


Defective chromatic and achromatic visual pathways in developmental dyslexia: Cues for an integrated intervention programme

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Abstract.

Purpose: As well as obtaining confirmation of the magnocellular system involvement in Developmental dyslexia (DD); the aim was primarily to search for a possible involvement of the parvocellular system; and, furthermore, to complete the assessment of the visual chromatic axis by also analysing the koniocellular system.

Methods: Visual evoked potentials (VEPs) in response to achromatic stimuli with low luminance contrast and low spatial frequency, and isoluminant red/green and blue/yellow stimuli with high spatial frequency were recorded in 10 dyslexic children and 10 age- and sex-matched, healthy subjects.

Results: Dyslexic children showed delayed VEPs to both achromatic stimuli (magnocellular-dorsal stream) and isoluminant red/green and blue/yellow stimuli (parvocellular-ventral and koniocellular streams). To our knowledge, this is the first time that a dysfunction of colour vision has been brought to light in an objective way (i.e., by means of electrophysiological methods) in children with DD.

Conclusion: These results give rise to speculation concerning the need for a putative approach for promoting both learning how to read and/or improving existing reading skills of children with or at risk of DD. The working hypothesis would be to combine two integrated interventions in a single programme aimed at fostering the function of both the magnocellular and the parvocellular streams.

Keywords: Reading disorder, letter recognition, parvocellular, magnocellular, koniocellular, chromatic contrast, luminance contrast, VEPs

1. Introduction

1.1. Human visual system

The human visual system consists of three parallel pathways that originate from different retinal ganglion cells and, after making a relay on specific areas

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of the lateral geniculate nucleus, converge on visual cortical areas V1 and V2: a) the magnocellular pathway, which originates from the *parasol* ganglion cells and projects onto large magnocellular neurons of the dorsal lateral geniculate nucleus; b) the parvocellular pathway, which originates from the *midget* ganglion cells and projects onto small parvocellular neurons of the ventral lateral geniculate nucleus; c) the koniocellular pathway, which originates from the *bistratified* ganglion cells and projects onto the neurons of the intercalated layers of the lateral geniculate nucleus (Chatterjee & Callaway, 2002; Sumner et al., 2008; Ribeiro & Castelo-Branco, 2010).

Beyond the early visual cortical areas, the two main systems form two segregated projective pathways towards extrastriate and associative visual cortical areas: magnocellular inputs are conveyed to V5/MT and the posterior parietal cortex (magnocellular-dorsal stream) (Born & Bradley, 2005), while parvocellular inputs to V4 and the inferotemporal cortex (parvocellular-ventral stream) (Heywood et al., 1992).

The magnocellular system, being composed of large neurons characterised by high excitability and conduction velocity, is a phasic system capable of producing rapid and short responses. It is mainly sensitive to object movements and achromatic stimuli with low spatial frequency, high temporal frequency and low luminance contrast. It is basically concerned with parafoveal and peripheral vision and its main physiological function is to direct attention to the spatial characteristics (localisation) of the object (*where* pathway) (Vidyasagar, 1999; Laycock et al., 2008; Brown, 2009).

The parvocellular system, however, being composed of small neurons characterised by lower excitability and conduction velocity, is a tonic system capable of producing late responses, which are however more sustained over time. It is mainly sensitive to isoluminant red/green chromatic stimuli with high spatial frequency, low temporal frequency and high chromatic contrast. It is basically concerned with foveal or central vision and its main physiological function is to focus attention on the proper characteristics (recognition) of the object (*what* pathway) (Vidyasagar, 1999; Laycock et al., 2008; Brown, 2009).

The koniocellular system is mainly sensitive to isoluminant blue/yellow chromatic stimuli, but also contributes to object movement perception through projections onto V5/MT, which is part of the magnocellular-dorsal stream (Shipp, 2006).

1.2. Developmental dyslexia

Developmental Dyslexia (DD) consists of a deficit in acquiring adequate reading skills and occurs despite the lack of any neurological, cognitive, sensorial and social disability in subjects with normal intelligence and normal educational opportunities (Lyon et al., 2003; Peterson & Pennington, 2012).

At present, the main problem of DD is believed to depend on an impaired processing of auditory and phonological stimuli (phonological awareness theory) (Peterson & Pennington, 2012; Hornickel & Kraus, 2013), however, several theories have been called into play over the years to explain the pathogenesis of DD, some of which still retain their validity (for review see Paulesu et al., 2014).

One of them is the magnocellular theory, which is based on a perceptual defect of the magnocellular system (Lovegrove et al., 1982; Galaburda & Livingstone, 1993). The original assumption of this theory was based on the belief that, in normal reading, the parvocellular system is active during fixations, as opposed to the magnocellular system which is active during saccades. In this way, the magnocellular stream would seemingly exert an inhibitory effect on the parvocellular system only during saccades, thus preventing parvocellular neuronal activity from continuing until the next fixation. In DD, however, given the impaired inhibition of the parvocellular system due to the magnocellular weakness, an overlay of images arising from two subsequent fixations would appear to occur, with the final effect of confusing the reading.

This hypothesis, however, has already been called into question by Burr and colleagues (Burr et al., 1994), who have shown that during saccades the magnocellular system actually inhibits its own previous activity, rather than that of the parvocellular system, thus confuting the intimate mechanism from which the reading impairment was thought to arise. Nevertheless, this finding was not incompatible with the possibility that a deficit of the magnocellular system contributes, to some extent, to the neurobiological substrate of DD. Since the early 90s, in fact, several confirmations of a magnocellular dysfunction have been recorded for children with dyslexia: abnormal responses have been reported for those characteristics of visual stimuli that are specifically targeted by the magnocellular system, namely, achromatic vision, low spatial frequency, low contrast (May et al., 1991; Maddock et al., 1992; Romani et al., 2001; Samar et al., 2002; Vaegan & Hollows, 2006),

high temporal frequency and object movement perception (Kubová et al., 1996; Kuba et al., 2001; Schulte-Körne et al., 2004). These findings fit with anatomico-structural observations that magnocellular neurons of the lateral geniculate nucleus of dyslexic children are both smaller and dystrophic compared with those of age-matched normal readers (Livingstone et al., 1991). Recently, a reduced activation of V5/MT during visual processing of moving objects has been demonstrated in dyslexic children (Oluade et al., 2013). However, no activation deficit has emerged with respect to younger children matched for reading skills (i.e., with the same reading skills), suggesting that the magnocellular dysfunction can be considered an effect rather than the cause of DD. At present, the magnocellular dysfunction is considered to be connected to an impairment of visual attentional shifting, both in its spatial and temporal aspects (Hari & Renvall, 2001). Indeed, this is a crucial skill in the segmentation of letter strings into grapheme constituents (graphemic parsing) (Gori & Facoetti, 2014) and is, therefore, a preliminary condition to the letter-to-speech sound integration.

As far as the parvocellular system is concerned, for many years it has been investigated using visual stimuli with high spatial-frequency (i.e., within the characteristic sensitivity range of the parvocellular system) but achromatic (i.e., stimuli towards which the parvocellular system is substantially blind, whilst the magnocellular system is maximally sensitive) (Victor et al., 1998). In this way, a unique electrophysiological study was able to demonstrate delayed responses in dyslexic children compared to controls (Farrag et al., 2002), although the lack of selectivity of the visual stimuli employed did not exclude the possibility of a contribution from the magnocellular system to the abnormal responses. On the other hand, more recently Ahmadi et al. (2015) have been able to demonstrate an impairment of the parvocellular system in dyslexic children. They did this by presenting coloured visual stimuli (i.e., highly selective for the parvocellular system) in the form of images of natural scenes and by determining the red-green isoluminant point using the psychophysical (subjective) method.

1.3. Aims and scope

In the present study we used an electrophysiological (objective) method. This is represented by visual evoked potentials (VEPs), capable of disclosing even subtle functional impairments of the investigated

systems. This is more sensitive even than the self-awareness that tested subjects may have of their own dysfunctions (for rev. see Tobimatsu & Celesia, 2006).

