



US 20160272951A1

(19) **United States**

(12) **Patent Application Publication**
Senger et al.

(10) **Pub. No.: US 2016/0272951 A1**

(43) **Pub. Date: Sep. 22, 2016**

(54) **ACYLTRANSFERASES AND USES THEREOF
IN FATTY ACID PRODUCTION**

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(21) Appl. No.: **15/159,931**

(22) Filed: **May 20, 2016**

Related U.S. Application Data

(62) Division of application No. 13/806,269, filed on Dec.
21, 2012, now Pat. No. 9,388,437, filed as application
No. PCT/EP2011/060315 on Jun. 21, 2011.

(60) Provisional application No. 61/358,431, filed on Jun.
25, 2010.

(30) **Foreign Application Priority Data**

Jun. 25, 2010 (EP) 10167342.4

Publication Classification

(51) **Int. Cl.**

C12N 9/10 (2006.01)

A23K 20/158 (2006.01)

A23D 9/02 (2006.01)

C12P 7/64 (2006.01)

C12N 15/82 (2006.01)

(52) **U.S. Cl.**

CPC **C12N 9/1029** (2013.01); **C12P 7/6427**
(2013.01); **C12N 15/8247** (2013.01); **A23D**
9/02 (2013.01); **A23L 1/3006** (2013.01); **A23L**
1/3008 (2013.01); **A23L 1/3002** (2013.01);
A23K 20/158 (2016.05); **C12Y 203/0102**
(2013.01); **A23V 2002/00** (2013.01)

(57)

ABSTRACT

The present invention relates to the recombinant manufacture of polyunsaturated fatty acids. Specifically, it relates to acyltransferase polypeptides, polynucleotides encoding said acyltransferases as well as vectors, host cells, non-human transgenic organisms containing said polynucleotides. Moreover, the present invention contemplates methods for the manufacture of polyunsaturated fatty acids as well as oils obtained by such methods.

Fig 1:

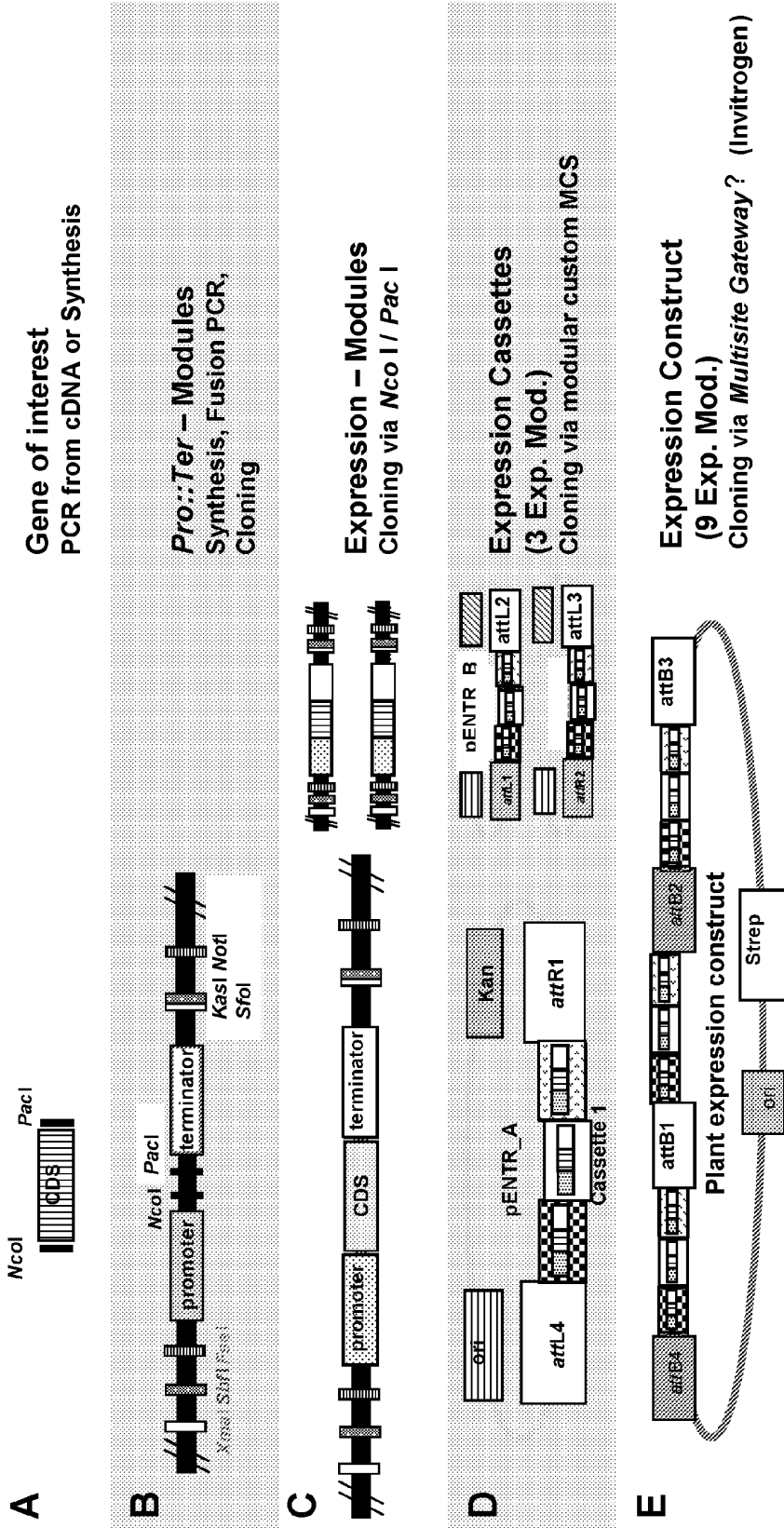


Fig 2:

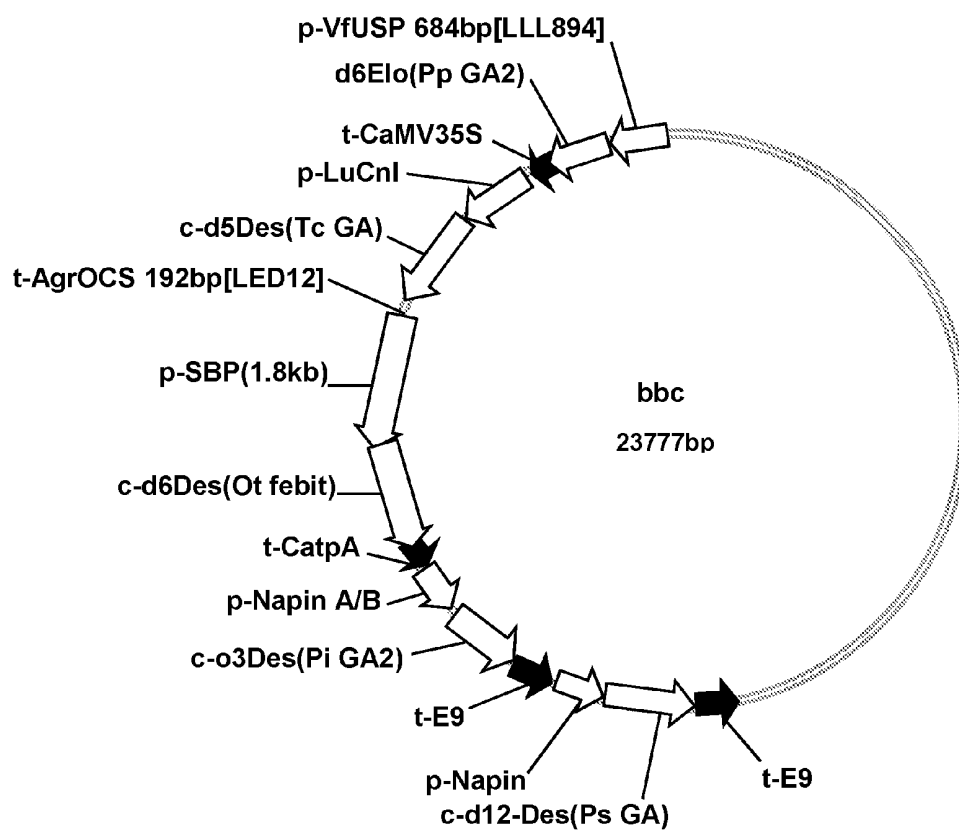
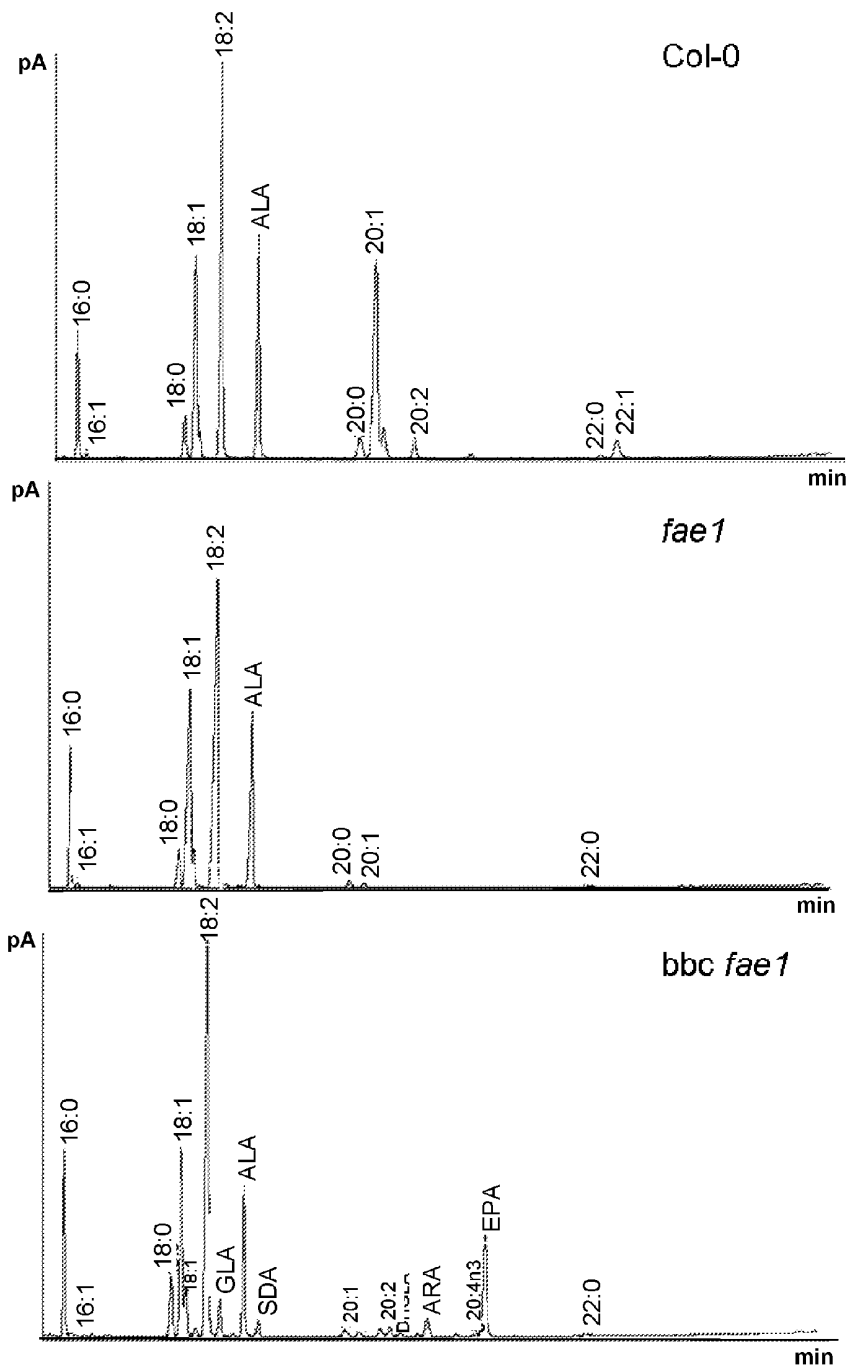


Fig 3:



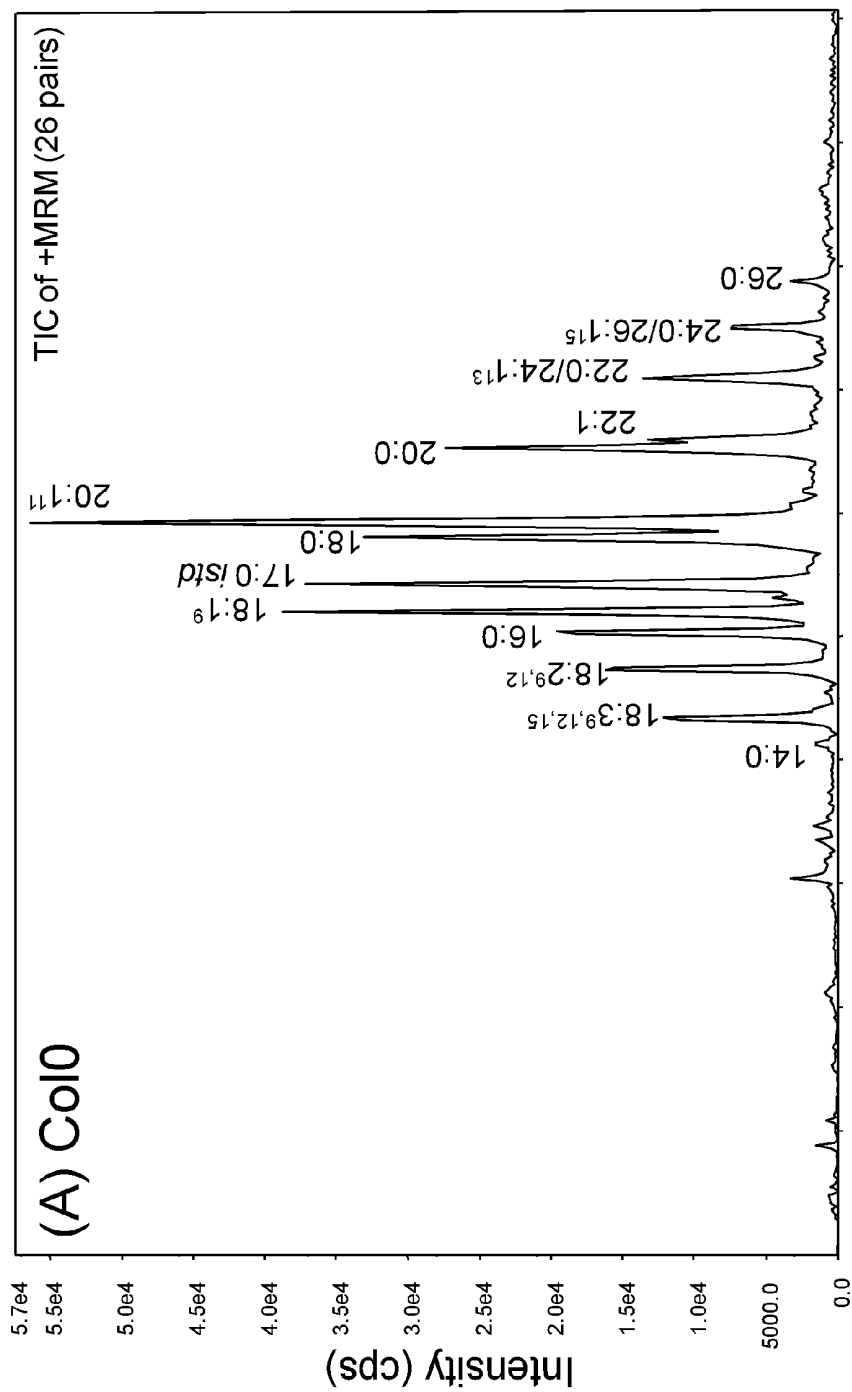


Fig 4:

Fig 4 (continued):

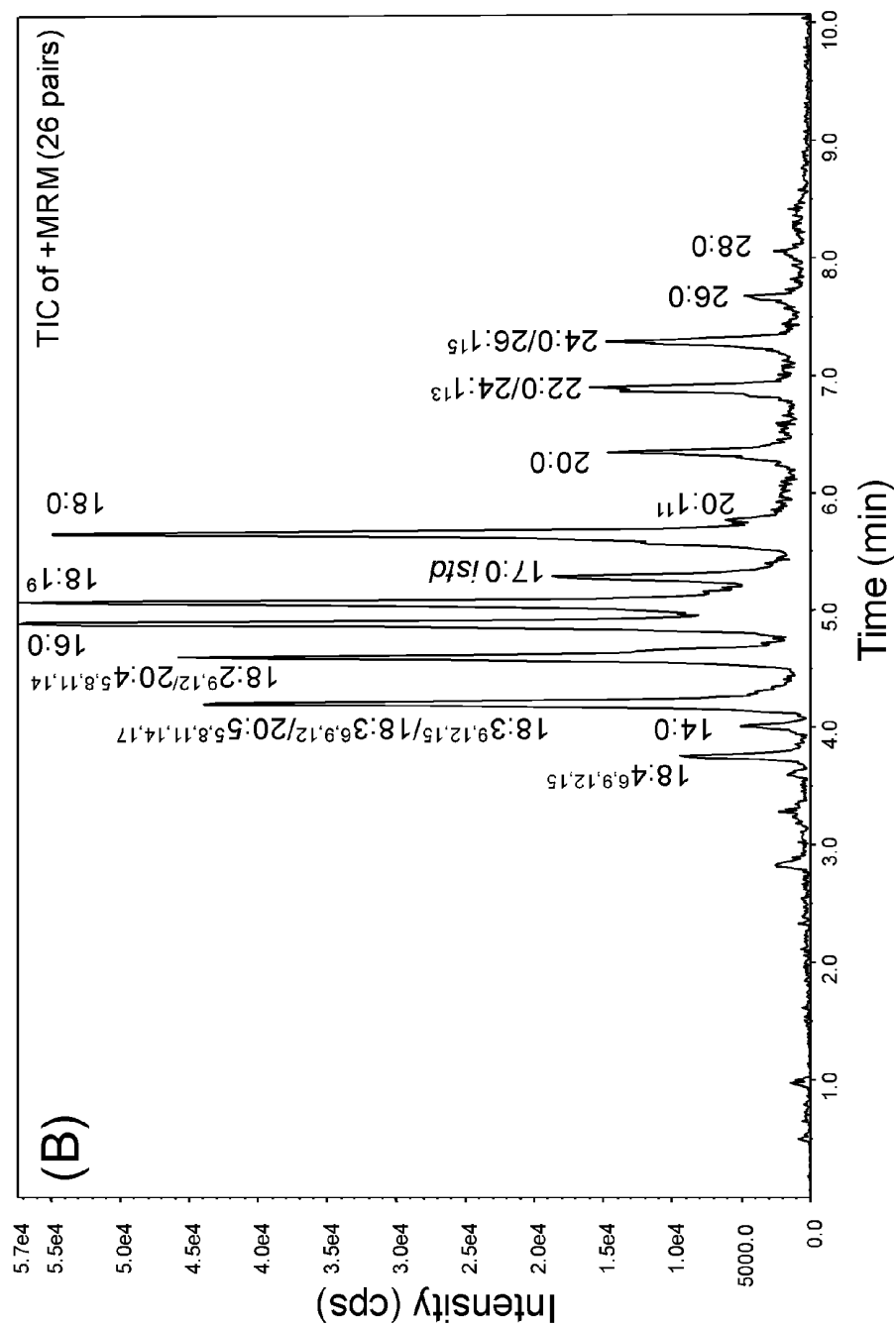


Fig 5:

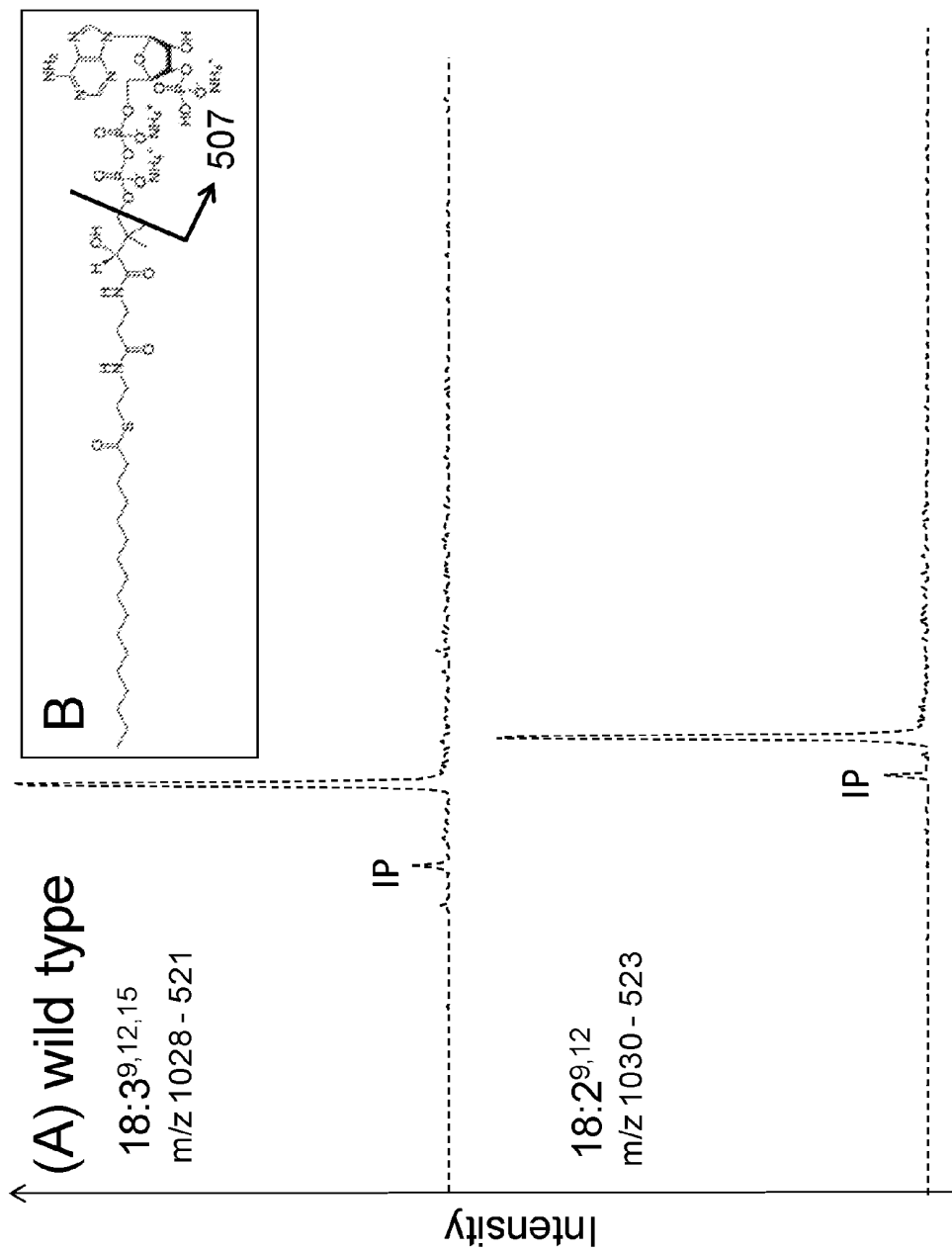


Fig 5 (continued):

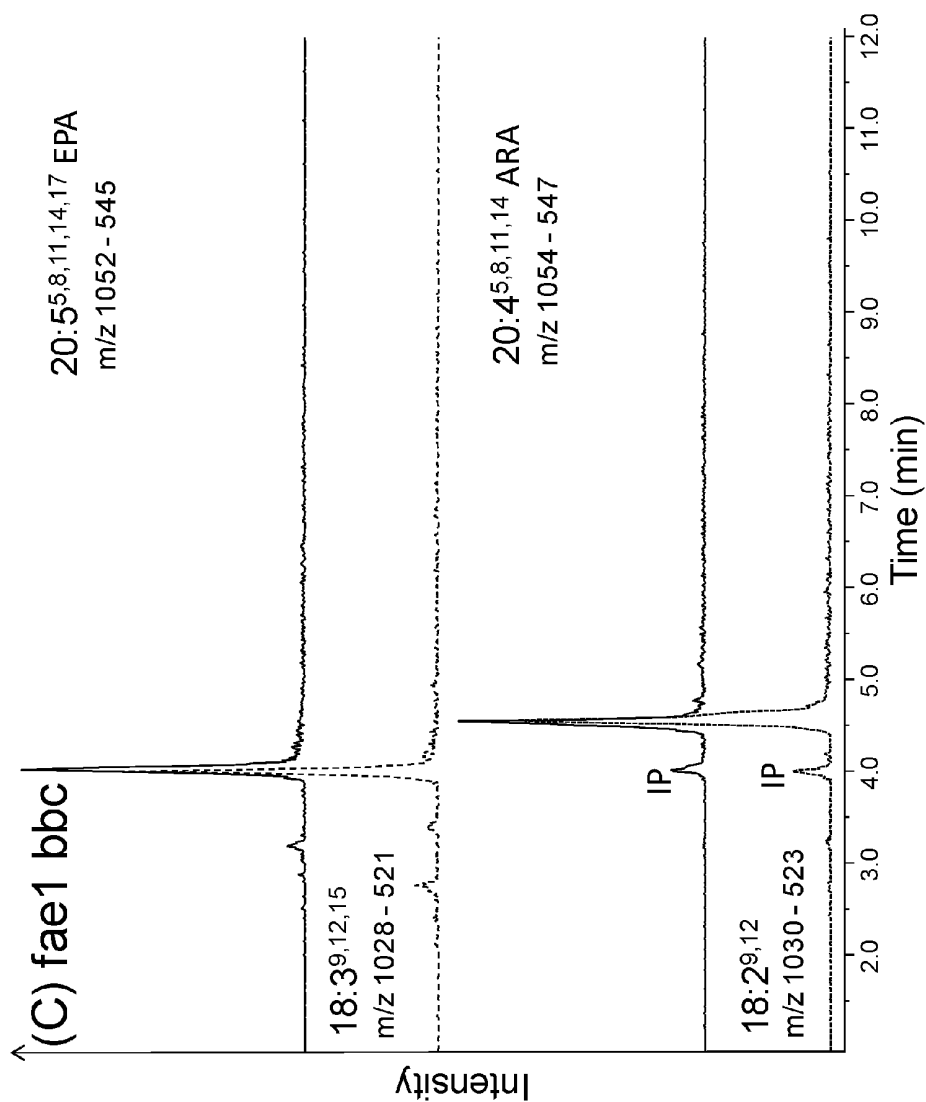


Fig 6:

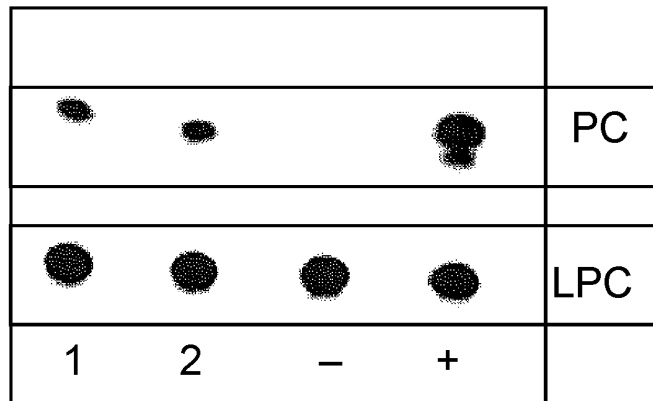


Fig 7:

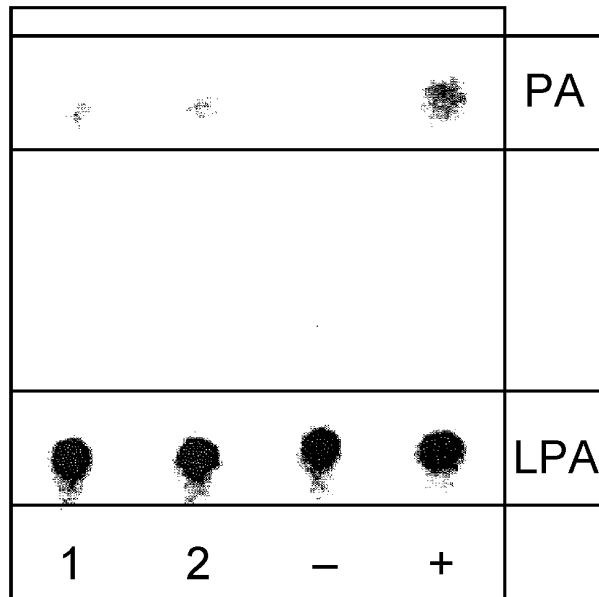
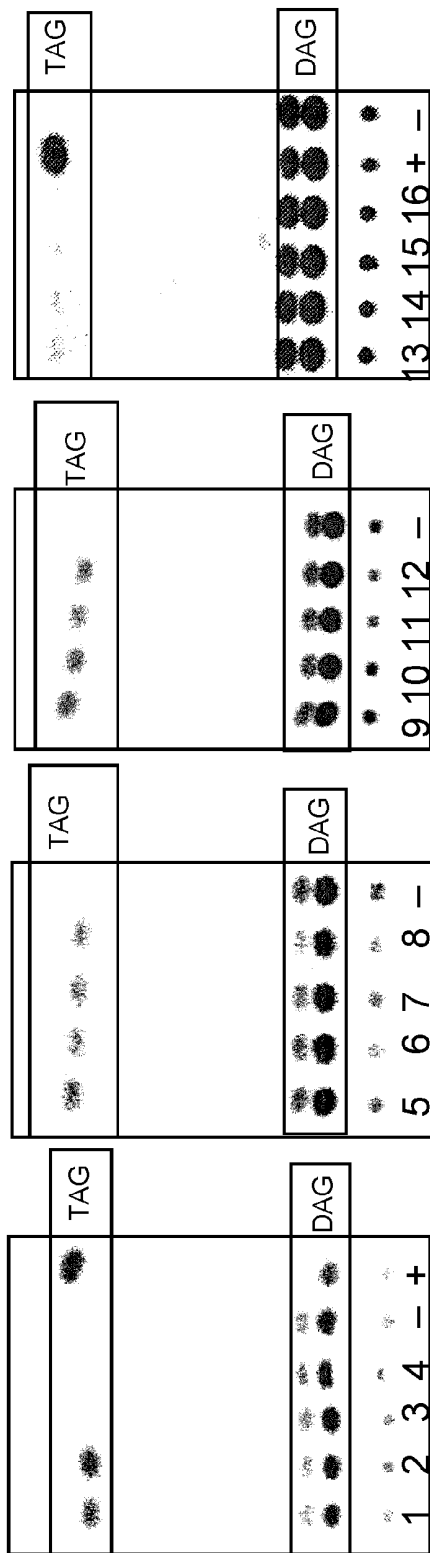


Fig 8



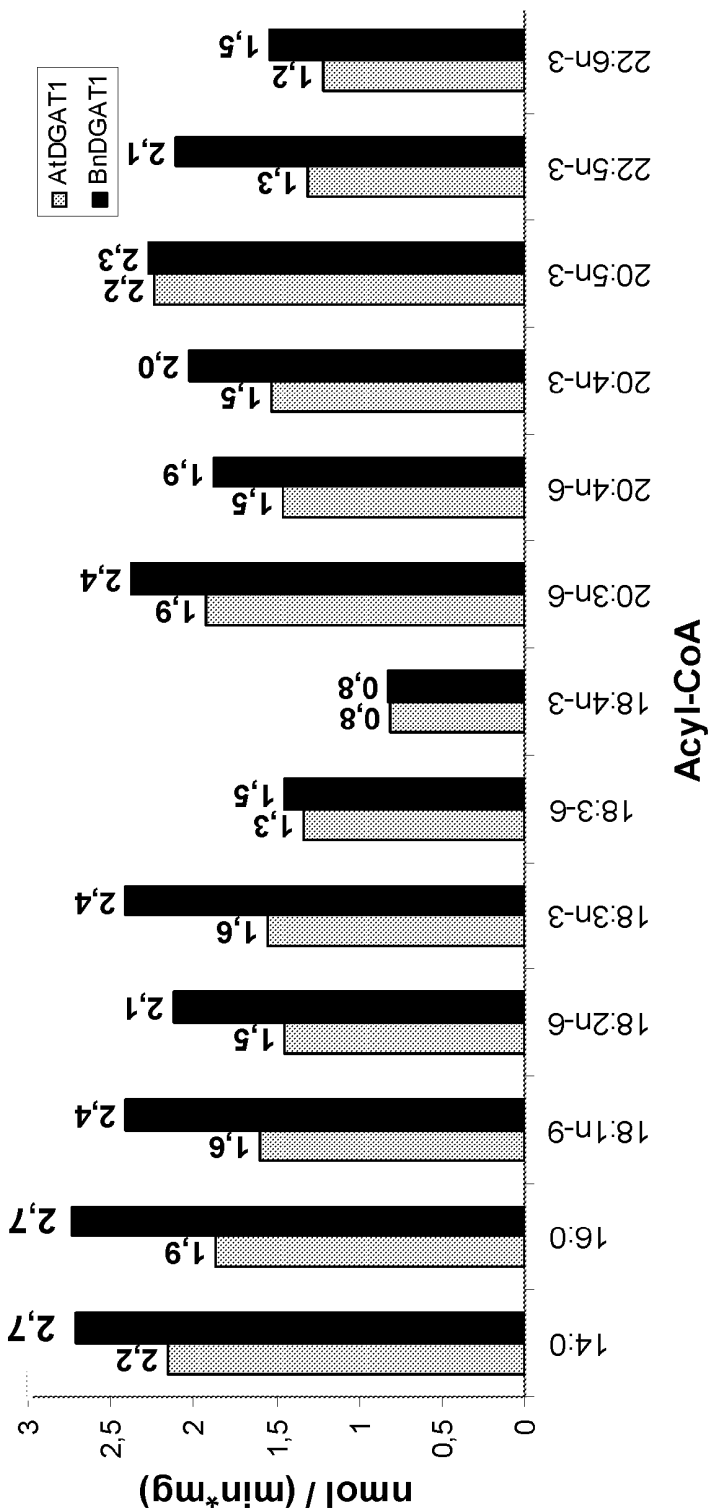
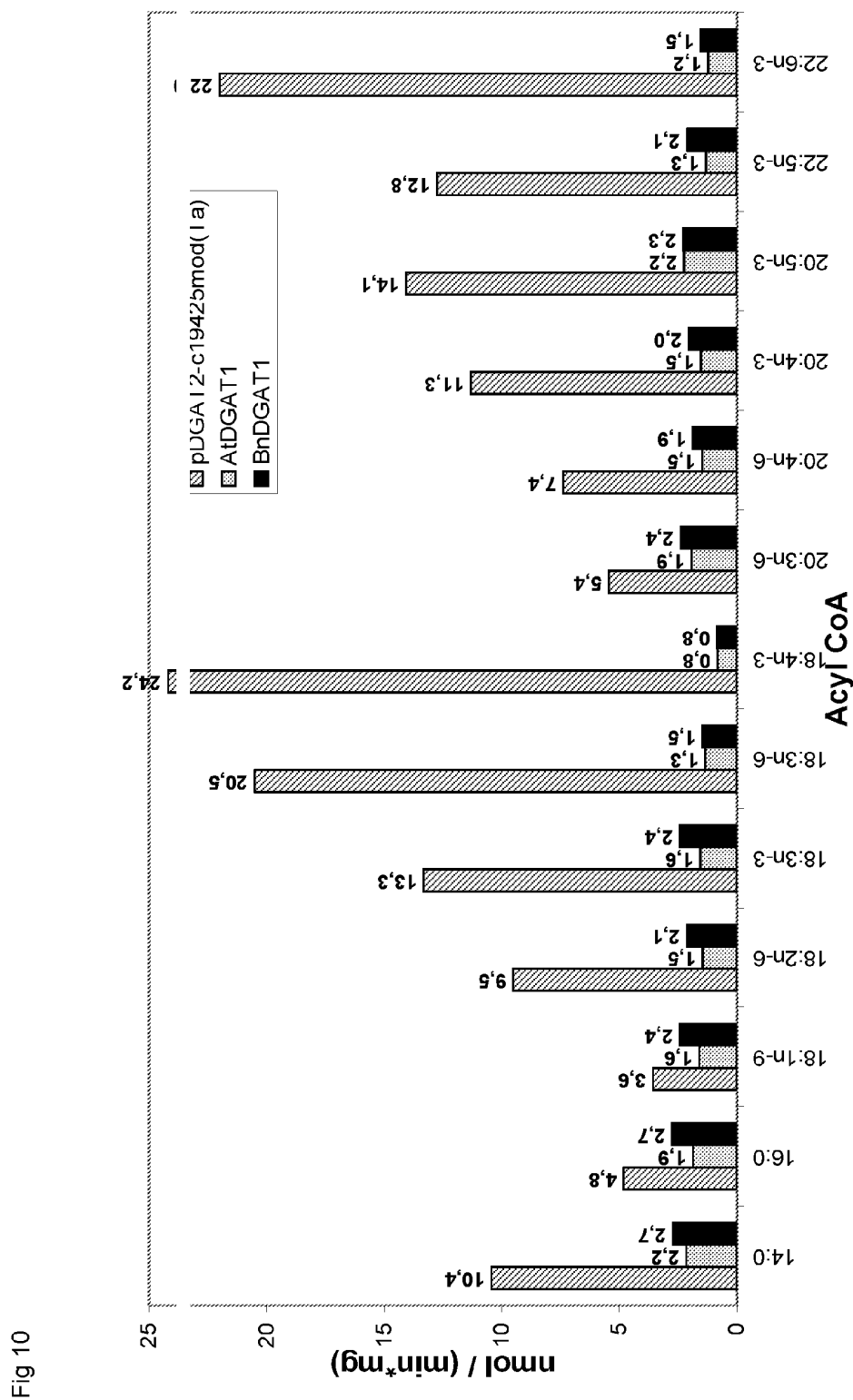


Fig 9



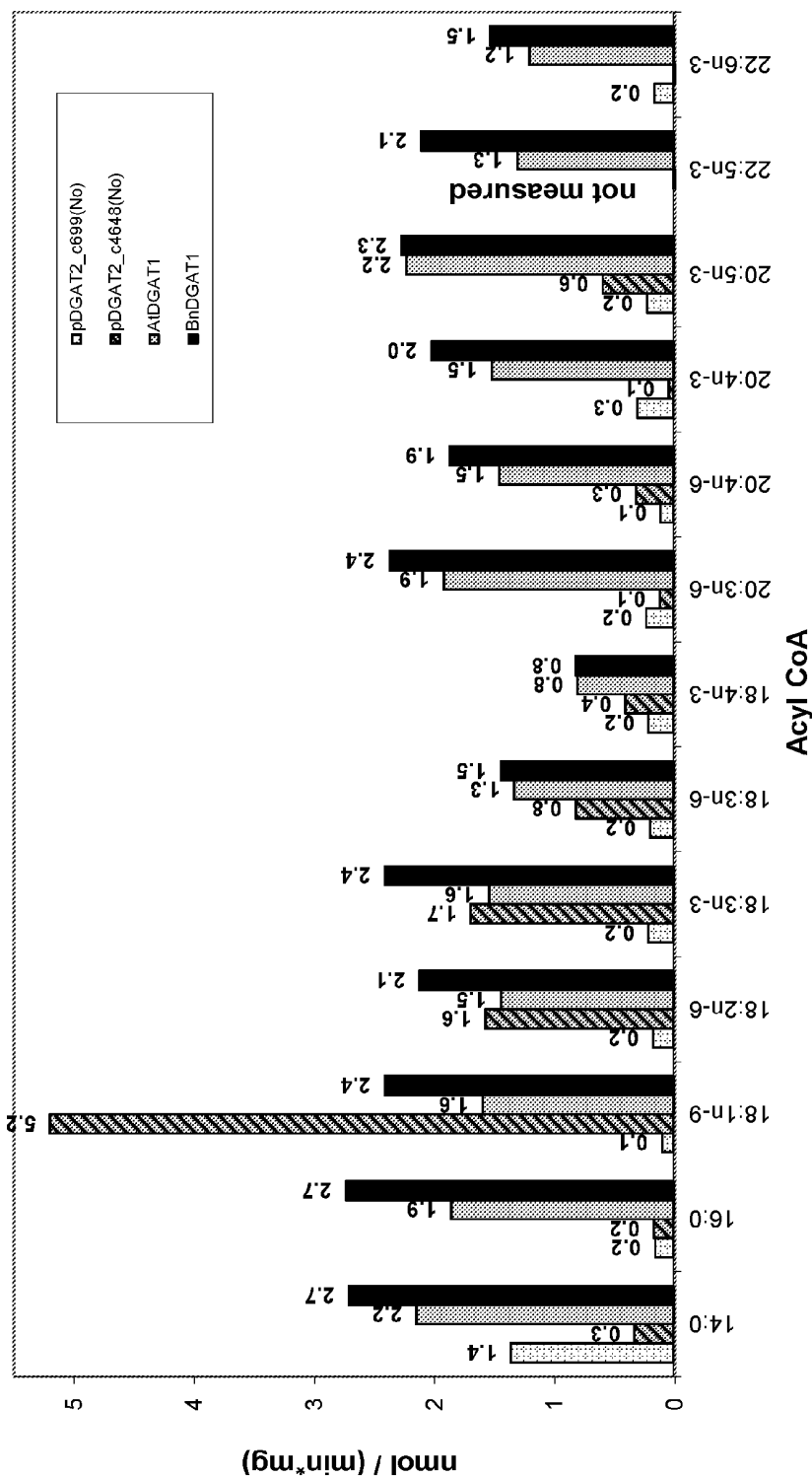


Fig 11

ACYLTRANSFERASES AND USES THEREOF IN FATTY ACID PRODUCTION

RELATED APPLICATIONS

[0001] This application is a divisional application of U.S. application Ser. No. 13/806,269 filed Dec. 21, 2012, which is a national stage application (under 35 U.S.C. §371) of PCT/EP2011/060315, filed Jun. 21, 2011 which claims benefit of European Application No. 10167342.4 filed Jun. 25, 2010, and U.S. Provisional Application No. 61/358,431, filed Jun. 25, 2010. The entire contents of each of these applications are hereby incorporated by reference herein in their entirety.

SUBMISSION OF SEQUENCE LISTING

[0002] The Sequence Listing associated with this application is filed in electronic format via EFS-Web and hereby incorporated by reference into the specification in its entirety. The name of the text file containing the Sequence Listing is Sequence_Listing_074021_0214_01. The size of the text file is 216 KB and the text file was created on May 19, 2016.

[0003] The present invention relates to the recombinant manufacture of polyunsaturated fatty acids. Specifically, it relates to acyltransferase polypeptides, polynucleotides encoding said acyltransferase polypeptides as well to vectors, host cells, non-human transgenic organisms containing said polynucleotides. Moreover, the present invention contemplates methods for the manufacture of polyunsaturated fatty acids as well as oils obtained by such methods.

[0004] Fatty acids and triacylglycerides have a various applications in the food industry, in animal feed, supplement nutrition, and in the cosmetic and pharmacological and pharmaceutical field. The individual applications may either require free fatty acids or triacylglycerides. In both cases, however, polyunsaturated fatty acids either free or esterified are of pivotal interest for many of the aforementioned applications. In particular, polyunsaturated omega-3-fatty acids and omega-6-fatty acids are important constituents in animal and human food. These fatty acids are supposed to have beneficial effects on the overall health and, in particular, on the central nervous system, the cardiovascular system, the immune system, and the general metabolism. Within traditional food, the polyunsaturated omega-3-fatty acids are mainly found in fish and plant oils. However, in comparison with the needs of the industry and the need for a beneficial diet, this source is rather limited.

[0005] The various polyunsaturated fatty acids (PUFA) and PUFA-containing triglycerides are also mainly obtained from microorganisms such as *Mortierella* and *Schizochytrium* or from oil-producing plants such as soybean or oilseed rape, algae such as *Cryptothecodinium* or *Phaeodactylum* and others, where they are usually obtained in the form of their triacylglycerides. The free PUFA are usually prepared from the triacylglycerides by hydrolysis. However, long chain polyunsaturated fatty acids (LCPUFA) having a C-18, C-20, C-22 or C-24 fatty acid body, such as docosahexaenoic acid (DHA), eicosapentaenoic acid (EPA), arachidonic acid (ARA), dihomo-gamma-linolenic acid or docosapentaenoic acid (DPA) can not be efficiently isolated from natural oil crop plants such as oilseed rape, soybean, sunflower or safflower. Conventional natural sources of these fatty acids are, thus, merely fish, such as herring, salmon, sardine, redfish, eel, carp, trout, halibut, mackerel, zander or tuna, or from algae.

[0006] Especially suitable microorganisms for the production of PUFA in industrial scale are microalgae such as *Phaeodactylum tricornutum*, *Porphoridium* species, *Thraustochytrium* species, *Nannochloropsis* species, *Schizochytrium* species or *Cryptothecodinium* species, ciliates such as *Stylonychia* or *Colpidium*, fungi such as *Mortierella*, *Entomophthora* or *Mucor* and/or mosses such as *Physcomitrella*, *Ceratodon* and *Marchantia* (Vazhappilly 1998, Botanica Marina 41: 553-558; Totani 1987, Lipids 22: 1060-1062; Akimoto 1998, Appl. Biochemistry and Biotechnology 73: 269-278). Strain selection has resulted in the development of a number of mutant strains of the microorganisms in question which produce a series of desirable compounds including PUFA. However, the mutation and selection of strains with an improved production of a particular molecule such as the polyunsaturated fatty acids is a time-consuming and difficult process. This is why recombinant methods as described above are preferred whenever possible. However, only limited amounts of the desired PUFA or LCPUFA and, in particular, DHA or EPA, can be produced with the aid of the above mentioned microorganisms, and, depending on the microorganism used, these are generally obtained as fatty acid mixtures of, for example, EPA, DPA and DHA.

[0007] Many attempts in the past have been made to make available genes which are involved in the synthesis of fatty acids or triglycerides for the production of oils in various organisms. Various desaturases have been described in the art; see, e.g., documents WO 91/13972, WO 93/11245, WO 94/11516, EP-A-0 550 162, WO 94/18337, WO 97/30582, WO 97/21340, WO 95/18222, EP-A-0 794 250, Stukey 1990, J. Biol. Chem., 265: 20144-20149, Wada 1990, Nature 347: 200-203, Huang 1999, Lipids 34: 649-659, WO 93/06712, U.S. Pat. No. 5,614,393, WO 96/21022, WO 00/21557, WO 99/27111, WO 98/46763, WO 98/46764, WO 98/46765, WO 99/64616 or WO 98/46776. These enzymes can be used for the production of unsaturated fatty acids. Thus, due to modern molecular biology, it has become possible to increase at least to some extent the content of the desired polyunsaturated fatty acids and, in particular, the PUFA or LCPUFA in a given organism. Elongases for the production of fatty acids are disclosed in the document WO2009/016202.

[0008] The biosynthesis of LCPUFA and the incorporation of LCPUFA into membrane lipids or triacylglycerides proceeds via various metabolic pathways (Abadi 2001, European Journal of Lipid Science & Technology 103:106-113). In bacteria such as *Vibrio*, and microalgae, such as *Schizochytrium*, malonyl-CoA is converted into LCPUFA via an LCPUFA-producing polyketide synthase (Metz 2001, Science 293: 290-293; WO 00/42195; WO 98/27203; WO 98/55625). In microalgae, such as *Phaeodactylum*, and mosses, such as *Physcomitrella*, unsaturated fatty acids such as linoleic acid or linolenic acid are converted in a plurality of desaturation and elongation steps to give LCPUFA (Zank 2000, Biochemical Society Transactions 28: 654-658). Desaturation takes place either on acyl groups bound to Coenzyme A (acyl-CoA) or on acyl groups of membrane lipids, whereas elongation is biochemically restricted to acyl chains bound to CoA. In mammals, the biosynthesis of DHA comprises a chain shortening via beta-oxidation, in addition to desaturation and elongation steps. In microorganisms and lower plants, LCPUFA are present either exclusively in the form of membrane lipids, as is the case in *Physcomitrella* and *Phaeodactylum*, or in membrane lipids and triacylglycerides, as is the case in *Schizochytrium* and *Mortierella*. Incorporation

tion of LCPUFA into lipids and oils, as well as the transfer of the fatty acid moiety (acyl group) between lipids and other molecular species such as acyl-CoA, is catalyzed by various acyltransferases and transacylases. These enzymes are, known to carry out the incorporation or interexchange of saturated and unsaturated fatty acids (Slabas 2001, J. Plant Physiology 158: 505-513, Frentzen 1998, Fett/Lipid 100: 161-166, Cases 1998, Proc. Nat. Acad. Sci. USA 95: 13018-13023). One group of acyltransferases having three distinct enzymatic activities are enzymes of the "Kennedy pathway", which are located on the cytoplasmic side of the membrane system of the endoplasmic reticulum (ER). The ER-bound acyltransferases in the microsomal fraction use acyl-CoA as the activated form of fatty acids. Glycerol-3-phosphate acyltransferase (GPAT) catalyzes the incorporation of acyl groups at the sn-1 position of glycerol-3-phosphate. 1-Acylglycerol-3-phosphate acyltransferase, also known as lysophosphatidic acid acyltransferase (LPAAT), catalyze the incorporation of acyl groups at the sn-2 position of lysophosphatidic acid (LPA). After dephosphorylation of phosphatidic acid by phosphatidic acid phosphatase (PAP), diacylglycerol acyltransferase (DGAT) catalyzes the incorporation of acyl groups at the sn-3 position of diacylglycerols. Further enzymes directly involved in TAG biosynthesis—apart from the said Kennedy pathway enzymes—are the phospholipid diacylglycerol acyltransferase (PDAT), an enzyme that transfers acyl groups from the sn-2 position of membrane lipids to the sn-3 position of diacylglycerols, and diacylglyceroldiacylglycerol transacylase (DDAT), an enzyme that transfers acylgroups from the sn-2 position of one diacylglycerol-molecule to the sn-3 position of another diacylglycerol-molecule. Lysophospholipid acyltransferase (LPLAT) represents a class of acyltransferases that are capable of incorporating activated acyl groups from acyl-CoA to membrane lipids, and possibly catalyze also the reverse reaction. More specifically, LPLATs can have activity as lysophosphatidylethanolamine acyltransferase (LPEAT) and lysophosphatidylcholine acyltransferase (LPCAT). Further enzymes, such as lecithin cholesterol acyltransferase (LCAT) can be involved in the transfer of acyl groups from membrane lipids into triacylglycerides, as well.

[0009] The documents WO 98/54302 and WO 98/54303 disclose a human LPAAT and its potential use for the therapy of diseases, as a diagnostic, and a method for identifying modulators of the human LPAAT. Moreover, a variety of acyltransferases with a wide range of enzymatic functions have been described in the documents WO 98/55632, WO 98/55631, WO 94/13814, WO 96/24674, WO 95/27791, WO 00/18889, WO 00/18889, WO 93/10241, Akermoun 2000, Biochemical Society Transactions 28: 713-715, Tumaney 1999, Biochimica et Biophysica Acta 1439: 47-56, Fraser 2000, Biochemical Society Transactions 28: 715-7718, Szymne 1984, Biochem. J. 223: 305-314, Yamashita 2001, Journal of Biological Chemistry 276: 26745-26752, and WO 00/18889.

[0010] Higher plants comprise PUFA, such as linoleic acid and linolenic acid. However, the LCPUFA ARA, EPA and DHA are not present in the seed oils of higher plants or only in traces (Ucciani: Nouveau Dictionnaire des Huiles Végétales. Technique & Documentation-Lavoisier, 1995. ISBN: 2-7430-0009-0). It is nevertheless highly desirable to produce LCPUFA in higher plants, preferably in oil seeds such as oilseed rape, linseed, sunflower and soybean, since large

amounts of high-quality LCPUFA for the various aforementioned applications may be obtained thereby at low costs.

