

1 **Occurrence of deoxynivalenol in an elderly cohort in the UK: a**  
2 **biomonitoring approach**

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29 **Occurrence of deoxynivalenol in an elderly cohort in the UK: a**  
30 **biomonitoring approach**

31 Deoxynivalenol (DON) is a *Fusarium* toxin, to which humans are frequently exposed via diet.  
32 Despite elderly are speculated to be sensitive to the toxic effects of DON as a result of age-  
33 related conditions, disease and altered DON metabolism, there is lack of available data on DON  
34 biomarkers in this age group. This study characterised urinary DON concentrations and its  
35 metabolites in elderly aged  $\geq 65$ years (n=20) residing in Hull, UK. Morning urinary specimens  
36 were collected over two consecutive days together with food records to assess dietary intake  
37 over a 24h-period prior to each urinary collection. Free DON (un-metabolised), total DON  
38 (sum of free DON and DON-glucuronides or DON-GlcA) and de-epoxy deoxynivalenol  
39 (DOM-1) were analysed using a validated LC-MS/MS methodology. Total DON was detected  
40 in 90% of elderly men and women on both days. Mean total DON concentrations on day 1 were  
41 not different from those on day 2 (elderly men, day 1:  $22.2 \pm 26.3$  ng/mg creat, day 2:  $28.0 \pm 34.4$   
42 ng/mg creat,  $p=0.95$ ; elderly women, day 1:  $22.4 \pm 14.6$  ng/mg creat, day 2:  $29.1 \pm 22.8$  ng/mg  
43 creat,  $p= 0.58$ ). Free DON and DON-GlcA were detected in 60-70% and 90% of total urine  
44 samples respectively. DOM-1 was absent from all samples. Estimated dietary intake of DON  
45 suggested that 10% elderly exceeded the maximum provisional tolerable daily intake for DON.  
46 In this single-site, UK-based cohort, elderly were frequently exposed to DON, although mean  
47 total DON concentrations were reported at moderate levels. Future larger studies are required  
48 to investigate DON exposure in elderly from different regions of the UK, but also from different  
49 counties worldwide.

50 Keywords: mycotoxins; deoxynivalenol; *Fusarium graminearum*; biomonitoring; elderly

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## 53 **Introduction**

54 The trichothecene deoxynivalenol (DON), also known as vomitoxin, is a secondary  
55 metabolite of *Fusarium graminearum* and *Fusarium culmorum*, to which humans and  
56 animals are predominantly exposed via food and feed (Pestka and Smolinski 2005; Pestka  
57 2010; European Food Safety Authority 2013). As a result of its solubility in water and  
58 stability during cooking (temperatures 120 °C), storage conditions and milling processes,  
59 DON contaminates cereal grains and remains to a large extent unaffected during manufacture  
60 (Bretz et al. 2006; Scudamore et al. 2009). Thus, it can be also found in cereal-based  
61 derivative products including breakfast cereals, bread, confectionary, beer, infant formulas,  
62 and baby foods (Scudamore et al. 2009; European Food Safety Authority 2013). Several  
63 animal studies have shown that acute DON exposure cause anorexia, vomiting, abdominal  
64 pain and diarrhoea, while lifetime exposure to DON has also been associated with  
65 reproductive impairments, neuro- and immuno-toxicity (Pestka 2010). Acutely, humans  
66 appear to be affected in a similar way as animals following high DON intake, confirming that  
67 the gastrointestinal track is a main target for DON toxicity (Pestka and Smolinski 2005;  
68 Pestka 2010; European Food Safety Authority 2013). In contrast, the chronic effects of DON  
69 exposure in humans are uncertain, and their study is hampered by challenges in assessing risk  
70 exposure, together with difficulties in differentially identifying symptoms of DON toxicity  
71 from those due to other illnesses (Etzel 2006; Marin et al. 2013).

72 Advances in biomonitoring with the development of highly sensitive analytical  
73 procedures have allowed the assessment of DON and its main metabolites in urine (Turner  
74 et al. 2008). DON can be excreted in urine in its un-metabolised form (free DON) or after

75 being converted to its metabolites (Turner et al. 2008). In brief, DON can be conjugated to  
76 glucuronides (DON-GlcA) in the liver and possibly in the intestine and kidneys, with  
77 deoxynivalenol-3-glucuronide (DON-3-GlcA) and deoxynivalenol-15-glucuronide (DON-  
78 15-GlcA) being two of the main DON metabolites currently identified (Pestka and Smolinski  
79 2005; Pestka 2010; European Food Safety Authority 2013). Deepoxy-DON (DOM-1) has  
80 been characterised as another major metabolite of DON produced by gut microbiota with  
81 deepoxidase activity in mammals (Pestka and Smolinski 2005; Pestka 2010; European Food  
82 Safety Authority 2013). DOM-1 has been inconsistently detected in human biological  
83 samples (Turner et al. 2010a; 2010b; Follmann et al. 2016; Wells et al. 2017), and remains  
84 uncertain, if human microbiota naturally possesses de-epoxidase activity, or whether rumen  
85 microbiota, which effectively converts DON to DOM-1, is transferred to humans during their  
86 contact with animals (Turner et al. 2010a; Wu et al. 2010). Biomonitoring studies in human  
87 adults have shown great variability in exposure to DON, as evaluated by the frequency of  
88 detection and biomarker concentrations in urine specimens (Turner, et al. 2010a; 2010b;  
89 Turner et al. 2011; Warth et al. 2012; Shephard et al. 2013; Heyndrickx et al. 2015; Gerding  
90 et al. 2014; Rodriguez-Carrasco et al. 2014; Solfrizzo et al. 2014; Gerding et al. 2015; Wallin  
91 et al. 2015; Follmann et al. 2016; Wells et al. 2017). These differences can be largely  
92 explained by geographical differences and eating patterns, whilst differences in population  
93 characteristics (e.g., age, physiological/disease status) may also contribute to these results.

94       Elderly may be an adult subpopulation at increased risk of experiencing toxicological  
95 effects when they are exposed to fungal toxins, including DON. DON toxic effects including  
96 anorexia, nausea, diarrhoea and stomach pain are similar to and may even exacerbate

97 symptoms associated with age-related conditions and diseases common amongst elderly  
98 (Talley et al. 1992; Martone et al. 2013). Although data from human studies are currently  
99 lacking, this notion is supported by findings in animal models suggesting increased  
100 susceptibility of aged animals to the negative side effects (*i.e.*, anorexia) of DON exposure  
101 compared to young adult animals (Clark et al. 2015). Ageing is accompanied by several  
102 physiological and metabolic changes including a reduction in renal and hepatic clearance  
103 (Mangoni and Jackson 2004), which in turn, may modify the way DON is absorbed,  
104 distributed, metabolised and excreted. Alterations in intestinal microbiota composition with  
105 increasing age have also been reported in humans (Claesson et al. 2011), albeit it remains  
106 unknown if such changes would affect the detoxification of DON. Conversely, a significant  
107 reduction in food intake has been reported with advancing age (Drewnowski 2000;  
108 Wakimoto and Block 2001), which suggests a lower DON intake in elderly through this main  
109 route of exposure. To our knowledge, elderly have not been previously considered as a  
110 separate population group with distinct characteristics compared to those of their younger  
111 counterparts within DON-related biomonitoring research.

112         This study aimed to characterise DON concentrations and its metabolites in urine of  
113 elderly individuals aged  $\geq 65$  years residing in Hull, UK. To allow comparison with younger  
114 adults, a dataset of UK adults aged 18-64 years residing in the same area, previously  
115 published as part of our ongoing work in this area of research, was used (Wells et al. 2017).

## 116 **Materials and Methods**

### 117 *Participants selection and recruitment*

118 Elderly aged  $\geq 65$  years (n=20) and adults aged 18-64 years living in the Hull and East  
119 Yorkshire were recruited via word of mouth, by an announcement in the local newspapers  
120 and an email distributed by the University of Hull and Hull and East Yorkshire Hospitals  
121 NHS.

