



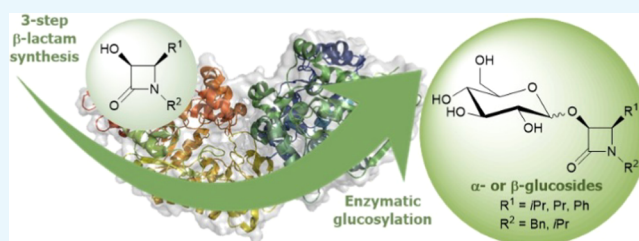
Chemoenzymatic Approach toward the Synthesis of 3-O-(α/β)-Glucosylated 3-Hydroxy- β -lactams

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Supporting Information

ABSTRACT: Glycosylation significantly alters the biological and physicochemical properties of small molecules. β -Lactam alcohols comprise eligible substrates for such a transformation based on their distinct relevance in the chemical and medicinal community. In this framework, the unprecedented enzymatic glycosylation of the rigid and highly strained four-membered β -lactam azaheterocycle was studied. For this purpose, *cis*-3-hydroxy- β -lactams were efficiently prepared in three steps by means of a classical organic synthesis approach, while a biocatalytic step was implemented for the selective formation of the corresponding 3-O- α - and - β -glucosides, hence overcoming the complexities typically encountered in synthetic glycochemistry and contributing to the increasing demand for sustainable processes in the framework of green chemistry. Two carbohydrate-active enzymes were selected based on their broad acceptor specificity and subsequently applied for the α - or β -selective formation of β -lactam-sugar adducts, using sucrose as a glucosyl donor.



INTRODUCTION

The very first β -lactam synthesis by Staudinger back in 1907,¹ followed by the accidental discovery of the antibacterial properties of penicillin 1 (penicillin G, R = Bn) by Fleming 20 years later,² marked the beginning of a revolution in the science of medicine (Figure 1). Up to now, penicillins and more recently discovered classes of β -lactam antibiotics are considered to be the major means of defense against pathogenic bacteria and represent the world's major biotechnology products.³ Over the years, besides their antibacterial properties, other pharmacological features of β -lactams were also unveiled (e.g., cholesterol absorption inhibition,⁴ inhibition of human tryptase,⁵ thrombin⁶ and chymase,⁷ vasopressin V1a antagonist activity,⁸ antiviral,⁹ antitumor,¹⁰ antidiabetic,¹¹ anti-inflammatory,¹² antiparkinsonian¹³ and antitubercular activity,¹⁴ etc.), which, in combination with their considerable synthetic importance as four-membered, highly strained cyclic amides,^{15,16} promoted β -lactams to become one of the most studied classes of azaheterocycles to date.

In general, monocyclic β -lactams unfairly comprise an underexplored subclass, though they are particularly interesting with respect to the treatment of bacterial infections (e.g., commercial 3-aminomonobactam aztreonam 2, Figure 1).¹⁷ 3-Hydroxy- β -lactams 3, such as glutamine synthetase-inhibiting tabtoxinine β -lactam 4, can also possess antimicrobial activity,^{18,19} whereas others have been proven to exhibit significant antitumor effects. For example, several 1,4-diary-

lactidin-2-ones (such as compound 5), which are conformationally restricted analogues of (*Z*)-combretastatin A-4 6, a natural product that inhibits tubulin polymerization, showed strong potencies against human neuroblastoma cell growth.¹⁰ Furthermore, the renowned " β -lactam synthon method" has been applied repeatedly to convert versatile 3-hydroxy- β -lactam precursors into a variety of highly functionalized scaffolds of biological and medicinal interest,¹⁵ such as the side chain of the anticancer drug taxol,²⁰ antibiotics,²¹ alkaloids,²² amino acids, peptides,²³ etc.

In the framework of the biological and synthetic relevance of the class of 3-hydroxy- β -lactams (3, Figure 1), we proposed to subject them to a glycosylation reaction, i.e. glycosylation with glucose.²⁴ Glycosylation can be defined as the attachment of a carbohydrate onto a chemical scaffold, known as the aglycone, via an O-, N-, S-, or C-atom. Hence, in this study, the β -lactam 3-hydroxy-entity serves as the anchor point. Glycosylation is a very common phenomenon in nature and probably concerns one of the most frequent and important modifications of biomolecules.²⁵ It is, for instance, believed to play a key role in cellular communication in plants through accumulation, storage, and transport of hydrophobic substances.²⁶ Moreover, glycosylation has numerous practical applications in synthesis, where it might act as a chiral source or lead to bioactive

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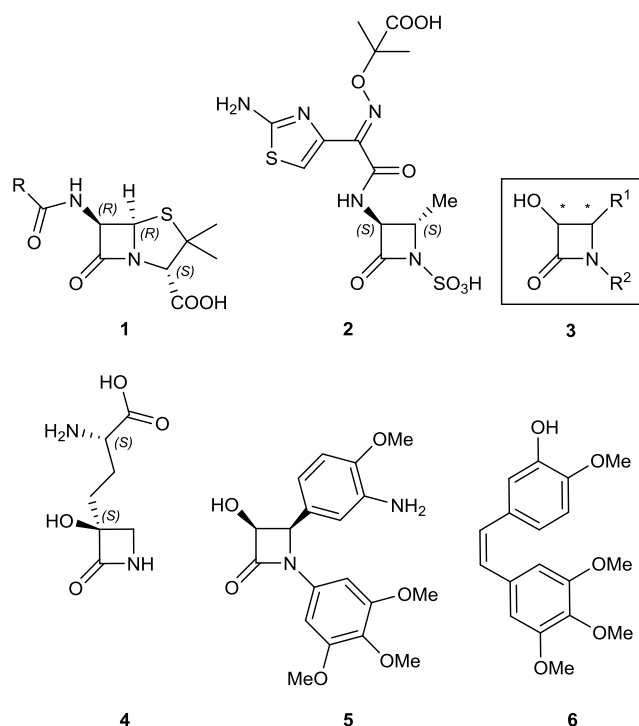


Figure 1. Bioactive (β -lactam-containing) compounds.

precursors,^{27–29} or in industry, where it can be applied to detergents and cosmetics (e.g., long alkyl chain aglycones),^{30,31} antifungal and antimicrobial compounds (e.g., unsaturated alkyl chain aglycones),³² antitumor formulations and cardiac-related drugs (e.g., peptide and steroid aglycones)^{32,33} and “controlled release” compounds (e.g., flavor and fragrance aglycones).³⁴

This endless series of examples arises from the influence glycosylation can exert on various physicochemical and biological properties. The most obvious advantage concerns the solubility and bioavailability increase of hydrophobic aglycones, augmenting their potential in pharmacological applications,²⁴ as is for example the case for glycopeptides and curcumin (20 million-fold better aqueous solubility of the latter).^{35,36} Besides the effect on solubility, glycosylation can be used to alter the bioactivity of the compound. Although so far not a widespread mechanism of antibiotic resistance, glycosylation does play a role in self-protection of antibiotic-producing organisms,³⁷ while bringing about a spectacular increase in activity, as is well illustrated by vancomycin, a glycopeptide effective against methicillin-resistant *Staphylococcus aureus*. By means of glycorandomization, which is a way to acquire a library of analogues by changing the carbohydrate moiety, several derivatives have been found to be up to 500 times more effective than vancomycin itself.³⁸ Related to this, and even more triggering for the purpose of this study, is the finding that phenolic *O*-glycosylated ezetimibe, a β -lactam with cholesterol absorption inhibitory activity, was found to be 4 times as potent as its corresponding aglycone **7** (Figure 2).³⁹ Moreover, glycosylation enables tuning of the oxidative and thermal stability, as is the case for vitamin C,⁴⁰ or the toxicity of the aglycone, illustrated by a reduced cardiotoxicity of anthracycline antibiotics.⁴¹

Undoubtedly, glycosides are of high commercial interest,²⁶ though their synthesis is still far from trivial. Chemical glycosylation reactions using these complex substrates typically

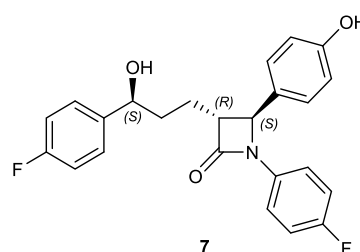


Figure 2. Ezetimibe **7**.

