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Use of a Robust Dehydrogenase from an Archael Hyperthermophile in Asymmetric Catalysis-Dynamic Reductive Kinetic Resolution Entry into (S)-Profens

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Hyperthermophilic archaea are of great interest in evolutionary microbiology, owing to their ability to withstand high temperatures, and often extremes of pressure, pH and salinity. Enzymes from these organisms¹ may offer particular opportunities for asymmetric synthesis, complementary to approaches with mesophilic enzymes,² or those involving enzyme³ and pathway⁴ reengineering. However, perhaps due to a bias that hyperthermophilic enzymes have "narrow substrate specificities,"⁵ archaeal extremophiles remain a largely untapped resource in asymmetric synthesis.⁶

Herein, we disclose a remarkably general Dynamic Reductive Kinetic Resolution (DYRKR) entry into (*S*)-profens, including several important NSAIDs. The enzyme employed is alcohol dehydrogenase (ADH)-10, one of 13 annotated ADHs in the hyperthermophile *Sulfolobus solfataricus*. Protein phylogenetic analysis of this paralogous family indicates SsADH-10 is most closely related to homologues in distant taxa (Fig. 1). The highest identity between SsADH-10 and any other SsADHs is only 34%, suggesting that the SsADH family was established prior to the emergence of other archaeal lineages. Though not described as such, the SsADH-10 appears to be the only SsADH isozyme for which structural information is available in the pdb.⁷

The requisite, 2-arylpropionaldehydes were readily assembled via Pd(0)-catalyzed arylation of *t*-butyl propionate under Buchwald-Hartwig-type⁸ conditions, followed by reduction to the aldehyde (LDBBA⁹ or LAH/DMP oxid-*see* SI). Optimal DYRKR conditions (Table 1-80°C, pH 9) led to efficient throughput of *rac*-aldehyde to the (*S*)-2-arylpropionaldehyde, particularly with *m*- and *p*-substitution. Notably, (*S*)-profenols corresponding to the NSAIDs naproxen (**3b**, scaled to 1 gram @ 98% yield and 95% ee), ibuprofen (**3d**, IP), flurbiprofen (**3h**, FIP), fenoprofen (**3j**, FP) and ketoprofen (**3l**, KP) were obtained in excellent yields (up to 96%) and high enantioselectivity (up to 99 %).

Naproxen is FDA-approved as the active (S)-antipode. While most individuals can invert (R)-ibuprofen to the (S)-antipode, the pathway is inefficient for KP^{10a} and FIP. ^{10b} Moreover, the recent observation that the profen-CoA thioester intermediates inhibit G6PDH, ^{10c} argues for "chiral switching" to single (S)-antipodes. ^{10d} Entries into (S)-profens ^{11,12} include asymmetric

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hydrogenation (NP 98% ee, IP 97% ee), ^{11g} and hydroformylation (IP, 92% ee). ^{11f} DKR processes include enantioselective crystallization (NP >99% ee), ^{11b} DYRKR with H₂ as reductant under Ru(II) catalysis (IP 92% ee), ^{11d} and lipase/Ru(II)-mediated-DKR of allylic acetates, followed by Cu-mediated Grignard-arylation (FIP 97% ee; ^{11c} Knochel arylation: ^{11e} IP 97% ee). The hydrovinylation/oxidation approach is impressive (IP, FP, FIP, NP >96% ee), ^{11a} but access to KP requires late stage arylation. Thus, the broad side chain tolerance of SsADH-10 makes the method presented here among the most generally (*S*)-selective.

To explore how these extended hydrophobic substrates bind to SsADH-10, docking was carried out (Fig. 2) for the (*S*)-antipodes of flurbiprofenal, naproxenal, ketoprofenal and fenoprofenal. A detailed discussion of the approach and results is provided in the SI. Briefly, W95 is seen as enforcing (*S*)-selectivity, with ligands clustering into two distinct distal ring binding modes. "Channel-gating"-L272 and L295 appear to form a hydrophobic pocket for naproxenal and flurbiprofenal. For the more flexible ketoprofenal and fenoprofenal, edge-to-face interactions with W117 and F49 are proposed.

From a practical viewpoint, we have also found that SsADH-10 may be engaged in a "thermal recycling" approach that may be generalizable to other hyperthermophilic enzymes. Namely, while 30 vol% cosolvent is often needed to dissolve hydrophobic DH substrates, ¹³ we use a higher T (80°C) @ just 5% EtOH (solvent and biorenewable reductant). Importantly, upon completion of the reaction, cooling to rt allows the product to precipitate and be collected by filtration (*see TOC graphic and* SI). Reclaimed SsADH may be recycled (5 cycles @ 94-96 % ee). Given the growing interest in thermophilic enzymes in synthesis, ^{1,14} and in engineering thermostability into mesophilic enzymes, ¹⁵ this "thermal switching" approach is likely to find broad application, well beyond the domain of geothermal dehydrogenases.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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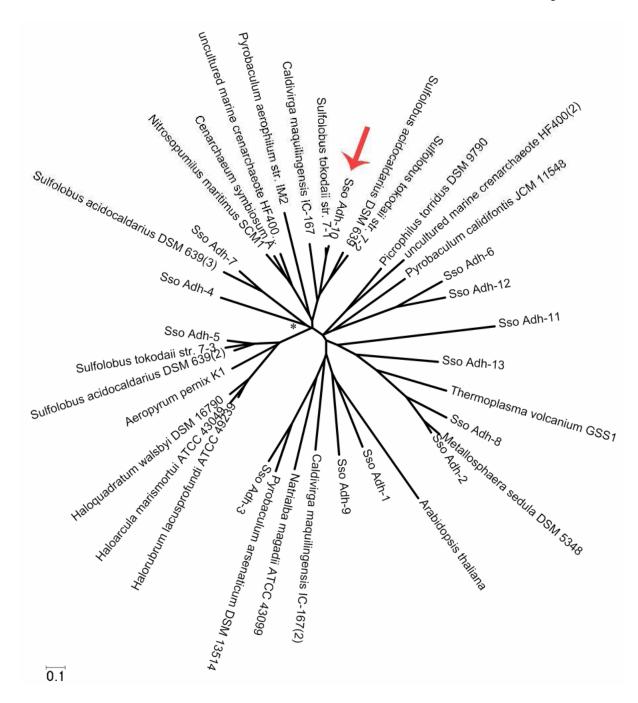
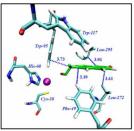
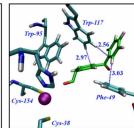


