

1 **Periconceptional maternal one-carbon biomarkers are associated with embryonic**
2 **development according to the Carnegie stages**

3 **Running title: One-carbon metabolism and Carnegie stages**

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20 **ABSTRACT**

21 **Study question:** Is periconceptional maternal one-carbon (I-C) metabolism associated
22 with embryonic morphological development in non-malformed ongoing pregnancies?

23 **Summary answer:** Serum vitamin B12, red blood cell (RBC) folate and plasma total
24 homocysteine (tHcy) are associated with embryonic development according to the
25 Carnegie stages.

26 **What is known already:** Derangements in maternal I-C metabolism affect reproductive
27 and pregnancy outcomes, as well as future health of the offspring.

28 **Study design, size, duration:** Between 2010 and 2014, women with singleton ongoing
29 pregnancies were enrolled in a prospective periconceptional cohort study.

30 **Participants/materials, setting, methods:** 234 pregnancies, including 138 spontaneous
31 pregnancies with strict pregnancy dating and 96 pregnancies derived from *in vitro*
32 fertilization (IVF), intracytoplasmic sperm injection (ICSI) or cryo-embryo transfer
33 (IVF/ICSI pregnancies), underwent longitudinal transvaginal three-dimensional
34 ultrasound (3D US) scans from 6⁺⁰ up to 10⁺² weeks of gestation. Carnegie stages were
35 defined using internal and external morphologic criteria in a virtual reality system.
36 Maternal venous blood samples were collected at enrolment for serum vitamin B12, RBC
37 folate and plasma total homocysteine (tHcy) assessment. Associations between
38 biomarker concentrations and longitudinal Carnegie stages were investigated using linear
39 mixed models.

40 **Main results and the role of chance:** We performed a median of three 3D US scans per
41 pregnancy (range 1-5) resulting in 600 good quality datasets for the Carnegie stage
42 annotation (80.5%). Vitamin B12 was positively associated with embryonic development
43 in the total study population ($\beta=0.001$ (95% CI: 0.000; 0.002), $p<0.05$) and in the subgroup

44 of strictly dated spontaneous pregnancies ($\beta=0.002$ (95% CI: 0.001; 0.003), $p<0.05$). Low
45 vitamin B12 concentrations (-2 standard deviation (SD), 73.4 pmol/l) **are associated with**
46 delayed embryonic development by 1.4 days (95% CI: 1.3-1.4) compared to high
47 concentrations (+2SD, 563.1 pmol/l). RBC folate was positively associated with Carnegie
48 stages only in IVF/ICSI pregnancies ($\beta=0.001$ (95% CI: 0.0005; 0.0015), $p<0.05$). Low
49 RBC folate concentrations (-2SD, 875.4 nmol/l) were associated with a 1.8-day delay
50 (95% CI: 1.7-1.8) in development compared to high concentrations (+2SD, 2119.9
51 nmol/l). tHcy was negatively associated with embryonic development in the total study
52 population ($\beta = -0.08$ (95% CI: -0.14; -0.02), $p<0.01$), as well as in the IVF/ICSI subgroup
53 ($\beta= -0.08$ (95% CI: -0.15; -0.01), $p<0.05$). High tHcy concentrations (+2SD, 10.4 $\mu\text{mol/l}$)
54 were associated with a delay of 1.6 days (95% CI: 1.5-1.7) in embryonic development
55 compared to low concentrations (-2 SD, 3.0 $\mu\text{mol/l}$).

56 **Limitations, reasons for caution:** The study was performed in a tertiary care centre,
57 resulting in high rates of folic acid supplement use and comorbidity that may reduce the
58 external validity of our findings.

59 **Wider implications of the findings:** In periconceptional care, maternal I-C biomarkers
60 should be taken into account as predictors of embryonic morphological development.

61 **Combining embryonic size measurements with morphological assessment could better**
62 **define normal embryonic development.**

63 **Study funding/competing interest(s):** The work was funded by the Department of
64 Obstetrics and Gynaecology, Erasmus MC, University Medical Centre, Rotterdam, The
65 Netherlands. RPMST is CSO of the startup company Slimmere Zorg and CEO of eHealth
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67 **Trial registration number:** not applicable.

68 **Key words:** Carnegie stage, maternal one-carbon metabolism, homocysteine, folate,

69 vitamin B12.

