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39. HIGH-RESOLUTION PLIOCENE–PLEISTOCENE BIOSTRATIGRAPHY OF SITE 959, EASTERN EQUATORIAL ATLANTIC OCEAN¹

Im Chul Shin,^{2,3} Samir Shafik,⁴ and David K. Watkins²

ABSTRACT

High-resolution calcareous nannofossil biostratigraphy was examined from Cores 159-959C-1H through 8H in 20-cm intervals for the Ocean Drilling Program in the eastern equatorial Atlantic Ocean. Well-preserved marker species occur continuously and are relatively abundant in Hole 959C. Six zones (CN10 through CN15) are identified. All calcareous nannofossils are well preserved. Late Neogene (Pliocene–Pleistocene) sediment is dominated by *Florispheera profunda*, *Gephyrocapsa caribbeanica*, *G. oceanica*, *Gephyrocapsa* spp., *Reticulofenestra pseudoumbilica*, *R. minutula*, and small *Reticulofenestra*. The sedimentation rate varies from 0.4 cm/k.y. to 13.5 cm/k.y. This strong variations are related to unconformities. The lowest sedimentation rate occurs in the early late Pliocene (Subzone CN12a; 0.4 cm/k.y.), and the highest sedimentation rate in the early early Pleistocene (Subzone CN13b; 13.5 cm/k.y.). The average sedimentation rate from Cores 159-959C-1H through 8H is 1.5 cm/k.y.

INTRODUCTION

The objective of this paper is to describe more accurately the age of Pliocene to Pleistocene sediment based on the calcareous nannofossils in the equatorial Atlantic Ocean. Leg 159 drilled four sites (Sites 959–962) on the Marginal Ridge of the Cote d'Ivoire Transform Margin in the eastern equatorial Atlantic. Four holes were drilled at Site 959: Holes 959A, 959B, 959C, and 959D. Pliocene to Pleistocene sediment from Hole 959C was chosen for the purposes of this study. Hole 959C was drilled at 3°37.669'N, 2°44.116'W, in a water depth of 2090 m (Fig. 1), using advanced hydraulic piston coring (APC) until refusal at 179.6 meters below seafloor (mbsf).

Core recovery from Hole 959C was complete, with a relatively high sediment accumulation rate. Hole 959C also contains abundant well-preserved calcareous nannofossils; therefore, it is an ideal site for the study of high-resolution calcareous nannofossil biostratigraphy.

MATERIALS AND METHODS

Conventionally, the smear-slide method is used by paleontologists for the biostratigraphy and quantitative study of calcareous nannofossils. However, size fractionation on smear slides has been observed as a result of tooth-pick action on a slide (Wei, 1988). Beaufort (1991; p. 415) stressed that “a count of relative abundance using smear slides is accurate only for species of similar size unless a large number of view fields are examined.” To avoid the size fractionation of calcareous nannofossils during the preparation of smear slides, the settling method developed by Beaufort (1991) and Ehrendorfer (1993) is used.

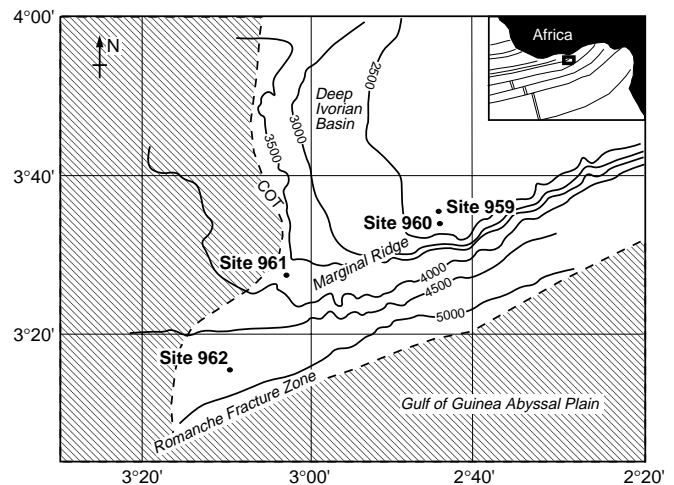


Figure 1. Site 959 location map.

Neogene zonation of Okada and Bukry (1980) was used for the biostratigraphy (Table 1). Biostratigraphy was based on analysis of 371 Neogene (Pliocene to Pleistocene) samples from Cores 1H through 8H. Samples were selected at 20-cm sampling intervals. Cores 1H through 8H are composed of nannofossil ooze with foraminifers (Shipboard Scientific Party, 1996). Only the presence or absence of the biostratigraphic marker species are checked in the 371 samples. Relative abundances of calcareous nannoplankton species per section (1.5-m intervals) among the above mentioned 371 samples are reported based on 500 countings from the settled slide (Table 2). The light microscope with a magnification 1000× was used for this study. Species that do not occur within 500 countings are designated with a “R” in the species percentage chart (Table 2).

The sedimentation rate curve is drawn based on the known age of calcareous nannofossil marker species.

RESULTS AND DISCUSSION

Biostratigraphy

The results of biostratigraphy analysis of each sample are shown in Figure 2 and Tables 2 and 3. Six zones (Zone CN15–CN10) from

¹Masclé, J., Lohmann, G.P., and Moullade, M. (Eds.), 1998. *Proc. ODP, Sci. Results*, 159: College Station, TX (Ocean Drilling Program).