[0011] However, one drawback of using transgenic plants expressing various of the aforementioned desaturases and elongases involved in the synthesis of PUFA and LCPUFA is that the latter are not efficiently incorporated into triacylglycerides, but rather into membranes. Furthermore, efficient processing of a given acyl molecule-substrate, e.g. linoleic acid, by a plurality of desaturation and elongation steps towards the desired LCPUFA, e.g. ARA, EPA and/or DHA, is hindered by the requirement to transfer the acyl molecule and its derivatives generated by the elongation and desaturation reactions back and forth between membrane lipids and acyl-CoA. For this reason, intermediates towards desired LCPUFA are incorporated into oil before the synthesis of the desired LCPUFA is complete. These two problems are undesired for the following reasons: First, the main lipid fraction in oil seeds are triacylglycerides. This is why, for economical reasons, it is necessary to concentrate LCPUFA in triacylglycerides. Second, LCPUFA which are incorporated into membranes can modify the physical characteristics of the membranes and thus have harmful effects on the integrity and transport characteristics of the membranes and on the stress tolerance of plants. Third, for efficient LCPUFA synthesis, it is desirable to increase the flux of intermediate-LCPUFA between the two sites of biosynthesis—that are membrane lipids and acyl-CoA—and/or decrease the flux of intermediate-PUFA/-LCPUFA into oil. Transgenic plants which comprise and express genes coding for enzymes of LCPUFA biosynthesis and produce LCPUFA have been described, e.g., in DE 102 19 203 or WO2004/087902. However, these plants produce LCPUFA in amounts which require further optimization for processing the oils present in said plants. Moreover, it was proposed that delta 6 desaturated fatty acids may be shifted into the acyl-CoA pool for increasing efficiency of fatty acid elongation in plants (Singh 2005, *Curr. Opin. Plant Biol.*, 8: 197-203). Another publication demonstrated in *Arabidopsis*, that the additional expression of RcDGAT2 from *Ricinus communis* increase the storage of hydroxyfatty acids produced by a *Ricinus communis* fatty acid hydroxylase 12 (FAH12) from 17% to 30% in the seed oil.

[0012] Accordingly, means for increasing the content of PUFA or LCPUFA, such as EPA and DHA, in triglycerides in, e.g., plant seed oils, are still highly desirable.

[0013] Thus, the present invention relates to a polynucleotide comprising a nucleic acid sequence elected from the group consisting of:

[0014] a) a nucleic acid sequence having a nucleotide sequence as shown in any one of SEQ ID NOs: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, and 55;

[0015] b) a nucleic acid sequence encoding a polypeptide having an amino acid sequence as shown in any one of SEQ ID NOs: 53, 2, 5, 8, 11, 14, 17, 20, 23, 26, 29, 32, 35, 38, 41, 44, 47, 50, and 56;

[0016] c) a nucleic acid sequence being at least 40% identical to the nucleic acid sequence of a) or b), wherein said nucleic acid sequence encodes a polypeptide having acyltransferase activity;

[0017] d) a nucleic acid sequence encoding a polypeptide having acyltransferase activity and having an amino acid sequence which is at least 45% identical to the amino acid sequence of b); and

[0018] e) a nucleic acid sequence which is capable of hybridizing under one of the following sets of conditions to

any one of a) to d), wherein said nucleic acid sequence encodes a polypeptide having acyltransferase activity:

[0019] f) hybridization in 50 mM Tris, pH 7.6, 6×SSC, 5×Denhardt's, 1.0% sodium dodecyl sulfate (SDS) 100 μg denaturated calf thymus DNA at 34° C. overnight and wash twice with 2×SSC, 0.5% SDS at room temperature for 15 min each, repeat twice with 0.2×SSC, 0.5% SDS at room temperature for 15 min each and then repeat twice with 0.2 SSC, 0.5% SDS at 50° C. for 15 min;

[0020] g) hybridization in 6×SSPE (Sodium chloride Sodium Phosphate-EDTA), 5×Denhardt's solution, 0.5% SDS 100 μg denaturated calf thymus DNA at 34° C. overnight and wash twice with 2×SSC, 0.5% SDS at room temperature for 15 min each, repeat twice with 0.2×SSC, 0.5% SDS at room temperature for 15 min each and then repeat twice with 0.2 SSC, 0.5% SDS at 50° C. for 15 min;

[0021] h) hybridization in 20-30% formamide, 5×SSPE, 5×Denhardt's solution, 1% SDS 100 μg denaturated salmon sperm DNA at 34° C. overnight and wash twice with 2×SSPE, 0.2% SDS at 42° C. for 15 min each, repeat twice with 2×SSPE, 0.2% SDS at 55° C. for 30 min each and repeat twice with 0.2 SSC, 0.5% SDS at 50° C. for 15 min;

[0022] i) hybridization in 7% SDS, 0.5 M NaPO₄, 1 mM EDTA at 50° C. overnight and wash in 2×SSC, 0.1% SDS at 50° C. or 65° C.;

[0023] j) hybridization in 7% SDS, 0.5 M NaPO₄, 1 mM EDTA at 50° C. overnight and wash in 1×SSC, 0.1% SDS at 50° C. or 65° C.; or

[0024] k) hybridization in 7% SDS, 0.5 M NaPO₄, 1 mM EDTA at 50° C. overnight and wash in 0.1×SSC, 0.1% SDS at 50° C. or 65° C.

[0025] The term "polynucleotide" as used in accordance with the present invention relates to a polynucleotide comprising a nucleic acid sequence which encodes a polypeptide having acyltransferase activity. Preferably, the polypeptide encoded by the polynucleotide of the present invention having acyltransferase activity upon expression in a plant shall be capable of increasing the amount of PUFA and, in particular, LCPUFA esterified to triglycerides in, e.g., seed oils or the entire plant or parts thereof. Such an increase is, preferably, statistically significant when compared to a LCPUFA producing transgenic control plant which expresses the minimal set of desaturases and elongases required for LCPUFA synthesis but does not express the polynucleotide of the present invention. Such a transgenic plant may, preferably, express desaturases and elongases comprised by the vector LJB765 listed in table 11 of example 5 in WO2009/016202 or a similar set of desaturases and elongases required for DHA synthesis. Whether an increase is significant can be determined by statistical tests well known in the art including, e.g., Student's t-test. More preferably, the increase is an increase of the amount of triglycerides containing LCPUFA of at least 5%, at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45% or at least 50% compared to the said control. Preferably, the LCPUFA referred to before is a polyunsaturated fatty acid having a C-20, C-22 or C24 fatty acid body, more preferably, EPA or DHA, most preferably, DHA. Suitable assays for measuring the activities mentioned before are described in the accompanying Examples.

[0026] The term "acyltransferase activity" or "acyltransferase" as used herein encompasses all enzymatic activities and enzymes which are capable of transferring or are involved

in the transfer of PUFA and, in particular; LCPUFA from the acyl-CoA pool or the membrane phospholipids to the triglycerides, from the acyl-CoA pool to membrane lipids and from membrane lipids to the acyl-CoA pool by a transesterification process. It will be understood that this acyltransferase activity will result in an increase of the LCPUFA esterified to triglycerides in, e.g., seed oils. In particular, it is envisaged that these acyltransferases are capable of producing triglycerides having esterified EPA or even DHA, or that these acyltransferases are capable of enhancing synthesis of desired PUFA by increasing the flux for specific intermediates of the desired PUFA between the acyl-CoA pool (the site of elongation) and membrane lipids (the predominant site of desaturation). Specifically, acyltransferase activity as used herein relates to lysophospholipid acyltransferase (LPLAT) activity, preferably, lysophosphatidylcholine acyltransferase (LPCAT) or Lysophosphatidylethanolamine acyltransferase (LPEAT) activity, lysophosphatidic acid acyltransferase (LPAAT) activity, glycerol-3-phosphate acyltransferase (GPAT) activity or diacylglycerol acyltransferase (DGAT), and, more preferably, to LPLAT, LPAAT, DGAT or GPAT activity.

[0027] More preferably, polynucleotides having a nucleic acid sequence as shown in SEQ ID NOs: 1, 4, and 7, encoding polypeptides having amino acid sequences as shown in SEQ ID NOs: 2, 5, and 8 or variants thereof, preferably, exhibit LPLAT activity. Polynucleotides having a nucleic acid sequence as shown in SEQ ID NOs: 10, and 13, encoding polypeptides having amino acid sequences as shown in SEQ ID NOs: 11, and 14 or variants thereof, preferably, exhibit LPAAT activity. Polynucleotides having a nucleic acid sequence as shown in SEQ ID NOs: 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, 52, and 55, encoding polypeptides having amino acid sequences as shown in SEQ ID NOs: 17, 20, 23, 26, 29, 32, 35, 38, 41, 44, 47, 50, 53, and 56 or variants thereof, preferably, exhibit DGAT activity. A polynucleotide having a nucleic acid sequence as shown in SEQ ID NO: 55, encoding a polypeptide having amino acid sequences as shown in SEQ ID NO: 56 or variants thereof, preferably, exhibit GPAT activity.

[0028] A polynucleotide encoding a polypeptide having a acyltransferase activity as specified above has been obtained in accordance with the present invention, preferably, from *Nannochloropsis oculata* and/or *Thraustochytrium aureum*. However, orthologs, paralogs or other homologs may be identified from other species.

[0029] Thus, the term "polynucleotide" as used in accordance with the present invention further encompasses variants of the aforementioned specific polynucleotides representing orthologs, paralogs or other homologs of the polynucleotide of the present invention. Moreover, variants of the polynucleotide of the present invention also include artificially generated muteins. Said muteins include, e.g., enzymes which are generated by mutagenesis techniques and which exhibit improved or altered substrate specificity, or codon optimized polynucleotides. The polynucleotide variants, preferably, comprise a nucleic acid sequence characterized in that the sequence can be derived from the aforementioned specific nucleic acid sequences shown in any one of SEQ ID NOs: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, and 55 or by a polynucleotide encoding a polypeptide having an amino acid sequence as shown in any one of SEQ ID NOs: 53, 2, 5, 8, 11, 14, 17, 20, 23, 26, 29, 32, 35, 38, 41, 44, 47, 50, and 56 by at least one nucleotide substitution, addition and/or deletion, whereby the variant

nucleic acid sequence shall still encode a polypeptide having a acyltransferase activity as specified above. Variants also encompass polynucleotides comprising a nucleic acid sequence which is capable of hybridizing to the aforementioned specific nucleic acid sequences, preferably, under stringent hybridization conditions. These stringent conditions are known to the skilled artisan and can be found in Current Protocols in Molecular Biology, John Wiley & Sons, N. Y. (1989), 6.3.1-6.3.6. A preferred example for stringent hybridization conditions are hybridization conditions in 6× sodium chloride/sodium citrate (=SSC) at approximately 45° C., followed by one or more wash steps in 0.2×SSC, 0.1% SDS at 50 to 65° C. The skilled artisan knows that these hybridization conditions differ depending on the type of nucleic acid and, for example when organic solvents are present, with regard to the temperature and concentration of the buffer. For example, under “standard hybridization conditions” the temperature differs depending on the type of nucleic acid between 42° C. and 58° C. in aqueous buffer with a concentration of 0.1 to 6×SSC (pH 7.2). If organic solvent is present in the abovementioned buffer, for example 50% formamide, the temperature under standard conditions is approximately 42° C. The hybridization conditions for DNA:DNA hybrids are, preferably, 0.1×SSC and 20° C. to 45° C., preferably between 30° C. and 45° C. and more preferably between 45° C. and 65° C. The hybridization conditions for DNA:RNA hybrids are, more preferably, 0.1×SSC and 30° C. to 55° C., most preferably between 45° C. and 65° C. The abovementioned hybridization temperatures are determined for example for a nucleic acid with approximately 100 bp (=base pairs) in length and a G+C content of 50% in the absence of formamide. The skilled artisan knows how to determine the hybridization conditions required by referring to textbooks such as the textbook mentioned above, or the following textbooks: Sambrook et al., “Molecular Cloning”, Cold Spring Harbor Laboratory, 1989; Hames and Higgins (Ed.) 1985, “Nucleic Acids Hybridization: A Practical Approach”, IRL Press at Oxford University Press, Oxford; Brown (Ed.) 1991, “Essential Molecular Biology: A Practical Approach”, IRL Press at Oxford University Press, Oxford.

[0030] In detail variants of polynucleotides still encode a polypeptide having a acyltransferase activity as specified above comprising a nucleic acid sequence which is capable of hybridizing preferably under conditions equivalent to hybridization in 50 mM Tris, pH 7.6, 6×SSC, 5×Denhardt’s, 1.0% sodium dodecyl sulfat (SDS) 100 µg denaturated calf thymus DNA at 34° C. overnight, followed by washing twice with 2×SSC, 0.5% SDS at room temperature for 15 min each, then wash twice with 0.2×SSC, 0.5% SDS at room temperature for 15 min each and then wash twice with 0.2 SSC, 0.5% SDS at 50° C. for 15 min each to a nucleic acid described by any one of SEQ ID NOs: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, and 55 or the complement thereof.

[0031] More preferably, said variants of polynucleotides comprising a nucleic acid sequence which is capable of hybridizing under conditions equivalent to hybridization in 6×SSPE (Sodium chloride Sodium Phosphate-EDTA), 5×Denhardt’s solution, 0.5% sodium dodecyl sulfat (SDS) 100 µg denaturated calf thymus DNA at 34° C. overnight, followed by washing twice with 2×SSC, 0.5% SDS at room temperature for 15 min each, then wash twice with 0.2×SSC, 0.5% SDS at room temperature for 15 min each and then wash twice with 0.2 SSC, 0.5% SDS at 50° C. for 15 min each to a

nucleic acid described by any one of SEQ ID NOs: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, and 55 or the complement thereof.

[0032] Most preferably, said variants of polynucleotides comprising a nucleic acid sequence which is capable of hybridizing under conditions equivalent to hybridization in 20-30% formamide, 5×SSPE (Sodium chloride Sodium Phosphate-EDTA), 5×Denhardt’s solution, 1% sodium dodecyl sulfat (SDS) 100 µg denaturated salmon sperm DNA at 34° C. overnight, followed by washing twice with 2×SSPE, 0.2% SDS at 42° C. for 15 min each, then wash twice with 2×SSPE, 0.2% SDS at 55° C. for 30 min each and then wash twice with 0.2 SSC, 0.5% SDS at 50° C. for 15 min each to a nucleic acid described by any one of SEQ ID NOs: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, and 55 or the complement thereof.

[0033] In another preferred embodiment aforementioned variants of polynucleotides still encode a polypeptide having a acyltransferase activity as specified above comprising a nucleic acid sequence which is capable of hybridizing under conditions equivalent to hybridization in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50° C. overnight with washing in 2×SSC, 0.1% SDS at 50° C. or 65° C., preferably 65° C. to a nucleic acid described by any one of SEQ ID NOs: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, and 55 or the complement thereof. In still another preferred embodiment, said variants of polynucleotides comprising a nucleic acid sequence which is capable of hybridizing under conditions equivalent to hybridization in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50° C. overnight with washing in 1×SSC, 0.1% SDS at 50° C. or 65° C., preferably 65° C. to a nucleotide sequence described by any one of SEQ ID NOs: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, and 55 or the complement thereof, most preferably, said variants of polynucleotides comprising a nucleic acid sequence which is capable of hybridizing under conditions equivalent to hybridization in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50° C. overnight with washing in 0.1×SSC, 0.1% SDS at 50° C. or 65° C., preferably 65° C. to a nucleic acid sequence described by any one of SEQ ID NOs: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, and 55 or the complement thereof.

[0034] The term “hybridization” as used herein includes “any process by which a strand of nucleic acid molecule joins with a complementary strand through base pairing.” (J. Coombs (1994) Dictionary of Biotechnology, Stockton Press, New York). Hybridization and the strength of hybridization (i.e., the strength of the association between the nucleic acid molecules) is impacted by such factors as the degree of complementarity between the nucleic acid molecules, stringency of the conditions involved, the T_m of the formed hybrid, and the G:C ratio within the nucleic acid molecules. As used herein, the term “T_m” is used in reference to the “melting temperature.” The melting temperature is the temperature at which a population of double-stranded nucleic acid molecules becomes half dissociated into single strands. The equation for calculating the T_m of nucleic acid molecules is well known in the art. As indicated by standard references, a simple estimate of the T_m value may be calculated by the equation: T_m=81.5+0.41 (% G+C), when a nucleic acid molecule is in aqueous solution at 1 M NaCl [see e.g., Anderson and Young, Quantitative Filter Hybridization, in Nucleic Acid Hybridization (1985)]. Other references include more sophis-

ticated computations, which take structural as well as sequence characteristics into account for the calculation of Tm. Stringent conditions, are known to those skilled in the art and can be found in Current Protocols in Molecular Biology, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6.

[0035] A “complement” of a nucleic acid sequence as used herein refers to a nucleotide sequence whose nucleic acid molecules show total complementarity to the nucleic acid molecules of the nucleic acid sequence.

[0036] The term “Complementary” or “complementarity” refers to two nucleotide sequences which comprise antiparallel nucleotide sequences capable of pairing with one another (by the base-pairing rules) upon formation of hydrogen bonds between the complementary base residues in the antiparallel nucleotide sequences. For example, the sequence 5'-AGT-3' is complementary to the sequence 5'-ACT-3'. Complementarity can be “partial” or “total.” “Partial” complementarity is where one or more nucleic acid bases are not matched according to the base pairing rules. “Total” or “complete” complementarity between nucleic acid molecules is where each and every nucleic acid base is matched with another base under the base pairing rules. The degree of complementarity between nucleic acid molecule strands has significant effects on the efficiency and strength of hybridization between nucleic acid molecule strands.

[0037] Alternatively, polynucleotide variants are obtainable by PCR-based techniques such as mixed oligonucleotide primer-based amplification of DNA, i.e. using degenerated primers against conserved domains of the polypeptides of the present invention. Conserved domains of the polypeptide of the present invention may be identified by a sequence comparison of the nucleic acid sequences of the polynucleotides or the amino acid sequences of the polypeptides of the present invention. Oligonucleotides suitable as PCR primers as well as suitable PCR conditions are described in the accompanying Examples. As a template, DNA or cDNA from bacteria, fungi, plants or animals may be used.

[0038] Further, variants include polynucleotides comprising nucleic acid sequences which are at least up to 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% identical to the nucleic acid sequences shown in any one of SEQ ID NOS: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, and 55, preferably, encoding polypeptides retaining an acyltransferase activity as specified above.

[0039] Moreover, also encompassed are polynucleotides (derivatives) which comprise nucleic acid sequences encoding a polypeptide having an amino acid sequences which are at least up to 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% identical to the amino acid sequences shown in any one of SEQ ID NOS: 53, 2, 5, 8, 11, 14, 17, 20, 23, 26, 29, 32, 35, 38, 41, 44, 47, 50, and 56, wherein the polypeptide, preferably, retains acyltransferase activity as specified above. The percent identity values are, preferably, calculated over the entire amino acid or nucleic acid sequence region. A series of programs based on a variety of algorithms is available to the skilled artisan for comparing different sequences. In a preferred embodiment, the percent identity

between two amino acid sequences is determined using the Needleman and Wunsch algorithm (Needleman 1970, J. Mol. Biol. (48):444-453) which has been incorporated into the needle program in the EMBOSS software package (*EMBOSS: The European Molecular Biology Open Software Suite*, Rice, P., Longden, I., and Bleasby, A, Trends in Genetics 16(6), 276-277, 2000), using either a BLOSUM 45 or PAM250 scoring matrix for distantly related proteins, or either a BLOSUM 62 or PAM 160 scoring matrix for closer related proteins, and a gap opening penalty of 16, 14, 12, 10, 8, 6, or 4 and a gap extension penalty of 0.5, 1, 2, 3, 4, 5, or 6. Guides for local installation of the EMBOSS package as well as links to WEB-Services can be found at <http://emboss.sourceforge.net>. A preferred, non-limiting example of parameters to be used for aligning two amino acid sequences using the needle program are the default parameters, including the EBLOSUM62 scoring matrix, a gap opening penalty of 10 and a gap extension penalty of 0.5. In yet another preferred embodiment, the percent identity between two nucleotide sequences is determined using the needle program in the EMBOSS software package (*EMBOSS: The European Molecular Biology Open Software Suite*, Rice, P., Longden, I., and Bleasby, A, Trends in Genetics 16(6), 276-277, 2000), using the EDNAFULL scoring matrix and a gap opening penalty of 16, 14, 12, 10, 8, 6, or 4 and a gap extension penalty of 0.5, 1, 2, 3, 4, 5, or 6. A preferred, non-limiting example of parameters to be used in conjunction for aligning two amino acid sequences using the needle program are the default parameters, including the EDNAFULL scoring matrix, a gap opening penalty of 10 and a gap extension penalty of 0.5. The nucleic acid and protein sequences of the present invention can further be used as a “query sequence” to perform a search against public databases to, for example, identify other family members or related sequences. Such searches can be performed using the BLAST series of programs (version 2.2) of Altschul et al. (Altschul 1990, J. Mol. Biol. 215:403-10). BLAST using acyltransferase nucleic acid sequences of the invention as query sequence can be performed with the BLASTn, BLASTx or the tBLASTx program using default parameters to obtain either nucleotide sequences (BLASTn, tBLASTx) or amino acid sequences (BLASTx) homologous to acyltransferase sequences of the invention. BLAST using acyltransferase protein sequences of the invention as query sequence can be performed with the BLASTp or the tBLASTn program using default parameters to obtain either amino acid sequences (BLASTp) or nucleic acid sequences (tBLASTn) homologous to acyltransferase sequences of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST using default parameters can be utilized as described in Altschul et al. (Altschul 1997, Nucleic Acids Res. 25(17):3389-3402).

TABLE 1

Relation of sequence types of query and hit sequences for various BLAST programs				
Input query sequence	Converted Query	Algorithm	Converted Hit	Actual Database
DNA		BLASTn		DNA
PRT		BLASTp		PRT
DNA	PRT	BLASTx		PRT
PRT		tBLASTn	PRT	DNA
DNA	PRT	tBLASTx	PRT	DNA

[0040] A polynucleotide comprising a fragment of any of the aforementioned nucleic acid sequences is also encompassed as a polynucleotide of the present invention. The fragment shall encode a polypeptide which still has acyltransferase activity as specified above. Accordingly, the polypeptide may comprise or consist of the domains of the polypeptide of the present invention conferring the said biological activity. A fragment as meant herein, preferably, comprises at least 50, at least 100, at least 250 or at least 500 consecutive nucleotides of any one of the aforementioned nucleic acid sequences or encodes an amino acid sequence comprising at least 20, at least 30, at least 50, at least 80, at least 100 or at least 150 consecutive amino acids of any one of the aforementioned amino acid sequences.

[0041] The variant polynucleotides or fragments referred to above, preferably, encode polypeptides retaining acyltransferase activity to a significant extent, preferably, at least 10%, at least 20%, at least 30%, at least 40%, at least 50%, at least 60%, at least 70%, at least 80% or at least 90% of the acyltransferase activity exhibited by any of the polypeptide shown in any one of SEQ ID NOs: 2, 5, 8, 11, 14, 17, 20, 23, 26, 29, 32, 35, 38, 41, 44, 47, 50, 53, and 56 or derivative of any of these polypeptides. The activity may be tested as described in the accompanying examples.

[0042] The polynucleotides of the present invention either essentially consist of the aforementioned nucleic acid sequences or comprise the aforementioned nucleic acid sequences. Thus, they may contain further nucleic acid sequences as well. Preferably, the polynucleotide of the present invention may comprise in addition to an open reading frame further untranslated sequence at the 3' and at the 5' terminus of the coding gene region: at least 500, preferably 200, more preferably 100 nucleotides of the sequence upstream of the 5' terminus of the coding region and at least 100, preferably 50, more preferably 20 nucleotides of the sequence downstream of the 3' terminus of the coding gene region. Furthermore, the polynucleotides of the present invention may encode fusion proteins wherein one partner of the fusion protein is a polypeptide being encoded by a nucleic acid sequence recited above. Such fusion proteins may comprise as additional part other enzymes of the fatty acid or PUFA biosynthesis pathways, polypeptides for monitoring expression (e.g., green, yellow, blue or red fluorescent proteins, alkaline phosphatase and the like) or so called "tags" which may serve as a detectable marker or as an auxiliary measure for purification purposes. Tags for the different purposes are well known in the art and comprise FLAG-tags, 6-histidine-tags, MYC-tags and the like.

[0043] The polynucleotide of the present invention shall be provided, preferably, either as an isolated polynucleotide (i.e. purified or at least isolated from its natural context such as its natural gene locus) or in genetically modified or exogenously (i.e. artificially) manipulated form. An isolated polynucleotide can, for example, comprise less than approximately 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in the genomic DNA of the cell from which the nucleic acid is derived. The polynucleotide, preferably, is provided in the form of double or single stranded molecule. It will be understood that the present invention by referring to any of the aforementioned polynucleotides of the invention also refers to complementary or reverse complementary strands of the specific sequences or variants thereof referred to before. The

polynucleotide encompasses DNA, including cDNA and genomic DNA, or RNA polynucleotides.

[0044] However, the present invention also pertains to polynucleotide variants which are derived from the polynucleotides of the present invention and are capable of interfering with the transcription or translation of the polynucleotides of the present invention. Such variant polynucleotides include anti-sense nucleic acids, ribozymes, siRNA molecules, morpholino nucleic acids (phosphorodiamidate morpholino oligos), triple-helix forming oligonucleotides, inhibitory oligonucleotides, or micro RNA molecules all of which shall specifically recognize the polynucleotide of the invention due to the presence of complementary or substantially complementary sequences. These techniques are well known to the skilled artisan. Suitable variant polynucleotides of the aforementioned kind can be readily designed based on the structure of the polynucleotides of this invention.

[0045] Moreover, comprised are also chemically modified polynucleotides including naturally occurring modified polynucleotides such as glycosylated or methylated polynucleotides or artificial modified ones such as biotinylated polynucleotides.

[0046] Advantageously, it has been found in accordance with the present invention that the polynucleotides encoding the above mentioned polypeptides having acyltransferase activity and, in particular, LPLAT, LPAAT, DGAT and/or GPAT activity, can be used for the manufacture of PUFA and, in particular, LCPUFA when expressed in a transgenic host organism or cell. Specifically, the aforementioned acyltransferase activities will allow for an increase of LCPUFA esterified to triglycerides in seed oils by shifting the said LCPUFA from the acyl-CoA pool (by polypeptides having LPAAT, DGAT or GPAT activity as specified above) and/or from the acyl-CoA pool/phospholipid pool to the phospholipid pool/acyl-CoA pool (by polypeptides having LPLAT as specified above) via transesterification. Surprisingly, it was found that the acyltransferases encoded by the polynucleotides of the present invention are also capable of efficiently shifting rather long and highly unsaturated LCPUFA towards the triglyceride pool or between the phospholipid pool and the acyl-CoA pool, in particular, even the long chain intermediates. More surprisingly even, DHA which is known to be incorporated in triglycerides only in very low amounts, if at all, can be efficiently transesterified to triglycerides by the acyltransferases of the invention.

[0047] In particular the LPLAT of the present invention can efficiently catalyse the transesterification of 18:2n-6 from the sn2 position of phosphatidylcholine (PC) to CoA and/or from CoA to lysophosphatidylcholine (LPC), the transesterification of 18:2n-6 from the sn2 position of phosphatidylethanolamine (PE) to CoA and/or from CoA to lysophosphatidylethanolamine (LPE), the transesterification of 18:2n-6 from the sn2 position of phosphatidylserine (PS) to CoA and/or from CoA to lysophosphatidylserine (LPS), the transesterification of 18:3n-6 from the sn2 position of phosphatidylcholine (PC) to CoA and/or from CoA to lysophosphatidylcholine (LPC), the transesterification of 18:3n-6 from the sn2 position of phosphatidylethanolamine (PE) to CoA and/or from CoA to lysophosphatidylethanolamine (LPE), the transesterification of 18:3n-6 from the sn2 position of phosphatidylserine (PS) to CoA and/or from CoA to lysophosphatidylserine (LPS), the transesterification of 18:3n-3 from the sn2 position of phosphatidylcholine (PC) to CoA and/or from CoA to lysophosphatidylcholine (LPC), the transesterification of 18:3n-3

ity of the LPLAT, LPAAT, GPAT or DGAT of the present invention is useful to generate an artificially EPA-specificity. Most preferably the activity of the LPLAT, LPAAT, GPAT or DGAT of the present invention is useful to generate an artificially DHA-specificity.

[0058] In a preferred embodiment of the polynucleotide of the present invention, said polynucleotide further comprises an expression control sequence operatively linked to the said nucleic acid sequence.

[0059] The term “expression control sequence” as used herein refers to a nucleic acid sequence which is capable of governing, i.e. initiating and controlling, transcription of a nucleic acid sequence of interest, in the present case the nucleic sequences recited above. Such a sequence usually comprises or consists of a promoter or a combination of a promoter and enhancer sequences. Expression of a polynucleotide comprises transcription of the nucleic acid molecule, preferably, into a translatable mRNA. Additional regulatory elements may include transcriptional as well as translational enhancers. The following promoters and expression control sequences may be, preferably, used in an expression vector according to the present invention. The *cos*, *tac*, *trp*, *tet*, *trp-tet*, *lpp*, *lac*, *lpp-lac*, *lacIq*, *T7*, *T5*, *T3*, *gal*, *trc*, *ara*, *SP6*, λ -PR or λ -PL promoters are, preferably, used in Gram-negative bacteria. For Gram-positive bacteria, promoters *amy* and *SPO2* may be used. From yeast or fungal promoters *ADC1*, *AOX1r*, *GAL1*, *MF α* , *AC*, *P-60*, *CYC1*, *GAPDH*, *TEF*, *rp28*, *ADH* are, preferably, used. For animal cell or organism expression, the promoters *CMV-*, *SV40-*, *RSV-promoter* (Rous sarcoma virus), *CMV-enhancer*, *SV40-enhancer* are preferably used. From plants the promoters *CaMV/35S* (Franck 1980, *Cell* 21: 285-294), *PRP1* (Ward 1993, *Plant. Mol. Biol.* 22), *SSU*, *OCS*, *lib4*, *usp*, *STLS1*, *B33*, *nos* or the ubiquitin or phaseolin promoter. Also preferred in this context are inducible promoters, such as the promoters described in EP 0 388 186 A1 (i.e. a benzylsulfonamide-inducible promoter), Gatz 1992, *Plant J.* 2:397-404 (i.e. a tetracyclin-inducible promoter), EP 0 335 528 A1 (i.e. an abscisic-acid-inducible promoter) or WO 93/21334 (i.e. an ethanol- or cyclohexenol-inducible promoter). Further suitable plant promoters are the promoter of cytosolic FBPase or the *ST-LSI* promoter from potato (Stockhaus 1989, *EMBO J.* 8, 2445), the phosphoribosyl-pyrophosphate amidotransferase promoter from *Glycine max* (Genbank accession No. U87999) or the node-specific promoter described in EP 0 249 676 A1. Particularly preferred are promoters which enable the expression in tissues which are involved in the biosynthesis of fatty acids. Also particularly preferred are seed-specific promoters such as the *USP* promoter in accordance with the practice, but also other promoters such as the *LeB4*, *DC3*, phaseolin or napin promoters. Further especially preferred promoters are seed-specific promoters which can be used for monocotyledonous or dicotyledonous plants and which are described in U.S. Pat. No. 5,608,152 (napin promoter from oilseed rape), WO 98/45461 (oleosin promoter from *Arabidopsis*, U.S. Pat. No. 5,504,200 (phaseolin promoter from *Phaseolus vulgaris*), WO 91/13980 (Bce4 promoter from *Brassica*), by Baumlein et al., *Plant J.*, 2, 2, 1992:233-239 (*LeB4* promoter from a legume), these promoters being suitable for dicots. The following promoters are suitable for monocots: *Ipt-2* or *Ipt-1* promoter from barley (WO 95/15389 and WO 95/23230), hordein promoter from barley and other promoters which are suitable and which are described in WO 99/16890. In principle, it is possible to use all natural pro-

motors together with their regulatory sequences, such as those mentioned above, for the novel process. Likewise, it is possible and advantageous to use synthetic promoters, either additionally or alone, especially when they mediate a seed-specific expression, such as, for example, as described in WO 99/16890. In a particular embodiment, seed-specific promoters are utilized to enhance the production of the desired PUFA or LCPUFA.

[0060] The term “operatively linked” as used herein means that the expression control sequence and the nucleic acid of interest are linked so that the expression of the said nucleic acid of interest can be governed by the said expression control sequence, i.e. the expression control sequence shall be functionally linked to the said nucleic acid sequence to be expressed. Accordingly, the expression control sequence and, the nucleic acid sequence to be expressed may be physically linked to each other, e.g., by inserting the expression control sequence at the 5'end of the nucleic acid sequence to be expressed. Alternatively, the expression control sequence and the nucleic acid to be expressed may be merely in physical proximity so that the expression control sequence is capable of governing the expression of at least one nucleic acid sequence of interest. The expression control sequence and the nucleic acid to be expressed are, preferably, separated by not more than 500 bp, 300 bp, 100 bp, 80 bp, 60 bp, 40 bp, 20 bp, 10 bp or 5 bp.

[0061] In a further preferred embodiment of the polynucleotide of the present invention, said polynucleotide further comprises a terminator sequence operatively linked to the nucleic acid sequence.

[0062] The term “terminator” as used herein refers to a nucleic acid sequence which is capable of terminating transcription. These sequences will cause dissociation of the transcription machinery from the nucleic acid sequence to be transcribed. Preferably, the terminator shall be active in plants and, in particular, in plant seeds. Suitable terminators are known in the art and, preferably, include polyadenylation signals such as the *SV40-poly-A* site or the *tk-poly-A* site or one of the plant specific signals indicated in Loke et al. 2005, *Plant Physiol* 138, pp. 1457-1468, downstream of the nucleic acid sequence to be expressed.

[0063] The present invention also relates to a vector comprising the polynucleotide of the present invention.

[0064] The term “vector”, preferably, encompasses phage, plasmid, viral vectors as well as artificial chromosomes, such as bacterial or yeast artificial chromosomes. Moreover, the term also relates to targeting constructs which allow for random or site-directed integration of the targeting construct into genomic DNA. Such target constructs, preferably, comprise DNA of sufficient length for either homologous or heterologous recombination as described in detail below. The vector encompassing the polynucleotide of the present invention, preferably, further comprises selectable markers for propagation and/or selection in a host. The vector may be incorporated into a host cell by various techniques well known in the art. If introduced into a host cell, the vector may reside in the cytoplasm or may be incorporated into the genome. In the latter case, it is to be understood that the vector may further comprise nucleic acid sequences which allow for homologous recombination or heterologous insertion. Vectors can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. The terms “transformation” and “transfection”, conjugation and transduction, as used in the present context, are intended to com-

prise a multiplicity of prior-art processes for introducing foreign nucleic acid (for example DNA) into a host cell, including calcium phosphate, rubidium chloride or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, natural competence, carbon-based clusters, chemically mediated transfer, electroporation or particle bombardment. Suitable methods for the transformation or transfection of host cells, including plant cells, can be found in Sambrook et al. (Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989) and other laboratory manuals, such as Methods in Molecular Biology, 1995, Vol. 44, *Agrobacterium* protocols, Ed.: Gartland and Davey, Humana Press, Totowa, N.J. Alternatively, a plasmid vector may be introduced by heat shock or electroporation techniques. Should the vector be a virus, it may be packaged in vitro using an appropriate packaging cell line prior to application to host cells.

[0065] Preferably, the vector referred to herein is suitable as a cloning vector, i.e. replicable in microbial systems. Such vectors ensure efficient cloning in bacteria and, preferably, yeasts or fungi and make possible the stable transformation of plants. Those which must be mentioned are, in particular, various binary and co-integrated vector systems which are suitable for the T-DNA-mediated transformation. Such vector systems are, as a rule, characterized in that they contain at least the *vir* genes, which are required for the *Agrobacterium*-mediated transformation, and the sequences which delimit the T-DNA (T-DNA border). These vector systems, preferably, also comprise further cis-regulatory regions such as promoters and terminators and/or selection markers with which suitable transformed host cells or organisms can be identified. While co-integrated vector systems have *vir* genes and T-DNA sequences arranged on the same vector, binary systems are based on at least two vectors, one of which bears *vir* genes, but no T-DNA, while a second one bears T-DNA, but no *vir* gene. As a consequence, the last-mentioned vectors are relatively small, easy to manipulate and can be replicated both in *E. coli* and in *Agrobacterium*. These binary vectors include vectors from the pBIB-HYG, pPZP, pBecks, pGreen series. Preferably used in accordance with the invention are Bin19, pB1101, pBinAR, pGPTV and pCAMBIA. An overview of binary vectors and their use can be found in Hellens et al, Trends in Plant Science (2000) 5, 446-451. Furthermore, by using appropriate cloning vectors, the polynucleotides can be introduced into host cells or organisms such as plants or animals and, thus, be used in the transformation of plants, such as those which are published, and cited, in: Plant Molecular Biology and Biotechnology (CRC Press, Boca Raton, Fla.), chapter 6/7, pp. 71-119 (1993); F. F. White, Vectors for Gene Transfer in Higher Plants; in: Transgenic Plants, vol. 1, Engineering and Utilization, Ed.: Kung and R. Wu, Academic Press, 1993, 15-38; B. Jenes et al., Techniques for Gene Transfer, in: Transgenic Plants, vol. 1, Engineering and Utilization, Ed.: Kung and R. Wu, Academic Press (1993), 128-143; Potrykus 1991, Annu. Rev. Plant Physiol. Plant Molec. Biol. 42, 205-225.

[0066] More preferably, the vector of the present invention is an expression vector. In such an expression vector, i.e. a vector which comprises the polynucleotide of the invention having the nucleic acid sequence operatively linked to an expression control sequence (also called "expression cassette") allowing expression in prokaryotic or eukaryotic cells or isolated fractions thereof. Suitable expression vectors are

known in the art such as Okayama-Berg cDNA expression vector pcDV1 (Pharmacia), pCDM8, pRc/CMV, pcDNA1, pcDNA3 (Invitrogen) or pSPORT1 (GIBCO BRL). Further examples of typical fusion expression vectors are pGEX (Pharmacia Biotech Inc; Smith 1988, Gene 67:31-40), pMAL (New England Biolabs, Beverly, Mass.) and pRIT5 (Pharmacia, Piscataway, N.J.), where glutathione S-transferase (GST), maltose E-binding protein and protein A, respectively, are fused with the recombinant target protein. Examples of suitable inducible non-fusion *E. coli* expression vectors are, inter alia, pTrc (Amann 1988, Gene 69:301-315) and pET 11d (Studier 1990, Methods in Enzymology 185, 60-89). The target gene expression of the pTrc vector is based on the transcription from a hybrid *trp*-lac fusion promoter by host RNA polymerase. The target gene expression from the pET 11d vector is based on the transcription of a T7-gn10-lac fusion promoter, which is mediated by a coexpressed viral RNA polymerase (T7 gn1). This viral polymerase is provided by the host strains BL21 (DE3) or HMS174 (DE3) from a resident λ -prophage which harbors a T7 gn1 gene under the transcriptional control of the lacUV 5 promoter. The skilled artisan is familiar with other vectors which are suitable in prokaryotic organisms; these vectors are, for example, in *E. coli*, pLG338, pACYC184, the pBR series such as pBR322, the pUC series such as pUC18 or pUC19, the M113mp series, pKC30, pRep4, pHS1, pHS2, pPLc236, pMBL24, pLG200, pUR290, pIN-III113-B1, λ gt11 or pBdCl, in *Streptomyces* pIJ101, pIJ364, pIJ702 or pIJ361, in *Bacillus* pUB110, pC194 or pBD214, in *Corynebacterium* pSA77 or pAJ667. Examples of vectors for expression in the yeast *S. cerevisiae* comprise pYep Sec1 (Baldari 1987, Embo J. 6:229-234), pMfa (Kurjan 1982, Cell 30:933-943), pJRY88 (Schultz 1987, Gene 54:113-123) and pYES2 (Invitrogen Corporation, San Diego, Calif.). Vectors and processes for the construction of vectors which are suitable for use in other fungi, such as the filamentous fungi, comprise those which are described in detail in: van den Hondel, C. A. M. J. J., & Punt, P. J. (1991) "Gene transfer systems and vector development for filamentous fungi, in: Applied Molecular Genetics of fungi, J. F. Peberdy et al., Ed., pp. 1-28, Cambridge University Press: Cambridge, or in: More Gene Manipulations in Fungi (J. W. Bennett & L. L. Lasure, Ed., pp. 396-428: Academic Press: San Diego). Further suitable yeast vectors are, for example, pAG-1, YE6, YE13 or pEMBLye23. As an alternative, the polynucleotides of the present invention can be also expressed in insect cells using baculovirus expression vectors. Baculovirus vectors which are available for the expression of proteins in cultured insect cells (for example Sf9 cells) comprise the pAc series (Smith 1983, Mol. Cell Biol. 3:2156-2165) and the pVL series (Lucklow 1989, Virology 170:31-39).

[0067] The polynucleotide of the present invention can be expressed in single-cell plant cells (such as algae), see Falcitore 1999, Marine Biotechnology 1 (3):239-251 and the references cited therein, and plant cells from higher plants (for example Spermatophytes, such as arable crops) by using plant expression vectors. Examples of plant expression vectors comprise those which are described in detail in: Becker 1992, Plant Mol. Biol. 20:1195-1197; Bevan 1984, Nucl. Acids Res. 12:8711-8721; Vectors for Gene Transfer in Higher Plants; in: Transgenic Plants, Vol. 1, Engineering and Utilization, Ed.: Kung and R. Wu, Academic Press, 1993, p. 15-38. A plant expression cassette, preferably, comprises regulatory sequences which are capable of controlling the

gene expression in plant cells and which are functionally linked so that each sequence can fulfill its function, such as transcriptional termination, for example polyadenylation signals. Preferred polyadenylation signals are those which are derived from *Agrobacterium tumefaciens* T-DNA, such as the gene 3 of the Ti plasmid pTiACH5, which is known as octopine synthase (Gielen 1984, EMBO J. 3, 835) or functional equivalents of these, but all other terminators which are functionally active in plants are also suitable. Since plant gene expression is very often not limited to transcriptional levels, a plant expression cassette preferably comprises other functionally linked sequences such as translation enhancers, for example the override sequence, which comprises the 5'-untranslated tobacco mosaic virus leader sequence, which increases the protein/RNA ratio (Gallie 1987, Nucl. Acids Research 15:8693-8711). As described above, plant gene expression must be functionally linked to a suitable promoter which performs the expression of the gene in a timely, cell-specific or tissue-specific manner. Promoters which can be used are constitutive promoters (Benfey 1989, EMBO J. 8:2195-2202) such as those which are derived from plant viruses such as 35S CAMV (Franck 1980, Cell 21:285-294), 19S CaMV (see U.S. Pat. No. 5,352,605 and WO 84/02913) or plant promoters such as the promoter of the Rubisco small subunit, which is described in U.S. Pat. No. 4,962,028. Other preferred sequences for the use in functional linkage in plant gene expression cassettes are targeting sequences which are required for targeting the gene product into its relevant cell compartment (for a review, see Kermodé 1996, Crit. Rev. Plant Sci. 15, 4: 285-423 and references cited therein), for example into the vacuole, the nucleus, all types of plastids, such as amyloplasts, chloroplasts, chromoplasts, the extracellular space, the mitochondria, the endoplasmic reticulum, oil bodies, peroxisomes and other compartments of plant cells. As described above, plant gene expression can also be facilitated via a chemically inducible promoter (for a review, see Gatz 1997, Annu. Rev. Plant Physiol. Plant Mol. Biol., 48:89-108). Chemically inducible promoters are particularly suitable if it is desired that genes are expressed in a time-specific manner. Examples of such promoters are a salicylic-acid-inducible promoter (WO 95/19443), a tetracyclin-inducible promoter (Gatz 1992, Plant J. 2, 397-404) and an ethanol-inducible promoter. Promoters which respond to biotic or abiotic stress conditions are also suitable promoters, for example the pathogen-induced PRP1-gene promoter (Ward 1993, Plant Mol. Biol. 22:361-366), the heat-inducible hsp80 promoter from tomato (U.S. Pat. No. 5,187,267), the cold-inducible alpha-amylase promoter from potato (WO 96/12814) or the wound-inducible pinII promoter (EP 0 375 091 A). The promoters which are especially preferred are those which bring about the expression of genes in tissues and organs in which fatty acid, lipid and oil biosynthesis takes place, in seed cells such as the cells of endosperm and of the developing embryo.

[0068] Suitable promoters are the napin gene promoter from oilseed rape (U.S. Pat. No. 5,608,152), the USP promoter from *Vicia faba* (Baumlein 1991, Mol. Gen. Genet. 225 (3):459-67), the oleosin promoter from *Arabidopsis* (WO 98/45461), the phaseolin promoter from *Phaseolus vulgaris* (U.S. Pat. No. 5,504,200), the Bce4 promoter from *Brassica* (WO 91/13980) or the legumin B4 promoter (LeB4; Baumlein 1992, Plant Journal, 2 (2):233-9), and promoters which bring about the seed-specific expression in monocotyledonous plants such as maize, barley, wheat, rye, rice and the like.

Suitable promoters to be taken into consideration are the Ipt2 or Ipt1 gene promoter from barley (WO 95/15389 and WO 95/23230) or those which are described in WO 99/16890 (promoters from the barley hordein gene, the rice glutelin gene, the rice oryza gene, the rice prolamin gene, the wheat gliadin gene, wheat glutelin gene, the maize zein gene, the oat glutelin gene, the *sorghum* kasirin gene, the rye secalin gene). Likewise, especially suitable are promoters which bring about the plastid-specific expression since plastids are the compartment in which the precursors and some end products of lipid biosynthesis are synthesized. Suitable promoters such as the viral RNA-polymerase promoter are described in WO 95/16783 and WO 97/06250, and the clpP promoter from *Arabidopsis*, described in WO 99/46394.

[0069] The abovementioned vectors are only a small overview of vectors to be used in accordance with the present invention. Further vectors are known to the skilled artisan and are described, for example, in: Cloning Vectors (Ed., Pouwels, P. H., et al., Elsevier, Amsterdam-New York-Oxford, 1985, ISBN 0 444 904018). For further suitable expression systems for prokaryotic and eukaryotic cells see the chapters 16 and 17 of Sambrook, loc. cit.

[0070] It follows from the above that, preferably, said vector is an expression vector. More preferably, the said polynucleotide of the present invention is under the control of a seed-specific promoter in the vector of the present invention. A preferred seed-specific promoter as meant herein is selected from the group consisting of Conlinin 1, Conlinin 2, napin, LuFad3, USP, LeB4, Arc, Fae, ACP, LuPXR, and SBP. For details, see, e.g., US 2003-0159174.

[0071] Moreover, the present invention relates to a host cell comprising the polynucleotide or the vector of the present invention.

[0072] Preferably, said host cell is a plant cell and, more preferably, a plant cell obtained from an oilseed crop. More preferably, said oilseed crop is selected from the group consisting of flax (*Linum* sp.), rapeseed (*Brassica* sp.), soybean (*Glycine* and *Soja* sp.), sunflower (*Helianthus* sp.), cotton (*Gossypium* sp.), corn (*Zea mays*), olive (*Olea* sp.), safflower (*Carthamus* sp.), cocoa (*Theobroma cacao*), peanut (*Arachis* sp.), hemp, camelina, *crambe*, oil palm, coconuts, groundnuts, sesame seed, castor bean, *lesquerella*, tallow tree, sheanuts, tungnuts, kapok fruit, poppy seed, jojoba seeds and *perilla*.

[0073] Also preferably, said host cell is a microorganism. More preferably, said microorganism is a bacterium, a fungus or algae. More preferably, it is selected from the group consisting of *Candida*, *Cryptococcus*, *Lipomyces*, *Rhodospiridium*, *Yarrowia* and *Schizochytrium*.

[0074] Moreover, a host cell according to the present invention may also be an animal cell. Preferably, said animal host cell is a host cell of a fish or a cell line obtained therefrom. More preferably, the fish host cell is from herring, salmon, sardine, redfish, eel, carp, trout, halibut, mackerel, zander or tuna.

[0075] It will be understood that if the host cell of the invention shall be applied for LCPUFA production, it shall be capable of carrying out desaturation and elongation steps on fatty acids. To produce the LCPUFA according to the invention, the C16- or C18-fatty acids must first be desaturated by the enzymatic activity of a desaturase and subsequently be elongated by at least two carbon atoms via an elongase. After one elongation cycle, this enzyme activity gives C18- or C20-fatty acids and after two or three elongation cycles C22-

or C24-fatty acids. The activity of the desaturases and elongases used in the process according to the invention preferably leads to C18-, C20-, C22- and/or C24-fatty acids, advantageously with at least two double bonds in the fatty acid molecule, preferably with three, four or five double bonds, especially preferably to give C20- and/or C22-fatty acids with at least two double bonds in the fatty acid molecule, preferably with three, four or five double bonds in the molecule. After a first desaturation and the elongation have taken place, further desaturation steps such as, for example, one in the delta-5 position may take place. Products of the process according to the invention which are especially preferred are DGLA, ARA, EPA DPA and/or DHA, most preferably EPA and/or DHA. Desaturases and elongases which are required for this process may not always be present naturally in the host cell. Accordingly, the present invention, preferably, envisages a host cell which in addition to the polynucleotide of the present invention comprises polynucleotides encoding such desaturases and/or elongases as required depending on the selected organism. Preferred desaturases and/or elongases which shall be present in the host cell are at least one enzyme selected from the group consisting of: Δ -4-desaturase, Δ -5-desaturase, Δ -5-elongase, Δ -6-desaturase, Δ 12-desaturase, Δ 15-desaturase, ω 3-desaturase and Δ -6-elongase. Especially preferred are the bifunctional d12d15-Desaturases d12d15Des(Ac) from *Acanthamoeba castellanii* (WO2007042510), d12d15Des(Cp) from *Claviceps purpurea* (WO2008006202) and d12d15Des(Lg)1 from *Lottia gigantea* (WO2009016202), the d12-Desaturases d12Des(Co) from *Calendula officinalis* (WO200185968), d12Des(Lb) from *Laccaria bicolor* (WO2009016202), d12Des(Mb) from *Monosiga brevicollis* (WO2009016202), d12Des(Mg) from *Mycosphaerella graminicola* (WO2009016202), d12Des(Nh) from *Nectria haematococca* (WO2009016202), d12Des(Ol) from *Ostreococcus lucimarinus* (WO2008040787), d12Des(Pb) from *Phycomyces blakesleeanus* (WO2009016202), d12Des(Ps) from *Phytophthora sojae* (WO2006100241) and d12Des(Tp) from *Thalassiosira pseudonana* (WO2006069710), the d15-Desaturases d15Des(Hr) from *Helobdella robusta* (WO2009016202), d15Des(Mc) from *Microcoleus chthonoplastes* (WO2009016202), d15Des(Mf) from *Mycosphaerella fijiensis* (WO2009016202), d15Des(Mg) from *Mycosphaerella graminicola* (WO2009016202) and d15Des(Nh)2 from *Nectria haematococca* (WO2009016202), the d4-Desaturases d4Des(Eg) from *Euglena gracilis* (WO2004090123), d4Des(Tc) from *Thraustochytrium* sp. (WO2002026946) and d4Des(Tp) from *Thalassiosira pseudonana* (WO2006069710), the d5-Desaturases d5Des(Ol)2 from *Ostreococcus lucimarinus* (WO2008040787), d5Des(Pp) from *Physcomitrella patens* (WO2004057001), d5Des(Pt) from *Phaeodactylum tricornutum* (WO2002057465), d5Des(Tc) from *Thraustochytrium* sp. (WO2002026946), d5Des(Tp) from *Thalassiosira pseudonana* (WO2006069710) and the d6-Desaturases d6Des(Cp) from *Ceratodon purpureus* (WO2000075341), d6Des(Ol) from *Ostreococcus lucimarinus* (WO2008040787), d6Des(Ot) from *Ostreococcus tauri* (WO2006069710), d6Des(Pf) from *Primula farinosa* (WO2003072784), d6Des(Pir)_BO from *Pythium irregulare* (WO2002026946), d6Des(Pir) from *Pythium irregulare* (WO2002026946), d6Des(Plu) from *Primula luteola* (WO2003072784), d6Des(Pp) from *Physcomitrella patens* (WO200102591), d6Des(Pt) from *Phaeodactylum tricornutum* (WO2002057465), d6Des(Pv) from

Primula vialii (WO2003072784) and d6Des(Tp) from *Thalassiosira pseudonana* (WO2006069710), the d8-Desaturases d8Des(Ac) from *Acanthamoeba castellanii* (EP1790731), d8Des(Eg) from *Euglena gracilis* (WO200034439) and d8Des(Pm) from *Perkinsus marinus* (WO2007093776), the ω 3-Desaturases ω 3Des(Pi) from *Phytophthora infestans* (WO2005083053), ω 3Des(Pir) from *Pythium irregulare* (WO2008022963), ω 3Des(Pir)2 from *Pythium irregulare* (WO2008022963) and ω 3Des(Ps) from *Phytophthora sojae* (WO2006100241), the bifunctional d5d6-elongases d5d6Elo(Om)2 from *Oncorhynchus mykiss* (WO2005012316), d5d6Elo(Ta) from *Thraustochytrium aureum* (WO2005012316) and d5d6Elo(Tc) from *Thraustochytrium* sp. (WO2005012316), the d5-elongases d5Elo(At) from *Arabidopsis thaliana* (WO2005012316), d5Elo(At)2 from *Arabidopsis thaliana* (WO2005012316), d5Elo(Ci) from *Ciona intestinalis* (WO2005012316), d5Elo(Ol) from *Ostreococcus lucimarinus* (WO2008040787), d5Elo(Ot) from *Ostreococcus tauri* (WO2005012316), d5Elo(Tp) from *Thalassiosira pseudonana* (WO2005012316) and d5Elo(XI) from *Xenopus laevis* (WO2005012316), the d6-elongases d6Elo(Ol) from *Ostreococcus lucimarinus* (WO2008040787), d6Elo(Ot) from *Ostreococcus tauri* (WO2005012316), d6Elo(Pi) from *Phytophthora infestans* (WO2003064638), d6Elo(Pir) from *Pythium irregulare* (WO2009016208), d6Elo(Pp) from *Physcomitrella patens* (WO2001059128), d6Elo(Ps) from *Phytophthora sojae* (WO2006100241), d6Elo(Ps)2 from *Phytophthora sojae* (WO2006100241), d6Elo(Ps)3 from *Phytophthora sojae* (WO2006100241), d6Elo(Pt) from *Phaeodactylum tricornutum* (WO2005012316), d6Elo(Tc) from *Thraustochytrium* sp. (WO2005012316) and d6Elo(Tp) from *Thalassiosira pseudonana* (WO2005012316), the d9-elongases d9Elo(Ig) from *Isochrysis galbana* (WO2002077213), d9Elo(Pm) from *Perkinsus marinus* (WO2007093776) and d9Elo(Ro) from *Rhizopus oryzae* (WO2009016208).

