# 1 The energetics of fish growth and how it constrains food-web trophic structure

- 2 Diego R. Barneche<sup>1,\*</sup>, Andrew P. Allen<sup>2</sup>
- 3 <sup>1</sup>Centre for Geometric Biology, School of Biological Sciences, Monash University, Clayton Vic 3800, Australia
- 4 <sup>2</sup>Department of Biological Sciences, Macquarie University, Sydney NSW 2109, Australia
- <sup>5</sup> Present address: School of Life and Environmental Sciences, The University of Sydney, Camperdown NSW
   6 2006, Australia
- 7 Statement of authorship: D.R.B. compiled the data, D.R.B. and A.P.A. conceived the study,
- 8 analyzed the data, and wrote the manuscript.
- 9 Data accessibility statement: All data and R code (data manipulation, analyses, figures and
- 10 tables) can be downloaded from a GitHub repository (https://github.com/dbarneche/FishGrowth),
- 11 which will be made publicly available upon publication.
- 12 Running title: Energetics from individuals to ecosystems
- 13 Keywords: physiology, climate change, fisheries, efficiency, biomass, productivity, trophic
- 14 pyramids
- 15 **Type of article:** Letters
- 16 Word Count: 4,910 (main text), 147 (Abstract)
- 17 Number of references: 48
- 18 Number of figures: 4
- 19 Number of tables: 1
- 20 Correspondence should be sent to: Diego R. Barneche, Centre for Geometric Biology /
- 21 School of Biological Sciences, Monash University, Clayton, Vic. 3800, Australia (Tel: +61
- 22 (3) 9905 5100; Email: barnechedr@gmail.com).

# 23 ABSTRACT

The allocation of metabolic energy to growth fundamentally influences all levels of biologi-24 25 cal organization. Here we use a first-principles theoretical model to characterize the energetics of fish growth at distinct ontogenetic stages and in distinct thermal regimes. Empirically, 26 we show that the mass scaling of growth rates follows that of metabolic rate, and is some-27 what steeper at earlier ontogenetic stages. We also demonstrate that the cost of growth,  $E_m$ , 28 varies substantially among fishes, and that it may increase with temperature, trophic level, and 29 level of activity. Theoretically, we show that  $E_m$  is a primary determinant of the efficiency of 30 energy transfer across trophic levels, and that energy is transferred more efficiently between 31 trophic levels if the prey are young and sedentary. Overall, our study demonstrates the impor-32 tance of characterizing the energetics of individual growth in order to understand constraints 33 on the structure of food webs and ecosystems. 34

# 35 INTRODUCTION

Organisms must expend energy to gather, consume, and transform the resources necessary to 36 produce biomass, so the rate of biomass production is fundamental at multiple biological lev-37 els (Lindeman 1942). At the individual level, it influences fitness by constraining how quickly 38 an organism reaches maturity and subsequently produces offspring (Brown et al. 1993). At 39 the population level, it constrains the intrinsic rate of population increase (Savage et al. 2004). 40 At the community level, it constrains how much energy and materials can flow to the next 41 trophic level in a food web (Andersen et al. 2009; Irigoien et al. 2013). And, at the ecosystem 42 level, the fraction of assimilated energy lost in producing that biomass (through respiration) 43 limits the total heterotrophic metabolism, and hence the number of trophic levels, that can be 44 supported in a food web (Lindeman 1942; Pauly & Christensen 1995). 45

We still lack a comprehensive theoretical understanding of the energetics of growth, despite recent progress (e.g. West *et al.* 2001; Hou *et al.* 2008; Kooijman 2009). Fish are excellent model organisms for evaluating such a theory because they encompass the highest global species richness among vertebrates (> 33,000 species), they range in body mass by over > 8 orders of magnitude (~0.1 g – 1 × 10<sup>7</sup> g), and they occupy diverse freshwater and marine habitats that vary substantially in thermal regime (~0 – 40°C; Froese & Pauly 2017).

Understanding the energetics of growth is of immense practical importance. For example, in 52 fisheries management, knowing how long wild fish stocks take to achieve maturity, and how 53 much food they need to do so, is crucial for establishing sustainable yields in fisheries man-54 agement (Szuwalski et al. 2017). Moreover, sustainable yields at ocean-basin scales may in 55 the future be affected by changes in environmental variables such as temperature through their 56 effects on the growth energetics of individual fish (e.g. van Rijn et al. 2017). Extensive em-57 58 pirical work has documented determinants of fish growth (e.g. Houde 1989); however, these studies have typically focused on how one or few species at a particular life stage respond to 59 a particular set of environmental conditions. With few notable exceptions (see, for example, 60 Pauly & Pullin 1988; Houde 1989; Charnov & Gillooly 2004; Sibly et al. 2015), there has 61 been little attempt to generalize determinants of individual-level biomass production across 62 multiple fish species within a theoretical framework. 63

64 Theoretical work on the mechanisms underlying growth dynamics has focused primarily on understanding why individuals tend to follow a sigmoid growth trajectory over ontogeny 65 such that mass-specific growth rate is rapid during early life stages, but slows down as indi-66 67 viduals approach an asymptotic adult size. More than half a century ago, Ludwig von Bertalanffy (1938) proposed that these sigmoid growth trajectories arise because the overall rate 68 of catabolism increases more rapidly with size than the rate of anabolism. While the mecha-69 nistic basis of this model (hereafter BGM) has been questioned by many, including fisheries 70 scientists (e.g. Enberg et al. 2008), the BGM is frequently employed for fish because it of-71 ten provides a reasonable statistical fit to ontogenetic growth data. Importantly, however, the 72 73 BGM is usually fitted using widely-available data on fish length, rather than less-available data on mass, which is unfortunate given that growth is fundamentally an energetic process, 74 and that the energetic costs of growth are related to changes in mass (West et al. 2001; Hou et 75 al. 2008; Moses et al. 2008). 76

West *et al.* (2001) proposed an ontogenetic growth model (see also Gillooly *et al.* 2002;
Moses *et al.* 2008) that is based on principles of allometry and mass and energy balance. Hou *et al.* (2008) subsequently expanded this model (hereafter OGM) by explicitly partitioning
the total metabolic rate into active, growth, and maintenance components. Consequently,
the OGM predicts that growth rates are inextricably linked to the size- and temperature-

dependencies of metabolic rate, consistent with empirical data (Pauly & Pullin 1988; Houde
1989; Atkinson 1994; Gillooly *et al.* 2002; Brown *et al.* 2004). Although the OGM can be
mathematically similar in functional form to the BGM (Makarieva *et al.* 2004), its conceptual
foundations are fundamentally different. In particular, while growth is an anabolic process,
maintenance in the OGM involves both anabolism and catabolism because, for example,
protein turnover is fueled by respiration.