122 Inclusion criteria were being in general good health, not taking any current  
123 medication initiated within the last 3 months or were on stable medication (over a duration  
124  $>3$  months). Exclusion criteria were inability to provide informed consent, acute or chronic  
125 illness (chronic renal, hepatic or cardiac problems, cancer), chronic gastrointestinal  
126 conditions (*e.g.*, coeliac disease), gluten sensitivity, eating disorders, depression, psychosis  
127 or hospitalisation within the last three months prior to enrolment in the trial or participation  
128 in a weight loss programme. Individuals on stable medication, which may influence appetite  
129 such as oral steroids, were not included in the study.

130 Ethical approval was granted by the National Health Service (NHS), National  
131 Research Ethics Service (NRES) Committee Yorkshire & the Humber-Leeds West (IRAS  
132 project code: 147707).

### 133 *Study design*

134 This dataset analysed in the present study consists part of a more comprehensive investigation  
135 entitled “Experimental study of deoxynivalenol biomarkers in urine” performed for the  
136 European Food Safety Authority GP/EFSA/CONTAM/2013/04 (Brera et al. 2015), which

137 investigated the presence of DON and DON metabolites in urine samples collected from  
138 children (aged 3-9 years), adolescents (aged 10-17 years), adults (aged 18-64 years), pregnant  
139 women and elderly (aged  $\geq 65$  years) (total n=635) in the UK, Italy and Norway.

140 A validated, semi-quantitative food frequency questionnaire (FFQ) (previously used  
141 in (Brera et al. 2015; Wells et al. 2016; Wells et al. 2017), was designed to assess food  
142 consumption of cereal grains and cereal-based products that commonly contribute to DON  
143 dietary exposure over a month recall period, while a food record was used to collect detailed  
144 information about food items consumed 24hours preceding the collection of each urine  
145 sample. Age, height, weight and physical activity were self-reported as part of the FFQ.  
146 Participants were supplied with four urine collection containers of 50 mL each and written  
147 guidance for collecting their first morning urine samples at home, on two consecutive days.  
148 This timeframe was intended to cover potential between-day variability within participant  
149 and enhance the repeatability of our findings. Participants returned the urine samples at the  
150 Hull Royal Infirmary (Hull, UK) on the same day of collection. All samples were centrifuged  
151 at 2000 rpm for 10 minutes and stored at  $-80^{\circ}\text{C}$  until further analysis. DON and its  
152 metabolites were tested using the validated HPLC-MS/MS methodology as previously  
153 described in (Turner et al. 2008).

#### 154 ***Laboratory analysis***

155 DON and its metabolites were analysed using  $^{13}\text{C}$  labelled DON standard (Sigma, Saint  
156 Louis, MI, USA; product number: 34128, 1.2 mL), DON (Sigma, Saint Louis, MI, USA;  
157 product number: D0156, 1 mg),  $\beta$ -glucuronidase (Type IX-A from E. coli; Sigma, Saint

158 Louis, MI, USA; product number: G7396 - 2MU), DOM-1 (Sigma, Saint Louis, MI, USA;  
159 product number: 34135, 2 mL), and DON test WB<sup>TM</sup> immunoaffinity columns (Vicam,  
160 Milford, MA, USA; product number: G1066). All analyses were conducted on a Waters 2795  
161 HPC Separation Module (Waters Corp., Milford, MA, USA) with a Quattro Micro Triple  
162 Quadrupole Mass Spectrometer (Micromass UK Ltd., Manchester, UK).

### 163 *Sample preparation*

164 Stored urine samples were centrifuged at 2000 rpm at -4 °C for 15 minutes. For each  
165 participant, two aliquots (1 mL) were prepared by mixing <sup>13</sup>C-DON internal standard solution  
166 to a final concentration of 20 ng/mL. In the first aliquot, total DON was determined as the  
167 sum of DON-GlcAs and free DON. For measuring the combined DON-GlcAs (DON-3-GlcA  
168 and DON-15-GlcA) and free DON, each sample was set to pH 6.8 and treated with β-  
169 glucuronidase solution (23,000 units, in KH<sub>2</sub>PO<sub>4</sub> 75 mM) in a shaking water bath at 37 °C  
170 for 18 hours. The samples were then centrifuged (2000 rpm; -4 °C; 15 min), and the  
171 supernatant was diluted to a final 4 mL with phosphate buffered saline (PBS, pH 7.4), before  
172 being passed through a wide bore DON immunoaffinity column. DON was removed from  
173 columns with methanol (4 mL) and extracts were dried under vacuum using a Savant<sup>TM</sup>  
174 SpeedVac<sup>TM</sup> (Thermo Fisher Scientific Inc., Waltham, MA, USA) or equivalent and  
175 dissolved in 10% ethanol (250 μL) for LC-MS analysis. DOM-1 was quantified on the same  
176 aliquot analysed for DON-GlcA. Free DON was assessed in the second aliquot using the  
177 same procedures, but no β-glucuronidase treatment was performed.



178 *HPLC-MS analysis: DON determination*

179 The separation of DON was performed by utilising reversed phase chromatography using a  
180 Luna C<sub>18</sub> column (150 × 4.6 mm, 5-µm particle size) (Phenomonex, Macclesfield, UK) with  
181 a mobile phase sequence of 27 minutes 20% methanol, reconstructed to a wash of 75%  
182 methanol after 10 minutes followed by 20% methanol after 16 minutes (flow rate 1 mL/min;  
183 injection volume 25µL). One fifth of the eluent was placed into the desolvation chamber of  
184 the MS. Selective ion recording (SIR) was used to quantify DON with respect to <sup>13</sup>C-DON  
185 internal standard. The following mass spectrometer conditions were kept stable: capillary  
186 voltage 3.5 kV, desolvation temperature: 300 °C, extraction cone voltage: 3.00 V, sampling  
187 cone voltage: 35.00 V, source temperature: 100 °C, cone gas flow: 50 L/h, collision energy:  
188 1.0 and desolvation gas flow 500 L/h. Two masses of DON ([DON-H]<sup>+</sup>, m/z 297.2 and  
189 [DON-Na]<sup>+</sup>, m/z 319.2) and <sup>13</sup>C-DON ([<sup>13</sup>C-DON-H]<sup>+</sup>, m/z 312.2 and [<sup>13</sup>C-DON-Na]<sup>+</sup>, m/z  
190 334.2) were monitored for 0.25 seconds (each mass) and were then summed to form a total  
191 ion current peak for the internal standard and each analyte. The calibration curve (range 2-  
192 250 ng/mL) was established by injecting DON and <sup>13</sup>C-DON standard solutions (prepared in  
193 10% ethanol) DON-GlcA concentrations were estimated indirectly, by subtracting free DON  
194 from total DON values.

195 *LC-MS analysis: DOM-1 determination*

196 DOM-1 was separated by utilising the same chromatographic column used for DON  
197 separation combined with a mobile phase sequence of 35 minutes 20% methanol, changed to  
198 a wash of 75% methanol after 20 minutes and then to a phase of 20% methanol after 26

199 minutes (injection volume 25 L; flow rate 1 mL/min). Part of the eluent was driven into the  
200 MS desolvation chamber. Similar to DON analysis, DOM-1 was quantified by SIR with  
201 reference to the calibration curve (range of 2–200 ng/mL), which resulted from the injection  
202 of DOM-1 standard solutions (prepared in 10% ethanol). Each of two masses of DOM-1,  
203 [DOM-1-H]<sup>+</sup>, m/z 281.3 and [DOM-1-Na]<sup>+</sup>, m/z 303.3 were monitored (0.25 seconds) and  
204 summed to obtain a total ion current peak for DOM-1.