[0076] The present invention also relates to a cell, preferably a host cell as specified above or a cell of a non-human organism specified elsewhere herein, said cell comprising a polynucleotide which is obtained from the polynucleotide of the present invention by a point mutation, a truncation, an inversion, a deletion, an addition, a substitution and homologous recombination. How to carry out such modifications to a polynucleotide is well known to the skilled artisan and has been described elsewhere in this specification in detail.

[0077] The present invention furthermore relates to a method for the manufacture of a polypeptide encoded by a polynucleotide of any the present invention comprising

[0078] a) cultivating the host cell of the invention under conditions which allow for the production of said polypeptide; and

[0079] b) obtaining the polypeptide from the host cell of step a).

[0080] Suitable conditions which allow for expression of the polynucleotide of the invention comprised by the host cell depend on the host cell as well as the expression control sequence used for governing expression of the said polynucleotide. These conditions and how to select them are very well known to those skilled in the art. The expressed polypeptide may be obtained, for example, by all conventional purification techniques including affinity chromatography, size exclusion chromatography, high pressure liquid chromatography (HPLC) and precipitation techniques including antibody precipitation. It is to be understood that the method

may—although preferred—not necessarily yield an essentially pure preparation of the polypeptide. It is to be understood that depending on the host cell which is used for the aforementioned method, the polypeptides produced thereby may become posttranslationally modified or processed otherwise.

[0081] The present invention encompasses a polypeptide encoded by the polynucleotide of the present invention or which is obtainable by the aforementioned method.

[0082] The term “polypeptide” as used herein encompasses essentially purified polypeptides or polypeptide preparations comprising other proteins in addition. Further, the term also relates to the fusion proteins or polypeptide fragments being at least partially encoded by the polynucleotide of the present invention referred to above. Moreover, it includes chemically modified polypeptides. Such modifications may be artificial modifications or naturally occurring modifications such as phosphorylation, glycosylation, myristylation and the like (Review in Mann 2003, Nat. Biotechnol. 21, 255-261, review with focus on plants in Huber 2004, Curr. Opin. Plant Biol. 7, 318-322). Currently, more than 300 posttranslational modifications are known (see full ABFC Delta mass list at <http://www.abrf.org/index.cfm/dm.home>). The polypeptide of the present invention shall exhibit the acyltransferase activities referred to above.

[0083] The present invention furthermore relates to an antibody or a fragment derived thereof as an antigen which specifically recognizes a polypeptide encoded by the nucleic acid sequences of the invention.

[0084] Antibodies against the polypeptides of the invention can be prepared by well known methods using a purified polypeptide according to the invention or a suitable fragment derived therefrom as an antigen. A fragment which is suitable as an antigen may be identified by antigenicity determining algorithms well known in the art. Such fragments may be obtained either from the polypeptide of the invention by proteolytic digestion or may be a synthetic peptide. Preferably, the antibody of the present invention is a monoclonal antibody, a polyclonal antibody, a single chain antibody, a chimerized antibody or a fragment of any of these antibodies, such as Fab, Fv or scFv fragments etc. Also comprised as antibodies by the present invention are bispecific antibodies, synthetic antibodies or chemically modified derivatives of any of the aforementioned antibodies. The antibody of the present invention shall specifically bind (i.e. does significantly not cross react with other polypeptides or peptides) to the polypeptide of the invention. Specific binding can be tested by various well known techniques. Antibodies or fragments thereof can be obtained by using methods which are described, e.g., in Harlow and Lane “Antibodies, A Laboratory Manual”, CSH Press, Cold Spring Harbor, 1988. Monoclonal antibodies can be prepared by the techniques originally described in Köhler 1975, Nature 256, 495, and Galfré 1981, Meth. Enzymol. 73, 3, which comprise the fusion of mouse myeloma cells to spleen cells derived from immunized mammals. The antibodies can be used, for example, for the immunoprecipitation, immunolocalization or purification (e.g., by affinity chromatography) of the polypeptides of the invention as well as for the monitoring of the presence of said variant polypeptides, for example, in recombinant organisms, and for the identification of proteins or compounds interacting with the proteins according to the invention.

[0085] Moreover, the present invention contemplates a non-human transgenic organism comprising the polynucleotide or the vector of the present invention.

[0086] Preferably, the non-human transgenic organism is a microorganism, more preferably the non-human transgenic organism is a fungus and most preferably the non-human transgenic organism is a plant, plant part, or plant seed. Preferred plants to be used for introducing the polynucleotide or the vector of the invention are plants which are capable of synthesizing fatty acids, such as all dicotyledonous or monocotyledonous plants, algae or mosses. It is to be understood that host cells derived from a plant may also be used for producing a plant according to the present invention. Preferred plants are selected from the group of the plant families Adolotheciaceae, Anacardiaceae, Asteraceae, Apiaceae, Betulaceae, Boraginaceae, Brassicaceae, Bromeliaceae, Caricaceae, Cannabaceae, Convolvulaceae, Chenopodiaceae, Crypthecodiniaceae, Cucurbitaceae, Ditrachaceae, Elaeagnaceae, Ericaceae, Euphorbiaceae, Fabaceae, Geraniaceae, Gramineae, Juglandaceae, Lauraceae, Leguminosae, Linaceae, Prasinophyceae or vegetable plants or ornamentals such as *Tagetes*. Examples which may be mentioned are the following plants selected from the group consisting of: Adolotheciaceae such as the genera *Physcomitrella*, such as the genus and species *Physcomitrella patens*, Anacardiaceae such as the genera *Pistacia*, *Mangifera*, *Anacardium*, for example the genus and species *Pistacia vera* [pistachio], *Mangifer indica* [mango] or *Anacardium occidentale* [cashew], Asteraceae, such as the genera *Calendula*, *Carthamus*, *Centaurea*, *Cichorium*, *Cynara*, *Helianthus*, *Lactuca*, *Locusta*, *Tagetes*, *Valeriana*, for example the genus and species *Calendula officinalis* [common marigold], *Carthamus tinctorius* [safflower], *Centaurea cyanus* [cornflower], *Cichorium intybus* [chicory], *Cynara scolymus* [artichoke], *Helianthus annuus* [sunflower], *Lactuca sativa*, *Lactuca crispata*, *Lactuca esculenta*, *Lactuca scariola* L. ssp. *sativa*, *Lactuca scariola* L. var. *integrata*, *Lactuca scariola* L. var. *integrifolia*, *Lactuca sativa* subsp. *romana*, *Locusta communis*, *Valeriana locusta* [salad vegetables], *Tagetes lucida*, *Tagetes erecta* or *Tagetes tenuifolia* [african or french marigold], Apiaceae, such as the genus *Daucus*, for example the genus and species *Daucus carota* [carrot], Betulaceae, such as the genus *Corylus*, for example the genera and species *Corylus avellana* or *Corylus colurna* [hazelnut], Boraginaceae, such as the genus *Borago*, for example the genus and species *Borago officinalis* [borage], Brassicaceae, such as the genera *Brassica*, *Melanosinapis*, *Sinapis*, *Arabidopsis*, for example the genera and species *Brassica napus*, *Brassica rapa* ssp. [oilseed rape], *Sinapis arvensis* *Brassica juncea*, *Brassica juncea* var. *juncea*, *Brassica juncea* var. *crispifolia*, *Brassica juncea* var. *foliosa*, *Brassica nigra*, *Brassica sinapioides*, *Melanosinapis communis* [mustard], *Brassica oleracea* [fodder beet] or *Arabidopsis thaliana*, Bromeliaceae, such as the genera *Anana*, *Bromelia* (pineapple), for example the genera and species *Anana comosus*, *Ananas ananas* or *Bromelia comosa* [pineapple], Caricaceae, such as the genus *Carica*, such as the genus and species *Carica papaya* [pawpaw], Cannabaceae, such as the genus *Cannabis*, such as the genus and species *Cannabis sativa* [hemp], Convolvulaceae, such as the genera *Ipomea*, *Convolvulus*, for example the genera and species *Ipomea batatas*, *Ipomea pandurata*, *Convolvulus batatas*, *Convolvulus tiliaceus*, *Ipomea fastigiata*, *Ipomea tiliacea*, *Ipomea triloba* or *Convolvulus panduratus* [sweet potato, batate], Chenopodiaceae, such as the genus

Beta, such as the genera and species *Beta vulgaris*, *Beta vulgaris* var. *altissima*, *Beta vulgaris* var. *Vulgaris*, *Beta maritima*, *Beta vulgaris* var. *perennis*, *Beta vulgaris* var. *conditiva* or *Beta vulgaris* var. *esculenta* [sugarbeet], Cryptocodiaceae, such as the genus *Cryptocodium*, for example the genus and species *Cryptocodium cohnii*, Cucurbitaceae, such as the genus *Cucurbita*, for example the genera and species *Cucurbita maxima*, *Cucurbita mixta*, *Cucurbita pepo* or *Cucurbita moschata* [pumpkin/squash], Cymbellaceae such as the genera *Amphora*, *Cymbella*, *Okedenia*, *Phaeodactylum*, *Reimeria*, for example the genus and species *Phaeodactylum tricorutum*, Ditrichaceae such as the genera Ditrichaceae, *Astomiopsis*, *Ceratodon*, *Chrysoblastella*, *Ditrichum*, *Distichium*, *Eccecidium*, *Lophidion*, *Philibertiella*, *Pleuridium*, *Saelania*, *Trichodon*, *Skottsbergia*, for example the genera and species *Ceratodon antarcticus*, *Ceratodon columbiae*, *Ceratodon heterophyllus*, *Ceratodon purpureus*, *Ceratodon purpureus*, *Ceratodon purpureus* ssp. *convolutus*, *Ceratodon purpureus* spp. *stenocarpus*, *Ceratodon purpureus* var. *rotundifolius*, *Ceratodon ratodon*, *Ceratodon stenocarpus*, *Chrysoblastella chilensis*, *Ditrichum ambiguum*, *Ditrichum brevisetum*, *Ditrichum crispatisimum*, *Ditrichum difficile*, *Ditrichum falcifolium*, *Ditrichum flexicaule*, *Ditrichum giganteum*, *Ditrichum heteromallum*, *Ditrichum lineare*, *Ditrichum lineare*, *Ditrichum montanum*, *Ditrichum montanum*, *Ditrichum pallidum*, *Ditrichum punctulatum*, *Ditrichum pusillum*, *Ditrichum pusillum* var. *tortile*, *Ditrichum rhynchostegium*, *Ditrichum schimperii*, *Ditrichum tortile*, *Distichium capillaceum*, *Distichium hagenii*, *Distichium inclinatum*, *Distichium macounii*, *Eccecidium floridanum*, *Eccecidium whiteleggei*, *Lophidion strictus*, *Pleuridium acuminatum*, *Pleuridium alternifolium*, *Pleuridium holdridgei*, *Pleuridium mexicanum*, *Pleuridium ravenelii*, *Pleuridium subulatum*, *Saelania glaucescens*, *Trichodon borealis*, *Trichodon cylindricus* or *Trichodon cylindricus* var. *oblongus*, Elaeagnaceae such as the genus *Elaeagnus*, for example the genus and species *Olea europaea* [olive], Ericaceae such as the genus *Kalmia*, for example the genera and species *Kalmia latifolia*, *Kalmia angustifolia*, *Kalmia microphylla*, *Kalmia polifolia*, *Kalmia occidentalis*, *Cistus chamaerhodendros* or *Kalmia lucida* [mountain laurel], Euphorbiaceae such as the genera *Manihot*, *Janipha*, *Jatropha*, *Ricinus*, for example the genera and species *Manihot utilisima*, *Janipha manihot*, *Jatropha manihot*, *Manihot aipil*, *Manihot dulcis*, *Manihot manihot*, *Manihot melanobasis*, *Manihot esculenta* [manihot] or *Ricinus communis* [castor-oil plant], Fabaceae such as the genera *Pisum*, *Albizia*, *Cathormion*, *Feuillea*, *Inga*, *Pithecolobium*, *Acacia*, *Mimosa*, *Medicago*, *Glycine*, *Dolichos*, *Phaseolus*, *Soja*, for example the genera and species *Pisum sativum*, *Pisum arvense*, *Pisum humile* [pea], *Albizia berteriana*, *Albizia julibrissin*, *Albizia lebbek*, *Acacia berteriana*, *Acacia littoralis*, *Albizia berteriana*, *Albizia berteriana*, *Cathormion berteriana*, *Feuillea berteriana*, *Inga fragrans*, *Pithecolobium berterianum*, *Pithecolobium fragrans*, *Pithecolobium berterianum*, *Pseudalbizia berteriana*, *Acacia julibrissin*, *Acacia nemu*, *Albizia nemu*, *Feuillea julibrissin*, *Mimosa julibrissin*, *Mimosa speciosa*, *Sericanrda julibrissin*, *Acacia lebbek*, *Acacia macrophylla*, *Albizia lebbek*, *Feuillea lebbek*, *Mimosa lebbek*, *Mimosa speciosa* [silk tree], *Medicago sativa*, *Medicago falcata*, *Medicago varia* [alfalfa], *Glycine max Dolichos soja*, *Glycine gracilis*, *Glycine hispida*, *Phaseolus max*, *Soja hispida* or *Soja max* [soybean], Funariaceae such as the genera *Aphanorrhagma*, *Entosthodon*, *Funaria*, *Physcomitrella*,

Physcomitrium, for example the genera and species *Aphanorrhagma serratum*, *Entosthodon attenuatus*, *Entosthodon bolanderi*, *Entosthodon bonplandii*, *Entosthodon californicus*, *Entosthodon drummondii*, *Entosthodon jamesonii*, *Entosthodon leibergii*, *Entosthodon neoscoticus*, *Entosthodon rubrisetus*, *Entosthodon spathulifolius*, *Entosthodon tucsoni*, *Funaria americana*, *Funaria bolanderi*, *Funaria calcarea*, *Funaria californica*, *Funaria calvescens*, *Funaria convoluta*, *Funaria flavicans*, *Funaria groutiana*, *Funaria hygrometrica*, *Funaria hygrometrica* var. *arctica*, *Funaria hygrometrica* var. *calvescens*, *Funaria hygrometrica* var. *convoluta*, *Funaria hygrometrica* var. *muralis*, *Funaria hygrometrica* var. *utahensis*, *Funaria microstoma*, *Funaria microstoma* var. *obtusifolia*, *Funaria muhlenbergii*, *Funaria orcuttii*, *Funaria plano-convexa*, *Funaria polaris*, *Funaria ravenelii*, *Funaria rubriseta*, *Funaria serrata*, *Funaria sonora*, *Funaria sublimbatus*, *Funaria tucsoni*, *Physcomitrella californica*, *Physcomitrella patens*, *Physcomitrella readeri*, *Physcomitrium australe*, *Physcomitrium californicum*, *Physcomitrium collenchymatum*, *Physcomitrium coloradense*, *Physcomitrium cupuliferum*, *Physcomitrium drummondii*, *Physcomitrium eurystomum*, *Physcomitrium flexifolium*, *Physcomitrium hookeri*, *Physcomitrium hookeri* var. *serratum*, *Physcomitrium immersum*, *Physcomitrium kellermanii*, *Physcomitrium megalocarpum*, *Physcomitrium pyriforme*, *Physcomitrium pyriforme* var. *serratum*, *Physcomitrium rufipes*, *Physcomitrium sandbergii*, *Physcomitrium subsphaericum*, *Physcomitrium washingtoniense*, Geraniaceae, such as the genera *Pelargonium*, *Cocos*, *Oleum*, for example the genera and species *Cocos nucifera*, *Pelargonium grossularioides* or *Oleum cocois* [coconut], Gramineae, such as the genus *Saccharum*, for example the genus and species *Saccharum officinarum*, Juglandaceae, such as the genera *Juglans*, *Wallia*, for example the genera and species *Juglans regia*, *Juglans ailanthifolia*, *Juglans sieboldiana*, *Juglans cinerea*, *Wallia cinerea*, *Juglans bixbyi*, *Juglans californica*, *Juglans hindii*, *Juglans intermedia*, *Juglans jamaicensis*, *Juglans major*, *Juglans microcarpa*, *Juglans nigra* or *Wallia nigra* [walnut], Lauraceae, such as the genera *Persea*, *Laurus*, for example the genera and species *Laurus nobilis* [bay], *Persea americana*, *Persea gratissima* or *Persea persea* [avocado], Leguminosae, such as the genus *Arachis*, for example the genus and species *Arachis hypogaea* [peanut], Linaceae, such as the genera *Linum*, *Adenolinum*, for example the genera and species *Linum usitatissimum*, *Linum humile*, *Linum austriacum*, *Linum bienne*, *Linum angustifolium*, *Linum catharticum*, *Linum flavum*, *Linum grandiflorum*, *Adenolinum grandiflorum*, *Linum lewisii*, *Linum narbonense*, *Linum perenne*, *Linum perenne* var. *lewisii*, *Linum pratense* or *Linum trigynum* [linseed], Lythraeae, such as the genus *Punica*, for example the genera and species *Punica granatum* [pomegranate], Malvaceae, such as the genus *Gossypium*, for example the genera and species *Gossypium hirsutum*, *Gossypium arboreum*, *Gossypium barbadense*, *Gossypium herbaceum* or *Gossypium thurberi* [cotton], Marchantiaceae, such as the genus *Marchantia*, for example the genera and species *Marchantia berteriana*, *Marchantia foliacea*, *Marchantia macropora*, Musaceae, such as the genus *Musa*, for example the genera and species *Musa nana*, *Musa acuminata*, *Musa paradisiaca*, *Musa* spp. [banana], Onagraceae, such as the genera *Camissonia*, *Oenothera*, for example the genera and species *Oenothera biennis* or *Camissonia brevipes* [evening primrose], Palmae, such as the genus *Elaeis*, for example the genus and species *Elaeis guineensis* [oil palm], Papaver-

aceae, such as the genus *Papaver*, for example the genera and species *Papaver orientale*, *Papaver rhoeas*, *Papaver dubium* [poppy], Pedaliaceae, such as the genus *Sesamum*, for example the genus and species *Sesamum indicum* [sesame], Piperaceae, such as the genera *Piper*, *Artanthe*, *Peperomia*, *Steffensia*, for example the genera and species *Piper aduncum*, *Piper amalago*, *Piper angustifolium*, *Piper auritum*, *Piper betel*, *Piper cubeba*, *Piper longum*, *Piper nigrum*, *Piper retrofractum*, *Artanthe adunca*, *Artanthe elongata*, *Peperomia elongata*, *Piper elongatum*, *Steffensia elongata* [cayenne pepper], Poaceae, such as the genera *Hordeum*, *Secale*, *Avena*, *Sorghum*, *Andropogon*, *Holcus*, *Panicum*, *Oryza*, *Zea* (maize), *Triticum*, for example the genera and species *Hordeum vulgare*, *Hordeum jubatum*, *Hordeum murinum*, *Hordeum secalinum*, *Hordeum distichon*, *Hordeum aegiceras*, *Hordeum hexastichon*, *Hordeum hexastichum*, *Hordeum irregulare*, *Hordeum sativum*, *Hordeum secalinum* [barley], *Secale cereale* [rye], *Avena sativa*, *Avena fatua*, *Avena byzantina*, *Avena fatua* var. *sativa*, *Avena hybrida* [oats], *Sorghum bicolor*, *Sorghum halepense*, *Sorghum saccharatum*, *Sorghum vulgare*, *Andropogon drummondii*, *Holcus bicolor*, *Holcus sorghum*, *Sorghum aethiopicum*, *Sorghum arundinaceum*, *Sorghum caffrorum*, *Sorghum cernuum*, *Sorghum dochna*, *Sorghum drummondii*, *Sorghum durra*, *Sorghum guineense*, *Sorghum lanceolatum*, *Sorghum nervosum*, *Sorghum saccharatum*, *Sorghum subglabrescens*, *Sorghum verticilliflorum*, *Sorghum vulgare*, *Holcus halepensis*, *Sorghum miliaceum*, *Panicum militaceum* [millet], *Oryza sativa*, *Oryza latifolia* [rice], *Zea mays* [maize], *Triticum aestivum*, *Triticum durum*, *Triticum turgidum*, *Triticum hybernium*, *Triticum macha*, *Triticum sativum* or *Triticum vulgare* [wheat], Porphyridiaceae, such as the genera *Chrootheca*, *Flintiella*, *Petrovanella*, *Porphyridium*, *Rhodella*, *Rhodorus*, *Vanhoeffenia*, for example the genus and species *Porphyridium cruentum*, Proteaceae, such as the genus *Macadamia*, for example the genus and species *Macadamia integrifolia* [macadamia], Prasinophyceae such as the genera *Nephroselmis*, *Prasinococcus*, *Scherffelia*, *Tetraselmis*, *Mantoniella*, *Ostreococcus*, for example the genera and species *Nephroselmis olivacea*, *Prasinococcus capsulatus*, *Scherffelia dubia*, *Tetraselmis chui*, *Tetraselmis suecica*, *Mantoniella squamata*, *Ostreococcus tauri*, Rubiaceae such as the genus *Cofea*, for example the genera and species *Cofea* spp., *Cofea arabica*, *Cofea canephora* or *Cofea liberica* [coffee], Scrophulariaceae such as the genus *Verbascum*, for example the genera and species *Verbascum blattaria*, *Verbascum chaxii*, *Verbascum densiflorum*, *Verbascum lagurus*, *Verbascum longifolium*, *Verbascum lychnitis*, *Verbascum nigrum*, *Verbascum olympicum*, *Verbascum phlomooides*, *Verbascum phoenicum*, *Verbascum pulverulentum* or *Verbascum thapsus* [mullein], Solanaceae such as the genera *Capsicum*, *Nicotiana*, *Solanum*, *Lycopersicon*, for example the genera and species *Capsicum annum*, *Capsicum annum* var. *glabriusculum*, *Capsicum frutescens* [pepper], *Capsicum annum* [paprika], *Nicotiana tabacum*, *Nicotiana alata*, *Nicotiana attenuata*, *Nicotiana glauca*, *Nicotiana langsdorffii*, *Nicotiana obtusifolia*, *Nicotiana quadrivalvis*, *Nicotiana repanda*, *Nicotiana rustica*, *Nicotiana sylvestris* [tobacco], *Solanum tuberosum* [potato], *Solanum melongena* [eggplant], *Lycopersicon esculentum*, *Lycopersicon lycopersicum*, *Lycopersicon pyriforme*, *Solanum integrifolium* or *Solanum lycopersicum* [tomato], Sterculiaceae, such as the genus *Theobroma*, for example the genus and species *Theobroma cacao* [cacao] or Theaceae, such as the genus *Camellia*, for example the

genus and species *Camellia sinensis* [tea]. In particular preferred plants to be used as transgenic plants in accordance with the present invention are oil fruit crops which comprise large amounts of lipid compounds, such as peanut, oilseed rape, canola, sunflower, safflower, poppy, mustard, hemp, castor-oil plant, olive, sesame, *Calendula*, *Punica*, evening primrose, mullein, thistle, wild roses, hazelnut, almond, *macadamia*, avocado, bay, pumpkin/squash, linseed, soybean, pistachios, borage, trees (oil palm, coconut, walnut) or crops such as maize, wheat, rye, oats, triticale, rice, barley, cotton, cassava, pepper, *Tagetes*, Solanaceae plants such as potato, tobacco, eggplant and tomato, *Vicia* species, pea, alfalfa or bushy plants (coffee, cacao, tea), *Salix* species, and perennial grasses and fodder crops. Preferred plants according to the invention are oil crop plants such as peanut, oilseed rape, canola, sunflower, safflower, poppy, mustard, hemp, castor-oil plant, olive, *Calendula*, *Punica*, evening primrose, pumpkin/squash, linseed, soybean, borage, trees (oil palm, coconut). Especially preferred plants are plants such as sunflower, safflower, tobacco, mullein, sesame, cotton, pumpkin/squash, poppy, evening primrose, walnut, linseed, hemp, thistle or safflower. Very especially preferred plants are plants such as safflower, sunflower, poppy, evening primrose, walnut, linseed, or hemp.

[0087] Preferred mosses are *Physcomitrella* or *Ceratodon*. Preferred algae are *Isochrysis*, *Mantoniella*, *Ostreococcus* or *Cryptocodinium*, and algae/diatoms such as *Phaeodactylum* or *Thraustochytrium*. More preferably, said algae or mosses are selected from the group consisting of: *Shewanella*, *Physcomitrella*, *Thraustochytrium*, *Nannochloropsis*, *Fusarium*, *Phytophthora*, *Ceratodon*, *Isochrysis*, *Aleurita*, *Muscarioides*, *Mortierella*, *Phaeodactylum*, *Cryptocodinium*, specifically from the genera and species *Thalassiosira pseudonona*, *Euglena gracilis*, *Physcomitrella patens*, *Phytophthora infestans*, *Fusarium gramineum*, *Cryptocodinium cohnii*, *Ceratodon purpureus*, *Isochrysis galbana*, *Aleurita farinosa*, *Thraustochytrium* sp., *Nannochloropsis oculata*, *Muscarioides viallii*, *Mortierella alpina*, *Phaeodactylum tricornerutum* or *Caenorhabditis elegans* or especially advantageously *Phytophthora infestans* and *Cryptocodinium cohnii*.

[0088] Transgenic plants may be obtained by transformation techniques as elsewhere in this specification. Preferably, transgenic plants can be obtained by T-DNA-mediated transformation. Such vector systems are, as a rule, characterized in that they contain at least the vir genes, which are required for the *Agrobacterium*-mediated transformation, and the sequences which delimit the T-DNA (T-DNA border). Suitable vectors are described elsewhere in the specification in detail.

[0089] Also encompassed are transgenic non-human animals comprising the vector or polynucleotide of the present invention. Preferred non-human transgenic animals envisaged by the present invention are fish, such as herring, salmon, sardine, redfish, eel, carp, trout, halibut, mackerel, zander or tuna.

[0090] It will be understood that in order to produce the LCPUFA according to the invention, the C16- or C18-fatty acids must first be desaturated by the enzymatic activity of a desaturase and subsequently be elongated by at least two carbon atoms via an elongase in the non-human transgenic organism. After one elongation cycle, this enzyme activity gives C18- or C20-fatty acids and after two or three elongation cycles C22- or C24-fatty acids. The activity of the desaturases and elongases used in the process according to the

invention preferably leads to C18-, C20-, C22- and/or C24-fatty acids, advantageously with at least two double bonds in the fatty acid molecule, preferably with three, four or five double bonds, especially preferably to give C20- and/or C22-fatty acids with at least two double bonds in the fatty acid molecule, preferably with three, four or five double bonds in the molecule. After a first desaturation and the elongation have taken place, further desaturation steps such as, for example, one in the delta-5 position may take place. Products of the process according to the invention which are especially preferred are DGLA, ARA, EPA DPA and/or DHA, most preferably EPA and/or DHA. Desaturases and elongases which are required for this process may not always be present naturally in the organism. Accordingly, the present invention, preferably, envisages a transgenic non-human organism which in addition to the polynucleotide of the present invention comprises polynucleotides encoding such desaturases and/or elongases as required depending on the selected organism. Preferred desaturases and/or elongases which shall be present in the organism are at least one enzyme selected from the group consisting of: Δ -4-desaturase, Δ -5-desaturase, Δ -5-elongase, Δ -6-desaturase, Δ 12-desaturase, Δ 15-desaturase, ω 3-desaturase and Δ -6-elongase. Especially preferred are the bifunctional d12d15-Desaturases d12d15Des(Ac) from *Acanthamoeba castellanii* (WO2007042510), d12d15Des(Cp) from *Claviceps purpurea* (WO2008006202) and d12d15Des(Lg)1 from *Lottia gigantea* (WO2009016202), the d12-Desaturases d12Des(Co) from *Calendula officinalis* (WO200185968), d12Des(Lb) from *Laccaria bicolor* (WO2009016202), d12Des(Mb) from *Monosiga brevicollis* (WO2009016202), d12Des(Mg) from *Mycosphaerella graminicola* (WO2009016202), d12Des(Nh) from *Nectria haematococca* (WO2009016202), d12Des(Ol) from *Ostreococcus lucimarinus* (WO2008040787), d12Des(Pb) from *Phycomyces blakesleeanus* (WO2009016202), d12Des(Ps) from *Phytophthora sojae* (WO2006100241) and d12Des(Tp) from *Thalassiosira pseudonana* (WO2006069710), the d15-Desaturases d15Des(Hr) from *Helobdella robusta* (WO2009016202), d15Des(Mc) from *Microcoleus chthonoplastes* (WO2009016202), d15Des(Mf) from *Mycosphaerella fijiensis* (WO2009016202), d15Des(Mg) from *Mycosphaerella graminicola* (WO2009016202) and d15Des(Nh)2 from *Nectria haematococca* (WO2009016202), the d4-Desaturases d4Des(Eg) from *Euglena gracilis* (WO2004090123), d4Des(Tc) from *Thraustochytrium* sp. (WO2002026946) and d4Des(Tp) from *Thalassiosira pseudonana* (WO2006069710), the d5-Desaturases d5Des(Ol)2 from *Ostreococcus lucimarinus* (WO2008040787), d5Des(Pp) from *Physcomitrella patens* (WO2004057001), d5Des(Pt) from *Phaeodactylum tricornerutum* (WO2002057465), d5Des(Tc) from *Thraustochytrium* sp. (WO2002026946), d5Des(Tp) from *Thalassiosira pseudonana* (WO2006069710) and the d6-Desaturases d6Des(Cp) from *Ceratodon purpureus* (WO2000075341), d6Des(Ol) from *Ostreococcus lucimarinus* (WO2008040787), d6Des(Ot) from *Ostreococcus tauri* (WO2006069710), d6Des(Pf) from *Primula farinosa* (WO2003072784), d6Des(Pir)_{BO} from *Pythium irregulare* (WO2002026946), d6Des(Pir) from *Pythium irregulare* (WO2002026946), d6Des(Plu) from *Primula luteola* (WO2003072784), d6Des(Pp) from *Physcomitrella patens* (WO200102591), d6Des(Pt) from *Phaeodactylum tricornerutum* (WO2002057465), d6Des(Pv) from *Primula vialii* (WO2003072784) and d6Des(Tp) from *Thalassiosira pseudonana* (WO2006069710), the d8-De-

saturases d8Des(Ac) from *Acanthamoeba castellanii* (EP1790731), d8Des(Eg) from *Euglena gracilis* (WO200034439) and d8Des(Pm) from *Perkinsus marinus* (WO2007093776), the ω 3-Desaturases ω 3Des(Pi) from *Phytophthora infestans* (WO2005083053), ω 3Des(Pir) from *Pythium irregulare* (WO2008022963), ω 3Des(Pir)2 from *Pythium irregulare* (WO2008022963) and ω 3Des(Ps) from *Phytophthora sojae* (WO2006100241), the bifunctional d5d6-elongases d5d6Elo(Om)2 from *Oncorhynchus mykiss* (WO2005012316), d5d6Elo(Ta) from *Thraustochytrium aureum* (WO2005012316) and d5d6Elo(Tc) from *Thraustochytrium* sp. (WO2005012316), the d5-elongases d5Elo(At) from *Arabidopsis thaliana* (WO2005012316), d5Elo(At)2 from *Arabidopsis thaliana* (WO2005012316), d5Elo(Ci) from *Ciona intestinalis* (WO2005012316), d5Elo(Ol) from *Ostreococcus lucimarinus* (WO2008040787), d5Elo(Ot) from *Ostreococcus tauri* (WO2005012316), d5Elo(Tp) from *Thalassiosira pseudonana* (WO2005012316) and d5Elo(XI) from *Xenopus laevis* (WO2005012316), the d6-elongases d6Elo(Ol) from *Ostreococcus lucimarinus* (WO2008040787), d6Elo(Ot) from *Ostreococcus tauri* (WO2005012316), d6Elo(Pi) from *Phytophthora infestans* (WO2003064638), d6Elo(Pir) from *Pythium irregulare* (WO2009016208), d6Elo(Pp) from *Physcomitrella patens* (WO2001059128), d6Elo(Ps) from *Phytophthora sojae* (WO2006100241), d6Elo(Ps)2 from *Phytophthora sojae* (WO2006100241), d6Elo(Ps)3 from *Phytophthora sojae* (WO2006100241), d6Elo(Pt) from *Phaeodactylum tricornerutum* (WO2005012316), d6Elo(Tc) from *Thraustochytrium* sp. (WO2005012316) and d6Elo(Tp) from *Thalassiosira pseudonana* (WO2005012316), the d9-elongases d9Elo(Ig) from *Isochrysis galbana* (WO2002077213), d9Elo(Pm) from *Perkinsus marinus* (WO2007093776) and d9Elo(Ro) from *Rhizopus oryzae* (WO2009016208).

[0091] Furthermore, the present invention encompasses a method for the manufacture of polyunsaturated fatty acids comprising:

[0092] a) cultivating the host cell of the invention under conditions which allow for the production of polyunsaturated fatty acids in said host cell; and

[0093] b) obtaining said polyunsaturated fatty acids from the said host cell.

[0094] The term "polyunsaturated fatty acids (PUFA)" as used herein refers to fatty acids comprising at least two, preferably, three, four, five or six, double bonds. Moreover, it is to be understood that such fatty acids comprise, preferably from 18 to 24 carbon atoms in the fatty acid chain. More preferably, the term polyunsaturated fatty acids relates to long chain PUFA (LCPUFA) having from 20 to 24 carbon atoms in the fatty acid chain. Preferred unsaturated fatty acids in the sense of the present invention are selected from the group consisting of arachidonic acid (ARA) 20:4 (5,8,11,14), eicosapentaenoic acid (EPA) 20:5 (5,8,11,14,17), and docosahexaenoic acid (DHA) 22:6 (4,7,10,13,16,19) and, more preferably, from EPA and DHA. Thus, it will be understood that most preferably, the methods provided by the present invention relating to the manufacture of EPA or DHA. Moreover, also encompassed are the intermediates of LCPUFA which occur during synthesis starting from oleic acid 18:1 (9), preferably, linoleic acid 18:2 (9,12), alpha-linolenic acid 18:3 (9,12,15), gamma-linolenic acid 18:3 (6,9,12), stearidonic acid 18:4 (6,9,12,15), dihomogamma-linolenic acid 20:3 (8,11,14), eicosadienoic acid 20:2 (11,14),

eicosatrienoic acid 20:3 (11,14,17), eicosatetraenoic acid 20:4 (8,11,14,17) and docosapentaenoic acid (DPA) 22:5 (4,7,10,13,16).

[0095] The term “cultivating” as used herein refers maintaining and growing the host cells under culture conditions which allow the cells to produce the said polyunsaturated fatty acid, i.e. the PUFA and/or LCPUFA referred to above, preferably, as triglyceride esters. This implies that the polynucleotide of the present invention is expressed in the host cell so that the acyltransferase activity is present. Suitable culture conditions for cultivating the host cell are described in more detail below.

[0096] The term “obtaining” as used herein encompasses the provision of the cell culture including the host cells and the culture medium as well as the provision of purified or partially purified preparations thereof comprising the polyunsaturated fatty acids, preferably, as triglyceride esters. More preferably, the PUFA and LCPUFA are to be obtained as triglyceride esters, e.g., in form of an oil. More details on purification techniques can be found elsewhere herein below.

[0097] The host cells to be used in the method of the invention are grown or cultured in the manner with which the skilled artisan is familiar, depending on the host organism. Usually, host cells are grown in a liquid medium comprising a carbon source, usually in the form of sugars, a nitrogen source, usually in the form of organic nitrogen sources such as yeast extract or salts such as ammonium sulfate, trace elements such as salts of iron, manganese and magnesium and, if appropriate, vitamins, at temperatures of between 0° C. and 100° C., preferably between 10° C. and 60° C. under oxygen or anaerobic atmosphere dependent on the type of organism. The pH of the liquid medium can either be kept constant, that is to say regulated during the culturing period, or not. The cultures can be grown batchwise, semibatchwise or continuously. Nutrients can be provided at the beginning of the fermentation or administered semicontinuously or continuously: The produced PUFA or LCPUFA can be isolated from the host cells as described above by processes known to the skilled artisan, e.g., by extraction, distillation, crystallization, if appropriate precipitation with salt, and/or chromatography. It might be required to disrupt the host cells prior to purification. To this end, the host cells can be disrupted beforehand. The culture medium to be used must suitably meet the requirements of the host cells in question. Descriptions of culture media for various microorganisms which can be used as host cells according to the present invention can be found in the textbook “Manual of Methods for General Bacteriology” of the American Society for Bacteriology (Washington D.C., USA, 1981). Culture media can also be obtained from various commercial suppliers. All media components are sterilized, either by heat or by filter sterilization. All media components may be present at the start of the cultivation or added continuously or batchwise, as desired. If the polynucleotide or vector of the invention which has been introduced in the host cell further comprises an expressible selection marker, such as an antibiotic resistance gene, it might be necessary to add a selection agent to the culture, such as an antibiotic in order to maintain the stability of the introduced polynucleotide. The culture is continued until formation of the desired product is at a maximum. This is normally achieved within 10 to 160 hours. The fermentation broths can be used directly or can be processed further. The biomass may, according to requirement, be removed completely or partially from the fermentation broth by separation methods

such as, for example, centrifugation, filtration, decanting or a combination of these methods or be left completely in said broth. The fatty acid preparations obtained by the method of the invention, e.g., oils, comprising the desired PUFA or LCPUFA as triglyceride esters are also suitable as starting material for the chemical synthesis of further products of interest. For example, they can be used in combination with one another or alone for the preparation of pharmaceutical or cosmetic compositions, foodstuffs, or animal feeds. Chemically pure triglycerides comprising the desired PUFA or LCPUFA can also be manufactured by the methods described above. To this end, the fatty acid preparations are further purified by extraction, distillation, crystallization, chromatography or combinations of these methods. In order to release the fatty acid moieties from the triglycerides, hydrolysis may be also required. The said chemically pure triglycerides or free fatty acids are, in particular, suitable for applications in the food industry or for cosmetic and pharmacological compositions.

[0098] Moreover, the present invention relates to a method for the manufacture of polyunsaturated fatty acids comprising:

[0099] a) cultivating the non-human transgenic organism of the invention under conditions which allow for the production of polyunsaturated fatty acids in said host cell; and

[0100] b) obtaining said polyunsaturated fatty acids from the said non-human transgenic organism.

[0101] Further, it follows from the above that a method for the manufacture of an oil, lipid or fatty acid composition is also envisaged by the present invention comprising the steps of any one of the aforementioned methods and the further step of formulating PUFA or LCPUFA as oil, lipid or fatty acid composition. Preferably, said oil, lipid or fatty acid composition is to be used for feed, foodstuffs, cosmetics or pharmaceuticals. Accordingly, the formulation of the PUFA or LCPUFA shall be carried out according to the GMP standards for the individual envisaged products. For example, oil may be obtained from plant seeds by an oil mill. However, for product safety reasons, sterilization may be required under the applicable GMP standard. Similar standards will apply for lipid or fatty acid compositions to be applied in cosmetic or pharmaceutical compositions. All these measures for formulating oil, lipid or fatty acid compositions as products are comprised by the aforementioned manufacture.

[0102] The present invention also relates to oil comprising a polyunsaturated fatty acid or a polyunsaturated fatty acid composition obtainable by the aforementioned methods.

[0103] The term “oil” refers to a fatty acid mixture comprising unsaturated and/or saturated fatty acids which are esterified to triglycerides. Preferably, the triglycerides in the oil of the invention comprise PUFA or LCPUFA as referred to above. The amount of esterified PUFA and/or LCPUFA is, preferably, approximately 30%, a content of 50% is more preferred, a content of 60%, 70%, 80% or more is even more preferred. The oil may further comprise free fatty acids, preferably, the PUFA and LCPUFA referred to above. For the analysis, the fatty acid content can be, e.g., determined by GC analysis after converting the fatty acids into the methyl esters by transesterification. The content of the various fatty acids in the oil or fat can vary, in particular depending on the source. The oil, however, shall have a non-naturally occurring composition with respect to the PUFA and/or LCPUFA composition and content. It will be understood that such a unique oil composition and the unique esterification pattern of PUFA

and LCPUFA in the triglycerides of the oil shall only be obtainable by applying the methods of the present invention specified above. Moreover, the oil of the invention may comprise other molecular species as well. Specifically, it may comprise minor impurities of the polynucleotide or vector of the invention. Such impurities, however, can be detected only by highly sensitive techniques such as PCR.

[0104] The contents of all references cited throughout this application are herewith incorporated by reference in general and with respect to their specific disclosure content referred to above.

[0105] This invention is further illustrated by the following figures and examples which should not be construed as limiting the scope of the invention.

FIGURES

[0106] FIG. 1: Cloning strategy employed for stepwise buildup of plant expression plasmids of the invention.

[0107] FIG. 2: Vector map of the *bbc* construct used for *Arabidopsis* transformation.

[0108] FIG. 3: GC chromatogram of fatty acids methyl esters of total fatty acids of Col-0, *fae1* mutant and *fae1* transformed with *bbc*. Total fatty acids were measured as described by Wu et al., 2005. The content of the different fatty is indicated in table 5.

[0109] FIG. 4: Total ion count of 26 acyl CoA ESI-MS/MS MRM pairs for *Arabidopsis* (A) Col-0 and (B) *fae1* harbouring EPA biosynthesis pathway. Maturing *Arabidopsis* seeds were harvested 18 days after flowering. Acyl-CoA was extracted according to Larson et al (2001) and LC conditions after Han et al. (2010).

[0110] FIG. 5: Identification of Eicosapentaenoic and Arachidonic-CoA's in the acyl CoA pool of *Arabidopsis* Col-0 and EPA producing plants. MRM chromatograms of co-eluting acyl-CoA of interest in (A) wild type and (C) *fae1* harbouring EPA biosynthetic pathway with recorded reactions shown for each transition, isotopic peaks (IP) of homologous long chain acyl CoA are shown. (B) Characteristic fragmentation of the protonated acyl-CoA by neutral loss of 507 to give the protonated acyl pantetheine group.

[0111] FIG. 6: LPCAT activity assay.

[0112] A yeast mutant lacking LPEAT and LPCAT activity (due to knockout of the gene YOR175c) was transformed with the empty vector pYES2.1 (lane marked “-”) and with pYES2.1 harboring the cDNA of pLPAAT_c6316(No) (lane 1 and 2, SEQ-ID: 13). Microsomal isolations of these transformants and the wildtype yeast strain BY4742 (lane marked “+”) containing 5 µg protein where incubated with alpha-linolenic acid-CoA and [¹⁴C]-18:1-lysophosphatidylcholine (LPC). Thin layer chromatography was performed to separate lipid classes. Like for wildtype yeast (lane marked “+”), phosphatidylcholine (PC) is observed for both yeast clones shown in lane 1 and 2, indicating the gene pLPAAT_c6316(No) has LPCAT activity and complements the missing LPCAT activity of the knockout strain.

[0113] FIG. 7: LPAAT activity assay.

[0114] A yeast mutant lacking LPAAT activity (due to knockout of the gene YDL052c) was transformed with the empty vector pYES2.1 (lane marked “-”) and with pYES2.1 harboring the cDNA of pLPAAT_c6316(No) (lane 1 and 2, SEQ-ID: 13). Microsomal isolations of these transformants and the wildtype yeast strain BY4742 (lane marked “+”) containing 5 µg protein where incubated with alpha-linolenic acid-CoA and [¹⁴C]-18:1-lysophosphatidic acid (LPA). Thin

layer chromatography was performed to separate lipid classes. Like for wildtype yeast (lane marked “+”), phosphatidic acid (PA) is observed for both yeast clones shown in lane 1 and 2, indicating the gene pLPAAT_c6316(No) has LPAAT activity and complements the missing LPAAT activity of the knockout strain.

[0115] FIG. 8: DGAT activity assay.

[0116] A yeast mutant lacking the capability to synthesis TAG (due to knockout of the four genes YCR048W, YNR019W, YOR245C and YNR008W) was transformed with the empty vector pYES2.1 (lane marked “-”) and with pYES2.1 harboring the cDNA of pDGAT2-c19425mod(Ta) (SEQ-ID 52, lane 1 and 2), pDGAT2_c4648(No) (SEQ-ID 34, lane 5 and 6), pDGAT2_c48271(No) (SEQ-ID 102, lane 7 and 8), BnDGAT1 (SEQ-ID 107, lane 9 and 10), AtDGAT1 (SEQ-ID 105, lane 11 and 12), pDGAT2_c699(No) (SEQ-ID 19, lane 13 and 14) and pDGAT2_c2959(No) (SEQ-ID 25, lane 15). Microsomal isolations of these transformants and the wildtype yeast strain G175 (lane marked “+”) where incubated with ¹⁴C-labeled oleic acid and diacylglycerole (DAG). Thin layer chromatography was performed to separate lipid classes. Like for wildtype yeast (lane marked “+”), triacylglycerole (TAG) is observed in lane 1, 2, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14 and 15, indicating pDGAT2-c19425mod(Ta), pDGAT2_c4648(No), pDGAT2_c48271(No), BnDGAT1, AtDGAT1, pDGAT2_c699(No) and pDGAT2_c2959(No) encode polypeptides having DGAT activity and complement the missing TAG-synthesis capability of the knockout.

[0117] FIG. 9: Substrate specificity of AtDGAT1 and BnDGAT1. The specific activity of the enzymes AtDGAT1 and BnDGAT1 using the substrates indicated at the x-axis is given as the amount (in nmol) of substrate consumed in one minute per mg total protein and was determined as described in example 10.

[0118] FIG. 10: Substrate specificity of pDGAT2-c19425(Ta) compared to AtDGAT1 and BnDGAT1. The specific activity of the enzymes pDGAT2-c19425(Ta), AtDGAT1 and BnDGAT1 using the substrates indicated at the x-axis is given as the amount (in nmol) of substrate consumed in one minute per mg total protein and was determined as described in example 10.

[0119] FIG. 11: Substrate specificity of pDGAT2_c699(No) and pDGAT2_c4648(No) compared to AtDGAT1 and BnDGAT1. The specific activity of the enzymes pDGAT2_c699(No) and pDGAT2_c4648(No), AtDGAT1 and BnDGAT1 using the substrates indicated at the x-axis is given as the amount (in nmol) of substrate consumed in one minute per mg total protein and was determined as described in example 10.

EXAMPLES

Example 1

General Cloning Methods

[0120] Cloning methods as e.g. use of restriction endonucleases to cut double stranded DNA at specific sites, agarose gel electrophoreses, purification of DNA fragments, transfer of nucleic acids onto nitrocellulose and nylon membranes, joining of DNA-fragments, transformation of *E. coli* cells and culture of bacteria where performed as described in Sambrook et al. (1989) (Cold Spring Harbor Laboratory Press: ISBN 0-87965-309-6).

Example 2

Sequence Analysis of Recombinant DNA

[0121] Sequencing of recombinant DNA-molecules was performed using a laser-fluorescence DNA sequencer (Applied Biosystems Inc, USA) employing the sanger method (Sanger et al. (1977) Proc. Natl. Acad. Sci. USA 74, 5463-5467). Expression constructs harboring fragments obtained by polymerase chain reactions were subjected to sequencing to confirm the correctness of expression cassettes consisting of promoter, nucleic acid molecule to be expressed and terminator to avoid mutations that might result from handling of the DNA during cloning, e.g. due to incorrect primers, mutations from exposure to UV-light or errors of polymerases.