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Pliocene to Pleistocene are identified in Hole 959C (Fig. 2, Table 3). The boundary between Zone CN15 and Subzone CN14b is identified by the interval from the first occurrence (FO) of *E. huxleyi*. In this study, Zone CN15 and Subzone 14b are combined because of the difficulties of identifying *E. huxleyi* under the light microscope. Subzone CN14b (*C. cristatus* Zone) is defined as the interval from the last occurrence (LO) of *P. lacunosa* to the FO of *E. huxleyi*. Samples 159-959C-1H-1, 0.0–2.5 cm, to 2H-2, 80.0–82.5 cm (0.01–4.61 mbsf), belong to Zone CN15 and Subzone CN14b. These zones are dominated by *E. huxleyi*, *Florisphaera profunda*, *G. oceanica*, *G. caribbeanica*, and small *Gephyrocapsa* spp. (Table 2). Samples from this zone contain no broken nannoplanktons, well-preserved bridges of *Gephyrocapsa* species (including *G. aperta*), and *Scapholithus fossilis*, indicating good preservation. Sample 159-959C-1H-1, 120.0–122.5 cm (1.21 mbsf), contains a trace amount of reworked *Discoasters*, which became extinct at the Pliocene/Pleistocene boundary. A trace amount of reworked *Calcidiscus macintyreii* also is observed in Samples 159-959C-2H-1, 0.0–2.5 cm (2.31 mbsf), 2H-1, 120.0–122.5 cm (3.51 mbsf), 2H-2, 0.0–2.5 cm (3.81 mbsf), and 2H-2, 40.0–42.5 cm (4.21 mbsf).

The *Emiliania ovata* Subzone (CN14a) is defined by the interval from the FO of *G. oceanica* to the LO of *P. lacunosa*. The *E. ovata* Subzone includes the interval from Sample 159-959C-2H-2, 100.0–102.5 cm, to 3H-1, 80.0–82.5 cm (4.81–12.61 mbsf). *Gephyrocapsa caribbeanica*, *G. oceanica*, *Gephyrocapsa* spp. (mostly *G. aperta*), and *F. profunda* are abundant in this zone (Table 2). The following five samples contain a trace amount of reworked nannofossils from this zone: Samples 159-959C-2H-2, 100.0–102.5 cm, contains reworked *C. macintyreii* (middle Miocene–lower Pleistocene) and *S. belemnus* (lower Miocene); Sample 159-959C-2H-2, 140.0–142.5 cm, contains *Coronocylus nitescens* (upper Eocene–Miocene); Sample 159-959C-2H-4, 40.0–42.5 cm, contains *D. berggrenii* (late Miocene); Sample 159-959C-2H-6, 80.0–82.5 cm, contains reworked *C. luminus* (Eocene–Oligocene). Sample 159-959C-3H-1, 60.0–62.5 cm, contains reworked *D. quinqueramus* (late Miocene). Many samples contain *S. fossilis*, suggesting good preservation. The LO of *P. lacunosa* shows very scarce distribution in the eastern equatorial Atlantic Ocean (Leg 108), and that makes the boundary between Subzones CN14a and CN14b imprecise (Manivit, 1989). However, this hole contains relatively abundant and well-preserved *P. lacunosa*, and the LO datum of *P. lacunosa* is clearly distinguished.

The interval from Sample 159-959C-3H-1, 100.0–102.5 cm, to 3H-3, 0.0–2.5 cm (12.81–14.81 mbsf; 13 samples), is assigned to Subzones CN13a and CN13b. These 13 samples do not contain any *G. caribbeanica* or *G. oceanica*. *G. aperta* is abundant in the above 13 samples. These 13 samples also do not contain *D. brouweri*. Therefore, these samples are assigned to Subzone CN13a. However, the samples below Sample 159-959C-3H-3, 0.0–2.5 cm (14.81 mbsf), contain *G. caribbeanica*, suggesting Subzone CN13b. Therefore, we assigned these 13 samples to Subzones CN13a–b. Young et al. (1994) mentioned that some ambiguity exists for the CN14a/CN13b boundary. Several samples from this interval contain *Scapholithus fossilis*, suggesting good preservation. Sample 159-959C-3H-2, 80.0–82.5 cm (14.11 mbsf), contains a trace amount of reworked *Fasciculithus tympaniformis* (late Paleocene).

The *G. caribbeanica* Subzone (CN13b) is identified by the FO of *G. caribbeanica* to the FO of *G. oceanica*. Samples between 159-959C-3H-3, 20.0–22.5 cm, and 3H-6, 140.0–142.5 cm (15.01–20.71 mbsf), are assigned to this zone. Well-preserved bridges of *G. caribbeanica* and *S. fossilis* indicate good preservation of calcareous nannofossils from this zone. *Florisphaera profunda* and *Gephyrocapsa* spp. are the most abundant species (Table 2).