#### 205 *Analysis of creatinine*

206 Adjustments of DON concentrations for creatinine were used to account for differences in  
207 dilution between individuals that may have resulted from the sole collection of a first morning  
208 urine samples. An in-house micro-titre plate assay was used to determine urinary creatinine.  
209 Samples were diluted in water (1:20) and 100 µL was added, in duplicate, to a 96-well plate.  
210 A duplicate standard curve of creatinine concentrations within the range 0-20 µg/mL was  
211 obtained for each plate. A further 100 µL of alkaline picric acid solution was added to each  
212 well, incubated at 25 °C for 30 minutes and measured at 490 nm using a plate  
213 spectrophotometer. Urinary total DON concentrations are presented as unadjusted (ng/mL)  
214 and adjusted for creatinine (ng/mg creatinine).

#### 215 *Estimated dietary exposure of DON based on urinary analysis*

216 The estimated dietary exposure of DON was calculated using formula 1 (Ezekiel et al. 2014;  
217 Heyndrickx et al. 2015):

$$218 \text{ Estimated dietary exposure of DON } (\mu\text{g/kg b.w./day}) = \frac{\text{total DON} \times V}{ER \times \text{b.w.}} * 1000 \text{ (1)}$$

219 **Total DON = unadjusted total DON concentrations in urine**

220 ER = urinary excretion rate of DON is 72% (Turner et al. 2010b).

221 V = daily urine output (mL/day) was estimated to be 1.5 L

222 b.w. = self-reported body weight (kg)

223 Based on the toxicological data in animals and humans, the Joint Food and Agriculture  
224 Organization of the United Nations (FAO)/World Health Organisation (WHO) Expert  
225 Committee on Food Additives (JECFA) have set a provisional maximum tolerable daily  
226 intake (PMTDI) at 1 µg/kg body weight (b.w.) day for DON and its main metabolites (Joint  
227 Expert Committee on Food and Additives, 2010). As such, we compared the estimated  
228 dietary exposure of DON with PMTDI at µg/kg b.w./day and characterised exposure as below  
229 or exceeding PMTDI.

### 230 *Dietary assessment and analysis*

231 In this report, the results of the food records covering the dietary intake over a 24-h period  
232 prior to each urinary samples collection are only reported. The food records included the  
233 following food categories that are commonly contaminated with DON: breakfast cereals and  
234 snacks, bread, products alternative to bread, flakes, cereals, other breads, biscuits, bakery  
235 goods, pizza, pasta, wheat germ, beer, pancakes and pita bread. These categories are in line  
236 with the validated FFQ designed to assess DON exposure through diet and used in previous  
237 DON-related research (Brera et al. 2015; Wells 2016; 2017). Each of the two food records  
238 included three main meals (breakfast, lunch, dinner) and snacks. For each of these eating  
239 occasions, examples of food items belonging to the aforementioned food categories were  
240 included. To enhance consistency and enable participants to record intake in an easy way,

241 each category was tabulated with tick boxes as to whether the portion consumed was small,  
242 medium or large. Photographic examples of portion sizes were also used to help participants  
243 to accurately quantify their dietary intake. Importantly, participants completed food records  
244 at the presence of research dietitians, who provided clarification and asked supplementary  
245 information. Portion sizes reported in the food records were converted into grams by research  
246 dietitians prior to data analysis.

### 247 *Statistical Analysis*

248 All numerical data were checked for normality using the Shapiro-Wilk test. Urinary DON  
249 levels between day 1 and day 2 within adults and elderly were compared using a paired *t-test*  
250 and a Wilcoxon signed-rank test for normally and non-normally distributed respectively.  
251 Comparisons between groups were performed using an independent *t-test* or Mann-Whitney  
252 test for normally and non-normally distributed, respectively. Fisher's exact test was used for  
253 comparing categorical variables. The association between urinary DON levels and its  
254 metabolites and sex, weight, BMI and dietary intake of commonly foods contaminated with  
255 DON were assessed using the Spearman's correlation coefficient (two-tailed). All analyses  
256 were carried out using IBM SPSS Statistics Version 24.0. Results were deemed significant  
257 at a *p*-value  $\leq 0.05$ .

## 258 **Results**

### 259 *Baseline characteristics*

260 A total of 50% of the 20 elderly (aged  $\geq 65$  years) and 52% of the 31 adults (aged 18-64 years)

261 were men. Baseline characteristics of elderly and adults by sex are shown in Table 1. Men  
262 were significantly taller (elderly,  $p=0.15$ ; adults,  $p<0.001$ ) and heavier (elderly,  $p=0.04$ ;  
263 adults,  $p<0.001$ ) than women in both the elderly and adult groups. There was a trend towards  
264 a higher BMI in elderly men than women ( $p=0.051$ ) and BMI was not different between adult  
265 men than women ( $p=0.10$ ). Elderly men were significantly shorter than adult men ( $p=0.03$ ),  
266 but there were no further differences in weight ( $p=0.28$ ) and BMI ( $p=0.63$ ). Height ( $p=0.34$ ),  
267 weight ( $p=0.22$ ) or BMI ( $p=0.36$ ) did not differ between adult and elderly women. In all  
268 groups, most of the participants reported moderate physical activity (elderly men, 80%;  
269 elderly women, 70%; adult men, 50%; adult women, 80%), and there were no significant  
270 differences for physical activity categories among groups ( $p$ -values from 0.14 to 0.49).  
271 Elderly women had significantly lower urinary creatinine levels (mg/dl) than elderly men  
272 ( $p=0.003$ ) and adult women ( $p=0.004$ ), which is in line with sex and age differences shown  
273 previously (Barr et al., 2005).

#### 274 *Total DON Concentrations in Urine Samples*

275 Table 2 presents unadjusted and creatinine-adjusted DON concentrations in urine samples on  
276 day 1 and 2 in elderly ( $n = 20$ ) and adults ( $n = 31$ ). Total DON was detected in 90% of elderly  
277 men and women on both days and 100% of adult men and women on days 1 and 2. Mean  
278 total creatinine-adjusted DON concentrations (ng/mg creat) on day 1 were not different from  
279 total DON concentrations on day 2 in elderly (men, day 1:  $22.2 \pm 26.3$ , day 2:  $28.0 \pm 34.4$ ,  
280  $p=0.95$ ; women, day 1:  $22.4 \pm 14.6$ , day 2:  $29.1 \pm 22.8$ ,  $p= 0.58$ ) or adults (men, day 1:  $24.3$   
281  $\pm 38.2$ , day 2:  $21.3 \pm 19.1$ ,  $p=0.41$ ; women, day 1:  $12.7 \pm 7.9$ , day 2:  $18.2 \pm 18.0$ ,  $p=0.82$ ) (Table

282 2). Mean total creatinine-adjusted DON concentrations (ng/mg creat) for both days (pooled  
283 data for day 1 and day 2) were 25.1 for elderly men ( $n = 10$ ) and 22.8 for adult men ( $n = 16$ ),  
284 with no differences found between the two groups ( $p = 0.70$ ). Although greater mean total  
285 DON concentrations (pooled data for day 1 and 2) were reported in elderly women  
286 ( $25.7 \pm 17.4$  ng/mg creat) compared to those in adult women ( $15.5 \pm 11.2$  ng/mg creat), these  
287 did not reach statistical significance ( $p = 0.079$ ).

### 288 *DON metabolite concentration in urine samples*

289 Free DON (>limit of quantification or LOQ 0.25 ng/mL) was detected in 60-70% of elderly  
290 men and women, and in all urine samples of adults. DON-GlcA (>LOQ 0.50 ng/mL) was  
291 present in elderly (90%) and adults (100%) samples. In contrast, DOM-1 was absent from all  
292 samples of both age groups. In elderly subjects, DON-GlcA made significant contribution  
293 ranging from 83 to 91% of the total DON and free DON represented 9-17% of total DON;  
294 with these contributions being very similar among elderly and adults (Table 2).