Example 3

Cloning of Yeast Expression Construct Via Homologous Recombination

[0122] The open reading frame listed in SEQ ID NOs: 52, 1, 4, 7, 10, 13, 16, 19, 22, 25, 28, 31, 34, 37, 40, 43, 46, 49, 55, 102, 105 and 107 encoding polypeptides with the amino acid sequence SEQ ID NOs: 53, 2, 5, 8, 11, 14, 17, 20, 23, 26, 29, 32, 35, 38, 41, 44, 47, 50, 56, 103, 106 and 108 that have acyltransferase activity can be amplified using the primer listed in table 2 in a polymerase chain reaction. By doing so, the open reading frame is 5' fused to about 60 nucleotides of the 3' end of the GAL1 promoter sequence with simultaneous introduction of and Asc I and/or Nco I restriction site between the fusion site and 3' fused to about 60 nucleotides of the 5' end of the CYC1 terminator sequence with simul-

aneous introduction of and Pac I restriction site. To integrate these fragments into pYES2.1 TOPO downstream of the galactose inducible GAL1 Promoter via homologous recombination, the vector pYES2.1 (Invitrogen) can be digested using the restriction endonucleases Pvu II and Xba I, and *Saccharomyces cerevisiae* can be transformed with 5 to 20 ng of linearized pYES2.1 TOPO vector and 20 to 100 ng PCR product per 50 µl competent cells using the transformation method described by Schiestl et al. (Schiestl et al. (1989) Curr. Genet. 16(5-6), pp. 339-346), to obtain pYES-pLPLAT_c1216(No), pYES-pLPLAT_c3052(No), pYES-pLPEAT-c7109(Ta), pYES-pLPAAT_c2283(No), pYES-pLPAAT_c6316(No), pYES-pDGAT2_irc24907(No), pYES-pDGAT2_c699(No), pYES-pDGAT2_c1910(No), pYES-pDGAT2_c2959(No), pYES-pDGAT2_c4857(No), pYES-pDGAT1_c21701(No), pYES-pDGAT2_c4648(No), pYES-pDGAT2_c1660(No), pYES-pDGAT2_c29432(No), pYES-pDGAT2_c1052(No), pYES-pDGAT2-c18182(Ta), pYES-pDGAT2-c5568(Ta), pYES-pDGAT2-c19425(Ta), pYES-pDGAT2_c48271(No), AtDGAT1, BnDGAT1 and pYES-pGPAT_c813(No) in various wildtype yeasts and yeast mutants. Positive transformants can be selected based on the complementation of the URA auxotrophy of the chosen *S. cerevisia* strain. To validate the correctness of the expression construct harbored by a particular yeast clone, plasmids can be isolated as described in Current Protocols in Molecular Biology (Hoffmann, Curr. Protoc. Mol. Biol. 2001 May; Chapter 13:Unit13.11), transformed into *E. coli* for amplification and subjected to sequencing of the expression cassette as described in example 2. For later cloning into plant expression plasmids, the introduced restrictions site for Asc I and/or Nco I in combination with Pac I can be used.

TABLE 2

Primer sequences for cloning acyltransferase-polynucleotides of the invention for yeast expression		
Gene-Name	Primer	SEQ-ID
pLPLAT_c1216 (No)	Forward: ataaaagtatcaacaaaaattgttaatatacctctatactttaacgt caaggagaaaaaaccccgatcgcgcgccaccatggacaa ggcactggcaccgtt	46
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaactaaactttcttccttccc tcta	47
pLPLAT_c3052 (No)	Forward: ataaaagtatcaacaaaaattgttaatatacctctatactttaacgt caaggagaaaaaaccccgatcgcgcgccaccatgaccacg actgtcatctctag	48
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaactaaagcctcccgcac aacgagc	49
pLPEAT-c7109 (Ta)	Forward: ataaaagtatcaacaaaaattgttaatatacctctatactttaacgt caaggagaaaaaaccccgatcgcgcgccaccatggaggg catcgagtcgatagt	50
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaactataaggcttctccc gcgagg	51
pLPAAT_c2283 (No)	Forward: ataaaagtatcaacaaaaattgttaatatacctctatactttaacgt caaggagaaaaaaccccgatcgcgcgccaccatgaagac gccacgagcctggc	52

TABLE 2-continued

Primer sequences for cloning acyltransferase-polynucleotides of the invention for yeast expression		
Gene-Name	Primer	SEQ-ID
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaac ccttccttttcggttagagcggatttaattaataagctctcgaatcgct cttct	53
pLPAAAT_c6316 (No)	Forward: ataaaagtatcaacaaaaattggttaataacacctctatactttaacgt caaggagaaaaaaccccgatcggcgcgccaccatggtcagg aggaagatggacgt	54
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaac ccttccttttcggttagagcggatttaattaacacgacgcggcgc cttgacgt	55
pDGAT2_lrc24907 (No)	Forward: ataaaagtatcaacaaaaattggttaataacacctctatactttaacgt caaggagaaaaaaccccgatcggcgcgccaccatggcacc tccccaccggcccc	56
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaac ccttccttttcggttagagcggatttaattaacatttgaccactaaggct ggcct	57
pDGAT2_c699 (No)	Forward: ataaaagtatcaacaaaaattggttaataacacctctatactttaacgt caaggagaaaaaaccccgatcggcgcgccaccatgggtctat ttggcagcgggat	58
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaac ccttccttttcggttagagcggatttaattaactaaaagaaattcaac gtccgat	59
pDGAT2_c1910 (No)	Forward: ataaaagtatcaacaaaaattggttaataacacctctatactttaacgt caaggagaaaaaaccccgatcggcgcgccaccatggtgagta tccccagtcgctc	60
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaac ccttccttttcggttagagcggatttaattaactaaaagaaattccagc tcctgt	61
pDGAT2_c2959 (No)	Forward: ataaaagtatcaacaaaaattggttaataacacctctatactttaacgt caaggagaaaaaaccccgatcggcgcgccaccatgacgccg caagccgatcac	62
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaac ccttccttttcggttagagcggatttaattaactcaatggacaacg ggcgcg	63
pDGAT2_c4857 (No)	Forward: ataaaagtatcaacaaaaattggttaataacacctctatactttaacgt caaggagaaaaaaccccgatcggcgcgccaccatggttacc tcttcgctcgtcg	64
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaac ccttccttttcggttagagcggatttaattaataggcgcgcgaatg aactcct	65
pDGAT1_c21701 (No)	Forward: ataaaagtatcaacaaaaattggttaataacacctctatactttaacgt caaggagaaaaaaccccgatcggcgcgccaccatgccttttg gacggctgcatc	66
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaac ccttccttttcggttagagcggatttaattaacacccgaaaatcct ccttct	67
pDGAT2_c4648 (No)	Forward: ataaaagtatcaacaaaaattggttaataacacctctatactttaacgt caaggagaaaaaaccccgatcggcgcgccaccatggccaa ggctaacttccccgc	68

TABLE 2-continued

Primer sequences for cloning acyltransferase-polynucleotides of the invention for yeast expression		
Gene-Name	Primer	SEQ- ID
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaatcacctttataagcagctt cttgc	69
pDGAT2_c1660 (No)	Forward: ataaaagtatcaacaaaaattggttaatatacctctatactttaacgt caaggagaaaaaaccgcatcgggcgccaccatggttgctgc agggattaagctg	70
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaatcacaaacaggaccaat ttatgat	71
pDGAT2_c29432 (No)	Forward: ataaaagtatcaacaaaaattggttaatatacctctatactttaacgt caaggagaaaaaaccgcatcgggcgccaccatggttgatgg cgccgtcggcg	72
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaatcagaagtggaagc gtcttgc	73
pDGAT2_c1052 (No)	Forward: ataaaagtatcaacaaaaattggttaatatacctctatactttaacgt caaggagaaaaaaccgcatcgggcgccaccatgggctgc accactgaccca	74
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaatcacgacttcggacagt ccaaaa	75
pDGAT2-c18182 (Ta)	Forward: ataaaagtatcaacaaaaattggttaatatacctctatactttaacgt caaggagaaaaaaccgcatcgggcgccaccatggttgctgc ttgagcacagcgc	76
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaactacacaaatcgcatc gtcttgc	77
pDGAT2-c5568 (Ta)	Forward: ataaaagtatcaacaaaaattggttaatatacctctatactttaacgt caaggagaaaaaaccgcatcgggcgccaccatggttctcct ctgccttccta	78
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaactacgagtcagccac ttgatgc	79
pDGAT2-c19425 (Ta)	Forward: ataaaagtatcaacaaaaattggttaatatacctctatactttaacgt caaggagaaaaaaccgcatcgggcgccaccatggttcttctc catcgaacggga	80
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaactaacctcggtgtaca gcgccg	81
pGPAT_c813 (No)	Forward: ataaaagtatcaacaaaaattggttaatatacctctatactttaacgt caaggagaaaaaaccgcatcgggcgccaccatgcatcc cgcagcaccattga	82
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaatcagacaagctcctctt cccct	83
pDGAT2_c48271 (No)	Forward: ataaaagtatcaacaaaaattggttaatatacctctatactttaacgt caaggagaaaaaaccgcatcgggcgccaccatggcgcc atctcaccgcgcaa	109

TABLE 2-continued

Primer sequences for cloning acyltransferase-polynucleotides of the invention for yeast expression		
Gene-Name	Primer	SEQ-ID
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaactaccacactccaact tcgccc	110
AtDGAT1	Forward: ataaaagtatcaacaaaaattgtaataatatacctctatactttaacgt caaggagaaaaaacccggatcggcgcgccaccatggcgattt tggattctgctgg	111
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaatcatgacatcgatcctttt cgggt	112
BnDGAT1	Forward: ataaaagtatcaacaaaaattgtaataatatacctctatactttaacgt caaggagaaaaaacccggatcggcgcgccaccatggagattt tggattctggagg	113
	Reverse: aactataaaaaataaataggacctagacttcaggttgctaaact ccttccttttcggttagagcggatttaattaatgacatcctttcctttg cgggt	114

TABLE 3

Coding polynucleotide sequences, amino acid sequences encoded thereby and expressed sequences (mRNA) of the acyltransferases of the invention							
Gene name	Organism	ORF in bp	SEQ-ID No.	Amino acids	SEQ-ID No.	mRNA in bp	SEQ-ID No.
pLPLAT_c1216(No)	<i>Nannochloropsis oculata</i>	1485	1	494	2	1908	3
pLPLAT_c3052(No)	<i>Nannochloropsis oculata</i>	1776	4	591	5	2247	6
pLPEAT-c7109(Ta)	<i>Thraustochytrium aureum</i>	1134	7	377	8	1288	9
pLPAAT_c2283(No)	<i>Nannochloropsis oculata</i>	1284	10	427	11	1826	12
pLPAAT_c6316(No)	<i>Nannochloropsis oculata</i>	1395	13	464	14	1771	15
pDGAT2_Irc24907(No)	<i>Nannochloropsis oculata</i>	1026	16	341	17	1100	18
pDGAT2_c699(No)	<i>Nannochloropsis oculata</i>	1206	19	401	20	1772	21
pDGAT2_c1910(No)	<i>Nannochloropsis oculata</i>	1173	22	390	23	1239	24
pDGAT2_c2959(No)	<i>Nannochloropsis oculata</i>	1089	25	362	26	1609	27
pDGAT2_c4857(No)	<i>Nannochloropsis oculata</i>	1464	28	487	29	1682	30
pDGAT1_c21701(No)	<i>Nannochloropsis oculata</i>	1539	31	512	32	1904	33
pDGAT2_c4648(No)	<i>Nannochloropsis oculata</i>	1083	34	360	35	1362	36
pDGAT2_c1660(No)	<i>Nannochloropsis oculata</i>	1695	37	564	38	2074	39
pDGAT2_c29432(No)	<i>Nannochloropsis oculata</i>	1029	40	342	41	1585	42
pDGAT2_c1052(No)	<i>Nannochloropsis oculata</i>	1251	43	416	44	1923	45
pDGAT2-c18182(Ta)	<i>Thraustochytrium aureum</i>	930	46	309	47	1134	48
pDGAT2-c5568(Ta)	<i>Thraustochytrium aureum</i>	1179	49	392	50	1303	51

TABLE 3-continued

Coding polymucleotide sequences, amino acid sequences encoded thereby and expressed sequences (mRNA) of the acyltransferases of the invention							
Gene name	Organism	ORF in bp	SEQ-ID No.	Amino acids	SEQ-ID No.	mRNA in bp	SEQ-ID No.
pDGAT2-c19425(Ta)	<i>Thraustochytrium aureum</i>	1389	52	462	53	1547	54
pGPAT_c813(No)	<i>Nannochloropsis oculata</i>	1977	55	658	56	2460	57
pDGAT2_c48271(No)	<i>Nannochloropsis oculata</i>	960	102	319	103	1265	104

Example 4

Assembly of Genes Required for PUFA Synthesis within a T-Plasmid

[0123] For synthesis of EPA in *Arabidopsis* seeds, the set of genes encoding the proteins of the metabolic EPA pathway (table 4) was combined with expression elements (promoter, terminators) and transferred into binary t-plasmids that were used for agrobacteria mediated transformation of plants as described in example 5. To this end, the general cloning strategy depicted in FIG. 1 was employed: Genes listed in table 4 were PCR-amplified using Phusion™ High-Fidelity DNA Polymerase (NEB, Frankfurt, Germany) according to the manufactures instructions from cDNA using primer introducing a Nco I and/or Asc I restriction site at the 5' terminus, and a Pac I restriction site at the 3' terminus (FIG. 1B). To obtain the final expression modules, PCR-amplified genes were cloned between promoter and terminator via Nco I and/or Pac I restriction sites (FIG. 1C). Up to three of those expression modules were combined as desired to expression cassettes harbored by either one of pENTR/A, pENTR/B or pENTR/C (FIG. 1D). Finally, the Multisite Gateway™ System (Invitrogen) was used to combine three expression cassette harbored by pENTR/A, pENTR/B and pENTR/C (FIG. 1E) to obtain the final binary T-plasmids bbc (SEQ-ID 101, FIG. 2).

TABLE 4

Genes of the bbc construct for synthesis of EPA (20:5n-3) in Arabidopsis seeds. The elements controlling the expression of the respective genes are as well indicated.					
Name	Source Organism	Activity	SEQ-ID	Promoter	Terminator
d12Des(Ps)	<i>Phytophthora sojae</i>	d-12 Desaturase	96	p-BnNapin	t-E9
d6Des(Ot)	<i>Ostreococcus tauri</i>	d-6 Desaturase	97	p-SBP	t-CatpA
d5Des(Te)	<i>Traustochytrium ssp.</i>	d-5 Desaturase	98	p-LuCnl	t-AgroOCS
d6Elo(Pp)	<i>Physcomitrella patens</i>	d-6 Elongase	99	p-VfUSP	t-CaMV35S
o-3Des(Pi)	<i>Phytophthora infestans</i>	o-3 Desaturase	100	p-Napin	t-E9

Example 5

Plant Transformation

[0124] The resulting binary vector bbc harboring the genes reconstituting EPA biosynthesis pathway were transformed into *Agrobacterium tumefaciens* (Hofgen and Willmitzer (1988) Nucl. Acids Res. 16: 9877). The transformation of *A. thaliana* was accomplished by means of the floral-dip method (Clough and Bent (1998) Plant Journal 16: 735-

743), this method is known to the skilled person. Wild-type *Arabidopsis* seeds contain considerable amounts of eicosenoic acid (20:1) (Table 5). Biosynthesis of 20:1 competes for the substrates of the PUFA biosynthesis pathway. This competition was circumvented by transforming bbc into the *Arabidopsis* fae1 mutant (James et al., (1995) The Plant Cell 7:309-319).

Example 6

Quantification of Metabolic Fatty Acyl-CoAs in Wild-Type and EPA Producing *Arabidopsis* Seeds

[0125] The selected transgenic *Arabidopsis* plants from example 3 were analyzed in respect to PUFA content in seeds. Seeds from wild-type, fae1 mutant and transgenics harboring the bbc construct were harvested 18 days after flowering. Total fatty acid, representing the fatty acids esterified to CoA, on lipids and as triacyl-glycerides were extracted and analyzed by gas chromatography as described in Wu et al., (2005) Nature Biotechnology 23(8): 1013-1017.

[0126] In seeds of fae1 transformed with bbc the EPA accumulation was 12.2%, the seeds contained small amounts of intermediate or side products: ARA (3.2%), SDA (0.8%), GLA (2.6%) which were not present in wild-type or fae1 (FIG. 3, Table 5).

TABLE 5

Content of fatty acids in seeds of wild-type (Col-0), fae1 mutant and fae1 transformed with bbc construct				
Fatty acid	Common name of FA	Col-0	fae1	bbc fae1
16:0	Palmitic acid	6.2	8.8	6.8
18:0	Stearic acid	3.1	4.1	5.3
18:1	Oleic acid	16.3	27.5	18.9
18:2	Linoleic acid	28.2	39.0	30.8

TABLE 5-continued

Content of fatty acids in seeds of wild-type (Col-0), fae1 mutant and fae1 transformed with bbc construct				
Fatty acid	Common name of FA	Col-0	fae1	bbc fae1
18:3n6	Gamma-Linolenic acid	0.0	0.0	2.6
18:3n3	Alpha-Linolenic acid	15.6	18.4	11.9
18:4n3	Stearidonic acid	0.0	0.0	0.8
20:1	Eicosenoic acid	22.8	0.4	0.3
20:4n6	Arachidonic acid	0.0	0.0	3.2
20:5n3	Eicosapentaenoic acid	0.0	0.0	12.2
Others		7.8	1.8	7.2

[0127] For PUFA biosynthesis the acyl-moiety has to be shuffled between different metabolic pools. For example, the elongation of the acyl chain by two carbon atoms occurs specifically on acyl-CoA (Zank et al., (2002) *The Plant Journal* 318(3):255-268. The efficiency of the transfer of the acyl-residue between the metabolic pools may represent a bottleneck for PUFA production in plants. Therefore the accumulation of EPA or intermediates of EPA biosynthesis as CoA species was analyzed by LC/MS². As a control CoA pool of wild-type seeds were as well analyzed. The Acyl-CoA metabolites were extracted from the seed tissue according to Larson and Graham, 2001. LC/MS² was applied as described by Magnes et al., 2005. Briefly, CoA were separated with high resolution by reversed-phase high performance liquid chromatography (HPLC) with a ammonium hydroxide and acetonitrile gradient. The acyl-CoA species were identified and quantified by multireaction monitoring using triple quadrupole mass spectrometry. Only a few methods using mass spectrometry for characterization of long chain acyl-CoA have been published, the majority of which employ negative ionisation mode showing abundant ions. In contrast, positive ionisation has only one abundant ion [M-H]⁺, furthermore the predominant ion in MS² spectra is the fatty acyl-pantetheine fragment (m/z 507—FIG. 5 B), characteristic of CoA-activated substances. In choosing the acyl-pantetheine of interest in multireaction monitoring mode (MRM) a very sensitive, selective and reproducible method was established. CoA-activated substances can be monitored by scanning for the neutral loss of phosphoadenosine diphosphate. Generally for reliable analysis, all interfering peaks must be chromatographically separated; in the case of EPA and ARA this is not possible (FIG. 4 B). However through the use of MRM, incorporating very short dwell times (15 ms), it is possible to follow the individual chromatograms of acyl-CoA of interest and demonstrate the presence of EPA and ARA in the acyl CoA pool (FIG. 5 C). According to internal standards the eicosapentaenoyl-CoA was in the range of . . . % of the total Co-A pool.

[0128] In conclusion these results show that PUFA accumulate in the metabolic CoA pool and are not transferred to DAG to be released as TAG into the seed oil. Such a bottleneck may be overcome by the co-expression of an acyltransferase from table 3, having the appropriate substrate specificity. The application of suitable acyltransferase may increase the flux of fatty acid between the metabolic pools and increase the PUFA biosynthesis rate.

Example 7

Activity Assays Using Yeast Extracts

[0129] To characterize the functions of the acyltransferase polypeptides of the invention, yeast mutants can be

employed that are defective in certain acyltransferase activities. For example, the yeast mutant Y13749 (Genotype: BY4742; Mat alpha; his3Δ1; leu2Δ0; lys2Δ0; ura3Δ0; YDL052c::kanMX4) lacking LPAAT activity can be transformed with expression constructs harboring candidate polypeptides to check for restoration (complementation) of LPAAT activity, the yeast mutant Y12431 (genotype BY4742; Mat alpha; his3Δ1; leu2Δ0; lys2Δ0; ura3Δ0; YOR175c::kanMX4) lacking LPLAT activity can be transformed with expression constructs harboring candidate polypeptides to check for restoration (complementation) of LPLAT activity, the yeast mutant H1246 (genotype MATa leu2-3,112 trp1-1 can1-100 ura3-1 ade2-1 his3-11,15 YOR245::KanMX4 YNR008W::TRP1 YCR048W::HIS3 YNR019W::LEU2) lacking the ability to synthesize triacylglycerole can be transformed with expression constructs harboring candidate polypeptides to check for restoration (complementation) of the ability to synthesize triacylglycerole. The yeast mutants can for example harbor the expression constructs listed in example 3 employing the transformation method described in example 3.

[0130] For LPAAT activity assay, clones of the yeast mutant Y13749 harboring pYES-pLPAAT_c6316(No) can be grown at 28° C. in 10 ml selective media (SC-URA) with 2% raffinose as carbon source over night. The next day, expression of the acyltransferase polypeptide can be induced by transferring the cells to fresh media containing 2% galactose, for example by inoculating 100 ml of fresh culture to an optical density (measure at 600 nm) of OD₆₀₀=0.1. Cells are harvested after 24 h incubation at 28° C. by centrifugation at 3000×g for 5 min and resuspended in 1 ml resuspension buffer (25 mM Tris/HCL pH 7.6) and disrupted using acid washed zirconium bead (200 μm average diameter) in a mill (Retsch, Germany) by three minutes agitation at 300 rpm. The supernatant is transferred to a fresh tube and centrifuged at 3000×g for 5 min. The obtained supernatant is the crude extract. Protein content is measured according to Bradford (Bradford, M. M. (1976), *Anal. Biochem.* Bd. 72, pp. 248-254) with bovine serum albumin as standard.

[0131] Assay mixtures contain 1 to 50 μg of protein, 10 μl of 100 nM [¹⁴C]-18:1-LPA (giving about 2000 dpm/nmol), 10 μl of 50 nM 18:1-CoA or 50 nM 18:3n-3-CoA in assay buffer (25 mM Tris/HCL pH 7.6, 0.5 mg/ml BSA) to give a total volume of 100 μl. Samples are incubated for 10 min at 30° C. The assays are terminated by extraction of the lipids into chloroform according to Blight and Dyer (Blight, E. G. and Dyer, W. J. (1959), *Can. J. Biochem. Physiol.* 37, pp. 911-917). Lipids are separated on thin layer chromatography (TLC) silica 60 plates (Merck) using chloroform/methanol/acetic acid/water (90:15:10:3), and autoradiographic pictures are taken using an instant imager (Packard). It can be seen by the formation of phosphatidic acid (PA) in FIG. 7, that pLPAAT_c6316(No) (SEQ-ID 13, lane 1 and 2) encodes a polypeptide having LPAAT activity.

[0132] For LPCAT and LPEAT activity assay, clones of the yeast mutant Y12431 harboring pYES-pLPAAT_c6316(No) can be grown at 28° C. in 10 ml selective media (SC-URA) with 2% raffinose as carbon source over night. The next day, expression of the acyltransferase polypeptide can be induced by transferring the cells to fresh media containing 2% galactose, for example by inoculating 100 ml of fresh culture to an optical density (measure at 600 nm) of OD₆₀₀=0.1. Cells are harvested after 24 h incubation at 28° C. by centrifugation at 3000×g for 5 min and resuspended in

1 ml resuspension buffer (25 mM Tris/HCL pH 7.6) and disrupted using acid washed zirconium bead (200 μ m average diameter) in a mill (Retsch, Germany) by three minutes agitation at 300 rpm. The supernatant is transferred to a fresh tube and centrifuged at 3000 \times g for 5 min. The obtained supernatant is the crude extract. Protein content is measured according to Bradford (Bradford, M. M. (1976), *Anal. Biochem.* Bd. 72, pp. 248-254) with bovine serum albumin as standard. Assay mixtures contain either 10 μ l 100 nM [14 C]-LPC (LPCAT activity assay) or 10 μ l 100 nM [14 C]-LPE (LPEAT activity assay), 1 to 50 μ g of protein, 10 μ l of 50 nM 18:1-CoA or 50 nM 18:3n-3-CoA in assay buffer (25 mM Tris/HCL pH 7.6, 0.5 mg/ml BSA) to give a total volume of 100 μ l. Samples are incubated for 10 min at 30 $^{\circ}$ C. The assays are terminated by extraction of the lipids into chloroform according to Bligh and Dyer (Bligh, E. G. and Dyer, W. J. (1959), *Can. J. Biochem. Physiol.* 37, pp. 911-917). Lipids are separated on thin layer chromatography (TLC) silica 60 plates (Merck) using chloroform/methanol/acetic acid/water (90:15:10:3), and autoradiographic pictures are taken using an instant imager (Packard). It can be seen by the formation of phosphatidylethanolamine (PC) in FIG. 6, that pLPAAT_c6316(No) (SEQ-ID 13, lane 1 and 2) encodes a polypeptide having LPCAT activity.

[0133] For DGAT activity assay, clones of the yeast mutant H1246 harboring either one of pYES-pDGAT2_c699(No), pYES-pDGAT2_c2959(No), pYES-pDGAT2_c4648(No), pYES-pDGAT2_c48271(No), pYES-pDGAT2_c19425(Ta), pYES-AtDGAT1, or pYES-BnDGAT1 can be grown at 28 $^{\circ}$ C. in 10 ml selective media (SC-URA) with 2% raffinose as carbon source over night. The next day, expression of the acyltransferase polypeptides can be induced by transferring the cells to fresh media containing 2% galactose, for example by inoculating 100 ml of fresh culture to an optical density (measure at 600 nm) of OD₆₀₀=0.1. Activity as indicated by the formation of TAG (as indicated, the mutant H1246 is unable to synthesize TAG) can be measured either by relying on yeast-endogenous substrate-DAG, or by providing substrate in an in vitro assay.

[0134] For the former type of assay, cells are harvested after reaching stationary phase during incubation at 28 $^{\circ}$ C. by centrifugation at 3000 \times g for 5 min and resuspended in 2 ml resuspension buffer (phosphate buffered saline (PBS) pH 7.4, see Sambrook et al., "Molecular Cloning", Cold Spring

Harbor Laboratory, 1989). The equivalent of 200 mg cell pellet is taken, the volume adjusted to 210 μ l using PBS and 790 μ l of methanol:chloroform (2:1) are added. Cells are disrupted using acid washed zirconium bead (200 μ m average diameter) in a mill (Retsch, Germany) by three minutes agitation at 300 rpm and lipids are extracted according to Bligh and Dyer (Bligh, E. G. and Dyer, W. J. (1959), *Can. J. Biochem. Physiol.* 37, pp. 911-917).

[0135] The in vitro assay is the preferred way of testing for DGAT activity, when activity is known or expected to be weak when relying on endogenous substrate. Instead, both the type and concentration of the DAG acceptor molecule, as well as the type and concentration of the fatty acid-CoA can be controlled. To do so, cells are harvested after 24 h incubation at 28 $^{\circ}$ C. by centrifugation at 3000 \times g for 5 min and resuspended in 1 ml resuspension buffer (25 mM Tris/HCL pH 7.6) and disrupted using acid washed zirconium bead (200 μ m average diameter) in a mill (Retsch, Germany) by three minutes agitation at 300 rpm. The supernatant is transferred to a fresh tube and centrifuged at 3000 \times g for 5 min. The obtained supernatant is the crude extract. Protein content is measured according to Bradford (Bradford, M. M. (1976), *Anal. Biochem.* Bd. 72, pp. 248-254) with bovine serum albumin as standard. Assay mixtures contain 10 μ l 50 nM [14 C]-6:0-DAG (giving about 3000 dpm/nmol), 50 μ g of microsomal protein (the amount can be adjusted to stay within linear conditions without substrate limitation), 10 μ l of 50 nM 18:3n-3-CoA or 50 nM 22:6n-3-CoA in assay buffer (50 mM Hepes buffer pH 7.2, 1 mg/ml BSA) to give a total volume of 100 μ l. Samples are incubated for 10 min at 30 $^{\circ}$ C.

[0136] In either case—in vivo or in vitro assay—lipids are separated on thin layer chromatography (TLC) silica 60 plates (Merck) using hexane:diethylether:acetic acid (70:30:1), and stained in iodine vapor. It can be seen by the formation of triacylglycerole (TAG) using the in vitro assay-conditions in FIG. 8, that pDGAT2-c19425mod(Ta) (SEQ-ID 52, lane 1 and 2), pDGAT2_c4648(No) (SEQ-ID 34, lane 5 and 6), pDGAT2_c48271(No) (SEQ-ID 102, lane 7 and 8), BnDGAT1 (SEQ-ID 107, lane 9 and 10), AtDGAT1 (SEQ-ID 105, lane 11 and 12), pDGAT2_c699(No) (SEQ-ID 19, lane 13 and 14) and pDGAT2_c2959(No) (SEQ-ID 25, lane 15) encode polypeptides having DGAT activity.

[0137] Table 6 summarizes the results of the LPCAT, LPAAT and DGAT activity tests.

TABLE 6

Measured with microsomal protein and [14 C]-18:1-LPA, [14 C]-18:1-LPC or [14 C]-6:0-1,2-DAG. Of the in vitro DGAT assay, 1 mg/ml of BSA was added to reduce activity for staying in the linear range.

Enzyme Class	Candidate	SEQ-IDs (ORF/- protein/mRNA)	Activity in vitro using 18:3-CoA nmol/(mg*min)	Activity in vitro using 22:6-CoA nmol/(mg*min)	Activity in vivo
LPAAT	pLPAAT_c6316(No)	13/14/15	81	64	
LPCAT	pLPAAT_c6316(No)	13/14/15	38	9	
DGAT	pDGAT2_c699(No)	19/20/21	0.22	0.17	Yes
DGAT	pDGAT2_c2959(No)	25/26/27	0.95	0.67	Yes
DGAT	pDGAT2_c4648(No)	34/35/36	1.4	0.17	Yes
DGAT	pDGAT2_c48271(No)	102/103/104	1.6	0	Yes
DGAT	pDGAT2-c19425(Ta)	52/53/54	4.0	5.6	Yes
DGAT	AtDGAT1	105/106/—	1.6	1.2	Yes
DGAT	BnDGAT1	107/108/—	2.4	1.5	Yes

Example 8

Determination of Substrate Specificity for LPAAT

[0138] For determination of substrate specificities of the LPAAT enzymes, clones of the yeast mutant Y13749 (described in example 7) harboring LPAAT genes in the pYES plasmid can be grown at 28° C. in 10 ml selective media (SC-URA) with 2% raffinose as carbon source over night. The next day, expression of the acyltransferase polypeptides can be induced by transferring the cells to fresh media containing 2% galactose, for example by inoculating 100 ml of fresh culture to an optical density (measure at 600 nm) of $OD_{600}=0.1$. Cells are harvested after 24 h incubation at 28° C. by centrifugation at 3000×g for 5 min and resuspended in 1 ml disruption buffer (20 mM Tris/HCL pH 7.6, 10 mM MgCl₂, 1 mM EDTA, 5% glycerol, 0.3 M (NH₄)₂SO₄) and disrupted using acid washed zirconium beads (200 μm average diameter) in a mill (Retsch, Germany) by three minutes agitation at 300 rpm. The supernatant and the beads are transferred to a fresh tube. Disruption buffer is added up to 20 ml and the tube is centrifuged at 8000×g for 5 min. The obtained supernatant is centrifuged for 2 hrs at 42000 rpm at 4° C. The pellet (microsomal fraction) is resuspended in a small volume of 0.1 M phosphate buffer pH 7.2. Protein content in the microsomal fraction is measured according to Bradford (Bradford, M. M. (1976), Anal. Biochem. Bd. 72, pp. 248-254) with bovine serum albumin as standard. Assay mixtures contain 1-5 μg of microsomal protein (the amount is adjusted to achieve linear conditions without substrate limitation), 10 μl of 1 mM [¹⁴C]-18:1-LPA (5000 dpm/nmol), 10 μl of 1 mM acyl-CoA in assay buffer (0.1 M phosphate buffer pH 7.2, 10 mg/ml Bovine Serum Albumine (BSA)) to give a total volume of 100 μl. Like to amount of microsomal protein added to the assay, also the amount of BSA has influence on observed enzyme activities, where higher amounts of BSA result on lower activities and lower amounts of BSA result in higher activities. The enzyme specificity can be tested for different acyl-CoA:s, e.g. 14:0-CoA, 16:0-CoA, 18:1-CoA, 18:2-CoA, 18:3-CoA, 18:3-CoA, 18:3-CoA, 18:4-CoA, 20:3-CoA, 20:4-CoA, 20:4-CoA, 20:4(n-3)-CoA, 20:5-CoA, 22:5-CoA, 22:6-CoA. Samples are incubated for 4 min at 30° C. The assays are terminated by extraction of the lipids into chloroform according to Bligh and Dyer (Bligh, E. G. and Dyer, W. J. (1959), Can. J. Biochem. Physiol. 37, pp. 911-917). Lipids are separated on thin layer chromatography (TLC) silica 60 plates (Merck) using chloroform/methanol/acetic acid/water (90:15:10:3), and autoradiographic pictures are taken using an instant imager (Packard). The amount of phosphatidic acid (PA) produced in the reaction (and hence the enzyme activity) can be determined from the picture.

Example 9

Determination of Substrate Specificity for LPLAT

[0139] For LPCAT and LPEAT activity assay, clones of the yeast mutant Y12431 harboring LPLAT genes in the pYES plasmid can be grown at 28° C. in 10 ml selective media (SC-URA) with 2% raffinose as carbon source over night. The next day, expression of the acyltransferase polypeptides can be induced by transferring the cells to fresh media containing 2% galactose, for example by inoculating 100 ml of fresh culture to an optical density (measure at 600

nm) of $OD_{600}=0.1$. Cells are harvested after 24 h incubation at 28° C. by centrifugation at 3000×g for 5 min and resuspended in 1 ml disruption buffer (20 mM Tris/HCL pH 7.6, 10 mM MgCl₂, 1 mM EDTA, 5% glycerol, 0.3 M (NH₄)₂SO₄) and disrupted using acid washed zirconium beads (200 μm average diameter) in a mill (Retsch, Germany) by three minutes agitation at 300 rpm. The supernatant and the beads are transferred to a fresh tube. Disruption buffer is added up to 20 ml and the tube is centrifuged at 8000×g for 5 min. The obtained supernatant is centrifuged for 2 hrs at 42000 rpm at 4° C. The pellet (microsomal fraction) is resuspended in a small volume of 0.1 M phosphate buffer pH 7.2. Protein content in the microsomal fraction is measured according to Bradford (Bradford, M. M. (1976), Anal. Biochem. Bd. 72, pp. 248-254) with bovine serum albumin as standard. Assay mixtures contain either 10 μl 1 mM [¹⁴C]-18:1-Lysophosphatidylcholine (-LPC), 5000 dpm/nmol (LPCAT assay) or 10 μl 1 mM [¹⁴C]-18:1-Lysophosphatidylethanolamine (-LPE), 5000 dpm/nmol (LPEAT assay), 1-10 μg of microsomal protein (the amount is adjusted to achieve linear conditions without substrate limitation), 10 μl of 1 mM acyl-CoA in assay buffer (0.1 M phosphate buffer pH 7.2., 10 mg/ml BSA) to give a total volume of 100 μl. Like to amount of microsomal protein added to the assay, also the amount of BSA has influence on observed enzyme activities, where higher amounts of BSA result on lower activities and lower amounts of BSA result in higher activities. The enzyme specificity can be tested for different acyl-CoA:s, e.g. 14:0-CoA, 16:0-CoA, 18:1-CoA, 18:2-CoA, 18:3-CoA, 18:3-CoA, 18:3-CoA, 18:4-CoA, 20:3-CoA, 20:4-CoA, 20:4(n-3)-CoA, 20:5-CoA, 22:5-CoA, 22:6-CoA. Samples are incubated for 4 min at 30° C. The assays are terminated by extraction of the lipids into chloroform according to Bligh and Dyer (Bligh, E. G. and Dyer, W. J. (1959), Can. J. Biochem. Physiol. 37, pp. 911-917). Lipids are separated on thin layer chromatography (TLC) silica 60 plates (Merck) using chloroform/methanol/acetic acid/water (90:15:10:3), and autoradiographic pictures are taken using an instant imager (Packard). The amount of phosphatidyl choline (PC) or phosphatidyl ethanol amine (PE) produced in the reaction (and hence the enzyme activity) can be determined from the picture.

Example 10

Determination of Substrate Specificity for DGAT

[0140] For DGAT activity assay, clones of the yeast mutant H1246 harboring either one of pYES-pDGAT2_c699 (No), pYES-pDGAT2_c2959(No), pYES-pDGAT2_c4648 (No), pYES-pDGAT2_c48271(No), pYES-pDGAT2-c19425(Ta), pYES-AtDGAT1, or pYES-BnDGAT1 can be grown at 28° C. in 10 ml selective media (SC-URA) with 2% raffinose as carbon source over night. The next day, expression of the acyltransferase polypeptides can be induced by transferring the cells to fresh media containing 2% galactose, for example by inoculating 100 ml of fresh culture to an optical density (measure at 600 nm) of $OD_{600}=0.1$. Cells are harvested after 24 h incubation at 28° C. by centrifugation at 3000×g for 5 min and resuspended in 1 ml disruption buffer (20 mM Tris/HCL pH 7.6, 10 mM MgCl₂, 1 mM EDTA, 5% glycerol, 0.3 M (NH₄)₂SO₄) and disrupted using acid washed zirconium beads (200 μm average diameter) in a mill (Retsch, Germany) by three minutes agitation at 300 rpm. The supernatant and the beads

are transferred to a fresh tube. Disruption buffer is added up to 20 ml and the tube is centrifuged at 8000×g for 5 min. The obtained supernatant is centrifuged for 2 hrs at 42000 rpm at 4° C. The pellet (microsomal fraction) is resuspended in a small volume of 0.1 M phosphate buffer pH 7.2. Protein content in the microsomal fraction is measured according to Bradford (Bradford, M. M. (1976), *Anal. Biochem.* Bd. 72, pp. 248-254) with bovine serum albumin as standard. Assay mixtures contain 5 µl 1 mM [¹⁴C]-6:0-DAG, 3000 dpm/nmol, 1-100 µg of microsomal protein (the amount is adjusted to achieve linear conditions without substrate limitation), 5 µl of 1 mM acyl-CoA in assay buffer (50 mM Hepes buffer pH 7.2, 1 mg/ml BSA) to give a total volume of 100 µl. The enzyme specificity can be tested for different acyl-CoA:s, e.g. 14:0-CoA, 16:0-CoA, 18:1-CoA, 18:2-CoA, 18:3-CoA, γ18:3-CoA, 18:4-CoA, 20:3-CoA, 20:4-CoA, 20:4(n-3)-CoA, 20:5-CoA, 22:5-CoA, 22:6-CoA. Samples are incubated for 4 min at 30° C. The assays are terminated by extraction of the lipids into chloroform according to Bligh and Dyer (Bligh, E. G. and Dyer, W. J. (1959), *Can. J. Biochem. Physiol.* 37, pp. 911-917). Lipids are separated on thin layer chromatography (TLC) silica 60 plates (Merck) using hexane:diethylether:acetic acid (70:30:1), and autoradiographic pictures are taken using an instant imager (Packard). The amount of triacylglycerol (TAG) produced in the reaction (and hence the enzyme activity) can be determined from the picture. In *Brassica napus* and *Arabidopsis*, the DGAT involved in TAG-formation in seeds are of the DGAT1 type. The enzyme activity AtDGAT1 and BnDGAT1 for the different substrates can be seen in FIG. 9. The enzyme activity of pDGAT2-c19425(Ta) for the different substrates, compared to AtDGAT1 and BnDGAT1 is shown in FIG. 10. The enzyme activity of pDGAT2_c699 (No) and pDGAT2_c4648(No) for the different substrates, compared to AtDGAT1 and BnDGAT1 is shown in FIG. 11. The data in FIGS. 10 and 11 show clearly, that all DGAT2 enzymes shown in these figures vary strongly towards their activities for the various substrates, whereas the DGAT1 involved in TAG formation in *Arabidopsis* and *Brassica napus* exhibit less variability towards these different substrates.

Example 11

Determination of Substrate Selectivity for LPAAT

[0141] For determination of substrate selectivities of the LPAAT enzymes, clones of the yeast mutant Y13749 (described in example 7) harboring LPAAT genes can be grown at 28° C. in 10 ml selective media (SC-URA) with 2% raffinose as carbon source over night. The next day, expression of the acyltransferase polypeptides can be induced by transferring the cells to fresh media containing 2% galactose, for example by inoculating 100 ml of fresh culture to an optical density (measure at 600 nm) of OD₆₀₀=0.1. Cells are harvested after 24 h incubation at 28° C. by centrifugation at 3000×g for 5 min and resuspended in 1 ml disruption buffer (20 mM Tris/HCL pH 7.6, 10 mM MgCl₂, 1 mM EDTA, 5% glycerol, 0.3 M (NH₄)₂SO₄) and disrupted using acid washed zirconium beads (200 µm average diameter) in a mill (Retsch, Germany) by three minutes agitation at 300 rpm. The supernatant and the beads are transferred to a fresh tube. Disruption buffer is added up to 20 ml and the tube is centrifuged at 8000×g for 5 min. The obtained supernatant is centrifuged for 2 hrs at 42000 rpm at 4° C. The pellet

(microsomal fraction) is resuspended in a small volume of 0.1 M phosphate buffer pH 7.2. Protein content in the microsomal fraction is measured according to Bradford (Bradford, M. M. (1976), *Anal. Biochem.* Bd. 72, pp. 248-254) with bovine serum albumin as standard. The substrate selectivity can be determined by mixing equimolar amounts of different acyl-CoA:s in the same reaction and measure the preference for using the different acyl groups as substrates. The assay is run as in the specificity studies (Example 5) but scaled up 18 times to get sufficient amount of PA for detection. Up to 4 different acyl-CoA:s can be used in the assay in equimolar amount instead of one single acyl-CoA. Samples are incubated for 4 min at 30° C. The assays are terminated by extraction of the lipids into chloroform according to Bligh and Dyer (Bligh, E. G. and Dyer, W. J. (1959), *Can. J. Biochem. Physiol.* 37, pp. 911-917). Lipids are separated on thin layer chromatography (TLC) silica 60 plates (Merck) using chloroform/methanol/acetic acid/water (90:15:10:3), and autoradiographic pictures are taken using an instant imager (Packard). The phosphatidic acid (PA) is recovered from the plate and the fatty acids methylated in situ on the gel with sulphuric acid (2%) in methanol. Fatty acid methyl esters are extracted with hexane and separated by gas-liquid chromatography (GLC) using a WCOT fused silica 50 m×0.32 mm ID capillary column coated with CP-Wax 58-CB DF=0.3 (Chrompack inc., The Netherlands) and quantified relative to methyl-heptadecanoate added as an internal standard. The selectivity can be determined by calculating the amount of each acyl group that has been acylated to LPA.

Example 12

Determination of Substrate Selectivity for LPLAT

[0142] For LPCAT and LPEAT activity assay, clones of the yeast mutant Y12431 harboring LPLAT genes can be grown at 28° C. in 10 ml selective media (SC-URA) with 2% raffinose as carbon source over night. The next day, expression of the acyltransferase polypeptides can be induced by transferring the cells to fresh media containing 2% galactose, for example by inoculating 100 ml of fresh culture to an optical density (measure at 600 nm) of OD₆₀₀=0.1. Cells are harvested after 24 h incubation at 28° C. by centrifugation at 3000×g for 5 min and resuspended in 1 ml disruption buffer (20 mM Tris/HCL pH 7.6, 10 mM MgCl₂, 1 mM EDTA, 5% glycerol, 0.3 M (NH₄)₂SO₄) and disrupted using acid washed zirconium beads (200 µm average diameter) in a mill (Retsch, Germany) by three minutes agitation at 300 rpm. The supernatant and the beads are transferred to a fresh tube. Disruption buffer is added up to 20 ml and the tube is centrifuged at 8000×g for 5 min. The obtained supernatant is centrifuged for 2 hrs at 42000 rpm at 4° C. The pellet (microsomal fraction) is resuspended in a small volume of 0.1 M phosphate buffer pH 7.2. Protein content in the microsomal fraction is measured according to Bradford (Bradford, M. M. (1976), *Anal. Biochem.* Bd. 72, pp. 248-254) with bovine serum albumin as standard. The substrate selectivity can be determined by mixing equimolar amounts of different acyl-CoA:s in the same reaction and measure the preference for using the different acyl groups as substrates. The assay is run as in the specificity studies (Example 6) but scaled up 18 times to get sufficient amount of PC or PE for detection. Up to 4 different acyl-CoA:s can be used in the assay in equimolar amount instead of one

single acyl-CoA. Samples are incubated for 4 min at 30° C. The assays are terminated by extraction of the lipids into chloroform according to Bligh and Dyer (Bligh, E. G. and Dyer, W. J. (1959), *Can. J. Biochem. Physiol.* 37, pp. 911-917). Lipids are separated on thin layer chromatography (TLC) silica 60 plates (Merck) using chloroform/methanol/acetic acid/water (90:15:10:3), and autoradiographic pictures are taken using an instant imager (Packard). The PC or PE is recovered from the plate and the fatty acids methylated in situ on the gel with sulphuric acid (2%) in methanol. Fatty acid methyl esters are extracted with hexane and separated by gas-liquid chromatography (GLC) using a WCOT fused silica 50 m×0.32 mm ID capillary column coated with CP-Wax 58-CB DF=0.3 (Chrompack inc., The Netherlands) and quantified relative to methyl-heptadecanoate added as an internal standard. The selectivity can be determined by calculating the amount of each acyl group that has been acylated to LPC or LPE.

Example 13

Determination of Substrate Selectivity for DGAT

[0143] For DGAT activity assay, clones of the yeast mutant H1246 harboring DGAT genes can be grown at 28° C. in 10 ml selective media (SC-URA) with 2% raffinose as carbon source over night. The next day, expression of the acyltransferase polypeptides can be induced by transferring the cells to fresh media containing 2% galactose, for example by inoculating 100 ml of fresh culture to an optical density (measure at 600 nm) of OD₆₀₀=0.1. Cells are harvested after 24 h incubation at 28° C. by centrifugation at 3000×g for 5 min and resuspended in 1 ml disruption buffer (20 mM Tris/HCL pH 7.6, 10 mM MgCl₂, 1 mM EDTA, 5% glycerol, 0.3 M (NH₄)₂SO₄) and disrupted using acid washed zirconium beads (200 μm average diameter) in a

mill (Retsch, Germany) by three minutes agitation at 300 rpm. The supernatant and the beads are transferred to a fresh tube. Disruption buffer is added up to 20 ml and the tube is centrifuged at 8000×g for 5 min. The obtained supernatant is centrifuged for 2 hrs at 42000 rpm at 4° C. The pellet (microsomal fraction) is resuspended in a small volume of 0.1 M phosphate buffer pH 7.2. Protein content in the microsomal fraction is measured according to Bradford (Bradford, M. M. (1976), *Anal. Biochem. Bd.* 72, pp. 248-254) with bovine serum albumin as standard. The substrate selectivity can be determined by mixing equimolar amounts of different acyl-CoA:s in the same reaction and measure the preference for using the different acyl groups as substrates. The assay is run as in the specificity studies (Example 7) but scaled up 18 times to get sufficient amount of TAG for detection. Up to 4 different acyl-CoA:s can be used in the assay in equimolar amount instead of one single acyl-CoA. Samples are incubated for 4 min at 30° C. The assays are terminated by extraction of the lipids into chloroform according to Bligh and Dyer (Bligh, E. G. and Dyer, W. J. (1959), *Can. J. Biochem. Physiol.* 37, pp. 911-917). Lipids are separated on thin layer chromatography (TLC) silica 60 plates (Merck) using chloroform/methanol/acetic acid/water (90:15:10:3), and autoradiographic pictures are taken using an instant imager (Packard). The TAG is recovered from the plate and the fatty acids methylated in situ on the gel with sulphuric acid (2%) in methanol. Fatty acid methyl esters are extracted with hexane and separated by gas-liquid chromatography (GLC) using a WCOT fused silica 50 m×0.32 mm ID capillary column coated with CP-Wax 58-CB DF=0.3 (Chrompack inc., The Netherlands) and quantified relative to methyl-heptadecanoate added as an internal standard. The selectivity can be determined by calculating the amount of each acyl group that has been acylated to TAG.

