

# ***Escherichia coli* with virulence factors and multidrug resistance in the Plankenburg River**

## **AUTHORS:**

Corne Lamprecht<sup>1</sup>  
Marco Romanis<sup>1</sup>  
Nicola Huisamen<sup>1</sup>  
Anneri Carinus<sup>1</sup>  
Nika Schoeman<sup>1</sup>  
Gunnar O. Sigge<sup>1</sup>  
Trevor J. Britz<sup>1</sup>

## **AFFILIATION:**

<sup>1</sup>Department of Food Science,  
Stellenbosch University,  
Stellenbosch, South Africa

## **CORRESPONDENCE TO:**

Trevor Britz

## **EMAIL:**

tjbritz@sun.ac.za

## **POSTAL ADDRESS:**

Department of Food Science,  
Stellenbosch University, Private  
Bag X1, Matieland 7600,  
South Africa

## **DATES:**

**Received:** 06 Nov. 2013

**Revised:** 06 Feb. 2014

**Accepted:** 18 Feb. 2014

## **KEYWORDS:**

enteropathogenic;  
enteroaggregative;  
virulence genes; antibiotic  
resistance; pollution

## **HOW TO CITE:**

Lamprecht C, Romanis M,  
Huisamen N, Carinus A,  
Schoeman N, Sigge GO, et al.  
*Escherichia coli* with virulence  
factors and multidrug resistance  
in the Plankenburg River. S  
Afr J Sci. 2014;110(9/10),  
Art. #2013-0347, 6 pages.  
[http://dx.doi.org/10.1590/  
sajs.2014/20130347](http://dx.doi.org/10.1590/sajs.2014/20130347)

*Escherichia coli* is a natural inhabitant of the gut and *E. coli* levels in water are considered internationally to be an indication of faecal contamination. Although not usually pathogenic, *E. coli* has been linked to numerous foodborne disease outbreaks, especially those associated with fresh produce. One of the most common ways through which *E. coli* can be transferred onto fresh produce is if contaminated water is used for irrigation. In this study, a total of 81 confirmed *E. coli* strains were isolated from the Plankenburg River as part of three separate studies over 3 years. During sampling, *E. coli* levels in the river were above the accepted levels set by the World Health Organization and the South African Department of Water Affairs and Forestry for safe irrigation of fresh produce, which indicates that transfer of *E. coli* during irrigation is highly probable. Multiplex polymerase chain reaction screening for pathogenic gene sequences revealed one enteroaggregative positive strain and four enteropathogenic positive strains. The four enteropathogenic strains were also found to be resistant to three or more critically and highly important antibiotics and were therefore classified as multidrug resistant strains. These results show that *E. coli* with enteropathogenic potential and multiple antimicrobial resistance properties has persisted over time in the Plankenburg River.

## **Introduction**

*Escherichia coli* is a natural inhabitant of the gut of humans, birds and other warm-blooded animals and is widely accepted as an indicator of faecal contamination of water. It is a robust bacterium which is genetically highly adaptable to environmental stresses, and has been shown to survive and multiply in the environment.<sup>1,2</sup> Based on the aforementioned, concerns have been raised in recent years regarding the status of *E. coli* as just a faecal indicator organism.<sup>3</sup>

Although most strains are commensal, pathogenic *E. coli* strains can contain various virulence factors and can be responsible for a variety of infections.<sup>4</sup> Based on the specific virulence factors present, pathogenic *E. coli* can be classified as either extra-intestinal pathogenic *E. coli* (ExPEC) or intestinal pathogenic *E. coli* (InPEC). ExPEC strains are usually able to cause infections in anatomical sites outside of the intestinal tract and are associated with urinary tract infections, neonatal meningitis and septicaemia. ExPEC, like commensal *E. coli*, can colonise the intestinal tract without causing gastroenteritis. In contrast, InPEC strains can cause different types of gastroenteritis and can be divided into six pathogenic groups: enterohaemorrhagic (EHEC); enteropathogenic (EPEC); enteroaggregative (EAEC); enterotoxigenic (ETEC); enteroinvasive (EIEC) and diffusely adherent (DAEC) *E. coli*. Each of the InPEC types has different infection mechanisms and symptoms.<sup>4</sup>

Foodborne disease outbreaks linked to pathogenic *E. coli*, specifically those derived from fresh produce, are increasing both in number and intensity.<sup>5</sup> As a result, *E. coli* is considered to be an emerging pathogen. One of the most common means by which *E. coli* can be transferred to fresh produce is via contaminated irrigation water. Recognising this potential danger, the World Health Organization (WHO) and the South African Department of Water Affairs (DWA) have set a recommended limit for irrigation water used for fresh produce of 1000 faecal coliforms/100 mL.<sup>6,7</sup>

Long-term monitoring studies of the Plankenburg River have revealed that although faecal coliform loads vary depending on the season, the loads are above the recommended limits.<sup>8</sup> The Plankenburg River flows past an informal settlement as well as through an industrial area of the town of Stellenbosch before it converges with the Eerste River and flows through an agricultural region, where the water is withdrawn for irrigation. Constant high faecal coliform levels might therefore contribute to the possible transfer of *E. coli* during irrigation, if river water is not treated prior to irrigation.

Although the presence of several potential pathogens in the Plankenburg River has been reported,<sup>8</sup> the occurrence and types of pathogenic *E. coli* are not known. The aim of this study was therefore to determine the number, types and antibiotic susceptibility of potential pathogenic *E. coli* present in the Plankenburg River.