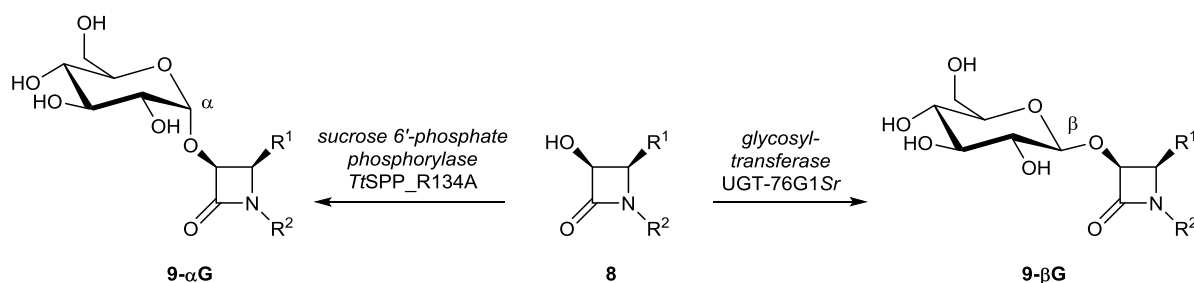
suffer from the lack of regioselectivity (primary, secondary, and hemiacetalic hydroxyl groups) and stereoselectivity with respect to the anomeric carbon center. Protection and deprotection steps have to be taken into account, involving additional synthetic manipulations, reagents, catalysts, solvents and hence toxic chemical waste streams, besides low overall yields. The development of economically and ecologically feasible procedures in the field of glycochemistry thus remains challenging.^{24,25,42} Biocatalytic routes have therefore received increasing attention as efficient and green alternatives, by circumventing the disadvantages associated with chemical glycosylation protocols.^{43,44} Enzymes are capable of regio-, stereo-, and enantioselectively glycosylating various acceptors under mild conditions using renewable starting materials, rendering additional synthetic steps and the concomitant requirements redundant. However, despite considerable efforts to improve enzymatic glycosylation reactions, no universal enzymatic glycosylation platform exists to date. Furthermore, the methodology still suffers from the lack of long-term enzymatic stability, acceptor solubility issues in the aqueous systems used and mostly expensive donors, hence hampering its broad use.^{24,45,46}

Yet, convinced by the assets of biocatalysis and its implementation in organic synthesis, as was also demonstrated by our preliminary studies on the chemoenzymatic syntheses of biologically relevant azaheterocyclic compounds,^{47,48} the enzymatic glycosylation of chemically prepared *cis*-3-hydroxy- β -lactams **8**, without affecting the sensitive four-membered ring system, is envisioned in the current work. In fact, enzymatic glycosylation of 3-hydroxy- β -lactams as challenging substrates based on their rigidity and steric bulk, has never been described before. As the configuration of the anomeric center is known to alter the biological properties of the molecule,^{49,50} the selective formation of both α - and β -glucosides **9** are contemplated, as presented in Scheme 1.

RESULTS AND DISCUSSION

In a first phase, the selective synthesis of the starting *cis*-3-hydroxy- β -lactams **8** was performed based on preceding research performed by our group, by means of three consecutive steps.⁵¹ Though reported more than 100 years ago, the Staudinger synthesis remains the most general and most frequently applied methodology for the preparation of β -lactams and derived compounds based on its simplicity in reaction procedures and predictability of the stereochemical outcome. The mechanism concerns a [2 + 2]-cyclocondensation between functionalized aldimines, obtained through imination of the corresponding aldehydes, and in situ generated ketenes.¹ To that end, primary amines **11** were condensed with aldehydes **10** in the presence of magnesium sulfate under various conditions, depending on the nature of the substrates (Scheme 2 and Table 1). The (*E*)-*N*-(alkyl/

Scheme 1. α - and β -Glucosylation of *cis*-3-Hydroxy- β -lactams **8 by a Mutant Glycoside Phosphorylase from *Thermoanaerobacterium thermosaccharolyticum* (TtSPP) or by a Glycosyltransferase from *Stevia rebaudiana*, Respectively**



Scheme 2. Synthesis of 3-Hydroxyazetidin-2-ones **8**

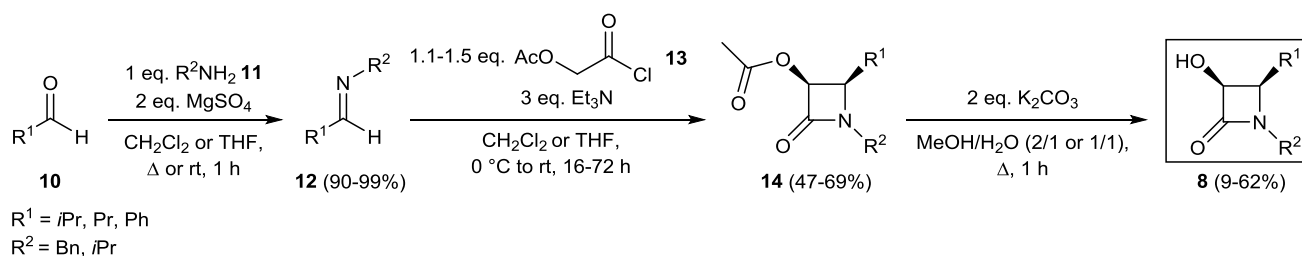


Table 1. Synthesis of 3-Hydroxyazetidin-2-ones **8**

Cpd	R^1	R^2	solvent, T	12 (%)	eq 13 , solvent, t	14 (%) (<i>cis/trans</i>) ^a	MeOH/H ₂ O	8 (%) ^d (<i>cis/trans</i>)
10a	<i>i</i> Pr	Bn	THF, Δ	99	1.3 eq 13 , CH_2Cl_2 , 16 h	47 ^b (91:9)	2:1	47 (100:0)
10b	Pr	<i>i</i> Pr	CH_2Cl_2 , rt	90	1.1 eq 13 , CH_2Cl_2 , 16 h	84 ^c (100:0)	1:1	9 ^c (100:0)
10c	Ph	Bn	THF, Δ	99	1.5 eq 13 , THF, 72 h	69 ^b (100:0)	1:1	62 (100:0)

^aDetermined via 1H NMR analysis of the crude reaction mixture. ^bAfter column chromatography (SiO_2). ^cCrude yield. ^dAfter recrystallization from EtOAc/hexane (1:1). ^eOverall yield from imine **12b**.

arylmethylidene)amines **12**, thus obtained were reacted with acetoxyacetyl chloride **13**, transformed in situ to the corresponding ketene through treatment with triethylamine, affording 3-acetoxyazetidin-2-ones **14**. The appropriate alkylic/arylic starting materials were selected ($R^1 = iPr, Pr, Ph$; $R^2 = Bn, iPr$), thus offering a variety of β -lactams with respect to steric bulk and polarity. Almost complete diastereoselectivity toward the *cis*- β -lactams **14** was observed, as deduced from the 1H NMR spectra ($CDCl_3$). The vicinal coupling constants J_{H3-H4} between the C3- and C4-protons of the azetidin-2-one ring systems varied between 4.6 and 5.0 Hz, values typical for *cis*-isomers,⁵² while for derivative **14a**, a minor amount (9%) of *trans*- β -lactam was formed as well ($J_{H3-H4} = 1.7$ Hz, $CDCl_3$). Pure *cis*- β -lactams **14** were obtained after column chromatography-mediated isolation on silica gel (except for alkyl-derivative **14b**, posing difficulties during purification and hence further used in a crude form) and subjected to alkaline conditions in a mixture of methanol and water (1:1–2:1) for deacetylation of the C3-group, thus releasing the premised free alcohols **8** for subsequent glycosylation without affecting the relative *cis*-stereochemistry.