88 Of particular relevance, the OGM reveals the importance of a particular parameter,  $E_m$ , which is the amount of energy that must be expended in respiration to produce a fixed quan-89 tity of biomass (West et al. 2001; Hou et al. 2008; Moses et al. 2008). This quantity has 90 91 been shown to vary substantially within and among different clades of animals (Wieser 1994; Moses et al. 2008; Sears et al. 2012). However, to our knowledge, is still unknown to what 92 extent this variation reflects differences in physiology versus environmental and/or ecological 93 factors. Understanding how and why  $E_m$  varies is fundamental because, as we will demon-94 strate later, this quantity constrains food-web structure via its effects on the efficiency of en-95 96 ergy transfer between trophic levels.

97 In this study, we use the OGM as a framework to characterize the mass and temperature dependence of growth rates for marine and freshwater fishes. In so doing, we first quantify 98 99 the fraction of total metabolic energy allocated to biomass production across different species 100 at differing life stages and temperature regimes. Using these data, we then test three hypothe-101 ses: H1: Growth rates exhibit the same mass scaling exponent as metabolic rate at different 102 ontogenetic stages; H2:  $E_m$  is temperature independent; and H3:  $E_m$  is independent of ecological variables such as trophic level and level of activity. Finally, we combine our empirical 103 estimates of  $E_m$  with novel theory to explore how it constrains food-web structure through its 104 effects on the efficiency of energy transfer between trophic levels. 105

#### 106 MATERIALS AND METHODS

# 107 1. Theory and hypotheses

## 108 Hypothesis H1

109 The OGM (West et al. 2001; Hou et al. 2008) is derived based on energy balance for an or-

110 ganism with a wet mass of *m* (in g), a resting metabolic rate of  $B_r$  (i.e. J d<sup>-1</sup>), and a growth rate 111 per unit time, *t* (d), of dm/dt (g d<sup>-1</sup>),

$$\frac{dm}{dt} = \frac{B_r}{E_m} \left[ 1 - \left(\frac{m}{M}\right)^{1-\alpha} \right] = \frac{B_o(T)}{E_m} m^\alpha \left[ 1 - \left(\frac{m}{M}\right)^{1-\alpha} \right],\tag{1}$$

where  $B_r = B_o(T)m^{\alpha}$ ,  $B_o(T)$  is a metabolic normalization constant independent of body mass 112  $(J g^{-\alpha} d^{-1})$  that varies among fish taxa and with absolute body temperature (Barneche *et al.*) 113 2014), T(K),  $\alpha$  is a dimensionless mass-scaling exponent that is theoretically predicted to take 114 a value of 3/4 in the fractal-like distribution model of West *et al.* (1997), and  $E_m$  is the en-115 ergy expended to produce one unit of biomass (J g<sup>-1</sup>). The expression  $[1 - (m/M)^{1-\alpha}]$  is the 116 fraction of resting metabolic energy that is allocated to growth. This fraction approaches 0 as 117 118 the organism approaches its asymptotic adult mass, M, at which point all resting metabolic 119 energy is allocated to maintenance, which is assumed to remain constant on a mass-specific 120 basis over ontogeny.

121 Hypothesis H1: Growth rates exhibit the same mass scaling exponent as metabolic rate at 122 different ontogenetic stages. The OGM states that the mass scaling of growth rates are gov-123 erned by the effects of body mass on resting metabolic rates,  $B_r$ . Empirically, it has been observed that early in ontogeny, the scaling of metabolic rates in fish can be substantially steeper 124 125 (Bochdansky & Leggett 2001) than the canonical 3/4 value observed for adult fish (Barneche et al. 2014). Whether these shifts in metabolic mass scaling over ontogeny are also observed 126 for growth rates remains poorly explored. However, this phenomenon can be empirically 127 assessed by fitting growth-rate data at different ontogenetic stages to a function of the same 128 129 form as that commonly fitted to metabolic rates (eqns S1–S3),

$$\frac{dm}{dt} = g_o m^{\gamma} e^{\frac{E_g}{k} \left(\frac{1}{T_s} - \frac{1}{T}\right)} I(T)^{-1}, \qquad (2)$$

130 where  $g_o$  is a normalized growth rate that is independent of mass and temperature (g 131  $^{1-\gamma} d^{-1}$ ) at some arbitrary standardized temperature,  $T_s$  (K), k is the Boltzmann con-132 stant (8.62 × 10<sup>-5</sup> eV K<sup>-1</sup>),  $\gamma$  is a dimensionless mass-scaling exponent, and  $E_g$  (eV) 133 is an activation energy (Barneche *et al.* 2014). With respect to size, eqn 1 predicts that  $g_o \propto [1 - (m/M)^{1-\alpha}]$ , implying a decline in normalized rates with increasing ontogenetic 135 stage, m/M. With respect to temperature,  $e^{\frac{E_g}{k}(\frac{1}{T_s} - \frac{1}{T})}$  is the Boltzmann relationship that char-136 acterizes temperature-induced enhancement of growth rates below the temperature optimum,  $T_{opt}$  (K), using an activation energy,  $E_g$  (eV) (Gillooly *et al.* 2002). The inactivation term,  $I(T) = \left(1 + \left(\frac{E_g}{E_i - E_g}\right)e^{\frac{E_i}{k}\left(\frac{1}{T_{opt}} - \frac{1}{T}\right)}\right)$ , characterizes declines in rates at temperature *T* above  $T_{opt}$  using an inactivation parameter,  $E_i$  (Schoolfield *et al.* 1981; Barneche *et al.* 2014).

#### 140 Hypotheses H2–H3

141 The respiratory energy expended in producing a unit of biomass,  $E_m$ , can be calculated by 142 rearranging eqn 1:

$$E_m = B_r \frac{dt}{dm} [1 - (m/M)^{1-\alpha}].$$
 (3)

143 In practice, calculating  $E_m$  using eqn 3 requires ontogenetic growth data in order to estimate 144  $[1 - (m/M)^{1-\alpha}]$ . In the absence of such data, an upper bound estimate for  $E_m$ ,  $E_m^*$ , can be 145 calculated using estimates of growth rate taken early in ontogeny

$$E_m^* \approx B_r \frac{dt}{dm},\tag{4}$$

146 when the mass of an individual is negligible compared to the asymptotic adult mass,  $m \ll M$ 147 (Moses *et al.* 2008).

The quantity  $E_m$  reflects energy allocated to growth via both direct and indirect paths (Hou 148 149 et al. 2008). Thus, its magnitude does not correspond to the combustion energy (i.e. chemical-150 energy content) of assimilated biomass (Makarieva et al. 2004). Rather, it corresponds to 151 the sum of all direct and indirect energy costs that an individual must expend in producing biomass. The direct costs of synthesizing biomass may, in fact, be lower than the combustion 152 153 energy because, for example, proteins may be constructed from pre-formed amino acids assimilated from food. The indirect costs include other processes that are not directly related to 154 155 biomass production, but that are nevertheless essential for this production to occur (e.g. digestion). While disentangling and quantifying the processes contributing to  $E_m$  are challenging, 156 the overall magnitude of  $E_m$  can nevertheless be quantified based on eqns 3 and 4. 157

*Hypothesis H2:*  $E_m$  *is temperature independent.* In the OGM, the temperature dependence of growth rates is assumed to be governed by the effects of temperature on the resting metabolic rate normalization,  $B_o(T)$  (eqn 1; Gillooly *et al.* 2002). A corollary of this assumption is that  $E_m$  is invariant with respect to environmental temperature. This hypothesis can be readily assessed by investigating the relationship between empirically-estimated values of  $E_m$ (eqns 3 and 4) and the environmental temperature at which organisms grow.