## **Materials and methods**

### *Sampling*

Water samples from which *E. coli* was isolated were collected from the Plankenburg River (Stellenbosch, Western Cape Province) at three sites (P-0, P-1 and P-2).<sup>8</sup> The P-0 sampling site was about 5 km upstream of the P-1 site and was selected specifically to assess the impact of an informal settlement and industrial area on the water quality of the Plankenburg River. However, for large parts of each year, especially during the dry summer months, this site had no flowing water and therefore it was not sampled at the same frequency as the other sites. Two of the sites – P-1 and P-2 – are situated downstream of an informal settlement and industrial area and have shown high levels of faecal contamination.<sup>8</sup> River water sampling was done in accordance with the SANS 5667-6 method<sup>9</sup>; all samples were 1 L in volume.

*E. coli* isolation and identification

Study 1 (year 1):

Daily, for 9 days, 1 L of water was collected aseptically at site P-1. The presence of coliforms, faecal coliforms and *E. coli* were determined using the multiple tube fermentation (MTF) technique.<sup>10</sup> The *E. coli* broth tubes that exhibited gas production as well as fluorescence after 24 h incubation at 44.5 °C were streaked onto Eosin Methylene-blue Lactose Sucrose (L-EMB) agar (Oxoid, Hampshire, UK). Five typical *E. coli* colonies (dark purple to black colonies with a distinct metallic sheen) from each of the water samples were selected using Harrison's disc method<sup>11</sup> and further purified using Brilliance™ *E. coli*/coliform selective agar (Oxoid).

Study 2 (year 2):

Daily, for 14 days, 1 L of water was collected aseptically at site P-2. *E. coli* enumeration was done according to the American Public Health Association Standard Method<sup>12</sup> and Violet Red Bile Agar (Merck, Johannesburg, South Africa) was used for the enumeration. For further analysis, representative *Enterobacteriaceae* colonies (red and pink) were selected using Harrison's disc method.<sup>11</sup> The selected colonies were further purified using Brilliance™ *E. coli*/coliform selective agar (Oxoid).

Study 3 (year 3):

As part of a larger environmental study, sites P-0, P-1 and P-2 were sampled twice. Enumeration of total coliforms and *E. coli* was done according to SANS 9308<sup>13</sup> using Colilert 18 (IDEXX, Cape Town, South Africa). The Colilert 18 method is considered more user-friendly than the traditional MTF method used in Study 1, and it has been reported that Colilert results compare well with MTF results.<sup>14</sup> After incubation,<sup>14</sup> Quantitrays were divided into quarters and two fluorescent wells per quarter were pooled (1 mL per well to yield a maximum of 8 mL). A dilution was made from the pooled extract and spread plates were prepared on L-EMB agar (Oxoid). Five typical *E. coli* colonies from each of the water samples were selected at the highest dilution using Harrison's disc method<sup>11</sup> and purified using Brilliance™ *E. coli*/coliform selective agar (Oxoid).

Characterisation and confirmation of *E. coli* identification

Each isolate was streaked out on nutrient agar (Biolab, Johannesburg, South Africa), and the analytical profile index (API) 20E system (BioMérieux, Johannesburg, South Africa) was used in conjunction with Gram staining, a catalase test and growth on MacConkey medium.<sup>15</sup> The profile was then entered into the API Web database (BioMérieux) for species identification. Isolates were stored in 40% (v/v) glycerol at -80 °C.

Polymerase chain reaction analyses

Cell extracts of all isolates were prepared prior to the polymerase chain reaction (PCR) using the method of Altalhi and Hassen<sup>16</sup>. Isolates identified as *E. coli* with API were tested for the presence of the *E. coli uidA* household gene using the primer sequences of Heijnen and Medema<sup>17</sup> and KAPATaq™ HotStart DNA polymerase (KAPABiosystems, Cape Town, South Africa). Strains that tested positive for *uidA* were subjected to a multiplex PCR method as modified from Omar and Barnard<sup>18</sup> to screen for the presence of InPEC genes (Table 1). Strains identified as InPECs were also screened for the presence of ExPEC genes<sup>19</sup> and classified phylogenetically using the triplex PCR method of Clermont et al.<sup>20</sup> for *E. coli* phylogenetic group (genogroup) identification. The Kapa2G™ Fast Multiplex PCR kit (KAPABiosystems) was used for the InPEC and ExPEC multiplex methods as well as for the triplex PCR method. Positive and negative controls were included in all PCRs, and all reactions were performed in a G-storm thermal cycler (Vacutec, Johannesburg, South Africa). Primer sequences and concentrations are presented in Table 1. Reaction conditions are presented in Table 2. PCR products were all visualised with UV illumination after gel electrophoresis in agarose gels containing 1 µg/mL ethidium bromide and 0.5 x TBE

buffer. Gel electrophoresis was performed at 210V/20 min for the *uidA* PCR products and the triplex PCR products, using 1.25% agarose gels and 2% agarose gels, respectively. Gel electrophoresis of the InPEC and ExPEC PCR products were conducted at 120V/90 min using 1.25% agarose gels. A 100-bp marker (Promega, Madison, WI, USA) was included for size estimation purposes.