The *E. annula* Subzone (CN13a) is identified by the LO of *D. brouweri* to the FO of *G. caribbeanica*. Samples 159-959C-3H-7, 0.0–2.5 cm, to 4H-3, 60.0–62.5 cm (20.81–24.91 mbsf), belong to this zone. Sample 959C-3H-7, 20.0–22.5 cm (21.01 mbsf), contains

Table 1. Standard Neogene zonation of Okada and Bukry (1980).

	NN	CN		
Pleistocene	21	<i>E. huxleyi</i>	15	
	20	<i>G. oceanica</i>	14	b <i>C. cristatus</i> a <i>E. ovata</i>
	19	<i>C. doronicoides</i>	13	b <i>G. caribbeanica</i> a <i>E. annula</i>
Pliocene	18	<i>D. brouweri</i>	d	<i>C. macintyreii</i>
	17		c	<i>D. pentaradiatus</i>
	16		b	<i>D. surculus</i>
			a	<i>D. tamalis</i>
	15	<i>R. pseudoubili</i>	11	b <i>D. asymmetri</i> a <i>S. neobabies</i>
	14	<i>A. tricorniculatus</i>	d	<i>A. delicatus</i>
	13		c	<i>C. rugosus</i>
12	b		<i>C. acutus</i>	
Miocene	11	<i>D. quinqueramus</i>	a	<i>T. rugosus</i>
			b	<i>A. primus</i>
			a	<i>D. berggrenii</i>
10	<i>D. neohamatus</i>	b	<i>D. neorectus</i>	
		a	<i>D. bellus</i>	

Note: * = first occurrence; + = last occurrence.

a trace amount of reworked *Discoaster*. Samples 159-959C-4H-2, 80.0–82.5 cm (23.61 mbsf), and 4H-2, 140.0–142.4 cm (24.21 mbsf), also contain reworked *D. brouweri*. *Florisphaera profunda*, *P. lacunosa*, and *R. minutula* are the most abundant species (Table 2). The relative abundance of *Gephyrocapsa* spp. starts to decrease from this zone (Table 2). Most samples contain dissolution-susceptible species of *S. fossilis*. On the other hand, three samples (Samples 159-959C-4H-2, 100.0–102.5 cm, 4H-2, 120.0–122.5 cm, and 4H-3, 60.0–62.5 cm) contain several nannofossil fragments.

The *C. macintyreii* Subzone (CN12d) is defined by the LO of *D. pentaradiatus* to the LO of *D. brouweri*. Samples 159-959C-4H-3, 80.0–82.5 cm, to 4H-5, 140.0–142.5 cm (25.11–28.71 mbsf), are assigned to this zone. *Florisphaera profunda* and *R. minutula* are the most abundant species (Table 2). Samples 159-959C-4H-3, 80.0–82.5 cm (25.11 mbsf), and 4H-3, 100.0–102.5 cm (25.31 mbsf), contain several nannofossil fragments.

The *D. pentaradiatus* Subzone (CN12c) is defined by the LO of *D. surculus* to the LO of *D. pentaradiatus*. Sample 159-959C-4H-6, 0.0–2.5 cm, to 5H-7, 40.0–42.5 cm (28.81–40.21 mbsf), are assigned to this zone. This zone is dominated by *F. profunda*, *R. minutula*, *R. minuta*, and small specimens of *Reticulofenestra*. Nannofossils from this zone are well preserved.

The *D. surculus* Subzone (CN12b) is defined by the LO of *D. tamalis* to the LO of *D. surculus*. Samples 159-959C-5H-7, 60.0–62.5 cm, to 6H-1, 80.0–82.5 cm (41.31–46.21 mbsf), are assigned to this zone. The central arm of *D. surculus* is well preserved in all samples from this interval. This zone, which has a duration of 0.4 m.y. (Okada and Bukry, 1980) is only 1.8 m thick. *F. profunda*, *R. minuta*, and *R. minutula* are the most abundant species.

The *D. tamalis* Subzone (CN12a) is divided into upper and lower by the LO of *D. challengeri* and *D. variabilis*. Samples 159-959C-6H-1, 100.0–102.5 cm, through 6H-4, 140.0–142.5 cm (41.31–46.21 mbsf), belong to the upper part of Subzone CN12a, based on the absence of *D. challengeri* and *D. variabilis*. However, Samples 159-959C-6H-5, 0.0–2.5 cm, through 6H-6, 100.0–102.5 cm (46.31–48.81 mbsf), belong to the lower part of CN12a, based on the presence of *D. challengeri*, *D. variabilis*, and *R. pseudoubilica*. This zone (both upper and lower CN12a) is dominated by *F. profunda*, *R. minutula*, and *R. minuta*. A trace amount of reworked *S. abies* occurs in Sample 159-959C-6H-6, 80.0–82.5 cm (48.61 mbsf).

The boundary between *D. asymmetricus* Subzone (CN11b) and *S. neobabies* Subzone (CN11a) is defined by the LO of *D. asymmetricus*

Table 2. Relative abundances of calcareous nannofossils with one sample per section, Hole 959C.