164 Hypothesis H3:  $E_m$  is independent of ecological variables such as trophic level and level of activity. While the OGM assumes invariance of  $E_m$  over ontogeny, it currently makes no 165 166 predictions regarding variation in  $E_m$  among species. In fact, it is known empirically that  $E_m$ 167 varies across multiple clades of animals (Wieser 1994; Moses et al. 2008; Sears et al. 2012). For fish, the possible causes for this variation are still unknown, but may be related to ecologi-168 cal variables. For instance, fishes at higher trophic levels may have generally higher  $E_m$  values 169 at least in part because trophic level is positively associated with muscle protein (Killen et al. 170 2016), which may be expensive to generate. For similar reasons, caudal aspect ratio (=  $h^2/s$ , 171 172 where h is the height of the caudal fin and s its surface area), a proxy for activity level (Froese 173 & Pauly 2017), may also be positively correlated with  $E_m$ . Here we explore these ideas using Hypothesis H3, invariance of  $E_m$ , as a null expectation. 174

# 175 The role of $E_m$ in constraining trophic efficiency and food-web structure

Understanding how and why  $E_m$  varies is of fundamental importance to predicting constraints on the efficiency of energy transfer between trophic levels (Economo *et al.* 2005; Andersen *et al.* 2009). In fact,  $E_m$  is a primary determinant of the fraction of assimilated food that is transferable to the next trophic level, as we will now show. The rate of energy assimilation by an individual of size m, A(m), is (Hou *et al.* 2008)

$$A(m) = B_{tot} + E_c \frac{dm}{dt} = fB_r + E_c \frac{B_r}{E_m} \left[ 1 - \left(\frac{m}{M}\right)^{1-\alpha} \right] = B_o(T)m^{\alpha} \left[ f + \frac{E_c}{E_m} \left[ 1 - \left(\frac{m}{M}\right)^{1-\alpha} \right] \right],$$
(5)

181 where  $E_c$  is the combustion energy of biomass (~24 kJ g<sup>-1</sup> dry mass; Hou *et al.* 2008), and

 $B_{tot}$  is the total (i.e. field) metabolic rate, which is defined as the sum of the resting metabolic 182 rate,  $B_r$ , and the active rate that sustains locomotion, feeding, and other activities. Following 183 Hou *et al.* (2008), we assume that  $B_{tot} = fB_r$  in the expression above, where f is a dimen-184 sionless parameter referred to as 'activity scope', which is constrained to be greater than 1 and 185 less than the ratio of maximum metabolic rate to resting metabolic rate (i.e. "factorial aero-186 bic scope" sensu Killen et al. 2016). Using eqns 1, 3 and 5, we can calculate the fraction of 187 188 energy assimilated by a prey individual,  $\varepsilon$ , that is transferred to the next trophic level through consumption of prey biomass by a predator. This fraction is calculated by taking the ratio of 189 190 the amount of energy contained in a prey item consumed at size m,  $E_cm$ , and the total lifetime-191 *integrated* amount of energy that the prey item has assimilated from birth to its age at consumption,  $t^*$ , 192

$$\varepsilon = \frac{E_c m}{\int_{t=0}^{t^*} A(m(t)) dt},\tag{6}$$

where m(t) is mass at age t (eqns S4–S6), and  $t^* = -E_m \ln \left[1 - \left(\frac{m}{M}\right)^{1-\alpha}\right] / (B_o(T)M^{1-\alpha}(1-t))$ 193  $\alpha$ )) is an approximation to the age at which the prey was consumed, assuming that the prey's 194 195 mass at birth  $m_0$ , is negligible relative to its asymptotic size ( $m_0/M \approx 0$  in eqn S6). Eqn 6 196 quantitatively predicts that if a prey individual is consumed by a predator at an earlier life his-197 tory stage,  $m/M \approx 0$ , a larger fraction of the prey's assimilated energy,  $\varepsilon$ , will be transferable 198 to its predator. It is important to note that these efficiencies represent upper-bound estimates because they only incorporate energy losses attributable to the respiration of the prey, and thus 199 exclude other factors affecting the energy transfer (e.g. food assimilation efficiency by preda-200 201 tors). Eqn 6 can be readily integrated numerically to calculate how maximum transfer efficiency,  $\varepsilon$ , changes with  $E_m$ , relative ontogenetic stage, m/M, and activity scope, f. 202

We can also use eqn 6 to expand theoretical predictions obtained from a static sizespectrum model (Brown & Gillooly 2003). This model predicts a specific relationship between total biomass, W, and individual body mass, m,

$$W \propto m^{1 + \frac{\ln(\varepsilon)}{\ln(PPMR)} - \alpha},\tag{7}$$

- 206 based on the size-scaling of metabolic rate ( $\alpha$ ) and the assumptions that the predator-prey
- 207 body-mass ratio, *PPMR*, and the efficiency of energy transfer between trophic levels,  $\varepsilon$ , are
- 208 held constant moving between trophic levels (Brown & Gillooly 2003).

#### 209 2. Data compilation

We use two independent datasets to test hypotheses *H1–H3*. Dataset I comprises 179 experimentally measured paired estimates of growth rate (g wet mass d<sup>-1</sup>) and metabolic rate (J d<sup>-1</sup>) taken early in ontogeny (25 measurements for embryos and 154 measurements for larvae), meaning that  $m/M \approx 0$ , and hence that  $[1 - (m/M)^{1-\alpha}] \approx 1$ . These data encompass 18 species of marine and freshwater fishes with body masses of 9  $\mu$ g – 0.5 g and temperatures of 3 – 28°C (Supplementary Information).