**Table 1:** Primer sequences used for *uidA*,<sup>17</sup> triplex,<sup>20</sup> InPEC<sup>18</sup> and ExPEC<sup>19</sup> polymerase chain reactions (PCRs)

Primer <sup>†</sup>	Primer sequence (5' - 3')	Size (bp)	Primer concentration
<b><i>E. coli</i> confirmation PCR<sup>17</sup></b>			
UAL 1939b	ATGGAATTCGCCGATTTGTC	187	0.4 µM
UAL 2105b	ATTGTTTGCCTCCCTGCTGC		
<b>InPEC PCR<sup>18</sup></b>			
<b><i>E. coli</i> control</b>			
Mdh01 (F)	GGTATGGATCGTTCCGACCT <sup>21</sup>	300	0.2 µM
Mdh02 (R)	GGCAGAATGGTAACACCAGAGT <sup>21</sup>		
<b>EIEC</b>			
L-lial (F)	GGTATGATGATGATGAGTCCA <sup>22</sup>	650	0.2 µM
lal (R)	GGAGGCCAACAAATATTTC <sup>22</sup>		
<b>EPEC &amp; EHEC<sup>†</sup></b>			
L-eaeA (F)	GACCCGGCACAAGCATAAGC <sup>22</sup>	384	0.2 µM
L-eaeA (R)	CCACCTGCAGCAACAGAGG <sup>22</sup>		
<b>EHEC</b>			
Stx1 (F)	ACACTGGATGATCTCAGTGG <sup>23</sup>	614	0.2 µM
Stx1 (R)	CTGAATCCCCTCCATTATG <sup>23</sup>		
Stx2 (F)	CCATGACAACGGACAGCAGT <sup>23</sup>	779	0.2 µM
Stx2 (R)	CCTGTCAACTGAGCACTTTG <sup>23</sup>		
<b>ETEC</b>			
LT (F)	GGCGACAGATTATACCGTGC <sup>22</sup>	450	0.2 µM
LT (R)	CGGTCTCTATATCCCTGTT <sup>22</sup>		
ST (F)	TTTCCCCTCTTTAGTCAGTCAACT <sup>18</sup>	160	0.2 µM
ST (R)	GGCAGGATTACAACAAGTTCA <sup>18</sup>		
<b>EAEC</b>			
Eagg (F)	AGACTCTGGCGAAAGACTGTATC <sup>24</sup>	194	0.2 µM
Eagg (R)	ATGGCTGTCTGTAATAGATGAGAAC <sup>24</sup>		
<b>Phylogenetic group PCR<sup>20</sup></b>			
chuA.1 (F)	GACGAACCAACGGTCAGGAT	279	0.2 µM
chuA.2 (R)	TGCCGCCAGTACCAAGACA		
yjaA.1 (F)	TGAAGTGTGAGGAGACGCTG	211	0.2 µM
yjaA.2 (R)	ATGGAGAATGCGTCTCTCAAC		
TSPE4.C2.1 (F)	GAGTAATGTCGGGGCATTCA	152	0.2 µM
TSPE4.C2.2 (R)	CGCGCCAACAAGTATTACG		
<b>ExPEC PCR<sup>19</sup></b>			
papA (F)	ATGGCAGTGGTGTCTTTGGTG	717	0.2 µM
papA (R)	CGTCCCACCATACGTCTCTTC		
papC (F)	GTGGCAGTATGAGTAATGACCGTTA	205	0.2 µM
papC (R)	ATATCCTTTCTGCAGGGATGCAATA		
sfa (F)	CTCCGGAGAACTGGGTGCATCTTAC	410	0.2 µM
sfa (R)	CGGAGGAGTAATTACAACCTGGCA		
iutA (F)	GGCTGGACATCATGGGAAGTGG	302	0.2 µM
iutA (R)	CGTCGGGAACGGGTAGAATCG		
kpsMT (F)	GCGCATTGCTGATACTGTTG	272	0.2 µM
kpsMT (R)	CATCCAGACGATAAGCATGAGCA		

<sup>†</sup>F, forward primer; R, reverse primer

<sup>†</sup>EHEC strains should be eaeA positive as well as stx 1 and/or stx 2 positive

**Table 2:** Thermocycling conditions used for *uidA*, triplex, InPEC and ExPEC polymerase chain reactions (PCRs)

PCR method	Thermocycling conditions
<i>uidA</i>	95 °C for 3 min, 35 cycles of 95 °C for 30 s, 59.7 °C for 30 s, 72 °C for 30 s, final extension at 72 °C for 5 min
InPEC multiplex	95 °C for 3 min, 30 cycles of 95 °C for 15 s, 55 °C for 30 s, 68 °C for 30 s, final extension at 72 °C for 5 min
Genogroup triplex	95 °C for 3 min, 30 cycles of 95 °C for 30 s, 60 °C for 30 s, 72 °C for 30 s, final extension 72 °C for 5 min
ExPEC multiplex	95 °C for 3 min, 30 cycles of 95 °C for 15 s, 61 °C for 30 s, 68 °C for 30 s, final extension at 72 °C for 5 min

### Antimicrobial resistance testing

The *E. coli* pathogens identified with PCR analysis were also screened for antimicrobial susceptibility against Ampicillin (10 µg), Cephalothin (30 µg), Chloramphenicol (30 µg), Ciprofloxacin (5 µg), Tetracycline (30 µg), Trimethoprim (2.5 µg), and Streptomycin (S) (10 µg). This screening was done using the standard antimicrobial disc susceptibility test described by the US Clinical and Laboratory Standards Institute (CLSI).<sup>25</sup> Inhibition zones were interpreted using the performance standards of CLSI<sup>26</sup> (for Ampicillin, Cephalothin, Chloramphenicol, Ciprofloxacin, Tetracycline and Streptomycin) and Andrews<sup>27</sup> (for Trimethoprim) and strains were classified as susceptible, intermediate or resistant. The American Type Culture Collection (ATCC) strain 25922 was included as a susceptible control in all antimicrobial resistance screening tests.

### Standard cultures for PCR analysis

*E. coli* ATCC 25922 was used as a positive control in the *uidA* PCR and the triplex PCR. ATCC 25922 was also combined with ATCC 35218 and used as a positive control for the ExPEC multiplex PCR after the identity of bands amplified in the expected regions for *papA*, *sfa/foc*, *iutA*, *kpsMT* II and *papC* were confirmed with sequencing and BLAST identification. A combination of EPEC, EHEC, ETEC, EIEC and EAEC positive strains was used as a positive control for the InPEC multiplex PCR.