The primary aims of the present study were: a) to obtain a confirmation of the magnocellular system involvement, by using achromatic stimuli with low luminance contrast and low spatial frequency; b) to search for any potential involvement of the parvocellular system, by using isoluminant red/green stimuli with high spatial frequency, to which this system is specifically sensitive, (i.e. the most suitable stimuli for selectively stimulating this system).

An additional aim was c) to complete the functional assessment of the visual chromatic axis by using isoluminant blue/yellow chromatic stimuli in order to analyse also the koniocellular system.

2. Materials and methods

2.1. Participants

Ten dyslexic children (5 girls; mean age $142,3 \pm 14,3$ months) were selected from a sample of children referred to the Children's Neuropsychiatric Medical Facility of Viareggio (Italy). These children had been diagnosed as dyslexic by an expert paediatric neuropsychiatrist (D.M.S.) on the basis of the Italian National Recommendations (VV.AA.). These require that the child have (a) a full-scale IQ within the normal range (i.e., greater than 85), as measured by the Wechsler Intelligence Scale for Children (3rd edition; WISC-III) (Orsini & Picone, 2006), and (b) a performance two negative SDs below age group norms in one reading task, or one negative SD in at least two reading tasks of the standard Italian test for assessment of reading skills. This test consists of the following three reading tasks: (1) MT battery (Cornoldi & Colpo, 1998); (2) word reading task (Sartori et al., 1995); (3) pseudo-word reading task (Sartori et al., 1995).

The MT battery was used to obtain a measurement of the children's reading speed and accuracy while reading aloud age-standardised Italian prose passages (i.e., ecological-context reading). This was done by computing respectively the mean number of syllables/sec read as well as the number of errors made by the children.

The ability to read aloud was also measured using the word reading task (Sartori et al., 1995) consisting of four standardised clinical lists of 112 Italian words.

Furthermore, the phonological decoding ability was then measured using the pseudo-word task (Sartori et al., 1995) consisting of three standardised clinical lists of 48 Italian pseudo-words (Sartori et al., 1995). Also in these cases, both accuracy and reading speed were scored.

Dyslexic children suffering from attention deficit hyperactivity disorder (ADHD) were excluded from the experiment, to avoid interfering and confounding effects. None of the participants had been treated for any neurological or psychiatric disorder, nor were they under pharmacological treatment at the time of the experimental session.

Ten normal readers, with no reported academic difficulties, matched to the dyslexics in age [mean age $134,3 \pm 26,0$ months, $t(18) = -0.852$, $P = 0.405$] and gender [4 girls; Fisher's exact test, $P = 1.00$], served as the control group.

All children were native Italian speakers and had normal or corrected-to-normal vision. No subjects exhibited any colour deficits, as determined by Ishihara colour plates (Ishihara, 1997). All children's parents gave their informed consent to the study, in accordance with the Declaration of Helsinki.

Table 1 shows the mean and SD of age and text reading tests for the control and dyslexic groups. Controls and dyslexics were comparable in chronological age [$t(18) = -8.852$, n.s.], but were significantly different on accuracy and speed of word, pseudo-word and text reading.

2.2. Visual stimuli

Stimuli were designed to preferentially activate functionally separate pathways in the visual system, traditionally described as magno-cellular (M), parvo-cellular (P) and konio-cellular (K) streams.

Chromatic visual stimuli were equiluminant horizontal sinusoidal gratings, modulated both in luminance (Y-Bk) and chromaticity (R-G and B-Y). Stimuli were obtained by combining red and green

gratings of identical contrast and luminance. Chromatic contrast patterns (red-green or blue-yellow) were obtained by superimposing (out-of-phase by 180 deg) red-black and green-black gratings (or blue-black and yellow-black, respectively) of identical contrast. Luminance contrast patterns (white-black) were obtained by superimposing the same gratings in-phase (Porciatti & Sartucci 1999). Gratings were generated by a VSG/2 graphic card (Cambridge Research[®], UK), displayed full-field on a colour monitor (Samsung Sync Master1100DF[®], 21 inches) at a frame rate of 120 Hz and 14 bits per colour per pixel, suitably linearised by gamma correction (Porciatti & Sartucci, 1996; Porciatti & Sartucci, 1999).

The equiluminant point was measured by assessing contrast sensitivity with the method of ascending limits for a 1 c/deg red-green or black-yellow grating, counterphased at 15 Hz (Fiorentini et al., 1996; Porciatti & Sartucci, 1999). The point of minimum sensitivity was taken as the equiluminant value for the subject. The relative luminance (r) is easily defined by the usual formula $r = Lum_{red} / (Lum_{red} + Lum_{green})$, where values of $r = 0$, $r = 0.5$ (equiluminant point, at maximum chromatic contrast) and $r = 1.0$ respectively define G-Bk, R-G and R-Bk patterns (Mullen, 1985). The extreme values (i.e. $r = 0$ and $r = 1$) characterise gratings with a pure luminance contrast and a poor chromatic contrast.

To minimise the contribution of short-wavelength cones, for red-green stimuli the patterns were viewed through yellow filters (Kodak Wratten 16), thus attenuating wavelengths below 500 nm.

Chromatic contrast stimuli with a transient-onset presentation and a peak spatial frequency of about 2 c/deg (single bar width = 15 arcmin) with a 14×14 deg field size were adopted, as previous studies have shown that larger fields introduce luminance contamination, due both to chromatic aberration and retinal inhomogeneity (Stabell & Stabell, 1980; Porciatti & Sartucci, 1999). Luminance contrast stimuli were employed at two different peak spatial frequencies:

Table 1
Mean (M) and standard deviation (SD) of age and reading abilities in both control and dyslexic groups

	Controls (N = 10)		Dyslexics (N = 10)		Comparison	
	M	SD	M	SD	T(18)	P
Age (months)	134.30	26.02	142.30	14.29	-8.852	0.405
Text reading errors (number)	2.2	1.55	9.8	5.23	-4.407	<0.001
Text reading speed (syll/sec)	4.004	1.044	1.541	0.758	6.037	<0.001
Word reading errors (Z-score)	0.230	0.400	-1.178	0.851	4.735	<0.001
Word reading speed (Z-score)	0.215	0.379	-1.990	0.874	7.318	<0.001
Pseudoword reading errors (Z-score)	0.191	0.317	-1.373	0.917	5.099	<0.001
Pseudoword reading speed (Z-score)	0.249	0.441	-1.927	0.643	8.822	<0.001

2c/deg (single bar width = 15 arcmin, i.e. small bar size) and 0.5c/deg (single bar width = 60 arcmin, i.e. large bar size). Two different contrast levels (K90% and K20%) were used for both luminance contrast and chromatic contrast for red-green VEPs recordings, while only the higher contrast level (K90%) was used for chromatic blue-yellow VEPs recordings.

The visible screen was 26 cm wide and 24 cm high and the viewing distance 100 cm. Mean luminance was kept at 17 cd m⁻² with a retinal luminance of 330 Troland when viewed through natural pupils, measured to be about 5 mm in all subjects.

For any other technical information see previous works on the topic by Porciatti and Sartucci (Porciatti & Sartucci, 1996, 1999).

2.3. Electrophysiological recordings

Transient VEPs were recorded on-line using a BM 623 device (Biomedica Mangoni, Pisa).

The recording AgCl electrode was placed on the Oz position of the 10–20 International EEG System, while the reference electrode was positioned over Cz and the ground was located on the forehead (Harding et al., 1996; Porciatti & Sartucci, 1999; Tobimatsu et al., 2000).

VEPs were recorded in response to abrupt reversal (1 reversal/sec = 1 Hz) of a horizontal square wave grating (see above for spatial frequency and contrast features). As a consequence, the duration of each stimulus as well as the recording time-window was 500 ms.

Subjects maintained stable fixation on a dot (diameter, 0.2°) throughout stimulus presentation. The display was centred on the vertical meridian (central stimulation). In accordance with the international recommendations for visual system testing (Holder et al., 2010), both eyes were stimulated for each participant, one eye at a time (other eye patched) in random order, in order to avoid a binocular summation of the amplitudes of evoked responses.

