www.sospublication.co.in

Journal of Advanced Laboratory Research in Biology



We- together to save yourself society

Volume 2, Issue 4, October 2011



e-ISSN 0976-7614

Research Article

Antagonistic Bioactivity of Endophytic Actinomycetes Isolated from Medicinal Plants

M. Gangwar^{1*}, S. Dogra² and N. Sharma³

^{1,2*}Department of Microbiology, Punjab Agricultural University, Ludhiana-141004, India. ³Department of Plant Breeding and Genetics, Punjab Agricultural University, Ludhiana-141004, India.

Abstract: Endophytic actinomycetes are promising biocontrol agents for use in agriculture and have been isolated from various plant species. In the present study, 40 endophytic actinomycetes were isolated from roots, stems and leaves of three medicinal plants viz. *Aloe vera, Mentha arvensis* and *Ocimum sanctum*. The identification revealed that the majority of the isolates were *Streptomyces* spp. and the rest were identified as *Saccharopolyspora* spp., *Micromonospora* spp. and *Actinopolyspora* spp. The dual tests revealed that nine endophytic actinomycete isolates displayed a wide spectrum activity against nine fungal phytopathogens. Out of 8 isolates, 90% inhibited the growth of at least one or more phytopathogenic fungi and *Saccharopolyspora* 0-9 (Out of 8 isolates, 90% inhibited the growth of at least one or more phytopathogenic fungi and *Saccharopolyspora* 0-9 exhibited antagonistic activity against *Aspergillus niger, Aspergillus flavus, Alternaria brassicicola, Botrytis cinerea, Penicillium digitatum, Fusarium oxysporum, Penicillium pinophilum, Phytophthora dresclea and Colletotrichum falcatum.*

Keywords: Endophytic actinomycetes, Phytopathogenic fungi, Biocontrol agents, Antagonistic activity.

1. Introduction

Actinomycetes are known as producers of antibiotics and other biologically active substances with high commercial value such as vitamins, alkaloids, plant growth factors, enzymes and enzyme inhibitors (Tanaka and Omura, 1993). Approximately two-thirds of naturally occurring antibiotics, including some of agricultural importance, have also been isolated from these soil microorganisms. Evidence indicated that actinomycetes are important in the rhizosphere because they can influence plant growth and protect plant roots against invasion by root pathogenic fungi (Crawford et al., 1993; Tokala et al., 2002). Actinomycetes are found also as endophytes that colonize the plant tissues. Actinomycetes have been isolated from surface sterilized roots of Italian native plants (Sardi et al., 1992), from roots and leaves of maize (De-Araujo et

al., 2000), from roots and leaves of banana plants (Cao *et al.*, 2004) and from surface sterilized wheat roots (Coombs and Franco, 2003). *In vitro* and *in vivo* antagonistic activities of endophytic actinomycetes against plant pathogens have been reported (Cao *et al.*, 2005; Taechowisan *et al.*, 2003; Tian *et al.*, 2004). The introduction of endophytic actinomycetes into plants with the ability to colonize the internal tissue would further enhance the stability and increase their potential effectiveness as biocontrol agents (Coombs *et al.*, 2004).

In order to find effective biocontrol agents for fungal plant pathogens, endophytic actinomycetes were isolated from surface-sterilized roots, stems and leaves of medicinal plants. The present study was undertaken with a view to testing the potential of endophytic actinomycetes as biocontrol agents against various phytopathogenic fungi.

2. Materials and Methods

2.1 In vitro antagonistic bioassay

The actinomycete isolates were evaluated for their activity towards nine pathogenic fungi: Aspergillus niger, Aspergillus flavus, Alternaria brassicicola, **Botrytis** cinerea, Penicillium digitatum, Penicillium Fusarium oxysporum, pinophilum, *Phytophthora dresclea* and *Colletotrichum falcatum* by dual-culture in vitro assay. Fungal discs (8mm in diameter), 5 days old on PDA at 28°C were placed at the center of PDA plates. Two actinomycete discs (8mm) 5 days old, grown on yeast malt extract (YM) incubated at 28°C were placed on opposite sides of the plates, 3cm away from fungal disc. Plates without the actinomycete disc serve as controls. All the plates were incubated at 28°C for 14 days and colony growth inhibition (%) was calculated by using the formula: C – $T/C \ge 100$, where C is the colony growth of pathogen in control and T is the colony growth of pathogen in dual culture. The zone of inhibition was measured between the pathogen and actinomycete isolates (Khamna et al., 2009).

3. Results and Discussion

3.1 Antifungal activities

Eight (20%) of actinomycete isolates were active against at least one of the nine phytopathogenic fungi (Table 1).