## Results and discussion

### *E. coli* loads

The *E. coli* loads in the Plankenburg River samples and the number of confirmed *E. coli* strains are presented in Table 3. In all instances, the *E. coli* counts exceeded the WHO recommended limit for irrigation water used for fresh produce of 1000 faecal coliforms/100 mL.<sup>6</sup> National guidelines set by the DWA associate risk with different *E. coli* levels in irrigation water: 'no risk' is associated with <1 *E. coli*/100 mL, 'low risk' is associated with 1–999 *E. coli*/100 mL and 'high risk' is associated with 1000–3999 *E. coli*/100 mL.<sup>7</sup> Considering these guidelines, the *E. coli* levels at the time of analysis qualify as unacceptably high for the most part. It would therefore be a fair assumption to conclude that, under these conditions, the transfer of microbes from the Plankenburg River via irrigation onto the surface of fresh produce is highly probable.

**Table 3:** *E. coli* load ranges in the Plankenburg River and number of confirmed *E. coli* isolates

Study (sampling site)	<i>E. coli</i> load range	Number of isolates
Study 1 (P-1)	29 000–790 000 MPN/100 mL	27
Study 2 (P-2)	3900–118 500 cfu/mL	30
Study 3 (P-0)	1600–20 000 MPN/100 mL	10
Study 3 (P-1)	250 000–1 000 000 MPN/100 mL	8
Study 3 (P-2)	2500–310 000 MPN/100 mL	6

### *E. coli* with InPEC virulence genes

The 81 *E. coli* strains isolated from the river (Table 3) were positively identified by the API system and their identity was then confirmed using *uidA* PCR. The strains were then screened for InPEC gene sequences and the characteristics of the pathogenic *E. coli* strains identified are presented in Table 4. Of the 81 strains screened (Table 3), five pathogens were identified (one EAEC and four EPEC strains) (Table 4). This result concurs with previous reports that *E. coli* strains that carry pathogenic gene sequences represent 0.9–10% of *E. coli* present in surface waters.<sup>28</sup> It is interesting to note that the three EPEC strains isolated during Study 2 (Table 4) had different biochemical (API) profiles, which suggests that they are not clones.

**Table 4:** Intestinal pathogenic (InPEC) *E. coli* strains isolated

Strain	Source	API profile code	Genogroup	Pathotype
<i>E. coli</i> H45	Study 1 (P-1)	5 0 4 4 5 5 2 5 7	A <sub>1</sub>	EAEC
<i>E. coli</i> A95	Study 2 (P-2)	1 0 4 4 5 7 2 5 7	B2 <sub>3</sub>	EPEC
<i>E. coli</i> A118	Study 2 (P-2)	5 1 4 4 5 5 2 5 7	B1	EPEC
<i>E. coli</i> A132	Study 2 (P-2)	5 0 4 4 5 5 2 5 7	B2 <sub>3</sub>	EPEC
<i>E. coli</i> S49a	Study 3 (P-2)	1 0 4 4 5 7 2 5 7	B2 <sub>3</sub>	EPEC

EAEC, enteroaggregative *E. coli*; EPEC, enteropathogenic *E. coli*

The API profiles for all the pathogenic strains showed that they differed in three respects: production of lysine decarboxylase (LDC) and L-ornithine decarboxylase (ODC) and utilisation of saccharose. Variation in these three properties has been observed before in *E. coli* isolated from untreated surface waters and soil.<sup>29,30</sup> It has furthermore been reported that *E. coli* can induce production of amino acid decarboxylases (such as LDC and ODC) in response to reduced pH conditions.<sup>31</sup> This report illustrates the highly adaptable nature of *E. coli* which helps it survive in more acidic environments.<sup>32</sup>

Because the PCR-based detection method of Clermont<sup>20</sup> was used to identify *E. coli* phylogenetic groups (genogroups), the four main groups (A, B1, B2 and D) can further be subdivided into seven subgroups to increase discrimination: A<sub>0</sub>, A<sub>1</sub>, B1, B2<sub>1</sub>, B2<sub>2</sub>, B2<sub>3</sub>, D<sub>1</sub> and D<sub>2</sub>.<sup>33</sup> It has been reported that ExPEC strains usually belong to genogroups B2 and D, while InPEC strains that cause severe diarrhoea-related diseases (EIEC, EHEC and ETEC) are most commonly classified into genogroups A and B1.<sup>34</sup> *E. coli* strains that cause mild and chronic diarrhoea (EAEC, EPEC and DAEC) can belong to any of the four main phylogenetic groups.<sup>34</sup> It has, however, been reported that most human EPEC strains belong to genogroups B1 or B2.<sup>35</sup> Carlos et al.<sup>36</sup> examined *E. coli* from a variety of different primary hosts (human, goat, chicken, cow, sheep and pig) and found that B2<sub>3</sub> strains in particular could only be found in human samples. It was therefore concluded that the EPEC strains isolated from the river in this study were possibly of human origin, which suggests human faecal contamination of the Plankenburg River. The isolation data

indicates that the contamination might also be constant over time, as similar EPEC strains were isolated during Studies 2 and 3 (Table 4).