Fifty stimuli were delivered for each stimulus condition (block) and for each eye. Block sequences occurred in random order. The whole protocol had a duration of about 45' including pauses between blocks. All participants were naïve to VEP recordings and were only admitted into the experimental room immediately before the recording session.

Signals were filtered (0.3 ± 100 Hz, 26 dB/oct) amplified (50 000 fold), digitised (2 kHz, 12 bit resolution) and averaged (at least 50 sums), with a rejection of signals exceeding a threshold voltage.

Partial averages (blocks of 10 sums) of total average were used to evaluate response consistency (Porciatti & Sartucci, 1999; Caleo et al., 2007; Bocci et al., 2014). At least two series of 50 events (total: 100 traces) were averaged with the stimulus contrast reversal.

VEPs were measured in terms of both latency from the stimulus onset (in ms) and amplitude (in μ V). The amplitude of the P1 component (obtained from achromatic stimuli) was measured peak-to-peak (i.e., with respect to the peak of the preceding negative wave), while the amplitude of the N1 component (obtained from chromatic stimuli) was measured baseline-to-peak (i.e., with respect to the first 50 ms of the recording trace taken as baseline) (for ref. see Holder et al., 2010).

For any other technical information see previous works on the topic by Porciatti and Sartucci (Porciatti & Sartucci, 1996, 1999).

2.4. Statistical analysis

Statistical analysis was carried out using the Statistical Package for the Social Sciences (SPSS, Inc., Chicago, IL, USA).

Differences between the two groups regarding demographic items and reading abilities were analysed by means of Student's *t* tests (or Fisher's exact test when comparing gender proportions).

A series of two-way ANOVAs with a 2 × 2 factorial design (two factors and two levels for each factor) was carried out in order to assess the possible presence of main and/or interaction effects on VEPs latency and amplitude. The reading ability factor (with normal and dyslexic reading as levels) was tested from time to time with the following factors: (a) luminance contrast (with low and high contrast as levels), (b) stimulus size (with small and large bar size as levels), for black/white stimuli; (c) chromatic contrast (with low and high contrast as levels), for isoluminant red/green stimuli; (d) chromatic channel (with red/green and blue/yellow opponent channel as levels). The normality of distribution of each variable was tested and transformed data were used when necessary. Between- and within-group multiple comparisons were made by means of the Tukey *post-hoc* test. Data were presented as mean ± SEM; a level of 5% probability ($p < 0.05$) was considered significant.

Effect sizes were also computed in order to provide a measurement of the magnitude of the observed effects and, then, to aid their practical interpretation. Effect size relative to the two-way ANOVA tests

was estimated using the partial Eta-Squared (η^2_p). A commonly used interpretation is to refer to effect size as small ($\eta^2_p = 0.01$), medium ($d = \eta^2_p = 0.06$), and large ($\eta^2_p = 0.14$) based on η^2_p benchmarks suggested by Cohen (1988). When an interaction was present, the value of the interaction coefficient was also reported. Effect size relative to the *post-hoc* (Tukey) tests was estimated by using the appropriate Cohen's index (d). Cohen (1988) has provided d benchmarks to define small ($d = 0.2$), medium ($d = 0.5$), and large ($d = 0.8$) effects. The between-group difference of means was also reported.

Power ($1-\beta$) of all the performed tests with $\alpha = 0.050$ were also calculated and reported. A generally accepted minimum level of power is 0.80 (Cohen, 1988).

3. Results

Results are summarized in Table 2, where data are reported as mean values \pm SD of VEPs latency and amplitude in response to either luminance contrast (P1 component from black/white pattern) or chromatic contrast stimuli (N1 component from red/green and blue/yellow equiluminant patterns) for both normal and dyslexic readers.

As a representative example, Fig. 1 depicts both grand average and individual VEPs traces recorded from either normal or dyslexic readers to red-green equiluminant patterns at high chromatic contrast.

3.1. Luminance contrast

A two-way ANOVA was performed to determine whether the reading ability and the luminance contrast of the black/white stimulus affected P1 latency (Fig. 2A). A main effect of luminance contrast was found, $F(1,65) = 39.232$, $p < 0.001$, $\eta^2_p = 0.38$, $1-\beta = 1$, indicating that the low contrast level yielded

P1 latencies that were delayed with respect to the high one. There was also a main effect on the reading ability, $F(1,65) = 10.688$, $p < 0.01$, $\eta^2_p = 0.14$, $1-\beta = 0.89$, showing that dyslexic readers produced overall later responses than normal readers. Finally, there was an interaction between luminance contrast and reading ability, $F(1,65) = 6.147$, $p < 0.05$, $\eta^2_p = 0.09$, $1-\beta = 0.69$, interaction coefficient = 10.89, so that the contrast effect depended on what kind of reading ability was present. In particular, *post-hoc* tests (Tukey) showed that, at low contrast, dyslexic readers generated significantly later responses than normal readers, $p < 0.001$. Furthermore, a simple effect of the luminance contrast was found at each level of reading ability, being stronger in dyslexic (difference of means = 19.2, $p < 0.001$, $d = 1.27$) than in normal readers (difference of means = 8.32, $p < 0.01$, $d = 0.66$).

The reading ability and the luminance contrast were also tested to determine whether they were able to affect P1 amplitude (Fig. 2B). A two-way ANOVA found only a main effect of luminance contrast, $F(1,65) = 5.951$, $p < 0.05$, $\eta^2_p = 0.08$, $1-\beta = 0.67$, indicating that the low contrast level yielded P1 amplitudes that were smaller with respect to the high one. However, no simple effects were found between treatment groups.

In addition, the reading ability and the size of the black/white stimulus (set at the high contrast level) were tested to determine whether they were able to affect both P1 latency and amplitude, but no effects were found (Fig. 2C,D).

3.2. Chromatic contrast

A two-way ANOVA was performed to determine whether the reading ability and the chromatic contrast of the red/green stimulus affected N1 latency (Fig. 3A).

Table 2

Synoptic view of VEP latencies and amplitudes in both control and dyslexic groups

Stimulus type	Wave	Latency (ms)		Amplitude (μ V)	
		Normal readers	Dyslexic readers	Normal readers	Dyslexic readers
B/W Sz 60' K 20%	P1	118.96 \pm 7.6	131.54 \pm 16.55	10.44 \pm 6.41	10.24 \pm 2.88
B/W Sz 60' K 90%	P1	110.61 \pm 5.72	112.34 \pm 6.64	14.04 \pm 5.74	13.69 \pm 6.1
B/W Sz 15' K 90%	P1	112.43 \pm 7.83	116.37 \pm 8.39	12.75 \pm 5.17	10.57 \pm 5.72
R/G Sz 15' K 20%	N1	182.88 \pm 13.83	199.10 \pm 29.17	6.82 \pm 3.45	7.07 \pm 3.17
R/G Sz 15' K 90%	N1	166.77 \pm 8.11	199.50 \pm 17.08	10.83 \pm 4.96	9.44 \pm 5.15
B/Y Sz 15' K 90%	N1	180.98 \pm 12.53	192.33 \pm 14.97	9.01 \pm 6.25	8.66 \pm 3.78

Data are reported as mean values \pm SD. B/W = Black/White; R/G = Red/Green; B/Y = Blue/Yellow. Sz = stimulus size (in arcminutes); K = stimulus contrast.

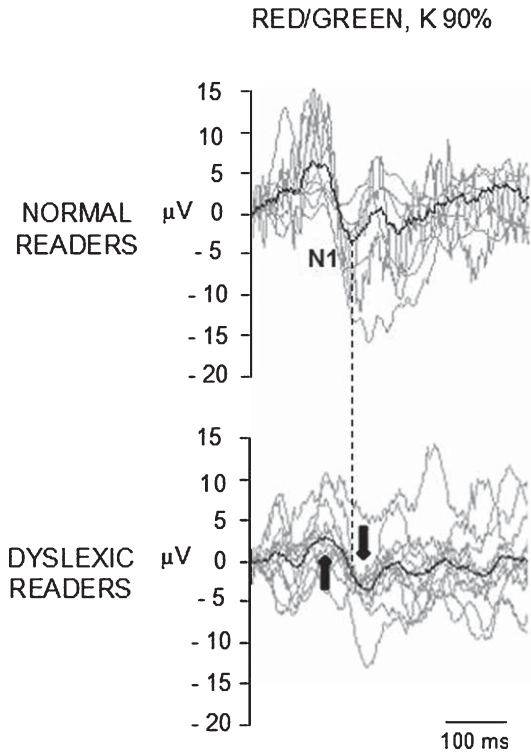


Fig. 1. VEPs recorded in response to red/green high contrast sinusoidal gratings. Note the reduced amplitude (even if not statistically significant, black arrows) and the delayed latency (dotted lines) of dyslexic readers (lower panel) compared to normal readers (upper panel).