Different isolates of *Streptomyces* spp. displayed an array of activity against pathogenic fungi, particularly *S. albosporus* A4, an *Aloe vera* isolate and O-11, an *Ocimum sanctum* isolate. This is in conformity with the results of several studies carried out by other investigators (Crawford *et al.*, 1993; Taechowisan and Lumyong, 2003; Tian *et al.*, 2004; Verma *et al.*, 2009). *Saccharopolyspora* O-9 from *Ocimum sanctum* strongly inhibited all of the pathogenic fungi (Fig. 1) with maximum percent inhibition was observed against the fungus *Penicillium digitatum* (71.4%).

Actinomycetes-fungus antagonism has been demonstrated for a variety of plant pathogens such as Alternaria, Rhizoctonia, Verticillium, Fusarium, Phytophthora and Pythium spp. (Yuan and Crawford, 1995; Aghighi et al., 2004). The ability of isolates to inhibit the growth of fungal pathogens is implication of the volatile secondary metabolites secreted by actinomycetes. So, in the present study, the potential of endophytic actinomycetes to inhibit the growth of pathogens has been studied. Dual-culture assays showed that some actinomycetes isolated can be developed as potential biocontrol agents. Therefore, further studies are necessary to assess the ability of the isolates to confer protection against pathogens and their role in enhancing growth and yield of plants under field conditions.

In conclusion, all but one of our Streptomyces isolates tested displayed antifungal activity. In addition, to Streptomyces, the medicinal plants carried rare actinomycetes all of which displayed antifungal activity with *Saccharopolyspora* being the potent antagonist. Our survey suggested that medicinal plants are a potent source of endophytic actinomycetes with wide biological activity against pathogenic fungi.

Table 1. Antifunga	activity of actinomycete isolates.	
--------------------	------------------------------------	--

	Percentage (%) inhibition ^a									
Actinomycete isolates	Asper gillus niger	Colletotrichum falcatum	Asper gillus flavus	Alternaria brassicicola	Penicillium digitatum	Fusarium oxysporum	Penicillium pinophilum	Phytophthora dresclea	Botrytis cinerea	
Micromonospora 0-14	0	0	0	62.2 ± 0.1	19.5 ± 0.1	12.6 ± 0.1	0	16 ± 0.1	16.1 ± 0.1	
S. viridis A3	0	0	0	39.6 ± 0.2	22.8 ± 0.1	45.8 ± 0.1	0	0	17.4 ± 0.1	
S. albosporus A4	11.4 ± 0.2	14.5 ± 0.1	0	63.5 ± 0.1	60.6 ± 0.1	19.8 ± 0.2	0	69.3 ± 0.3	57.9 ± 0.1	
S. cinereus A6	0	0	0	42.4 ± 0.08	0	17.6 ± 0.1	34.5 ± 0.2	0	0	
Micromonospora A9	0	0	0	40.3 ± 0.2	0	48.5 ± 0.2	0	13.9 ± 0.1	0	
S. cinereus O-1	0	0	0	25.3 ± 0.1	15.2 ± 0.1	0	0	0	42.6 ± 0.1	
Saccharopolyspora O-9	13.2 ± 0.2	28.6 ± 0.1	26.5 ± 0.2	30.5 ± 0.1	71.4 ± 0.2	51.4 ± 0.1	17.8 ± 0.1	56.4 ± 0.1	49.9 ± 0.1	
S. albosporus 0-11	16.7 ± 0.1	17.5 ± 0.2	0	44.6 ± 0.2	18.6 ± 0.1	59.5 ± 0.2	0	53.5 ± 0.2	53.4 ± 0.2	

^aAverage ± standard error from triplicate samples

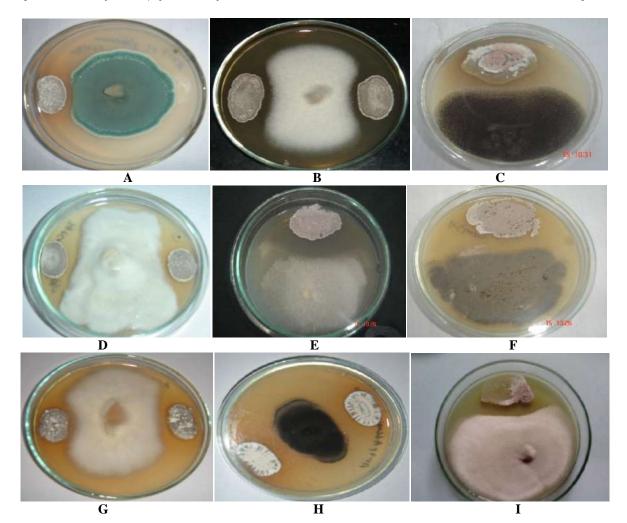


Fig. 1. Inhibition effect of Saccharopolyspora O-9 against Penicillium digitatum (A), Fusarium oxysporum (B), Aspergillus niger (C), Colletotrichum falcatum (D), Botrytis cinerea (E), Aspergillus flavus (F), Phytophthora dresclea (G), Alternaria brassicicola (H) and Penicillium pinophilum (I), Penicillium digitatum