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atgaacaata cttgggatat ctccagggtg gaacgcgtta aatgtgcgat attcgggtcca 420
atgctcatcc cccccgtct gctcctgctc tttgtgtcac ttcttggtgc ctacgggttc 480
ggcaagctct ctaccattgg cgcagaacta gagcgcccct tgcctcgatg gcgcatcgac 540

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ctgcagcacc ccatgaagtt ttttgcccgc gggattatgt ttgcattggg ctaccattgg 600
atctccatca aaggaaagca agcaagcccc caacacgctc ctatcgttgt ctccaatcat 660
tgctecttct gtgaagccat ctatctgect gggcgectct tgtccatggc tgtttcccgc 720
cgggagaatg cegctatecc tttttttgga gggctgatgc aacaagtcca atgcatcttc 780
gtctcgcgca cgcacaaaga ctcccggacc actgtcgcca acgagatctt gagacgctcc 840
aaaatagaaa gggggcagtg gcaccgctca ctctctgtct tcccagaagg gaccaccacg 900
aacgggagtg ccgtgatcag cttcaaagtc ggctccttgg cgggtggggg aagcgtgcag 960
ccagtcgctg taccctacc ttccaaccaa atctgcgac catcatgggt cagtgtgggg 1020
ccgcatcccg gcgagattct gtttaaattg ctgtgtcagc catggaacag tatgaatgtt 1080
actttcctgc ctgtgtataa tcccgcagcc gctgaaattg atgatcccgt gctgttttagc 1140
acaaatgtca ggcggttgat agccgcagag ttgggcgtgc ctgccagtga tcacacatte 1200
gatgacgttt tgttgttaat ggaggcaaag aagctagggt accagggggg tcttcgtgat 1260
tgcattctct agctgaaaaa tatgcgaaa attctagaaa ttgacctggc aaaagcgaaa 1320
gaatatttgc atgaattttc tcagcttgac acaaacagga aggggctgtt atcatacccc 1380
caattcatta aagccttcgg ctccgaggat tcagacgcac ttcggagtct atttgtgtg 1440
ttagacgtgc aagatcgggg agtgcataat ttggtggagt acaccacagg gttagcactg 1500
ttgaatgagc aaggcaccca tggttttgat ggggccatgc gcttgatttt caaagttcaa 1560
gattcgagtg gggagggggc gctgtcgaag gaagacacgg caaaggtgct gcggcggtg 1620
tggcctgacg tgacgacgga gctgttcgac tcgacgtttg ctgcggcgga cacagataat 1680
aacgggacgt tgagcgctga tgagtttctg gcgttggcga ggtcaaatca acacttgtgc 1740
ccgtcgtcct agagctcgtt gtgcgggagg ctttga 1776

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<210> SEQ ID NO 5
<211> LENGTH: 591
<212> TYPE: PRT
<213> ORGANISM: Nannochloropsis oculata

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<400> SEQUENCE: 5

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Met Thr Thr Thr Val Ile Ser Ser Ser Met Gly Pro Ile Leu Ala Tyr
1          5          10          15
Tyr Thr Cys Ala Thr Ile Thr Ile Tyr Val Val Leu Gly Arg Phe Ser
20          25          30
Ser Pro Asn Pro Arg Leu Arg Trp Leu Lys Leu Lys Asp Leu Glu Asn
35          40          45
Ile Glu Thr Ala Asn Pro Ala Ala His Pro Ser Glu Ser Asp Ser Met
50          55          60
Pro Leu Asn Ser Gly Asn Leu Ser Ser Ser Lys Pro Ile Ala Ala Ala
65          70          75          80
Glu Met Leu Gln Thr Pro Ser Ala Ser Ser Ser Ser Pro Ser Ala Ser
85          90          95
Pro Glu Arg Lys Ala Pro Met Met Arg Lys Leu Ser Phe Leu Ala Thr
100         105         110
Thr Gly Val Ile Glu Asn Pro Phe Met Asn Asn Thr Trp Asp Ile Ser
115         120         125
Arg Leu Glu Arg Val Lys Cys Ala Ile Phe Gly Pro Met Leu Ile Pro
130         135         140

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Pro Arg Leu Leu Leu Leu Phe Val Ser Leu Leu Gly Ala Tyr Gly Phe
 145 150 155 160
 Gly Lys Leu Ser Thr Ile Gly Ala Glu Leu Glu Arg Pro Leu Pro Arg
 165 170 175
 Trp Arg Ile Asp Leu Gln His Pro Met Lys Phe Phe Ala Arg Gly Ile
 180 185 190
 Met Phe Ala Leu Gly Tyr His Trp Ile Ser Ile Lys Gly Lys Gln Ala
 195 200 205
 Ser Pro Gln His Ala Pro Ile Val Val Ser Asn His Cys Ser Phe Cys
 210 215 220
 Glu Ala Ile Tyr Leu Pro Gly Arg Leu Leu Ser Met Ala Val Ser Arg
 225 230 235 240
 Arg Glu Asn Ala Ala Ile Pro Phe Phe Gly Gly Leu Met Gln Gln Val
 245 250 255
 Gln Cys Ile Phe Val Ser Arg Thr Asp Lys Asp Ser Arg Thr Thr Val
 260 265 270
 Ala Asn Glu Ile Leu Arg Arg Ser Lys Ile Glu Arg Gly Gln Trp His
 275 280 285
 Arg Gln Leu Leu Val Phe Pro Glu Gly Thr Thr Thr Asn Gly Ser Ala
 290 295 300
 Val Ile Ser Phe Lys Val Gly Ser Phe Ala Gly Gly Val Ser Val Gln
 305 310 315 320
 Pro Val Ala Val Ser Tyr Pro Ser Asn Gln Ile Cys Asp Pro Ser Trp
 325 330 335
 Val Ser Gly Gly Pro His Pro Gly Glu Ile Leu Phe Lys Leu Leu Cys
 340 345 350
 Gln Pro Trp Asn Ser Met Asn Val Thr Phe Leu Pro Val Tyr Asn Pro
 355 360 365
 Asp Ala Ala Glu Ile Asp Asp Pro Val Leu Phe Ser Thr Asn Val Arg
 370 375 380
 Arg Leu Ile Ala Ala Glu Leu Gly Val Pro Ala Ser Asp His Thr Phe
 385 390 395 400
 Asp Asp Val Leu Leu Leu Met Glu Ala Lys Lys Leu Gly Tyr Gln Gly
 405 410 415
 Gly Leu Arg Asp Cys Ile Ser Glu Leu Lys Asn Met Arg Lys Ile Leu
 420 425 430
 Glu Ile Asp Leu Ala Lys Ala Lys Glu Tyr Leu His Glu Phe Ser Gln
 435 440 445
 Leu Asp Thr Asn Arg Lys Gly Leu Leu Ser Tyr Pro Gln Phe Ile Lys
 450 455 460
 Ala Phe Gly Ser Gln Asp Ser Asp Ala Leu Arg Ser Leu Phe Cys Val
 465 470 475 480
 Leu Asp Val Gln Asp Arg Gly Val Ile Asn Leu Val Glu Tyr Thr Thr
 485 490 495
 Gly Leu Ala Leu Leu Asn Glu Gln Gly Thr Asp Gly Phe Asp Gly Ala
 500 505 510
 Met Arg Leu Ile Phe Lys Val Gln Asp Ser Ser Gly Glu Gly Arg Leu
 515 520 525
 Ser Lys Glu Asp Thr Ala Lys Val Leu Arg Arg Leu Trp Pro Asp Val
 530 535 540

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Thr Thr Glu Leu Phe Asp Ser Thr Phe Ala Ala Ala Asp Thr Asp Asn
545 550 555 560

Asn Gly Thr Leu Ser Ala Asp Glu Phe Leu Ala Leu Ala Arg Ser Asn
565 570 575

Gln His Leu Cys Pro Ser Leu Lys Ser Ser Leu Cys Gly Arg Leu
580 585 590

<210> SEQ ID NO 6
 <211> LENGTH: 2247
 <212> TYPE: DNA
 <213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 6

```

aaaaagtttg agattttcag caaagtaatc aagataataa acaaaaacaa tcctataaag    60
gaaaaacaac agggactatt tgcctcgcct cctcacgcct gcccaattag gggaccaacg    120
atcacaacta tgaccacgac tgctcatctct agctcgatgg ggcccatcct ggcctattat    180
acgtgtgcc aatcaccat ctacgtagtg ctcgccgcct tttccagtcc aaaccgcgc    240
ttgagatggc tgaagctcaa agacctggag aacattgaga ctgcgaacc ggccgcgcac    300
ccttcagagt ctgattctat gcctcttaat tetggcaatc tatcgtcttc caagcccatt    360
gccgcagctg agatgcttca aactccctcg gcacgcctct cctcgccctc ggcacccca    420
gagcgcgaaag ctccatgatg gcggaagctt tctttctcgc ccacgactgg agtcatcgaa    480
aatcccttta tgaacaatac ttgggataac tccaggttgg aacgcgttaa atgtgcgata    540
ttcgggtcaa tgctcatccc ccccgcctcg ctctctctct ttgtgtcact tcttggtgcc    600
tacgggttcg gcaagctctc taccattggc gcagaactag agcgcgccctt gcctcgatgg    660
cgcatcgacc tgcagcacc ccatgaagttt tttgcccgcg ggattatggt tgcattgggc    720
taccattgga tctccatcaa aggaaagcaa gcaagcccgc aacacgctcc tatcgttgte    780
tccaatcatt gctcctcttg tgaagccatc tatctgcctg ggcgcctctt gtccatggct    840
gtttcccgcc gggagaatcg cgctatccct ttttttgagg ggctgatgca acaagtccaa    900
tgcatcttcg tctcgcgcac cgacaaagac tcccggacca ctgtcgccaa cgagatcttg    960
agacgctcca aaatagaaa ggggcagtg caccgtcaac tctcgtctt cccagaaggg    1020
accaccacga acgggagtg cgtgatcagc ttcaaagtcg gctcctttgc cgggtgggta    1080
agcgtgcagc cagtcgctgt atcctaccct tccaacaaa tctgcatcc atcatgggtc    1140
agtgggtggc cgcatecccg cgagattctg tttaaattgc tgtgtcagcc atggaacagt    1200
atgaatgta ctttctctgc tgtgtataat cccgacgcgc ctgaaattga tgatcccgtg    1260
ctgtttagca caaatgtcag cgggttgata gccgcagagt tgggcgtgcc tgccagtgat    1320
cacacattcg atgacgtttt gttgttaatg gaggcaaaga agctagggtg ccaggggggt    1380
cttcgtgatt gcatctctga gctgaaaaat atgcaaaga ttctagaaat tgacctggca    1440
aaagcgaag aatatttgca tgaattttct cagcttgaca caaacaggaa ggggctgtta    1500
tcatacccc aattcattaa agccttcggc tcgcaggatt cagacgcact tcggagtcta    1560
ttttgtgtg tagactgca agatcgggga gtgatcaatt tgggtggagta caccacaggg    1620
ttagcactgt tgaatgagca aggcaccgat ggttttgatg gggccatgcg cttgattttc    1680
aaagttcaag attcagagtg ggagggggcg ctgtcgaagg aagacacggc aaaggtgctg    1740
cggcgctgt ggcctgacgt gacgacggag ctgttcgact cgacgtttgc tgcggcggac    1800

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acagataata acgggacgtt gagcgtgat gagtttctgg cgttggcgag gtcaaatcaa 1860
cacttggtcc cgtcgtcaa gagctcgttg tgcgggaggc tttgagtaa tgttttatgc 1920
tgcattgttt ataagaagca tgtatgtgaa aatgtaaata gattagacct ggtgtagatt 1980
ggctaggagt ttaataggca aggttcctatg tcgaaaaaaa atgtgcccgcg attaaagtga 2040
ggaaaacaca ctcaatttctt tacacaattt ggaacacttt gttcctctat ttcgcataaa 2100
acagcgacca gcaattcaac cgcacgagcg tctcatagca ccaaaccctc ctgttcatcc 2160
ctccaacctt cctcctcccc ccttcgacct tetgtctctc cactttcatt cctcctcaac 2220
catttactca tgcaatcctc tcggcct 2247

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<210> SEQ ID NO 7
<211> LENGTH: 1134
<212> TYPE: DNA
<213> ORGANISM: Thraustochytrium aureum

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<400> SEQUENCE: 7

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```

atggagggca tcgagtcgat agtggacgac gacttttggg agtgcttcca gagccggaaa 60
ccgcgacctt ggaactggaa tgcctacttg tggccgctgt gggetgcccgg tgtctttatc 120
cggtaactttg tccttttccc gatccgctt gcgatttttg cgatgggctg gattctgttc 180
ggaatcggga tgttggtcac gcaaaccctc tttccgcacg ggccgctcgc cacctcgttt 240
gagcacggac tgatctcgat gatgtgcggc gtgttctgta tcacctgggg ggcggtcctc 300
cggtagcacg ggtcgcgggt caagccgcga gagggcgagt gccagcccgt gtacgttgcc 360
aaccacactt cgatgatcga cgtcatcctc ttgcagcaga tgcgctgctt ttcgctcgtg 420
ggccagcgcc acaaaaggcat cgtcgggttt ttgcaagagg tcgtgctggg ctgtttgacg 480
tgcgtctggt tcgaccgccc cgagatcaag gacagggcag ccgtggcgcg caagctcaac 540
gagcatcgca acgaccgac tcgcaaccct ctgctcgtgt ttccggaggg aacgtgctg 600
aacaatgagt acgtgatcca gttcaagaag ggcactcttg agatcggcgc ccccgtggtc 660
ccagtcgcca tcaagtacaa caaaatgttc gtggaccctt tctggaactc gcgcgcgcag 720
tcgttcccga tgcacctcgt agagctcatg acctcgtggt gcctcatttg cgaggtttgg 780
tacctcaagc cgtcgcgagc catggagcgc gagtcgtcca ccgattttgc agcacgcgtg 840
aagaaggcga ttgcggacca ggcggcctt aagaacgtca actgggacgg ctacatgaag 900
tattggaagc catcggagcg ttacttgcgc gcgcgccagg cgatcttcgc caaaaactctc 960
cgcaaatcc actcgcgctc tttggagcag gacaaggctg accggcaggc cattctgcac 1020
gacctggacg gcgcgttccc ggattctggg acacaccgcg gcgagcgcga gtcgccaaga 1080
gagccgggtc tgcggcgcgc ccaggcggcc tccgcgccgg gagaagcctt atag 1134

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<210> SEQ ID NO 8
<211> LENGTH: 377
<212> TYPE: PRT
<213> ORGANISM: Thraustochytrium aureum

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<400> SEQUENCE: 8

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```

Met Glu Gly Ile Glu Ser Ile Val Asp Asp Asp Phe Trp Lys Cys Phe
1           5           10           15
Gln Ser Arg Lys Pro Arg Pro Trp Asn Trp Asn Ala Tyr Leu Trp Pro
20           25           30

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Leu Trp Ala Ala Gly Val Phe Ile Arg Tyr Phe Val Leu Phe Pro Ile
 35 40 45
 Arg Leu Ala Ile Phe Ala Met Gly Trp Ile Leu Phe Gly Ile Gly Met
 50 55 60
 Leu Val Thr Gln Thr Cys Phe Pro His Gly Pro Arg Arg Thr Ser Leu
 65 70 75 80
 Glu His Gly Leu Ile Ser Met Met Cys Gly Val Phe Cys Ile Thr Trp
 85 90 95
 Gly Ala Val Ile Arg Tyr His Gly Ser Pro Val Lys Pro Arg Glu Gly
 100 105 110
 Glu Cys Gln Pro Val Tyr Val Ala Asn His Thr Ser Met Ile Asp Val
 115 120 125
 Ile Ile Leu Gln Gln Met Arg Cys Phe Ser Leu Val Gly Gln Arg His
 130 135 140
 Lys Gly Ile Val Arg Phe Leu Gln Glu Val Val Leu Gly Cys Leu Gln
 145 150 155 160
 Cys Val Trp Phe Asp Arg Gly Glu Ile Lys Asp Arg Ala Ala Val Ala
 165 170 175
 Arg Lys Leu Asn Glu His Ala Asn Asp Pro Thr Arg Asn Pro Leu Leu
 180 185 190
 Val Phe Pro Glu Gly Thr Cys Val Asn Asn Glu Tyr Val Ile Gln Phe
 195 200 205
 Lys Lys Gly Ile Phe Glu Ile Gly Ala Pro Val Val Pro Val Ala Ile
 210 215 220
 Lys Tyr Asn Lys Met Phe Val Asp Pro Phe Trp Asn Ser Arg Ala Gln
 225 230 235 240
 Ser Phe Pro Met His Leu Val Glu Leu Met Thr Ser Trp Cys Leu Ile
 245 250 255
 Cys Glu Val Trp Tyr Leu Lys Pro Leu Glu Arg Met Glu Arg Glu Ser
 260 265 270
 Ser Thr Asp Phe Ala Ala Arg Val Lys Lys Ala Ile Ala Asp Gln Ala
 275 280 285
 Gly Leu Lys Asn Val Asn Trp Asp Gly Tyr Met Lys Tyr Trp Lys Pro
 290 295 300
 Ser Glu Arg Tyr Leu Arg Ala Arg Gln Ala Ile Phe Ala Lys Thr Leu
 305 310 315 320
 Arg Lys Ile His Ser Arg Ser Leu Glu Gln Asp Lys Ala Asp Arg Gln
 325 330 335
 Ala Ile Leu His Asp Leu Asp Gly Ala Phe Pro Asp Ser Gly Thr His
 340 345 350
 Arg Gly Glu Arg Glu Ser Pro Arg Glu Pro Gly Leu Arg Arg Arg Gln
 355 360 365
 Ala Ala Ser Ala Pro Gly Glu Ala Leu
 370 375

<210> SEQ ID NO 9

<211> LENGTH: 1288

<212> TYPE: DNA

<213> ORGANISM: Thraustochytrium aureum

<400> SEQUENCE: 9

atggagggca tcgagtcgat agtggacgac gacttttggg agtgcttcca gagccggaaa 60

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ccgcgacct ggaactggaa tgcctacttg tggccgctgt gggctgcggg tgtctttatc 120
cggtaactttg tecttttccc gatccggctt gegatttttg cgatgggctg gattctgttc 180
ggaatcggga tgttggtcac gcaaacctgc tttccgcacg ggccgcgtcg cacctcgctt 240
gagcacggac tgatctcgat gatgtgcggc gtgttctgta tcaactgggg ggccggtcacc 300
cggtagccacg ggtcgcgggt caagcccgca gagggcgagt gccagcccgt gtacgttgcc 360
aaccacactt cgatgatcga cgtcatcacc ttgcagcaga tgcctgctt ttcgctcgtg 420
ggccagcgc acaaaaggcat cgtcgggttt ttgcaagagg tcgtgctggg ctgtttgacg 480
tgcgtctggt tcgaccgcgg cgagatcaag gacagggcag ccgtggcgcg caagctcaac 540
gagcatgcga acgaccgcga tcgcaaccgg ctgctcgtgt ttcggagggg aacgtgcgtg 600
aacaatgagt acgtgatcca gttcaagaag ggcacttttg agatcggcgc ccccggtggtc 660
ccagtcgcca tcaagtacaa caaaatgttc gtggaccctt tctggaactc gcgcgcgcag 720
tcgttcccga tgcacctcgt agagctcatg acctcgtggt gcctcatttg cgaggtttgg 780
tacctcaagc cgctcgagcg catggagcgc gagtcgtcca ccgattttgc agcacgcgtg 840
aagaaggcga ttgcggacca ggccggcctt aagaacgtca actgggacgg ctacatgaag 900
tattggaagc catcggagcg ttacttgccc gcgcgcacag cgatcttcgc caaaactctc 960
cgcaaaatcc actcgcgctc tttggagcag gacaaggctg accggcaggg cattctgcac 1020
gacctggacg gcgcggtccc ggattctggg acacaccgcg gcgagcgcga gtcgccaaaga 1080
gagccggggtc tgcggcgccg ccagggcgcc tccgcgcccg gagaagcctt atagcggcgt 1140
ttgccttgca cgctgatcaa cgtggggcat gtgggtgctc tgtggccaag agcaggccct 1200
gcgctcggca ctgcagcgt acgctcagac ttttcgcggt ggggcatgca tgcatecaaa 1260
cattttcttc cttcttccaa aaaaaaaaa 1288

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<210> SEQ ID NO 10

<211> LENGTH: 1284

<212> TYPE: DNA

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 10

```

atgaagacgc ccacgagcct ggcgtgcgga gcctgcacgg cagccgtggt aatgtgtttc 60
acaacaacag cagatgcctt tgccagcaca tcacaaccgg gcagcgttgg cgtggctgtc 120
gcgcggcggc caccaggctt ccaactgata gggcgatcat cagccacgac taggagaata 180
agcaggggag ggatagagga tctcggaaac catcacacgt gggcgggcag gatgtcgcag 240
cagcaccagc agcaccagca gcaccagcag caccgtcggc gtaggaggac acccactatg 300
ctagtggaga cagacgtgaa ggtaaaagag gaagcgggga ttggccacgg atcaggaagc 360
aacgaaagtg gcaacaggag cggcaagagc gggctctcgg cggcagacgc ctcagaaggt 420
acaggcccac cgccagtgcc cgtggatacc ttccgcaca agagcttggc ggaggtcccg 480
acggactatg gacctacct gaccattaaa gggttcaaga tcaatgcctt tggcttctat 540
ttctgcttcg tggccctatt ctgggggatc cctgggggtg tottctcat cctgtacaag 600
gogagtttgg agttcatgga caagatcgat cctcgcgggt acaacgtgga ccgctccagt 660
tccctatggg gctggtgac cagtatcagt actgactcct taccgacat taeggcatg 720
gagaacatc ccaagggacc ggcggtcttc gtcgccaacc acgctcctg gatggacgtg 780

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ccctacactg cccaactgcc catccgcgcc aagtacctag cgaaagctga cctggccaag   840
atcccaatcc tgggcaacgc catgagcatg gctcagcagc tcctcctcga tegagacgac   900
aagcgcagtc aaatggaagc cctgcgctct gctctcctga tcctcaagac aggcaccccc   960
atcttcgtct tcccgcaggg caccctgggg cctcaaggcc gaatgcagac cttaagatg  1020
gggtgcattca aggtggcgac caagggcgggc gtgcctatag tgctgtatc tatcgcgggg  1080
acgcatgtca tgatgcccac ggaggtgatc atgcctcaat gtgctggcgc gggaaatcacc  1140
gccattcatg tccacctccc catctccatc aagggcgcga cggaccagga gctgtcggat  1200
ctggcgtttg atactattaa caatgcattg tcagatgagc agcgggctat gcctagcagg  1260
aagaaggacg attcgagagc ttaa                                     1284

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<210> SEQ ID NO 11
<211> LENGTH: 427
<212> TYPE: PRT
<213> ORGANISM: Nannochloropsis oculata

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<400> SEQUENCE: 11

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```

Met Lys Thr Pro Thr Ser Leu Ala Cys Gly Ala Cys Thr Ala Ala Val
 1                               5 10 15
Leu Met Cys Phe Thr Thr Thr Ala Asp Ala Leu Ala Ser Thr Ser Gln
 20 25 30
Pro Gly Ser Val Gly Val Ala Val Ala Arg Arg Pro Pro Gly Phe His
 35 40 45
Ser Ile Gly Arg Ser Ser Ala Thr Thr Arg Arg Ile Ser Arg Gly Gly
 50 55 60
Ile Glu Asp Leu Gly Thr His His Thr Trp Gly Gly Arg Met Ser Gln
 65 70 75 80
Gln His Gln Gln His Gln Gln His Gln Gln His Arg Arg Arg Arg Arg
 85 90 95
Thr Pro Thr Met Leu Val Glu Thr Asp Val Lys Val Lys Glu Glu Ala
 100 105 110
Gly Ile Gly His Gly Ser Gly Ser Asn Glu Ser Gly Asn Arg Ser Gly
 115 120 125
Lys Ser Gly Ser Ala Ala Ala Asp Ala Ser Glu Gly Thr Gly Pro Pro
 130 135 140
Pro Val Pro Val Asp Thr Phe Arg His Lys Ser Leu Ala Glu Val Pro
 145 150 155 160
Thr Asp Tyr Gly Pro Tyr Leu Thr Ile Lys Gly Phe Lys Ile Asn Ala
 165 170 175
Phe Gly Phe Tyr Phe Cys Phe Val Ala Leu Phe Trp Ala Ile Pro Trp
 180 185 190
Gly Val Phe Leu Ile Leu Tyr Lys Ala Ser Leu Glu Phe Met Asp Lys
 195 200 205
Ile Asp Pro Arg Arg Tyr Asn Val Asp Arg Ser Ser Ser Leu Trp Gly
 210 215 220
Trp Leu Thr Ser Ile Ser Thr Asp Ser Leu Pro Asp Ile Thr Gly Met
 225 230 235 240
Glu Asn Ile Pro Lys Gly Pro Ala Val Phe Val Ala Asn His Ala Ser
 245 250 255
Trp Met Asp Val Pro Tyr Thr Ala Gln Leu Pro Ile Arg Ala Lys Tyr

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	260		265		270	
Leu	Ala	Lys	Ala	Asp	Leu	Ala
	275		Lys	Ile	Pro	Ile
			280		Leu	Gly
					285	Asn
						Ala
						Met
Ser	Met	Ala	Gln	His	Val	Leu
290					295	Leu
						Asp
						Arg
						Asp
						300
						Lys
						Arg
						Ser
						Gln
Met	Glu	Ala	Leu	Arg	Ser	Ala
305				310		Leu
						Leu
						Ile
						Leu
						Lys
						315
						Thr
						Gly
						Thr
						Pro
320						
Ile	Phe	Val	Phe	Pro	Glu	Gly
				325		Thr
						Arg
						Gly
						Pro
						Gln
						Gly
						Arg
						Met
						335
						Gln
Thr	Phe	Lys	Met	Gly	Ala	Phe
			340			Lys
						Val
						Ala
						Thr
						Lys
						Ala
						Gly
						Val
						Pro
						350
Ile	Val	Pro	Val	Ser	Ile	Ala
						Gly
						Thr
						His
						Val
						Met
						Met
						Pro
						Lys
						Glu
						365
Val	Ile	Met	Pro	Gln	Cys	Ala
						Gly
						Arg
						Gly
						Ile
						Thr
						Ala
						Ile
						His
						Val
						370
						375
His	Pro	Pro	Ile	Ser	Ile	Lys
385						Gly
						Arg
						Thr
						Asp
						395
						Gln
						Glu
						Leu
						Ser
						Asp
						400
Leu	Ala	Phe	Asp	Thr	Ile	Asn
						Asn
						Ala
						Leu
						Ser
						Asp
						410
						Glu
						Gln
						Arg
						Ala
						415
Met	Pro	Ser	Arg	Lys	Lys	Asp
						Asp
						Ser
						Arg
						Ala
						425

<210> SEQ ID NO 12
 <211> LENGTH: 1826
 <212> TYPE: DNA
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 12

```

aagataataa caaaaacaat cctctaaaag gaaaacaaca ggtgtacaat tccaggacag      60
acgacaagtg attcatgaag acgcccacga gcctggcgtg cggagcctgc acggcagccg      120
tgtaaatgtg tttcacaaca acagcagatg cccttgccag cacatcacia cggggcagcg      180
ttggcgtggc tgtcgcgcgg cggccaccag gcttccactc gatagggcga tcatcagcca      240
cgactaggag aataagcagg ggagggatag aggatctcgg aaccatcac acgtggggcg      300
gcaggatgtc gcagcagcac cagcagcacc agcagcacca gcagcaccgt cggcgtagga      360
ggacaccac tatgctagtg gagacagacg tgaagtaaa agaggaagcg gggattggcc      420
acggatcagg aagcaacgaa agtggaaca ggagcggcaa gagcgggtct gcggcggcag      480
acgcctcaga aggtacaggc ccaccgccag tgcccgtgga taccttcggg cacaagagct      540
tggcggaggt cccgacggac tatggaccct acctgaccat taaagggttc aagatcaatg      600
cctttggett ctatttctgc ttcgtggccc tattctgggc gatcccctgg ggtgtettcc      660
tcatcctgta caaggcgagt ttggagtta tggacaagat cgatcctcgc cggtacaacg      720
tggaccgctc cagttcccta tggggctggc tgaccagtat cagtactgac tccttaccgg      780
acattacggg catggagaac attcccagg gaccggcggc cttcgtcggc aaccacgcct      840
cctggatgga cgtgccttac actgcccac tgccatccg cgccaagtac cttagcгааg      900
ctgacctggc caagatccca atcctgggca acgccatgag catggctcag cagctectcc      960
tcgatcgaga cgacaagcgc agtcaaagg aagccctgcg ctctgctctc ctgatcctca     1020
agacaggcac ccccatcttc gtcttcccc agggcaccgg tgggcctcaa ggccgaatgc     1080
    
```

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agacctttaa gatgggtgca ttcaaggtgg cgaccaaggc gggcgtgcct atagtgcctg 1140
tatctatcgc ggggacgcat gtcattgatgc ccaaggaggt gatcatgcct caatgtgctg 1200
gccggggaat caccgccatt catgtccacc ctcccatctc catcaagggc cgcacggacc 1260
aggagctgtc ggatctggcg tttgatacta ttaacaatgc attgtcagat gacgagcggg 1320
ctatgcctag caggaagaag gacgattcga gagcttaaga agaaggaaaa gagaagatgt 1380
gaaggaatga ggtgaaggca tgtcaacaat aggagataga gatcatgaag agatgagagc 1440
gagggaatca aaacccttc agtaagcctt gtgtagatca tatgcaggaa aagtgagcaa 1500
caggagcggc aggagaagca gttggggcga tcgagaaga caattaccaa gcaggaggca 1560
ataaaaggca attatcgaat agatttggag cgggggggtca gcgcacagcc gaacaagatg 1620
ccgtgtgctt agcagcagca gaatccgacc atagcgtaaa cctcacgaat gtttgtggtg 1680
agaagatggc aaatcaaat cttcatcgtt tgtttgcaat tggatgatga tgagattcct 1740
atagaccaga gagactggga agcttcacct ggagtaacag aaagaaagac taacagacga 1800
caacaaaaaa aaaaaaaaaa aaaaaaa 1826

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<210> SEQ ID NO 13

<211> LENGTH: 1395

<212> TYPE: DNA

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 13

```

atggtcagga ggaagatgga cgtggacagc tggcgcggc gcgaagcggc gtcagctacg 60
agcaacggcg ccaacgtccc gtcgtccacc tcctctacag cctccgcttc ttctctctcc 120
aaaggcacc taccgcagc tgctccaggcc ctgcaaacga aggcgcgacc attgcctcag 180
cctttatcga atgtggcaaa acgcgccttg tactacgagg cggaatgct ctggcaatca 240
atcaaggatg agctgcccgc cgagcaccgc gaccaggcct ctttacttgc ggcaatcgac 300
cagttcgaga ccaaccttct acgcatcagt cccgctcagc tcgccaccac ctctttacga 360
cggatcctac aacaactcga catgctcctg cgaatcatta cttgctcct ctacctctgc 420
cttctagggg tcatcacatt tttgccatg atcaactctg ttcccatcct cgaccgcctc 480
ctcgtaatcc tgggctggcc ccgtcgtttc ctcatctacg aactggcaca aaaggcatct 540
gcacgtggat ttctctacat ggccgggtgt ttctacacgg aagaagggaa gcaagccaat 600
gggatgaaa ccccccttgt cctcctcttt caaacggct cgaaccttga tggtctcttg 660
atcttgatt ccttctctca attctttaa tcaatcggga aagacgacat ctttctcatg 720
ccttacgtag ggtggatggc atatgtgtac ggcattctac ctatcgaccg caagcatcgt 780
aacgaagcaa tcaaacagct aggaacgacc acccgcgtct gtacctctgg tgtggcctc 840
gctctttccc ccgaggggac acgtagcaag accggacaat tgatgcgatt caagaaaggg 900
ccgttttact tacaagccga gacatcggct actgtcacc ctcttgcct cgttggaat 960
tacgagttgt ggcctccaaa ctatttcttt acctgtcctg ggcaggtggt gatgaggtat 1020
ctcccccca ttgaccatc ctccctcctt ccctcgggtg gtcggaacaa agacgagttc 1080
agtcgatatg tgcgaagca gatgtttgag gccattgatg atatcatggc tggttccgag 1140
gagggagggg aggaggtagg ggagaagagg aaaaaatag cgcggggggg gaaattgacc 1200
tgggtggtgc ggggagtgaa tttggcatgc atgtgcctgt tttggtgat ggtaaaggcg 1260

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gcgtggatgg tggtaacggg ggtgagtgac gcgtatgggt tcagtagggg ggcgttggeg 1320
gggggattcg ttgcatacac ggtgagtggt actgctggcc tgtatatatt gtactgcaag 1380
gcgccggcgt cgtga 1395

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<210> SEQ ID NO 14
<211> LENGTH: 464
<212> TYPE: PRT
<213> ORGANISM: Nannochloropsis oculata

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<400> SEQUENCE: 14

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```

Met Val Arg Arg Lys Met Asp Val Asp Ser Ser Ala Ala Gly Glu Ala
1          5          10          15
Ala Ser Ala Thr Ser Asn Gly Ala Asn Val Pro Ser Ser Thr Ser Ser
20          25          30
Thr Ala Ser Ala Ser Ser Ser Ser Lys Gly Thr Leu Pro Ala Arg Val
35          40          45
Gln Ala Leu Gln Thr Lys Ala Ala Thr Leu Pro Gln Pro Leu Ser Asn
50          55          60
Val Ala Lys Arg Ala Leu Tyr Tyr Glu Ala Glu Met Leu Trp Gln Ser
65          70          75          80
Ile Lys Asp Glu Leu Pro Ala Glu His Pro Asp Gln Ala Ser Leu Leu
85          90          95
Ala Ala Ile Asp Gln Phe Glu Thr Asn Leu Leu Arg Ile Ser Pro Ala
100         105         110
Gln Leu Ala Thr Thr Ser Leu Arg Arg Ile Leu Gln Gln Leu Asp Met
115         120         125
Leu Leu Arg Ile Ile Thr Cys Ser Leu Tyr Leu Cys Leu Leu Gly Val
130         135         140
Ile Thr Phe Leu Pro Met Ile Thr Leu Val Pro Ile Leu Asp Arg Leu
145         150         155         160
Leu Val Ile Leu Gly Trp Pro Arg Arg Phe Leu Ile Tyr Glu Leu Ala
165         170         175
Lys Lys Ala Ser Ala Arg Gly Phe Leu Tyr Leu Ala Gly Val Phe Tyr
180         185         190
Thr Glu Glu Gly Lys Gln Ala Asn Gly Tyr Glu Thr Pro Leu Val Leu
195         200         205
Leu Phe Gln His Gly Ser Asn Leu Asp Gly Phe Leu Ile Leu Asp Ser
210         215         220
Phe Pro Gln Phe Phe Lys Ser Ile Gly Lys Asp Asp Ile Phe Leu Met
225         230         235         240
Pro Tyr Val Gly Trp Met Ala Tyr Val Tyr Gly Ile Leu Pro Ile Asp
245         250         255
Arg Lys His Arg Asn Glu Ala Ile Lys Gln Leu Gly Arg Ala Thr Arg
260         265         270
Val Cys Thr Ser Gly Val Ala Val Ala Leu Ser Pro Glu Gly Thr Arg
275         280         285
Ser Lys Thr Gly Gln Leu Met Arg Phe Lys Lys Gly Pro Phe Tyr Leu
290         295         300
Gln Ala Glu Thr Ser Ala Thr Val Thr Pro Leu Val Ile Val Gly Asn
305         310         315         320
Tyr Glu Leu Trp Pro Pro Asn Tyr Phe Phe Thr Cys Pro Gly Gln Val
325         330         335

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Val Met Arg Tyr Leu Pro Pro Ile Asp His Ser Ser Leu Pro Pro Ser
 340 345 350

Val Gly Arg Asn Lys Asp Glu Phe Ser Arg Tyr Val Arg Lys Gln Met
 355 360 365

Phe Glu Ala Ile Asp Asp Ile Met Ala Gly Ser Glu Glu Gly Gly Lys
 370 375 380

Glu Val Gly Glu Lys Arg Lys Lys Tyr Ala Pro Gly Gly Lys Leu Thr
 385 390 395 400

Trp Trp Leu Arg Gly Val Asn Leu Ala Cys Met Cys Leu Phe Trp Leu
 405 410 415

Met Val Lys Ala Ala Trp Met Val Val Thr Gly Val Ser Asp Ala Tyr
 420 425 430

Gly Phe Ser Arg Gly Ala Leu Ala Gly Gly Phe Val Ala Tyr Thr Val
 435 440 445

Ser Val Thr Ala Gly Leu Tyr Ile Leu Tyr Cys Lys Ala Pro Ala Ser
 450 455 460

<210> SEQ ID NO 15

<211> LENGTH: 1771

<212> TYPE: DNA

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 15

```

atttttcagc aaagtaatca agataataaa caaaaacaat cctataaagg aaaaacaaca    60
ggacaaaatca atggtcagga ggaagatgga cgtggacagc tcggccgccg gccaagcggc    120
gtcagctacg agcaacggcg ccaacgtccc gtcgtccacc tcctctacag cctccgcttc    180
ttcctcctcc aaaggeaccc taccgcacg tgtccaggcc ctgcaaacga aggccgccac    240
attgctcag cctttatcga atgtggcaaa acgcgccttg tactacgagg cggaaatgct    300
ctggcaatca atcaaggatg agctgcccgc cgagcaccgc gaccaggcct ctttacttgc    360
ggcaatcgac cagttecgaga ccaaccttct acgcatcagt cccgctcage tegccaccac    420
ctctttacga cggatcctac aacaactcga catgctcctg cgaatcatta cttgctcct    480
ctactctgc cttctagggg tcatcacatt tttgcccatt atcactctcg ttccatecct    540
cgaccgcctc ctcgtaatcc tgggctggcc ccgtegttcc ctcatctacg aactggccaa    600
aaaggcatct gcacgtggat ttctctacct ggccggtggt ttctacacgg aagaagggaa    660
gcaagccaat gggatgaaa cccccctgt cctcctcttt caacacggct cgaaccttga    720
tggcttcttg atcttggatt cctttcctca attctttaa tcaatcggga aagacgacat    780
cttctcatg ccttaactag ggtggatggc atatgtgtac ggcatctac ctatcgaccg    840
caagcatcgt aacgaagcaa tcaaacagct aggacgagcc acccgcgtct gtacctctgg    900
tgtggccgtc gctctttccc ccgagggggc acgtagcaag accggacaat tgatgcgatt    960
caagaaaggg ccgctttact tacaagccga gacatcggt actgtcacc ctcttgtcat   1020
cgttgaaat tacgagttgt ggctccaaa ctatttcttt acctgtcctg ggcaggtggt   1080
gatgaggtat ctcccccca ttgaccattc ctccctcct cctcgggttg gtcggaacaa   1140
agacgagttc agtcgatatg tgcccaagca gatgtttgag gccattgatg atatcatggc   1200
tggttccgag gagggaggga aggaggtagg ggagaagagg aaaaaatag cgcggggggg   1260
gaaattgacc tgggtggtgc ggggagtga tttggcatgc atgtgcctgt tttggtgat   1320

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ggtaaaggcg gcgtggatgg tggtaacggg ggtgagtgac gcgtatgggt tcagtagggg 1380
ggcgttgccg gggggattcg ttgcatacac ggtgagtgtg actgctggcc tgtatatatt 1440
gtactgcaag gcgccggcgt cgtgagaggg gggaaaggag gggggaagga gagatagaag 1500
acgaggtaga ggtagatgtg agtgtgagat agcgcgagta ttatctttaa gaaaagagat 1560
gaattgtagt agaagagtcg ggtattttag caggagagaga atattgtatg gagggtaaac 1620
gtgtgggaaa gaggaggag ggacctgaga tggataatga aagaatacta gagagagcgc 1680
gtgacacggt cattgcttcc tcggattagt tgctgtgca taagttaaag ataatagaga 1740
ggaatggcgc tcgcatgctc ctctttacac t 1771

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<210> SEQ ID NO 16
<211> LENGTH: 1026
<212> TYPE: DNA
<213> ORGANISM: Nannochloropsis oculata

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<400> SEQUENCE: 16

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```

atggcacect cccaaccggc cccgccact gcacccgaga acccctacaa cctattgcca 60
cccaagcgc ccaatecgc gtaactggcg tatgcaagcc ttgccgctt ccttctcaet 120
tgettctcgg ccccttccag taactcgtgg gccaccacco tccgccgcgc ctgctgggcg 180
gcgtactgga cgacctacat ggacacaagc tataaggacg gctcacgggc ctggccctgg 240
tttcagcgat tgcgaatctg gcgtatgtat tgcggctatt tgcaggcaa agtcatttgc 300
acggtgcct tggaccggc gcagcaattt atcttcgcg cccatcccca cggcattggt 360
acctggaacc atttctgac catgactgac ggctgtcgat ttctctctc ctctaccoc 420
cgcccgcggc tcgacctggg tgcgacagta cttttcttca tccccttctt aaaggaaatt 480
ctgctttggc taggctgtgt ggatgctgga ggggccacgg ctcatgcggt tttggcgcg 540
ggctactcct cctcattta catcggtgga gaaaagagc agatttgac acggcgaggc 600
aaagacatcg tgggtgtaac tccccgcaag ggtttttgca agctggcctt ccagcataac 660
tgcccatcgc taccggtcta cgcatttggg gaaaacgac tgtatcgac gttcaaccac 720
ctcaaggact tccagctgtg ggtggctagc gccttcaagc tcgcttttcc tccttgttgg 780
ggcgtctct tctccctt cctccccct cccgtctcta tcacggtggt gatggcgag 840
cccttgctac ccagagcaca aaaaggaagt gcgagaagga gtggtggagg aaaaggggtg 900
gagccgacga gggaggaggt ggaggagctg cacttccgat acgtggaggc cttgcagaag 960
ttgtttgacg cacacaaagt caggcaggga gggaggagcg aagaggccac cttagtgttc 1020
aatga 1026

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<210> SEQ ID NO 17
<211> LENGTH: 341
<212> TYPE: PRT
<213> ORGANISM: Nannochloropsis oculata

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<400> SEQUENCE: 17

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Met Ala Pro Ser Pro Pro Ala Pro Pro Pro Ala Pro Glu Asn Pro Tyr
1           5           10           15
Asn Leu Leu Pro Pro Lys Arg Pro Asn Pro Gln Tyr Trp Arg Tyr Ala
20           25           30
Ser Leu Ala Ala Phe Leu Leu Thr Cys Phe Leu Ala Pro Ser Ser Asn

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	35		40		45										
Ser	Trp	Ala	Thr	Thr	Leu	Arg	Arg	Ala	Cys	Trp	Ala	Ala	Tyr	Trp	Thr
	50					55					60				
Thr	Tyr	Leu	Asp	Thr	Ser	Tyr	Lys	Asp	Gly	Ser	Arg	Ala	Trp	Pro	Trp
	65				70				75						80
Phe	Gln	Arg	Leu	Arg	Ile	Trp	Arg	Met	Tyr	Cys	Gly	Tyr	Leu	Gln	Gly
				85					90					95	
Lys	Val	Ile	Cys	Thr	Val	Pro	Leu	Asp	Pro	Ala	Gln	Gln	Phe	Ile	Phe
			100					105					110		
Ala	Ala	His	Pro	His	Gly	Ile	Gly	Thr	Trp	Asn	His	Phe	Leu	Thr	Met
		115					120					125			
Thr	Asp	Gly	Cys	Arg	Phe	Leu	Ser	Ser	Ser	Tyr	Pro	Arg	Pro	Arg	Leu
	130					135					140				
Asp	Leu	Gly	Ala	Thr	Val	Leu	Phe	Phe	Ile	Pro	Phe	Leu	Lys	Glu	Ile
	145				150					155				160	
Leu	Leu	Trp	Leu	Gly	Cys	Val	Asp	Ala	Gly	Ala	Ala	Thr	Ala	His	Ala
			165						170					175	
Val	Leu	Ala	Arg	Gly	Tyr	Ser	Ser	Leu	Ile	Tyr	Ile	Gly	Gly	Glu	Lys
			180					185					190		
Glu	Gln	Ile	Trp	Thr	Arg	Arg	Gly	Lys	Asp	Ile	Val	Val	Val	Arg	Pro
	195						200				205				
Arg	Lys	Gly	Phe	Cys	Lys	Leu	Ala	Leu	Gln	His	Asn	Cys	Pro	Ile	Val
	210					215					220				
Pro	Val	Tyr	Ala	Phe	Gly	Glu	Asn	Asp	Leu	Tyr	Arg	Thr	Phe	Asn	His
	225				230				235					240	
Leu	Lys	Asp	Phe	Gln	Leu	Trp	Val	Ala	Ser	Ala	Phe	Lys	Leu	Ala	Phe
			245						250					255	
Pro	Pro	Cys	Trp	Gly	Val	Leu	Phe	Leu	Pro	Phe	Leu	Pro	Leu	Pro	Val
		260						265					270		
Ser	Ile	Thr	Val	Val	Met	Gly	Glu	Pro	Leu	Leu	Pro	Arg	Ala	Gln	Lys
	275					280						285			
Gly	Ser	Ala	Arg	Arg	Ser	Gly	Gly	Gly	Lys	Gly	Val	Glu	Pro	Thr	Arg
	290					295					300				
Glu	Glu	Val	Glu	Glu	Leu	His	Phe	Arg	Tyr	Val	Glu	Ala	Leu	Gln	Lys
	305				310				315					320	
Leu	Phe	Asp	Ala	His	Lys	Val	Arg	Gln	Gly	Gly	Arg	Ser	Glu	Glu	Ala
				325					330					335	
Thr	Leu	Val	Val	Lys											
		340													

<210> SEQ ID NO 18
 <211> LENGTH: 1100
 <212> TYPE: DNA
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 18

atthtcagca aagtaatcaa gataataaac aaaaacaatc ctataaagga aaaacaacag	60
aacgatggca cctcctccac cggccccgcc acctgcaccc gagaaccctt acaacctatt	120
gccaccaag cgccccaatc cgcagtactg gcggtatgca agccttgccg ccttccttct	180
cacttgcttc ctggcccctt ccagtaactc gtgggccacc accctccgcc ggcctgctg	240
ggcggcgtac tggacgacct acctggacac aagctataag gacggctcac ggcctggcc	300

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ctggtttcag cgattgCGaa tctggcgtat gtattgcggc tatttgcagg gcaaagtcac 360
ttgcacggtg cecttggacc eggcgCagca atttatcttc gcggcccatc cccaecggcat 420
tggtacctgg aaccatttcc tgaccatgac tgacggctgt cgatttctct cctcctccta 480
ccccgccecg cggtctgacc tgggtgCgac agtacttttc ttcacccct tcttaaagga 540
aattctgctt tggctaggct gtgtggatgc tggagcggcc acggctcatg cggttttggc 600
gCGgggttac tctcctca tttacatcgg tggagaaaa gagcagattt ggacacggcg 660
aggcaaaGac atcgTggtg tacgtcccc caagggtttt tgcaagctgg cctccagca 720
taactgcccc atcgTaccgg tctacgatt tgggGaaac gatctgTatc gcacgttcaa 780
ccacctcaag gacttccagc tgtgggtggc tagcgccttc aagctcgctt tctcctctg 840
ttggggcgtc ctcttctcc ccttctccc cctccccgtc tctatcagc tggTgatggg 900
cgagcccttg ctaccagag caaaaaagg aagtgcgaga aggagtggTg gagGaaagg 960
ggtggagccg acgagggagg aggtggagga gctgcacttc cgatacgtgg aggccttgca 1020
gaagtgttt gacgcacaca aagtcaggca gggagggagg agcgaagagg ccaccttagt 1080
ggtcaaatga gGaaacacc 1100

