# Mutagenicity induced in *Salmonella* strains TA98 and TA100 by diphenylthiophenes

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#### ABSTRACT

Mutagenic properties of four different diphenylthiophenes: 3,4diphenylthiophene, 3,4-di(4'-methylphenyl)thiophene, 3,4-di(4'methoxyphenyl)thiophene and 3,4-di(4'-pentoxyphenyl)thiophene were investigated applying the *Salmonella* test. The research was done on two strains of *Salmonella* Typhimurium: TA98 and TA100,

#### INTRODUCTION

Sulfur heterocycles found in the environment often originate from fossil fuels (Bessler et al. 1997). Some secondary metabolites of microorganisms and plants also constitute another source of sulfur heterocycles, with thiophenes being the most common (Speight 1980).

Developments in organic electronics have caused growing interest in new molecular components for the construction of organic field-effect transistors (OFETs), organic light-emitting diodes (OLEDs) and photovoltaic cells (Perepichka and Perepichka 2009; Skotheim and Reynolds 2007). In this context, materials derived from linear organic  $\pi$ -conjugated monomers, e.g. diphenylthiopenes, are particularly appealing and commonly described (Helgesen and Krebs 2010; Henckens et al. 2005; Parvez et al. 2009). These materials, when suitably modified, enable low-cost fabrication processes of stable, large-area, and lightweight electronic devices (Odaci et al. 2008). Even though the thiophene monomers and polymers are now commonly applied in mobile phone and computer displays, their toxicity or mutagenicity remain unknown.

tested in two variants: with (+S9) and without (-S9) enzymatic activation. Only one compound 3,4-di(4'-methylphenyl)thiophene showed mutagenic activity when studied with metabolic activation (+S9) and its mutagenic rate (MR) score was 3.41 for the dose of 10.00 $\mu$ g-plate<sup>-1</sup>. Other studied compounds did not show any mutagenic activity (±S9) and their MR score did not exceed the threshold value of 2.0.

Numerous medicines contain thiophene rings in their structure. There are reports that thiophene compounds were activated to electrophilic intermediates by cytochrome P450 (Lin et al. 2011). The resulting sulfoxides can be then covalently modified by glutathione and other enzymes' compounds containing the thiol group. The mechanism for thiophene toxicity is not known, although it has been suggested that thiophene metabolism may lead to the formation of oxidative intermediates that could function as the ultimate toxicants (Dansette et al. 2005; Du et al. 2008).

## MATERIAL AND METHODS

#### Synthesis of diphenylthiophenes

Diphenylthiophenes were synthesized from the corresponding sulfides by McMurry coupling followed by aromatization according to a modified Nakayama protocol (Nakayama et al. 1985). The general experimental procedure is given below (Scheme 1).



A solution of 10.0mMol of sulfide 1(a-d) in dry THF (40ml) was added to 60.0mMol of activated Zn powder and placed in a flask equipped with a thermometer and gas inlet. The mixture was cooled to -20°C with vigorous stirring and 30.0mMol of TiCl<sub>4</sub> was added drop-wise over 3h in an argon atmosphere. The mixture was allowed to warm to 10-15°C and stirred for another 3h. The reaction was then quenched by pouring on crushed ice (10ml), neutralized with a saturated solution of K2CO3 and extracted with 30cm<sup>3</sup> of CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was evaporated and the remaining residue was dissolved in 20ml of benzene and placed in a flask equipped with a Dean-Stark trap. 5.0mMol of p-toluenesulfonic acid was added and the solution was refluxed for 4h. The chilled solution was washed with saturated NaHCO3 solution, dried over MgSO<sub>4</sub> and evaporated. The solvent was removed in vacuo and the residue was purified by column chromatography (silica gel/toluene). The yields of isolated diarylthiophenes 2(a-d) were within the range 81-95%.

The structures of the synthesized and characterized compounds are given in Table 1. All chemicals were specially synthesized for this experiment because they were not available commercially. The purity of samples was confirmed by NMR spectroscopy and was greater than 95%. Gas chromatography (GC-FID) analysis showed no impurities.

## Salmonella mutagenicity assays

Histidine-dependent strains of TA98 and TA100 of *Salmonella* Typhimurium were purchased from TRINOVA Biochem GmbH Germany and were cultured as described by Maron and Ames (1983) and Mortelmans and Zeiger (2000). Confirmation of genotypes of the tester strains were routinely carried out, including crystal violet, UV and ampicillin sensitivities. The number of

Table 1. The structure and characteristics of the examined compounds.