Age	Nannofossil zone	Core, section, interval (cm)	Depth (mbsf)	<i>Angulolithina arca</i>	<i>Calcidiscus leptoporus</i>	<i>Calcidiscus macintyreii</i>	<i>Ceratolithus cristatus</i>	<i>Ceratolithus rugosus</i>	<i>Coccolithus pelagicus</i>	<i>Dicryococites productus</i>	<i>Discoaster asymmetricus</i>	<i>Discoaster brouweri</i>	<i>Discoaster challengeri</i>	<i>Discoaster pentaradiatus</i>	<i>Discoaster quadramus</i>	<i>Discoaster surculus</i>	<i>Discoaster tamalis</i>	<i>Discoaster triradiatus</i>	<i>Discoaster variabilis</i>	<i>Emiliania huxleyi</i>	<i>Florisphaera profunda</i>	<i>Gephyrocapsa caribbeanica</i>	<i>Gephyrocapsa oceanica</i>	<i>Gephyrocapsa</i> spp.	
late/middle Pleistocene	<i>E. huxleyi</i> / <i>C. cristatus</i> (CN15/CN14b)	1H-1, 0-2.5	0.01		1.63															20.56	26.04	8.88	14.50	21.50	
		1H-1, 80-82.5	0.81		1.22																15.41	32.57	15.14	10.00	20.95
		2H-1, 80-82.5	3.11		0.35																4.79	6.55	24.25	1.42	61.59
		2H-2, 80-82.5	4.61		0.34																4.11	6.44	31.84	0.18	55.64
		2H-3, 80-82.5	6.11		1.11																	20.38	16.72	0.96	59.71
		2H-4, 80-82.5	7.61		1.06																	33.29	8.09	9.28	45.23
early Pleistocene	<i>E. ovata</i> (CN14a)	2H-5, 80-82.5	9.11		0.31																49.14	3.46	9.05	35.40	
		2H-6, 80-82.5	10.61		1.31																27.95	4.80	5.24	55.31	
		3H-1, 80-82.5	12.61		2.45					0.87												35.78		0.13	43.50
		3H-2, 80-82.5	14.11		1.51	0.17				3.73												16.08			74.54
		3H-3, 80-82.5	15.61		1.50					0.17												16.53	0.17		79.80
		3H-4, 80-82.5	17.11		1.52					0.33												23.78	3.20		58.08
	<i>G. caribbeanica</i> (CN13b)	3H-5, 80-82.5	18.61		0.85	R	0.14			1.07	7.62											29.12	11.36		47.02
		3H-6, 80-82.5	20.11		1.33	0.48				0.28	6.68											39.35	5.81		45.04
		4H-1, 80-82.5	22.11		2.00	1.00				1.00												55.40			10.80
		4H-2, 80-82.5	23.61		2.62	0.20	0.20			1.01												56.25			4.84
		4H-3, 80-82.5	25.11		0.80	0.20				0.40			0.60									44.40			8.40
		4H-4, 80-82.5	26.61		0.80	0.20	0.20			0.20		1.20	1.20									66.00			3.20
late Pliocene	<i>C. macintyreii</i> (CN12d)	4H-5, 80-82.5	28.11		1.80	0.20															42.00			2.00	
		4H-6, 80-82.5	29.61		0.80	0.20	0.20		0.40												47.60			1.60	
		5H-1, 80-82.5	31.61		0.80							0.60			0.20							58.50			2.60
	<i>D. pentaradiatus</i> (CN12c)	5H-2, 80-82.5	33.11		1.00	0.80									0.20							55.40			3.60
		5H-3, 80-82.5	34.61		0.20	0.40									0.20							58.40			1.00
		5H-4, 80-82.5	36.11		0.60							0.60			0.20							70.34			0.20
		5H-5, 80-82.5	37.61		0.40	0.20						0.60			1.20							55.60			0.20
		5H-6, 80-82.5	39.11		0.80	0.40	R				0.20		0.60		0.60							62.40			0.20
		6H-1, 80-82.5	41.11		1.17	1.17				0.20		0.20	0.20		2.35		R					58.71			1.17
	<i>D. surculus</i> (CN12b)	6H-2, 80-82.5	42.61		0.60	0.20					0.20	R			0.60		R					75.90			1.99
		6H-3, 80-82.5	44.11		0.99	0.60				0.20		0.40			2.39		R	R				40.76			14.71
		6H-4, 80-82.5	45.61		0.80							0.60					0.2					27.15			0.60
		6H-5, 80-82.5	47.11		0.40	0.20					0.79			R			0.2			R		38.98			1.57
		6H-6, 80-82.5	48.61		1.80	0.40							R	R	0.60		R					42.40			0.80
		7H-1, 80-82.5	50.61		1.00	0.20							0.20		0.80		R					35.00			4.00
	early Pliocene	<i>D. asymmetricus</i> (upper CN11b)	7H-2, 80-82.5	52.11		1.40	0.20					R	1.00		2.99		R					57.49			7.58
			7H-3, 80-82.5	53.61		1.20	R					0.20	0.80		0.80				R			45.40			6.20
			7H-4, 80-82.5	55.11		R	0.20						R	R		0.40						55.22			12.65
7H-5, 80-82.5			56.61		1.00	1.00						R	R		1.20						52.40			2.00	
7H-6, 80-82.5			58.11		1.20	0.80		0.20			0.20	0.60			1.00							49.60			1.40
8H-1, 80-82.5			60.11		1.40	0.40						R			0.80							50.10			0.60
<i>D. asymmetricus</i> (lower CN11)		8H-2, 80-82.5	61.61		1.40	0.20			0.20			0.20			1.40		R					39.00			0.60
		8H-3, 80-82.5	63.11		1.01	0.61						R			2.42		0.4				R	40.40			0.40
		8H-4, 80-82.5	64.61	0.20	1.43	1.22		R				1.43	0.81		2.44		R					32.79			0.41
		8H-5, 80-82.5	66.11		1.20	0.40		R			R	0.80	0.40		2.20		R					33.00			1.40
		8H-6, 80-82.5	67.61		1.19	0.99						0.20	0.40		3.58		R					23.46			0.80