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<210> SEQ ID NO 19

<211> LENGTH: 1206

<212> TYPE: DNA

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 19

```

atgggtctat ttggcagcgg gatcaaggaa aagacggagg ctgagaccgc gcaggtggag 60
cagcaagagc agcgcaagct gaagcaaaa ccttctctac tgcgggagcg caagggaggt 120
aatataacca aggagcccca gacgcccctg agtaatctga ggctgcccg tcccccgacc 180
gaggtggact ggagctcctt cctgagggc agctacacgc gcttcgggca tggcggggac 240
tggTggaCgC taatcaaggg gacgattgcc attttgttca cgtgggggac ctggctggct 300
ggcggcttgt ctccctttg gatgacttgg ttgtatacgc acggatacaa gaggacattc 360
tattcgatca taggccttt gctttaccgc cttttcttgc cgtgccagc ttggcctgga 420
tttgtccgat tcatTTtaaa catggctgga tattttgagg gcggtgcggc gatgtacgtc 480
gaaaactctt tcaaaggccg caatgtgaat ggtcctatca tgttgcccat gcaccccat 540
ggcatcatgc ctactcctt ccttctcaac ggtgcggggc ggatccacgc gcagaaaccg 600
gaggtattcc tccctccaca ctatcaagat atgtctctta aatcgacggg cgtggcggag 660
ccgttgttgt ttcgattcc gtttatttcg geatttctt attttttgg gtgtcgggag 720
cctgcgtcga aggagatgat gcacgacatc ttggggagge aggtgccgtt tgggatcctg 780
gtgggtggct ccgaggaaat cctcctcatg gagtaccaga agGaaacat ctacatcctc 840
gaacgtaaag gttttattaa atacgcctt cagcatggct acaccatcgc cattggctac 900
ctcttcggcg agtccaacct ctaccacacc atcactggg gacgcaagac ccgcctcgcc 960
ctcttcaaaa aattcaagat tccgttatt ttggcttggg gacgttggtt ctttccctta 1020
ctccctgagc gagcagcgc tttgaatgct gtcgttgga accctattga tttgccagc 1080
atagccaacc caagccaggc ggacattgac aaataccatg cgatgtacat tgagaaattg 1140
acagatttgt ttgaacggaa taaggcggcc tttgggtatt cagatcggac gttgaatttc 1200

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tttttag

1206

<210> SEQ ID NO 20

<211> LENGTH: 401

<212> TYPE: PRT

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 20

Met Gly Leu Phe Gly Ser Gly Ile Lys Glu Lys Thr Glu Ala Glu Thr
 1 5 10 15

Ala Gln Val Glu Gln Gln Glu Gln Ala Lys Leu Lys Gln Lys Pro Ser
 20 25 30

Leu Leu Arg Glu Arg Lys Gly Gly Asn Ile Thr Lys Glu Pro Gln Thr
 35 40 45

Pro Ser Ser Asn Leu Arg Pro Ala Arg Ser Pro Thr Glu Val Asp Trp
 50 55 60

Ser Ser Phe Pro Glu Gly Ser Tyr Thr Arg Phe Gly His Gly Gly Asp
 65 70 75 80

Trp Trp Thr Leu Ile Lys Gly Thr Ile Ala Ile Leu Phe Thr Trp Gly
 85 90 95

Thr Trp Leu Ala Gly Gly Leu Ser Pro Phe Trp Met Thr Trp Leu Tyr
 100 105 110

Thr His Gly Tyr Lys Arg Thr Phe Tyr Ser Ile Ile Gly Pro Leu Leu
 115 120 125

Tyr Pro Leu Phe Leu Pro Val Pro Ala Trp Pro Gly Phe Val Arg Phe
 130 135 140

Ile Leu Asn Met Ala Gly Tyr Phe Glu Gly Gly Ala Ala Met Tyr Val
 145 150 155 160

Glu Asn Ser Phe Lys Gly Arg Asn Val Asn Gly Pro Ile Met Leu Ala
 165 170 175

Met His Pro His Gly Ile Met Pro His Ser Phe Leu Leu Asn Gly Ala
 180 185 190

Gly Arg Ile His Ala Gln Lys Pro Glu Val Phe Leu Pro Pro His Tyr
 195 200 205

Gln Asp Met Ser Leu Lys Ser Thr Gly Val Ala Glu Pro Leu Leu Phe
 210 215 220

Arg Ile Pro Phe Ile Ser Ala Phe Leu Tyr Phe Phe Gly Cys Ala Glu
 225 230 235 240

Pro Ala Ser Lys Glu Met Met His Asp Ile Leu Gly Arg Gln Val Pro
 245 250 255

Phe Gly Ile Leu Val Gly Gly Ser Glu Glu Ile Leu Leu Met Glu Tyr
 260 265 270

Gln Lys Glu Asn Ile Tyr Ile Leu Glu Arg Lys Gly Phe Ile Lys Tyr
 275 280 285

Ala Leu Gln His Gly Tyr Thr Ile Ala Ile Gly Tyr Leu Phe Gly Glu
 290 295 300

Ser Asn Leu Tyr His Thr Ile Thr Trp Gly Arg Lys Thr Arg Leu Ala
 305 310 315 320

Leu Phe Lys Lys Phe Lys Ile Pro Leu Phe Leu Ala Trp Gly Arg Trp
 325 330 335

Phe Phe Pro Leu Leu Pro Glu Arg Ala Ala Pro Leu Asn Ala Val Val
 340 345 350

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Gly Asn Pro Ile Asp Leu Pro Arg Ile Ala Asn Pro Ser Gln Ala Asp
 355 360 365

Ile Asp Lys Tyr His Ala Met Tyr Ile Glu Lys Leu Thr Asp Leu Phe
 370 375 380

Glu Arg Asn Lys Ala Ala Phe Gly Tyr Ser Asp Arg Thr Leu Asn Phe
 385 390 395 400

Phe

<210> SEQ ID NO 21

<211> LENGTH: 1772

<212> TYPE: DNA

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 21

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atattcagca aagtaatcaa gataataaac aaaaacaatc ctataaagga aaaacaacag   60
acatcaacac aggtacttgc agccaccact gcagcaatta tagcaccatc acgaccacta   120
tgggtctatt tggcagcggg atcaaggaaa agacggaggc tgagaccgcg cagggtggagc   180
agcaagagca ggcgaagctg aagcaaaaac cttctctact gcgggagcgc aaggagggta   240
atataaccaa ggagccccag acgcctctga gtaatctgag gcctgcccgt tccccgaccg   300
aggtggactg gagctccttc cctgagggca gctacacgcg cttcgggcat ggcggggact   360
gggtggcgtc aatcaagggg acgattgcca ttttgttcac gtgggggacc tggctggctg   420
gcggtttgtc tcccttttgg atgacttggg tgtatacgca cggatacaag aggacattct   480
attcgatcat aggccctttg ctttaccgcg tttcttgcc cgtgccagct tggcctggat   540
ttgtccgatt cattttaaac atggctggat attttgaggg cgggtcggcg atgtacgtcg   600
aaaactcttt caaagccgcg aatgtgaatg gtcctatcat gttggccatg caccctcatg   660
gcatcatgcc tcactcttct cttctcaacg gtgcccggcg gatccacgcg cagaaaccgg   720
aggtattcct cctccacac tatcaagata tgtctcttaa atcgacgggc gtggcgggagc   780
cgttgttgtt tcgatttccg tttatttcgg catttcttta ttttttggg tgtgcccggagc   840
ctgcgtcgaa ggagatgatg cagcacatct tggggaggca ggtgccgttt gggatcctgg   900
tgggtggctc cgaggaaatc ctcctcatgg agtaccagaa ggaaaacatc tacatcctcg   960
aacgtaaagg ttttattaaa tacgcccttc agcatggcta caccatcgcc attggtacc   1020
tcttcggcga gtccaacctc taccacacca tcacctgggg acgcaagacc cgcctcgccc   1080
tctcaaaaa attcaagatt ccgttatttt tggcttgggg acgttggttc tttcccttac   1140
tccctgagcg agcagcgctt ttgaatgctg tegtggcaa cctattgat ttgccagga   1200
tagccaacc aagccaggcg gacattgaca aataccatgc gatgtacatt gagaaattga   1260
cagatttgtt tgaacggaat aaggcggcct ttgggtatc agatcggacg ttgaatttct   1320
tttaggtggg tgggaggaaa ggagggtaag agggagggtg ggaaggtgtg tgtagggggt   1380
gagtgttcag gcattgttgt tcaggcatgg aaagagactg acccaaccaa ctgaaaagga   1440
gatagacaag caagcacacc atggggcaca tgatcgtgat tagagagaag atgggcaaga   1500
gggagggact gatccggtgt aaatatagac acatgactga atgaagaagc aaggagagaa   1560
tggagaggaa tcagcagcag cagcagcagc agcagcagag aacaatagct cttaaggcag   1620
cagctacaac aatcaaaaac cgaacaagag cgaaaagtcc aaacgctaag attcgacacg   1680

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gagaacaaga acgaagaacg gtgatatcaa cagggataaa ttgtacgaac gaagcatgag 1740
tctagtgaaa acaacaaaaa aaaacaaaaa aa 1772

<210> SEQ ID NO 22
<211> LENGTH: 1173
<212> TYPE: DNA
<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 22
atgttgagta tccccgagtc gtctctgccc ctctcggacc ggactctggt gaagaatgga 60
ggcaaggaga cagagctttc cacgccggtc accgctccca cttcggaccg ctccgctacc 120
tacagtgatg gctattcgac cccaagtcc tacacattgg aggtcgcatt caaatattat 180
aagcgggtat gcatgctgta tgacgtgtgg acacgcacac aggggtgcatt tgctcttctc 240
atgctctggg gcgtctggct tgccgggtcc tttctgtgt tttggtgccc ctatttagta 300
gtgaaggggt attatactgc tgccctagct atggcagtga tcatggcata tccgtatgtg 360
gtcaagggtca agcaaaagccc ggcatttatt cgcttcatct tgagcggcgc gggatggttt 420
aagggcggga cgtgtttgta tttggaggag tcgatgaagc agatcgacac cagcaggtct 480
gtcctctctc gtcagcatcc gcatggtctc ttcacctatg gcttcatcca aaacgggtct 540
gctgcccgca tcgatgcccg caaacccgag gtttatgtgc ctgccgcatt tcgtcacatg 600
aaacccaacg ccaaggcctt cgtggaacct ttgctattca aaatcccgtc tatccgtcac 660
tttatcaccg ccttcggcaa cgccgccccg ggcacaaaa aagagatgca ccgtctcatg 720
tccactaaaa tccccctggg gctgttaccg ggtgggtcgg aagagatcat cttaagccac 780
catggccatg agcgggtgta catcctcaaa cggaaaggct tcctcaagta cgcattacaa 840
catggttaca cgatttgcat tggttacaca ttcggggagt ccgactcgta ccgcacctg 900
gactggggcg tgaagtcttc tacgtgttac ctgaagacct tccgcgttcc actctttgcg 960
tgctggggga cgtggtggtg ccccctcttg ccacggggga aggtggcgcct tgagacagtc 1020
gttgggaacc catttcggtt gcccaagatt gtatgaccca gccaggagga tattgataag 1080
tggcatcggg tgtatgtgca aaaacttgta gatttgttg atcggaacaa ggccaagtcc 1140
gggtatgggg acagggagct ggatttcttt tag 1173

<210> SEQ ID NO 23
<211> LENGTH: 390
<212> TYPE: PRT
<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 23
Met Leu Ser Ile Pro Glu Ser Ser Ser Pro Leu Ser Asp Arg Thr Leu
1 5 10 15
Val Lys Asn Gly Gly Lys Glu Thr Glu Leu Ser Thr Pro Val Thr Ala
20 25 30
Pro Thr Ser Asp Arg Ser Arg Thr Tyr Ser Asp Gly Tyr Ser Thr Pro
35 40 45
Lys Ser Tyr Thr Leu Glu Val Asp Pro Lys Phe Tyr Lys Arg Val Cys
50 55 60
Asp Ala Asp Asp Val Trp Thr Arg Thr Gln Gly Ala Phe Ala Leu Leu
65 70 75 80
Met Leu Trp Gly Val Trp Leu Ala Gly Ser Phe Ser Val Phe Trp Trp

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85					90					95						
Pro	Tyr	Leu	Val	Val	Lys	Gly	Tyr	Tyr	Thr	Ala	Ala	Ala	Leu	Ala	Met	Ala
			100					105						110		
Val	Ile	Met	Ala	Tyr	Pro	Tyr	Val	Val	Lys	Val	Lys	Gln	Ser	Pro	Ala	
		115					120					125				
Phe	Ile	Arg	Phe	Ile	Leu	Ser	Gly	Ala	Gly	Trp	Phe	Lys	Gly	Gly	Thr	
	130					135					140					
Cys	Leu	Tyr	Leu	Glu	Glu	Ser	Met	Lys	Gln	Ile	Asp	Thr	Ser	Glu	Ser	
	145					150					155				160	
Val	Leu	Leu	Cys	Gln	His	Pro	His	Gly	Leu	Phe	Thr	Tyr	Gly	Phe	Ile	
				165					170					175		
Gln	Asn	Gly	Ser	Ala	Ala	Arg	Ile	Asp	Ala	Arg	Lys	Pro	Glu	Val	Tyr	
			180					185					190			
Val	Pro	Ala	Ala	Phe	Arg	His	Met	Lys	Pro	Asn	Ala	Lys	Ala	Phe	Val	
		195					200					205				
Glu	Pro	Leu	Leu	Phe	Lys	Ile	Pro	Leu	Ile	Arg	His	Phe	Ile	Thr	Ala	
	210					215					220					
Phe	Gly	Asn	Ala	Ala	Pro	Ala	Thr	Lys	Lys	Glu	Met	His	Arg	Leu	Met	
	225					230					235				240	
Ser	Thr	Lys	Ile	Pro	Leu	Gly	Leu	Leu	Pro	Gly	Gly	Ser	Glu	Glu	Ile	
				245					250					255		
Ile	Leu	Ser	His	His	Gly	His	Glu	Arg	Val	Tyr	Ile	Leu	Lys	Arg	Lys	
			260					265						270		
Gly	Phe	Leu	Lys	Tyr	Ala	Leu	Gln	His	Gly	Tyr	Thr	Ile	Cys	Ile	Gly	
		275					280					285				
Tyr	Thr	Phe	Gly	Glu	Ser	Asp	Ser	Tyr	Arg	Thr	Leu	Asp	Trp	Gly	Val	
	290					295					300					
Lys	Phe	Arg	Thr	Trp	Tyr	Leu	Lys	Thr	Phe	Arg	Val	Pro	Leu	Phe	Ala	
	305					310					315				320	
Cys	Trp	Gly	Thr	Trp	Trp	Cys	Pro	Leu	Leu	Pro	Arg	Gly	Lys	Val	Ala	
				325					330					335		
Leu	Glu	Thr	Val	Val	Gly	Asn	Pro	Phe	Arg	Leu	Pro	Lys	Ile	Val	Asp	
			340					345					350			
Pro	Ser	Gln	Glu	Asp	Ile	Asp	Lys	Trp	His	Ala	Val	Tyr	Val	Gln	Lys	
		355					360					365				
Leu	Val	Asp	Leu	Phe	Asp	Arg	Asn	Lys	Ala	Lys	Phe	Gly	Tyr	Gly	Asp	
	370					375					380					
Arg	Glu	Leu	Asp	Phe	Phe											
	385					390										

<210> SEQ ID NO 24

<211> LENGTH: 1239

<212> TYPE: DNA

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 24

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attttcagca aagtaatcaa gataataaac aaaaacaatc ctataaaagg aaaaacaaca    60
ggtagaatgt tgagatcccc cgagtcgtcc tcgcccctct cggaccggac tctggtgaag    120
aatggaggca aggagaccga gctttccacg cgggtcaccg ctcccacttc ggaccgctcg    180
cgtacctaca gtgatggcta ttcgaccccc aagtectaca cattggaggt cgateccaaa    240
ttttataagc gggtatgcga tgctgatgac gtgtggacac gcacacaggg tgcatttgct    300

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cttctcatgc tctggggcgt ctggettgcc gggtcctttt ctgtgttttg gtggccctat 360
ttagtagtga aggggtatta tactgctgcc ctagnetatgg cagtgatcat ggcatatccg 420
tatgtggtca aggtcaagca aagcccggca tttattcgct tcatcttgag cggcgcggga 480
tggtttaagg gcgggacgtg tttgtatttg gaggagtcca tgaagcagat cgacaccagc 540
gagtcgtgcc tctctgtca gcatccgcat ggtctcttca cctatggctt catccaaaac 600
gggtctgctg cccgcacgca tgcccgcgaaa cccgagggtt atgtgcctgc cgcatttcgt 660
cacatgaaac ccaacgcaa ggccttcgtg gaacctttgc tattcaaaat cccgcttacc 720
cgtoacttta tcaccgcctt cggcaacgcc gccccggcga ccaaaaaaga gatgcaccgt 780
ctcatgtcca ctaaaattcc cctggggctg ttaccgggtg ggtcggaaga gatcatctta 840
agccaccatg gccatgagcg ggtgtacatc ctcaaacgga aaggcttccct caagtacgca 900
ttacaacatg gctacacgat ttgcattggt tacacattcg gggagtccga ctcgtaccgc 960
accttgact ggggctgtaa gtttcgtacg tggtaacctga agaccttccg cgttccactc 1020
tttgcgtgct gggggacgtg gtggtgcccc ctcttgccac gggggaagggt ggcgcttgag 1080
acagtcgttg ggaaccattc tcggttgccc aagattgtag atccgagcca ggaggatatt 1140
gataagtggc atgcccgtga tgtgcaaaaa cttgtagatt tgtttgatcg gaacaaggcc 1200
aagttcgggt atggggacag ggagctggat ttcttttag 1239

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<210> SEQ ID NO 25

<211> LENGTH: 1089

<212> TYPE: DNA

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 25

```

atgacgcgcg aagccgatat caccagcaag acgacatcca accccaagac ggetgcatcc 60
tccccctcca agacctcgcc ccccgcggtt caatacaaaag cagggaatgg caaggtgatc 120
acggtggcca tggccgagca agacgacggg aacatgggca ttttccgca gtgttggtgcg 180
atggtgacaa tggggataat catgtcgtgg tactacatcg tcgtcgttct ctccctcctg 240
tgcttggtgg ggatctcctt ctccctgcc tgccgggccc tggcggcgac ggtttttgta 300
ctcatgtgga gtgcggcgct tttgcgcctc gactaccagg ggtgggacgc tttctgcaac 360
tcattgatct tcaggctgtg gcgggactac ttccactacg aatacgtcct ggaagaaatg 420
atcgacceca acaagcgtca cctcttcgct gagatgcccc acggaatcct cccctgggga 480
gaggtgattt ccaattctat caccaaagcag cttttccccg ggagccgctg cggctccatt 540
gggtgcgagt teatcttctt ccttccgggc ctccggcaact tcttcgctg gatcgggtgt 600
cggccccgca gcccgagaaa tatcaaaaag atttttgatg atgggcagga ttgtgccgtg 660
acggtgggag gggtcgcccga gatgtttctg gttggaggag agaaggagcg gctctacctc 720
aaaaagcaca agggtttctg tcgagagggc atgaagaacg gcgcggaact ggtccctgtc 780
ttctgcttcg gcaacagcaa gttgttcaat gtgggtggggg agagcagtcg ggtgtccatg 840
ggcctgatga agcgtctctc gaggaggctc aaagccagcg tctctatctt ctacggccgt 900
ctcttcttac ccattccgat ccgcccaccg ctcttgctcg tgggtggaaa gccctgcccg 960
gtcgtgcaga atgcagagcc gaccaaggag gagatcgccg cgacgcacgc actcttttgc 1020
gagaagggtg aggagcttta ctacaaatc aggccggaat gggagacgcg cccgttgctc 1080

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attgagtaa

1089

<210> SEQ ID NO 26

<211> LENGTH: 362

<212> TYPE: PRT

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 26

Met Thr Pro Gln Ala Asp Ile Thr Ser Lys Thr Thr Ser Asn Pro Lys
 1 5 10 15

Thr Ala Ala Ser Ser Pro Ser Lys Thr Ser Pro Pro Ala Val Gln Tyr
 20 25 30

Lys Ala Gly Asn Gly Lys Val Ile Thr Val Ala Met Ala Glu Gln Asp
 35 40 45

Asp Gly Asn Met Gly Ile Phe Arg Glu Cys Cys Ala Met Val Thr Met
 50 55 60

Gly Ile Ile Met Ser Trp Tyr Tyr Ile Val Val Val Leu Ser Leu Leu
 65 70 75 80

Cys Leu Val Gly Ile Ser Phe Phe Pro Ala Trp Arg Ala Val Ala Ala
 85 90 95

Thr Val Phe Val Leu Met Trp Ser Ala Ala Leu Leu Pro Leu Asp Tyr
 100 105 110

Gln Gly Trp Asp Ala Phe Cys Asn Ser Cys Ile Phe Arg Leu Trp Arg
 115 120 125

Asp Tyr Phe His Tyr Glu Tyr Val Leu Glu Glu Met Ile Asp Pro Asn
 130 135 140

Lys Arg Tyr Leu Phe Ala Glu Met Pro His Gly Ile Phe Pro Trp Gly
 145 150 155 160

Glu Val Ile Ser Ile Ser Ile Thr Lys Gln Leu Phe Pro Gly Ser Arg
 165 170 175

Val Gly Ser Ile Gly Ala Ser Val Ile Phe Leu Leu Pro Gly Leu Arg
 180 185 190

His Phe Phe Ala Trp Ile Gly Cys Arg Pro Ala Ser Pro Glu Asn Ile
 195 200 205

Lys Lys Ile Phe Asp Asp Gly Gln Asp Cys Ala Val Thr Val Gly Gly
 210 215 220

Val Ala Glu Met Phe Leu Val Gly Gly Glu Lys Glu Arg Leu Tyr Leu
 225 230 235 240

Lys Lys His Lys Gly Phe Val Arg Glu Ala Met Lys Asn Gly Ala Asp
 245 250 255

Leu Val Pro Val Phe Cys Phe Gly Asn Ser Lys Leu Phe Asn Val Val
 260 265 270

Gly Glu Ser Ser Arg Val Ser Met Gly Leu Met Lys Arg Leu Ser Arg
 275 280 285

Arg Leu Lys Ala Ser Val Leu Ile Phe Tyr Gly Arg Leu Phe Leu Pro
 290 295 300

Ile Pro Ile Arg His Pro Leu Leu Phe Val Val Gly Lys Pro Leu Pro
 305 310 315 320

Val Val Gln Asn Ala Glu Pro Thr Lys Glu Glu Ile Ala Ala Thr His
 325 330 335

Ala Leu Phe Cys Glu Lys Val Glu Glu Leu Tyr Tyr Lys Phe Arg Pro
 340 345 350

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Glu Trp Glu Thr Arg Pro Leu Ser Ile Glu
 355 360

<210> SEQ ID NO 27
 <211> LENGTH: 1609
 <212> TYPE: DNA
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 27

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attttcagca aagtaatcaa gataataaac aaaaacaatc ctataaagga aaaacaacag    60
agagacaagt aggccaccag cattggtttc caccatgacg cgcgaagccg atatcaccag    120
caagacgaca tccaacccca agacggctgc atcctcccc tccaagacct cgccccccgc    180
cgttcaatac aaagcaggga atggcaaggt gatcacggtg gccatggccg agcaagacga    240
cgggaacatg ggcattttcc gcgagtgttg tgcgatggtg acaatgggga taatcatgtc    300
gtggtactac atcgtcgtcg ttctctccct cctgtgcttg gtggggatct ccttcttccc    360
tgcctggcgg geggtggcgg gcacggtttt tgtactcatg tggagtgcgg cgcttttgcc    420
gctcgactac caggggtggg acgctttctg caactcatgt atcttcaggc tgtggcggga    480
ctacttccac tacgaatacg tccctggaaga aatgatcgac cccaacaagc gctacctctt    540
cgctgagatg ccccacggaa tcttcccctg gggagagggt atttccattt ctatcaccaa    600
gcagcttttc cccgggagcc gcgtcgctc cattgggtgcg agtgtcatct tcctccttcc    660
gggcctccgg cacttcttcg cctggatcgg gtgtcggccc gcgagcccg agaatatcaa    720
aaagattttt gatgatgggc aggattgtgc cgtgacggtg ggaggggtcg ccgagatgtt    780
tctggttga ggagagaagg agcggctcta cctaaaaaag cacaagggtt tegtctgaga    840
ggccatgaag aacggcgcgg acctggctcc tgtcttctgc ttggcaaca gcaagtgtt    900
caatgtggtg ggggagagca gtcgggtgtc catgggcctg atgaagcgtc tctcgaggag    960
gctcaaagcc agcgtcctca ttttctacgg ccgtctcttc ctaccattc cgatccgcca   1020
cccgtcttg ttcgtggtgg gaaagccctt gccggctcgt cagaatgcag agccgaccaa   1080
ggaggagatc gcggcgacgc acgcactctt ttgcgagaag gtggaggagc tttactacaa   1140
attcaggccg gaatgggaga cgcgccggtt gtccattgag taaaatacgt ggacggagaa   1200
agcagggggc gtgtgtttga gtatctgatt gtgattgtga ttgtctgtgt ctgcacgtgt   1260
gtgtgtacga ttactctggt tgcctgtgcg gttttgaaag taactgtaaa ggtcagaaga   1320
gattagaaga cgagacttgg atacgatgaa ggggtgaagaa gaaatttaa acaattttga   1380
gattttatc atgtctgagg aataaatgta gatgtagaa aatttgaggt agttctcggt   1440
acttgcccc tatcatccgt gtttagtaac gaggtacatc cgtgcgacgg gtcggtgaa   1500
gtagccagcg teatcagaga gaggtctcac acacgatcgt gtgtccttgc acatgtcttt   1560
tccatttaac acgaattact tttttttaa aaaaataata aaaaaata   1609

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<210> SEQ ID NO 28
 <211> LENGTH: 1464
 <212> TYPE: DNA
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 28

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atggcttacc tcttccgctc tcgaagcaaa ggcgagggca acagcactag cagcagctgc    60

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tcttctctgt cggaagataa taagggcacg tccatccact cttecgaaat cgagccgcgc 120
gctcccgccca cgtccaaagc caccgacaagc agcataaagg agattgggaa gccctcattg 180
cccaccgcgc cacatttatac accaccagc ataagcaagg cagatagaaa tttcgccatt 240
gccgcagtag cagcaggagc actggagggg gctgcagcag gcgccgtgac agcaccaccc 300
accgaccaat ctccgaagaa gcagtacggg cagggtggta ctggggagcg agggaaggag 360
gcagaagtg gacgagaacg aagtgaagc gtcggcaacc tttactgtc atcaattaat 420
tcgttttcaa gctgcacgct cctatccttt ttggccggcg aggacgagac cccgtctcct 480
cccgagacag ggccgtctgg gattgatttc tcgacaccgg ctcacccgac catgcaactt 540
gtggacttca tcataccttt tctcttggtg cattatattc aagtcttcta ctccctagtc 600
ctcctcttca tctacctcgt caagcacggc cacagatggc cgtacctct cgctgccatc 660
tacgcccctt cgtacttcat tcttttacag cgattgggcg gatggccgtt caaaggatte 720
atgcgtcggc ccttttggcg gtgtgtccaa aggaccttag ctctccaggt gaaagagag 780
gtcgagctgc gtcagacga acagtacatt tttggtggc acccccacgg gatcttgctc 840
ttgtcccggc ttgcaatcta tgggggtctg tgggaaaagc tttttccggg tattcatttc 900
aagacgctag cggcaagtcc tctgttttg attccaccta ttcgcgaagt gtcgatcttg 960
ctgggtgggg tggatgcagg caggggcatca gcagcacggg cactcacaga cggctactcc 1020
gtctctcttt atccgggggg aagcaaggaa atctacacca ctgatcccta cactcctgaa 1080
acgaccctgg tctgaaaaat ccgcaaagc ttcattcgca tggccctccg ctatggctgt 1140
ccactcgtgc ctgtgtacac gtttgagaa aaatacgcct accatcggct agggccggcc 1200
acgggctttg cgcgctggct gttggcagtg ctgaaagtcc ctttcttgat cttttgggga 1260
cgatggggca cattcatgcc gctcaaggag acgcaggtgt cagtgggtgt gggcaagcca 1320
ctgcgcgtgc ccaaaatcga tggagatcct gccctgagg tgggtggagga atggttgac 1380
agatactgcg acgaagtcca ggcgttgctc cagcgacaca agaacaaata cgcaaagcct 1440
gaggagtcca ttgcatcgc ctaa 1464

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<210> SEQ ID NO 29

<211> LENGTH: 487

<212> TYPE: PRT

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 29

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Met Ala Tyr Leu Phe Arg Arg Arg Ser Lys Gly Glu Gly Asn Ser Thr
1           5           10          15
Ser Ser Ser Cys Ser Ser Leu Ser Glu Asp Asn Lys Gly Thr Ser Ile
20          25          30
His Ser Ser Glu Ile Glu Pro Arg Ala Pro Ala Thr Ser Lys Ala Thr
35          40          45
Thr Ser Ser Ile Lys Glu Ile Gly Lys Pro Ser Leu Pro Thr Ala Ala
50          55          60
His Leu Ser Pro Pro Ser Ile Ser Lys Ala Asp Arg Asn Phe Ala Ile
65          70          75          80
Ala Ala Val Ala Ala Gly Ala Leu Glu Gly Ala Ala Ala Gly Ala Val
85          90          95
Thr Ala Pro Pro Thr Asp Gln Ser Pro Lys Lys Gln Tyr Gly Gln Gly
100         105         110

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Gly Thr Gly Glu Arg Gly Lys Glu Ala Glu Gly Gly Arg Glu Arg Ser
 115 120 125
 Gly Ser Val Gly Asn Leu Leu Leu Ser Ser Ile Asn Ser Phe Ser Ser
 130 135 140
 Cys Thr Ser Leu Ser Phe Leu Ala Gly Glu Asp Glu Thr Pro Ser Pro
 145 150 155 160
 Pro Glu Thr Gly Pro Ala Gly Ile Asp Phe Ser Thr Pro Ala His Pro
 165 170 175
 Thr Met Gln Leu Val Asp Phe Ile Ile Thr Phe Leu Leu Val His Tyr
 180 185 190
 Ile Gln Val Phe Tyr Ser Leu Val Leu Leu Phe Ile Tyr Leu Val Lys
 195 200 205
 His Gly His Arg Trp Pro Tyr Leu Leu Ala Ala Ile Tyr Ala Pro Ser
 210 215 220
 Tyr Phe Ile Pro Leu Gln Arg Leu Gly Gly Trp Pro Phe Lys Gly Phe
 225 230 235 240
 Met Arg Arg Pro Phe Trp Arg Cys Val Gln Arg Thr Leu Ala Leu Gln
 245 250 255
 Val Glu Arg Glu Val Glu Leu Arg Pro Asp Glu Gln Tyr Ile Phe Gly
 260 265 270
 Trp His Pro His Gly Ile Leu Leu Leu Ser Arg Phe Ala Ile Tyr Gly
 275 280 285
 Gly Leu Trp Glu Lys Leu Phe Pro Gly Ile His Phe Lys Thr Leu Ala
 290 295 300
 Ala Ser Pro Leu Phe Trp Ile Pro Pro Ile Arg Glu Val Ser Ile Leu
 305 310 315 320
 Leu Gly Gly Val Asp Ala Gly Arg Ala Ser Ala Ala Arg Ala Leu Thr
 325 330 335
 Asp Gly Tyr Ser Val Ser Leu Tyr Pro Gly Gly Ser Lys Glu Ile Tyr
 340 345 350
 Thr Thr Asp Pro Tyr Thr Pro Glu Thr Thr Leu Val Leu Lys Ile Arg
 355 360 365
 Lys Gly Phe Ile Arg Met Ala Leu Arg Tyr Gly Cys Pro Leu Val Pro
 370 375 380
 Val Tyr Thr Phe Gly Glu Lys Tyr Ala Tyr His Arg Leu Gly Pro Ala
 385 390 395 400
 Thr Gly Phe Ala Arg Trp Leu Leu Ala Val Leu Lys Val Pro Phe Leu
 405 410 415
 Ile Phe Trp Gly Arg Trp Gly Thr Phe Met Pro Leu Lys Glu Thr Gln
 420 425 430
 Val Ser Val Val Val Gly Lys Pro Leu Arg Val Pro Lys Ile Asp Gly
 435 440 445
 Asp Pro Ala Pro Glu Val Val Glu Glu Trp Leu His Arg Tyr Cys Asp
 450 455 460
 Glu Val Gln Ala Leu Phe Gln Arg His Lys Asn Lys Tyr Ala Lys Pro
 465 470 475 480
 Glu Glu Phe Ile Ala Ile Ala
 485

<210> SEQ ID NO 30

<211> LENGTH: 1682

-continued

<212> TYPE: DNA

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 30

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attttcagca aaagtaatca agataataaa caaaaacaat cctataaagg aaaaacaaca    60
gggacaccag ggtgacgccc gcgaccccaa cactatggct tacctcttcc gtcgtcgaag    120
caaaggcgag ggcaacagca cttagcagcag ctgctcttct ctgtcgggaag ataataaggg    180
cacgtccatc cactcttccg aaatcgagcc gcgcgctccc gccacgtcca aagccacgac    240
aagcagcata aaggagattg ggaagccctc attgcccacc gccgcacatt tatcaccacc    300
cagcataagc aaggcagata gaaatcttgc cattgcccga gtagcagcag gagcactgga    360
gggggctgca gcaggcgccc tgacagcacc acccaccgac caatctccga agaagcagta    420
cgggcagggt ggtactgggg agcgagggaa ggaggcagaa ggtggacgag aacgaagtgg    480
aagcgtcggc aaccttttac tgatcaaat taattcgttt tcaagctgca cgtccctatc    540
ctttttggcc ggcgaggacg agaccccgtc tctctccgag acagggcctg ctgggattga    600
tttctcgaca ccggctcctc cgaccatgca acttgtggac ttcacatca cttttctctt    660
ggtgcattat attcaagtct tctactcctc agtccctcct ttcacatcacc tcgtcaagca    720
cggtcacaga tggccgtacc tctctgctgc catctacgcc cctctgact tcattccttt    780
acagcgattg ggcggatggc cgttcaaagg attcattgct cggccctttt ggcggtgtgt    840
ccaaaggacc ttactctctc aggtggaag agaggtcgag ctgctccag acgaacagta    900
catttttggg tggcaccctc acgggatctt gctcttgctc cggtttgcaa tctatggggg    960
tctgtgggaa aagcttttct cgggtattca tttcaagacg ctagcggcaa gtcctctgtt   1020
ttggattcca cctattcgcg aagtgtcgat cttgtctggg ggggtggatg caggcagggc   1080
atcagcagca cgggcactca cagacggeta ctccgtctct ctttatccgg ggggaagcaa   1140
ggaaatctac accactgato cctacactcc tgaaacgacc ctggctctga aaatccgcaa   1200
aggettcatt cgcattggccc tccgctatgg ctgtccactc gtgctctgtg acacgtttgg   1260
agaaaaatac gcctaccatc ggctagggcc ggccacgggc tttgcgcgct ggctgttggc   1320
agtgtgaaa gtccctttct tgatcttttg gggacgatgg ggcacattca tgccgctcaa   1380
ggagacgcag gtgtcagtgg ttgtgggcaa gccactgcgc gtgccccaaa tcgatggaga   1440
tctgcccctc gaggtggtgg aggaatgggt gcacagatac tgcgacgaag tccaggcgtt   1500
gttcacagca cacaagaaca aatacgcgaa gcctgaggag ttcattgaga tcgcctaaaa   1560
gggaaaaaaaa gtaaaacctc tccctccctt ccttctctct tttattacac atgcccttgc   1620
accaaccacg cgacatgagg ggacggaagg agctggatgc ggtgtggttt gtctgttcag   1680
ga                                                                                   1682

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<210> SEQ ID NO 31

<211> LENGTH: 1539

<212> TYPE: DNA

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 31

```

atgccttttg gaeggctgc atcagcctgg atttcggcct cagcattggt gccagccttg    60
gcggaaccaa ctttctcttg eggcaccgcc atcgtgggccc tcgtcgttat gtactacatt    120
gtcagcggcc aaaggtgtgc acgagctttg cgtccttccc caggggtgat tcgaaggaaa    180

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atgagttttt gttcggcggc ctgtgcggat ggtcccatgc ctgagcacgc caagatgaac 240
cctgtcgatc ctattatcaa tgccgtgggtg cttttcgagg gggaggcgcc cacgcgtgcg 300
gcggtggaat cggccatctt gccgctcttt gaattcgaac ggtttcgctc ccggaaggtt 360
aagattgggtg atgattggta ttgggaagtg ctgccttctt ttgacgctag gacgcattgtg 420
attgaagact ctttcaaggg tgccagcacc gatgacttgt ttcttcgctt ggagggtgtgg 480
tcccagaaac ccctgcatgt accggtggac gggcccgcct ttgaattgc tttgcttcgg 540
aatcaggata agaagggggc ctctgctgtg atttgcgta tcaaccatgc gattggtgat 600
gggtgtctctc tggccaagtt gatccccac gtgttcaagg acattgacgg ccagtcactg 660
ccgatcgggg agaagtttcg ccggcgggaa gcagggttca agccgacttt ccgcaaccct 720
tttaacctgc tggcttcgct tttcaaggtt ttgggtacgc ctactacggc gtttgatact 780
gacgtggggg tgacgattcc ggataaaaaa aatattacct ttacggggcg tcggtgcatt 840
gtgcgtatcc ccaccgtgaa gctttcgttc atcaagagca ttaaaaatgc ggcgaatgtg 900
actgtgaacg atgtggtgat gagcgcgggt gctggggcgg tgcacgatt tcggtgcgcg 960
caaaaagatc ctgcaatgct cgacccttta tcccattgta aagtccgtac acgcgctttg 1020
atgcctgtgg ctttgcctcg ggaggaggga gatcctgtca aggccttgcg aaacaagtgg 1080
agttttgctt ccgtggcgat gcccgtgggg gtcaagggga gtttgaacg cttgcatgca 1140
gcgaatgcca cgatgactgc gttgaaaaac agtccgatag tgatcgtgca gaatatggtg 1200
gaggctaacc taggggcacg cttgccgtgg acagtggcaa aacaaaccgc gtttgactcg 1260
tttgtgagge acacgtttgt gtttagcaat gtaccgggtc cgaacatgcc tataacattt 1320
gccggtcggg aagtgtcggg actgtatatg gcgtttgca atttgattcc tcagggtggc 1380
gctctgtcct tgaacggcaa gatcttcaac tgtctggtgc tggacgacga ggtcacgccc 1440
ggggcacgtg aactaggaga gcattttatt gacgagtga tggacttggc tcgaaggacg 1500
gggctggaaa atgtaaagaa ggaggatatt ttcggtga 1539

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<210> SEQ ID NO 32

<211> LENGTH: 512

<212> TYPE: PRT

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 32

```

Met Pro Phe Gly Arg Ala Ala Ser Ala Trp Ile Ser Ala Ser Ala Leu
1           5           10          15
Leu Pro Ala Leu Ala Asp Pro Thr Phe Leu Cys Gly Thr Ala Ile Val
20          25          30
Gly Leu Val Val Met Tyr Tyr Ile Val Ser Gly Gln Arg Cys Ala Arg
35          40          45
Ala Leu Arg Pro Ser Pro Gly Val Ile Arg Arg Lys Met Ser Phe Cys
50          55          60
Ser Ala Ala Cys Ala Asp Gly Pro Met Pro Glu His Ala Lys Met Asn
65          70          75          80
Pro Val Asp Pro Ile Ile Asn Ala Val Val Leu Phe Glu Gly Glu Ala
85          90          95
Pro Thr Arg Ala Ala Val Glu Ser Ala Ile Leu Pro Leu Phe Glu Phe
100         105         110

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Glu	Arg	Phe	Arg	Ser	Arg	Lys	Val	Lys	Ile	Gly	Asp	Asp	Trp	Tyr	Trp
	115						120					125			
Glu	Val	Leu	Pro	Ser	Phe	Asp	Ala	Arg	Thr	His	Val	Ile	Glu	Asp	Ser
	130					135					140				
Phe	Lys	Gly	Ala	Ser	Ile	Asp	Asp	Leu	Phe	Leu	Arg	Leu	Glu	Val	Trp
145					150					155					160
Ser	Gln	Lys	Pro	Leu	His	Val	Pro	Val	Asp	Gly	Pro	Ala	Phe	Glu	Phe
				165					170					175	
Ala	Leu	Leu	Arg	Asn	Gln	Asp	Lys	Lys	Gly	Pro	Ser	Ala	Val	Ile	Cys
			180					185					190		
Arg	Ile	Asn	His	Ala	Ile	Gly	Asp	Gly	Val	Ser	Leu	Ala	Lys	Leu	Ile
	195						200					205			
Pro	His	Val	Phe	Lys	Asp	Ile	Asp	Gly	Gln	Ser	Leu	Pro	Ile	Gly	Glu
	210					215					220				
Lys	Phe	Arg	Arg	Arg	Glu	Ala	Gly	Phe	Lys	Pro	Thr	Phe	Arg	Thr	Pro
225					230					235					240
Phe	Thr	Leu	Leu	Ala	Ser	Leu	Phe	Lys	Val	Leu	Gly	Thr	Pro	Thr	Thr
				245					250						255
Ala	Phe	Asp	Thr	Asp	Val	Gly	Leu	Thr	Ile	Pro	Asp	Lys	Lys	Asn	Ile
				260					265					270	
Thr	Phe	Thr	Gly	Arg	Arg	Cys	Ile	Val	Arg	Ile	Pro	Thr	Val	Lys	Leu
			275				280						285		
Ser	Phe	Ile	Lys	Ser	Ile	Lys	Asn	Ala	Ala	Asn	Val	Thr	Val	Asn	Asp
	290					295					300				
Val	Val	Met	Ser	Ala	Val	Ala	Gly	Ala	Val	His	Arg	Phe	Arg	Cys	Ala
305					310					315					320
Gln	Lys	Asp	Pro	Ala	Met	Leu	Asp	Pro	Leu	Ser	His	Cys	Lys	Val	Arg
				325					330					335	
Thr	Arg	Ala	Leu	Met	Pro	Val	Ala	Leu	Pro	Arg	Glu	Glu	Gly	Asp	Pro
				340					345					350	
Val	Lys	Ala	Leu	Arg	Asn	Lys	Trp	Ser	Phe	Ala	Ser	Val	Ala	Met	Pro
		355					360					365			
Val	Gly	Val	Lys	Gly	Ser	Leu	Glu	Arg	Leu	His	Ala	Ala	Asn	Ala	Thr
	370					375					380				
Met	Thr	Ala	Leu	Lys	Asn	Ser	Pro	Ile	Val	Ile	Val	Gln	Asn	Met	Val
385					390					395					400
Glu	Ala	Asn	Leu	Gly	Ala	Arg	Leu	Pro	Trp	Thr	Val	Ala	Lys	Gln	Thr
				405					410					415	
Ala	Phe	Asp	Ser	Phe	Val	Arg	His	Thr	Phe	Val	Phe	Ser	Asn	Val	Pro
				420					425					430	
Gly	Pro	Asn	Met	Pro	Ile	Thr	Phe	Ala	Gly	Arg	Glu	Val	Ser	Gly	Leu
		435					440					445			
Tyr	Met	Ala	Phe	Ala	Asn	Leu	Ile	Pro	Gln	Val	Gly	Ala	Leu	Ser	Leu
	450					455						460			
Asn	Gly	Lys	Ile	Phe	Thr	Cys	Leu	Val	Leu	Asp	Asp	Glu	Val	Thr	Pro
465					470						475				480
Gly	Ala	Arg	Glu	Leu	Gly	Glu	His	Phe	Ile	Asp	Glu	Leu	Met	Asp	Leu
				485					490					495	
Ala	Arg	Arg	Thr	Gly	Leu	Glu	Asn	Val	Lys	Lys	Glu	Asp	Ile	Phe	Gly
				500					505					510	

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<210> SEQ ID NO 33
<211> LENGTH: 1904
<212> TYPE: DNA
<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 33
attttcagca aagtaatcaa gataataaac aaaaacaatc ctataaagga aaaacaacag    60
ccacacagac gccccagctt caactctcca cacacgattt gccagtgagg gtcgtgcacc    120
ctccgcaacc acgagccttt tccacagtag tcatcctgcc catcacgctt aaaatcatgc    180
cttttgacg ggtgcatca gcctggattt cggcctcagc attgttgcca gccttgaggc    240
acccaacttt cctttgcggc accgccatcg tgggcctcgt cgttatgtac tacattgtca    300
gcggccaaag gttgacacga gctttgcgtc cttccccagg ggtgattcga aggaaaatga    360
gtttttgttc ggcggcctgt gcggatggtc ccatgcctga gcacgccaag atgaaccctg    420
tcgatcctat tatcaatgcc tgggtgcttt tcgaggggga ggcgcccacg cgtgcgccgg    480
tggaatcggc catcctgccc ctctttgaat tcgaacgggt tcgctcccgg aaggttaaga    540
ttggtgatga ttggtattgg gaagtgtgct cttcctttga cgtaggacg catgtgattg    600
aagactcttt caagggtgcc agcatcgatg acttgtttct tcgcctggag gtgtggtecc    660
agaaaccctt gcattgaccg ttggacgggc ccgcctttga atttgctttg cttcggaatc    720
aggataagaa ggggcccctc cctgtgattt gtcgtatcaa ccatgcgatt ggtgatggtg    780
tctctctggc caagttgatc ccccacgtgt tcaaggacat tgacggccag tcaactgccg    840
tcggggagaa gtttcgccgg cgggaagcag ggttcaagcc gactttccgc acccctttta    900
ccttgctggc ttcgcttttc aaggtattgg gtacgcctac tacggcgctt gatactgacg    960
tggggttgac gattccggat aaaaagaata ttaccctttac ggggcgctcg tgcaattgtc   1020
gtatcccac cgtgaagctt tcgttcatca agagcattaa aaatgcggcg aatgtgactg   1080
tgaacgatgt ggtgatgagc gcggttgctg gggccgtgca tcgatttcgt tcgcgcgaaa   1140
aagatcctgc aatgctcgac cctttatccc attgtaaagt ccgtacacgc gctttgatgc   1200
ctgtggcttt gccccgggag gagggagatc ctgtcaaggc tttgcgaaac aagtggagt   1260
ttgcttcctg ggcgatgccc tgggggtgca aggggagttt ggaacgcttg catgcagcga   1320
atgccacgat gactgcgttg aaaaacagtc cgatagtgat cgtgcagaat atggtggagg   1380
ctaacctagg ggcacgcttg ccgtggacag tggcaaaaca aaccgcgctt gactcgtttg   1440
tgaggcacac gtttgtggtt agcaatgtac cgggtccgaa catgcctata acatttgccc   1500
gtcgggaagt gtcgggactg tatatggcgt ttgcgaattt gattcctcag gtggcgctc   1560
tgctcctgaa cggcaagatc ttcacctgct tgggtgctgga cgacgaggtc acgcccgggg   1620
cacgtgaact aggagagcat tttattgacg agttgatgga cttggctcga aggacggggc   1680
tggaaaatgt aaagaaggag gatattttcg ggtgagaagc ctagaggaga gagggataga   1740
aggagggaa gatggagatg gttttgtgac atgcgcgtgt cgggtggctgc cgcggctgtc   1800
attggtgagg cgatcggtag ggtaaataga atgaactcat aagagaatga agagtgagaa   1860
agaagagcat ccgtaagcgg gaaacaaaaa aaaaaaaaaa aaaa                    1904

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<210> SEQ ID NO 34
<211> LENGTH: 1083
<212> TYPE: DNA
<213> ORGANISM: Nannochloropsis oculata

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-continued

<400> SEQUENCE: 34

```

atggccaagg ctaacttccc gcccgcgcg cgctatgtta atatgacgca ggtctatgcg    60
acaggcgctc acaatatgcc ggacgaggac cgcgtcaagg tcatgaacgg gctgtccaag    120
cccgtgacgg aggccaaggc aggtgatttg gggtttgggg atgttgagtc catgacggcc    180
tgggaagagt ttgtggcggc tatgttcttg ttgatcattg tgggaagcat gctttggatt    240
ccgattgceg tggtcggttt tgtcctgtgt gtccgcagcg cggtgccgtg ggtggtgatg    300
ctcatcgtgt tcttcgccct gagcctgcac ccagtcgccg gcattcatga tatggttcat    360
tcgcctttga atcactttat attcaagtac ttcagtctta aaatggcgag tgatgcacca    420
ctggatagtg ctggggcgcta tatctttggt gctccgccgc atggggtgct gccgatgggg    480
aatcttatga cgtgacacgc gatgaaggct tgtggtggat tggagtccg tgggctgacg    540
acagatgtcg cgtcaggct gcctttatct cgacattact taggcgcat tggactattt    600
gccgcgactg ggcacgtggc gaagcagtac ctgcacgaag ggtggtcaat aggcatactt    660
tcgggcccgg tcgcccgaat tttcgaggta aataataagg atgaagtggg gttgatgaag    720
gagaggaagg gctttgtgaa gctcgcctt cgcacgggaa ctccgctggt gcttgttat    780
atatttggga ataccaagct gttgtcggcg tggtatgatg atggaggtgt gttgcagggt    840
ctttcacggt atttgaaatg tgggtgtgtg ccactttggg gtcggtttgg attgcccgtt    900
atgcaccgcc atccggtgct gggcgcgatg gcaaagccga ttgtggtccc caaggtggag    960
ggggagccta cgcaggagat gatagatgat taccataatc tctctgtca gacgctggtc   1020
gatctctttg ataggtacaa gggcttatat ggctggccgg acaagaagct gcttataaag   1080
tga                                                                    1083

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<210> SEQ ID NO 35

<211> LENGTH: 360

<212> TYPE: PRT

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 35

```

Met Ala Lys Ala Asn Phe Pro Pro Ala Ala Arg Tyr Val Asn Met Thr
1          5          10          15
Gln Val Tyr Ala Thr Gly Ala His Asn Met Pro Asp Glu Asp Arg Val
20         25         30
Lys Val Met Asn Gly Leu Ser Lys Pro Val Thr Glu Ala Lys Ala Gly
35         40         45
Asp Leu Gly Phe Gly Asp Val Glu Ser Met Thr Ala Trp Glu Glu Phe
50         55         60
Val Ala Ala Met Phe Leu Leu Ile Ile Val Gly Ser Met Leu Trp Ile
65         70         75         80
Pro Ile Ala Val Val Gly Phe Val Leu Cys Val Arg Ser Ala Val Ala
85         90         95
Trp Val Val Met Leu Ile Val Phe Phe Ala Leu Ser Leu His Pro Val
100        105        110
Pro Arg Ile His Asp Met Val His Ser Pro Leu Asn His Phe Ile Phe
115        120        125
Lys Tyr Phe Ser Leu Lys Met Ala Ser Asp Ala Pro Leu Asp Ser Ala
130        135        140

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Gly Arg Tyr Ile Phe Val Ala Pro Pro His Gly Val Leu Pro Met Gly
 145 150 155 160
 Asn Leu Met Thr Val His Ala Met Lys Ala Cys Gly Gly Leu Glu Phe
 165 170 175
 Arg Gly Leu Thr Thr Asp Val Ala Leu Arg Leu Pro Leu Phe Arg His
 180 185 190
 Tyr Leu Gly Ala Ile Gly Thr Ile Ala Ala Thr Gly His Val Ala Lys
 195 200 205
 Gln Tyr Leu Asp Glu Gly Trp Ser Ile Gly Ile Ser Ser Gly Gly Val
 210 215 220
 Ala Glu Ile Phe Glu Val Asn Asn Lys Asp Glu Val Val Leu Met Lys
 225 230 235 240
 Glu Arg Lys Gly Phe Val Lys Leu Ala Leu Arg Thr Gly Thr Pro Leu
 245 250 255
 Val Ala Cys Tyr Ile Phe Gly Asn Thr Lys Leu Leu Ser Ala Trp Tyr
 260 265 270
 Asp Asp Gly Gly Val Leu Gln Gly Leu Ser Arg Tyr Leu Lys Cys Gly
 275 280 285
 Val Leu Pro Leu Trp Gly Arg Phe Gly Leu Pro Leu Met His Arg His
 290 295 300
 Pro Val Leu Gly Ala Met Ala Lys Pro Ile Val Val Pro Lys Val Glu
 305 310 315 320
 Gly Glu Pro Thr Gln Glu Met Ile Asp Asp Tyr His Asn Leu Phe Cys
 325 330 335
 Gln Thr Leu Val Asp Leu Phe Asp Arg Tyr Lys Gly Leu Tyr Gly Trp
 340 345 350
 Pro Asp Lys Lys Leu Leu Ile Lys
 355 360

<210> SEQ ID NO 36
 <211> LENGTH: 1362
 <212> TYPE: DNA
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 36

```

atthtcagca aagtaatcaa gataataaac aaaaacaatc ctataaagga aaaacaacag    60
gaggcatcac aagcaatatg gccaaaggcta acttcccgcc cgcggcgcgc tatgttaata    120
tgacgcaggt ctatgcaaca ggcgctcaca atatgccgga cgaggaccgc gtcaagggtca    180
tgaacgggct gtccaagccc gtgacggagg ccaaggcagg tgatttgggg tttggggatg    240
ttgagtccat gacggcctgg gaagagtttg tggcggctat gttcttgttg atcattgtgg    300
gaagcatgct ttgattcccg attgcggtgg tcggttttgt cctgtgtgtc cgcagecggg    360
tggegtgggt ggtgatgctc atcgtgttct tegcctgag cctgcacca gteccgcgca    420
ttcatgatat ggttcattcg cctttgaate actttatatt caagtacttc agtcttaaaa    480
tggcgagtga tgcaccactg gatagtgtg ggcgctatat ctttgttget ccgcccatg    540
gggtgctgcc gatggggaat cttatgacgg tgcacgcgat gaaggcttgt ggtggattgg    600
agttccgtgg gctgacgaca gatgtcgcgc tcaggctgcc tttatttcga cattaactag    660
gcgccattgg tactattgcc gcgactgggc acgtggcgaa gcagtacctc gacgaagggt    720
ggtaaatagg catatcttcg ggcggagtgc cggaaathtt cgaggtaaat aataaggatg    780
  
```


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aagtgggtgtt gatgaaggag aggaagggtt ttgtgaagct cgccttcgc acgggaactc	840
cgctgggtggc ttgttatata tttgggaata ccaagctgtt gtcggcgtgg tatgatgatg	900
gaggtgtgtt gcagggctctt tcacgttatt tgaatgtgg tgtgttgcca ctttggggtc	960
ggtttgatt gccgcttatg caccgccate cgggtgctgg cgcgatggca aagccgattg	1020
tggccccaa ggtggagggg gagcctacgc aggagatgat agatgattac cataatctct	1080
tctgtcagac gctggtcgat ctctttgata ggtacaaggg cttatatggc tggccggaca	1140
agaagctgct tataaagtga gtgggtaga gtagattgcg tgacgggggg gagaggggga	1200
tgaatgcaat tgtagaagga attctagga tttttcgta ggcgtttgt atctagtct	1260
gtagggatag gggcatttgt tcaggaggtg aaagtttgt cgggtgatcc aaagaccaa	1320
tgcagcacia caaatcaaag aaagcatgaa aacacaatcc aa	1362

<210> SEQ ID NO 37

<211> LENGTH: 1695

<212> TYPE: DNA

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 37

atgttgttc agggattaag ctggtctttt ttgacctgt cgattgtggt agaaatcttg	60
tttgtgatct cgacgtttgc tgtggggttt gagttgtttg ttggagcggc ggtgggtggc	120
ggcggcttct ttttggcttc ggaagtgttg atgattgtga gtttcattt ttatatgcct	180
acgacgacca cgactgtgac aacgaccggg ttggcgggta tggaggagaa ggtggaggag	240
gtggaggaga tgatgtggg gaaggaggga gtgggggaag aggacgagga gatggtggag	300
gaaaaggtgg acgtgacgc agcggcgacg acgaacgcac tcttaagaac cgaagcag	360
cggctgctct tggcgaaga gagtgtacg accactacta ctaccgcgac tgtgaccacg	420
gggcagacca gcaagcgtc tacttcattt atgcctgtcc gggtcgacga ggttccctt	480
gagcaattcc gccggctcac cgttataacc gttctgagta atatgcaata cctgcccttc	540
ctccttccca tctcctctt tgcctctca ggtcttctc tccctgtggc atcttttca	600
tggttcggcg ctttttgttg tctgacctca gcggtggtt taaacgcta tgtcaaaacc	660
acgttgcca aagctgggaa tcgtatttcc tccttccagc gtcctcctc taatgtcctc	720
cccacgctca tttatgccg gccgggtctt atttgctttt ttgcgtggag tcaaacccaa	780
ggtgggaggg aggacgggaa ggagcgcgcg gtgactgctg tcccggcttg ggcggcgcgc	840
acggccatgc attacctgta cctcttctc acgtttcgcg gaaatccgga agtaacggga	900
gagaggtact taggcgaaaa gctagagctg tggaaaggcg gttggtcatt gtactatttt	960
ttagaaggga tagatcaata ttttcaggcg aagtgtgtct tcatggaccc gaaactggat	1020
ctgaagggga aaccgcatgt gtttgcgttt caccacacgc gagtcagcc gtttacgacg	1080
ttttggattc agctttcgcg ggcctggagg gagggagtgg ggaagggaca gagattctgt	1140
gtgatgactg cgagtgttat gcattatgtg ccgttaatgc gcgatattt acagtggctc	1200
ggggggcggg aagtgagcag ggaagccatt tcgtacgcac tggaccgtaa acagtacgta	1260
ttgttggttc caggcgaca acaagagatg atggagtccc aatctcagat gggcgagatt	1320
cggatcatta cgaagcacgt cggcttcatt agattagcac tccagacagg cgcgccgcgc	1380
gtgcctgtgc tctcatttgg cgaagttgaa gtgatggatt ttgtccgta cccgcgtcta	1440

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cagcgtttct ttatctcgcg catcggtatt ccggttccct tcttcccata tggattgttt 1500
ggatttccca tcccaaggcc cgtgcccgtg acggtcgtgt ttggccgtcc gattgcagtg 1560
gagaaagtgg agcaaccgac gcaggaagag gtgcgtaaat tgcgaaaaa gtactttgaa 1620
agtatccagg aggtgtttga taaaaaag gcgaaggccc tggggcatgg aaatcataaa 1680
ttggtcctgt tgtga 1695

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<210> SEQ ID NO 38

<211> LENGTH: 564

<212> TYPE: PRT

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 38

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Met Leu Leu Gln Gly Leu Ser Trp Ser Phe Leu Thr Leu Ser Ile Val
1      5      10      15
Val Glu Ile Leu Phe Val Ile Ser Thr Phe Ala Val Gly Phe Glu Leu
20     25     30
Phe Val Gly Ala Ala Val Val Ala Gly Gly Phe Phe Leu Val Ser Glu
35     40     45
Val Leu Met Ile Val Ser Leu His Phe Tyr Met Pro Thr Thr Thr Thr
50     55     60
Thr Val Thr Thr Thr Gly Leu Ala Val Met Glu Glu Lys Val Glu Glu
65     70     75     80
Val Glu Glu Met Met Val Gly Lys Glu Gly Val Gly Glu Glu Asp Glu
85     90     95
Glu Met Val Glu Glu Lys Val Asp Val Thr Thr Ala Ala Thr Thr Asn
100    105    110
Ala Leu Leu Arg Thr Glu Lys Gln Arg Leu Leu Leu Ala Lys Glu Ser
115    120    125
Ala Thr Thr Thr Thr Thr Thr Ala Thr Val Thr Thr Gly Gln Thr Ser
130    135    140
Lys Thr Ser Thr Ser Phe Met Pro Val Arg Val Asp Glu Ala Ser Leu
145    150    155    160
Glu Gln Phe Arg Arg Leu Thr Val Ile Thr Val Leu Ser Asn Met Gln
165    170    175
Tyr Leu Pro Phe Leu Leu Pro Ile Leu Pro Phe Val Leu Ser Gly Leu
180    185    190
Pro Leu Pro Val Ala Ser Phe His Trp Phe Gly Ala Phe Cys Cys Leu
195    200    205
Thr Ser Ala Val Val Leu Asn Ala Tyr Val Lys Thr Thr Leu Ala Lys
210    215    220
Ala Gly Asn Arg Ile Ser Ser Phe Gln Arg Ser Leu Leu Asn Val Leu
225    230    235    240
Pro Thr Leu Ile Tyr Ala Ala Pro Gly Leu Ile Cys Phe Phe Ala Trp
245    250    255
Ser Gln His Gln Gly Gly Arg Glu Asp Gly Lys Glu Arg Ala Val Thr
260    265    270
Ala Phe Pro Ala Trp Ala Ala Leu Thr Ala Met His Tyr Leu Tyr Leu
275    280    285
Phe Leu Thr Phe Arg Gly Asn Pro Glu Val Thr Gly Glu Arg Tyr Leu
290    295    300
Gly Glu Lys Leu Glu Leu Trp Lys Gly Gly Trp Ser Leu Tyr Tyr Phe