Dataset II was obtained from FishBase (Froese & Pauly 2017). It contains 2,849 sets of pa-216 217 rameter estimates, corresponding to 2,849 ontogenetic growth curves, that were obtained by fitting the von Bertalanffy growth model (BGM) (von Bertalanffy 1957) to age and size data 218 collected from 503 species of marine and freshwater fish species. As is the tradition in fish-219 220 eries science, these growth curves were characterized using *length* rather than mass. To obtain standardized mass-based estimates of growth rate, we calculate optimum (i.e. maximum) 221 222 growth rates,  $g_{opt}$ , at mass  $m_{opt}$ , by combining these length-based data with species-specific 223 estimates of the mass-length conversion parameters. These  $g_{opt}$  estimates encompass 62 fam-224 ilies and a temperature range of -0.9–30°C. In the Supplementary Information, we derive an analytical approximation (eqns S4-S16) that justifies using BGM-estimated optimum growth 225 rates to assess hypotheses derived from the OGM. 226

#### 227 3. Testing hypotheses H1–H3

*H1*. The size and temperature dependence of growth rates from Dataset I were estimated byfitting eqn 2 to log-transformed data,

$$\ln\left[\frac{dm}{dt}\right] = \ln g_o + \gamma \ln m + \frac{E_g}{k} \left(\frac{1}{T_s} - \frac{1}{T}\right) - \ln I(T).$$
(8)

We fit eqn 8 using a Bayesian procedure implemented in the R package *brms* version 2.1.0 (Bürkner 2017) in order to derive posterior distributions and associated 95% credible intervals (CIs) for the fitted parameters (Table 1). A hierarchical modeling approach was adopted for this analysis with species-level random effects on the intercept (Supplementary Information). For the analysis of Dataset II, the size and temperature dependencies of optimum growth rates,  $g_{opt}$ , at size  $m_{opt}$  were estimated by fitting a simplified version of eqn 2, without the inactivation term for the temperature dependence to log-transformed data

$$\ln g_{opt} = \ln g_o + \gamma \ln m_{opt} + \frac{E_g}{k} \left( \frac{1}{T_s} - \frac{1}{T} \right), \tag{9}$$

which is equivalent to the simple Boltzmann relationship commonly used in the early 237 Metabolic Theory of Ecology (MTE) literature (Gillooly et al. 2002; Brown et al. 2004). In 238 239 contrast to Dataset I, the growth-rate data from Dataset II yielded little evidence of temperature optima (Supplementary Information), most likely because these data were obtained from 240 241 field-captured individuals, which are not expected to occur above their optimum temperatures in nature. While the lack of well-defined optima provides empirical justification for fitting 242 243 the simpler Boltzmann relationship to estimate an overall temperature dependence, this 244 approach obscures any changes in temperature dependence over the temperature range. It 245 also results in negative bias for estimated activation energy if a temperature optimum exists. 246 A hierarchical modeling approach was adopted for this analysis, with family-level random effects on the intercept, mass-scaling exponent and temperature-dependence slope ( $\Delta \gamma, \Delta E_g$ , 247  $\Delta \ln g_o$ ; Supplementary Information). 248

249 H2 and H3. In Dataset I, we used the paired measurements of metabolic and growth rates to obtain upper-bound estimates for the energy expended in growth,  $E_m^*$  (eqn 4), on a wet mass 250 basis. In Dataset II, estimates of  $E_m$  (wet mass basis) were calculated using eqn 3 by combin-251 252 ing a subset of the  $g_{opt}$  estimates (n = 582) with estimates of standard metabolic rate, which 253 account for the effects of body size and temperature (Barneche et al. 2014), and trophic level (Killen et al. 2016) (Supplementary Information). In the fish literature, resting rates are re-254 255 ferred to as 'standard rates' at some specified temperature because the metabolic rate of a resting fish will vary with ambient temperature. Calculations of  $E_m$  were restricted to 13 families 256

with growth rate and standard metabolic rate data. The maintenance-correction term in eqn 3,  $[1 - (m_{opt}/M)^{1-\alpha}]$ , was assumed to be 0.23 for these calculations, which corresponds to the overall size-scaling exponent for fish resting metabolic rates,  $\alpha = 0.77$  (Supplementary Information).

We assessed whether  $E_m^*$  (Dataset I) and  $E_m$  (Dataset II) exhibit temperature dependence (hypothesis *H2*), and whether  $E_m$  (Dataset II only) varies with aspect ratio and trophic level (hypothesis *H3*) by fitting a linear model of the form

$$\ln E_m = \ln \beta_0 + \beta_T T + \beta_L \ln L + \beta_A \ln(A+1), \tag{10}$$

where  $\beta_0$ ,  $\beta_T$ ,  $\beta_L$ , and  $\beta_A$  are respectively the fixed-effect model intercept and three slope pa-264 rameters, T is temperature in degrees Celsius, L (ranging from 2 to 4.4) is a continuous vari-265 able for trophic level, and A is the caudal fin aspect ratio (ranging from 0 to 2.99). Species-266 267 level estimates of the latter two ecological measures were downloaded from FishBase to assess hypothesis H3 for the  $E_m$  estimates in Dataset II. When species values were not available, 268 269 congeneric species values were used. The relationships of these ecological measures to  $E_m^*$ 270 were not investigated for Dataset I because the measures are not representative of trophic level or morphology for individuals at early ontogenetic stages. Model parameters were assigned 271 vague priors in a Bayesian modelling framework. We also included normally distributed ran-272 dom effects for  $\ln\beta_0$  and  $\beta_T$ , thereby allowing them to vary by species (Dataset I) and family 273 (Dataset II). The model fitting procedure in brms is similar to that described for eqns 8 and 9 274 275 (Supplementary Information).

### 276 RESULTS AND DISCUSSION

## 277 Hypothesis H1

The growth rate models (eqns 8 and 9) for both datasets reveal substantial species- (Dataset I) and family-level (Dataset II) variation in size- and temperature-corrected growth rates, characterized by standard deviations  $\sigma_{\Delta \ln g_o}$ . Particularly, the estimate from Dataset I implies a ~1.6fold change in growth rates among species ( $\approx e^{2 \times 0.22}$ , with  $\sigma_{\Delta \ln g_o} = 0.22$ ), while the estimate from Dataset II implies an ~2.2-fold change in growth rates among families ( $\approx e^{2 \times 0.39}$ , with  $\sigma_{\Delta \ln g_o} = 0.39$ ). After accounting for maintenance costs attributable to age in Dataset II (i.e.  $[1 - (m/M)^{1-\alpha}] = [1 - (m_{opt}/M)^{(1-0.77)}] \approx 0.23$ ), the average estimate of  $\ln g_o$  is only about 3-fold higher for Dataset I than for Dataset II (Table 1). Therefore, the mass dependencies of growth rates for Datasets I and II can be reasonably approximated by a single allometric function that spans 10 orders of magnitude in body mass (Fig. 1).

288 Contrary to hypothesis H1, the analyses of Datasets I and II yield contrasting estimates for 289 the mass scaling of growth rates, characterized by the scaling exponent  $\gamma$  (Table 1). The av-290 erage within-species  $\gamma$  is significantly steeper than 0.75 early in ontogeny (Dataset I average: 291 0.85; 95% CI: 0.80–0.91), but statistically indistinguishable from 0.75 at the size of optimum growth rate (Dataset II average: 0.77; 95% CI: 0.74–0.80). It is possible that the true scaling 292 293 exponent in Dataset I would approach a value of 1 (i.e. isometric mass scaling) if variation in energy allocation to maintenance with ontogenetic stage, m/M, were accounted for. One 294 295 would need full ontogenetic growth trajectories for these individuals (as we have for Dataset II) - including asymptotic adult mass, which can vary with temperature and other factors - in 296 297 order to estimate their maintenance costs.