Table 2 (continued).

Age	Nannofossil zone	Core, section, interval (cm)	Depth (mbsf)	<i>Helicosphaera inversa</i>	<i>Helicosphaera kampfneri</i>	<i>Helicosphaera sellii</i>	<i>Helicosphaera wallichii</i>	<i>Pontosphaera japonica</i>	<i>Pseudoemiliania lacunosa</i>	<i>Pyrocyclus ranghness</i>	<i>Reticulofenestra pseudoubilica</i>	<i>Reticulofenestra minutula</i>	<i>Reticulofenestra minuta</i>	<i>Reticulofenestra small</i>	<i>Rhabdosphaera clavivigera</i>	<i>Rhabdosphaera procerata</i>	<i>Scapholithus fossitis</i>	<i>Sphenolithus abies</i>	<i>Sphenolithus neobabies</i>	<i>Syracosphaera histrica</i>	<i>Syracosphaera pulchra</i>	<i>Umbellosphaera irregularis</i>	<i>Umbellosphaera sibogae</i>			
late/middle Pleistocene	<i>E. huxleyi/ C. cristatus</i> (CN15/CN14b)	1H-1, 0-2.5	0.01	0.89	5.18												R						0.89			
		1H-1, 80-82.5	0.81	0.41	2.84										0.54			0.14						0.14		
		2H-1, 80-82.5	3.11	0.18	0.88																	0.14		0.14		
		2H-2, 80-82.5	4.61	0.18	0.72																			0.14		
early Pleistocene	<i>E. ovata</i> (CN14a)	2H-3, 80-82.5	6.11	R	0.32				0.80									R					0.54			
		2H-4, 80-82.5	7.61	0.53	1.59	R	0.66																	0.54		
		2H-5, 80-82.5	9.11		1.42	0.10	0.31																		0.13	
		2H-6, 80-82.5	10.61	0.87	0.44	R	2.18																		0.10	
		3H-1, 80-82.5	12.61	1.16	1.42	0.26	10.30	0.13	0.13																0.87	
		3H-2, 80-82.5	14.11	0.43	0.43	R	4.69	0.17																	1.03	
	<i>G. caribbeanica</i> (CN13b)	3H-3, 80-82.5	15.61	0.17	1.00	R	R																		1.68	
		3H-4, 80-82.5	17.11	0.61	1.83	0.15	0.76						0.30												0.61	
		3H-5, 80-82.5	18.61	R	0.28	0.43	1.99						0.57												1.28	
		3H-6, 80-82.5	20.11	0.48	0.61	0.73	2.54	0.24					1.21												0.85	
		4H-1, 80-82.5	22.11	2.80		0.40	11.40						8.60												1.80	
late Pliocene	<i>E. annula</i> (CN13a)	4H-2, 80-82.5	23.61	2.82	0.60	0.20	3.02					14.31												3.43		
		4H-3, 80-82.5	25.11	0.60	2.00	1.40	0.80						10.20												1.40	
	<i>C. macintyreii</i> (CN12d)	4H-4, 80-82.5	26.61		3.00	0.20	0.60	0.20					1.80												4.20	
		4H-5, 80-82.5	28.11	1.40	1.40	0.20	1.00	0.40					45.40												2.00	
		4H-6, 80-82.5	29.61	0.20	0.40	0.40	0.40						4.60												0.20	
		5H-1, 80-82.5	31.61	0.40	2.60	0.40	0.60						0.80												0.40	
		5H-2, 80-82.5	33.11	0.20	2.40	0.20							1.60												0.20	
		5H-3, 80-82.5	34.61	0.60	1.60								1.40												0.20	
	<i>D. pentaradiatus</i> (CN12c)	5H-4, 80-82.5	36.11	0.40	1.60				0.20				R												0.20	
		5H-5, 80-82.5	37.61	0.40	0.80	0.20	0.20						1.80												0.40	
		5H-6, 80-82.5	39.11	0.60	2.00	0.60	0.40						2.20												0.80	
		6H-1, 80-82.5	41.11	1.37	0.20		R						1.96												0.39	
		6H-2, 80-82.5	42.61		2.19	0.20	0.20						4.58												0.60	
		6H-3, 80-82.5	44.11	0.40	1.19	0.20	0.40						6.16												0.80	
	<i>D. surculus</i> (CN12b)	6H-4, 80-82.5	45.61		0.80								5.59												0.20	
		6H-5, 80-82.5	47.11		1.18		0.40						2.36												1.38	
		6H-6, 80-82.5	48.61		3.20	0.20							8.00												0.20	
		7H-1, 80-82.5	50.61		1.40		0.80						12.20												1.20	
7H-2, 80-82.5		52.11		1.60		0.40						4.99												1.60		
7H-3, 80-82.5		53.61	2.80									7.00												1.60		
early Pliocene	<i>D. asymmetricus</i> (upper CN11b)	7H-4, 80-82.5	55.11		2.00			0.20				3.21	0.60	7.23	9.64									1.00		
		7H-5, 80-82.5	56.61		1.60		0.40					7.40	0.60	13.80	7.60									1.20		
		7H-6, 80-82.5	58.11		3.40	R	0.20						5.20	R	4.00	15.40								0.80		
		8H-1, 80-82.5	60.11		4.21		R						R	7.21	14.83									0.60		
		8H-2, 80-82.5	61.61		3.00								0.20	8.00	5.40									0.60		
		8H-3, 80-82.5	63.11	2.02									0.81	6.46	14.55										0.81	
	<i>D. asymmetricus</i> (lower CN11)	8H-4, 80-82.5	64.61		5.30				1.22				1.63	16.30	6.52										0.81	
		8H-5, 80-82.5	66.11		3.00		0.20						0.40	20.40	10.00										R	
		8H-6, 80-82.5	67.61		5.96		R						1.59	22.27	8.55										0.20	