### Health implications

The enteroaggregative (EAEC) *E. coli*, detected during the first study (Table 4), affects the small intestine and causes mild but persistent watery diarrhoea ( $\geq 14$  days) in people of all ages. It is frequently found in immunocompromised individuals. As a result of mucinase activity, it can cause mild but significant mucosal damage.<sup>4,37</sup>

Enteropathogenic (EPEC) *E. coli*, of which four were isolated during Studies 2 and 3 (Table 4), primarily affects children and infants, causing profuse watery diarrhoea, fever and nausea. It can also cause disease in certain animals. Outbreaks in developing countries can be quite severe, with reported mortality rates of up to 30%.<sup>37</sup> Adult infections are rare, but possible if high infective doses are combined with substances that can neutralise gastric pH. Certain medical conditions, such as diabetes, can also make adults more susceptible to EPEC infections.<sup>38</sup>

EPEC attachment to the small intestine is a multi-step process which starts with initial attachment to epithelial cells and microcolony stabilisation through bundle-forming pili (BFP), intimate adherence through the production of intimin, and the formation of attaching and effacing lesions and 'pedestal'-like structures.<sup>4,37</sup> Intimin production is facilitated by the locus of enterocyte effacement pathogenicity island which carries the *eae* gene, and is a prerequisite for EPEC and EHEC pathogenicity. BFP is coded for by the *E. coli* adherence factor plasmid, and, although it is considered a virulence contributing factor because of its stabilising role during initial microcolony formation, it is not an absolute requirement for EPEC pathogenicity.<sup>39</sup> Strains that are *eae*(+) and BFP(+) are referred to as typical EPEC (tEPEC), while *eae*(+) and BFP(-) strains are known as atypical EPEC (aEPEC) strains.<sup>40</sup> Although symptoms of aEPEC are milder, with non-inflammatory diarrhoea and no fever or vomiting, it is, however, associated with prolonged diarrhoea (> 14 days).<sup>39</sup> Prolonged diarrhoea for longer than 14 days is clinically associated with an increased risk for illness and death.<sup>40</sup> Although the EPEC strains identified in this study were all *eae* positive, no screening for BFP was done, so the strains could be either tEPEC or aEPEC. However, in both instances, the burden of disease would be significant.

### Antibiotic resistance profiles and ExPEC virulence genes

The presence of pathogenic *E. coli* in this river could result in waterborne or foodborne diseases. Treatment of these diseases could be further complicated if the pathogenic isolates are also resistant to medically important antibiotics. The five InPECs presented in Table 4 were additionally screened for antibiotic susceptibility to seven clinically important antibiotics, as well as for the presence of ExPEC genes. The results are presented in Table 5.

The results showed that the InPEC strains could not be positively confirmed as ExPEC, as they did not have two or more of the ExPEC gene sequences. The EAEC strain (*E. coli* H45) did carry the ExPEC gene sequence *iutA*, which codes for the aerobactin siderophore that

contributes to essential ferric iron uptake and transport in different iron-deficient environments.<sup>41,42</sup> It has been reported that the incidence of aerobactin genes correlates with the presence of highly virulent pathogenic *E. coli* strains.<sup>42</sup> It should, however, be stated that the pathogenic strains were only screened for the most abundant ExPEC gene sequences<sup>19</sup> and that more than 30 other ExPEC genes have been reported.<sup>41</sup> It is thus possible that *E. coli* H45 may be a carrier of other ExPEC gene sequences which were not tested for.

The antibiotics tested for in this study all represented different classes of antimicrobials: aminoglycosides (Streptomycin), fluoroquinolones (Ciprofloxacin), penicillins (Ampicillin), amphenicols (Chloramphenicol), cephalosporins (Cefalotin), dihydrofolate reductase inhibitors (Trimethoprim) and tetracyclines (Tetracycline). Antibiotic resistance testing revealed that, although all of the InPEC strains were resistant to multiple antibiotics, the four EPEC strains (Table 5) can be referred to as multidrug resistant (MDR) strains. This conclusion is based on the accepted definition of MDR which refers to the co-resistance that a strain can have to three or more classes of antimicrobials.<sup>43</sup> This finding concurs with previous observations that multiple antibiotic resistances are common for EPEC.<sup>37</sup>

The most abundant resistances were against Ampicillin (5/5) and Trimethoprim (4/5), followed by Tetracycline (3/5), Streptomycin (2/5) and Chloramphenicol (1/5). Trimethoprim and Ampicillin resistance were furthermore observed in the InPEC strains from all three studies, which showed that Trimethoprim and Ampicillin resistance persisted among bacteria in the river over time. Tetracycline and Streptomycin resistances were limited to strains from Study 2, while Chloramphenicol resistance was only observed in the isolate from Study 3. Antibiotic resistance can either be carried on mobile genetic elements, such as plasmids, which could be easily transferred horizontally between different bacteria, or be as a result of the environmental selection of a chromosomal mutation.<sup>44</sup> Whether chromosomal or plasmid-based, the results showed that antibiotic-resistant *E. coli* that also carries InPEC virulence genes is present in the Plankenburg River.

*E. coli* strains resistant to Tetracycline, Streptomycin, Chloramphenicol and Ampicillin have also been isolated from surface and groundwaters in KwaZulu-Natal and the North-West Province.<sup>45-47</sup> None of these studies tested for Trimethoprim resistance. The widespread occurrence of resistance to Tetracycline, Streptomycin, Chloramphenicol and Ampicillin is not surprising, as they are 'older' antibiotics, some of which have been in use since the 1940s. The fact that antibiotics such as Tetracycline have also been widely used as growth promoters in animal production, could also have contributed to the extent to which resistance has spread in the environment.<sup>48</sup>

The increased antibiotic resistance of Gram-negative bacteria such as *E. coli* is considered a serious global problem, because there is no foreseeable development of new antibiotic classes in the next 10 years.<sup>49</sup> This concern has led to the WHO classification of Ampicillin and Streptomycin as antimicrobials of critical importance for human medicine, while Chloramphenicol, Trimethoprim and Tetracycline are considered as highly important antimicrobials.<sup>49</sup> The occurrence of