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305	310	315	320
Leu Glu Gly Ile Asp Gln Tyr Phe Gln Ala Lys Leu Val Phe Met Asp	325	330	335
Pro Lys Leu Asp Leu Lys Gly Lys Pro His Val Phe Ala Phe His Pro	340	345	350
His Gly Val Gln Pro Phe Thr Thr Phe Trp Ile Gln Leu Ser Arg Ala	355	360	365
Trp Arg Glu Gly Val Gly Lys Gly Gln Arg Phe Cys Val Met Thr Ala	370	375	380
Ser Val Met His Tyr Val Pro Leu Met Arg Asp Ile Leu Gln Trp Leu	385	390	395
Gly Gly Arg Glu Val Ser Arg Glu Ala Ile Ser Tyr Ala Leu Asp Arg	405	410	415
Lys Gln Ser Val Leu Leu Val Pro Gly Gly Gln Gln Glu Met Met Glu	420	425	430
Ser Gln Ser Gln Met Gly Glu Ile Arg Ile Ile Thr Lys His Val Gly	435	440	445
Phe Ile Arg Leu Ala Leu Gln Thr Gly Ala Pro Leu Val Pro Val Leu	450	455	460
Ser Phe Gly Glu Val Glu Val Met Asp Phe Val Arg Tyr Pro Arg Leu	465	470	475
Gln Arg Phe Phe Ile Ser Arg Ile Gly Ile Pro Val Pro Phe Phe Pro	485	490	495
Tyr Gly Leu Phe Gly Phe Pro Ile Pro Arg Pro Val Pro Val Thr Val	500	505	510
Val Phe Gly Arg Pro Ile Ala Val Glu Lys Val Glu Gln Pro Thr Gln	515	520	525
Glu Glu Val Arg Lys Leu Ser Lys Lys Tyr Phe Glu Ser Ile Gln Glu	530	535	540
Val Phe Asp Lys Asn Lys Ala Lys Ala Leu Gly His Gly Asn His Lys	545	550	555
Leu Val Leu Leu			

<210> SEQ ID NO 39

<211> LENGTH: 2074

<212> TYPE: DNA

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 39

```

aagggaggga ggggaagagcg caccagaagg ccgtacgaaa gcaatggcgt ttttggcagc      60
catttttggg aggagccaag tttatgttgt tgcagggatt aagctggtct tttttgacct      120
tgtcgattgt ggtagaaatc ttgtttgtga tctcgacgtt tgctgtgggg tttgagttgt      180
ttgttgagc ggcggtggtg gcggggcgct tctttttggt ctcggaagtg ttgatgattg      240
tgagtttgea tttttatatt cctacgacga ccacgactgt gacaacgacc gggttggcgg      300
tgatggagga gaaggtggag gaggtggagg agatgatggt ggggaaggag ggagtggggg      360
aagaggacga ggagatggtg gaggaaaagg tggacgtgac gacagcggcg acgacgaacg      420
cactcttaag aaccgaaaag cagcggctgc tcttggcgaa agagagtgtc acgaccacta      480
ctactaccgc gactgtgacc acggggcaga ccagcaagac gtctacttca tttatgcctg      540
tccgggtcga cgaggcttcc cttgagcaat tccgccggtt caccgttata accgttctga      600
    
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gtaatatgca atacctgccc ttccctcttc ccacccctcc tttgtcctc tcaggtcttc 660
ctctccctgt ggcacatcttt cactgggttc gegctttttg ttgtctgacc teageggteg 720
ttttaaacgc ctatgtcaaa accacgttgg ccaaagctgg gaatcgtatt tcctccttcc 780
agcgctccct ccttaatgtc tccccacgc tcatttatgc cgcgccgggt cttattttgt 840
tttttgctg gagtcaacac caaggtggga gggaggacgg gaaggagcgc gcggtgactg 900
cgttcccggc ttggggggcg ctcaaggcca tgcattacct gtacctctt ctcacgttcc 960
gcggaatcc ggaagtaacg ggagagaggt acttaggcga aaagctagag ctgtggaaaag 1020
gcggttggtc attgtactat tttttagaag ggatagatca atattttcag gcgaagttgg 1080
tcttcatgga ccgaaaactg gatctgaagg gaaaaccgca tgtgtttgcg tttcaccac 1140
acggagtcca gccgtttacg acgttttggg ttcagcttcc gcgggcctgg agggagggag 1200
tggggaaggg acagagatcc tgtgtgatga ctgcgagtg tatgcattat gtgccgttaa 1260
tgccgatata attacagtgg ctccgggggc ggggaagtgg cagggaaacc atttcgtacg 1320
cactggaccg taaacagtca gtattgttgg ttccaggcgg acaacaagag atgatggagt 1380
cccaatctca gatggggcag attcggatca ttacgaagca cgtcggcttc attagattag 1440
cactccagac aggcgcgcgc ctccgtgcctg tgctctcatt tggcgaagtt gaagtgatgg 1500
attttgcctg gtaccgcgt ctacagcgtt tctttatctc gcgcacgggt attccggctc 1560
ccttcttccc atatggattg tttggatttc ccaccccaag gcccggtccc gtgacggctg 1620
tgtttgccg tccgattgca gtggagaaaag tggagcaacc gacgcaggaa gaggtgcgta 1680
aattgtcgaa aaagtacttt gaaagtatcc aggaggtggt tgataaaaaat aaggcgaagg 1740
ccctggggca tggaaatcat aaattggtcc tgttgtgagg gaggaagaga agcaaaaggg 1800
tgggagacag ggagatggat ggggagaagg aggtttgtgg gggtaggctt tcggagagag 1860
aacaacgga ctgatacaag acaaaagtgt aagatagaac ttcaggaaag cgaataaatg 1920
attgaacgac atagaaaaaa gaaagggcag cgagggaagg agggagggag gaagggagga 1980
cagtactgaa atgccaccaa tggcggctcc agcatcggag aatgcacaat aaagcaacaa 2040
agctagtcgg taatgaaaaa aaaaaaaaaa aaaa 2074

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<210> SEQ ID NO 40

<211> LENGTH: 1029

<212> TYPE: DNA

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 40

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atgttgatgg cgcgctcgcg gcggccagca tegtcttgg tggacccttt gccattgacg 60
gggaagctgc ctatcggggc aatcaggctc ttcacgtccc ggctgcttc atggcgtacc 120
actcccatgg tcgtggggcg ctccttctct gtggtgggat ccttcgtctg ggtgcccctt 180
gttatctggc tgggttgaa gaaatgtagg acacggaatc gacgcattgt ctacgtcctt 240
gttttgtgtg tcactctgac cctacctaca cggcgttggg acgcggtggg cttgaacggc 300
ctatggagcc gttttgtgga atatttttca gtccaggtgg taggggacga ccccttgccc 360
aaggaccgct ccgccgtcta cgccgtcatt cctcaccgca ccttcccctt tggctctcgc 420
gtggtctccc tcggctccct gaacaagatc ttcaataagg tccggcccgt ggtggcctcg 480
gcagtcttgc gctttccggg ctttgggtcaa ctaataggct tcgccggtgg ggtcgacgca 540

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gggccccaaag aagtaagcaa ggccatcaag aagggtgtt cagtgagtat ctgtcctggg 600
ggcatcgag agatgttctg gggatttcca aaggagggt gcttaccgcg ggaggaatat 660
ggtttcttac agtcgaggaa agggtttacc cgcattggcca tgaacacaaa tgtgcctgtg 720
gtcctctgtg actgttttgg taacacccac gcgatgcata aggcgaagac gccttgggtc 780
ttggaggcgc tatcaaggct tctcaagacc tctcttatct taacctgggg cgggtggggg 840
ctgccgatcc cctaccgtgt gcctctctc tacgcccgc gtaagcccct ccgcctcctg 900
cacgcagaaa atccaacccc tgctcagatt gaggcggcgc acgccagatt ctgcagggcc 960
ctttcggatt tgtttgatcg gtacaagttt tattatggat gggggcacia gacgcttcgc 1020
atcgtctga 1029

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<210> SEQ ID NO 41
<211> LENGTH: 342
<212> TYPE: PRT
<213> ORGANISM: Nannochloropsis oculata

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<400> SEQUENCE: 41

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```

Met Leu Met Ala Pro Ser Arg Arg Pro Ala Ser Ser Leu Val Asp Pro
1      5      10     15
Leu Pro Leu Thr Gly Lys Leu Pro Ile Gly Ala Ile Arg Leu Phe Thr
20     25     30
Ser Arg Pro Ala Ser Trp Arg Thr Thr Pro Met Val Val Gly Gly Ser
35     40     45
Leu Leu Val Val Gly Ser Phe Val Trp Val Pro Leu Val Ile Trp Leu
50     55     60
Gly Trp Lys Lys Cys Arg Thr Arg Asn Arg Arg Ile Val Tyr Val Leu
65     70     75     80
Val Leu Cys Val Ile Leu Thr Leu Pro Thr Arg Arg Trp Asp Ala Val
85     90     95
Val Leu Asn Gly Leu Trp Ser Arg Phe Val Glu Tyr Phe Ser Val Gln
100    105    110
Val Val Gly Asp Asp Pro Leu Pro Lys Asp Arg Ser Ala Val Tyr Ala
115    120    125
Val Ile Pro His Gly Thr Phe Pro Phe Gly Leu Gly Val Val Ser Leu
130    135    140
Gly Pro Leu Asn Lys Ile Phe Asn Lys Val Arg Pro Val Val Ala Ser
145    150    155    160
Ala Val Leu Arg Phe Pro Gly Phe Gly Gln Leu Ile Gly Phe Ala Gly
165    170    175
Gly Val Asp Ala Gly Pro Lys Glu Val Ser Lys Ala Ile Lys Lys Gly
180    185    190
Cys Ser Val Ser Ile Cys Pro Gly Gly Ile Ala Glu Met Phe Trp Gly
195    200    205
Phe Pro Lys Glu Gly Cys Leu Pro Arg Glu Glu Tyr Ala Phe Leu Gln
210    215    220
Ser Arg Lys Gly Phe Ile Arg Met Ala Met Lys His Asn Val Pro Val
225    230    235    240
Val Pro Val Tyr Cys Phe Gly Asn Thr His Ala Met His Lys Ala Lys
245    250    255
Thr Pro Trp Val Leu Glu Ala Leu Ser Arg Leu Leu Lys Thr Ser Leu

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	260		265		270	
Ile	Leu Thr Trp Gly Arg Trp Gly Leu Pro Ile Pro Tyr Arg Val Pro					
	275		280		285	
Leu	Leu Tyr Ala Val Gly Lys Pro Leu Arg Leu Leu His Ala Glu Asn					
	290		295		300	
Pro	Thr Pro Ala Gln Ile Glu Ala Ala His Ala Glu Phe Cys Arg Ala					
	305		310		315	320
Leu	Ser Asp Leu Phe Asp Arg Tyr Lys Phe Tyr Tyr Gly Trp Gly His					
		325		330		335
Lys	Thr Leu Arg Ile Val					
	340					

<210> SEQ ID NO 42
 <211> LENGTH: 1585
 <212> TYPE: DNA
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 42

```

attttcagca aagtaataca gataataaca aaaacaatcc tctaaaagga aaaacaacag    60
ctttaccctc agggagctca tgttgatggc gccgtcgccg cggccagcat cgtccttggg    120
ggaccctttg ccattgacgg ggaagctgcc tatcggggca atcaggctct tcacgtcccc    180
gcctgcttca tggcgtaaca ctcccatggt cgtggcgccg tccttgctgg tggtagggatc    240
cttcgtctgg gtgccccttg ttatctggct gggttggaag aaatgtagga cacggaatcg    300
acgcattgtc taegtccttg ttttgtgtgt catcttgacc ctacctacac ggcggtggga    360
cgcggtggtc ttgaacggcc tatggagccg ttttgtgtaa tatttttcag tccagtggtg    420
aggggacgac cccttgccca aggaccctc cgccgtctac gccgtcattc ctacggcac    480
cttccccctt ggtctcggcg tggctctcct cggctccttg aacaagatct tcaataaggt    540
cggccccgtg gtggcctcgg cagtcttgcg ctttccgggc tttggtaaac taataggctt    600
cgccgggtgg gtcgagcagc ggcccaaga agtaagcaag gccatcaaga agggctgttc    660
agtgagtatc tgtcctgggg gcatcgaga gatgttctgg ggatttcaa aggaggctg    720
cttaccgctg gaggaatatg cgttcttaca gtcgagggaaa gggtttatcc gcatggccat    780
gaaacacaat gtgcctgtgg tcctctgtga ctgttttggg aacacccacg cgatgcataa    840
ggcgaagacg ccttgggtct tggagcgct atcaaggtea gtcacggggg aatagtgggg    900
ttgagtggga gacggcgggg gaaaatatat cttgattttt attgtaccgc atctgcgagg    960
ctgtctctaa tcgctttcta cgcgagacca ttcaaaattt tcgctatttc tttgcgtcgt   1020
ctttccgtac gcattaggct tctcaagacc tctcttatct taacctgggg cgggtggggg   1080
ctgccgatcc cctaccgtgt gcctctcctc tacgccgtcg gtaagccct cgcctcctg   1140
cacgcagaaa atccaacccc tgctcagatt gaggcggcgc acgccagtt ctgcagggcc   1200
ctttcggatt tgtttgatcg gtacaagttt tattatggat gggggcacia gacgcttcgc   1260
atcgtctgag aacgggggga gggggggagg ggtcgttagg ttatgctgga aggaaagaga   1320
atgggagaga gggagagaga aagagtgggg aagatattga tggtagatgc ctgctctggg   1380
aggcaattgc tgcttgggga ggctcccgag ggagaatgag ggagcgaaga gtagggaaac   1440
caaattatta aatcttttct cttcgtaag acttaggaat aaatgtaaag taaaaagaag   1500
aagagcccgt ctcttgcctc aaattgaaag aaataaagat aaccaatgaa ctaaaaaaaa   1560
    
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 aaaaaaaaaa aaaaaaaaaa aaaaa 1585

<210> SEQ ID NO 43
 <211> LENGTH: 1251
 <212> TYPE: DNA
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 43

```

atgggcgcta ccactgcgac ccagactaaa aagacgttgg tcatgctggac agtcgcagtg   60
cgtaacgagg atatagtgcc ggaagcagcg acgggagacg gagcagcagg cgatgcaact   120
gctggtggcc tttctcgctc aacaccaaca gggctcccg aggctccac ttcgctttca   180
tcgcgactgg taccatcccc agcacaagtt tcatccatgc cccagcaca agcttcagcc   240
acgcctattg tggtgcggcc cgaggcacgc cccgcaggtc cacaaggccg tctacaagca   300
ttaggtgcgg tgctatTTTT ggggtctcatg gggctcgtcg tgtacctagt gatcgcgtca   360
gcgctttaca tcgtgattgg tttcgggtg ttgggccacc gcatttgccc ttcgatctta   420
ctcggggttt gggtaggaca agccctaatt tccgtcaagg tgctgcacca agaccggaa   480
ggtatcaagc ggtcgtggct tttccgagaa atggtgaact tttttgatgt gacactggtg   540
atggagcaga aattggacac ttccaagaag tacctatttg cacaacaccc gcacggtatc   600
cttcccctcg ccccctgtgt gtccgcttac tttgtctcgg acgtggtgcc cggcggaggc   660
aagatctttt gtttgataca tagcggcatc tttcacctgc ccatcgtccg ttttttcatg   720
ggtgaatggg gtgcactctc gcacaacaag gagtctgtcg ccgaagcaaa gcaacaagga   780
cagcattgct ccatcgtcgt cggcggggtc gcggagattt tctccaaaa cggagagacc   840
gagcaactgc aactcagaaa gggcttcatt cgtgaggcac ttcgtaatgg atatgacctt   900
gtgcccattg ttcactttgg ggccacgcgc atgtatcatt ttggtggccc tgtttcattt   960
tgggcgtcct tgtccaatta cctgcccttt ccctttttcc tcatgggggg atggggaaaa 1020
gggttgacct tgctcccaaa acctgtgcgt attgtaattg ctgctgggtc gcccataggc 1080
cttgccgctt tgtatggggg gccggaagga cagtcgggtc ctgatccaga cctggcgaaa 1140
gtggatttga tatatgagga gtggaagaag cacttgccgg gcctgtatta tcggcagcgg 1200
cctgagtggg aaacgcggga gttggagatt ttggactgtc cgaagtcgtg a 1251
  
```

<210> SEQ ID NO 44
 <211> LENGTH: 416
 <212> TYPE: PRT
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 44

```

Met Gly Ala Thr Thr Ala Thr Gln Thr Lys Lys Thr Leu Val Met Arg
1           5           10          15

Thr Val Ala Val Arg Asn Glu Asp Ile Val Pro Glu Ala Ala Thr Gly
          20          25          30

Asp Gly Ala Ala Gly Asp Ala Thr Ala Gly Gly Leu Ser Arg Ser Thr
          35          40          45

Pro Thr Ala Ala Pro Glu Ala Ser Thr Ser Leu Ser Ser Arg Leu Val
          50          55          60

Pro Ser Pro Ala Gln Val Ser Ser Met Pro Pro Ala Gln Ala Ser Ala
65          70          75          80
  
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Thr Pro Ile Val Val Arg Pro Glu Ala Arg Pro Ala Gly Pro Gln Gly
 85 90 95

Arg Leu Gln Ala Leu Gly Ala Val Leu Phe Leu Gly Leu Met Gly Ser
 100 105 110

Ser Leu Tyr Leu Val Ile Ala Ser Ala Leu Tyr Ile Val Ile Gly Phe
 115 120 125

Gly Val Leu Gly His Arg Ile Cys Pro Ser Ile Leu Leu Gly Val Trp
 130 135 140

Val Gly Gln Ala Leu Ile Ser Val Lys Val Leu His Gln Asp Pro Glu
 145 150 155 160

Gly Ile Lys Arg Ser Trp Leu Phe Arg Glu Met Val Asn Phe Phe Asp
 165 170 175

Val Thr Leu Val Met Glu Gln Lys Leu Asp Thr Ser Lys Lys Tyr Leu
 180 185 190

Phe Ala Gln His Pro His Gly Ile Leu Pro Leu Ala Pro Val Leu Ser
 195 200 205

Ala Tyr Phe Val Ser Asp Val Val Pro Gly Gly Gly Lys Ile Phe Cys
 210 215 220

Leu Ile His Ser Gly Ile Phe His Leu Pro Ile Val Arg Phe Phe Met
 225 230 235 240

Gly Glu Trp Gly Ala Leu Ser Ala Asn Lys Glu Ser Val Ala Glu Ala
 245 250 255

Lys Gln Gln Gly Gln His Cys Ser Ile Val Val Gly Gly Val Ala Glu
 260 265 270

Ile Phe Leu Gln Asn Gly Glu Thr Glu Gln Leu Gln Leu Arg Lys Gly
 275 280 285

Phe Ile Arg Glu Ala Leu Arg Asn Gly Tyr Asp Leu Val Pro Met Phe
 290 295 300

His Phe Gly Ala Thr Arg Met Tyr His Phe Val Gly Pro Val Ser Phe
 305 310 315 320

Trp Arg Ser Leu Ser Asn Tyr Leu Pro Phe Pro Phe Phe Leu Ile Gly
 325 330 335

Gly Trp Gly Lys Gly Leu Thr Leu Leu Pro Lys Pro Val Arg Ile Val
 340 345 350

Ile Ala Val Gly Ser Pro Ile Gly Leu Ala Ala Leu Tyr Gly Val Pro
 355 360 365

Glu Gly Gln Ser Val Pro Asp Pro Asp Leu Ala Lys Val Asp Leu Ile
 370 375 380

Tyr Glu Glu Trp Lys Lys His Leu Ala Gly Leu Tyr Tyr Arg Gln Arg
 385 390 395 400

Pro Glu Trp Glu Thr Arg Glu Leu Glu Ile Leu Asp Cys Pro Lys Ser
 405 410 415

<210> SEQ ID NO 45
 <211> LENGTH: 1923
 <212> TYPE: DNA
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 45

atcttcagca aagtaatacaa gataataaac aaaaacaatc ctataaagga aaaacaacag 60
 gaaagccaag ctgccacgct tgcataagaa caaagggggg catcaccacg cgacgctggg 120
 gacggagaag gacatcaaac aaggacacaa gcatggggcg taccactgcg acccagacta 180

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aaaagacggt ggtcatgctg acagtcgcag tgcgtaacga ggatatagtg ccggaagcag 240
cgacgggaga cggagcagca ggcgatgcaa ctgctgggtg cctttctcgc tcaacaccaa 300
cagcggctcc ggaggcctcc acttcgcttt catcgcgact ggtaccatcc ccagcacaag 360
tttcatccat gccccagca caagcttcag ccacgcctat tgtggtgctg cccgagggcac 420
gccccgcagg tccacaaggc cgtctacaag cattaggtgc ggtgctatct ttggggctca 480
tggggctgct gctgtacctg gtgatcgcgt cagcgcctta catcgtgatt ggtttcgggtg 540
tgttgggcca ccgcatcttc ccttcgatct tactcggggg ttgggttagga caagccctaa 600
tttccgcaaa ggtgctgcac caagaccggc aaggatcaa gcggtcgtgg cttttccgag 660
aatgggtgaa cttttttgat gtgacctggc tgatggagca gaaattggac acttccaaga 720
agtaacctatt tgcacaacac ccgcacggta tccttcccct cgcctccgtg ttgtccgctt 780
actttgtctc ggacgtgggt cccggcggag gcaagatctt ttgtttgata catagcggca 840
tctttcacct gcccatcgtc cgttttttca tgggtgaatg ggggtgactc tccgcaaaaca 900
aggagtctgt cgcgaagca aagcaacaag gacagcattg ctccatcgtc gtcggcgggg 960
tcgaggagat tttccctcaa aacggagaga ccgagcaact gcaactcaga aagggttca 1020
ttcgtgaggc acttcgtaat ggatatagac ttgtgccat gtttactttt ggggccacgc 1080
gcctgatca ttttgtggc cctgtttcat tttggcggtc cttgtccaat tacttgcctg 1140
ttcccttttt cctcattggg ggatggggaa aagggttgac cttgctcccc aaacctgtgc 1200
gtattgtaat tctgtctcgt tcgccatag gccttgcggc tttgtatggg gtgccggaag 1260
gacagtcggt gctgatcca gacctggcga aagtggattt gatataatgag gagggaaga 1320
agcacttggc gggcctgtat taccggcagc ggctgagtg ggaacgcgg gagttggaga 1380
ttttggactg tccgaagtcg tgagtgatta aaaagagatc gcactctgtc gacgaagtgc 1440
tttgtacagc agccgatag gggggaaggt aatatttga aaaggtcaaa aggtggagtg 1500
cagagtagga ggatttgaca aagattaaga cgtggcgcac atgacgacat gggagaaaga 1560
ctggtcgaat ttaacaaaa aaagagctac ccgagcaagc gtaacgcaga ggagcattta 1620
agtatgcatg ttcccaaggc aaggcaaggc aaaaggccat ccgagtagca ggcacacgca 1680
tgtaaagtgg cgacgcttac acttttggat attaacgaat aaaagacaca aggatgtcgc 1740
ttacagtgca gcagcagcaa ttacatgctt gtgcgaagtc tctaggggat acctccagca 1800
ctgtcatcaa cataagtaag atacgaaaga cacagaagga taagtgggag gatgggggtg 1860
agtaggaggg tggggaggtt ggatggaaaa ggggggttcg gcgagtgag ttggacaggg 1920
ccc 1923

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<210> SEQ ID NO 46

<211> LENGTH: 930

<212> TYPE: DNA

<213> ORGANISM: *Thraustochytrium aureum*

<400> SEQUENCE: 46

```

atgtcgttcg ttgagcacag cgcgggtggg ctctgcttg cctttgtgat gggcggcgca 60
ctgtactggt cctgggcccg gctcgcgggt ctcactctgg ggtcgtggtc gcaggtggct 120
acttatgtgg tctgacggc tgtgctggcc ctgcaccgca tcccggacat ctcgatgcc 180
gtgtacagct cgtggatcgt gcagcaattg tacaagtact ttacctaccg ctttgtgtac 240

```

-continued

```

tcggggaacg cgcgcgtact agcgcagacg cagggcgcgt tcateggcgc aggcgtcccg 300
cacggcgcga tgcggttctc caacctgctc teagtcctg ctgtcaactc gttttctccg 360
agccagaccg gggggcaatt tgctggggcg ccggcgagca ttgtgttccg cacgccttc 420
ctgcgctact ttaccatgtt caagtcggtc acggtgtcac gcgagagcct caccaaacag 480
ctggagctcg ggaacacggt tggcctggtt ggcgatggca tcgctgggat cttccaatgc 540
gaccacaacg acgaggctgt tgcgctccgg acgcgcaagg ggctcgaaa actggcgctg 600
cgaacggggc ggcccgtttt gccctgctac agcttgggaa acacggaagc gtttagcgtt 660
tggtttgacc gctggggcgt catggagcgc ctctcgcgca agctgcaggc gacgctggtt 720
ttctactggg gcaggtacgg cctccctggt cegtaccgtg tcaatatcac gatgatcctc 780
ggcgacatgg tctcgtcga ccaggtcgag aacccgacgc cggcacaggt cgatgcagtg 840
cacgagcgc tctctgctc catcgagaac gccttcaatc ggcacaaggc cgccttgggt 900
tggggccaca agacgatgcg atttgtgtag 930

```

<210> SEQ ID NO 47

<211> LENGTH: 309

<212> TYPE: PRT

<213> ORGANISM: *Thraustochytrium aureum*

<400> SEQUENCE: 47

```

Met Ser Phe Val Glu His Ser Ala Val Val Leu Val Leu Ala Phe Val
1          5          10          15
Met Gly Gly Ala Leu Tyr Trp Ser Trp Ala Gly Leu Ala Val Leu Ile
20          25          30
Trp Gly Ser Trp Ser Gln Val Ala Thr Tyr Val Val Leu Thr Ala Val
35          40          45
Leu Ala Leu His Pro Ile Pro Asp Ile Ser Asp Ala Val Tyr Ser Ser
50          55          60
Trp Ile Val Gln Gln Leu Tyr Lys Tyr Phe Thr Tyr Arg Phe Val Tyr
65          70          75          80
Ser Gly Asn Ala Arg Val Leu Ala Gln Thr Gln Ala Pro Phe Ile Gly
85          90          95
Ala Gly Val Pro His Gly Ala Met Pro Phe Ser Asn Leu Leu Ser Val
100         105         110
Pro Ala Val Asn Ser Phe Ser Pro Ser Gln Thr Gly Gly Glu Phe Val
115         120         125
Gly Ala Pro Ala Ser Ile Val Phe Arg Thr Pro Phe Leu Arg Tyr Phe
130         135         140
Thr Met Phe Lys Ser Val Thr Val Ser Arg Glu Ser Leu Thr Lys Gln
145         150         155         160
Leu Glu Leu Gly Asn Thr Val Gly Leu Val Gly Asp Gly Ile Ala Gly
165         170         175
Ile Phe Gln Cys Asp His Asn Asp Glu Val Val Ala Leu Arg Thr Arg
180         185         190
Lys Gly Leu Ala Lys Leu Ala Leu Arg Thr Gly Arg Pro Val Leu Pro
195         200         205
Cys Tyr Ser Leu Gly Asn Thr Glu Ala Phe Ser Val Trp Phe Asp Arg
210         215         220
Trp Gly Val Met Glu Arg Leu Ser Arg Lys Leu Gln Ala Ser Val Phe

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225	230	235	240
Phe Tyr Trp Gly Arg Tyr Gly Leu Pro Val Pro Tyr Arg Val Asn Ile	245	250	255
Thr Met Ile Leu Gly Asp Met Val Leu Val Asp Gln Val Glu Asn Pro	260	265	270
Thr Pro Ala Gln Val Asp Ala Val His Glu Arg Ile Leu Ala Ser Ile	275	280	285
Glu Asn Ala Phe Asn Arg His Lys Ala Ala Leu Gly Trp Gly His Lys	290	295	300
Thr Met Arg Phe Val			
305			

<210> SEQ ID NO 48
 <211> LENGTH: 1134
 <212> TYPE: DNA
 <213> ORGANISM: Thraustochytrium aureum

<400> SEQUENCE: 48

```

aagcgtttag cgtttggtt gaccgagcag gcgccaatgt cgttcgttga gcacagcgcg      60
gtggtgctcg tgettgcctt tgtgatgggc ggcgcactgt actggctctg ggcggggctc      120
gcggtgctca tctgggggtc gtggtcgcag gtggctactt atgtggtgct gacggctgtg      180
ctggccctgc acccgatccc ggacatctcg gatgcccgtg acagctcgtg gatcgtgcag      240
caattgtaca agtactttac ctaccgcttt gtgtactcgg ggaacgcgcg cgtactagcg      300
cagacgcagg cgccgttcat cggcgcaggc gtcccgcacg gcgcgatgcc gttctccaac      360
ctgctctcag tccctgctgt caactcgttt tctccgagcc agaccggggg cgaatttgtc      420
ggggcgccgg cgagcattgt gttccgcacg cctttcctgc gctactttac catggtcaag      480
tcgggtcagg tgtcaecgca gagcctcacc aaacagctgg agctcgggaa caeggttggc      540
ctggttggcg atggcatcgc tgggatcttc caatgcgacc acaacgacga ggtcgttgcg      600
ctccggacgc gcaaggggct cgcaaaaactg gcgctgcgaa cggggcggcc cgttttgccc      660
tgctacagct tgggaaacac ggaagcgttt agcgtttggt ttgaccgctg gggcgtcatg      720
gagcgcctct cgcgcaagct gcaggcgagc gtgtttttct actggggcag gtacggcctc      780
cctgttccgt accggtgcaa tatcacgatg atcctcggcg acatggtcct cgtcgaccag      840
gtcgagaacc cgacgcgggc acaggtcgat gcagtgacag agcgcattct tgcgtccatc      900
gagaacgect tcaateggca caaggccgcc cttggttggg gccacaagac gatgcgattt      960
gtgtaggagg tgctgtttgc caacaccaca cttggcctgg cctgggatgc ggctgggcca     1020
atcgtttcgg tgatecgcg tcgagctcga gctactcgag agtcaccgcc gagcgaggca     1080
gccataaaga gtcgaacgaa aatagcaaaa tgtgcaatc accaaaaaaaa aaaa          1134

```

<210> SEQ ID NO 49
 <211> LENGTH: 1179
 <212> TYPE: DNA
 <213> ORGANISM: Thraustochytrium aureum

<400> SEQUENCE: 49

```

atggtcttcc tctgccttcc ctacatgctc cccgaagcgc tgctcccttt cttggacacg      60
gcgacgctag gectcaccgc ggccctgccc ggagacaagg agaactttgt ccacacgttt      120
gccgtgtggt ggacgctctt gtggggcatt gcgttttggc cgatctttta cgccgcgctc      180

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-continued

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aagaattggg gcgtgcgagg gtggcggtc agcctggcgc tcgctgtctt cgcggctctgc 240
tcgttcggcg gcaactctcg gtaccactcg gagagcccac actaccgat ggcggttctc 300
atctgctcgc tcaactttgt ctacatctcc actacgttca ccaagaagcc agagtccaac 360
gcgtgcgggg agtggcccg gctcgcgag ctgcgcatct tgcccacat gtttgagcgc 420
ttcttcggcc tgcaggctct gctcaccgac ggtgccaagc gcgtcgcgca catgctgggc 480
gacgagtcga gcgcagaccc gcggatgcgc caggtaatgc tcctcttcca cccgcacagc 540
atcttcccag tctcgcacgc ggcgctgggt ctcaactctc tctggcgctc gcactttccc 600
cacctctcgg tcaaccccct aacagcgcgc attatccact ttgtgcccgt catgcccgcac 660
gttttgacgt ggctcggcat ctgcgacgct tccaaagcga gcgtgggtcaa cctcatcgcc 720
atggggcgca acgtccagat cgtgtgcggc gggcagaccg agatgttcga gtcccctcc 780
tgggacaagg agatttctgt ggtgcggcgc cgcgccttg gcgttttcaa gatcgcctc 840
cagcagggcc tcggtatcgt gccgatttac agcttcggag agccgctcac ctttgacaac 900
atatacatgc cccgcttgca aaacttttgc aagcgcgtgc tcggettccc ctgcccgttc 960
gtgatgctcg gtcagtaagg ccttcccatt ccgcgccgcg tcccaatttc ggtggtgtt 1020
ggcgagcccg tctttctcgc tcggcagacc gccgatcctt cgctcgagga ggtcaaagag 1080
tttcacagac gttactttga ggccctgcag gccctgttg accagttcaa ggaccaggcc 1140
gggcacggcc agttagcat caagtggctg gactcgtag 1179

```

<210> SEQ ID NO 50

<211> LENGTH: 392

<212> TYPE: PRT

<213> ORGANISM: *Thraustochytrium aureum*

<400> SEQUENCE: 50

```

Met Val Phe Leu Cys Leu Pro Tyr Met Leu Pro Glu Ala Leu Leu Pro
1           5           10           15
Phe Leu Asp Thr Ala Thr Leu Gly Leu Ile Pro Ala Leu Pro Gly Asp
20          25          30
Lys Glu Asn Phe Val His Thr Phe Ala Val Trp Trp Thr Leu Leu Trp
35          40          45
Ala Ile Ala Phe Trp Thr Ile Phe Tyr Ala Ala Leu Lys Asn Trp Gly
50          55          60
Val Arg Gly Trp Arg Leu Ser Leu Ala Leu Ala Val Phe Ala Val Cys
65          70          75          80
Ser Phe Gly Gly Thr Leu Arg Tyr His Ser Glu Ser Pro His Tyr Pro
85          90          95
Met Ala Val Leu Ile Cys Ser Leu Asn Phe Val Tyr Ile Ser Thr Thr
100         105         110
Phe Thr Lys Lys Pro Glu Ser Asn Ala Cys Arg Glu Trp Pro Glu Leu
115        120        125
Arg Glu Leu Arg Ile Leu Pro Asp Met Phe Glu Arg Phe Phe Gly Leu
130        135        140
Gln Val Leu Leu Thr Asp Gly Ala Lys Arg Val Ala His Met Leu Gly
145        150        155        160
Asp Glu Ser Ser Ala Asp Pro Arg Met Arg Gln Val Met Leu Leu Phe
165        170        175

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His Pro His Ser Ile Phe Pro Val Ser His Ala Ala Leu Gly Leu Thr
 180 185 190

Ser Leu Trp Arg Ser His Phe Pro His Leu Ser Val Asn Pro Leu Thr
 195 200 205

Ala Ser Ile Ile His Phe Val Pro Val Met Arg Asp Val Leu Gln Trp
 210 215 220

Leu Gly Ile Cys Asp Val Ser Lys Ala Ser Val Val Asn Leu Ile Gly
 225 230 235 240

Met Gly Arg Asn Val Gln Ile Val Cys Gly Gly Gln Thr Glu Met Phe
 245 250 255

Glu Ser Arg Ser Trp Asp Lys Glu Ile Ser Val Val Arg Ala Arg Arg
 260 265 270

Leu Gly Val Phe Lys Ile Ala Ile Gln Gln Gly Leu Gly Ile Val Pro
 275 280 285

Ile Tyr Ser Phe Gly Glu Pro Leu Thr Phe Asp Asn Ile Tyr Met Pro
 290 295 300

Arg Leu Gln Asn Phe Cys Lys Arg Val Leu Gly Phe Pro Cys Pro Phe
 305 310 315 320

Val Met Leu Gly Gln Tyr Gly Leu Pro Ile Pro Arg Arg Val Pro Ile
 325 330 335

Ser Val Ala Val Gly Glu Pro Val Phe Pro Ala Arg Gln Thr Ala Asp
 340 345 350

Pro Ser Leu Glu Glu Val Lys Glu Phe His Arg Arg Tyr Phe Glu Ala
 355 360 365

Leu Gln Ala Leu Phe Asp Gln Phe Lys Asp Gln Ala Gly His Gly Gln
 370 375 380

Cys Ser Ile Lys Trp Leu Asp Ser
 385 390

<210> SEQ ID NO 51
 <211> LENGTH: 1303
 <212> TYPE: DNA
 <213> ORGANISM: Thraustochytrium aureum

<400> SEQUENCE: 51

```

agctttacct gctacatggt cttcctctgc cttccctaca tgetccccga agcgctgctc    60
cctttcttgg acacggcgac gctaggcctc atccccggccc tgccccggaga caaggagaac    120
tttgccaca cgtttgccgt gtggtggacg ctcttggtgg cgattgcggt ttggacgatc    180
ttttacgccc cgctcaagaa ttggggcgctg cgaggggtggc ggctcagcct ggcgctcgct    240
gtcttcgccc tctgctcggt cggcggcact ctgcggtaac actcggagag cccacactac    300
ccgatggcgg ttctcatctg ctcgctcaac tttgtctaca tctccactac gttcaccaag    360
aagccagagt ccaacgctg cggggagtgg cccgagctgc gcgagctgcg catcttgccc    420
gacatgtttg agcgcttctt cggcctgcag gtcctgctca ccgacggtgc caagcgcgctc    480
gcgccatgac tgggacgacg gtcgagcgca gaccccgcca tggcgcaggt aatgctcctc    540
ttccaccgac acagcatctt cccagtctcg cacgcggcgc tgggtctcac ttcgctctgg    600
cgctcgcact tccccacact ctcggtcaac ccctaacag cgagcattat ccactttgtg    660
ccggtcatgc ggcagctttt cgagtggctc ggcactctgc acgtctcaa agcgagcgtg    720
gtcaacctca tcggcatggg gcgcaacgct cagatcgtgt gcggcgggca gaccgagatg    780
    
```

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ttcgagtccc gctcctggga caaggagatt tctgtggtgc gggcgcgccg ccttgcgctt 840
ttcaagatcg ccatccagca gggcctcggg atcgtgccga tttacagctt cggagagccg 900
ctcacctttg acaacatata catgcccgcg ttgcaaaact tttgcaagcg cgtgctcggc 960
ttcccctgcc cgttcgtgat gctcggtcag tacggccttc ccattccgcg ccgctgcccc 1020
atttcggtgg ctgttgccga gcccgctttt cctgctcggc agaccgcccga tccttcgctc 1080
gaggagggtca aagagtttca cagacgttac tttgaggccc tgcaggccct gtttgaccag 1140
ttcaaggacc aggccgggca cggccagtgt agcatcaagt ggctggactc gtagaggcag 1200
aaagcccgcg gcactgcttt tgcgcctgtg ccgttcccg tttgtagaac aaccttccaa 1260
cattcgttag ctttctctta aaaaaaaaaa aaaaaaaaaa aaa 1303

```

<210> SEQ ID NO 52

<211> LENGTH: 1389

<212> TYPE: DNA

<213> ORGANISM: *Thraustochytrium aureum*

<400> SEQUENCE: 52

```

atgtttcttc gcacgaacg ggaatggcga gaggaggacg agtgggccc aagcaggagccc 60
ggcgttgtct ccacgatgat ctggacccc atcctgatcg ggctccgctg cttcaacatc 120
tggtctcccg tggttacctg gccgctctcg tttctggctc gcgtcgtttt cggcatggag 180
atgaagaagg cgagcttctg ggacgtccct ctggagcggc gcaagcagac ggtggcagtt 240
gcggttctcg tgatgtgct cccctgcgtg ctgcttgcgt acgtctggtc gcttgtgctg 300
ctcgttttcc cgctgacgac gctgccaatg ctccggctact acatctggat cttcaagatc 360
gacaagagcc ccgagaacgg gcagcgcacg ccgttctctg gttactggtc ggctggcgcc 420
cacttcgctt cctacttccc gctgcgcctc atcaagacgc acaacctcga cccgagccgc 480
aagtacgtct tcgctgacca cccgcaacgc atcatcagca ttggcgcgct cggcaacttt 540
gccaccaacg cgacgggggt tagccgcaag tttcccggaa tcgacctccg cctcctcacc 600
ttggaaatga acttttggtg cccctggatc cgcgagttcc tgctgagcat gggcgtctgc 660
tcagccgcca agcgggtcctg caacaagatt ctcaagcagg ggcccggaa gcccacatg 720
ctggtcgttg gcggcggccc cgagtcgctc gacacggagc ccggcaccta caggctcacc 780
ttggggccca agggctttat ccgctgcgct ctcgacaacg gggccgacct cgtgctctgt 840
ctgccttctg gggagaacga catctttgac accatctact acgagtcagg caccgtgatg 900
cgcaagatcc aggaggtcgt gcgcaagcgc ctccgctttg ccacccctgt ttttccggc 960
cgcggttct tcaactacag ctttggtctc ctcccgcacc gggcccgggt cattgtcgtc 1020
tgccggcgcc ctatcaaggt cccaaaaact ccggaacacc tgccgggctc ggcgctctcg 1080
accacccctg aaggcgtcgc gcttgtcgac cagtaccacc aaaagtaagt cgcgagctg 1140
cgcccgctgt gggacctcta caagtccaag tgggcccgtc cgcgggcaga gtcgctcatg 1200
atcaagggtg tgcaaaaccc ggcctcccg cgctccccgt cgcgcccgat cccgcccggc 1260
cagcgcgttc ccgagagtgc cgcctcgtt tcgtttcgcg aggtcgacga ggccgaattt 1320
gaggccaagg aggacggcgc gacctcttcg ccgcagtcga tgtctgccc gctgtacacc 1380
gagggttag 1389

```

<210> SEQ ID NO 53

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```

<211> LENGTH: 462
<212> TYPE: PRT
<213> ORGANISM: Thraustochytrium aureum

<400> SEQUENCE: 53

Met Phe Leu Arg Ile Glu Arg Glu Trp Arg Glu Glu Asp Glu Trp Ala
1           5           10           15
Lys Gln Glu Pro Gly Val Val Ser Thr Met Ile Trp Thr Pro Ile Leu
20           25           30
Ile Gly Leu Arg Cys Phe Asn Ile Trp Leu Ser Val Val Thr Trp Pro
35           40           45
Leu Ser Phe Leu Ala Arg Val Val Phe Gly Met Glu Met Lys Lys Ala
50           55           60
Ser Phe Trp Asp Val Pro Leu Glu Arg Arg Lys Gln Thr Val Ala Val
65           70           75           80
Ala Gly Phe Val Met Leu Leu Pro Cys Val Leu Leu Ala Tyr Val Trp
85           90           95
Ser Leu Val Leu Leu Val Phe Pro Leu Thr Thr Leu Pro Met Leu Gly
100          105          110
Tyr Tyr Ile Trp Ile Phe Lys Ile Asp Lys Ser Pro Glu Asn Gly Gln
115          120          125
Arg Thr Pro Phe Leu Arg Tyr Trp Ser Ala Trp Arg His Phe Ala Ser
130          135          140
Tyr Phe Pro Leu Arg Leu Ile Lys Thr His Asn Leu Asp Pro Ser Arg
145          150          155          160
Lys Tyr Val Phe Ala Tyr His Pro His Gly Ile Ile Ser Ile Gly Ala
165          170          175
Phe Gly Asn Phe Ala Thr Asn Ala Thr Gly Phe Ser Arg Lys Phe Pro
180          185          190
Gly Ile Asp Leu Arg Leu Leu Thr Leu Glu Met Asn Phe Trp Cys Pro
195          200          205
Trp Ile Arg Glu Phe Leu Leu Ser Met Gly Val Cys Ser Ala Ala Lys
210          215          220
Arg Ser Cys Asn Lys Ile Leu Ser Lys Gly Pro Gly Ser Ala Ile Met
225          230          235          240
Leu Val Val Gly Gly Ala Ala Glu Ser Leu Asp Thr Glu Pro Gly Thr
245          250          255
Tyr Arg Leu Thr Leu Gly Arg Lys Gly Phe Ile Arg Val Ala Leu Asp
260          265          270
Asn Gly Ala Asp Leu Val Pro Val Leu Ala Phe Gly Glu Asn Asp Ile
275          280          285
Phe Asp Thr Ile Tyr Tyr Glu Ser Gly Thr Val Met Arg Lys Ile Gln
290          295          300
Glu Val Val Arg Lys Arg Leu Gly Phe Ala Thr Pro Val Phe Ser Gly
305          310          315          320
Arg Gly Phe Phe Asn Tyr Ser Phe Gly Phe Leu Pro His Arg Arg Pro
325          330          335
Val Ile Val Val Cys Gly Arg Pro Ile Lys Val Pro Lys Leu Pro Glu
340          345          350
His Leu Arg Gly Ser Ala Leu Ser Thr Thr Pro Glu Gly Val Ala Leu
355          360          365
Val Asp Gln Tyr His Gln Lys Tyr Val Ala Glu Leu Arg Arg Val Trp

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370	375	380														
Asp	Leu	Tyr	Lys	Ser	Lys	Trp	Ala	Val	Ser	Arg	Ala	Glu	Ser	Leu	Met	
385					390					395					400	
Ile	Lys	Gly	Val	Gln	Asn	Pro	Ala	Leu	Pro	Arg	Ser	Pro	Ser	Arg	Arg	
				405					410					415		
Ile	Pro	Pro	Ala	Gln	Arg	Val	Pro	Ala	Ser	Ala	Ala	Ser	Leu	Ser	Phe	
			420					425					430			
Arg	Glu	Val	Asp	Glu	Ala	Glu	Phe	Glu	Ala	Lys	Glu	Asp	Gly	Ala	Thr	
		435					440						445			
Ser	Ser	Pro	Gln	Ser	Met	Ser	Ala	Ala	Leu	Tyr	Thr	Glu	Gly			
	450					455					460					