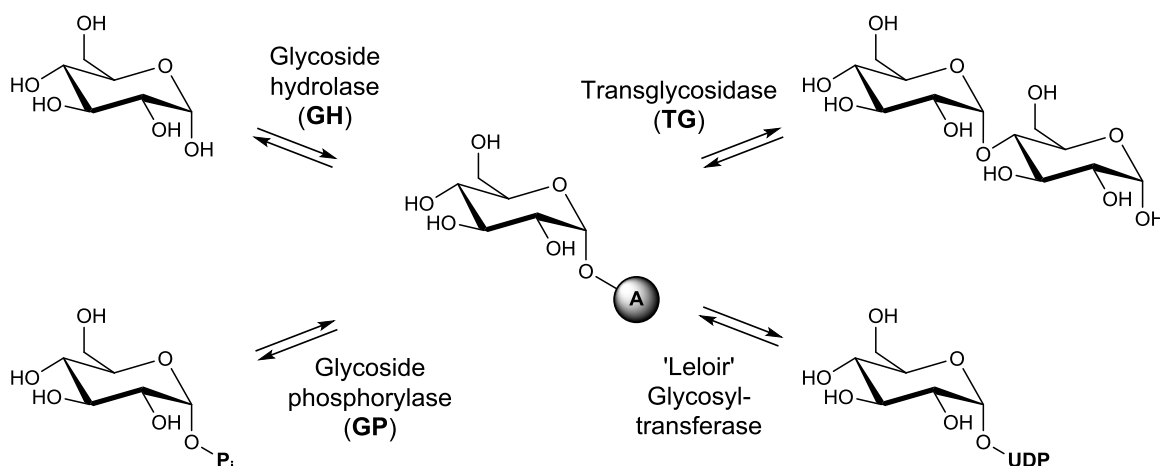
In the past century, organic chemists have developed various methods to glycosylate different acceptors, including hydroxylated β -lactams. Banik and co-workers led this research field when it comes to 3-*O*-glycosylazetidin-2-ones, more specifically the stereospecific glycosylation with glycals, cyclic enol ether derivatives of sugars. They reported an iodine-catalyzed α -glycosylation reaction of racemic 3-hydroxy- β -lactams with various 3-acetoxyglycals via the Ferrier rearrangement, providing access to an optically pure intermediate ((3*R*,4*S*)-

3-acetoxy-1-(*p*-anisyl)-4-phenylazetidin-2-one) for the preparation of the Taxol and Taxotere side chain by means of the optical resolution principle.⁵³ Toward the same objective, some research groups employed the Staudinger synthesis starting from protected glycosylated precursors of ketenes, equally affording 3-*O*- β -lactam-sugar adducts containing several chiral centers, which served as a source of chirality in asymmetric synthesis.^{54–56}

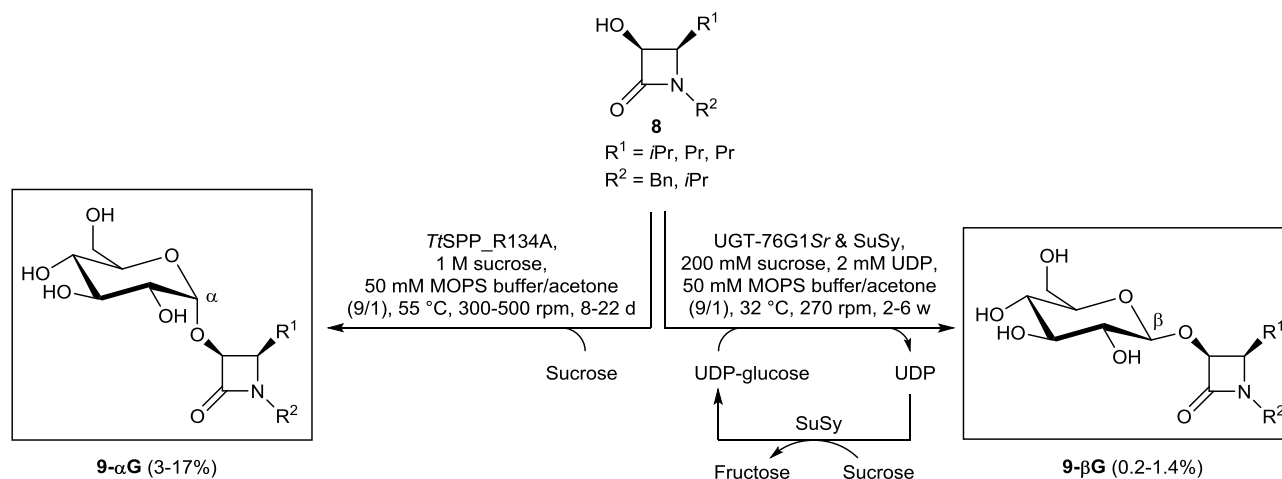
A variety of other chemical procedures are available enabling coupling of a sugar entity with a β -lactam at the C3-, C4-, or even the N1-position (or further away from the β -lactam nucleus) through carbon–carbon or carbon–heteroatom bond formation.⁵⁷ However, as mentioned before, these reactions typically suffer from low yields, low atom efficiencies, and excessive burden on the environment, making them barely viable from an economic and ecological point of view. Nonetheless, to be able to compare a chemical and enzymatic approach for the glycosylation of our specific substrates, the classical Koenigs–Knorr glycosylation was tested in this study for the attempted synthesis of 3-*O*- β -glucosyl- β -lactams **9**.⁵⁸ The mechanism comprises nucleophilic substitution of a glycosyl halide by an alcohol and still constitutes a preferred synthetic method in glycochemistry. Unfortunately, we encountered various obstacles when testing several glycosyl donors, acceptors, and reaction conditions, giving rise only to a minor amount of the expected glycoside and hence confirming the general conclusions reported in the literature with respect to chemical glycosylations.

On the other hand, many of the drawbacks of chemical glycosylation may be overcome by using biocatalysts.

Scheme 3. Carbohydrate-Active Enzymes Used in Glycosylation Reactions (A = Acceptor; P_i = Inorganic Phosphate; UDP = Uridine Diphosphate)²⁴



Scheme 4. Enzymatic Glucosylation of 3-Hydroxy- β -lactams **8** (*TtSPP_R134A*, Mutant R134A of the Sucrose 6'-Phosphate Phosphorylase from *T. thermosaccharolyticum*; UGT-76G1Sr, UDP-Glycosyltransferase from *S. rebaudiana*; SuSy, Sucrose Synthase from *Acidithiobacillus caldus*)



Nakayama and co-workers developed the first detailed synthesis of vancomycin and some derivatives through two enzymatic glycosylation steps,⁴⁵ while the enzymatic glucuronidation of ezetimibe **7** has been reported by Zaks and Dodds, in up to 92 and 88% isolated yield, respectively.⁵⁹ Although the synthesis of the donor substrates is somewhat demanding, careful comparison with their chemical syntheses, requiring tedious protection–deprotection steps of the glycosyl donors resulting in low overall yields, pointed to the enzymatic approach to be the most efficient one.^{39,60,61}

Four types of enzymes that are capable of catalyzing a glycosyl transfer can be distinguished (Scheme 3).⁶² The vast majority of glycosylation reactions in nature is performed by glycosyltransferases. Although these biocatalysts are highly efficient and provide excellent regio- and stereocontrol, their practical use is hampered due to the excessive cost of the nucleotide-activated carbohydrate donors they require. Transglycosidases are an attractive alternative in this regard, using cheap substrates such as sucrose and amylose, but the variety of different transglycosidase specificities that have been reported to date is rather limited. Finally, glycoside hydrolases and phosphorylases both perform a catabolic role in vivo, cleaving glycosidic bonds using water or phosphate,

respectively, but they can be exploited for synthetic purposes under specific process conditions as well.

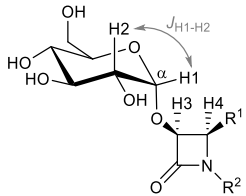
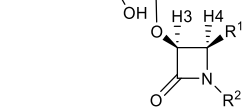
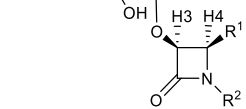
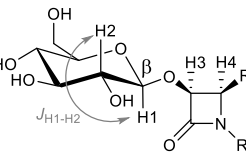
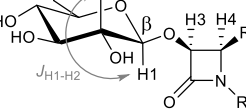
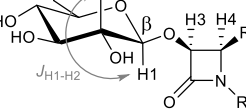
A critical property to consider when choosing a suitable candidate for catalyzing the envisioned glycosylation reactions is the substrate promiscuity. Indeed, acceptor sites that are strictly specific toward a narrow group of molecules will be less likely to bind the rather challenging *cis*-3-hydroxy- β -lactam targets. Therefore, two biocatalysts that are known to have a broad acceptor scope were selected. The sucrose 6'-phosphate phosphorylase from *T. thermosaccharolyticum* (*TtSPP*) is an appealing enzyme for the synthesis of α -glucosides considering the use of sucrose as a cheap and renewable donor substrate, and shows activity on a diverse range of small molecules with retention of the anomeric configuration.⁶³ Moreover, a mutant of *TtSPP* (R134A) was rationally designed previously in our laboratory to widen the entrance to the active site, triggering affinity for bulky polyphenols such as resveratrol.⁶⁴ The selected option for the synthesis of β -bonded adducts concerns the stevioside glucosyltransferase from *S. rebaudiana* (UGT-76G1Sr), which has been employed successfully on structures such as branched alcohols, flavonoids, terpenoids, and more.⁶⁵ Although its need for the expensive donor substrate uridine diphosphate glucose (UDP-glucose) is a major drawback, this