488 A main effect of reading ability was found,
 489 $F(1,59) = 30.383, p < 0.001, \eta^2_p = 0.34, 1-\beta = 1$, indi-
 490 cating that dyslexic readers produced overall later
 491 N1 responses than normal readers. A borderline
 492 significant main effect of chromatic contrast was
 493 also found, $F(1,59) = 3.12, p = 0.08, \eta^2_p = 0.05, 1-$
 494 $\beta = 0.41$, indicating that the low contrast level yielded
 495 N1 latencies that were delayed with respect to the
 496 high one. Finally, there was a borderline signifi-
 497 cant interaction between chromatic contrast and
 498 reading ability, $F(1,59) = 3.48, p = 0.065, \eta^2_p = 0.06,$
 499 $1-\beta = 0.45$, interaction coefficient = 16.55, so that the
 500 contrast effect depended on what kind of reading
 501 ability was present. *Post-hoc* tests (Tukey) showed
 502 that dyslexic readers generated significantly later N1
 503 responses than normal readers at both the high con-
 504 trast level (difference of means = 32.729, $p < 0.001,$
 505 $d = 1.37$) and the low contrast level (difference of
 506 means = 16.179, $p < 0.05, d = 0.65$). However, when
 507 switching from low to high chromatic contrast the
 508 dyslexic readers behaviour diverges from that of

509 normal readers, since the N1 latency does not
 510 decrease, but rather remains unchanged (Fig. 3A).
 511 In other words, increasing chromatic contrast in
 512 dyslexic readers does not mean improving the visual
 513 perception of the stimulus.

514 This result assumes even more significance if we
 515 consider that red/green high contrast stimuli yielded
 516 N1 latencies that were greater than 2 SD above the
 517 mean of controls in 16 out of 20 eyes tested (i. e., in
 518 80% of eyes). That is to say that N1 latencies were
 519 abnormal in all 10 subjects examined in at least one
 520 eye (i. e., in 100% of subjects).

521 Furthermore, a simple effect of chromatic con-
 522 trast was found in normal readers (difference of
 523 means = 16,11, $p < 0.05$), but not in dyslexic readers.

524 The reading ability and the chromatic contrast
 525 were also tested to determine whether they were
 526 able to affect N1 amplitude (Fig. 3B). A two-way
 527 ANOVA found only a main effect of chromatic
 528 contrast, $F(1,59) = 7.234, p < 0.01, \eta^2_p = 0.11, 1-$
 529 $\beta = 0.76$, indicating that the low contrast level yielded
 530 N1 amplitudes that were smaller with respect to the
 531 high contrast level.

532 3.3. Chromatic systems

533 A two-way ANOVA was performed to determine
 534 whether the reading ability and the kind of chromatic
 535 stimulus delivered (red/green or blue/yellow), and
 536 thus the kind of chromatic system involved (parvo-
 537 cellular or koniocellular, respectively), affected N1
 538 latency (Fig. 4A).

539 A main effect of reading ability was found,
 540 $F(1,67) = 43.830, p < 0.001, \eta^2_p = 0.39, 1-\beta = 1$, indi-
 541 cating that dyslexic readers produced overall later
 542 N1 responses than normal readers. Moreover, there
 543 was an interaction between the reading ability and
 544 the kind of chromatic stimulus, $F(1,67) = 10.316,$
 545 $p < 0.01, \eta^2_p = 0.14, 1-\beta = 0.89$, interaction coef-
 546 ficient = 21.38, so that the reading ability effect
 547 depended on the kind of chromatic stimulus deliv-
 548 ered. In particular, *post-hoc* tests (Tukey) showed
 549 that dyslexic readers generated significantly later N1
 550 responses than normal readers with both red/green
 551 stimuli (difference of means = 32.729, $p < 0.001,$
 552 $d = 1.73$) and blue/yellow stimuli (difference of
 553 means = 11.346, $p < 0.05, d = 0.57$). The so-called dif-
 554 ference of differences (32.729 - 11.346), which corre-
 555 sponds to the interaction coefficient (21.38), accounts
 556 for the N1 latency antithetical behaviour between
 557 dyslexic and normal readers ($t = 3.212, p < 0.01$)

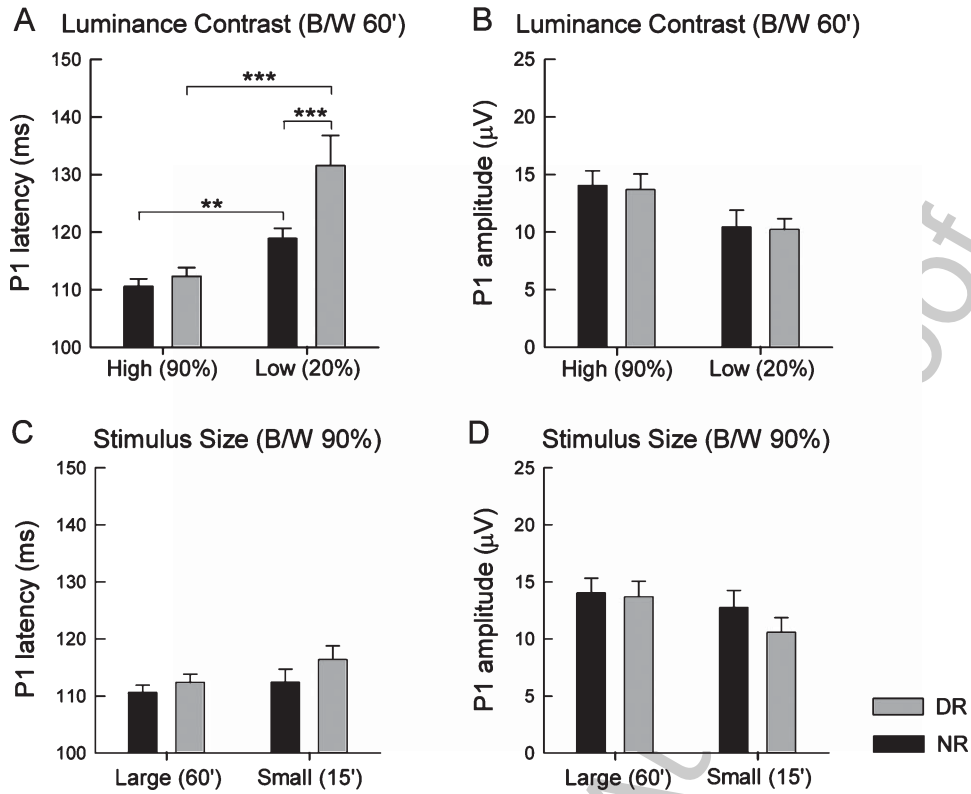


Fig. 2. Effects of luminance contrast and reading ability on P1 latency (panel 1A) and P1 amplitude (panel 1B). Effects of stimulus size (by setting luminance at high contrast) and reading ability on P1 latency (panel 1C) and P1 amplitude (panel 1D). Each bar represents the corresponding mean value \pm SEM. The different fill colours of the bars represent the two levels of reading ability: black indicates Normal Readers (NR) and grey Dyslexic Readers (DR). Data were analysed by two-way ANOVA and Tukey *post-hoc* test as described in Methods. *** <0.001 ; ** <0.01 ; * <0.05 . See Results for Main effects and Interactions.