**Table 5:** Antibiotic resistance profiles and presence of ExPEC genes in the InPEC strains

Strain	Pathotype	Source	Antibiotic resistance	ExPEC genes
<i>E. coli</i> H45	EAEC	Study 1 (P-1)	Ampicillin; Trimethoprim	<i>iutA</i>
<i>E. coli</i> A95	EPEC	Study 2 (P-2)	Ampicillin; Trimethoprim; Tetracycline	–
<i>E. coli</i> A118	EPEC	Study 2 (P-2)	Ampicillin; Trimethoprim; Tetracycline; Streptomycin	–
<i>E. coli</i> A132	EPEC	Study 2 (P-2)	Ampicillin; Tetracycline; Streptomycin	–
<i>E. coli</i> S49a	EPEC	Study 3 (P-2)	Ampicillin; Trimethoprim; Chloramphenicol	–

EAEC, enteroaggregative *E. coli*; EPEC, enteropathogenic *E. coli*

*E. coli* strains that show resistance to these important antibiotics in the Plankenburg River is therefore a matter of concern.

The presence of antibiotic substances in the aquatic environment is increasing mostly as a result of widespread application in the fields of human medical therapy and agriculture. Antibiotic substances are never fully metabolised in humans or other animals, which means that small amounts are regularly excreted and can enter water streams directly or indirectly via faecal pollution, agricultural run-off or through discharge from wastewater treatment plants.<sup>50,51</sup> The concentrations are usually at sub-inhibitory levels, which contribute to the development and spread of antibiotic resistance properties among environmental bacteria. If human pathogens acquire resistance to antibiotics, a serious situation can arise which, at the very least, can result in increased disease treatment costs. Antibiotic-resistant pathogens that are resistant to multiple antibiotics might, however, also lead to increased morbidity and mortality rates.<sup>51</sup>

## Conclusions

*Escherichia coli* counts in the Plankenburg River have been found to be unacceptably high and this river can thus be classified as a 'high-risk' irrigation water source. Under these conditions, transfer of *E. coli* from the water to produce during irrigation will be highly probable. MDR *E. coli* strains, harbouring intestinal pathogenic gene sequences, were isolated from the Plankenburg River on more than one occasion. Three of the *E. coli* strains carrying enteropathogenic sequences also belonged to the B<sub>2</sub> phylogenetic group, which could indicate human faecal contamination of the Plankenburg River. Subsequently, if irrigated fresh produce contaminated with MDR pathogens is consumed raw, it might act as a direct vehicle for the transmission of disease. Treatment of these MDR pathogenic bacterial related diseases will then be negatively impacted. It is therefore in the public's interest to report the existence of multiple antibiotic-resistant pathogenic *E. coli* in the Plankenburg River, so that corrective action, specifically in terms of treatment of irrigation water prior to irrigation, can be taken as soon as possible.

## Acknowledgements

We thank Dr T.G. Barnard (Water and Health Research Unit, University of Johannesburg) for kindly providing standard InPEC cultures. This study was part of a solicited research project (K5/1773) funded by the Water Research Commission and co-funded by the Department of Agriculture, Forestry and Fisheries (WRC Knowledge Review 2012\_13 KSA4). Financial support provided by the National Research Foundation (IFR2008112100010) is also gratefully acknowledged.

## Authors' contributions

T.J.B. and G.O.S. were the project leaders and were responsible for the project design; T.J.B. and C.L. were responsible for the experimental design and supervision; M.R., N.H., A.C., N.S. and C.L. performed the experiments; C.L. and T.J.B. wrote the manuscript; and G.O.S. made editorial contributions.

## References

1. Touchon M, Hoede C, Tenaillon O, Barbe V, Baeriswyl S, Bidet P, et al. Organised genome dynamics in the *Escherichia coli* species results in highly diverse adaptive paths. *PLoS Gen.* 2009;5:e1000344. <http://dx.doi.org/10.1371/journal.pgen.1000344>
2. Van Elsland JD, Semenov AV, Costa R, Trevors JT. Survival of *Escherichia coli* in the environment: Fundamental and public health aspects. *ISME J.* 2011;5:173–183. <http://dx.doi.org/10.1038/ismej.2010.80>
3. Liang Z, He Z, Powell CA, Stoffella PJ. Survival of *Escherichia coli* in soil with modified microbial community composition. *Soil Biol Biochem.* 2011;43:1591–1599. <http://dx.doi.org/10.1016/j.soilbio.2011.04.010>
4. Kaper JB, Nataro JP, Mobley HL. Pathogenic *Escherichia coli*. *Nature Rev.* 2004;2:123–140.
5. Painter JA, Hoekstra RM, Ayers T, Tauxe RV, Braden CR, Angulo FJ, et al. Attribution of foodborne illnesses, hospitalizations, and deaths to food commodities by using outbreak data, United States, 1998–2008. *Emerg Infect Dis.* 2013;19:407–415. <http://dx.doi.org/10.3201/eid1903.111866>

