biol Res.* (1994) 387:173–91.
 24. Cavalieri RR, Pitt-Rivers R. The effects of drugs on the distribution and metabolism of thyroid hormones. *Pharmacol Rev.* (1981) 33:55–80.
 25. Bianco AC, Kim BW. Deiodinases: implications of the local control of thyroid hormone action. *J Clin Invest.* (2006) 116:2571–9. doi: 10.1172/JCI29812
 26. Susic-Jurjevic B, Lutjohann D, Jaric I, Miler M, Vojnovic Milutinovic D, Filipovic B, et al. Effects of age and soybean isoflavones on hepatic cholesterol metabolism and thyroid hormone availability in acyclic female rats. *Exp Gerontol.* (2017) 92:74–81. doi: 10.1016/j.exger.2017.03.016
 27. Cettour-Rose P, Visser TJ, Burger AG, Rohner-Jeanrenaud F. Inhibition of pituitary type 2 deiodinase by reverse triiodothyronine does not alter thyroxine-induced inhibition of thyrotropin secretion in hypothyroid rats. *Eur J Endocrinol.* (2005) 153:429–34. doi: 10.1530/eje.1.01984
 28. Filipovic B, Susic-Jurjevic B, Ajdzanovic V, Zivanovic J, Manojlovic-Stojanoski M, Nestorovic N, et al. The phytoestrogen genistein prevents trabecular bone loss and affects thyroid follicular cells in a male rat model of osteoporosis. *J Anat.* (2018) 233:204–12. doi: 10.1111/joa.12828
 29. van den Beld AW, Visser TJ, Feelders RA, Grobbee DE, Lamberts SW. Thyroid hormone concentrations, disease, physical function, and mortality in elderly men. *J Clin Endocrinol Metab.* (2005) 90:6403–9. doi: 10.1210/jc.2005-0872
 30. Spencer CA, Lum SM, Wilber JF, Kaptein EM, Nicoloff JT. Dynamics of serum thyrotropin and thyroid hormone changes in fasting. *J Clin Endocrinol Metab.* (1983) 56:883–8. doi: 10.1210/jcem-56-5-883
 31. de Vries EM, Fliers E, Boelen A. The molecular basis of the non-thyroidal illness syndrome. *J Endocrinol.* (2015) 225:R67–81. doi: 10.1530/JOE-15-0133
 32. Pearce SH, Razvi S, Yadegarfar ME, Martin-Ruiz C, Kingston A, Collerton J, et al. Serum thyroid function, mortality and disability in advanced old age: the Newcastle 85+ study. *J Clin Endocrinol Metab.* (2016) 101:4385–94. doi: 10.1210/jc.2016-1935
 33. Gussekloo J, van Exel E, de Craen AJ, Meinders AE, Frolich M, Westendorp RG. Thyroid status, disability and cognitive function, and survival in old age. *JAMA* (2004) 292:2591–9. doi: 10.1001/jama.292.21.2591
 34. Waring AC, Arnold AM, Newman AB, Buzkova P, Hirsch C, Cappola AR. Longitudinal changes in thyroid function in the oldest old and survival: the cardiovascular health study all-stars study. *J Clin Endocrinol Metab.* (2012) 97:3944–50. doi: 10.1210/jc.2012-2481
 35. Wakai K, Egami I, Kato K, Kawamura T, Tamakoshi A, Lin Y, et al. Dietary intake and sources of isoflavones among Japanese. *Nutr Cancer* (1999) 33:139–45. doi: 10.1207/S15327914NC330204
 36. Messina M. Isoflavone intakes by Japanese were overestimated. *Am J Clin Nutr.* (1995) 62:645. doi: 10.1093/ajcn/62.3.645
 37. de Kleijn MJ, van der Schouw YT, Wilson PW, Adlercreutz H, Mazur W, Grobbee DE, et al. Intake of dietary phytoestrogens is low in postmenopausal women in the United States: the Framingham study(1–4). *J Nutr.* (2001) 131:1826–32. doi: 10.1093/jn/131.6.1826
- Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
- Copyright © 2018 Sathyapalan, Köhrle, Rijntjes, Rigby, Dargham, Kilpatrick and Atkin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.