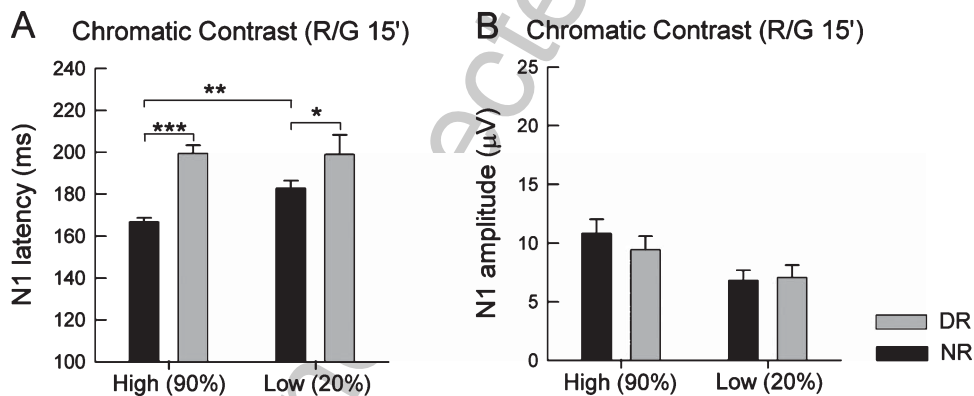


Fig. 3. Effects of chromatic contrast and reading ability on N1 latency (panel A) and N1 amplitude (panel B). Each bar represents the mean value \pm SEM. Fill colours of the bars as in Fig. 2. Data were analysed by two-way ANOVA and Tukey *post-hoc* test as described in Methods. *** <0.001 ; ** <0.01 ; * <0.05 . See Results for Main effects.

558 when switching from blue/yellow to red/green stimu-
 559 li (e.g., in dyslexic readers the N1 latency does not
 560 decrease, as in normal readers, but rather increases)
 561 (Fig. 4A).

562 Furthermore, a simple effect of the kind of chro-
 563 matic stimulus delivered was found in normal readers
 564 (difference of means = 14.208, $p < 0.01$), but not in
 565 dyslexic readers.

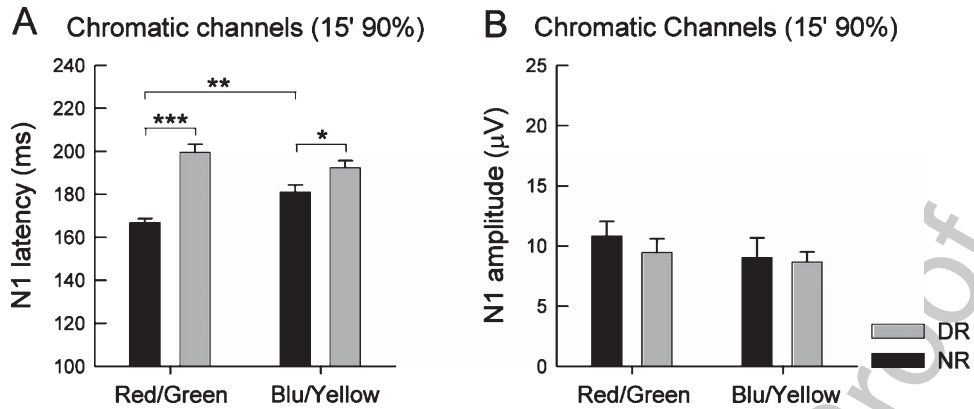


Fig. 4. Effects of chromatic channel and reading ability on N1 latency (panel A) and N1 amplitude (panel B). Each bar represents the mean value \pm SEM. Fill colours of the bars as in Fig. 2. Data were analysed by two-way ANOVA and Tukey *post-hoc* test as described in Methods. *** <0.001 ; ** <0.01 ; * <0.05 . See Results for Main effects and Interactions.

The reading ability and the kind of chromatic stimulus delivered were also tested to determine whether they were able to affect N1 amplitude, but no effects were found (Fig. 4B).

4. Discussion

4.1. General findings

At group level, dyslexic children showed delayed evoked responses to both achromatic stimuli (magnocellular-dorsal stream) and isoluminant red/green chromatic stimuli (parvocellular-ventral stream) compared with age-matched normal readers. Notably, dyslexic children also developed altered responses to isoluminant blue/yellow chromatic stimuli (koniocellular system). To our knowledge, this is the first time that a dysfunction of colour vision has been brought to light in an objective way (i.e., by means of electrophysiological methods) in children with DD.

Our results concerning poor perceptual sensitivity of dyslexic children towards achromatic stimuli with low spatial frequency and low luminance contrast confirm those obtained by others (May et al., 1991; Maddock et al., 1992; Romani et al., 2001; Samar et al., 2002; Vaegan & Hollows, 2006) supporting the hypothesis of a magnocellular-dorsal weakness as an integral part of the neurobiological substrate of developmental dyslexia. This is a hypothesis that still maintains its objective value, even though it has been reformulated with respect to the original magnocellular theory and is now known as “sluggish attentional shifting (SAS) hypothesis” (Hari &

Renvall, 2001). The magnocellular system dysfunction, in fact, is now placed in relation with a deficit of the (visual) attentional shifting. This is crucial, in its spatial and temporal aspects, in the parsing of sub-lexical orthographic units (graphemic parsing) (Gori & Facoetti, 2014). This latter aspect is necessary for both individual letter processing and letter-to-speech sound integration (Vidyasagar & Pammer, 2010).

Our results concerning the poor perceptual sensitivity of dyslexic children to isoluminant red/green chromatic stimuli with high chromatic contrast and high spatial frequency, strengthen and corroborate the hypothesis of an involvement of the parvocellular system firstly proposed by Farrag and colleagues (Farrag et al., 2002). In this work, delayed electrophysiological responses to achromatic stimuli with high spatial frequency were found in dyslexic children with respect to normal readers. Nevertheless, the fact that the stimuli used were achromatic, and as such not entirely selective for the parvocellular system, could leave the doubt that response delays depended to a certain extent on the co-activation of the magnocellular system. More recently, however, the parvocellular hypothesis has received a new impetus since, by employing colour stimuli (thus selective for the parvocellular system), higher red-green isoluminant points were shown in dyslexic children compared to controls (Ahmadi et al., 2015). Our results, obtained by means of an electrophysiological (objective) method, converge with those of Ahmadi and colleagues (Ahmadi et al., 2015), obtained by means of a psychophysiological (subjective) method. It follows that the reliability of both results is mutually reinforced.

632 Finally, the involvement we have found of the
 633 koniocellular system may be related to both of the
 634 following: a low sensitivity to opponent blue/yellow
 635 chromatic stimuli, supporting the notion of a visual
 636 impairment that extends to the entire chromatic axis;
 637 a poor perceptual sensitivity to motion, given the
 638 known contribution of the koniocellular system to the
 639 magnocellular-dorsal stream (Shipp, 2006).

640 4.2. *Colour vision in developmental dyslexia:* 641 *A brief historical review*

642 A growing body of literature exists about colour
 643 vision in developmental dyslexia, although it primar-
 644 ily focuses on several empirical findings regarding
 645 the possibility of obtaining an improvement in read-
 646 ing by means of colour lenses or colour filters (see
 647 Irlen, 1991), while the underlying neurobiological
 648 substrate has not been sufficiently explored. Ray and
 649 colleagues (Ray et al., 2005) showed that wearing yel-
 650 low filters for a period of three months may improve
 651 the reading abilities in dyslexic children. This booster
 652 effect seems to emerge through the removal of the
 653 S-cone inhibitory input on the magnocellular system.
 654 Yellow filters, in fact, seem to be able to cut off short
 655 wavelengths from the visible light spectrum (the so-
 656 called ‘negative blue’) and, in addition, to increase the
 657 phase coherence of the L- (red-) and M- (green-) cone
 658 input, thus normalising the cone contrast weighting.
 659 All this seems to result, on the whole, in an increase
 660 in efficiency of the magnocellular system (Ray et al.,
 661 2005). It should be noted, however, that these mea-
 662 sures, changing the background colour with respect
 663 to the text, do not change the chromatic contrast but
 664 rather the luminance contrast (non-opposing achro-
 665 matic channel) (Kremers & Link, 2008).