295 Mean free DON concentrations (ng/mg creat) were 3.8 and 3.2 on day 1 and day 2 in  
296 urinary specimens of elderly men and 2.3 and 2.8 for days 1 and 2 in samples of elderly  
297 women (Table 2). In adult men, mean urinary levels of free DON were reported to be 4.2 and  
298 3.2 ng/mg creat on days 1 and 2, while the respective values in adult women were 1.9 and  
299 2.8 ng/mg creat (Table 2). There were no differences in mean free DON concentrations  
300 between day 1 and day 2 in elderly (men,  $p = 0.50$ ; women,  $p = 0.58$ ) or adults (men,  $p = 0.50$ ;  
301 women,  $p = 0.45$ ). Furthermore, mean free DON concentrations (pooled data for days 1 and  
302 2), were not significantly different between elderly and adult men (elderly men:  $3.5 \pm 4.3$

303 ng/mg creat vs. adult men:  $3.7 \pm 4.8$  ng /mg creat;  $p=0.52$ ) or women (elderly women:  $2.3 \pm 1.6$   
304 ng/mg creat vs. adult women:  $2.6 \pm 1.6$  ng /mg creat;  $p=0.74$ ).

305       Amongst elderly individuals, mean DON-GlcA (ng/mg creat) concentrations were  
306 18.5 and 21.8 on days 1 and 2 in men and 20.1 and 26.3 for days 1 and 2 in women (Table  
307 2). Among adults, mean urinary levels of DON-GlcA (ng/mg creat) for days 1 and 2 were  
308 20.0 and 18.1 for men and 10.8 and 15.4 for women (Table 2). There were no differences  
309 between days 1 and 2 in any group (elderly men,  $p=0.95$ ; elderly women,  $p=0.21$ ; adult men,  
310  $p=0.41$ ; adult women,  $p=0.96$ ). When DON-GlcA (ng/mg creat) data were pooled for both  
311 days, no differences were seen between elderly and adult men (elderly men:  $20.1 \pm 15.1$  ng/mg  
312 creat vs. adult men:  $19.1 \pm 22.1$  ng /mg creat;  $p=0.62$ ) or elderly and adult women (elderly  
313 women:  $23.2 \pm 16.4$  ng/mg creat vs. adult women:  $13.1 \pm 9.7$  ng /mg creat;  $p=0.067$ ).

#### 314 *Estimated dietary DON exposure using urinary total DON concentrations*

315 We carried out a risk assessment by comparing the estimated dietary intake of DON with the  
316 PMTDI of  $1 \mu\text{g}/\text{kg b.w.}/\text{day}$ . The mean estimated dietary intake of DON was  $0.43$   
317  $\mu\text{g}/\text{kg}\cdot\text{b.w.}/\text{day}$  (minimum: 0, maximum: 2.33) for elderly and  $0.37 \mu\text{g}/\text{kg}\cdot\text{b.w.}/\text{day}$   
318 (minimum: 0.07, maximum: 1.33) for adults. In total, 10% of elderly and <3% of adults were  
319 estimated to exceed the PMTDI of  $1 \mu\text{g}/\text{kg b.w.}/\text{day}$ .

#### 320 *Correlation between urinary DON, demographic and anthropometric* 321 *characteristics*

322 In elderly, no correlations were found between urinary mean total DON (ng/mg creat) (pooled  
323 data for day 1 and day 2) and sex ( $r=-0.43$ ;  $p=0.86$ ), height ( $r=-0.128$ ;  $p=0.59$ ), weight ( $r=-$

324 0.294;  $p=0.21$ ) or BMI ( $r=-0.259$ ,  $p=0.27$ ).

### 325 ***Correlation between urinary total DON and Food Intake***

326 Table 3 presents main food categories commonly consumed by elderly and adults, which are  
327 often contaminated with DON. One adult participant had a total intake of all food categories  
328 equal to 0 on both experimental days, which is unlikely given the characteristics of our  
329 cohorts (healthy individuals, not under energy restriction). Therefore, this participant was  
330 excluded from this correlation analysis between urinary total DON and main food categories  
331 contributing to DON exposure. Analysis was performed in 20 elderly and the remaining 30  
332 adults with complete data. Elderly consumed significantly more flakes ( $p=0.016$ ), biscuits  
333 ( $p=0.033$ ) and baked good ( $p=0.044$ ) compared to adults. There were no other differences for  
334 any other food category between the two age groups ( $p$  values from 0.066 to 0.57). Total  
335 intake of commonly contaminated foods with DON did not differ between the elderly  
336 compared to adults ( $p=0.28$ ). In elderly, no significant correlations were found between  
337 urinary mean total DON (ng/mg creat) for both days and any food category that commonly  
338 contribute to dietary DON exposure ( $p$  values from 0.056-0.93). In a pooled analysis of  
339 elderly and adults, urinary mean total DON (ng/mg creat) for both days were positively  
340 correlated with flakes consumption ( $r=0.426$ ,  $p=0.02$ ), but not with any other food category  
341 or total intake of commonly contaminated foods with DON ( $p$  values from 0.13 to 0.95)  
342 (Table 3).

343



## 344 **Discussion**

345 Elderly individuals may be particularly susceptible to DON exposure due to age-related  
346 alterations in absorption, distribution, metabolism and excretion of toxins, but also as a result  
347 of diseases, which may resemble or increase sensitivity to the toxic effects of DON on bodily  
348 systems (Mangoni and Jackson 2004; Clark et al. 2015). Our study provides data on the  
349 concentrations of DON and its metabolites in urine samples of elderly individuals. A frequent  
350 detection of total DON (90%) at a mean concentration of 25.4 ng/mg creat, suggests DON  
351 exposure, although at relatively moderate levels in this small cohort in elderly. Compared to  
352 adults aged 18-64 years residing in the same area, elderly appeared to have similar urinary  
353 levels of DON, indicating no significant differences in DON exposure with age.

354 Comparative data in elderly are only available as part of our larger investigation,  
355 which in addition to the UK, included participants from Norway and Italy (Brera et al. 2015).  
356 Mean total DON concentrations in UK elderly (n=20, incidence=90%, mean levels=25.4  
357 ng/mg creat) were almost 3-fold to those detected in the elderly cohorts in Norway (n=20,  
358 incidence=100%, mean levels=8.9 ng/mg creat) and Italy (n=20, incidence=100%, mean  
359 levels=8.3 ng/mg creat) (Brera et al. 2015). Most available studies in adult populations have  
360 not included elderly individuals (aged  $\geq 65$  years), whereas those, that have included  
361 participants  $\geq 65$  years, have treated them as part of a mixed group of adults ( $\geq 18$  years)  
362 (Turner et al. 2010a; 2010b; Turner et al. 2011; Warth et al. 2012; Gerding et al. 2014;  
363 Heyndrickx et al. 2015; Follmann et al. 2016). Similarly to our study in elderly and adults  
364 based in a single urban area in the UK, current findings from European counties suggest that  
365 adults are frequently exposed to DON, however, the magnitude of exposure is highly variable

366 (Germany: n=30, incidence=100%, mean levels=6.0 ng/mg creat (Föllmann et al. 2016);  
367 Belgium: n=239, incidence 100%, mean levels: 87.9 ng/mg creat (Heyndrickx et al. 2015);  
368 Spain: n = 22; incidence: 73%, mean levels = 14.8 ng/mg creat (Rodriguez-Carrasco et al.  
369 2014); Sweden: n=252, incidence: 63%, mean levels=7.0 ng/mg (Wallin et al. 2015); Italy:  
370 n=52, incidence: 96%, mean levels=11.9 ng/mL (Solfrizzo et al. 2014); and Austria: n=27,  
371 incidence=96%, mean levels =20.4 ng/mL (Warth et al. 2012). Similar findings have also  
372 been reported in non-European countries including China (Turner et al. 2011), South Africa  
373 (Shephard et al. 2013) and Haiti (Gerding et al. 2015). The variability in the results may  
374 reflect differences in exposure as a result of dietary patterns, climate conditions, cultivating  
375 and processing or may be due to differences in study design, characteristics of study  
376 populations or analytical procedures and lower limits of quantification used for urinary DON  
377 assessment.