298 Overall, our results are consistent with previous observations that metabolic scaling is 299 steeper early in ontogeny for fish (e.g. Bochdansky & Leggett 2001). They also highlight the 300 importance of correction for ontogeny in order to properly evaluate mass scaling of growth 301 rates among species (Moses et al. 2008), and reinforce the idea that the scaling of biological 302 rates early in ontogeny may be steeper than the canonical value of 3/4 that is generally as-303 sumed by MTE (West et al. 1997), perhaps due to changes in the circulatory system during larval-post larval transition. The model of West et al. (1997), in fact, predicts positive de-304 viations from 3/4-power scaling if size range encompasses very small organisms that have 305 306 vascular distribution networks with only a few levels of branching. Evaluating more detailed predictions would require data on aorta and capillary diameters, and the numbers of branching 307 generations from aorta to capillary (West et al. 1997, 2001; Brummer et al. 2017). 308

## 309 Hypotheses H2–H3

310 The average temperature dependence of growth rates differ between Datasets I and II. While

311 it is statistically indistinguishable from the predicted 0.6–0.7 eV range for Dataset I (average for  $E_g$ : 0.65 eV; 95% CI: 0.48–0.93 eV), consistent with previous estimates of temperature 312 dependence of developmental times in fishes (Pauly & Pullin 1988; Gillooly et al. 2002), it is 313 significantly shallower than this value for Dataset II (average for  $E_g$ : 0.26 eV; 95% CI: 0.19– 314 0.34 eV). This unexpected low temperature dependence might be due in part to errors in age 315 estimation in seasonally-growing species (Campana 2001), statistical artifact of errors intro-316 317 duced by estimating masses from length-weight functions, and/or negative bias for the estimated activation energy due to our inability to account for temperature optima. It might also 318 reflect differences in how laboratory-grown (Dataset I) versus field-grown (Dataset II) fish re-319 320 spond to temperature, perhaps owing to (i) differences in resource availability, or (ii) changes 321 over ontogeny.

Estimates of  $E_m^*$  and  $E_m$  cover a similar range of values, and vary by ~3 orders of magnitude (Fig. 2). While these estimates highlight substantial variation in the calculated amount of energy an organism must expend in producing biomass, we note that they are comparable in magnitude to values reported for other groups of animals including insects, bivalves, anurans, birds and mammals (Wieser 1994; Moses *et al.* 2008; Sears *et al.* 2012).

327 Consistent with hypothesis H2, the estimated upper-bound  $E_m^*$  was independent of tem-328 perature in 78% of the 18 species in Dataset I (i.e. 95% credible intervals overlapped 0;  $\beta_T$ 329 = -0.04, 95% CI: -0.08-0.01; Fig. 2). In contrast, we observed a significant temperature dependence of  $E_m$  in 92% of the 13 families for which we could calculate  $E_m$  in Dataset II, with 330 an average 22-fold increase in  $E_m$  moving from 0°C to 30°C ( $\beta_T = 0.1032, 95\%$  CI: 0.0701– 331 0.1411; Fig. 2). Thus, these results suggest that a fish in the tropics requires substantially 332 333 more assimilated energy to produce a unit of biomass than a fish in the polar regions. The discrepancy we observe with regard to temperature dependence for  $E_m$  between Datasets I and II 334 335 warrants further investigation and careful scrutiny. Given this important caveat, our results for Dataset II are consistent with earlier accounts of decreasing growth efficiency with increasing 336 temperature in fishes (e.g. Houde 1989; Jobling 1997; McCarthy et al. 1998) and microbes 337 (Apple et al. 2006). 338

After accounting for the effects of temperature, contrary to hypothesis *H3*, we found substantial evidence (i.e. 95% posterior credible intervals did not overlap 0) of systematic in341 creases in  $E_m$  with fish trophic level ( $\beta_L = 0.73$ , 95% CI: 0.34–1.12), and aspect ratio ( $\beta_A =$ 342 0.41, 95% CI: 0.10–0.71). Therefore, our results are consistent with the fact that species with 343 higher muscle protein content might exhibit higher metabolic costs to build biomass. How-344 ever, top predators may have higher assimilation efficiencies (Sanders *et al.* 2016). Thus, our 345 findings may reveal some important physiological trade-offs (i.e. assimilation efficiency ver-346 sus cost of growth) that might occur across trophic levels, which will have fundamental effects 347 on the trophic structure of food webs (see below).

## 348 The role of $E_m$ in constraining trophic efficiency and food-web structure

349 We integrated the assimilation rate numerically in eqn 6 to calculate the curves for Fig. 3. 350 Results of these calculations demonstrate that the efficiency of energy transfer varies consider-351 ably with  $E_m$  over the range of values that were empirically observed in Fig. 2. In fact, the effects of  $E_m$  are of comparable magnitude to those of ontogenetic stage, highlighting the quan-352 353 titative importance of this variable for understanding energy transfers between trophic levels. Moreover, eqn 6 also predicts that higher values of activity scope (f) will have substantial 354 355 negative effects on the efficiency of energy transfer, highlighting the ecological importance that this variable has to food-web trophic structure (Killen et al. 2016). 356

357 The calculated range of efficiencies depicted in Fig. 3 were combined with parameters 358 from eqn 7 in order to predict how ecosystem-level size structure may change in relation to prey growth energetics,  $E_m$ , and ontogenetic stage, m/M (Fig. 4). For instance, our model 359 360 indicates that the existence of real closed-system top-heavy pyramids may be energetically and ecologically unlikely (e.g. Trebilco et al. 2016) - red area in Fig. 4 - because individu-361 als would need to be characterized by low values of  $E_m$ , and would have to prey exclusively 362 363 on very young offspring (i.e. high-efficiency prey resource). Moreover, compared to aquatic food-webs where the youngest prey available are comprised of eggs and larvae (i.e.  $m/M \approx$ 364 0), communities of viviparous animals (e.g. mammals with large offspring relative to adult 365 366 size), might be constrained to have a bottom-heavy trophic pyramid at least in part because the youngest prey items available (e.g. newborn cubs) are relatively large relative to their asymp-367 totic size and thus relatively inefficient at accruing biomass after birth. Future refinement and 368 tests of these predictions will require an assessment of ontogenetic stage of prey items in diets 369

370 of different species at the community level.