666 In addition, a study by Dain and colleagues
 667 (Dain et al., 2008), conducted with psychophysical
 668 (i.e., subjective) methods, revealed lower perceptual
 669 thresholds to yellow/blue stimuli in dyslexic children
 670 compared with controls, suggesting a dysfunction
 671 in colour vision in dyslexic children (dysfunc-
 672 tional hypersensitivity) restricted to the koniocellular
 673 system.

674 Finally, although further evidence of a colour
 675 vision dysfunction has been widely sought in dyslexic
 676 children, even in recent years (Gori et al., 2014),
 677 it has never been found before now, perhaps due
 678 to the inappropriateness of the visual stimuli used.
 679 On the contrary, both in the previously mentioned
 680 work of Ahmadi and colleagues (Ahmadi et al., 2015)
 681 and in the present study, red/green and blue/yellow

isoluminant stimuli, which are highly selective for
 either the parvocellular or the koniocellular path-
 ways, were employed.

682 4.3. *Hypotheses regarding parvocellular system* 683 *involvement*

684 Bearing in mind the specific functions that are
 685 classically attributed to each system, namely, for
 686 the magnocellular-dorsal system, the visual attention
 687 upon object spatial characteristics (visual localiza-
 688 tion) and, for the parvocellular-ventral system, the
 689 visual attention upon object specific features (visual
 690 recognition), a dysfunction of both systems would
 691 well explain impairments of both reading progres-
 692 sion and recognition of letter details that characterise
 693 reading disorders in dyslexics.

694 Moreover, the demonstration of an impairment of
 695 both systems would provide new elements to the
 696 hypothesis of a modulatory influence of the mag-
 697 nocellular system towards the parvocellular system.
 698 In fact, it is known that magnocellular inputs reach
 699 the primary visual cortex earlier than parvocellular
 700 ones (Laycock et al., 2008). This temporal advan-
 701 tage, according to recent studies, would give the
 702 opportunity to the magnocellular system to exert
 703 a top-down facilitatory control on the parvocellu-
 704 lar system through a reentrant loop of projection
 705 (upon the ventral system), via the orbitofrontal cor-
 706 tex and the fusiform gyrus (Kveraga et al., 2007;
 707 Tapia & Breitmeyer, 2011). Furthermore, consid-
 708 ering that the magnocellular system reaches full
 709 development at the age of 2-3 months (Crognale,
 710 2002), while the parvocellular one much later, at the
 711 turn of adolescence (Crognale, 2002; Pompe et al.,
 712 2006), one could assume that in the presence of an
 713 incomplete development of the magnocellular sys-
 714 tem the parvocellular system also suffers a delayed
 715 maturation.

716 4.4. *Possible cues for (re)habilitation*

717 The fact that both systems can contribute to the
 718 biological substrate of DD induces some speculation
 719 concerning activities to be promoted, and/or exer-
 720 cises to be undertaken, in real life environments in
 721 order to prevent and/or improve reading disorders
 722 in these children. A wellness program for reading
 723 abilities designed on the basis of what has emerged
 724 from this study should aim, in our opinion, toward
 725 the following two main objectives: a) to enhance
 726 the magnocellular-dorsal system function and, at the

730 same time, b) to support the parvocellular-ventral
731 system function and its development.

732 4.4.1. *Magnocellular-dorsal system*

733 As far as the magnocellular-dorsal system is con-
734 cerned, its main function is to direct visuospatial
735 attention and, consequently, to control the sequence
736 of saccades for reading progression (Iles et al., 2000;
737 Seassau et al., 2014). It has been shown recently
738 that playing action video games (AVG), which
739 implies dealing with quickly and unpredictably mov-
740 ing objects in the peripheral visual field, immediately
741 improves reading skills of dyslexic children, probably
742 by improving visual navigation skills (Franceschini
743 et al., 2013). As a consequence, (motion) percep-
744 tual learning and AVG has been proposed by Gori
745 and Facoetti for rehabilitation and/or educational pur-
746 poses (Gori e Facoetti, 2014; see also Karimpur &
747 Hamburger, 2015).

748 From the same perspective, in our opinion, propos-
749 ing figure-ground perception games (such as hidden
750 pictures or Where's Waldo) could have the same
751 effect of favouring scanning abilities that are cru-
752 cial for the successful acquisition of reading abilities.
753 Moreover, since the magnocellular-dorsal system is
754 deemed to be a perception-and-action system able to
755 mediate visually guided behaviour (such as reaching,
756 grasping and self-locomotion) (see Ashley, 2004), in
757 our opinion, children with or at risk of DD could also
758 benefit from practicing sports and games that train
759 such skills (for instance the so-called ball sports, such
760 as racquet-and-ball or net goal sports).

761 4.4.2. *Parvocellular-ventral system*

762 As far as the parvocellular-ventral system is con-
763 cerned, its main function is to discriminate and
764 recognise shapes and objects. It has been shown
765 recently that exercise in free-form printing of
766 manuscript letters triggers a writing-reading network
767 that includes both fronto-parietal regions (involved
768 in writing) and the visual word form area (part of
769 the ventral system and involved in reading and letter
770 processing). This would facilitate reading acquisi-
771 tion through an improved effectiveness in recruiting
772 the left fusiform gyrus during reading performance
773 alone (James & Engelhardt, 2012). As a consequence,
774 handwriting exercises have also been proposed by
775 James and Engelhardt (2012) so that children can
776 learn to recognise those attributes of letters (such as
777 shape and orientation) which are relevant for their
778 successful identification and categorisation.

779 Similarly, in our opinion, proposing puzzle games
780 (like Tetris or Tangram) could boost discrimination
781 and recognition abilities of shape, size and orientation
782 of geometric figures, which underlie individual letter
783 identification.

784 5. Concluding remarks

785 In conclusion, the group of dyslexic children
786 showed delayed evoked responses of all the visual
787 pathways examined and, in particular, a complete
788 involvement of the visual chromatic axis. The ampli-
789 tude of the evoked responses, on the contrary, was not
790 significantly reduced compared to normal readers.
791 This dissociation suggests a general slowing of visual
792 processing as a key feature of DD, consistent with
793 a delayed myelination (i.e., dysmaturation) rather
794 than with a reduced number of axons/neurons (Walsh
795 et al., 2005; American Clinical Neurophysiology
796 Society, 2006). This agrees with previous diffusion
797 tensor imaging studies that detected in dyslexic brains
798 abnormalities of fractional anisotropy and radial dif-
799 fusivity consistent with disrupted myelination in
800 the left superior longitudinal (arcuate) fasciculus
801 (Deutsch et al., 2005; Vandermosten et al., 2012).
802 However, the results obtained in the present study
803 suggest that dysmyelination might be a widespread
804 phenomenon in dyslexic brains, extending even out-
805 side the limits of the language network.

806 Our results do not necessarily imply that achro-
807 matic and chromatic visual impairments have to be
808 considered the cause of DD. In fact, they could sim-
809 ply represent an effect of DD or the product of third
810 factors. However, they can certainly be considered as
811 part of the neurobiological substrate of DD.

812 These results have led us to make some considera-
813 tions, no more than speculative at the present stage of
814 our research, concerning a putative wellness program
815 that could aim to promote learning how to read and/or
816 improve existing reading skills of children with, or
817 at risk of DD. The intention is to introduce enhanc-
818 ing interactions and/or additive or combined effects,
819 able to support the development of both visual sys-
820 tems as well as learning to read. This would occur
821 through the induction of synaptogenesis and myeli-
822 nation, in a period of life in which brain plasticity is
823 at its maximum (Kolb & Gibb, 2014).

824 Obviously, the effectiveness of our working
825 hypothesis will necessarily have to be tested by means
826 of specially designed studies and sufficiently large
827 numbers of patients before it can become part of

an accredited wellness program of reading prerequisites (i.e., a program aimed at the development and/or reinforcement of reading prerequisites); a wellness programme that is not to be considered as an alternative to the classic auditory-phonological approach but rather could usefully be associated to it.