378         Understanding the metabolism of DON in elderly is important in characterizing the  
379 risk of this population to the toxic effects of DON exposure. Towards this end, free DON  
380 (unmetabolised form of DON) was found in 60-70% of the urine samples of the elderly  
381 individuals. DON-GlcA was detected in 90% of all urine specimens of elderly individuals.  
382 In the positive samples, free DON accounted for 9-17% of the total DON concentrations;  
383 whilst DON-GlcA made the greatest contributions to total DON concentrations ranging from  
384 83-91%. Our results suggest that conjugation to glucuronides is the main detoxification route  
385 for DON in elderly humans. Similarly to our elderly cohort, contributions of DON-GlcA over  
386 80% of total DON concentrations were evident in our adult cohort. Our findings in elderly  
387 and adults individuals are in line with those of previous studies in adult populations (Turner

388 et al. 2011; Warth et al. 2012; Heyndrickx et al. 2015; Gerding et al. 2014). Few studies have  
389 also given insight into the different conjugates (Warth et al. 2012). For example, in a study  
390 in Austrian adults, DON-15GlcA accounted for about 75% of total DON-GlcA (Warth et al.  
391 2012). Heyndrickx et al., also confirmed that DON-GlcA at the 15-position is preferential  
392 than glucuronidation of DON at the 3-position (Heyndrickx et al. 2015). In our work, we  
393 were not able to differentiate between DON-GlcA at different positions since an enzymatic  
394 hydrolysis with  $\beta$ -glucuronidase allowed the estimation of DON-GlcA indirectly, by  
395 subtracting free DON from total DON concentrations.

396 In contrast to free DON and DON-GlcA, DOM-1 was not detected in any sample of  
397 our elderly or adult population. These results are consistent with previous findings in elderly  
398 from Italy (Brera et al. 2015), but also other studies in adults (Turner et al. 2011; Rodriguez-  
399 Carrasco et al. 2014) and support the idea that conversion of DON to DOM-1 may not be a  
400 main metabolic pathway in humans. This may be due to the natural absence of microbiota  
401 with the enzymatic activity required for this conversion (Turner et al. 2010a). Alternatively,  
402 if DON was converted to DOM-1, this metabolite was not significantly excreted in urine  
403 samples in the present investigation. Our results are strengthened by the analytical approach  
404 we followed for DOM-1 determination; given that urine samples were treated with  $\beta$ -  
405 glucuronidase, identification of both DOM-1 and DOM-1-glucuronide would be possible.  
406 DOM-1 has been detected in some other adult cohorts across Europe. In the experimental  
407 study of DON biomarkers in urine (Brera et al. 2015) performed in Italy, Norway and UK,  
408 DOM-1 (>LoQ) was detected in 12% of Norwegian samples, probably due to a lower LoQ  
409 compared to those used for the analysis of the samples in Italy and the UK. Similarly, 34%

410 of the samples of French farm workers (Turner et al. 2010a) and 38–60% of the total samples  
411 of German mil workers and controls ( Föllmann et al. 2016) were positive for DOM-1. These  
412 latter findings have been suggested to result from the transmission of rumen microbiota with  
413 de-epoxidase activity to humans (Turner et al. 2010a). Given that there is evidence to suggest  
414 changes in microbiota with increasing age, it will be useful to understand variations in the  
415 ability to detoxify DON by deepoxidation to DOM-1 in larger studies in elderly from  
416 different countries, and to consider differences by disease status.

417         Urinary biomarkers concentrations can be used to estimate the dietary intake of DON  
418 at an individual level, which can be subsequently be compared with the PMTDI (1 µg/kg  
419 b.w./day). By assuming a 72% urinary excretion ratio of DON and 1.5 L daily urine output  
420 (Turner et al. 2010b), we showed that the estimated dietary intake of DON varied between 0  
421 and 2.33 µg/kg b.w./day for elderly. In total 10% of the elderly were shown to exceed the  
422 PMTDI, but, notably, the 75% of the elderly individuals were estimated to have dietary intake  
423 of DON < 0.5 µg/kg b.w./day, which is below the half of the PMTDI. The percentage of the  
424 elderly exceeding the PMTDI is higher than the <3% of adults aged 18-64 who exceeded the  
425 PMTDI in this analysis, though the numbers in the study were low. Higher percentages  
426 exceeding the PMTDI than those reported amongst elderly in the present study have also  
427 been reported in other studies in adults in the UK (n=35, estimated dietary intake of DON:  
428 0.008–1.24 µg/kg·b.w./day, 17% participants exceeded PMTDI) (Turner et al. 2010b),  
429 Belgium (n=239, 16-29% participants exceeded PMTDI) (Heyndrickx et al. 2015), Italy  
430 (n=52, 40% participants exceeded PMTDI) (Solfrizzo et al. 2014) and Austria (n=27, 33%  
431 participants exceeded PMTDI) (Warth et al. 2012). In contrast, lower percentages were

432 shown in Spain (n=22; <4% exceeded PMTDI) (Rodriguez-Carrasco et al. 2014). Our results  
433 and their comparisons with the findings of previous studies should be interpreted after  
434 considering the limitations of current ways to estimate dietary intake of DON and the small  
435 number of participants in most available studies. For example, the uncertainty in these  
436 estimations was depicted by Heyndrickx and colleagues who showed 16 and 29% of total  
437 sample to exceed PMTDI depending on the formula used (Heyndrickx et al. 2015). In our  
438 previous work in children and adolescents, we also showed significant discrepancies in the  
439 percentages of participants exceeding PMTDI, when different assumption of daily urine  
440 output were tested (Papageorgiou et al. 2018). Assumptions of the urinary excretion of DON,  
441 reported to range 50-72% in previous human studies (Turner et al. 2010b; Shephard et al.  
442 2013), may further influence the estimations of dietary intake of DON based on urinary  
443 biomarkers. Other reasons that may explain the variable estimations of the dietary intake of  
444 DON include differences in dietary habits of population residing in different countries or  
445 seasonal variations in dietary intake within a population, but also discrepancies in DON  
446 contamination of food sources as a result of environmental conditions, cultivation, handling  
447 and processing (Bullerman & Biachini, 2007; Oldenburg et al., 2012; Gratz et al., 2014).  
448 Taken together, the findings of the available studies and their comparison underpin the need  
449 for further research in the area and standardization of the methodology used for estimating  
450 dietary exposure to DON. Future investigations using larger sample sizes of elderly  
451 populations are also required to confirm our findings and allow comparisons in this age  
452 population group.

453 To further explore the relationship between dietary exposure of DON and DON  
454 biomarkers, correlation analysis between the consumption of common foods contaminated  
455 with DON (based on food records over 24h prior to each urine sample collection) and total  
456 DON concentrations (pooled data for the 2 experimental days) in urine was performed. For  
457 our total samples of elderly and adults, this analysis revealed a significant positive association  
458 between flakes consumption and urinary mean total DON (ng/mg creat), but no significant  
459 correlations for any other food category were shown. The lack of pronounced associations  
460 may at least partially be explained by the fact that overall, DON exposure was relatively low  
461 amongst both elderly and adult participants and in this small sample. Limitations in the  
462 dietary assessment method (i.e., food records over a 24h period), challenges in estimating  
463 portion sizes or reporting bias may have also contributed to these non-significant  
464 associations. Nevertheless, given that urinary DON reflects to a large extent the DON dietary  
465 intake of the previous days (Turner et al. 2010b), food records over 24h prior to urine sample  
466 collection may more accurately reflect DON dietary exposure compared to FFQs, which  
467 typically capture dietary intake over a more prolonged period of time (e.g., last month)  
468 (Marin et al. 2013). For future studies investigating DON exposure, we recommend a longer  
469 recording period to get information on habitual dietary intake or intake of less frequently  
470 consumed foods. Inclusion of actual measurements of DON and metabolites in the recorded  
471 foods would be also useful in ascertaining associations between DON dietary exposure and  
472 DON biomarkers in biological samples. To minimize reporting bias and increase accuracy of  
473 portion sizes, research dietitians with experience in performing semi-structured interviews  
474 asked supplementary questions on the foods records and pictures with portion sizes were used

475 respectively. The food categories and example of food items used as the basis of the food  
476 records were comprehensive and previously used in DON research (Brera et al. 2015; Wells  
477 et al. 2017; Papageorgiou et al. 2018), however, some dietary sources contributing to the  
478 exposure of DON, which may be consumed less frequently at a local level, may have still  
479 been omitted (e.g., soups and sauces).