Table 2. Enzymatic Glucosylation of 3-Hydroxyazetidin-2-ones **8**

Cpd	R ¹	R ²	reaction conditions	conversion (%) ^d	9 (%)
α -glucosylation					
8a	ⁱ Pr	Bn	21 mM 8a , 0.06 mg/mL <i>Tt</i> SPP-R134A, 8 d	23	12 ^a
8b	Pr	ⁱ Pr	15 mM 8b , 0.04 mg/mL <i>Tt</i> SPP-R134A, 22 d	50	17 ^b
8c	Ph	Bn	15 mM 8c , 0.04 mg/mL <i>Tt</i> SPP-R134A, 20 d	12	3 ^b
β -glucosylation					
8a	ⁱ Pr	Bn	15 mM 8a , 0.13 mg/mL UGT-76G1 <i>Sr</i> , 0.36 mg/mL SuSy, 6 weeks	9	0.2 ^c
8b	Pr	ⁱ Pr	15 mM 8b , 0.19 mg/mL UGT-76G1 <i>Sr</i> , 0.31 mg/mL SuSy, 2 weeks	14	1.4 ^c
8c	Ph	Bn	15 mM 8c , 0.11 mg/mL UGT-76G1 <i>Sr</i> , 0.31 mg/mL SuSy, 2 weeks	4	1 ^c

^aAfter column chromatography (SiO₂). ^bAfter column chromatography (SiO₂) and prep. thin-layer chromatography (TLC). ^cAfter two-step column chromatography (SiO₂ & C₁₈). ^dDetermined via liquid chromatography–mass spectrometry (LC–MS)-analysis of the crude reaction mixture.

Table 3. Chemical Shift of Glycosidic Anomeric Carbon, Proton, and Vicinal Coupling Constant J_{H1-H2} (¹H and ¹³C NMR, D₂O)

α -Glucoside	R ¹	R ²	dr^a	δ_{CH1} (ppm)	J_{H1-H2} (Hz)	δ_{CH1} (ppm)
	<i>i</i> Pr	Bn	100/0	5.21 (d)	3.6	98.5
	Pr	<i>i</i> Pr	100/0	5.18 (d)	3.6	98.2
	Ph	Bn	100/0	4.97 (d)	4.0	101.6
β -Glucoside						
	<i>i</i> Pr	Bn	44/56	4.52 (d) 4.70 (d)	7.9 7.8	102.3 102.6
	Pr	<i>i</i> Pr	36/64	4.50 (d) 4.66 (d)	8.1 8.1	102.2 102.4
	Ph	Bn	41/59	4.10 (d) 4.52 (d)	8.4 8.4	101.1 102.6

^aDetermined via ¹H NMR analysis of purified glycosides.

problem is circumvented in this work by coupling the reaction to a sucrose synthase (SuSy).⁶⁶ Hence, UDP-glucose is regenerated from sucrose in situ while conveniently recycling the UDP byproduct of the transferase reaction (Scheme 4).

The solubility of the β -lactam acceptors **8** is quite poor in the aqueous solutions that these enzymes prefer. Therefore, the performance of various cosolvents (10% v/v) was assessed in small-scale glucosylation reactions. 2-Methylbutan-2-ol and acetonitrile appeared to be toxic to the enzymes, while 2-phenylethanol and isobutanol were unsuitable as they were found to act as acceptor substrates as well, resulting in side products. The desired glucoside was present in highest quantity when diethyl ether, acetone, methanol, dimethyl sulfoxide (DMSO), or methyl *t*-butyl ether were added. Acetone was finally selected as the most appropriate cosolvent due to its low toxicity, high volatility, and the absence of potentially competing hydroxyl groups. Additionally, the optimal acceptor concentration was determined for both the α - and β -glucosylation reactions. The highest conversions were observed at a concentration of 15 mM in all cases, except for the reaction of **8a** with *Tt*SPP_R134A (21 mM).

The α - and β -glucosylation of all three β -lactam acceptors were carried out at a preparative scale (0.375–2.1 mmol acceptor **8**) under these optimized conditions, at several degrees below the enzymes' optimal temperature to slow down irreversible denaturation. All glucosides were successfully

synthesized, although the reactions proceeded very slowly and took several days to reach a practical degree of conversion (Table 2). As a comparison, almost complete conversion of resveratrol could be achieved within 24 h with these enzymes.^{63,64} This difference may be explained by the rigid and bulky structure of the current compounds, which could cause steric clashes in the active site. Finally, after inactivation and removal of the enzymes, the target compounds **9** could be recovered and purified by means of (a sequence of) chromatographic techniques, which resulted in rather low yields. However, it should be noted that the main focus of this research was to obtain and isolate the glucosides in an analytically pure form to allow their spectroscopic analysis, hence the elaborate purification procedures to remove all impurities.

The successful synthesis of *cis*-3-O-D-glucopyranosyl-3-hydroxyazetidin-2-ones **9- α G** and **9- β G** was confirmed on the basis of the characteristic spectral data and their accordance with reported literature values (Scheme 4, Tables 2 and 3).⁶⁷ The typical anomeric proton signals are located as a doublet around δ 4.97–5.21 and 4.10–4.70 ppm for the α - and β -glucosides, respectively, and can easily be distinguished from the nearby doublet signals of the protons located on the β -lactam ring ($J = \pm 5$ Hz) (¹H NMR, D₂O). A clear difference is to be noticed between the respective values of the vicinal coupling constants J_{H1-H2} between the anomeric H1 and its

neighboring H2 proton. The values of 3.6–4.0 Hz correspond to α -glucosides, whereas β -glucosides show larger coupling constants of 7.8–8.4 Hz, thus, confirming the expected stereoselective formation of α - and β -isomers by *TtSPP_R134A* and *UGT-76G1Sr*, respectively. Finally, the glycosidic anomeric carbon signals are observable as a singlet at δ 98.2–101.6 and 101.1–102.6 ppm, respectively (^{13}C NMR, D_2O).

Upon coupling of racemic *cis*-3-hydroxy- β -lactams **8** with optically pure glucose, two *cis*-diastereomers (*R,R*)-**9-G** and (*S,S*)-**9-G** might be expected as enzymes are chiral catalysts and might therefore discriminate between either of the two enantiomeric acceptor substrates. Indeed, from the analysis of the NMR data presented in Table 3, it becomes clear that one single product is obtained after α -glucosylation with *TtSPP_R134A*. On the other hand, a very low enantioselectivity was observed in the formation of two isomeric β -glucosides **9- β G** using the *UGT-76G1Sr/SuSy* system (*dr* = 36:64–44:56, as determined via ^1H NMR). It should be noted, however, that these values are determined after the isolation of glycosides, implying the possibility that the recovery of the two diastereomers during purification might not be equal. On the basis of the low concentration of product, though, compared to the amount of non-reacted starting material, sucrose, fructose, UDP, and UDP-glucose in the crude mixture, assessment of the diastereomeric ratios based on the crude NMR data is not reliable and practically infeasible.

In the literature, some examples of glycosyltransferases catalyzing glycosylation reactions with high enantioselectivity are reported, though the reaction conditions such as temperature, (co-)solvent, enzyme formulation, additives, etc. clearly affect the isomeric ratios obtained.⁶⁸ Similarly, the β -glucosylation of compound **8a** at 32 °C with acetone as a cosolvent was found to furnish a more or less equimolar mixture of isomers, whereas a higher selectivity was observed upon incubation at 40 °C in combination with DMSO (*dr* = 44/56 and 71/29, respectively). A more detailed study on the influence of reaction conditions on the enantioselectivity of transferase-catalyzed transformations has not been published yet and could thus constitute a suitable topic for prospective research. Anyhow, it can be concluded that α -glucosylation of *cis*-3-hydroxy- β -lactams **8** with *TtSPP_R134A* takes place with a high enantioselectivity, and thus forms an efficient route toward optically pure α -glucosides **9- α G**, while opening up the opportunity to synthesize the corresponding chiral alcohols **8**.