371 Our approach builds on a simple static size spectrum model (Brown & Gillooly 2003), but 372 could inform more sophisticated contemporary dynamic size spectrum models that explicitly connect ontogenetic growth to population and ecosystem dynamics (e.g. Andersen et al. 2016; 373 374 Blanchard et al. 2017). For example, these models incorporate ontogenetic growth in a manner that is more closely related to the BGM. Recognizing that the efficiency of energy transfer 375 376 changes with the relative ontogenetic stage of the prey, and that this efficiency might change with temperature, trophic and activity levels through their effects on  $E_m$  (but see Barange *et al.* 377 2014 for recent integration of temperature effects) may therefore represent a fundamental step 378 379 forward.

380 Finally, our findings might also yield important insights in terms of fisheries management. 381 For example, in oligotrophic coastal marine communities such as coral reefs, mass-spawning events are very frequent (e.g. Harrison et al. 1984), and high-efficiency recycling of energy 382 and nutrients might be imperative. Thus, preserving large individuals that produce more (and 383 larger) eggs in aquatic communities (Birkeland & Dayton 2005) may be key to the mainte-384 nance of high-efficiency energy transfers between trophic levels. Understanding the energetics 385 of growth across different trophic levels (and/or functional groups) might therefore help estab-386 387 lish baselines of recovery potential in coastal fisheries (MacNeil et al. 2015).

# 388 FUTURE DIRECTIONS

Overall, our study demonstrates how growth rate and metabolic rate data can be synthesized 389 390 within a theoretical framework to obtain a deeper understanding of the energetics of growth 391 (Fig. 1). Particularly, our results reveal fundamental constraints (Fig. 2) on ecosystem dynamics that arise from individual-level metabolism (Figs. 3 and 4). Our analysis highlight general 392 393 trends, but also important differences among datasets, as well as among species, particularly with regards to the temperature dependence of  $E_m$ . New methods are needed to quantify and 394 predict  $E_m$  – across a broad range of taxa in both terrestrial and aquatic systems – based on the 395 underlying energetics of the biochemical processes involved in the construction of biomass. 396 397 Uncovering the drivers of variation in this single quantity (e.g. temperature, mode of activity, habitat, phylum) is a fruitful avenue forward: it offers an opportunity to bridge multiple dis-398

399 ciplines – from physiology to phylogenetics to ecosystem ecology – across space and time.

400 Such an interdisciplinary approach is needed if we are to predict how rising temperatures are

401 going to affect life-history evolution, ecosystem dynamics, the rehabilitation of economically

402 important fisheries stocks, and the global carbon cycle.

## 403 ACKNOWLEDGEMENTS

We thank the Associate Editor and two anonymous referees for constructive criticism that
helped improve earlier versions of this paper. This project was supported by Macquarie University (PhD scholarship to D.R.B.), and the Centre for Geometric Biology at Monash University (Post-doctoral fellowship to D.R.B.).

#### 408 **REFERENCES**

409 1. Al-Habsi, S.H., Sweeting, C.J., Polunin, N.V.C. & Graham, N.A.J. (2008). δ15N and δ13C

410 elucidation of size-structured food webs in a Western Arabian Sea demersal trawl assemblage.
411 *Marine Ecology Progress Series*, 353, 55–63.

412 2. Andersen, K., Beyer, J. & Lundberg, P. (2009). Trophic and individual efficiencies of size413 structured communities. *Proceedings of the Royal Society of London B: Biological Sciences*,
414 276, 109–114.

415 3. Andersen, K.H., Jacobsen, N.S. & Farnsworth, K.D. (2016). The theoretical foundations
416 for size spectrum models of fish communities. *Canadian Journal of Fisheries and Aquatic*417 *Sciences*, 73, 575–588.

418 4. Apple, J.K., del Giorgio, P.A. & Kemp, W.M. (2006). Temperature regulation of bacterial
419 production, respiration, and growth efficiency in a temperate salt-marsh estuary. *Aquatic Mi*420 *crobial Ecology*, 43, 243–254.

421 5. Atkinson, D. (1994). Temperature and organism size–A biological law for ectotherms? *Ad-*422 *vances in Ecological Research*, 25, 1–58.

423 6. Barange, M., Merino, G., Blanchard, J.L., Scholtens, J., Harle, J. & Allison, E.H. et al.

424 (2014). Impacts of climate change on marine ecosystem production in societies dependent

- 425 on fisheries. Nature Climate Change, 4, 211–216.
- 426 7. Barneche, D.R., Kulbicki, M., Floeter, S.R., Friedlander, A.M., Maina, J. & Allen, A.P.
- 427 (2014). Scaling metabolism from individuals to reef-fish communities at broad spatial scales.
- 428 *Ecology Letters*, 17, 1067–1076.
- 8. Birkeland, C. & Dayton, P.K. (2005). The importance in fishery management of leaving the
  big ones. *Trends in Ecology & Evolution*, 20, 356–358.
- 431 9. Blanchard, Julia L., Heneghan, Ryan F., Everett, Jason D., Trebilco, Rowan & Richardson,
- 432 A.J. (2017). From bacteria to whales: Using functional size spectra to model marine ecosys-
- 433 tems. Trends in Ecology & Evolution, 32, 174–186.
- 434 10. Bochdansky, A.B. & Leggett, W.C. (2001). Winberg revisited: Convergence of routine
- 435 metabolism in larval and juvenile fish. *Canadian Journal of Fisheries and Aquatic Sciences*,
  436 58, 220–230.
- 437 11. Brown, J.H. & Gillooly, J.F. (2003). Ecological food webs: High-quality data facilitate
  438 theoretical unification. *Proceedings of the National Academy of Sciences*, 100, 1467–1468.
- 439 12. Brown, J.H., Gillooly, J.F., Allen, A.P., Savage, V.M. & West, G.B. (2004). Toward a
  440 Metabolic Theory of Ecology. *Ecology*, 85, 1771–1789.
- 441 13. Brown, J.H., Marquet, P.A. & Taper, M.L. (1993). Evolution of body size: Consequences
  442 of an energetic definition of fitness. *The American Naturalist*, 142, 573–584.
- 443 14. Brummer, A.B., Savage, V.M. & Enquist, B.J. (2017). A general model for metabolic
- scaling in self-similar asymmetric networks. *PLOS Computational Biology*, 13, 1–25.
- 445 15. Bürkner, P. (2017). brms: An R package for Bayesian multilevel models using Stan. *Jour-*446 *nal of Statistical Software, Articles*, 80, 1–28.
- 16. Campana, S.E. (2001). Accuracy, precision and quality control in age determination, including a review of the use and abuse of age validation methods. *Journal of Fish Biology*, 59,
  197–242.
- 450 17. Charnov, E.L. & Gillooly, J.F. (2004). Size and temperature in the evolution of fish life
  451 histories. *Integrative and Comparative Biology*, 44, 494–497.
- 452 18. Economo, E.P., Kerkhoff, A.J. & Enquist, B.J. (2005). Allometric growth, life-history