Note: R = rare species that occur in the sample but were not seen in a count of 500 specimens.

Acme. A quantitative count of *D. asymmetricus* was not performed in this study. Therefore, the boundary between Subzones CN11a and CN11b is not determined in this report. However, in this study, Zone CN11 is divided into upper CN11b by the FO of *P. lacunosa* and the absence of *R. pseudumbilicata* and lower CN11 (meaning, the lower part of CN11b + CN11a), based on the absence of *P. lacunosa*, *A. primus*, and *A. tricorniculatus* (Table 1). Samples 159-959C-6H-6, 120.0–122.5 cm, through 8H-1, 100.0–102.5 cm (49.01–60.31 mbsf), are assigned to the upper part of Subzone CN11b. Most samples from this interval contain typical rounded *P. lacunosa*. However, Samples 159-959C-8H-1, 40.0–42.5 cm, to 8H-1, 100.0–102.5 cm (59.71–60.31 mbsf), contain typically rounded and elliptical *P. lacunosa* occurring together. This zone is dominated by *F. profunda*, *R. minutula*, and *R. minuta*. Sample 159-959C-7H-1, 20.0–22.5 cm (50.01 mbsf), contains seven-armed *D. brouweri*. All samples show good preservation of calcareous nannofossils.

Samples 159-959C-8H-1, 120.0–122.5 cm, to 8H-7, 20.0–22.5 cm (60.51–68.51 mbsf), are assigned to the lower part of Zone CN11. *Florisphaera profunda*, *R. minutula*, *R. minuta*, and small *Reticulofenestra* are the most abundant species. All samples show good preservation.

A. delicatus Subzone (CN10d) is defined by the FO of *D. asymmetricus* to the LO of *A. primus* and *A. tricorniculatus*. Two samples (159-959C-8H-7, 40.0–42.5 cm, and 8H-7, 60.0–62.5 cm; 68.71–68.8 mbsf) contain extremely rare specimens of *A. primus* and *A. tricorniculatus* and are assigned to Subzone CN10d.

Sedimentation Rate

A sedimentation rate curve is drawn based on the first appearance datum (FAD) and the last appearance datum (LAD) of marker species (Table 4). The age assignment of the marker species is based on

Berggren et al. (1985, 1995). The sedimentation rate varies in each nannofossil datum (0.4–13.5 cm/k.y.; see Fig. 3). Subzone CN13b (lower lower Pleistocene; 15.01–20.71 mbsf), which has a duration of 0.7 m.y. (Okada and Bukry, 1980), shows the highest sedimentation rate (13.5 cm/k.y.). The lower part of Subzone CN12a; (lower upper Pliocene; 46.31–48.81 mbsf) exhibits the lowest sedimentation rate (0.4 cm/k.y.). Overall, the Neogene sediments (interval from Core 159-959C-1H through 8H) show a very low sedimentation rate (1.5 cm/k.y.).

SUMMARY AND CONCLUSIONS

Six calcareous nannofossil zones (Zones CN10–CN15) were identified in Cores 159-959C-1H through 8H. Each zone contains well-preserved calcareous nannofossils, and marker species are relatively abundant. *Florisphaera profunda* is the most abundant species throughout all the samples investigated. The sedimentation rate varies from 0.4 cm/k.y. to 13.5 cm/k.y. This strong variation may indicate the existence of a disconformity. The average sedimentation rate for the late Neogene (Pliocene to Pleistocene) is 1.5 cm/k.y.