CONCLUSIONS

Glycosylation can have a profound impact on a compound's solubility, activity, and bioavailability. Preparing glycosylated molecules is, however, far from trivial. In this work, three *cis*-3-hydroxy- β -lactams were chemically synthesized via three straightforward steps, after which the potential of two carbohydrate-active enzymes for the production of their corresponding α - or β -glucosides was evaluated. These enzymes, which were selected based on their somewhat promiscuous nature, successfully catalyzed the intended reactions, albeit with low activity in comparison to the reported activities on other, less demanding substrates. However, the limited reaction rate is not surprising considering the bulky and rigid structure of these β -lactams, and was nonetheless sufficient to enable the production of six novel glucosides at a preparative scale and their subsequent isolation in analytically pure form. These findings highlight the potential

of the *T. thermosaccharolyticum* mutant phosphorylase and the *S. rebaudiana* glycosyltransferase as promising catalysts for the glycosylation of challenging molecules in general, and of *cis*-3-hydroxy- β -lactams in particular. Furthermore, the glucosyl moiety is transferred from the cheap and renewable sucrose with superior regio- and stereoselectivity (and even enantioselectivity for α -glucosylation) without the need for any protection or deprotection steps. Overall, the complimentary use of classical organic synthesis and biocatalysis for glycosylation reactions was once again shown to provide major advantages within the framework of green chemistry.

EXPERIMENTAL PART

General Methods. Commercially available solvents and reagents were purchased from common chemical suppliers and used without further purification. ^1H NMR and ^{13}C NMR spectra were recorded at 400 and 100 MHz (Bruker Avance III-400), respectively, in deuterated solvents with tetramethylsilane as the internal standard. IR spectra were obtained from samples in neat form with an attenuated total reflectance (ATR) accessory with a PerkinElmer Spectrum BX FT-IR or a Shimadzu IRAffinity-1S WL FT-IR spectrophotometer. Melting points were measured using a Kofler bench, type WME Heizbank of Wagner & Munz. High-performance liquid chromatography (HPLC) and HPLC-MS analyses were performed on an Agilent 1200 series HPLC system fitted with an Ascentis Express C_{18} column (particle size 2.7 μm , length 30 mm, internal diameter 4.6 mm) and connected to a UV–vis detector and an Agilent 1100 series LC/MSD-type SL mass spectrometer (ESI, 70 eV) using a mass-selective single-quadrupole detector. A mixture of acetonitrile/water (5 mM NH_4OAc) was used as the eluent. High-resolution electron spray (ES-TOF) mass spectra were obtained with Agilent Technologies 6210 Series Time of Flight. Automated column chromatography was performed on a Büchi Reveleris X2 flash chromatography system (normal phase) or Grace Reveleris X1 flash chromatography system (reversed phase), using Reveleris C_{18} or Reveleris silica cartridges. Centrifugation for harvesting cultures or performing buffer exchange was carried out in a Sorvall RC-6 Plus rotator. Cell extracts were sonicated in a Branson 250 sonifier (output control 3, 50% duty cycle).

Synthesis of 3-Acetoxyazetididin-2-ones **14.** The synthesis of *cis*-3-acetoxy-1-benzyl-4-phenylazetididin-2-one **14c** serves as a general example for the synthesis of 3-acetoxyazetididin-2-ones **14a**, **14b** (for details, see Table 1 of section “Results and Discussion”).

In a flask of 500 mL, 9.75 g (50 mmol) of (*E*)-*N*-(benzylidene)benzylamine **12c**, synthesized according to the literature,⁵¹ was dissolved in 120 mL dry tetrahydrofuran (THF), followed by the addition of 20.9 mL (150 mmol; 3 equiv) triethylamine. After cooling the reaction mixture to 0 °C using an ice bath, a solution of 8.0 mL (75 mmol; 1.5 equiv) of acetoxyacetyl chloride **13** in 30 mL of dry THF was added dropwise. This mixture was then stirred for 16 h at room temperature, whereupon the solvent was evaporated under reduced pressure. The residue was dissolved in dichloromethane (300 mL), washed twice with 360 mL water, which on its turn was extracted with 150 mL dichloromethane. Next, the combined organic phases were dried over magnesium sulfate. Filtration of the drying agent, evaporation of the solvent, and flash chromatography with petroleum ether/ethyl acetate (2:1) eventually yielded 10.18 g (69%) *cis*-3-acetoxy-1-benzyl-4-phenylazetididin-2-one **14c**.

cis-3-Acetoxy-1-benzyl-4-isopropylazetid-2-one 14a. The spectral data are derived from the mixture of *cis*- and *trans*-isomers (dr = 91:9). Only the major isomer (*cis*) is described.

¹H NMR (400 MHz, CDCl₃): δ 0.83 and 0.97 (2 × 3H, 2 × d, *J* = 6.8 Hz, (CH₃)₂CH); 1.95 (1H, ~octet, *J* = 6.8 Hz, (CH₃)₂CH); 2.14 (3H, s, CH₃C=O); 3.44 (1H, d × d, *J* = 6.8, 5.0 Hz, OCHCH); 4.11 and 4.86 (2 × 1H, 2 × d, *J* = 15.1 Hz, N(HCH)); 5.89 (1H, d, *J* = 5.0 Hz, CHO); 7.21–7.23 and 7.29–7.38 (2H and 3H, 2 × m, CH_{arom}). ¹³C NMR (100 MHz, CDCl₃): δ 19.2 and 19.4 ((CH₃)₂CH); 20.7 (CH₃C=O); 28.4 ((CH₃)₂CH); 46.0 (NCH₂); 62.9 (OCHCH); 74.5 (CHO); 128.0, 128.2, and 128.9 (5 × HC_{arom}); 135.1 (C_{arom,quat}); 166.1 (NC=O); 169.6 (CH₃C=O). MS (70 eV): *m/z* (%) 262 (M⁺ + 1, 24); 284 (M⁺ + Na, 27). High resolution electrospray ionization mass spectrometry (HRMS (ESI)) calculated for C₁₅H₂₀NO₃⁺: 262.1438 [M + H]⁺, found: 262.1440. Yellow oil. *R*_f = 0.16 (PE/EtOAc 4:1). Yield after column chromatography (SiO₂): 47%.

cis-3-Acetoxy-1-isopropyl-4-propylazetid-2-one 14b. β-Lactam **14b** was immediately used in the next reaction step, as purification proved to be impossible at this stage.

cis-3-Acetoxy-1-benzyl-4-phenylazetid-2-one 14c. The spectral data are in accordance with those reported in the literature.⁶⁹

Synthesis of 3-Hydroxyazetid-2-ones 8. The synthesis of *cis*-1-benzyl-3-hydroxy-4-phenylazetid-2-one **8c** serves as an example for the synthesis of 3-hydroxyazetid-2-ones **8a** and **8b**.

In a flask of 50 mL, 1.62 g (5.5 mmol) of *cis*-3-acetoxy-1-benzyl-4-phenylazetid-2-one **14c** was dissolved in 30 mL of a 1:1-solution of methanol and water (2:1 for **8a**). Potassium carbonate (1.52 g, 11 mmol; 2 equiv) was added and the reaction mixture was stirred for 1 h at reflux temperature. Hereafter, methanol was evaporated and an extraction was performed with 50 mL diethyl ether. The organic phase was washed twice with 50 mL water, whereupon the aqueous phase was extracted twice with 50 mL diethyl ether. After drying of the combined organic phases using magnesium sulfate, filtration of the drying agent, evaporation of the solvent and recrystallization from a mixture of ethyl acetate and hexane (1:1), 0.86 g (62%) *cis*-1-benzyl-3-hydroxy-4-phenylazetid-2-one **8c** was obtained.