480 We provided a detailed analysis of DON exposure in elderly, by determining DON  
481 and its main metabolites in urinary specimens and assessed intake of main food sources  
482 contaminated to DON. It is important to note that other factors including defence mechanisms  
483 and altered metabolism due to illness not investigated as part of this work may influence  
484 elderly's sensitivity to the toxic effects of DON. Our elderly population were relatively  
485 healthy (as indicated by inclusion and exclusion criteria), implying no major impairments in  
486 gastrointestinal, renal, hepatic or immune functions. As such, our results may be less  
487 generalizable in elderly individuals with disease (e.g., cancer, renal and liver disease,  
488 gastrointestinal disorders, or immunosuppression). Furthermore, it remains unknown how  
489 DON exposure (e.g., low *vs.* high) affects elderly individuals who experience symptoms that  
490 resemble DON toxicity such as anorexia of ageing or unintended weight loss and whether  
491 DON biomarkers can capture such effects.

## 492 **Conclusions**

493 Despite their potential sensitivity to DON toxicity, the elderly are an understudied population  
494 in mycotoxin research. Although elderly individuals in this single-site, UK-based cohort were  
495 commonly exposed to DON, mean total DON concentrations were reported at moderate

496 levels and PMTDI for DON (1 µg/kg b.w./day) was exceeded by 10% of the population  
497 studied. Furthermore, our findings support no significant differences in DON and its  
498 metabolites between elderly (aged ≥65 years) and younger adults (aged 18-64 years). Future  
499 larger studies are needed to explore DON exposure in elderly from different regions of the  
500 UK, but also from different countries worldwide. Simultaneous assessment of urinary DON  
501 biomarkers, dietary patterns, gastrointestinal distress (as a result of ageing, disease or DON  
502 toxicity) and disease status would be important to understand DON exposure, susceptibility  
503 and toxicity in this population.

504

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509

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516



517 **References**

- 518 Barr DB, Wilder LC, Caudill SP, Gonzalez AJ, Needham LL, Pirkle JL. 2005. Urinary creatinine  
519 concentrations in the U.S. population: implications for urinary biologic monitoring  
520 measurements. *Environ. Health Perspect.* 113(2):192-200. DOI: 10.1289/ehp.7337.
- 521 European Food Safety Authority. 2013. Deoxynivalenol in food and feed: occurrence and exposure.  
522 EFSA J. 11: 2013. DOI: 10.2903/j.efsa.2013.3379.
- 523 Brera C, Santis B, Debegnach F, Miano B, Moretti G, Lanzone A, Del Sordo G, Buonsenso D,  
524 Chiaretti A, Hardie L, White K. 2015. Experimental study of deoxynivalenol biomarkers in  
525 urine. EFSA Supporting Publications. EN-81. DOI: 10.2903/sp.efsa.2015.EN-818.
- 526 Bretz M, Beyer M, Cramer B, Knecht A, Humpf HU. 2006. Thermal degradation of the Fusarium  
527 mycotoxin deoxynivalenol. *J Agric Food Chem.* 54: 6445-51. DOI: 10.1021/jf061008g.
- 528 Bullerman, L. B.; Bianchini, A. 2007. Stability of mycotoxins during food processing. *Int J Food*  
529 *Microbiol.* 119 (1-2), 140-6. DOI: 10.1016/j.ijfoodmicro.2007.07.035.
- 530 Claesson MJ, Cusack S, O'Sullivan O, Greene-Diniz R, de Weerd H, Flannery E, Marchesi JR, Falush  
531 D, Dinan T, Fitzgerald G. et al. 2011. Composition, variability, and temporal stability of the  
532 intestinal microbiota of the elderly. *Proc Natl Acad Sci USA.* 108 Suppl 1: 4586-91. DOI:  
533 10.1073/pnas.1000097107.
- 534 Clark, ES, Flannery BM, Gardner EM, Pestka JJ. 2015. High Sensitivity of Aged Mice to  
535 Deoxynivalenol (Vomitoxin)-Induced Anorexia Corresponds to Elevated Proinflammatory  
536 Cytokine and Satiety Hormone Responses. *Toxins (Basel).* 7: 4199-215. DOI:  
537 10.3390/toxins7104199.
- 538 Drewnowski A. 2000. Sensory control of energy density at different life stages. *Proc Nutr Soc.* 59:  
539 239-44. DOI: 10.1017/S0029665100000264.
- 540 Etzel RA. 2006. What the primary care pediatrician should know about syndromes associated with  
541 exposures to mycotoxins. *Curr Probl Pediatr Adolesc Health Care.* 36: 282-305. DOI:  
542 10.1016/j.cppeds.2006.05.003.
- 543 Ezekiel CN, Warth B, Ogara IM, Abia WA, Ezekiel VC, Atehnkeng J, Sulyok M, Turne PC, Tayo  
544 GO, Krska R, Bandyopadhyay R. 2014. Mycotoxin exposure in rural residents in northern  
545 Nigeria: a pilot study using multi-urinary biomarkers. *Environ Int.* 66: 138-45. DOI:  
546 10.1016/j.envint.2014.02.003.
- 547 Föllmann W, Ali N, Blaszkewicz M, Degen GH. 2016. Biomonitoring of Mycotoxins in Urine: Pilot  
548 Study in Mill Workers. *J Toxicol Environ Health A.* 79: 1015-25. DOI:  
549 10.1080/15287394.2016.1219540.
- 550 Gerding J, Ali N, Schwartzbord J, Cramer B, Brown DL, Degen GH, Humpf HU. 2015. A  
551 comparative study of the human urinary mycotoxin excretion patterns in Bangladesh,  
552 Germany, and Haiti using a rapid and sensitive LC-MS/MS approach. *Mycotoxin Res.* 31:  
553 127-36. DOI: 10.1007/s12550-015-0223-9.
- 554 Gerding J, Cramer B, Humpf HU. 2014. Determination of mycotoxin exposure in Germany using an  
555 LC-MS/MS multibiomarker approach. *Mol Nutr Food Res.* 58: 2358-68. DOI:  
556 10.1002/mnfr.201400406.