Compound name and formula	Melting point, solvent	Spectral data	Yield
3,4-diphenylthiophene 2a	110°C (ethanol) (lit. m.p. 112-113°C) <sup>1</sup>	<sup>1</sup> HNMR (300 MHz, CDCl <sub>3</sub> ): 7.16-7.26 (m, 10H), 7.30 (s, 2H) <sup>13</sup> CNMR (75 MHz, CDCl <sub>3</sub> ): 124.2, 127.0, 128.3, 129.2, 136.7, 141.9	81%
3,4-di(4'-methylphenyl)thiophene 2b	69°C (ethanol) (lit. m.p. 70-72°C) <sup>2</sup>	$^{1}\mathrm{HNMR}$ (300 MHz, $\mathrm{CDCl}_{3}\mathrm{)}\mathrm{:}$ 2.36 (s, 6H), 7.10-7.15 (m, 8H), 7.29 (s, 2H) $^{13}\mathrm{CNMR}$ (75 MHz, $\mathrm{CDCl}_{3}\mathrm{)}\mathrm{:}$ 21.3, 123.7, 129.0, 133.9, 136.6, 141.8	85%
3,4-di(4'-metoxyphenyl)thiophene 2c $-\circ$	103-105°C (ethanol) (lit. m.p. 107-108°C) <sup>3</sup>	$^1\mathrm{HNMR}$ (300 MHz, $\mathrm{CDCl}_3$ ): 3.78 (s, 6H), 6.78-6.80 (m, 4H), 7.10-7.13 (m, 4H), 7.21 (s, 2H) $^{13}\mathrm{CNMR}$ (75 MHz, $\mathrm{CDCl}_3$ ): 55.3, 113.7, 123.2, 129.4, 130.2, 141.5, 158.7	84%
3,4-di(4'-pentoxyphenyl)thiophene 2d $\begin{bmatrix} 1 & 0 \\ 0 & 0 \end{bmatrix}_{4}^{\circ}$	46-48°C (petroleum ether)	$^{1}\mathrm{HNMR}$ (300 MHz, CDCl_3): 0.91-0.97 (6H, m), 1.35-1.47 (8H, m), 1.74-1.83 (4H, m), 3.94 (t, J=6.6Hz, 4H), 6.77-6.81 (m, 4H), 7.09-7.13 (m, 4H), 7.22 (s, 2H) $^{13}\mathrm{CNMR}$ (75 MHz, CDCl_3): 14.6, 23.1, 28.9, 29.6, 68.6, 114.8, 123.6, 129.7, 130.7, 142.0, 158.8 MS (m/z) 408,72	98%

<sup>1</sup> Grunwell et al. (1977)

<sup>2</sup> Dang and Chen (2007)

<sup>3</sup> Chadwick et al. (1972)

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spontaneous revertants and induced revertants following exposure to such diagnostic mutagens as B[a]P(10µg·plate<sup>-1</sup>) and NQNO (10µg·plate<sup>-1</sup>) were measured in each experiment and compared to control values. The assays were performed with and without an activation system. For activation, we used S9 mix, a "rat liver postmitochondrial supernatant" (TRINOVA Biochem). For this preliminary study, five concentrations (0.10, 1.00, 10.00, 50.00, 200.00µg per plate) in triplicate of each compound were fixed. In these studies, the tested compound was dissolved in 80µl of DMSO and added to 2.5ml molten top agar (at 42°C) with a 18h nutrient broth culture of an appropriate strain of S. Typhimurium and 0.5ml of S9 mix. The final mixture was poured on minimal glucose agar plates. Mutations from histidine-dependent to histidine-independent bacteria were assessed for 48h after plating by counting the colonies of bacteria on the Petri dishes. The compounds were assayed in triplicate at each dose level.

#### Statistical methods

The results are reported as mean numbers of revertant colonies per plate  $\pm$  the standard deviation for the test chemicals and the controls. The non-parametric method (Mann-Whitney) was applied for detecting the differences.

In order to compare the mutagenicity of selected compounds, their mutagenic rate (MR) were calculated. MR (as a relative measure of the mutagenic effect) is the ratio of the number of net revertants in a given dose and the average number of revertants in the negative control. A mutagenic effect occurs with a value of MR $\geq$ 2.