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REFERENCES

Beaufort, L., 1991. Adaptation of the random settling method for quantitative studies of calcareous nannofossils. *Micropaleontology*, 34:415-418.
 Berggren, W.A., Hilgen, F.J., Langereis, C.G., Kent, D.V., Obradovich, J.D., Raffi, I., Raymo, M.E., and Shackleton, N.J., 1995. Late Neogene chronology: new perspectives in high-resolution stratigraphy. *Geol. Soc. Am. Bull.*, 107:1272–1287.
 Berggren, W.A., Kent, D.V., and Van Couvering, J.A., 1985. The Neogene, Part 2. Neogene geochronology and chronostratigraphy. In Snelling, N.J. (Ed.), *The Chronology of the Geological Record*. Geol. Soc. London Mem., 10:211–260.
 Ehrendorfer, T.W., 1993. Late Cretaceous (Maestrichtian) calcareous nannoplankton biogeography with emphasis on events immediately preceding the Cretaceous/Paleocene boundary [Ph.D. dissert.]. Woods Hole Oceanographic Inst., Massachusetts Inst. of Technology.
 Manivit, H., 1989. Calcareous nannofossil biostratigraphy of Leg 108 sediments. In Ruddiman, W., Sarnthein, M., et al., *Proc. ODP, Sci. Results*, 108: College Station, TX (Ocean Drilling Program), 35–69.
 Okada, H., and Bukry, D., 1980. Supplementary modification and introduction of code numbers to the low-latitude coccolith biostratigraphic zonation (Bukry, 1973; 1975). *Mar. Micropaleontol.*, 5:321–325.
 Shipboard Scientific Party, 1996. Site 959. In Mascle, J., Lohmann, G.P., Clift, P.D., et al., *Proc. ODP, Init. Repts.*, 159: College Station, TX (Ocean Drilling Program), 65–150.
 Wei, W., 1988. A new technique for preparing quantitative nannofossil slides. *J. Paleontol.*, 62:472–473.
 Young, J.R., Flores, J.-A., and Wei, W., 1994. A summary chart of Neogene nannofossil magnetobiostratigraphy. *J. Nannoplankton Res.*, 16:21–27.

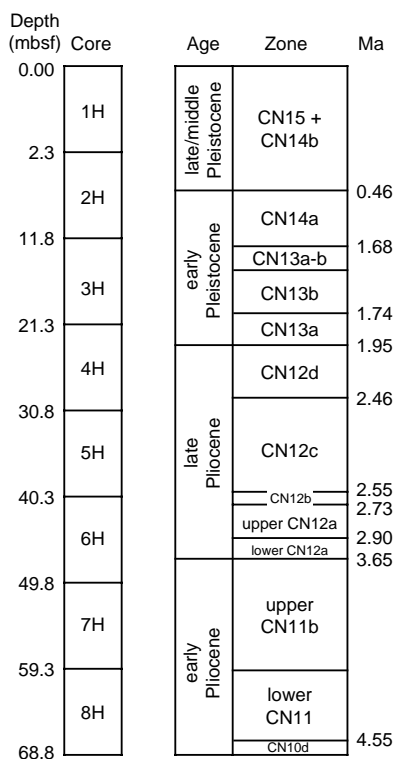


Figure 2. Biostratigraphic zonation of Hole 959C Cores 159-959C-1H through 8H. The age assignments are based on Berggren et al. (1985, 1995).

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APPENDIX

Calcareous Nannofossil Species Considered in this Report
(Listed Alphabetically by Genus Epithets)

- Angulolithina arca* Bukry, 1973
- Calcidiscus leptoporus* (Murray and Blackman, 1898) Loeblich and Tappan, 1978
- Calcidiscus macintyreii* (Bukry and Bramlette, 1969) Loeblich and Tappan, 1978
- Ceratolithus cristatus* Kamptner, 1950
- Ceratolithus rugosus* Bukry and Bramlette, 1968
- Coccolithus pelagicus* (Wallich) Schiller, 1930
- Dictyococcites productus* (Kamptner, 1963) Backman, 1980
- Discoaster asymmetricus* Gartner, 1969
- Discoaster brouweri* (Tan) emend. Bramlette and Riedel, 1963
- Discoaster challengerii* Bramlette and Riedel, 1954
- Discoaster pentaradiatus* (Tan) emend. Bramlette and Riedel, 1954
- Discoaster quadramus* Bukry, 1973
- Discoaster surculus* Martini and Bramlette, 1963
- Discoaster tamalis* Kamptner, 1967
- Discoaster triradiatus* Tan, 1927
- Discoaster variabilis* Martini and Bramlette, 1963