- 453 invariants and population energetics. *Ecology Letters*, 8, 353–360.
- 454 19. Enberg, K., Dunlop, E.S. & Jørgensen, C. (2008). Fish growth. In: Encyclopedia of ecol-
- 455 ogy (eds. Jørgensen, S.E. & Fath, B.D.). Elsevier, Oxford, pp. 1564–1572.
- 456 20. Froese, R. & Pauly, D. (2017). FishBase.
- 457 21. Gillooly, J.F., Charnov, E.L., West, G.B., Savage, V.M. & Brown, J.H. (2002). Effects of
- 458 size and temperature on developmental time. Nature, 417, 70-73.
- 459 22. Harrison, P.L., Babcock, R.C., Bull, G.D., Oliver, J.K., Wallace, C.C. & Willis, B.L.
- 460 (1984). Mass spawning in tropical reef corals. *Science*, 223, 1186–1189.
- 461 23. Hou, C., Zuo, W., Moses, M.E., Woodruff, W.H., Brown, J.H. & West, G.B. (2008). En-
- 462 ergy uptake and allocation during ontogeny. *Science*, 322, 736–739.
- 463 24. Houde, E.D. (1989). Comparative growth, mortality, and energetics of marine fish larvae:
- 464 Temperature and implied latitudinal effects. Fishery Bulletin, 87, 471–495.
- 465 25. Irigoien, X., Klevjer, T.A., Røstad, A., Martinez, U., Boyra, G. & Acuña, J.L. et al.
- 466 (2013). Large mesopelagic fishes biomass and trophic efficiency in the open ocean. *Nature*467 *Communications*, 5, 1–10.
- 468 26. Jobling, M. (1997). Temperature and growth: Modulation of growth rate via temperature
- 469 change. In: Society for experimental biology seminar series 61: Global warming: Implica-
- 470 tions for freshwater and marine fish (eds. Wood, C.M. & McDonald, D.G.). Book Section.
- 471 Cambridge University Press, Cambridge, pp. 225–254.
- 472 27. Killen, S.S., Glazier, D.S., Rezende, E.L., Clark, T.D., Atkinson, D. & Willener, A.S.T.
- 473 et al. (2016). Ecological influences and morphological correlates of resting and maximal
- 474 metabolic rates across teleost fish species. The American Naturalist, 187, 592-606.
- 475 28. Kooijman, S.A.L.M. (2009). Dynamic energy budget theory for metabolic organisation.
- 476 3rd edn. Cambridge University Press, Cambridge, UK.
- 477 29. Lindeman, R.L. (1942). The trophic-dynamic aspect of ecology. *Ecology*, 23, 399–417.
- 478 30. MacNeil, M.A., Graham, N.A.J., Cinner, J.E., Wilson, S.K., Williams, I.D. & Maina, J. et
- 479 al. (2015). Recovery potential of the world's coral reef fishes. Nature, 520, 341-344.
- 480 31. Makarieva, A.M., Gorshkov, V.G. & Li, B.-L. (2004). Ontogenetic growth: Models and

- 481 theory. *Ecological Modelling*, 176, 15–26.
- 482 32. McCarthy, I., Moksness, E. & Pavlov, D.A. (1998). The effects of temperature on growth
  483 rate and growth efficiency of juvenile common wolffish. *Aquaculture International*, 6, 207–
  484 218.
- 485 33. Moses, M.E., Hou, C., Woodruff, W.H., West, G.B., Nekola, J.C. & Zuo, Wenyun et al.
- 486 (2008). Revisiting a model of ontogenetic growth: Estimating model parameters from theory
  487 and data. *The American Naturalist*, 171, 632–645.
- 488 34. Pauly, D. & Christensen, V. (1995). Primary production required to sustain global fish489 eries. *Nature*, 374, 255–257.
- 490 35. Pauly, D. & Pullin, R.S.V. (1988). Hatching time in spherical, pelagic, marine fish eggs in
- 491 response to temperature and egg size. Environmental Biology of Fishes, 22, 261–271.
- 492 36. Sanders, D., Moser, A., Newton, J. & van Veen, F.J.F. (2016). Trophic assimilation ef-
- 493 ficiency markedly increases at higher trophic levels in four-level hostparasitoid food chain.
  494 *Proceedings of the Royal Society of London B: Biological Sciences*, 283.
- 495 37. Savage, V.M., Gillooly, J.F., Brown, J.H., West, Geoffrey B. & Charnov, E.L. (2004).
- 496 Effects of body size and temperature on population growth. *The American Naturalist*, 163,497 429–441.
- 498 38. Schoolfield, R.M., Sharpe, P.J.H. & Magnuson, C.E. (1981). Non-linear regression of bi-
- 499 ological temperature-dependent rate models based on absolute reaction-rate theory. *Journal of*500 *Theoretical Biology*, 88, 719–731.
- 39. Sears, K.E., Kerkhoff, A.J., Messerman, A. & Itagaki, H. (2012). Ontogenetic scaling of
  metabolism, growth, and assimilation: Testing metabolic scaling theory with *Manduca sexta*
- 503 larvae. *Physiological and Biochemical Zoology*, 85, 159–173.
- 40. Sibly, R.M., Baker, J., Grady, J.M., Luna, S.M., Kodric-Brown, A. & Venditti, C. et al.
- 505 (2015). Fundamental insights into ontogenetic growth from theory and fish. Proceedings of
- 506 the National Academy of Sciences, 112, 13934–13939.
- 507 41. Szuwalski, C.S., Burgess, M.G., Costello, C. & Gaines, S.D. (2017). High fishery catches
- 508 through trophic cascades in china. Proceedings of the National Academy of Sciences, 114,

509 717-721.

- 510 42. Trebilco, R., Dulvy, N.K., Anderson, S.C. & Salomon, A.K. (2016). The paradox of in-
- 511 verted biomass pyramids in kelp forest fish communities. *Proceedings of the Royal Society of*512 London B: Biological Sciences, 283.
- 513 43. van Rijn, I., Buba, Y., DeLong, J., Kiflawi, M. & Belmaker, J. (2017). Large but uneven
- reduction in fish size across species in relation to changing sea temperatures. *Global Change Biology*, 23, 3667–3674.
- 516 44. von Bertalanffy, L. (1938). A quantitative theory of organic growth (Inquiries on growth
- 517 laws. ii). Human Biology, 10, 181–213.
- 518 45. von Bertalanffy, L. (1957). Quantitative laws in metabolism and growth. *The Quarterly*519 *Review of Biology*, 32, 217–231.
- 46. West, G.B., Brown, J.H. & Enquist, B.J. (1997). A general model for the origin of allometric scaling laws in biology. *Science*, 276, 122–126.
- 47. West, G.B., Brown, J.H. & Enquist, B.J. (2001). A general model for ontogenetic growth. *Nature*, 413, 628–631.
- 48. Wieser, W. (1994). Cost of growth in cells and organisms: General rules and comparative
  aspects. *Biological Reviews*, 69, 1–33.