INTRODUCTION

70 One-carbon (I-C) metabolism is known to play a crucial role in cellular metabolism and
71 proliferation, as well as in the regulation of gene expression through epigenetic
72 mechanisms. Useful biomarkers of I-C metabolism for research and clinical practice are
73 serum vitamin B12 and folate, red blood cell (RBC) folate and plasma total homocysteine
74 (tHcy). Several studies linked maternal I-C biomarkers to reproductive, pregnancy and
75 health outcomes in the offspring (Stegers-Theunissen et al., 2013; Kalhan, 2016; Bergen
76 et al., 2012; Yajnik et al., 2014; van Uiter and Steegers-Theunissen, 2013a). Most
77 evidence is available on the association between maternal folate deficiency, folic acid
78 supplement use and congenital anomalies (Stegers-Theunissen et al., 2013).
79 Nevertheless, plasma tHcy concentration seems to be a more sensitive marker, with
80 increased concentrations strongly associated with miscarriage, hypertensive disorders,
81 preterm birth and birth defects (Ronnenberg et al., 2007; Steegers-Theunissen et al.,
82 1991; Hogeveen et al., 2012; Vollset et al., 2000). Due to the increased adherence to a
83 vegetarian diet and frequent association with vitamin deficiency, recent research also
84 focused on the associations between vitamin B12, birth defects and birth weight
85 (Finkelstein et al., 2015).

86 The introduction of high resolution three-dimensional ultrasound (3D US) scans combined
87 with visualization in immersive virtual reality (VR) systems, providing real depth
88 perception and more sensitive embryonic size measurements and morphological
89 evaluations, has markedly improved the opportunity to accurately study the
90 periconceptual period (time window: 14 weeks pre-conception to 10 weeks post-
91 conception) (Rousian et al., 2010; Baken et al., 2014; Steegers-Theunissen et al., 2013).
92 So far these innovative techniques were used to study embryonic crown-rump length

93 (CRL) and volume (EV) trajectories as non-invasive measures of first trimester embryonic
94 growth (Steegers-Theunissen et al., 2016). On the other hand, the Carnegie stages of
95 human embryonic development were introduced as a century old morphological
96 classification of fixated embryos dividing the embryonic period (58 post-conceptual
97 days) into 23 stages (O’Rahilly et al., 1987). The combination of 3D US and VR
98 visualization will allow us to investigate embryonic morphological development *in vivo*,
99 according to the longitudinal annotation of the Carnegie stages (O’Rahilly and Müller,
100 2010; Blaas et al., 1998; Verwoerd-Dikkeboom et al., 2008). Despite the fact that the
101 normal sequence of developmental events is constant and predictable in every embryo,
102 different times and velocities can occur, making comparisons possible and worthwhile.
103 Here, we aim to investigate the associations between periconceptual maternal
104 biomarkers of I-C metabolism and first trimester embryonic development, using serial
105 Carnegie stage annotation obtained by 3D US and VR.

106 **MATERIALS AND METHODS**

107 This study was performed in the setting of the Rotterdam Periconception Cohort (Predict
108 Study), a prospective periconceptual tertiary hospital-based cohort study started in 2009
109 at the Department of Obstetrics and Gynaecology of the Erasmus MC, University Medical
110 Centre, Rotterdam, with the aim to assess periconceptual determinants and predictors
111 of pregnancy outcome and offspring health (Steegers-Theunissen et al., 2016).

112 **Study population and sample**

113 All women before 8⁺⁰ weeks of gestation who conceived spontaneously, or after
114 intrauterine insemination (IUI), *in vitro* fertilization (IVF), intracytoplasmic sperm
115 injection (ICSI) or cryopreserved embryo transfer, were eligible for participation between

116 2010 and 2014 (figure 1). After exclusion for age below 18 years old, twins, miscarriage,
117 ectopic implantation, intrauterine fetal death, congenital anomalies and oocyte(s)
118 donation, 347 singleton ongoing pregnancies were enrolled. Since the Carnegie stages
119 describe embryonic development until the end of the embryonic period (10^{+2} weeks, 58
120 post-conceptual days), we excluded seven additional pregnancies for missing 3D US
121 scans before 10^{+2} weeks of gestation. Among spontaneously conceived pregnancies, we
122 selected pregnancies with known first day of last menstrual period (LMP), self-reported
123 regular cycle and observed crown-rump length (CRL) measurement corresponding to the
124 expected according to the Robinson curves (<7 days different) (Robinson and Fleming,
125 1975). The resulting total study population counted 234 pregnancies, consisting of 138
126 spontaneous or intrauterine insemination (IUI) pregnancies with strict pregnancy dating
127 and 96 IVF/ICSI pregnancies. Gestational age was defined from LMP for spontaneous
128 pregnancies (adjusted for duration of the menstrual cycle if <25 or >31 days), from LMP
129 or insemination date plus 14 days for IUI pregnancies, from the day of oocyte retrieval
130 plus 14 days for IVF/ICSI pregnancies, and from embryo transfer day plus 17 or 18 days
131 in pregnancies derived from transfer of cryopreserved embryos. Therefore, the total study
132 population included only pregnancies with strict and reliable dating by definition. Since a
133 possible influence of conception mode cannot be excluded, we performed the analysis
134 first in the total study population using conception mode as a confounder and we further
135 stratified the analysis to the two subgroups of strictly dated spontaneous and IVF/ICSI
136 pregnancies.