cis-1-Benzyl-3-hydroxy-4-isopropylazetid-2-one 8a. ¹H NMR (400 MHz, CDCl₃): δ 0.94 and 1.01 (2 × 3H, 2 × d, *J* = 6.9 Hz, (CH₃)₂CH); 2.06 (1H, ~octet, *J* = 6.9 Hz, (CH₃)₂CH); 3.28 (1H, d × d, *J* = 6.9, 5.2 Hz, OCHCH); 4.09 (1H, d, *J* = 5.9 Hz, OH); 4.11 and 4.81 (2 × 1H, 2 × d, *J* = 15.1 Hz, N(HCH)); 4.88 (1H, d × d, *J* = 5.9, 5.2 Hz, CHO); 7.20–7.22 and 7.27–7.37 (2H and 3H, 2 × m, CH_{arom}). ¹³C NMR (100 MHz, CDCl₃): δ 19.4 and 19.9 ((CH₃)₂CH); 28.5 ((CH₃)₂CH); 45.8 (NCH₂); 64.6 (OCHCH); 75.9 (CHO); 127.8, 128.2, and 128.8 (5 × HC_{arom}); 135.4 (C_{arom,quat}); 170.8 (C=O). MS (70 eV): *m/z* (%) 220 (M⁺ + 1, 100). HRMS (ESI) calculated for C₁₃H₁₈NO₂⁺: 220.1332 [M + H]⁺, found: 220.1334. White crystals. Yield after recrystallization from EtOAc/hexane (1:1): 47%. *T*_m = 76 °C.

cis-3-Hydroxy-1-isopropyl-4-propylazetid-2-one 8b. The spectral data are in accordance with those reported in the literature.¹

cis-1-Benzyl-3-hydroxy-4-phenylazetid-2-one 8c. The spectral data are in accordance with those reported in the literature.⁷⁰

Enzyme Production and Purification. The genes for mutant R134A of the *T. thermosaccharolyticum* sucrose 6'-phosphate phosphorylase (*TtSPP_R134A*; UniProt ID D9TT09), the *A. caldus* sucrose synthase (*SuSy*; UniProt ID A0A059ZV61) and the *S. rebaudiana* UDP-glucosyltransferase (*UGT-76G1Sr*; UniProt ID Q6VAB4) were ligated into expression vectors, transformed in *Escherichia coli* and expressed recombinantly as described in earlier work.^{64,65,71}

Cells containing the expressed proteins were harvested by centrifugation (20 000g, 5 min) and frozen overnight at –20 °C. Intracellular proteins were extracted by dissolving 5 g of frozen pellet in 25 mL of lysis buffer consisting of 50 mM 3-(*N*-morpholino)propanesulfonic acid (MOPS), 100 μM phenylmethylsulfonyl fluoride, 1 mg/mL lysozyme, pH 7.0. In the case of glucosyltransferase, MOPS was replaced by 50 mM sodium phosphate and 500 mM NaCl, pH 7.4. After incubation on ice for 30 min, the extract was exposed three times for 3 min of sonication and centrifuged to remove the insoluble fraction (20 000g, 30 min, 4 °C). Extracts containing sucrose synthase were used as such, while those containing sucrose 6'-phosphate phosphorylase were subjected to heat treatment (60 °C, 1 h) and centrifuged again. Extracts containing the glucosyltransferase were further purified by means of nickel–nitrilotriacetic acid affinity chromatography in gravity-flow columns as described by the supplier (MCLab). Subsequently, the buffer was exchanged to 50 mM MOPS, pH 7.0, in 30 kDa Amicon Ultra centrifugal filters (Merck).

The protein concentration was determined in triplicate using the BCA Protein Assay kit (Pierce) with bovine serum albumin as a standard.

Enzyme Activity Determination. The activity of *TtSPP_R134A* and *SuSy* was confirmed by performing reactions where sucrose is broken down (100 mM sucrose and 100 mM phosphate for *TtSPP_R134A*; 200 mM sucrose and 5 mM UDP for *SuSy*; 55 °C) and following the release of reducing sugars using the BCA method. The color reagent was prepared by combining 23 parts of a solution containing 1.5 g/L 4,4'-dicarboxy-2,2'-biquinoline dipotassium salt and 62.3 g/L anhydrous Na₂CO₃, one part of a solution composed of 23 g/L aspartic acid, 33 g/L anhydrous Na₂CO₃, and 7.3 g/L CuSO₄ and six parts ethanol. The reaction samples (25 μL) were incubated with 150 μL color reagent for 30 min at 70 °C. Absorbance was measured at 560 nm.

The activity of *UGT-76G1Sr* was confirmed through a colorimetric assay, where UDP released from UDP-glucose is converted to UTP by pyruvate kinase (PK) using phosphoenolpyruvate (PEP). This reaction generates pyruvate as well, which is in turn reduced to lactate while oxidizing NADH to NAD⁺ by lactate dehydrogenase (LDH).⁷² The assay mixture consisted of 100 times diluted PK/LDH enzymes from rabbit muscle (900–1400 U/mL LDH, 600–100 U/mL PK), 5 mM MgCl₂, 1.6 mg/mL bovine serum albumin, 0.3 mM NADH, and 0.3 mM PEP. A solution containing *UGT-76G1Sr* and substrates (100 μL; 10 mM UDP-glucose, 20 mM stevioside) was added to the assay mixture (100 μL) and absorbance at 340 nm was measured continuously.

Small-Scale Reactions and Optimization of Reaction Conditions. Small-scale reactions were carried out at 200 μL scale in Eppendorf tubes. The α-glucosylation reactions contained 1 M sucrose, 15 mM acceptor, 10% (v/v) organic solvent, 0.1 mg/mL *TtSPP_R134A*, and 50 mM MOPS (pH 7.0), and were incubated at 55 °C. The β-glucosylation reactions contained 200 mM sucrose, 15 mM acceptor, 10%

(v/v) organic solvent, 0.2 mg/mL UGT-76G1Sr, 1 mg/mL SuSy, and 50 mM MOPS (pH 7.0), and were incubated at 32 °C. A 1 μ L sample was taken after 24 h, spotted on silica gel 60 F₂₅₄ precoated plates (Merck) and analyzed by thin layer chromatography using a mixture of EtOAc/MeOH/H₂O (30:5:4) as the eluent. Spots were visualized by charring with 10% (v/v) H₂SO₄.

Reactions were optimized by assessing various solvents, i.e., 2-phenylethanol, methanol, isobutanol, 2-methylbutan-2-ol, acetone, methyl *t*-butyl ether, diethyl ether, and acetonitrile. Additionally, the acceptor concentration was varied (5–30 mM).

Synthesis of 3-O- α -D-Glucopyranosyl-3-hydroxyazetid-2-ones 9a-G and Downstream Processing. The production of α -glucosides 9a-G, 9b-G, and 9c-G was carried out using mutant *TtSPP_R134A* at a scale of 100, 25, or 50 mL, respectively. Each reaction contained 1 M sucrose, 10 v/v% acetone, 0.04–0.06 mg/mL *TtSPP_R134A*, 50 mM MOPS buffer (pH 7.0), and 21 mM of acceptor 8a or 15 mM of acceptor 8b or 8c. These reaction samples were incubated for 8–22 days at 55 °C with a reflux condenser, and stirred magnetically (IKA RCt Classic). The reaction samples (50 μ L) were inactivated by adding 50 μ L acetonitrile and 50 μ L ultrapure water, followed by centrifugation. These samples were analyzed by LC-MS (eluent: 10–100% CH₃CN in water (5 mM NH₄OAc)).

The α -glucosides were first purified by double extraction with isopropyl alcohol, to which NaCl was added (300 mg/mL) to obtain a biphasic system. The organic phase was washed with water to remove any remaining sugars, followed by evaporation under reduced pressure. The remaining product was dissolved in methanol, filtered for the removal of leftover salts and evaporated again. Subsequently, automatic column chromatography was performed (gradual increase in polarity of the eluent: petroleum ether/EtOAc (100:0) to petroleum ether/EtOAc (0:100) to EtOAc/MeOH/H₂O (30:5:4)). Minor impurities were finally removed via preparative TLC.