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References

- Ahmadi, K., Pouretamad, H.R., Esfandiari, J., Yoonessi, A., & Yoonessi, A. (2015). Psychophysical evidence for impaired magno, parvo, and konio-cellular pathways in dyslexic children. *Journal of Ophthalmic and Vision Research*, *10*(4), 433-440.
- American Clinical Neurophysiology Society (2006). Guideline 9B: Guidelines on visual evoked potentials. *American Journal of Electroneurodiagnostic Technology*, *46*(3), 254-274.
- Ashley, M.J. (2004). Traumatic brain injury: Rehabilitative treatment and case management. Boca Raton, FL: CRC Press.
- Bocci, T., Caleo, M., Tognazzi, S., Francini, N., Briscese, L., Maffei, L., Rossi, S., Priori, A., & Sartucci, F. (2014). Evidence for metaplasticity in the human visual cortex. *Journal of Neural Transmission*, *121*(3), 221-231.
- Born, R.T., & Bradley, D.C. (2005). Structure and function of visual area MT. *Annual Review of Neuroscience*, *28*, 157-189.
- Brown, J.M. (2009). Visual streams and shifting attention. *Progress in Brain Research*, *176*, 47-63.
- Burr, D.C., Morrone, M.C., & Ross, J. (1994). Selective suppression of the magnocellular visual pathway during saccadic eye movements. *Nature*, *371*(6497), 511-513.
- Caleo, M., Restani, L., Gianfranceschi, L., Costantini, L., Rossi, C., Rossetto, O., Montecucco, C., & Maffei, L. (2007). Transient synaptic silencing of developing striate cortex has persistent effects on visual function and plasticity. *The Journal of Neuroscience*, *27*(17), 4530-4540.
- Chatterjee, S., & Callaway, E.M. (2002). S cone contributions to the magnocellular visual pathway in macaque monkey. *Neuron*, *35*(6), 1135-1146.
- Cohen, J. (1988). Statistical power analysis for the behavioral sciences. New York, NY: Routledge Academic.
- Cornoldi, C., & Colpo, M. (1998). Prove di lettura MT per la scuola elementare-2 [Italian MT reading tests for primary school-2]. Firenze, Italy: Organizzazioni speciali.
- Crognale, M.A. (2002). Development, maturation, and aging of chromatic visual pathways: VEP results. *Journal of Vision*, *2*, 438-450.
- Dain, S.J., Floyd, R.A., & Elliot, R.T. (2008). Color and luminance increment thresholds in poor readers. *Visual Neuroscience*, *25*(3), 481-486.
- Deutsch, G.K., Dougherty, R.F., Bammer, R., Siok, W.T., Gabrieli, J.D., & Wandell, B. (2005). Children's reading performance is correlated with white matter structure measured by diffusion tensor imaging. *Cortex*, *41*(3), 354-363.
- Farrag, A.F., Khedr, E.M., & Abel-Naser, W. (2002). Impaired parvocellular pathway in dyslexic children. *European Journal of Neurology*, *9*(4), 359-363.
- Fiorentini, A., Porciatti, V., Morrone, M.C., & Burr, D.C. (1996). Visual ageing: Unspecific decline of the responses to luminance and colour. *Vision Research*, *36*(21), 3557-3566.
- Franceschini, S., Gori, S., Ruffino, M., Viola, S., Molteni, M., & Facoetti, A. (2013). Action video games make dyslexic children read better. *Current Biology*, *23*, 462-466.
- Galaburda, A., & Livingstone, M. (1993). Evidence for a magnocellular defect in developmental dyslexia. *Annals of the New York Academy of Sciences*, *682*, 70-82.
- Gori, S., & Facoetti, A. (2014). Perceptual learning as a possible new approach for remediation and prevention of developmental dyslexia. *Vision Research*, *99*, 78-87.
- Gori, S., Cecchini, P., Bigoni, A., Molteni, M., & Facoetti, A. (2014). Magnocellular-dorsal pathway and sub-lexical route in developmental dyslexia. *Frontiers in Human Neuroscience*, *8*, 460.
- Gori, S., Mascheretti, S., Giora, E., Ronconi, L., Ruffino, M., Quadrelli, E., Facoetti, A., & Marino, C. (2015). The DCDC2 intron 2 deletion impairs illusory motion perception unveiling the selective role of magnocellular-dorsal stream in reading (dis)ability. *Cerebral Cortex*, *25*(6), 1685-1695.
- Harding, G.F., Odom, J.V., Spileers, W., & Spekreijse, H. (1996). Standard for visual evoked potentials 1995. The International Society for Clinical Electrophysiology of Vision. *Vision Research*, *36*(21), 3567-3572.
- Hari, R., & Renvall, H. (2001). Impaired processing of rapid stimulus sequences in dyslexia. *Trends in Cognitive Sciences*, *5*(12), 525-532.
- Heywood, C.A., Gadotti, A., & Cowey, A. (1992). Cortical area V4 and its role in the perception of color. *Journal of Neuroscience*, *12*(10), 4056-4065.
- Holder, G.E., Celesia, G.G., Miyake, Y., Tobimatsu, S., & Weleber, R.G. (2010). International federation of Clinical Neurophysiology: Recommendations for visual system testing. *Clinical Neurophysiology*, *121*(9), 1393-1409.
- Hornickel, J., & Kraus, N. (2013). Unstable representation of sound: A biological marker of dyslexia. *The Journal of Neuroscience*, *33*(8), 3500-3504.
- Iles, J., Walsh, V., & Richardson, A. (2000). Visual search performance in dyslexia. *Dyslexia*, *6*(3), 163-177.
- Irlen, H. (1991). Reading by the colors: Overcoming dyslexia and other reading disabilities through the Irlen method. New York, NY: Avery Publishing Group.
- Ishihara, S. (1997). Tests for colour-deficiency. Tokyo, Japan: Kanehara.
- James, K.H., & Engelhardt, L. (2012). The effects of handwriting experience on functional brain development in pre-literate children. *Trends in Neuroscience and Education*, *1*(1), 32-42.