- Emiliania huxleyi* (Lohmann, 1902) Hay and Mohler, 1967
- Florisphaera profunda* Okada and Honjo, 1973
- Gephyrocapsa caribbeanica* Boudreaux and Hay, 1969
- Gephyrocapsa oceanica* Kamptner, 1943
- Helicosphaera inversa* Gartner, 1980
- Helicosphaera kamptneri* Bramlette and Martini, 1964
- Helicosphaera sellii* Bukry and Bramlette, 1969
- Helicosphaera wallichii* (Lohmann, 1902) Boudreaux and Hay, 1969
- Pontosphaera japonica* (Takayama) Nishida, 1971
- Pseudoemiliania lacunosa* (Kamptner, 1963) Gartner, 1969
- Pyrocyclus orgensis* (Bukry, 1971) Backman, 1980
- Reticulofenestra pseudoubilica* (Gartner, 1967) Gartner, 1969
- Reticulofenestra minutula* (Gartner, 1967) Haq and Berggren, 1978
- Reticulofenestra minuta* Roth, 1970
- Rhabdosphaera clavigera* Murray and Blackman, 1898
- Rhabdosphaera procera* Martini, 1969
- Scapholithus fossilis* Deflandre in Deflandre and Fert, 1954
- Sphenolithus abies* Deflandre in Deflandre and Fert, 1954
- Sphenolithus neoabies* Bukry and Bramlette, 1969
- Syracosphaera histrica* Kamptner, 1941
- Syracosphaera pulchra* Lohmann, 1902
- Umbellosphaera irregularis* Paasche in Markali & Paasche, 1955
- Umbellosphaera sibogae* (Weber-van Bosse, 1901) Gaarder, 1970

Table 3. Result of calcareous nannofossil zonation of Cores 159-959C-1H through 8H at 20-cm intervals.

Age	Nannofossil	Subzone	Core, section, interval (cm)	Depth (mbsf)
late/middle Pleistocene	<i>E. huxleyi</i> / <i>C. cristatus</i>	CN15/CN14b	1H-1, 0-2.5, to 2H-2, 80.0-82.5	0.01-4.61
	<i>E. ovata</i>	CN14a	2H-2, 100.0-102.5, to 3H-1, 80.0-82.5	4.81-12.61
early Pleistocene	<i>G. caribbeanica</i> / <i>E. annula</i>	CN13a-b	3H-1, 100.0-102.5, to 3H-3, 0.0-2.5	12.81-14.81
	<i>G. caribbeanica</i>	CN13b	3H-3, 20.0-22.5, to 3H-6, 140.0-142.5	15.01-20.71
	<i>E. annula</i>	CN13a	3H-7, 0.0-2.5 to 4H-3, 60.0-62.5	20.81-24.91
	<i>C. macintyreii</i>	CN12d	4H-3, 80.0-82.5, to 4H-5, 140.0-142.5	25.11-28.71
late Pliocene	<i>D. pentaradiatus</i>	CN12c	4H-6, 0.0-2.5, to 5H-7, 40.0-42.5	28.81-40.21
	<i>D. surculus</i>	CN12b	5H-7, 60.0-62.5, to 6H-1, 80.0-82.5	40.31-41.11
	<i>D. tamalis</i>	upper CN12a	6H-1, 100.0-102.5, to 6H-4, 140.0-142.5	41.31-46.21
		lower CN12a	6H-5, 0.0-2.5, to 6H-6, 100.0-102.5	46.31-48.81
early Pliocene	<i>D. asymmetricus</i>	upper CN11b	6H-6, 120.0-122.5, to 8H-1, 100.0-102.5	49.01-60.31
		lower CN11	8H-1, 120.0-122.5, to 8H-7, 20.0-22.5	60.51-68.51
			8H-7, 40.0-42.5, to 8H-7, 60.0-62.5	68.71-68.8
	<i>A. delicatus</i>	CN10d		

Table 4. Calcareous nannofossil biostratigraphic datums used to calculate sediment accumulation rates for Hole 959C.

Datum	Depth (mbsf)	Age (Ma)
LAD <i>P. lacunosa</i>	4.61	0.46
FAD <i>G. oceanica</i>	12.62	1.68
FAD <i>G. caribbeanica</i>	20.71	1.74
LAD <i>D. brouweri</i>	24.91	1.95
LAD <i>D. pentaradiatus</i>	28.71	2.46
LAD <i>D. surculus</i>	40.21	2.55
LAD <i>D. tamalis</i>	41.11	2.73
LAD <i>D. variabilis</i>	46.31	2.9
LAD <i>R. pseudoubilicata</i>	49.01	3.65
LAD <i>A. primus</i>	68.71	4.55

Notes: The age assignments of marker species are based on Berggren et al. (1985, 1995). FAD = first appearance datum; LAD = last appearance datum.

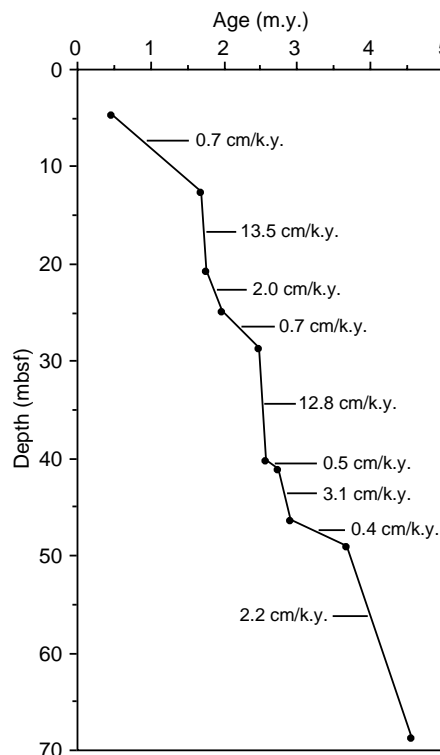


Figure 3. Late Neogene (Pliocene–Pleistocene) sedimentation rate curve for Hole 959C.