***cis*-1-Benzyl-3-O- α -D-glucopyranosyl-3-hydroxy-4-isopropylazetid-2-one 9a-G.** ¹H NMR (400 MHz, D₂O): δ 0.88 and 0.93 (2 \times 3H, 2 \times d, *J* = 6.8 Hz, (CH₃)₂CH); 2.07 (1H, octet, *J* = 6.8 Hz, (CH₃)₂CH); 3.39 (1H, d \times d, *J* = 9.9, 9.1 Hz, CHCHCH₂OH); 3.56 (1H, d \times d, *J* = 9.9, 3.6 Hz, CHCH₂); 3.56–3.72 (4H, m, CHⁱPr and CHCHCH(HCH)OH); 3.78 (1H, d \times d, *J* = 12.3, 2.2 Hz, O(HCH)); 4.27 and 4.68 (2 \times 1H, 2 \times d, *J* = 15.6 Hz, N(HCH)); 4.97 (1H, d, *J* = 5.0 Hz, CHCHⁱPr); 5.21 (1H, d, *J* = 3.6 Hz, CH₂); 7.26–7.28 and 7.31–7.41 (2H and 3H, 2 \times m, CH_{arom}). ¹³C NMR (100 MHz, D₂O): δ 18.3 and 19.2 ((CH₃)₂CH); 27.7 ((CH₃)₂CH); 45.7 (NCH₂); 60.4 (OCH₂); 64.1 (CHⁱPr); 69.4 (CHCHCH₂OH); 71.1 (CHCH₂); 72.7 and 72.8 (CHCHCHCH₂OH); 77.7 (CHCHⁱPr); 98.5 (CH₂); 128.0, 128.1 and 129.0 (5 \times HC_{arom}); 135.0 (C_{arom,quat}); 170.9 (C=O). IR (ATR, cm⁻¹): ν_{OH} = 3348; $\nu_{C=O}$ = 1730; ν_{max} = 2916, 2849, 2322, 1406, 1078, 1032, 924, 702. MS (70 eV): *m/z* (%) 404 (M⁺ + Na, 39); 382 (M⁺ + 1, 100). HRMS (ESI): calculated for C₁₉H₂₈NO₇⁺: 382.1860 [M + H]⁺, found: 382.1862. Yellow oil. *R*_f = 0.5 (EtOAc/MeOH/H₂O 30:5:4). Yield after normal phase column chromatography (SiO₂): 12%.

***cis*-3-O- α -D-Glucopyranosyl-3-hydroxy-1-isopropyl-4-propylazetid-2-one 9b-G.** ¹H NMR (400 MHz, D₂O): δ 0.91 (3H, t, *J* = 7.4 Hz, CH₂CH₃); 1.19 and 1.21 (2 \times 3H, 2 \times

d, *J* = 6.8 Hz, CH(CH₃)₂); 1.35 (2H, sextet, *J* = 7.4 Hz, CH₂CH₃); 1.71–1.77 (2H, m, CH₂CH₂CH₃); 3.39 (1H, ~t, *J* = 9.6 Hz, CHCHCH₂OH); 3.54 (1H, d \times d, *J* = 9.6, 3.6 Hz, CHCH₂); 3.57–3.77 (4H, m, CH(CH₃)₂ and CHCHCH(HCH)OH); 3.80 (1H, d \times d, *J* = 12.3, 2.1 Hz, O(HCH)); 3.99 (1H, ~q, *J* = 5.4 Hz, CHCHPr); 4.88 (1H, d, *J* = 5.4 Hz, CHCHPr); 5.18 (1H, d, *J* = 3.6 Hz, CH₂). ¹³C NMR (100 MHz, D₂O): δ 13.5 (CH₂CH₃); 18.8 (CH₂CH₃); 18.9 and 20.7 ((CH₃)₂CH); 30.7 (CH₂CH₂CH₃); 44.9 ((CH₃)₂CH); 57.6 (CHCHPr); 60.4 (OCH₂); 69.3 (CHCHCH₂OH); 71.2 (CHCH₂); 72.67 and 72.70 (CHCHCHCH₂OH); 76.7 (CHCHPr); 98.2 (CH₂); 169.3 (C=O). IR (ATR, cm⁻¹): ν_{OH} = 3339; $\nu_{C=O}$ = 1726; ν_{max} = 2922, 2851, 2108, 1452, 1155, 1103, 1024, 627. MS (70 eV): *m/z* (%) 356 (M⁺ + Na, 100); 334 (M⁺ + 1.59). *R*_f = 0.3 (EtOAc/MeOH/H₂O 30:5:4). Yield after normal phase column chromatography and preparative TLC (SiO₂): 17%.

***cis*-1-Benzyl-3-O- α -D-glucopyranosyl-3-hydroxy-4-phenylazetid-2-one 9c-G.** ¹H NMR (400 MHz, D₂O): δ 1.84–1.88 and 3.20–3.22 (1H and 2H, 2 \times m, CHCHCHCH₂OH); 3.16 (1H, d \times d, *J* = 12.9, 1.9 Hz, (HCH)O); 3.28 (1H, d \times d, *J* = 12.9, 3.1 Hz, (HCH)O); 3.40 (1H, d \times d, *J* = 9.1, 4.0 Hz, CHCH₂); 4.14 (1H, d, *J* = 15.0 Hz, N(HCH)); 4.68 (1H, d, *J* = 15.0 Hz, N(HCH)); 4.91 (1H, d, *J* = 4.5 Hz, CHPh); 4.97 (1H, d, *J* = 4.0 Hz, CH₂); 5.16 (1H, d, *J* = 4.5 Hz, CHCHPh); 7.18–7.22 (2H, m, CH_{arom}); 7.31–7.38 (6H, m, CH_{arom}); 7.40–7.44 (2H, m, CH_{arom}). ¹³C NMR (100 MHz, D₂O): δ 44.6 (NCH₂); 59.5 (OCH₂); 63.0 (CHPh); 68.1, 72.2, and 72.4 (CHCHCHCH₂OH); 71.2 (CHCH₂); 83.6 (CHCHPh); 101.6 (CH₂); 128.2, 128.7, 128.8, 128.9, 129.0, and 129.2 (10 \times HC_{arom}); 132.9 and 134.1 (2 \times C_{arom,quat}); 169.7 (C=O). IR (ATR, cm⁻¹): ν_{OH} = 3352; $\nu_{C=O}$ = 1743; ν_{max} = 2926, 1452, 1412, 1350, 1080, 1022, 772, 700. MS (70 eV): *m/z* (%) 416 (M⁺ + 1, 100). HRMS (ESI): calculated for C₂₂H₂₆NO₇⁺: 416.1704 [M + H]⁺, found: 416.1705. *R*_f = 0.4 (EtOAc/MeOH/H₂O 30:5:4). White crystals. Yield after normal phase column chromatography and preparative TLC (SiO₂): 3%.

Synthesis of 3-O- β -D-Glucopyranosyl-3-hydroxyazetid-2-ones 9a- β G, 9b- β G, and 9c- β G and Downstream Processing. The β -glucosides 9a- β G, 9b- β G, and 9c- β G were synthesized at 100 mL scale using 200 mM sucrose, 2 mM UDP, 15 mM 8a, 8b, or 8c, 10% (v/v) acetone, 0.11–0.19 mg/mL UGT-76G1Sr, 0.31–0.36 mg/mL SuSy, and 50 mM MOPS buffer (pH 7.0). The reaction samples were incubated at 32 °C in falcon tubes with continuous shaking at 200 rpm for 2 (8b, 8c) or 6 (8a) weeks. The reaction samples (50 μ L) were inactivated by adding 50 μ L acetonitrile and 50 μ L ultrapure water, followed by centrifugation. These samples were analyzed by LC-MS (eluent: 10–100% CH₃CN in water (5 mM NH₄OAc)).

The products were isolated by extraction and column chromatography as described for the α -glucosides. Finally, any remaining impurities were removed by reverse phase column chromatography (C₁₈) using a gradient of water and acetonitrile as the eluent: CH₃CN/H₂O (0:100) to CH₃CN/H₂O (100:0).

***cis*-1-Benzyl-3-O- β -D-glucopyranosyl-3-hydroxy-4-isopropylazetid-2-one 9a- β G.** The spectral data are derived from the mixture of diastereomers (*dr* = 44:56).