- 936 Karimpur, H., & Hamburger, K. (2015). The future of action video
937 games in psychological research and application. *Frontiers in*
938 *Psychology*, 6, 1747.
- 939 Kolb, B., & Gibb, R. (2014). Searching for the principles of brain
940 plasticity and behavior. *Cortex*, 58, 251-260.
- 941 Kremers, J., & Link, B. (2008). Electroretinographic responses
942 that may reflect activity of parvo- and magnocellular post-
943 receptor visual pathways. *Journal of Vision*, 8(15), 11-14.
- 944 Kuba, M., Szanyi, J., Gayer, D., Kremláček, J., & Kubová, Z.
945 (2001). Electrophysiological testing of dyslexia. *Acta Medica*
946 *(Hradec Kralove)*, 44(4), 131-134.
- 947 Kubová, Z., Kuba, M., Peregrin, J., & Nováková, V. (1996). Visual
948 evoked potential evidence for magnocellular system deficit
949 in dyslexia. *Physiological Research/Academia Scientiarum*
950 *Bohemoslovaca*, 45(1), 87-89.
- 951 Kveraga, K., Boshyan, J., & Bar, M. (2007). Magnocellular pro-
952 jections as the trigger of top-down facilitation in recognition.
953 *The Journal of Neuroscience*, 27(48), 13232-13240.
- 954 Laycock, R., Crewther, D.P., & Crewther, S.G. (2008). The advan-
955 tage in being magnocellular: A few more remarks on attention
956 and the magnocellular system. *Neuroscience & Biobehavioral*
957 *Reviews*, 32(8), 1409-1415.
- 958 Livingstone, M.S., Rosen, G.D., Drislane, F.W., & Galaburda,
959 A.M. (1991). Physiological and anatomical evidence for
960 a magnocellular defect in developmental dyslexia. *Proceed-*
961 *ings of the National Academy of Sciences of the United States*
962 *of America*, 88(18), 7943-7947.
- 963 Lovegrove, W., Martin, F., Bowling, A., Blackwood, M., Bad-
964 cock, D., & Paxton, S. (1982). Contrast sensitivity functions
965 and specific reading disability. *Neuropsychologia*, 20(3),
966 309-315.
- 967 Lyon, G.R., Shaywitz, S.E., & Shaywitz, B.A. (2003). A definition
968 of dyslexia. *Annals of Dyslexia*, 53(1), 1-14.
- 969 Maddock, H., Richardson, A.J., & Stein, J.F. (1992). Reduced
970 and delayed visual evoked potentials in dyslexics. *Journal*
971 *of Physiology*, 459, 130P.
- 972 May, J.G., Lovegrove, W.J., Martin, F., & Nelson, P. (1991).
973 Pattern-elicited visual evoked potentials in good and poor
974 readers. *Clinical Vision Sciences*, 6, 131-136.
- 975 Mullen, K.T. (1985). The contrast sensitivity of human colour
976 vision to red-green and blue-yellow chromatic gratings. *The*
977 *Journal of Physiology*, 359, 381-400.
- 978 Olulade, O.A., Napoliello, E.M., & Eden, G.F. (2013). Abnormal
979 visual motion processing is not a cause of dyslexia. *Neuron*,
980 79(1), 180-190.
- 981 Orsini, A., & Picone, L. (2006). WISC-III. Contributo alla taratura
982 italiana. [WISC-III. Contribution to the Italian calibration].
983 Firenze, Italy: Giunti O.S. Organizzazioni Speciali.
- 984 Paulesu, E., Danelli, L., & Berlinger, M. (2014). Reading the
985 dyslexic brain: Multiple dysfunctional routes revealed by
986 a new meta-analysis of PET and fMRI activation studies.
987 *Frontiers in Human Neuroscience*, 8, 830.
- 988 Peterson, R.L., & Pennington, B.F. (2012). Seminar: Developmen-
989 tal dyslexia. *Lancet*, 379(9830), 1997-2007.
- 990 Pompe, M.T., Kranjc, B.S., & Breclj, J. (2006). Visual evoked
991 potentials to red-green stimulation in schoolchildren. *Visual*
992 *Neuroscience*, 23(3-4), 447-451.
- Porciatti, V., & Sartucci, F. (1996). Retinal and cortical evoked
993 responses to chromatic contrast stimuli. Specific losses in both
994 eyes of patients with multiple sclerosis and unilateral optic
995 neuritis. *Brain*, 119(Pt 3), 723-740. 996
- Porciatti, V., & Sartucci, F. (1999). Normative data for onset VEPs
997 to red-green and blue-yellow chromatic contrast. *Clinical*
998 *Neurophysiology*, 110(4), 772-781. 999
- Ray, N.J., Fowler, S., & Stein, J.F. (2005). Yellow filters can
1000 improve magnocellular function: Motion sensitivity, conver-
1001 gence, accommodation, and reading. *Annals of the New York*
1002 *Academy of Sciences*, 1039, 283-293. 1003
- Ribeiro, M.J., & Castelo-Branco, M. (2010). Psychophysical chan-
1004 nels and ERP population responses in human visual cortex:
1005 Area summation across chromatic and achromatic pathways.
1006 *Vision Research*, 50(13), 1283-1291. 1007
- Romani, A., Conte, S., Callicco, R., Bergamaschi, R., Versino, M.,
1008 Lanzi, G., Zambrino, C.A., & Cosi, V. (2001). Visual evoked
1009 potential abnormalities in dyslexic children. *Functional Neu-*
1010 *rology*, 16(3), 219-229. 1011
- Samar, V.J., Parasnis, I., & Berent, G.P. (2002). Deaf poor readers'
1012 pattern reversal visual evoked potentials suggest magnocellu-
1013 lar system deficits: Implication for diagnostic neuroimaging
1014 of dyslexia in deaf individuals. *Brain and Language*, 80(1),
1015 21-44. 1016
- Sartori, G., Job, R., & Tressoldi, P.E. (1995). Batteria per la valu-
1017 tazione della dislessia e della disortografia evolutiva [Battery
1018 for the assessment of developmental dyslexia and dysorthog-
1019 raphy]. Firenze, Italy: Giunti O.S. Organizzazioni Speciali. 1020
- Schulte-Körne, G., Bartling, J., Deimel, W., & Remschmidt, H.
1021 (2004). Motion-onset VEPs in dyslexia. Evidence for visual
1022 perceptual deficit. *Neuroreport*, 15(6), 1075-1078. 1023
- Seassau, M., Gárad, C.L., Bui-Quoc, E., & Bucci, M.P. (2014).
1024 Binocular saccade coordination in reading and visual search:
1025 A developmental study in typical reader and dyslexic children.
1026 *Frontiers in Integrative Neuroscience*, 8, 85. 1027
- Shipp, S.D. (2006). Parallel visual pathways. *Advances in Clinical*
1028 *Neuroscience & Rehabilitation*, 6(1), 21-23. 1029
- Stabell, U., & Stabell, B. (1980). Variation in density of mac-
1030 ular pigmentation and in short-wave cone sensitivity with
1031 eccentricity. *Journal of the Optical Society of America*, 70(6),
1032 706-711. 1033
- Sumner, P., Anderson, E.J., Sylvester, R., Haynes, J.D., & Rees,
1034 G. (2008). Combined orientation and colour information
1035 in human V1 for both L-M and S-cone chromatic axes.
1036 *Neuroimage*, 39(2), 814-824. 1037
- Tapia, E., & Breitmeyer, B.G. (2011). Visual consciousness
1038 revisited: Magnocellular and parvocellular contributions to
1039 conscious and nonconscious vision. *Psychological Science*,
1040 22(7), 934-942. 1041
- Tobimatsu, S., & Celesia, G.G. (2006). Studies of human visual
1042 pathophysiology with visual evoked potentials. *Clinical Neu-*
1043 *rophysiology*, 117(7), 1414-1433. 1044
- Tobimatsu, S., Celesia, G.G., Haug, B.A., Onofri, M., Sartucci, F.,
1045 & Porciatti, V. (2000). Recent advances in clinical neurophys-
1046 iology of vision. *Supplements to Clinical Neurophysiology*,
1047 53, 312-322. 1048
- Vaegan, & Hollows, F.C. (2006). Visual-evoked response, pattern
1049 electroretinogram, and psychophysical magnocellular thresh-

- 1050 olds in glaucoma, optic atrophy, and dyslexia. *Optometry & Vision Science*, 83(7), 486-498. 1062
- 1051 1063
- 1052 Vandermosten, M., Boets, B., Poelmans, H., Sunaert, S., Wouters, 1064
- 1053 J., & Ghesquière, P. (2012). A tractography study in dyslexia: 1065
- 1054 Neuroanatomic correlates of orthographic, phonological and 1066
- 1055 speech processing. *Brain*, 135(Pt 3), 935-948. 1067
- 1056 Victor, J.D., Purpura, K.P., & Conte, M.M. (1998). Chromatic 1068
- 1057 and luminance interactions in spatial contrast signals. *Visual Neuroscience*, 15(4), 607-624. 1069
- 1058 1070
- 1059 Vidyasagar, T.R. (1999). A neuronal model of attentional spotlight: 1071
- 1060 Parietal guiding the temporal. *Brain Research Brain Research Reviews*, 30(1), 66-76. 1072
- 1061 1073
- Vidyasagar, T.R., & Pammer, K. (2010). Dyslexia: A deficit 1074
- in visuo-spatial attention, not in phonological processing. *Trends in Cognitive Sciences*, 14(2), 57-63.
- VV.AA. (2009). Disturbi evolutivi specifici di apprendimento. Raccomandazioni per la pratica clinica dei disturbi evolutivi specifici di apprendimento: Dislessia, disortografia, disgrafia e discalculia. Promosso da Associazione Italiana Dislessia. [Learning disabilities: Recommendations for clinical practice on dyslexia, dysorthographia, and dyscalculia]. Trento, Italy: Erickson.
- Walsh, P., Kane, N., & Butler, S. (2005). The clinical role of evoked potentials. *Journal of Neurology, Neurosurgery, and Psychiatry*, 76(Suppl 2), ii16-ii22.

Uncorrected Author Proof