6. World Health Organization (WHO). Health guidelines for the use of wastewater in agriculture and aquaculture. Technical report series no. 778. Geneva: WHO; 1989.
7. Department of Water Affairs and Forestry (DWA). South African water quality guidelines. Volume 4. In: Holmes S, editor. Agricultural use: Irrigation. 2nd ed. Pretoria: CSIR Environmental Services; 1996. p. 63–68.
8. Britz TJ, Sigge GO, Huisamen N, Kikine T, Ackermann A, Loetter M, et al. Fluctuations of indicator and index microbes as indication of pollution over three years in the Plankenburg and Eerste Rivers, Western Cape, South Africa. *Water SA.* 2013;39:457–465.
9. SANS. Method 5667-6. Water quality – Sampling, Part 6: Guidance on sampling of rivers and streams. Pretoria: Standards South Africa Printers; 2006.
10. Cristensen D, Crawford C, Szabo R. Method MFHPB-19: Enumeration of coliforms, faecal coliforms and of *E. coli* in foods using the MPN method. Health Products and Food Branch. Ottawa: Food Directorate, Health Canada; 2002.
11. Harrigan W, McCance ME. Statistical methods for the selection and examination of microbial colonies. In: Harrigan W, McCance ME, editors. Laboratory methods in food and dairy microbiology. London: Academic Press; 1976. p. 47–49.
12. American Public Health Association (APHA). Standard methods for the examination of water and wastewater. 21st ed. Washington DC: APHA; 2005.
13. SANS. Method 9308-2. Water quality: Detection and enumeration of coliform organisms, thermotolerant coliform and presumptive *Escherichia coli*. Part 2: Multiple tube (most probable number) method. Pretoria: Standards South Africa Printers; 2004.
14. Noble RT, Weisberg SB, Leecaster MK, McGee CD, Ritter K, Walker KO, et al. Comparison of beach bacterial water quality indicator measurement methods. *Environ Monit Assess.* 2003;81:301–312. <http://dx.doi.org/10.1023/A:1021397529041>
15. Gerhardt P, Murray RGE, Costilow RN, Nester EW, Wood WA, Krieg NR, et al., editors. Manual of methods for general bacteriology. Washington DC: American Society for Microbiology; 1981.
16. Aitalhi AD, Hassan SA. Bacterial quality of raw milk investigated by *Escherichia coli* and isolates analysis for specific virulence-gene markers. *Food Cont.* 2009;20:913–917. <http://dx.doi.org/10.1016/j.foodcont.2009.01.005>
17. Heijnen L, Medema G. Quantitative detection of *E. coli*, *E. coli* O157 and other shiga toxin producing *E. coli* in water samples using a culture method combined with real-time PCR. *J Water Health.* 2006;4:487–498.
18. Omar KB, Barnard TG. The occurrence of pathogenic *Escherichia coli* in South African waste water treatment plants as detected by multiplex PCR. *Water SA.* 2010;36:172–179.
19. Xia X, Meng J, Zhao S, Jones S, Gaines S, Ayers S, et al. Identification and antimicrobial resistance of extraintestinal pathogenic *Escherichia coli* from retail meats. *J Food Protec.* 2011;74:38–44. <http://dx.doi.org/10.4315/0362-028X.JFP-10-251>
20. Clermont O, Bonacorsi S, Bingen E. Rapid and simple determination of the *Escherichia coli* phylogenetic group. *Appl Environ Microbiol.* 2000;66:4555–4558. <http://dx.doi.org/10.1128/AEM.66.10.4555-4558.2000>
21. Tarr CL, Large TM, Moeller CL, Lacher DW, Tarr PI, Acheson DW, et al. Molecular characterization of a serotype O121:H19 clone, a distinct shiga toxin-producing clone of pathogenic *Escherichia coli*. *Infect Immun.* 2002;70:6853–6859. <http://dx.doi.org/10.1128/IAI.70.12.6853-6859.2002>
22. Lopez-Saucedo C, Cerna JF, Villegas-Sepulveda N, Thompson R, Velazquez FR, Torres J, et al. Single multiplex polymerase chain reaction to detect diverse loci associated with diarrheagenic *Escherichia coli*. *Emerg Infect Dis.* 2003;9:127–131. <http://dx.doi.org/10.3201/eid901.010507>
23. Moses AE, Garbati MA, Egwu GO, Ameh JA. Detection of *E. coli* O157 and O26 serogroups in human immunodeficiency virus-infected patients with clinical manifestation of diarrhea in Maiduguri, Nigeria. *Res J Med Med Sci.* 2006;1(4):140–145.
24. Pass MA, Odedra R, Batt RM. Multiplex PCRs for identification of *Escherichia coli* virulence genes. *J Clinical Microbiol.* 2000;38:2001–2004.
25. Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial disk susceptibility tests; approved standard. 10th ed. Volume 29 number 1. CLSI document M02-A10. Wayne, PA: CLSI; 2009.

26. Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing; twenty-first informational supplement. 31(1). CLSI document M100-S21. Wayne, PA: CLSI; 2011.
27. Andrews JM. BSAC standardized disc susceptibility testing method (version 8). BSAC Working Party on Susceptibility Testing. J Antimicrob Chemother. 2009;64:454–489. <http://dx.doi.org/10.1093/jac/dkp244>
28. Masters N, Wiegand A, Ahmed W, Katouli M. *Escherichia coli* virulence genes profile of surface waters as an indicator of water quality. Water Res. 2011;45:6321–6333. <http://dx.doi.org/10.1016/j.watres.2011.09.018>
29. Janezic KJ, Ferry B, Hendricks EW, Janiga BA, Johnson T, Murphy S, et al. Phenotypic and genotypic characterization of *Escherichia coli* isolated from untreated surface waters. Open Microbiol J. 2013;7:9–19. <http://dx.doi.org/10.2174/1874285801307010009>
30. Brennan FP, Abram F, Chinalia FA, Richards KG, O'Flaherty V. Characterization of environmentally persistent *Escherichia coli* isolates leached from an Irish soil. Appl Environ Microbiol. 2010;76 (7):2175–2180. <http://dx.doi.org/10.1128/AEM.01944-09>
31. Kanjee U, Gutsche I, Alexopoulos E, Zhao B, El Bakkouri M, Thibault G, et al. Linkage between the bacterial acid stress and stringent responses: The structure of the inducible lysine decarboxylase. EMBO J. 2011;30:931–944. <http://dx.doi.org/10.1038/emboj.2011.5>
32. Maurelli AT, Fernandez RE, Bloch CA, Rode CK, Fasano A. 'Black holes' and bacterial pathogenicity: A large genomic deletion that enhances the virulence of *Shigella* spp. and enteroinvasive *Escherichia coli*. Proc Natl Acad Sci USA. 1998;95:3943–3948. <http://dx.doi.org/10.1073/pnas.95.7.3943>
33. Escobar-Paramo P, Le Menac'h A, Le Gall T, Amorin C, Gouriou S, Picard B, et al. Identification of forces shaping the commensal *Escherichia coli* genetic structure by comparing animal and human isolates. Environ Microbiol. 2006;8:1975–1984. <http://dx.doi.org/10.1111/j.1462-2920.2006.01077.x>
34. Hamelin K, Bruant G, El-Shaarawi A, Hill S, Edge TA, Fairbrother J, et al. Occurrence of virulence and antimicrobial resistance genes in *Escherichia coli* isolates from different aquatic ecosystems within the St. Clair river and Detroit river areas. Appl Environ Microbiol. 2007;73:477–484. <http://dx.doi.org/10.1128/AEM.01445-06>
35. Tramuta C, Robino P, Nebbia P. Phylogenetic background of attaching and effacing *Escherichia coli* isolates from animals. Vet Res Commun. 2008;32:433–437. <http://dx.doi.org/10.1007/s11259-008-9042-1>
36. Carlos C, Pires MM, Stoppe NC, Hachich EM, Sato MIZ, Gomes TAT, et al. *Escherichia coli* phylogenetic group determination and its application in the identification of the major animal source of fecal contamination. BMC Microbiol. 2010;10:161–172. <http://dx.doi.org/10.1186/1471-2180-10-161>
37. Torres AG. Intestinal pathogenic *Escherichia coli*. In: Barrett ADT, Stanberry LR, editors. Vaccines for biodefence and emerging and neglected diseases. New York: Elsevier; 2009. p. 1013–1029. <http://dx.doi.org/10.1016/B978-0-12-369408-9.00051-2>
38. Nataro JP, Kaper JB. Diarrheagenic *Escherichia coli*. Clin Microbiol Rev. 1998;11(1):142–201.
39. Nguyen RN, Taylor LS, Tauschek M, Robins-Browne RM. Atypical enteropathogenic *Escherichia coli* infection and prolonged diarrhea in children. Emerg Infect Dis. 2006;12:597–603. <http://dx.doi.org/10.3201/eid1204.051112>
40. Nataro JP. Atypical enteropathogenic *Escherichia coli*: Typical pathogens? Emerg Infect Dis. 2006;12:696–699. <http://dx.doi.org/10.3201/eid1204.060125>
41. Chapman TA, Wu XY, Barchia I, Bettelheim KA, Driesen S, Trott D, et al. Comparison of virulence gene profiles of *Escherichia coli* strains isolated from healthy and diarrheic swine. Appl Environ Microbiol. 2006;72:4782–4795. <http://dx.doi.org/10.1128/AEM.02885-05>
42. Garénaux A, Caza M, Dozois CM. The ins and outs of siderophore mediated iron uptake by extra-intestinal pathogenic *Escherichia coli*. Vet Microb. 2011;153:89–98. <http://dx.doi.org/10.1016/j.vetmic.2011.05.023>
43. Doyle MP, Loneragan GH, Scott HM, Singer RS. Antimicrobial resistance: Challenges and perspectives (Institute of Food Technologists - Expert Report). Comp Rev Food Sci Food Safety. 2013;12:234–248. <http://dx.doi.org/10.1111/1541-4337.12008>
44. Akter S, Rafiq-Un N, Rupa FA, Bari MDL, Hossain MAC. Antibiotic resistance and plasmid profiles in bacteria isolated from market fresh vegetables. Agric Food Anal Bacteriol. 2011;1(2):140–149.
45. Olaniran AO, Naicker K, Pillay B. Antibiotic resistance profiles of *Escherichia coli* isolates from river sources in Durban, South Africa. Water SA. 2009;25:174–183.
46. Wose Kinge CN, Ateba CN, Kawadza DT. Antibiotic resistance profiles of *Escherichia coli* isolated from different water sources in the Mmabatho locality, North-West Province, South Africa. S Afr J Sci. 2010;106(1/2):44–49.
47. Phokela PT, Ateba CN, Kawadza TK. Assessing antibiotic resistance profiles in *Escherichia coli* and *Salmonella* species from groundwater in the Mafikeng area, South Africa. African J Microbiol Res. 2011;5(32):5902–5909.
48. Tadesse DA, Zhao S, Tong E, Ayers S, Singh A, Bartholomew MJ, et al. Antimicrobial drug resistance in *Escherichia coli* from humans and food animals, United States, 1950–2002. Emerg Infect Dis. 2012;18(5):741–749. <http://dx.doi.org/10.3201/eid1805.111153>
49. World Health Organization (WHO). Critically important antimicrobials for human medicine, 3rd revision. Geneva: WHO; 2012.
50. Lupo A, Coyne S, Berendonk TU. Origin and evolution of antibiotic resistance: The common mechanisms of emergence and spread in water bodies. Front Microbiol. 2012;3(18):1–13.
51. Li B, Zhang T. Different removal behaviours of multiple trace antibiotics in municipal wastewater chlorination. Water Res. 2013;47:2970–2982. <http://dx.doi.org/10.1016/j.watres.2013.03.001>