¹H NMR (400 MHz, D₂O): δ 0.92–0.99 (2 \times 6H, m, 2 \times (CH₃)₂CH); 2.10 (2 \times 1H, octet, *J* = 6.5 Hz, 2 \times (CH₃)₂CH); 3.33–3.41 (2 \times 1H, m, 2 \times CHCH₂); 3.42–3.55 (2 \times 3H, m, 2 \times CHCHCHCHCH₂); 3.65–3.72 (2 \times 1H, m, CHⁱPr);

3.73–3.78 (2 × 1H, m, 2 × O(HCH)); 3.89–3.95 (2 × 1H, m, 2 × O(HCH)); 4.34 (1H, d, J_{AB} = 15.8 Hz, N(HCH)); 4.36 (1H, d, J_{AB} = 15.4 Hz, N(HCH)); 4.52 (1H, d, J = 7.9 Hz, CH_β); 4.70 (1H, d, J = 7.8 Hz, CH_β); 4.73 (1H, d, J_{AB} = 15.4 Hz, N(HCH)); 4.75 (1H, d, J_{AB} = 15.8 Hz, N(HCH)); 5.08 (1H, d, J = 4.8 Hz, $CHCH^iPr$); 5.14 (1H, d, J = 4.9 Hz, $CHCH^iPr$); 7.33–7.47 (2 × 5H, m, CH_{arom}). ^{13}C NMR (100 MHz, D_2O): δ 18.0, 18.3, 18.8, and 19.1 (2 × (CH_3)₂CH); 27.5 ((CH_3)₂CH); 27.6 ((CH_3)₂CH); 45.8 (NCH₂); 45.9 (NCH₂); 60.7 (OCH₂); 60.8 (OCH₂); 64.7 (CH^iPr); 64.9 (CH^iPr); 69.6, 69.7, 75.69, 75.71, 76.0, and 76.2 (2 × $CHCHCHCHCH_\beta$); 73.0 ($CHCH_\beta$); 73.2 ($CHCH_\beta$); 79.7 ($CHCH^iPr$); 80.7 ($CHCH^iPr$); 102.3 (CH_β); 102.6 (CH_β); 128.0, 128.1, 128.2, and 129.0 (2 × (5 × HC_{arom})); 135.09 ($C_{arom,quat}$); 135.15 ($C_{arom,quat}$); 170.68 (C=O); 170.72 (C=O). IR (ATR, cm^{-1}): ν_{OH} = 3356; $\nu_{C=O}$ = 1724; ν_{max} = 2927, 1680, 1415, 1227, 1078, 1043, 993, 419. MS (70 eV): m/z (%) 382 ($M^+ + 1$, 100). HRMS (ESI): calculated for $C_{19}H_{28}NO_7^+$: 382.1860 [$M + H$]⁺, found: 382.1863. Colorless oil. R_f = 0.4 (EtOAc/MeOH/H₂O 30:5:4). Yield after column chromatography (SiO₂ and C₁₈): 0.2%.

cis-3-O- β -D-Glucopyranosyl-3-hydroxy-1-isopropyl-4-propylazetidin-2-one 9b- β G. The spectral data are derived from the mixture of diastereomers (dr = 36:64).

1H NMR (400 MHz, D_2O): δ 0.95 (2 × 3H, t, J = 7.3 Hz, 2 × CH_2CH_3); 1.24–1.31 (2 × 6H, m, 2 × (CH_3)₂CH); 1.33–1.50 (2 × 2H, m, 2 × CH_2CH_3); 1.68–1.82 (2 × 2H, m, 2 × $CH_2CH_2CH_3$); 3.35 (2 × 1H, d, J = 8.4, 8.1 Hz, 2 × $CHCH_\beta$); 3.38–3.55 (2 × 3H, m, 2 × $CHCHCHCHCH_\beta$); 3.72–3.84 (2 × (2 × 1H), m, 2 × CH^iPr and 2 × O(HCH)); 3.93 (2 × 1H, d, J = 12.2 Hz, O(HCH)); 4.00–4.05 (2 × 1H, m, 2 × $CHCH^iPr$); 4.50 (1H, d, J = 8.1 Hz, CH_β); 4.66 (1H, d, J = 8.1 Hz, CH_β); 5.01 (1H, d, J = 4.8 Hz, $CHCH^iPr$); 5.03 (1H, d, J = 4.8 Hz, $CHCH^iPr$). ^{13}C NMR (100 MHz, D_2O): δ 13.4 (CH_2CH_3); 13.5 (CH_2CH_3); 18.5 (CH_2CH_3); 18.7 (CH_2CH_3); 18.9 and 20.7 ((CH_3)₂CH); 19.0 and 20.6 ((CH_3)₂CH); 30.6 ($CH_2CH_2CH_3$); 30.9 ($CH_2CH_2CH_3$); 45.0 ((CH_3)₂CH); 45.1 ((CH_3)₂CH); 58.5 ($CHPr$); 58.7 ($CHPr$); 60.7 (2 × OCH₂); 69.58, 69.66, 75.66, 75.69, 75.9 and 76.2 (2 × $CHCHCHCHCH_\beta$); 73.0 ($CHCH_\beta$); 73.2 ($CHCH_\beta$); 79.4 ($CHCH^iPr$); 79.6 ($CHCH^iPr$); 102.2 (CH_β); 102.4 (CH_β); 168.8 (C = O); 169.0 (C = O). IR (ATR, cm^{-1}): ν_{OH} = 3356; $\nu_{C=O}$ = 1722; ν_{max} = 2932, 1680, 1416, 1225, 1175, 1078, 1043, 419. MS (70 eV): m/z (%) 334 ($M^+ + 1$, 100). Colorless oil. R_f = 0.38 (EtOAc/MeOH/H₂O 30:5:4). Yield after column chromatography (SiO₂ and C₁₈): 1.4%.

cis-1-Benzyl-3-O- β -D-glucopyranosyl-3-hydroxy-4-phenylazetidin-2-one 9c- β G. The spectral data are derived from the mixture of diastereomers (dr = 41:59).

1H NMR (400 MHz, D_2O): δ 3.01 (1H, d × d, J = 8.8, 8.4 Hz, $CHCH_\beta$); 3.07 (1H, d × d, J = 8.5, 8.4 Hz, $CHCH_\beta$); 3.13–3.41 (2 × 3H, m, 2 × $CHCHCHCHCH_\beta$); 3.53 (1H, d × d, J = 11.8, 5.5 Hz, O(HCH)); 3.70 (1H, d × d, J = 11.8, 5.7 Hz, O(HCH)); 3.73 (1H, d, J = 12.0 Hz, O(HCH)); 3.89 (1H, d, J = 12.0 Hz, O(HCH)); 4.10 (1H, d, J = 8.4 Hz, CH_β); 4.13 (1H, d, J_{AB} = 14.8 Hz, N(HCH)); 4.17 (1H, d, J_{AB} = 15.1 Hz, N(HCH)); 4.52 (1H, d, J = 8.4 Hz, CH_β); 4.72 (1H, d, J_{AB} = 14.8 Hz, N(HCH)); 4.73 (1H, d, J_{AB} = 15.1 Hz, N(HCH)); 4.91 (1H, d, J = 4.6 Hz, $CHPh$); 4.93 (1H, d, J = 4.3 Hz, $CHPh$); 5.31 (1H, d, J = 4.6 Hz, $CHCHPh$); 5.37 (1H, d, J = 4.3 Hz, $CHCHPh$); 7.20–7.21, 7.32–7.35, and 7.41–7.43 (2 × 10H, 3 × m, 2 × (10 × HC_{arom})). ^{13}C NMR (100 MHz, D_2O): δ 44.6 (NCH₂); 44.7 (NCH₂); 60.66 (OCH₂); 60.72

(OCH₂); 61.9 ($CHPh$); 62.6 ($CHPh$); 69.2, 69.5, 75.1, 75.5, and 75.9 (2 × $CHCHCHCHCH_\beta$); 72.5 ($CHCH_\beta$); 72.8 ($CHCH_\beta$); 80.6 ($CHCHPh$); 82.3 ($CHCHPh$); 101.1 (CH_β); 102.6 (CH_β); 128.2, 128.4, 128.66, 128.68, 128.73, 128.75, 128.79, 128.9, and 128.98 (2 × (10 × HC_{arom})); 132.7 ($C_{arom,quat}$); 133.3 ($C_{arom,quat}$); 134.2 (2 × $C_{arom,quat}$); 169.1 (C=O); 169.3 (C=O). IR (ATR, cm^{-1}): ν_{OH} = 3341; $\nu_{C=O}$ = 1738; ν_{max} = 2924, 1672, 1452, 1416, 1233, 1076, 1045, 700. MS (70 eV): m/z (%) 416 ($M^+ + 1$, 100). Colorless oil. R_f = 0.36 (EtOAc/MeOH/H₂O 30:5:4). Yield after column chromatography (SiO₂ and C₁₈): 1%.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acsomega.8b01969.

1H NMR and ^{13}C NMR spectra of compounds 8a, 9a-c- α G, 9a-c- β G, 14a (PDF)

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Notes

The authors declare no competing financial interest.

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