- 557 Gratz SW, Richardson AJ, Duncan G, Holtrop G. 2014. Annual variation of dietary deoxynivalenol  
558 exposure during years of different Fusarium prevalence: a pilot biomonitoring study. *Food*  
559 *Addit Contam Part A Chem Anal Control Expo Risk Assess.* 31 (9), 1579-85.
- 560 Heyndrickx E, Sioen I, Huybrechts B, Callebaut A, De Henauw S, De Saeger S. 2015. Human  
561 biomonitoring of multiple mycotoxins in the Belgian population: Results of the BIOMYCO  
562 study. *Environ Int.* 84: 82-9. DOI: 10.1016/j.envint.2015.06.011.
- 563 Joint Expert Committee on Food and Additives (JECFA). 2010. Evaluation of certain food additives  
564 and contaminants. In Report of the Seventy-Second Meeting of the Joint FAO/WHO Expert  
565 Committee on Food Additives. In WHO Technical Report Series, edited by World Health  
566 Organization.
- 567 Mangoni AA, Jackson SH. 2004. Age-related changes in pharmacokinetics and pharmacodynamics:  
568 basic principles and practical applications, *Br J Clin Pharmacol.* 57: 6-14.  
569 DOI:10.1046/j.1365-2125.2003.02007.x.
- 570 Marin S, Ramos AJ, Cano-Sancho G, Sanchis V. 2013. Mycotoxins: occurrence, toxicology, and  
571 exposure assessment, *Food Chem Toxicol.* 60: 218-37. DOI: 10.1016/j.fct.2013.07.047.
- 572 Martone AM, Onder G, Vetrano DL, Ortolani E, Tosato M, Marzetti E, Landi F. 2013. Anorexia of  
573 aging: a modifiable risk factor for frailty. *Nutrients.* 5: 4126-33. DOI: 10.3390/nu5104126.
- 574 Njumbe Ediage E, Diana Di Mavungu J, Song S, Sioen I, De Saeger S. 2013. Multimycotoxin analysis  
575 in urines to assess infant exposure: a case study in Cameroon. *Environ Int.* 57-58: 50-9. DOI:  
576 10.1016/j.envint.2013.04.002.
- 577 Oldenburg E, Schittenhelm S. 2012. Effect of plant water deficit on the deoxynivalenol concentration  
578 in Fusarium-infected maize kernels. *Mycotoxin Res.* 28 (4), 229-36. DOI: 10.1007/s12550-  
579 012-0136-9.
- 580 Papageorgiou M, Wells L, Williams C, White K, De Santis B, Liu Y, Debegnach F, Miano B, Moretti  
581 G, Greetham S, Brera C. et al. 2018. Assessment of Urinary Deoxynivalenol Biomarkers in  
582 UK Children and Adolescents. *Toxins (Basel).* 10. DOI: 10.3390/toxins10020050.
- 583 Pestka JJ. 2010. Deoxynivalenol: mechanisms of action, human exposure, and toxicological  
584 relevance. *Arch Toxicol* 84: 663-79. DOI: 10.1007/s00204-010-0579-8.
- 585 Pestka JJ, Smolinski AT. 2005. Deoxynivalenol: toxicology and potential effects on humans. *J*  
586 *Toxicol Environ Health B Crit Rev.* 8: 39-69. DOI: 10.1080/10937400590889458.
- 587 Rodriguez-Carrasco Y, Molto JC, Manes J, Berrada H. 2014. Development of a GC-MS/MS strategy  
588 to determine 15 mycotoxins and metabolites in human urine. *Talanta.* 128: 125-31. DOI:  
589 10.1016/j.talanta.2014.04.072.
- 590 Scudamore, KA, Hazel CM, Patel S, Scriven F. 2009. Deoxynivalenol and other Fusarium  
591 mycotoxins in bread, cake, and biscuits produced from UK-grown wheat under commercial  
592 and pilot scale conditions. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess.*  
593 26: 1191-98. DOI: 10.1080/02652030902919426.
- 594 Shephard GS, Burger HM, Gambacorta L, Gong YY, Krska R, Rheeder JP, Solfrizzo M, Srey C,  
595 Sulyok M, Visconti A, et al. 2013. Multiple mycotoxin exposure determined by urinary  
596 biomarkers in rural subsistence farmers in the former Transkei, South Africa. *Food Chem*  
597 *Toxicol.* 62: 217-25. DOI: 10.1016/j.fct.2013.08.040.

- 598 Solfrizzo M, Gambacorta L, Visconti A. 2014. Assessment of multi-mycotoxin exposure in southern  
599 Italy by urinary multi-biomarker determination. *Toxins (Basel)*. 6: 523-38. DOI:  
600 [10.3390/toxins6020523](https://doi.org/10.3390/toxins6020523).
- 601 Talley NJ, O'Keefe EA, Zinsmeister AR, Melton LJ. 1992. Prevalence of gastrointestinal symptoms  
602 in the elderly: a population-based study. *Gastroenterology*. 102: 895-901. DOI:  
603 [10.1016/0016-5085\(92\)90175-X](https://doi.org/10.1016/0016-5085(92)90175-X).
- 604 Turner PC, Burley VJ, Rothwell JA, White KL, Cade JE, Wild CP. 2008. Deoxynivalenol: rationale  
605 for development and application of a urinary biomarker. *Food Addit Contam Part A Chem*  
606 *Anal Control Expo Risk Assess*. 25: 864-71. DOI: [10.1080/02652030801895040](https://doi.org/10.1080/02652030801895040).
- 607 Turner PC, Hopton RP, Lecluse Y, White KL, Fisher J, Lebailly P. 2010a. Determinants of urinary  
608 deoxynivalenol and de-epoxy deoxynivalenol in male farmers from Normandy, France. *J*  
609 *Agric Food Chem*. 58: 5206-12. DOI: [10.1021/jf100892v](https://doi.org/10.1021/jf100892v).
- 610 Turner PC, Hopton RP, Lecluse Y, White KL, Fisher J, Cade JE, Wild CP. 2011. Assessment of  
611 deoxynivalenol metabolite profiles in UK adults. *Food Chem Toxicol*. 49: 132-5. DOI:  
612 [10.1016/j.fct.2010.10.007](https://doi.org/10.1016/j.fct.2010.10.007).
- 613 Turner PC, Ji BT, Shu XO, Zheng W, Chow WH, Gao YT, Hardie LJ. 2011. A biomarker survey of  
614 urinary deoxynivalenol in China: the Shanghai Women's Health Study. *Food Addit Contam*  
615 *Part A Chem Anal Control Expo Risk Assess*. 28: 1220-3. DOI:  
616 [10.1080/19440049.2011.584070](https://doi.org/10.1080/19440049.2011.584070).
- 617 Turner PC, White KL, Burley VJ, Hopton RP, Rajendram A, Fisher J, Cade JE, Wild CP. 2010b. A  
618 comparison of deoxynivalenol intake and urinary deoxynivalenol in UK adults. *Biomarkers*.  
619 15: 553-62. DOI: [10.3109/1354750X.2010.495787](https://doi.org/10.3109/1354750X.2010.495787).
- 620 Wakimoto P, Block G. 2001. Dietary intake, dietary patterns, and changes with age: an  
621 epidemiological perspective. *J Gerontol A Biol Sci Med Sci*. 56: 65-80. DOI:  
622 [10.1093/gerona/56.suppl\\_2.65](https://doi.org/10.1093/gerona/56.suppl_2.65)
- 623 Wallin S, Gambacorta L, Kotova N, Lemming EW, Nalsen C, Solfrizzo M, Olsen M. 2015.  
624 Biomonitoring of concurrent mycotoxin exposure among adults in Sweden through urinary  
625 multi-biomarker analysis. *Food Chem Toxicol*. 83: 133-9. DOI: [10.1016/j.fct.2015.05.023](https://doi.org/10.1016/j.fct.2015.05.023).
- 626 Warth B, Sulyok M, Fruhmann P, Berthiller F, Schuhmacher R, Hametner C, Adam G, Frohlich J,  
627 Krska R. 2012. Assessment of human deoxynivalenol exposure using an LC-MS/MS based  
628 biomarker method. *Toxicol Lett*. 211: 85-90. DOI: [10.1016/j.toxlet.2012.02.023](https://doi.org/10.1016/j.toxlet.2012.02.023).
- 629 Wells L, Hardie L, Williams C, White K, Liu Y, De Santis B, Debegnach F, Moretti G, Greetham S,  
630 Brera C, et al. 2017. Deoxynivalenol Biomarkers in the Urine of UK Vegetarians. *Toxins*  
631 *(Basel)* 7:196. DOI: [10.3390/toxins9070196](https://doi.org/10.3390/toxins9070196).
- 632 Wells L, Hardie L, Williams C, White K, Liu Y, De Santis B, Debegnach F, Moretti G, Greetham S,  
633 Brera C, Rigby A, et al. 2016. Determination of Deoxynivalenol in the Urine of Pregnant  
634 Women in the UK. *Toxins (Basel)*. 8:306. DOI: [10.3390/toxins8110306](https://doi.org/10.3390/toxins8110306).
- 635 Wu Q, Dohnal V, Huang L, Kuca K, Yuan Z. 2010. Metabolic pathways of trichothecenes. *Drug*  
636 *Metab Rev*. 42: 250-67. DOI:[10.1080/03602530903125807](https://doi.org/10.1080/03602530903125807).
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639 **Table 1.** Baseline characteristics of elderly ( $\geq 65$  years) and adults (18-64 years) by sex