### 526 TABLES

527 Table 1. Point estimates and 95% credible intervals (as determined using Bayesian methods) for fitted parameters in the growth rate models. Fixed-effect parameters include:  $\gamma$ , the 528 average for the mass-dependence of growth rate;  $E_g$ , the average for the temperature depen-529 530 dence of growth rate;  $\ln g_o$ , the average for the mass-corrected growth rate at temperature  $T_s$ = 15°C;  $T_{opt}$  (Dataset I only), the temperature optimum of fish growth rates and  $E_i$  (Dataset I 531 only), the inactivation parameter describing the rate of decline in growth rate at temperatures 532  $> T_{opt}$ . Random-effects include the standard deviations for species- (Dataset I) and family-533 level (Dataset II) variation in size- and temperature-corrected rates at  $T_s$  ( $\Delta \ln g_o$ ) as well as 534 standard deviation for family-level size dependence ( $\Delta \gamma$ ) and temperature dependence  $\Delta E_g$ 535 536 (Dataset II only).

I Π Parameter Mean 2.5% 97.5% Mean 2.5% 97.5% **Fixed Effects** Mass,  $\gamma$ 0.85 0.80 0.91 0.77 0.74 0.80 Activation energy,  $E_g$  (eV) 0.65 0.48 0.93 0.26 0.19 0.34 Inactivation parameter,  $E_i$  (eV) 2.67 1.18 4.74 Optimum temperature,  $T_{opt}$  (K) 302.47 296.91 309.69 Normalization,  $\ln g_o$  (g g<sup>- $\gamma$ </sup> d<sup>-1</sup>) -3.04 -3.38 -2.57 -5.52 -5.67 -5.34 537 **Random effects** Std. Deviation of  $\Delta \ln g_o$ 0.22 0.04 0.47 0.39 0.20 0.61 Std. Deviation of  $\Delta \gamma$ 0.07 0.04 0.11 Std. Deviation of  $\Delta E_{g}$ 0.24 0.32 0.17 Correlation of  $\Delta \ln g_o$  and  $\Delta \gamma$ -0.70-0.90 -0.28 Correlation of  $\Delta \ln g_o$  and  $E_g$ 0.04 -0.43 0.53 0.08 -0.34 Correlation of  $\Delta \gamma$  and  $\Delta E_g$ 0.49

#### 538 FIGURE LEGENDS

Figure 1. Scaling of growth rates of fish with respect to (left) body mass and (right) and tem-539 540 perature in Datasets I (white squares) and II (grey circles). Parameter estimates (listed in Table 1) were obtained using Bayesian methods. The effect of temperature on growth rate was con-541 trolled for by standardizing the temperature measures, T (in K), to  $T_s = 288.15$  K (= 15°C) 542 based on temperature scaling relationships, where k is the Boltzmann constant (8.62  $\times$  10<sup>-5</sup> 543 eV K<sup>-1</sup>). The effect of body mass was controlled for by standardizing measures to 1 gram 544 based on the mass scaling relationships. The mass-corrected rates at temperature  $T_s$ ,  $\ln g_o$ , 545 for Dataset I (-3.04  $g^{1-\gamma} d^{-1}$ ) and Dataset II (-5.52  $g^{1-\gamma} d^{-1}$ ), correspond to averages across 546 547 species and families, respectively. Dataset I was fitted to a model that allowed  $lng_{0}$  to vary among species, whereas Dataset II was fitted to a model that allowed  $\ln g_o$ ,  $\gamma$ , and  $E_r$  to vary 548 among families. Growth rates from Dataset II are optimum growth rates  $(g_{opt})$  and mass is 549 mass at optimum growth rates  $(m_{opt})$ . Thus, the ontogenetic stage was controlled for (eqn 1) 550 by expressing growth rates as  $(dm/dt)[1 - (m/M)^{1-\alpha}]$ , assuming  $[1 - (m/M)^{1-\alpha}] = 0.23$  for 551 Dataset II. For Dataset I, it was assumed that  $[1 - (m/M)^{1-\alpha}] = 1$  because growth rates were 552 measured at an early ontogenetic stage. 553

Figure 2. Distributions of values for  $E_m$  (in J g<sup>-1</sup> wet mass), the amount of energy necessary 554 to produce a unit of biomass at both embryo/larval growth rate (white squares, Dataset I), and 555 556 maximum growth rate (grey circles, Dataset II). Values from Dataset I are upper-bound estimates of  $E_m^*$  obtained from 179 direct measurements of growth rates and metabolic rates 557 (assuming  $[1 - (m/M)^{1-\alpha}] = 1$ , eqn 4). Values from Dataset II are  $E_m$  estimates (eqn 3) cal-558 culated for 582 growth-rate measurements, which encompass 13 families that overlap with the 559 560 standard metabolic-rate data from FishBase (Barneche et al. 2014). Equations in the top-left represent the fixed-effect estimates of the temperature dependence of  $E_m^*$  and  $E_m$  for Datasets I 561 562 and II, respectively.

**Figure 3.** Upper-bound estimates for the efficiency of energy transfer given different values for ontogenetic stages, m/M, and  $E_m$  (J g<sup>-1</sup>) (eqns 1, 3, 4, 5 and 6). Efficiencies only incorporate energy losses due to respiration, and thus exclude losses attributable to other processes. Polygons were calculated for different values of  $E_m$  (wet mass basis) which encompass the range of values estimated in Fig. 2, assuming that  $m_o = 0$  g,  $\alpha = 0.77$ ,  $B_o = 18.67$  J g<sup>-0.77</sup> d<sup>-1</sup> for fish at 15°C (Table S1), and  $E_c = (24 \text{ kJ g}^{-1} \text{ dry mass}) (0.15 \text{ g dry mass g}^{-1} \text{ wet mass}) = 3,600 \text{ J g}^{-1}$  wet mass (Hou *et al.* 2008). Polygon upper and lower bounds represent the efficiencies of individuals either at permanent resting (f = 1) or active (f = 4) state. An average f= 4 is consistent with evidence reported for fishes (Killen *et al.* 2016). The values for *m* (size at the time of consumption) and *M* (asymptotic adult size of the prey individual) are arbitrary because  $\varepsilon$  depends only on their ratio.

Figure 4. Relationship between ontogenetic stage of prey at time of predation and energy nec-574 essary to produce a unit of biomass  $E_m$ , and the resulting effects on the size structuring of bi-575 ological communities and the energy transfer efficiency between trophic levels ( $\varepsilon$ , eqns 5, 6 576 and 7). Different colors indicate different resulting community biomass (W) – individual body 577 mass (m) scaling relationships, with blue areas depicting bottom-heavy pyramids and red areas 578 depicting top-heavy pyramids. The black solid line represents an area where  $W \propto m^0$ , i.e. a 579 biomass stack, which corresponds to an average energy transfer efficiency of 0.14. The values 580 581 in the figure were calculated assuming PPMR = 2327:1 (following Al-Habsi et al. 2008),  $\alpha$  = 582 0.77, and f = 2.4 in eqn 5.