#### RESULTS

The compounds showed mutagenic activity only in strain TA100 with metabolic activation. 3,4-di(4'-methylphenyl)thiophene 2b induced the highest number of TA100 revertants, for the dose of  $10.00\mu$ g·plate<sup>-1</sup> and caused a high mutagenic rate (MR=3.41). For the dose  $50\mu$ g·plate<sup>-1</sup>, a significant decrease in the number of revertants was observed. For the next two examined compounds (3,4di(4'-metoxyphenyl)) -thiophene 2c and 3,4-di (4'-pentoxyphenyl)thiophene 2d), a weak toxic effect for the lowest concentration was observed. A small toxic effect was observed in both variants (with and without metabolic activation) for 3,4-di(4'-pentoxyphenyl)-thiophene 2d. The combined data for all four compounds for the strain TA100 are shown in Figure 1.

Diphenylthiophens in both variants showed no mutagenic effect in strain TA98. The number of revertants in strain TA98 for each tested dose is presented in Table 2.



Figure 1. Mean number of revertants induced by four different diphenylthiophenes in *Salmonella* Typhimurium strain TA100.

#### DISCUSSION

In the structure of the examined diphenylthiophenes, different aromatic functionalities or substitutes are attached to the thiophene ring. Our study was intended to determine the relationship between simple modification of the diphenylthiophene structure and mutagenicity or toxicity of the compound. The metabolism of a symmetricallysubstituted thiophene molecule may include oxidation of sulfoxides. In contrast to benzothiophene S-oxides, thiophene S-oxides are a very reactive species. Although few of them have been described to date (Dansette et al. 2005), recent results show that oxidation of the thiophene ring leads to adducts. These adducts are obtained in a Michael-type addition of a nucleophilic reagent containing a thiol group to a thiophene-oxide intermediate (Dreiem and Fonnum 2004).

Among the four examined diphenylthiophenes, only 3,4-di(4'-methylphenyl)thiophene induced mutations in the

Chemical	Dose (µg·plate·1)	Mean number of revertants	
		Strain TA98-S9	Strain TA98+S9
3,4-diphenylthiophene <b>2a</b>	0.1	32.6	34.6
	1.0	35.6	38.3
	10.0	39.3	35.6
	50.0	32.6	33.3
	200.0	29.6	29.6
3,4-di (4'-methylphenyl) thi ophene ${\bf 2b}$	0.1	31.3	35.6
	1.0	34.6	36.0
	10.0	32.0	31.0
	50.0	34.3	36.0
	200.0	33.0	33.0
3,4-di(4'-metoxyphenyl)thiophene $2c$	0.1	28.6	38.6
	1.0	30.0	35.6
	10.0	31.6	38.6
	50.0	29.6	36.6
	200.0	32.6	31.6
3,4-di (4'-pentoxyphenyl) thiophene ${\bf 2d}$	0.1	36.6	33.6
	1.0	32.0	32.6
	10.0	34.6	31.3
	50.0	32.3	31.0
	200.0	31.6	31.6

Table 1. Mean number of revertants for four tested compounds without (-S9) and with (+S9) activation in *Salmonella* Typhimurium strain TA98.

Salmonella Typhimurium strain TA100. It is possible that this compound may be oxidized by cytochrome P450 into a more mutagenic species. Du et al. (2008) have suggested that sulfur compounds could be susceptible to form nucleophilic attack and form DNA adducts. The strain TA100 is sensitive to base-pair mutations which are provoked by small DNA adducts (Benigni and Bossa 2008).

3,4-di(4'-metoxyphenyl)thiophene and 3,4-di(4'-pentoxyphenyl)thiophene were toxic for the strain TA100 in a variant with S9 activation. It is probable that greater functional groups with different conformations could inhibit the formation of DNA adducts (Boctor et al. 1995). It is also possible that derivatives of these two compounds did not match to the binding side of the enzyme. The active site of cytochrome P450 is non-specific but it is only suitable for uniplanar molecules (Oritz de Montellano 2005).

We have not observed any mutagenic rates for *Salmonella* Typhimurium TA98 because the tested compounds and their derivatives were too small to provoke frameshift mutations (Benigni and Bossa 2008).

According to the results of our study, we conclude that derivatives of diphenylthiophenes cause only base-pairexchange types of mutation and not the frame-shift mutation.

It is difficult to define the metabolic pathways for thiophene derivatives. It is possible that there are different paths correlated with interaction connections (Dansette et al. 2005), molecular shape (Dreiem and Fonnum 2004), branching, symmetry (Du et al. 2008), distribution of charge or quantum-chemical properties of the compound.