<210> SEQ ID NO 54

<211> LENGTH: 1547

<212> TYPE: DNA

<213> ORGANISM: Thraustochytrium aureum

<400> SEQUENCE: 54

```

aggctgacct gcgaagagcg cgagatgttt ctctgcatcg aacgggaatg gcgagaggag    60
gacgagtggg ccaagcagga gccccggcgtt gtctccaacga tgatctggac cccgatcctg    120
atcgggctcc gctgcttcaa catctggctc tccgtgggta cctggccgct ctctttctg    180
gctcgcgtcg ttttcggcat ggagatgaag aaggcgagct tctgggacgt cctctggag    240
cggcgcaagc agacgggtgc agttgcgggc ttcgtgatgc tgctcccctg cgtgctgctt    300
gcgtaactct ggtcgtttgt gctgctgctt tccccgctga cgacgctgcc aatgctcggg    360
tactacatct ggatcttcaa gatcgacaag agccccgaga acgggcagcg cacgcccgtt    420
ctgcgttact ggtcggcgtg gcgccacttc gcctcctact tcccgtgctg cctcatcaag    480
acgcacaacc tcgacccgag ccgcaagtac gtcttgcgct accaccgca cggcatcatt    540
agcattggcg cgttcggcaa ctttgccacc aacgcgacgg ggtttagccg caagtttccc    600
ggaatcgacc tcgcctcct caccttgaa atgaactttt ggtgcccctg gatccgagag    660
ttctctgctg gcatgggctg ctgctcagcc gccaaagcgt cctgcaacaa gattctcagc    720
aaggggcccg gaagcgccat catgctggtc gttggcggcg ccgcgagtc gctcgacacg    780
gagcccgcca cctacaggct cacgttgggc cgcaagggtt ttatccgctg cgcgctcgac    840
aacggggcgc acctcgtgcc tgtgctgccc ttcggggaga acgacatctt tgacaccatc    900
tactacgagt ccggcaccgt gatgcgcaag atccaggagg tctgctgcaa gcgcccctgc    960
tttgccacc ctgttttttc cggcccgggc ttcttcaact acagctttgg ctctcctccc    1020
caccggcgcc cggtcattgt cgtctgctgg cgccctatca aggtcccaaa actcccggaa    1080
cactctgctg gctcggcgtc ctgaccacc cctgaaggcg tctcgttctg cgaccagtac    1140
caccaaaaag acgtcgccga gctgcgcgcg gtgtgggacc tctacaagtc caagtgggccc    1200
gtctcggcgg cagagctcgt catgatcaag ggtgtgcaaa acccggcgct cccgctctcc    1260
ccgtcgcgcc gcacctccgc ggccgagcgc gttcccgcga gtgcccctc gctttctggtt    1320
cgcgaggtcg acgaggccga atttgaggcc aaggaggacg gcgcgacctc ttcccgcag    1380
tccatgtctg ccgctctgta caccgaggtt tagtctctat cagcttgccg atctcgccat    1440
cccgccctg cctcgcgtcc gcgcccggcg agttttgtca tgcaccagcg ccttctggtt    1500
gttgaagtaa caaacgtaaa cgttttttct ttctttcaaa aaaaaaa    1547

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-continued

<210> SEQ ID NO 55

<211> LENGTH: 1977

<212> TYPE: DNA

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 55

atgccatccc gcagcaccat tgaggctcatt aaggccgata agaaccagaa taatctggcg 60
tatggcctga ttgttgcat cctcctggcc attgaeccca acccgcgtaa agtcatcgcc 120
gcctctctcg gcactcccct tcgatggttc gcctaccctt gcctgggtcat gcttggccac 180
ctattctctca cccactccca ggaattttctc tacgacggcg tcgggtctt cttccgctcc 240
atcctttcga tcttcttccg tcaagtgcac attgtgggca tcgacaacat cccgaaacac 300
ggcctgtca tcttctcccg gaaccactcg aaccaatttg tcgacgggat catggtcctc 360
accaccgccc aacaccgctt cggtctcctt atcgccgaaa agtctacaa ccaccctggt 420
gtcggcacat ttgcaaaact cgcggggcgc gtgcccgtca cccgccctca agacagcgct 480
aagctcatgc aaggtaccat tatcatgtcc ggccgctctg tcaagggaca aggaaccgcc 540
tttagtcacg agctcgtccc cggcgacaag ctacgtctaa aaggtggtgc tgatcaatte 600
aaagtcgagt ccatcacctc cgataccgag ctgatgctct cggagaacgg ccccttctct 660
ccccctctct ctacctcccg ctccgccctt gaaaaactag ggaaggtgga ccagaccctg 720
gtctacaatg ccgtgttcga gcaccttaag cacgggaaat gcacggtat cttccccgaa 780
ggcggctcgc acgateggac agacctccta cccctcaagg tagggattgc actcatcgcc 840
tgcggcatgg tcgataaata caatatcaca gtgcccatcg tccccgtggg tttgaactac 900
ttccgaggcc accggtttctg tggacgggtg gtagtagaat tcgggcccagc aattcgcgtg 960
ccggaagagt tggcggagtt gtacaagacc aatcgacgcg aggcgatca ccagtttctg 1020
accaacgtgg aggaagggat cggggcgacg cttgtgacgg cgcctgatta ccacgcgttg 1080
catttggtgt acacggcacc gaggttattt cagaaggata attggattcc gagcccacgg 1140
gagaagatgg atttgaaccg gcggtttgcc gaggggtata aaattttgat gaataagtat 1200
ggggagcaga ggcggcgccg gttggtggag ttggagagga ggttgaatga ttacaaaaa 1260
actctgcata cgttgggttt gagggattac caagtgccga cgttgagga ggatgataac 1320
ttaaagtgtg gttacacgat agcgcatttg tttttggtgt tgacgctggc gatgatgccg 1380
agcttggtgt tgaacgcgcc ggtggggttg attgcccgga ttgtttcgag tcgggagcag 1440
aaaaaggcct tggcggcgct cgggtaaaag atcgaggcga gggatgtggt tatgagcaaa 1500
aaaatcacgt tgtcgattgt cttggttccg accctatgga tcgtgtacgc catcctcctc 1560
cttcggtaca cctccctcca gccctccacc gtcgcccgtc tcttcttctc ctgtcccctc 1620
tttctctatc ttgggtcat ggcacagaa gctggcatgg ttgacgcaa ggatctcaaa 1680
cccgtcgtta tgcgtctttt acccggagct cgtaagaaaa tggcgaccct cctgcgagg 1740
cgcgcgacgc tacaagaga aatccgcgcc tacatacacc agatcggccc tgaacttggg 1800
agtctctaca ccgacaaaac cgtcaagtgg gaagaatcgc tccgcaagtc ctcatcggcg 1860
gctgacttgc aatcgttggt gaacgaagcg acccaacca agatgcaagg aagtcagacg 1920
gaaggaggga atggtggaga aaaaggggga aggaaggggg aagaggagct tgtctga 1977

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<210> SEQ ID NO 56
<211> LENGTH: 658
<212> TYPE: PRT
<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 56
Met Pro Ser Arg Ser Thr Ile Glu Val Ile Lys Ala Asp Lys Asn Gln
1      5      10
Asn Asn Leu Ala Tyr Gly Leu Ile Val Val Ile Leu Leu Ala Ile Asp
20     25     30
Pro Asn Pro Val Lys Val Ile Ala Ala Ser Leu Gly Ile Pro Ser Arg
35     40     45
Trp Phe Ala Tyr Pro Cys Leu Val Met Leu Gly His Leu Phe Leu Thr
50     55     60
His Ser Gln Glu Phe Leu Tyr Asp Gly Val Arg Val Phe Phe Arg Ser
65     70     75     80
Ile Leu Ser Ile Phe Phe Arg Gln Val Asp Ile Val Gly Ile Asp Asn
85     90     95
Ile Pro Lys His Gly Pro Val Ile Phe Ser Gly Asn His Ser Asn Gln
100    105    110
Phe Val Asp Gly Ile Met Val Leu Thr Thr Ala Gln His Arg Val Gly
115    120    125
Phe Leu Ile Ala Glu Lys Ser Tyr Asn His Pro Val Val Gly Thr Phe
130    135    140
Ala Lys Leu Ala Gly Ala Val Pro Val Thr Arg Pro Gln Asp Ser Ala
145    150    155    160
Lys Leu Met Gln Gly Thr Ile Ile Met Ser Gly Arg Ser Val Lys Gly
165    170    175
Gln Gly Thr Ala Phe Ser His Glu Leu Val Pro Gly Asp Lys Leu Arg
180    185    190
Leu Lys Gly Gly Ala Asp Gln Phe Lys Val Glu Ser Ile Thr Ser Asp
195    200    205
Thr Glu Leu Met Leu Ser Glu Asn Gly Pro Leu Pro Pro Pro Ser Ser
210    215    220
Thr Ser Ala Ser Pro Phe Glu Lys Leu Gly Lys Val Asp Gln Thr Arg
225    230    235    240
Val Tyr Asn Ala Val Phe Glu His Leu Lys His Gly Lys Cys Ile Gly
245    250    255
Ile Phe Pro Glu Gly Gly Ser His Asp Arg Thr Asp Leu Leu Pro Leu
260    265    270
Lys Val Gly Ile Ala Leu Ile Ala Cys Gly Met Val Asp Lys Tyr Asn
275    280    285
Ile Thr Val Pro Ile Val Pro Val Gly Leu Asn Tyr Phe Arg Gly His
290    295    300
Arg Phe Arg Gly Arg Val Val Val Glu Phe Gly Pro Ala Ile Arg Val
305    310    315    320
Pro Glu Glu Leu Ala Glu Leu Tyr Lys Thr Asn Arg Arg Glu Ala Tyr
325    330    335
His Gln Phe Leu Thr Asn Val Glu Glu Gly Met Arg Ala Thr Leu Val
340    345    350
Thr Ala Pro Asp Tyr His Ala Leu His Leu Val Tyr Thr Ala Arg Arg
355    360    365

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Leu Phe Gln Lys Asp Asn Trp Ile Pro Ser Pro Arg Glu Lys Met Asp
 370 375 380

Leu Asn Arg Arg Phe Ala Glu Gly Tyr Lys Ile Leu Met Asn Lys Tyr
 385 390 395 400

Gly Glu Gln Arg Pro Ala Ala Leu Val Glu Leu Glu Arg Arg Leu Asn
 405 410 415

Asp Tyr Gln Lys Thr Leu His Thr Leu Gly Leu Arg Asp Tyr Gln Val
 420 425 430

Pro Thr Leu Glu Glu Asp Asp Asn Leu Lys Leu Cys Tyr Thr Ile Ala
 435 440 445

His Leu Phe Leu Val Leu Thr Leu Ala Met Met Pro Ser Leu Val Leu
 450 455 460

Asn Ala Pro Val Gly Leu Ile Ala Arg Ile Val Ser Ser Arg Glu Gln
 465 470 475 480

Lys Lys Ala Leu Ala Ala Ser Arg Val Lys Ile Glu Ala Arg Asp Val
 485 490 495

Val Met Ser Lys Lys Ile Thr Leu Ser Ile Val Leu Val Pro Thr Leu
 500 505 510

Trp Ile Val Tyr Ala Ile Leu Leu Arg Tyr Thr Ser Leu Gln Pro
 515 520 525

Ser Thr Val Ala Val Leu Phe Phe Ser Cys Pro Leu Phe Ser Tyr Leu
 530 535 540

Gly Val Met Ala Thr Glu Ala Gly Met Val Asp Ala Lys Asp Leu Lys
 545 550 555 560

Pro Val Val Met Arg Leu Leu Pro Gly Ala Arg Lys Lys Met Ala Thr
 565 570 575

Leu Pro Ala Glu Arg Ala Gln Leu Gln Arg Glu Ile Arg Ala Tyr Ile
 580 585 590

His Gln Ile Gly Pro Glu Leu Gly Ser Leu Tyr Thr Asp Lys Thr Val
 595 600 605

Lys Trp Glu Glu Tyr Val Arg Lys Ser Ser Ser Ala Ala Asp Leu Gln
 610 615 620

Ser Leu Leu Asn Glu Ala Thr Gln Pro Lys Met Gln Gly Ser Gln Thr
 625 630 635 640

Glu Gly Gly Asn Gly Gly Glu Lys Gly Gly Arg Lys Gly Glu Glu Glu
 645 650 655

Leu Val

<210> SEQ ID NO 57
 <211> LENGTH: 2460
 <212> TYPE: DNA
 <213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 57

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attttcagca aagtaatcaa gataataaac aaaaacaatc ctataaagga aaaacaacag    60
gcacgcgtcc tgaggtgccc gtgcctgtaa ttttctcctc tgggactgtc ggccatcgtc    120
aggaacaagc gcggccacca gggctcattt cgaatcaagc acatccgttc cacaccggg    180
caacaaaacc atgccatccc gcagcaccat tgaggtcatt aaggccgata agaaccagaa    240
taatctggcg tatggcctga ttgtgtcat cctcctggcc attgacccca acccctgtaa    300
agtcacgcc gcctctctcg gcateccctc tcgatggttc gcctaccctc gcctgggtcat    360
    
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gcttggccac ctattcctca cccactccca ggaatttctc tacgacggcg tccgggtctt	420
cttccgctcc atcctttcga tcttcttccg tcaagtcgac attgtgggca tcgacaacat	480
cccgaaacac ggcctctgca tcttctcggg gaaccactcg aaccaatttg tcgacgggat	540
catggtcctc accacegccc aacaccgegt cggcttcctt atcgccgaaa agtcctacaa	600
ccaccctggt gtcggcacat ttgcaaaaact cgcggggcgc gtgcccgtca cccgcctca	660
agacagcgtc aagctcatgc aaggtaccat tatcatgtcc ggccgctctg tcaagggaca	720
aggaaccgcc tttagtcacg agctcgtccc cggcgacaag ctacgtctaa aaggtggtgc	780
tgatcaattc aaagtcgagt ccatcacctc cgataccgag ctgatgctct ccgagaacgg	840
cccccttct cccccctct ctacctccgc ctgcctctt gaaaaactag ggaagtgga	900
ccagaccctg gtctacaatg ccgtgttcga gcaccttaag cacgggaaat gcatcgggat	960
cttccccgaa ggcggctcgc acgatcggac agacctccta cccctcaagg tagggattgc	1020
actcatcgcc tgcggcatgg tcgataaata caatcacaca gtgcccctcg tccccgtggg	1080
tttgaactac ttcgaggcc accggtttcg tggacgggtg gtagtagaat tggggccagc	1140
aattcgcgtg ccggaagagt tggcggagtt gtacaagacc aatcgacgcg aggcgtatca	1200
ccagtttctg accaacgtgg aggaagggat gcgggcgacg cttgtgacgg cgcctgatta	1260
ccacgcgttg catttgggtg acacggcacg gaggttattt cagaaggata attggattcc	1320
gagccccgag gagaagatgg atttgaaccg gcggtttgcc gaggggtata aaattttgat	1380
gaataagtat ggggagcaga ggcgggcggc gttggtggag ttggagagga ggttgaatga	1440
ttacaaaaaa actctgcata cgttggggtt gagggattac caagtgccga cgttgaggga	1500
ggatgataac ttaaaattgt gttcacacgat agcgcatttg tttttggtgt tgacgctggc	1560
gatgatgccg agcttgggtg tgaacgcgcc ggtgggggtg attgcccgga ttgtttcgag	1620
tcgggagcag aaaaaggcct tggcggcgtc ccgggtaaag atcgaggcga gggatgtggt	1680
tatgagcaaa aaaatcacgt tgtcgtattg cttggttccg accctatgga tcgtgtacgc	1740
catctctc cttcggta caacctccca gccctccacc gtcgcccgtc tcttcttctc	1800
ctgtcccctc ttttctatc ttgggggtat ggccacagaa gctggcatgg ttgacgcaa	1860
ggatctcaaa cccgctgcta tgcgtctttt acccgagct cgtaagaaaa tggcgaccct	1920
ccctgcggag cgcgcgcagc tacaagaga aatccgcgcc tacatacacc agatcggccc	1980
tgaacttggg agtctctaca ccgacaaaac cgtcaagtgg gaagaatacg tccgcaagtc	2040
ctcatcggcg gctgacttgc aatcgttgtt gaacgaagcg acccaacca agatgcaagg	2100
aagtcagacg gaaggaggga atggtggaga aaaaggggga aggaaggggg aagaggagct	2160
tgtctgatac gtcaccgaaa ttgtcgcacg cgatgaatgg aagagagacg ccgccaccag	2220
ttaagatgac tcaaaaccgg ctggtgacgg ggaagaagga tgcataaggag ggattatgag	2280
ggagggaggg caggggtgat gagttagaat tcgatgcaca tagagaagga tgttctctggc	2340
tgggactgta aattggttag ggtaaatatt gtgtgtgctg catcgtcttt gtcacgtacg	2400
tgaaggaaa cggaaaggaa aaaaagtgga atacaagaca aaaaaaaaaa aaaaaaaaaa	2460

<210> SEQ ID NO 58

<211> LENGTH: 104

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

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<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 58

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccggate ggcgcccac catggacaag gcactggcac cggt 104

<210> SEQ ID NO 59

<211> LENGTH: 101

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 59

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaact aaactttcct ccttccctct a 101

<210> SEQ ID NO 60

<211> LENGTH: 104

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 60

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccggate ggcgcccac catgaccacg actgtcatct cttag 104

<210> SEQ ID NO 61

<211> LENGTH: 101

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 61

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaatc aaagcctccc gcacaacgag c 101

<210> SEQ ID NO 62

<211> LENGTH: 104

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 62

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccggate ggcgcccac catggagggc atcgagtcca tagt 104

<210> SEQ ID NO 63

<211> LENGTH: 101

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 63

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60

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tagagcggat ttaattaact ataaggcttc tcccggcgcg g 101

<210> SEQ ID NO 64
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 64

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60

accccggatc ggcgcgccac catgaagacg cccacgagcc tggc 104

<210> SEQ ID NO 65
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 65

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60

tagagcggat ttaattaatt aagctctcga atcgctcttc t 101

<210> SEQ ID NO 66
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 66

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60

accccggatc ggcgcgccac catggtcagg aggaagatgg acgt 104

<210> SEQ ID NO 67
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 67

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60

tagagcggat ttaattaatc acgacgcgg cgcttgcag t 101

<210> SEQ ID NO 68
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 68

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60

accccggatc ggcgcgccac catggcacc ccccaccg cccc 104

<210> SEQ ID NO 69
<211> LENGTH: 101
<212> TYPE: DNA

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<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 69

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcett ccttttcggt 60
tagagcggat ttaattaatc atttgaccac taaggtggcc t 101

<210> SEQ ID NO 70
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 70

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccgatc ggcgcgccac catgggtcta tttggcagcg ggat 104

<210> SEQ ID NO 71
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 71

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcett ccttttcggt 60
tagagcggat ttaattaact aaaagaaatt caacgtccga t 101

<210> SEQ ID NO 72
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 72

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccgatc ggcgcgccac catgttgagt atcccgagt cgtc 104

<210> SEQ ID NO 73
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 73

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcett ccttttcggt 60
tagagcggat ttaattaact aaaagaaatc cagctccctg t 101

<210> SEQ ID NO 74
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 74

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ataaaagtat caacaaaaaa ttgtaatat acctctatac ttaacgtca aggagaaaa 60
accccgatc ggcgcccac catgacgcc caagccgata tcac 104

<210> SEQ ID NO 75
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 75

aactataaaa aaataaatag ggacctagac ttcaggtgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaatt actcaatgga caacggggcg g 101

<210> SEQ ID NO 76
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 76

ataaaagtat caacaaaaaa ttgtaatat acctctatac ttaacgtca aggagaaaa 60
accccgatc ggcgcccac catggcttac ctctccgtc gtcg 104

<210> SEQ ID NO 77
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 77

aactataaaa aaataaatag ggacctagac ttcaggtgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaatt aggcgatcgc aatgaactcc t 101

<210> SEQ ID NO 78
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 78

ataaaagtat caacaaaaaa ttgtaatat acctctatac ttaacgtca aggagaaaa 60
accccgatc ggcgcccac catgcctttt ggacgggctg catc 104

<210> SEQ ID NO 79
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 79

aactataaaa aaataaatag ggacctagac ttcaggtgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaatc acccgaaaat atcctccttc t 101

<210> SEQ ID NO 80

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<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 80

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaa 60
accccgatc ggcgcccac catggccaag gctaacttc cgcc 104

<210> SEQ ID NO 81
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 81

aactataaaa aaataaatag ggacctagac ttcaggtgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaatc actttataag cagcttcttg t 101

<210> SEQ ID NO 82
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 82

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaa 60
accccgatc ggcgcccac catgttcttg cagggattaa gctg 104

<210> SEQ ID NO 83
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 83

aactataaaa aaataaatag ggacctagac ttcaggtgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaatc acaacaggac caatttatga t 101

<210> SEQ ID NO 84
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 84

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaa 60
accccgatc ggcgcccac catgttgatg ggcgctgcg ggcg 104

<210> SEQ ID NO 85
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

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<400> SEQUENCE: 85

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaatc agacgatgcg aagcgtcttg t 101

<210> SEQ ID NO 86

<211> LENGTH: 104

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 86

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccgatc ggcgcgccac catggcgctt accactgcga ccca 104

<210> SEQ ID NO 87

<211> LENGTH: 101

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 87

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaatc acgacttcgg acagtccaaa a 101

<210> SEQ ID NO 88

<211> LENGTH: 104

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 88

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccgatc ggcgcgccac catgtcgttc gttgagcaca ggc 104

<210> SEQ ID NO 89

<211> LENGTH: 101

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 89

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaact acacaaatcg catcgtcttg t 101

<210> SEQ ID NO 90

<211> LENGTH: 104

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 90

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccgatc ggcgcgccac catggtcttc ctctgccttc ccta 104

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<210> SEQ ID NO 91
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 91

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaact acgagtcag ccacttgatg c 101

<210> SEQ ID NO 92
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 92

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccgatc ggcgcgccac catgtttctt cgcacgaac ggga 104

<210> SEQ ID NO 93
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 93

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaact aaccctcggg gtacagcgcc g 101

<210> SEQ ID NO 94
<211> LENGTH: 104
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 94

ataaaagtat caacaaaaaa ttgtaatat acctctatac tttaacgtca aggagaaaaa 60
accccgatc ggcgcgccac catgccatcc cgcagcacca ttga 104

<210> SEQ ID NO 95
<211> LENGTH: 101
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer

<400> SEQUENCE: 95

aactataaaa aaataaatag ggacctagac ttcaggttgt ctaactcctt ccttttcggt 60
tagagcggat ttaattaatc agacaagtc ccttcccc t 101

<210> SEQ ID NO 96
<211> LENGTH: 1197
<212> TYPE: DNA
<213> ORGANISM: Phytophthora sojae

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<400> SEQUENCE: 96

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gcactgtttt atgcagcaac ctttattgat cgtgctggtg cagcagccta tgttctgtgg    300
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cattcagcac tgctggttcc gtatcattct tggcgtatta gccatcgtaa acatcattca    480
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<210> SEQ ID NO 97

<211> LENGTH: 1371

<212> TYPE: DNA

<213> ORGANISM: *Ostreococcus tauri*

<400> SEQUENCE: 97

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ttggctaaga ccttcgctag aagatacgtg gttatcgagg gagttgagta cgatgtgacc    180
gatttcaaac accctggagg aaccgtgatt ttctacgctc tctetaacac tggagctgat    240
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cacaacaagc accacgctac tctcaaaaa gtgaggcagc atatggattt ggataccact    720
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aactacaagg tgatgactta tgctggagct tggaggcta ctttgggaaa cctcgataat	1320
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<210> SEQ ID NO 98

<211> LENGTH: 1320

<212> TYPE: DNA

<213> ORGANISM: Thraustochytrium sp.

<400> SEQUENCE: 98

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ccaggaggtt ccattattaa ttcctcacc gagggagaag ctggagtga tgctacccaa	180
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aagttggatg cttctaaggt ggagtctagg ttctctgcta aggagcaggc tagaagggac	300
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<210> SEQ ID NO 99

<211> LENGTH: 873

<212> TYPE: DNA

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<213> ORGANISM: *Physcomitrella patens*

<400> SEQUENCE: 99

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<210> SEQ ID NO 100

<211> LENGTH: 1086

<212> TYPE: DNA

<213> ORGANISM: *Phytophthora infestans*

<400> SEQUENCE: 100

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<210> SEQ ID NO 101
<211> LENGTH: 23777
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Plasmid

<400> SEQUENCE: 101
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acgtgcacaa	cagaattgaa	agcaaatatc	atgcatcat	aggcgtctcg	catatctcat	19500
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tgtccaacag	agtagaggtt	agcgaaggtg	gtagaaacag	cagaggtgta	tggaacatcg	19620
tagtaaggga	ggttatgtct	cttgaagaga	gcctcaactc	ttggagagat	ctccttgaac	19680
ctgaattgtg	gagcgggttg	gaacaaatga	tgctcgattt	ggaagttgag	gttagacatc	19740
caccaggtaa	ccaaccaaga	cttggtagag	atgttcacgg	tatgatcagc	agcgtactca	19800
agccaatgca	attgatcctc	tggtgttgta	actggcaaat	gggtatgaga	cacagcgaat	19860
tggaggaaga	tgtagatgca	tccaagtcgc	aaagagcaga	ggtacattcc	cacagaagtt	19920
ccaggagaat	atcccaaaagc	tcccatcaag	gagaaccatc	cgatataatc	agcgaagatc	19980
cacacaaact	ccatatgtct	cttggctctg	agcatatata	ttgggtgcaa	gtacaaggtc	20040
catcccaatc	cgatcaacaa	gcaagacact	ggagcgaaca	aataagcctg	aactctgagc	20100

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cacaaagcca	acaagatcc	tggetttaacc	ttcctcacia	ctctctcggt	gaaagcaacg	20160
agtggcaagg	tgttcaaate	cacatcgtgc	tccaatctgt	ttggagcagc	atggtgctta	20220
gaatgctgg	tcttccagta	gtgtccagac	attccacate	caactccgta	gaagaactcg	20280
cacatcctat	catcgagcca	gataactcca	gtgaaagac	cgtgtcccat	ctcatgcata	20340
accatccgc	atcttctctg	agcgattccg	ttcatcacca	ctccaaaac	caaagagggt	20400
ggagaagcct	tagacatcaa	ccagaaagac	aaagcgaaca	aagccacaat	ctccaccact	20460
ctgtagatca	tgtgtgggat	agatggatcg	aagtatccct	cagcaaccaa	ctcctctctg	20520
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gactccacct	tagaagcate	caactttggg	agggacttga	ggtacttate	agcctttccg	20640
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gctttaattt	ggattat	tattctctta	cettggccgt	tcattattcac	atccctaaag	21180
gcaagacaga	attgaaatgg	ggccaaaaat	taaaacgatg	gatatgacct	acatagtgt	21240
ggatcaatta	acgtcgaagg	aaaatactga	ttctattagg	ggtagagatt	gatcggttaa	21300
ttatccaata	catgccgttg	gttaattagg	attatataaa	aaatcgatca	tctattagaa	21360
tcgattacgg	ttaaatagg	ataaaaatgg	agagaattga	atcagttata	aatttgtttt	21420
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taatctaaaa	acattaatga	ggtaaagact	ctctcaaatg	ggatattctt	cgaaaatttt	21720
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actctttgac	accctatcag	atcccaacgt	tgtccctgg	ttcgaaaacca	ccatttcaaa	21840
catgaacata	tcacaaaata	aacattttaga	caccaaata	ctgctaattg	ccggcctaac	21900
ctgcaggat	cccggaatt	accggtagta	ggcgcctact	ttggccggcc	ctgaattaac	21960
gccgaattaa	ttcgggggat	ctggatttta	gtactggatt	ttggtttttag	gaattagaaa	22020
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acacatgagc	gaaaccctat	aggaacccta	attcccttat	ctgggaacta	ctcacacatt	22140
attatggaga	aactcgagct	tgctgatcac	tcggctcttag	ctcccttttg	ctttccatcg	22200
gatggcttga	tgtacttttg	cacgtagaag	tttccgaaga	ggaacaagag	ggagatcatg	22260
tagtagaaga	ggatcttgat	gagccattgt	ggatattggag	cgttggtttt	catatcgtag	22320
taagcttgca	ccaagttgag	catgaaactgg	aacatctgga	attgggtgag	gtatcttccc	22380

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cagaagaggt acttgttctt gagctttggg gaagatctca agcaagcagc caagaagtag 22440
taagcgtaca tcaacacgtg cactccagag ttgagagcag cactccaata agcctctcct 22500
cctggagcgt ggtgagcaat agcccaccag ataagggaga tagaagagtg gtggtacacg 22560
tggaggaaag aaatctgtct ggtggatctc ttgaggatca tgatecaggt atccatgaac 22620
tccacgtact tggacatgta gaagaggtaa acgaggatag ccatctcctt gtgctttggg 22680
ttataagcgt tccccacaaa ggaatatctc caggatagag cttggtaagc gataccacag 22740
cacatgtaaa gagacaaaagc gaagcagaac aagttgtgca ccaacaccaa agcttgcaac 22800
aagaatggct cagaagctct tggcttgaga tctctagcct tgatecaaaag caatcctccg 22860
atcacgatgg tcaagtaaac agacactccc aacacaattg gagttggaga atcaacgagt 22920
ggcaatcctc tagtagttgg ggtatcagtc aactcaactc cgaagatcc caacaaagcg 22980
ttcactcctt gggaaacctt tccatcaac tctccgtaga acctctcaac aacttccatg 23040
gtactggcta tgaagaaatt ataatcgtgt aaaacttagt gagtgtgtat gaatgaaagt 23100
attgcaaaat cctcattata tagactacat gcataactag ttgcatgtaa atttgtagtt 23160
ttcttcatta ttgcatcctc caagtggatg tcatggtttt acacatggct tccatgcaaa 23220
tcatttccaa aatattttta aactttccac agggcatcca tgcattgcacc tcaaaacttg 23280
tgtgtggtaa cattgtgtc ttgaaaaatt actaaacctt ttgtccacgt gacgttcatg 23340
cacctcaaat cttgtgtggt accattatta tcctcaagaa ttattgaatg tttggtgtat 23400
atgccatcca tgcagcattg caacaattaa atctccaaac cttgtggtac catattcact 23460
cactttaatt ctctatagt agaaatatta gcaaatattt acatttccag ttgattagta 23520
tatgtattta gaagacaaaa ataatttaga atcaattaat caacttgcaa attgctaagt 23580
gttggaacac gttagcataa aaggtgttat aaatttagta ccaaatataa aaatttatcg 23640
caaatcaaat acataacaca catagtaaaa caaaaacaaa ttacaagggt ttagacgttt 23700
agtggcaatg tgtaaatttg ctgcagggcg cgaattggc cttagtggcc aagcttggcg 23760
taatcatggc aactttt 23777

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<210> SEQ ID NO 102

<211> LENGTH: 960

<212> TYPE: DNA

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 102

```

atggccgcca tctcaccgcg caaacatcct cgcctgato ttaaggagcg catgatcggg 60
ggctctgctc tggcttcgct catctacgta tggctctttg gtgtcattgt tgtacccttg 120
gctacgtaca agatgtggc acagggcgac tatcgctcg cctcggcct cctcctttat 180
tacgcctacc gttgggtcta tccgaccaag gaatgggccc tegtgcgca catctaccga 240
gccggcaacc gatatttcta cccacaagag gtcctttttg atggcttcaa ggagatcaaa 300
cccgaatcga ggtcattgat ttgcatgca cgcgatgaa tottgactat tggttgggcg 360
ttgaccagca cgagtcacc catgacgca gccaatgtga agtggctggt gacggaggct 420
ttgttgctt tgccttttat cagcgacttc ctgtcctgga acggctgtgc acacgctagc 480
aagagctaca tgcaaaaccg tatgacgaag ggtgcgaatc ttgcctgct ccccgaggcg 540
tttgaagagg cttccctcta tcaacacagc tcttaccgtg tctacatccg aaagcgca 600

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ggctttgttg tgtatgccct cagatatggt tataagattt atccttcggt cgtctttggg 660
gaggagaagt gttattttctc tttgatgccc gactgggggt ggctaacggc ggcgaggcta 720
tggttgaatc agttccgggt cccggcagtt gcgtttgctg gaaagttggt tttggtgcct 780
gggtgggatt cgcatttgat cacggtgate ggcgcccccg tgggtgtgce gaggetagag 840
aagccaacgg aagaggaggt gaggaagtac cattcgttgt atgtgcgtgc attgatggaa 900
ttgtttgaga agcacaanaac ccaatattgt gagaaggggg cgaagttgga ggtgtggtag 960

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<210> SEQ ID NO 103

<211> LENGTH: 319

<212> TYPE: PRT

<213> ORGANISM: *Nannochloropsis oculata*

<400> SEQUENCE: 103

```

Met Ala Ala Ile Ser Pro Arg Lys His Pro Pro Pro Asp Leu Lys Glu
1           5           10           15
Arg Met Ile Gly Gly Leu Leu Leu Ala Ser Leu Ile Tyr Val Trp Leu
20          25          30
Phe Gly Val Ile Val Val Pro Leu Ala Thr Tyr Lys Met Leu Ala Gln
35          40          45
Gly Asp Tyr Arg Leu Ala Leu Gly Leu Leu Leu Tyr Tyr Ala Tyr Arg
50          55          60
Trp Val Tyr Pro Thr Lys Glu Trp Ala Leu Val Arg Asp Ile Tyr Arg
65          70          75          80
Ala Gly Asn Arg Tyr Phe Tyr Pro Gln Glu Val Leu Phe Asp Gly Phe
85          90          95
Lys Glu Ile Lys Pro Glu Ser Arg Ser Leu Ile Cys Met His Pro His
100         105         110
Gly Ile Leu Thr Ile Gly Trp Ala Leu Thr Ser Thr Ser Pro Thr Met
115        120        125
Thr His Ala Asn Val Lys Trp Leu Val Thr Glu Ala Leu Leu Arg Leu
130        135        140
Pro Phe Ile Ser Asp Phe Leu Ser Trp Asn Gly Cys Ala His Ala Ser
145        150        155        160
Lys Ser Tyr Met Gln Asn Arg Met Thr Lys Gly Ala Asn Leu Ala Leu
165        170        175
Leu Pro Gly Gly Phe Glu Glu Ala Ser Leu Tyr Gln His Ser Ser Tyr
180        185        190
Arg Val Tyr Ile Arg Lys Arg Thr Gly Phe Val Val Tyr Ala Leu Arg
195        200        205
Tyr Gly Tyr Lys Ile Tyr Pro Ser Phe Val Phe Gly Glu Glu Lys Cys
210        215        220
Tyr Phe Ser Leu Met Pro Asp Trp Gly Trp Leu Thr Ala Ala Arg Leu
225        230        235        240
Trp Leu Asn Gln Phe Arg Phe Pro Ala Val Ala Phe Val Gly Lys Leu
245        250        255
Phe Leu Val Pro Gly Trp Asp Ser His Leu Ile Thr Val Ile Gly Ala
260        265        270
Pro Val Val Leu Pro Arg Leu Glu Lys Pro Thr Glu Glu Glu Val Arg
275        280        285
Lys Tyr His Ser Leu Tyr Val Arg Ala Leu Met Glu Leu Phe Glu Lys
290        295        300

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His Lys Thr Gln Tyr Cys Glu Lys Gly Ala Lys Leu Glu Val Trp
305 310 315

<210> SEQ ID NO 104

<211> LENGTH: 1265

<212> TYPE: DNA

<213> ORGANISM: Nannochloropsis oculata

<400> SEQUENCE: 104

```

atcttcagca aagtaatcaa gataataaac aaaaacaatc ctataaagga aaaacaacag    60
gcgcatcatc tacgaagata gccatggcgc ccatctcacc gcgcaaacat cctccgcctg    120
atcttaagga gcgcatgatc gggggctctgc tcttggtctc gctcatctac gtagggctct    180
ttgggtgcat tgttgtaccc ttggctacgt acaagatgct ggcacagggc gactatcgcc    240
tcgccctcgg cctcctcctt tattacgect accggtgggt ctatccgacc aaggaatggg    300
ccctcgtgcg cgacatctac cgagccggca accgatattt ctaccacaaa gaggtccttt    360
ttgatggcct caaggagatc aaacccgaat cgaggtcatt gatttgcatt caccgcgatg    420
gaatcttgac tattggttgg cggttgacca gcacgagtc caccatgacg cagccaatg    480
tgaagtggct ggtgacggag gctttgttgc gcttgccttt tatcagcgac ttctgtcct    540
ggaacggctg tgcacacgct agcaagagct acatgcaaaa ccgtatgacg aagggtgcca    600
atcttgccct gctccccgga gggtttgaag aggcctccct ctatcaacac agctcttacc    660
gtgtctacat ccgaaagcgc acaggctttg ttgtgtatgc cctcagatat ggttataaga    720
tttatccttc gttcgtcttt ggggaggaga agtggtattt ctctttgatg cccgactggg    780
ggtaggtaac gggggcgagg ctatggttga atcagttccg gttcccgcca gttgcgttg    840
tcggaaagt gtttttggg cctgggtggg attcgcattt gatcacggtg atcgcgccc    900
ccgtgggtgt gccgaggcta gagaagccaa cggaagagga ggtgaggaag taccattcgt    960
tgtatgtgcg tgcattgatg gaattgtttg agaagcacia aaccaatat tgtgagaagg    1020
ggcgcaagt ggaggtgtgg taggataggg agagagggaa gggaaagtaa cacacatgta    1080
cagagctatg accaaagtaa tcgactgatg ggaggaggga gagggaaagt gaaagggaga    1140
aagaaagaga gagggggagg ctgccacacc gcgacgctgc gtgagtgctg ggtgtgtgtg    1200
tgtggagccc ttgatatttg aaataaaaat taaaaataaa aaaaaaaaaa aaaaaaaaaa    1260
aaaaa                                           1265

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<210> SEQ ID NO 105

<211> LENGTH: 1563

<212> TYPE: DNA

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 105

```

atggcgattt tggattctgc tggcggtact acggtgacgg agaacggtgg cggagagttc    60
gtcgatcttg ataggcttcg tcgacggaaa tcgagatcgg attcttctaa cggacttctt    120
ctctctgggt ccgataataa ttctccttcg gatgatgttg gagctcccgc cgacgttagg    180
gatcggattg attcctgttg taacgatgac gctcagggaa cagccaattt ggccggagat    240
aataacggtg gtggcgataa taacggtggt ggaagaggcg gcggagaagg aagaggaaac    300
gccgatgcta cgtttacgta tcgaccgtcg gttccagctc atcggagggc gagagagagt    360

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ccacttagct ccgacgcaat cttcaaacag agccatgccg gattattcaa cctctgtgta 420
gtagtcttta ttgctgtaaa cagtagactc atcatcgaaa atcttatgaa gtatggttgg 480
ttgatcagaa cggatttctg gtttagttca agatcgctgc gagattggcc gcttttcatg 540
tgttgatat ccttttcgat ctttctttg getgccttta cggttgagaa attggtactt 600
cagaaatata taccagaacc tggttgcatc tttcttcata ttattatcac catgacagag 660
gttttgtatc cagtttacgt caccctaagg tgtgattctg cttttttatc aggtgtcact 720
ttgatgctcc tcacttgcac tgtgtggcta aagttgggtt cttatgctca tactagctat 780
gacataagat ccttagccaa tgcagctgat aaggccaatc ctgaagtctc ctactacgtt 840
agcttgaaga gcttggcata tttcatggtc gctcccacat tgtgttatca gccaaagtat 900
ccacgttctg catgtatacg gaagggttgg gtggctcgto aatttgcaaa actggtcata 960
ttcaccggat tcattggatt tataatagaa caatatataa atcctattgt caggaactca 1020
aagcatcctt tgaaggcga tcttctatat gctattgaaa gagtgttgaa gctttcagtt 1080
ccaaatttat atgtgtggct ctgcatgttc tactgcttct tccacctttg gttaaacata 1140
ttggcagagc ttctctgctt cggggatcgt gaattctaca aagattggtg gaatgcaaaa 1200
agtgtgggag attactggag aatgtggaat atgcctgttc ataaatggat ggttcgacat 1260
atatacttcc cgtgcttgcg cagcaagata ccaaagacac tcgccattat cattgctttc 1320
ctagtctctg cagtctttca tgagcatatc atcgcagtto cttgtcgtct cttcaagcta 1380
tgggcttttc ttgggattat gtttcagggt cctttgggtc tcatacaaaa ctatctacag 1440
gaaaggtttg gctcaacggg ggggaacatg atcttctggt tcattctctg cattttcgga 1500
caaccgatgt gtgtgcttct ttattaccac gacctgatga accgaaaagg atcgatgtca 1560
tga 1563

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<210> SEQ ID NO 106

<211> LENGTH: 520

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 106

```

Met Ala Ile Leu Asp Ser Ala Gly Val Thr Thr Val Thr Glu Asn Gly
1          5          10          15
Gly Gly Glu Phe Val Asp Leu Asp Arg Leu Arg Arg Arg Lys Ser Arg
20          25          30
Ser Asp Ser Ser Asn Gly Leu Leu Leu Ser Gly Ser Asp Asn Asn Ser
35          40          45
Pro Ser Asp Asp Val Gly Ala Pro Ala Asp Val Arg Asp Arg Ile Asp
50          55          60
Ser Val Val Asn Asp Asp Ala Gln Gly Thr Ala Asn Leu Ala Gly Asp
65          70          75          80
Asn Asn Gly Gly Gly Asp Asn Asn Gly Gly Gly Arg Gly Gly Gly Glu
85          90          95
Gly Arg Gly Asn Ala Asp Ala Thr Phe Thr Tyr Arg Pro Ser Val Pro
100         105         110
Ala His Arg Arg Ala Arg Glu Ser Pro Leu Ser Ser Asp Ala Ile Phe
115         120         125
Lys Gln Ser His Ala Gly Leu Phe Asn Leu Cys Val Val Val Leu Ile
130         135         140

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Ala Val Asn Ser Arg Leu Ile Ile Glu Asn Leu Met Lys Tyr Gly Trp
145 150 155 160

Leu Ile Arg Thr Asp Phe Trp Phe Ser Ser Arg Ser Leu Arg Asp Trp
165 170 175

Pro Leu Phe Met Cys Cys Ile Ser Leu Ser Ile Phe Pro Leu Ala Ala
180 185 190

Phe Thr Val Glu Lys Leu Val Leu Gln Lys Tyr Ile Ser Glu Pro Val
195 200 205

Val Ile Phe Leu His Ile Ile Ile Thr Met Thr Glu Val Leu Tyr Pro
210 215 220

Val Tyr Val Thr Leu Arg Cys Asp Ser Ala Phe Leu Ser Gly Val Thr
225 230 235 240

Leu Met Leu Leu Thr Cys Ile Val Trp Leu Lys Leu Val Ser Tyr Ala
245 250 255

His Thr Ser Tyr Asp Ile Arg Ser Leu Ala Asn Ala Ala Asp Lys Ala
260 265 270

Asn Pro Glu Val Ser Tyr Tyr Val Ser Leu Lys Ser Leu Ala Tyr Phe
275 280 285

Met Val Ala Pro Thr Leu Cys Tyr Gln Pro Ser Tyr Pro Arg Ser Ala
290 295 300

Cys Ile Arg Lys Gly Trp Val Ala Arg Gln Phe Ala Lys Leu Val Ile
305 310 315 320

Phe Thr Gly Phe Met Gly Phe Ile Ile Glu Gln Tyr Ile Asn Pro Ile
325 330 335

Val Arg Asn Ser Lys His Pro Leu Lys Gly Asp Leu Leu Tyr Ala Ile
340 345 350

Glu Arg Val Leu Lys Leu Ser Val Pro Asn Leu Tyr Val Trp Leu Cys
355 360 365

Met Phe Tyr Cys Phe Phe His Leu Trp Leu Asn Ile Leu Ala Glu Leu
370 375 380

Leu Cys Phe Gly Asp Arg Glu Phe Tyr Lys Asp Trp Trp Asn Ala Lys
385 390 395 400

Ser Val Gly Asp Tyr Trp Arg Met Trp Asn Met Pro Val His Lys Trp
405 410 415

Met Val Arg His Ile Tyr Phe Pro Cys Leu Arg Ser Lys Ile Pro Lys
420 425 430

Thr Leu Ala Ile Ile Ile Ala Phe Leu Val Ser Ala Val Phe His Glu
435 440 445

Leu Cys Ile Ala Val Pro Cys Arg Leu Phe Lys Leu Trp Ala Phe Leu
450 455 460

Gly Ile Met Phe Gln Val Pro Leu Val Phe Ile Thr Asn Tyr Leu Gln
465 470 475 480

Glu Arg Phe Gly Ser Thr Val Gly Asn Met Ile Phe Trp Phe Ile Phe
485 490 495

Cys Ile Phe Gly Gln Pro Met Cys Val Leu Leu Tyr Tyr His Asp Leu
500 505 510

Met Asn Arg Lys Gly Ser Met Ser
515 520

<210> SEQ ID NO 107

<211> LENGTH: 1506

-continued

<212> TYPE: DNA

<213> ORGANISM: Brassica napus

<400> SEQUENCE: 107

```

atggagattt tggattctgg aggcgtcact atgccgacgg agaacgggtg tgccgatctc   60
gatacgcctc gtcaccggaa accgagatcg gattcttcca atggacttct tcttgattcc   120
gtaactgttt cccgatgctga cgtgagggat cggggtgatt cagctgttga ggatactcaa   180
ggaaaagcca atttggccgg agaaaacgaa attaggaat cccgtggaga agcggggggga   240
aacgtggatg taaggtacac gtatcggccg tcggttccag ctcatcggag ggtgcggggag   300
agtccactca gctctgacgc catcttcaaa cagagccatg ctggactatt caacctgtgt   360
gtagtgttc ttgttgctgt aaacagtaga ctcatcatcg aaaatctcat gaagtacggt   420
tggttgatca gaactgattt ctggtttagt tcaacgtctc tgcgagattg gcccttttc   480
atgtgttgtc tctcccttcc aatcttctct ttggctgcct ttaccgtcga gaaattagta   540
cttcagaaat gcatatctga acctgttgtc atcattcttc atattattat caccatgacc   600
gaggctctgt atccagtcta tgtcacteta aggtgtgatt ccgccttctt atcaggtgtc   660
acgttgatgc tctcacttgc cattgtgtgg ctgaagttgg tttcttacgc tcactaac   720
tatgacataa gaaccctaga taattcatct gataaggcca atcctgaagt ctctactat   780
gttagcttga agagcttggc gtatttcatg cttgctccca cattgtgtta tcagccgagc   840
tatccacgtt ctccatgat cccgaagggt tgggtggctc gtcaatttgc aaagctgatc   900
atattcactg gattcatggg atttataata gagcaatata taaatcctat tgttaggaac   960
tcaaaacatc ctttgaagg ggatctctta tacgggtgtg aaagagtgtt gaagctttca  1020
gttccaaatt tatacgtgtg gctctgcatg ttctactgct tcttccacct ttggttaaac  1080
atattggcag agctcctctg cttcggggat cgtgaattct acaagattg gtggaatgca  1140
aaaagcgtgg gagattattg gagaatgtgg aatatgcctg ttcataaatg gatggttcga  1200
catgtatact ttccgtgcct tcgcagaaat ataccgaaag taccgctat tacccttgcct  1260
ttcttagtct ctgcagtctt tcatgagtta tgcacgcag ttccttgtcg tctcttcaaa  1320
ctatgggctt tcttggggat tatgtttcag gtgcctttgg tatttatcac aaactaccta  1380
caagaaagggt ttggctccat ggtgggaaac atgatattct ggtttacctt ctgcattttc  1440
ggacaaccga tgtgtgtgct tctttattat caccacttga tgaaccgcaa aggaaagatg  1500
tcatag                                     1506

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<210> SEQ ID NO 108

<211> LENGTH: 501

<212> TYPE: PRT

<213> ORGANISM: Brassica napus

<400> SEQUENCE: 108

```

Met Glu Ile Leu Asp Ser Gly Gly Val Thr Met Pro Thr Glu Asn Gly
 1             5             10             15

Gly Ala Asp Leu Asp Thr Leu Arg His Arg Lys Pro Arg Ser Asp Ser
 20             25             30

Ser Asn Gly Leu Leu Pro Asp Ser Val Thr Val Ser Asp Ala Asp Val
 35             40             45

Arg Asp Arg Val Asp Ser Ala Val Glu Asp Thr Gln Gly Lys Ala Asn
 50             55             60

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Leu Ala Gly Glu Asn Glu Ile Arg Glu Ser Gly Gly Glu Ala Gly Gly
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 Arg Val Arg Glu Ser Pro Leu Ser Ser Asp Ala Ile Phe Lys Gln Ser
 100 105 110
 His Ala Gly Leu Phe Asn Leu Cys Val Val Val Leu Val Ala Val Asn
 115 120 125
 Ser Arg Leu Ile Ile Glu Asn Leu Met Lys Tyr Gly Trp Leu Ile Arg
 130 135 140
 Thr Asp Phe Trp Phe Ser Ser Thr Ser Leu Arg Asp Trp Pro Leu Phe
 145 150 155 160
 Met Cys Cys Leu Ser Leu Ser Ile Phe Pro Leu Ala Ala Phe Thr Val
 165 170 175
 Glu Lys Leu Val Leu Gln Lys Cys Ile Ser Glu Pro Val Val Ile Ile
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 Leu His Ile Ile Ile Thr Met Thr Glu Val Leu Tyr Pro Val Tyr Val
 195 200 205
 Thr Leu Arg Cys Asp Ser Ala Phe Leu Ser Gly Val Thr Leu Met Leu
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 Leu Thr Cys Ile Val Trp Leu Lys Leu Val Ser Tyr Ala His Thr Asn
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 Tyr Asp Ile Arg Thr Leu Ala Asn Ser Ser Asp Lys Ala Asn Pro Glu
 245 250 255
 Val Ser Tyr Tyr Val Ser Leu Lys Ser Leu Ala Tyr Phe Met Leu Ala
 260 265 270
 Pro Thr Leu Cys Tyr Gln Pro Ser Tyr Pro Arg Ser Pro Cys Ile Arg
 275 280 285
 Lys Gly Trp Val Ala Arg Gln Phe Ala Lys Leu Ile Ile Phe Thr Gly
 290 295 300
 Phe Met Gly Phe Ile Ile Glu Gln Tyr Ile Asn Pro Ile Val Arg Asn
 305 310 315 320
 Ser Lys His Pro Leu Lys Gly Asp Leu Leu Tyr Gly Val Glu Arg Val
 325 330 335
 Leu Lys Leu Ser Val Pro Asn Leu Tyr Val Trp Leu Cys Met Phe Tyr
 340 345 350
 Cys Phe Phe His Leu Trp Leu Asn Ile Leu Ala Glu Leu Leu Cys Phe
 355 360 365
 Gly Asp Arg Glu Phe Tyr Lys Asp Trp Trp Asn Ala Lys Ser Val Gly
 370 375 380
 Asp Tyr Trp Arg Met Trp Asn Met Pro Val His Lys Trp Met Val Arg
 385 390 395 400
 His Val Tyr Phe Pro Cys Leu Arg Arg Asn Ile Pro Lys Val Pro Ala
 405 410 415
 Ile Ile Leu Ala Phe Leu Val Ser Ala Val Phe His Glu Leu Cys Ile
 420 425 430
 Ala Val Pro Cys Arg Leu Phe Lys Leu Trp Ala Phe Leu Gly Ile Met
 435 440 445
 Phe Gln Val Pro Leu Val Phe Ile Thr Asn Tyr Leu Gln Glu Arg Phe
 450 455 460

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Gly Ser Met Val Gly Asn Met Ile Phe Trp Phe Thr Phe Cys Ile Phe
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Gly Gln Pro Met Cys Val Leu Leu Tyr Tyr His Asp Leu Met Asn Arg
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Lys Gly Lys Met Ser
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tagagcggat ttaattaact atgacatcctt tcctttgcgg t          101

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We claim:

1. A polynucleotide comprising an expression control sequence operatively linked to a heterologous nucleic acid sequence selected from the group consisting of:

- a) the nucleic acid sequence of SEQ ID NO: 52 or 54;
- b) a nucleic acid sequence encoding a polypeptide comprising the amino acid sequence of SEQ ID NO: 53;
- c) a nucleic acid sequence having at least 60% sequence identity to the nucleic acid sequence of a) or b), wherein said nucleic acid sequence encodes a polypeptide having diacylglycerol acyltransferase activity; and
- d) a nucleic acid sequence encoding a polypeptide having at least 60% sequence identity to the amino acid sequence of SEQ ID NO: 53, wherein said polypeptide has diacylglycerol acyltransferase activity.

2. The polynucleotide of claim 1, wherein said polynucleotide further comprises a terminator sequence operatively linked to the nucleic acid sequence.

3. A vector comprising the polynucleotide of claim 1.

4. A host cell comprising:

- a) the polynucleotide of claim 1; or
- b) a vector comprising said polynucleotide.

5. The host cell of claim 4, wherein the host cell is a plant cell or a microorganism.

6. The host cell of claim 4, wherein the host cell is yeast, fungus, algae, moss, or an insect cell.

7. A method for the manufacture of a polypeptide, comprising:

- a) cultivating the host cell of claim 4 under conditions which allow for the production of said polypeptide; and
- b) obtaining the polypeptide from said host cell.

8. A non-human transgenic organism comprising:

- a) the polynucleotide of claim 1; or
- b) a vector comprising said polynucleotide, wherein the non-human transgenic organism is a plant or a microorganism.

9. The non-human transgenic organism of claim 8, wherein the microorganism is a fungus, algae, moss, or yeast.

10. A method for the manufacture of polyunsaturated fatty acids, comprising:

- a) cultivating the host cell of claim 4 under conditions which allow for the production of polyunsaturated fatty acids in said host cell; and
- b) obtaining said polyunsaturated fatty acids from said host cell.

11. A method for the manufacture of polyunsaturated fatty acids, comprising:

- a) cultivating the non-human transgenic organism of claim 8 under conditions which allow for the production of polyunsaturated fatty acids in said host cell; and
- b) obtaining said polyunsaturated fatty acids from said non-human transgenic organism.

12. The method of claim 11, wherein said polyunsaturated fatty acid is arachidonic acid (ARA), eicosapentaenoic acid (EPA), and/or docosahexaenoic acid (DHA).

13. A method for the manufacture of an oil, lipid, or fatty acid composition, comprising:

- a) cultivating the host cell of claim 4 under conditions which allow for the production of polyunsaturated fatty acids in said host cell;
- b) obtaining said polyunsaturated fatty acids from said host cell; and
- c) formulating the polyunsaturated fatty acid as an oil, lipid, or fatty acid composition.

14. The method of claim 13, wherein said oil, lipid, or fatty acid composition is to be used for feed, foodstuffs, cosmetics, or pharmaceuticals.

15. A method for the manufacture of polyunsaturated fatty acids, comprising:

- a) cultivating a plant comprising the polynucleotide of claim 1 or a vector comprising said polynucleotide under conditions which allow for the production of polyunsaturated fatty acids in said plant or seeds thereof; and
- b) obtaining said polyunsaturated fatty acids from said plant or seeds thereof.

16. The method of claim 15, wherein the polyunsaturated fatty acids are obtained from the seeds of said plant.

17. A method for the manufacture of an oil, lipid or fatty acid composition, comprising:

- a) providing a polyunsaturated fatty acid produced by the method of claim 15; and
- b) formulating said polyunsaturated fatty acid as an oil, lipid or fatty acid composition.

18. A method for the manufacture of an oil, lipid or fatty acid composition, comprising:

- a) cultivating a plant comprising the polynucleotide of claim 1 or a vector comprising said polynucleotide under conditions which allow for the production of polyunsaturated fatty acids in said plant or seeds thereof; and

b) obtaining an oil, lipid or fatty acid composition from said plant or seeds thereof.

19. The method of claim **18**, wherein the oil, lipid or fatty acid composition is obtained from the seeds of said plant.

20. A method for the production of feed, foodstuffs, cosmetics or pharmaceuticals, comprising:

a) obtaining an oil, lipid or fatty acid composition produced by the method of claim **18**; and

b) processing said oil, lipid or fatty acid composition to produce feed, foodstuffs, cosmetics or pharmaceuticals.

21. A method for the manufacture of polyunsaturated fatty acids, comprising:

a) obtaining an oil, lipid or fatty acid composition produced by the method of claim **18**; and

b) obtaining polyunsaturated fatty acids from said oil, lipid or fatty acid composition.

22. A plant, or a plant part, plant cell, or seed thereof, wherein said plant, or said plant part, plant cell, or seed thereof, comprises:

a) the polynucleotide of claim **1**; or

b) a vector comprising said polynucleotide.

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