	Elderly ( $\geq 65$ years)		Adults (18-64 years)	
	Men (n=10)	Women (n=10)	Men (n=16)	Women(n=15)
<b>Height (cm)</b>	170.1 (5.9)*,†	163.1 (6.6)	177.6 (5.3)*	165.5 (5.7)
<b>Weight (kg)</b>	85.3 (13.3)*	67.1 (11.2)	91.1 (12.8)*	73.1 (12.1)
<b>BMI (kg/m<sup>2</sup>)</b>	29.6 (5.6)	25.2 (3.6)	28.8 (3.2)*	26.7 (3.9)
<b>Physical activity</b>				
<b>Sedentary (n, %)</b>	1 (10)	1(10)	0 (0)	0 (0)
<b>Light (n, %)</b>	0 (0)	2 (20)	6 (38)	2 (13)
<b>Moderate (n, %)</b>	8 (80)	7 (70)	8 (50)	12 (80)
<b>Heavy (n, %)</b>	1 (10)	0 (0)	1 (6)	1 (7)
<b>Exceptional (n, %)</b>	0 (0)	0 (0)	1 (6)	0 (0)
<b>Urine creatinine (mg/dl)</b>	110 (57)*	38 (24)‡	124 (88)	114 (84)

640 The Table includes adult data previously presented in Wells et al. 2017. \*,  $p < 0.05$ , significant different from women in the  
641 same age group; †,  $p < 0.05$ , significant different from adult men; ‡,  $p < 0.05$ , significant different from adult women.  
642

643 **Table 2.** Unadjusted and creatinine-adjusted DON and metabolites concentrations by day  
 644 and sex in elderly and adults.

Age Group	Day	Total DON		Free DON		DON-GlcA		DOM-1		
		1	2	1	2	1	2	1	2	
Elderly Men (n=10)	Incidence	90	90	70	60	90	90	nd	nd	
	%Total DON	N/A	N/A	17	9	83	91	nd	nd	
	Unadjusted (ng/mL)									
	Mean	26.7	20.1	5.0	2.2	21.7	17.8	nd	nd	
	Median	7.2	13.9	0.6	1.0	5.5	12.0	nd	nd	
	Min-Max	0-186	0-65.1	0-42.0	0-11.8	0-144	0-53.3	nd	nd	
	Adjusted (ng/mg creatinine)									
	Mean	22.2	28.0	3.8	3.2	18.5	21.8	nd	nd	
	Median	11.2	11.6	1.6	0.7	8.8	10.3	nd	nd	
	Min-Max	0-82.9	0-97.0	0.0-18.7	0.0-12.5	0-64.1	0-56.8	nd	nd	
Elderly Women (n=10)	Incidence	90	90	90	70	90	90	nd	nd	
	%Total DON	N/A	N/A	11	11	89	89	nd	nd	
	Unadjusted (ng/mL)									
	Mean	8.0	8.8	0.8	1.0	7.3	7.8	nd	nd	
	Median	6.2	8.1	0.9	0.8	5.7	7.8	nd	nd	
	Min-Max	0-28.8	0-17.5	0.1-1.8	0-3.3	0-27.0	0.0-14.2	nd	nd	
	Adjusted (ng/mg creatinine)									
	Mean	22.4	29.1	2.3	2.8	20.1	26.3	nd	nd	
	Median	15.7	24.3	2.0	2.5	13.8	22.7	nd	nd	
	Min-Max	0.0-42.5	0-81.0	1.1-5.7	0.0-7.8	0-39.9	0-76.3	nd	nd	
Adults Men (n=16)	Incidence	100	100	100	100	100	100	nd	nd	
	%Total DON	N/A	N/A	16	15	84	85	nd	nd	
	Unadjusted (ng/mL)									
	Mean	13.1	18.1	2.1	2.9	11.0	15.2	nd	nd	
	Median	11.9	12.6	1.8	1.9	7.1	11.0	nd	nd	
	Min-Max	2.2-19.4	5.1-58.8	0.3-4.2	0.5-13.8	1.8-26.8	4.3-45.0	nd	nd	
	Adjusted (ng/mg creatinine)									
	Mean	24.3	21.3	4.2	3.2	20.0	18.1	nd	nd	
	Median	10.9	14.4	1.8	2.1	9.8	12.2	nd	nd	
	Min-Max	0.5-153	2.5-62.3	0.1-31.5	0.4-12.7	0.4-122	2.1-55.4	nd	nd	
Adult Women (n=15)	Incidence	100	100	87	87	100	100	nd	nd	
	%Total DON	N/A	N/A	15	16	85	84	nd	nd	
	Unadjusted (ng/mL)									
	Mean	12.4	14.1	2.0	2.4	10.4	11.8	nd	nd	
	Median	10.7	10.1	1.8	1.3	9.0	9.6	nd	nd	
	Min-Max	3.3-40.6	0.9-36.0	0.3-4.2	0.1-8.6	2.9-31.9	0.7-27.4	nd	nd	
	Adjusted (ng/mg creatinine)									
	Mean	12.7	18.2	1.9	2.8	10.8	15.4	nd	nd	
	Median	11.3	11.3	1.6	2.5	10.0	10.5	nd	nd	
	Min-Max	3.9-27.8	1.0-66.2	0.2-4.7	0.2-9.3	3.3-23.1	0.8-56.8	nd	nd	

The Table includes adult data previously presented in Wells et al. 2017. Data are presented as mean, medium and min (minimum) - max (maximum); DON: deoxynivalenol; DON-GlcA: deoxynivalenol glucuronide; DOM-1: deoxy-deoxynivalenol; min: minimum; max: maximum; nd: not detected. LOQ for free DON was 0.25 ng/mL and for DON-GlcA and DOM-1 0.50 ng/mL.

**Table 3.** Consumption of food categories that commonly contribute to dietary DON exposure in elderly (n=20) and adults (n=30).

<b>Food category</b>	<b>Elderly (n = 20)</b>	<b>Adults (n=30)</b>
<b>Bread (g/d)</b>	101±49 (0-203)	89±60 (0-203)
<b>Flakes (g/d)</b>	22±16 (0-60)*	12±14 (0-50)
<b>Breakfast cereals (g/d)</b>	4±8 (0-23)	5±8 (0-30)
<b>Other breads (g/d)</b>	17±30 (0-90)	34±45 (0-135)
<b>Biscuits (g/d)</b>	15±15 (0-53)*	8±13 (0-60)
<b>Baked goods (g/d)</b>	28±32 (0-112)*	15±32 (0-112)
<b>Pizza (g/d)</b>	9±30 (0-128)	17±41 (0-128)
<b>Pasta (g/d)</b>	23±47 (0-115)	60±77 (0-230)
<b>Wheat germ (g/d)</b>	2±5 (0-15)	1±3 (0-15)
<b>Beer (g/d)</b>	43±105 (0-431)	91±153 (0-431)
<b>Pancakes (g/d)</b>	6±28 (0-124)	7±23 (0-93)
<b>Total (g/d)</b>	269±148 (65-625)	339±180 (45-658)

Data are presented as mean±1SD (range: minimum-maximum). Analysis was performed in 20 elderly and 30 adults with complete data. \*,  $p < 0.05$ , significant different from adults.