Figure 1. Protein phylogeny of the SsADH proteins. A consensus neighbor joining distance tree is shown of all SsADHs and homologues of highest sequence identity in related taxa. Distances are indicated by the bar (lower left corner) and represent 10 substitutions per 100 residues. Percent occurrence among 100 trees was greater than 50% for all nodes except those indicated with an asterisk.







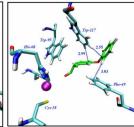
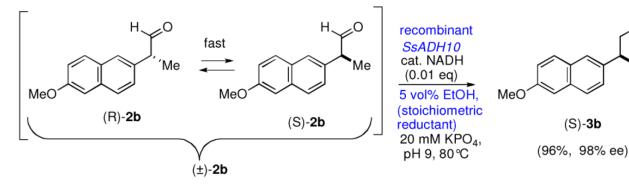


Figure 2. Structures of thermally relaxed (GROMACS 4.07) SsADH-10 (from 1R37) to which has been docked (Autodock Vina - left to right-): (i) (*S*)-flurbiprofenal (ii) (*S*)-naproxenal (iii) (*S*)-ketoprofenal and (iv) (*S*)-fenoprofenal (Zn ligation sphere: H68, C38, C154 and substrate carbonyl)

Table 1

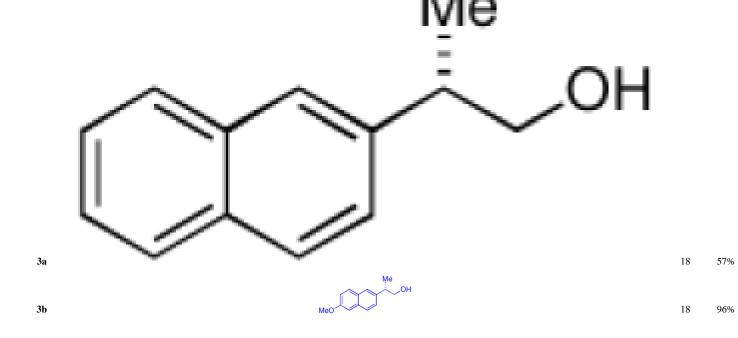
SsADH10-Mediated DYRKR Entry into Profenols



Cpd No. DYRKR Yield (%)b Product^a t(h)

HO.

(S)-3b



3e

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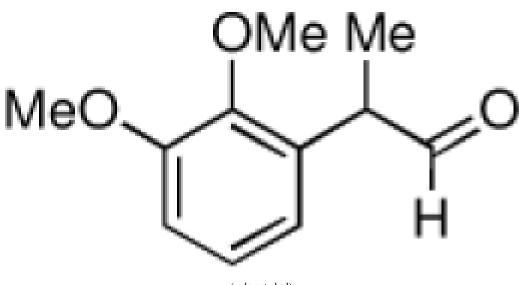
PYRR Product^d (b) (%)

Me, OH

3c 18 90%

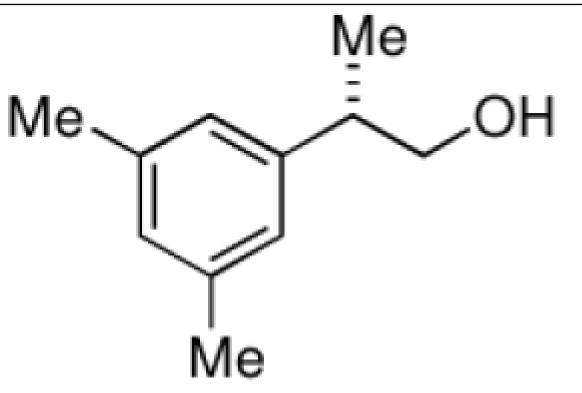
18

74%



3g

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18

99%

Cpd No.	DYRKR Product ^a	t(h)	Yield $(\%)^b$
	F OH		
3h	<u> M</u> e	18	77%
3i	F ₃ C OH	24	55%
3j	ме он Он	12	85%
3k	Me OH	18	95%
31		18	85%

 $^a\mathrm{DYRKR}$ performed on a 1 mmol scale (1 mol% NADH; 5 vol% EtOH)

 $^{^{}b}$ Isolated yields

 $^{^{\}it C}_{\rm }$ ee's by chiral LC or GC. Blue - profen drug precursor.