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#### REFERENCES

- Benigni, R., C. Bossa. 2008. Structure alerts for carcinogenicity, and the *Salmonella* assay system: A novel insight through the chemical relational databases technology. Mutation Research 695: 248-261.
- Bressler, D.C., J.A. Norman, P.M. Fedorak. 1997. Ring cleavage of sulfur heterocycles: how does it happen? Biodegradation 8: 297–311.
- Boctor, S., M.K. Ross, T. Salib, C.R. Shank. 1995. Modulation of DNA adduct formation by successive exposures of DNA to small and bulky chemical carcinogens. Carcinogenesis 16: 3057-3062.
- Chadwick, D.J., J. Chambers, G.D. Meakins, R.L. Snowden. 1972. Preparation of thiophene esters by the Hinsberg reaction. Journal of the Chemical Society Perkin Transactions 1: 2079-2081.
- Dang, Y., Y. Chen. 2007. Oxidation of 3,4-diaryl-2,5dihydrothiophenes to 3,4-diarylthiophenes using CuBr2: Simple and efficient preparation of 3,4-diarylthiophenes. European Journal of Organic Chemistry 34: 5661-5666.
- Dansette, M.P., G. Bertho, D. Mansuy. 2005. First evidence that cytochrome P450 may catalyze both S-oxidation and epoxidation of thiophene derivatives. Biochemical and Biophysical Research Communications 338: 450-455.

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- Dreiem, A., F. Fonnum. 2004. Thiophene is toxic to cerebellar granule cells in culture after bioactivation by rat liver enzymes. NeuroToxicology 25: 959-966.
- Du, H., J. Wang, J. Watzl, X. Zhang, Z. Hu. 2008. Classification structure-activity relationship (CSAR) studies for prediction of genotoxicity of thiophene derivatives. Toxicology Letters 177: 10-19.
- Grunwell, J.R., D.L. Foerst, M.J. Sanders. 1977. Mechanism of thiophene formation upon photolysis of enethiol esters. Journal of Organic Chemistry 42: 1142-1145.
- Helgesen, M., F.C. Krebs. 2010. Photovoltaic performance of polymers based on dithienylthienopyrazines bearing thermocleavable benzoate esters. Macromolecules 43: 1253–1260.
- Henckens, A., K. Colladet, S. Fourier, T.J. Cleij, L. Lutsen, J. Gelan, D. Vanderzande. 2005. Synthesis of 3,4-diphenyl-substituted poly(thienylene vinylene), low-band-gap polymers via the dithiocarbamate route. Macromolecules 38: 19-26.
- Lin, H.L, H. Zhang, C. Medower, P.F. Hollenberg, W.W. Johnson. 2011. Inactivation of cytochrome P450 (P450) 3A4 but not P450 3A5 by OSI-930, a thiophene-containing anticancer drug. Drug Metabolism and Disposition 39: 345-350.
- Maron, D.M., B.N. Ames. 1983. Revised methods for the Salmonella mutagenicity test. Mutation Research 113: 173-215.

- Mortelmans, K., E. Zeiger. 2000. The Ames Salmonella/microsome mutagenicity assay. Mutation Research 455: 29-60.
- Nakayama, J., H. Machina, M. Hoshino. 1985. General synthesis of 2,5-dihydrothiophenes (3-thiolenes) from diketo sulfides. Tetrahedron Letters 26: 1981-1982.
- Odaci, D., S. Kiralp Kayahan, S. Timur, L. Toppare. 2008. Use of a thiophene-based conducting polymer in microbial biosensing. Electrochimica Acta 53: 4104-4108.
- Ortiz de Montellano, P.R. 2005. Cytochrome P450: Structure, Mechanism and Biochemistry. 689 p. Kluwer Academic/Plenum Publishers, New York.
- Perepichka, F.I, D.F. Perepichka. 2009. Handbook of Thiophene-Based Materiales vol 1: Synthesis and Theory. 865 p. John Wiley & Sons Ltd, Chichester, West Sussex.
- Parvez, M., R.P. Jiminez, T.C. Sutherland, J. Viccars. 2009. Synthesis and optical and electronic properties of thiophene derivatives. European Journal of Organic Chemistry 32: 5635-5646.
- Skotheim, T., J. Reynolds. 2007. Conjugated Polymers Theory, Synthesis, Properties and Characterization. 992p. CRC Press. Boca Raton.
- Speight, J.G. 1980. The Chemistry and Technology of Petroleum. 498p. Marcel Dekker Inc., New York.