References

- [1]. Aghighi, S., Bonjar, G.H.S., Rawashdeh, R., Batayneh, S. and Saadoun, I. (2004). First Report of Antifungal Spectra of Activity of Iranian Actinomycetes Strains Against Alternaria solani, Alternaria alternate, Fusarium solani, Phytophthora megasperma, Verticillium dahliae and Saccharomyces cerevisiae. Asian J. Plant Sci., 3: 463-71.
- [2]. Cao, L., Qiu, Z., You, J., Tan, H. and Zhou, S. (2004). Isolation and characterization of endophytic *Streptomyces* strain from surface sterilized tomato (*Lycopersicon esculentum*) roots. *Lett. Appl. Microbiol.*, **39:** 425-30.
- [3]. Cao, L., Qiu, Z., You, J., Tan, H. and Zhou, S. (2005). Isolation and Characterization of endophytic streptomycete antagonists of Fusarium wilt pathogen from surface-sterilized banana roots. *FEMS Microbiol. Lett.*, **247**: 147-52.

- [4]. Coombs, J.T. and Franco, C.M.M. (2003). Isolation and identification of actinobacteria from surface-sterilized wheat roots. *Appl. Environ. Microbiol.*, 69: 5603-08.
- [5]. Coombs, J.T., Michelsen, P.P. and Franco, C.M.M. (2004). Evaluation of endophytic bacteria as antagonists of *Gaeumannomyces graminis* var. *tritici* in wheat. *Biol. Control*, **29**: 359-66.
- [6]. Crawford, D.L., Lynch, J.M., Whipps, J.M. and Ousley, M.A. (1993). Isolation and characterization of actinomycete antagonistic of a fungal root pathogen. *Appl. Environ. Microbiol.*, 59: 3899-05.
- [7]. Araujo, J.M., Silva, A.C. & Azevedo, J.L. (2000). Isolation of endophytic actinomycetes from roots and leaves of maize (*Zea mays L.*). *Brazilian Archives of Biology and Technology*, 43: 447– 451.
- [8]. Khamna, S., Yokota, A. and Lumyong, S. (2009). Actinomycetes isolated from medicinal plant

rhizosphere soil: diversity and screening of antifungal compound, indole-3-acetic acid and siderophore production. *World J. Microbiol. Biotechnol.*, **25**: 649-55.

- [9]. Sardi, P., Saracchi, M., Quaroni, S., Petrolini, B., Borgonovi, G.E. and Merli, S. (1992). Isolation of endophytic *Streptomyces* strains from surfacesterilized roots. *Appl. Environ. Microbiol.*, 58: 2691-93.
- [10]. Taechowisan, T. and Lumyong, S. (2003). Activity of endophytic actinomycetes from roots of *Zingiber officinale* and *Alpinia galanga* against phytopathogenic fungi. *Ann. Microbiol.*, **53**: 291-98.
- [11]. Taechowisan, T., Peberdy, J.F. and Lumyong, S. (2003). Isolation of endophytic actinomycetes from selected plants and their antifungal activity. *World J. Microbiol. Biotechnol.*, **19**: 381-85.
- [12]. Tanaka, Y. and Omura, S. (1993). Agroactive compounds of microbial origin. Ann. Rev. Microbiol., 47: 57-87.

- [13]. Tian, X.L., Cao, L.X., Tan, H.M., Zeng, Q.G., Jia, Y.Y., Han, W.Q. & Zhou, S.N. (2004). Study on the communities of endophytic fungi and endophytic actinomycetes from rice and their antipathogenic activities *in vitro*. World J. Microbiol. Biotechnol., **20**: 303-09.
- [14]. Tokala, R.K., Strap, J.L., Jung, C.M., Crawford, D.L., Salove, M.H., Deobald, L.A., Bailey, J.F. and Morra, M.J. (2002). Novel Plant microbe rhizosphere interaction involving *Streptomyces lydicus* WYEC108 and the Pea plant (*Pisum sativum*). Appl. Environ. Microbiol., 68: 2161-71.
- [15]. Verma, V.C., Gond, S.K., Kumar, A., Mishra, A., Kharwar, R.N. and Gange, A.C. (2009). Endophytic actinomycetes from *Azadirachta indica* A. Juss.: Isolation, diversity and antimicrobial activity. *Microb. Ecol.*, **57**: 749-56.
- [16]. Yuan, W.M. and Crawford, D.L. (1995). Characterization of *Streptomyces lydicus* WYEC108 as a potential biocontrol agent against fungal root and seed rots. *Appl. Environ. Microbiol.*, 61: 3119-28.