137 **General data**

138 Self-administered general questionnaires reporting items on age, geographical origin,
139 education, obstetric and medical history, and periconceptional lifestyle (smoking, alcohol
140 consumption, folic acid and multivitamin supplement use) were collected at enrolment.
141 Anthropometric measures were recorded by trained researchers.

142 **Blood sample analysis**

143 One first trimester fasting venous blood sample for serum vitamin B12, RBC folate and
144 plasma tHcy assessment was collected at enrolment and drawn in a vacutainer
145 ethylenediamine tetraacetate (EDTA) tube and in a dry vacutainer tube (BD diagnostics,
146 Plymouth, UK). The dry vacutainer tubes were centrifuged at 2,000 xg, serum was
147 collected and analyzed for vitamin B12 measurement using an immunoelectro-
148 chemoluminescence assay (E170; Roche Diagnostics GmbH, Mannheim, Germany).
149 Plasma was separated by centrifugation within one hour for determination of tHcy by
150 using a sensitive liquid chromatography tandem mass spectrum method (HPLC-Tandem
151 MS, Waters Micromass Quattro Premier XE Mass Spectrometer with Acquity UPLC
152 system, Milford, Massachusetts, United States). EDTA-blood was kept on ice and 0.1 ml
153 EDTA blood was hemolysed with 0.9 ml freshly prepared 1.0% ascorbic acid. The
154 hematocrit was determined with the ADVIA 120 Hematology Analyzer (Bayer Diagnostics,
155 Leverkusen, Germany). RBC folate was calculated with the following formula: (nM
156 hemolysate folate × 10/hematocrit) - [nM serum folate × (1 - hematocrit) /hematocrit] = nM
157 RBC folate.

158 **Ultrasound data**

159 From 6⁺⁰ up to 10⁺² weeks of gestation, all included women underwent serial 3D US scans
160 performed by trained researchers using the high frequency (4.5 – 11.9 MHz) vaginal

161 probe of a GE Voluson E8 (GE Healthcare, Zipf, Austria). Ultrasound scans were
162 performed on a weekly basis until 2013 and then reduced to a two weekly-basis after the
163 pilot study showed an accurate modelling of growth trajectory obtained with 3 scans per
164 pregnancy (at 7, 9, 11 weeks of gestation) (van Uitert et al., 2013b). The obtained 3D
165 datasets were stored as Cartesian volumes and transferred to the BARCO I-Space VR
166 system at the Department of Bioinformatics, Erasmus University Medical Centre,
167 Rotterdam. This system, running the V-Scope volume rendering application, aims to
168 improve dataset visualization by projecting a hologram in a 4-walled CAVE-like (Cave
169 Automatic Virtual Environment) VR system, allowing full depth perception and intuitive
170 interaction with the volume (Verwoerd-Dikkeboom et al., 2010; Koning et al., 2009). The
171 Carnegie criteria for external and internal morphological characteristics were used by one
172 trained researcher to stage all embryos, as previously described (Verwoerd-Dikkeboom
173 et al., 2008; O'Rahilly and Müller, 2010). As external morphological characteristics we
174 used the Carnegie criteria for the development of limbs (arms and legs) and embryonic
175 curvature. Internal morphological characteristics primarily included the criteria for the
176 brain cavity development. The assessment of Carnegie stages required 1 to 2 minutes
177 per embryo.

178 **Statistical analysis**

179 In order to evaluate selection bias, we compared maternal baseline characteristics and
180 biomarker concentrations between excluded and included pregnancies **using Chi-square**
181 **or exact tests for ordinal variables and Mann-Whitney U test for continuous variables.**
182 Univariable linear regression was performed to evaluate associations between maternal
183 baseline characteristics and biomarker concentrations.

184 To estimate associations between maternal biomarkers of I-C metabolism and embryonic
185 development, we treated the Carnegie stages as a continuous variable that was censored
186 at its maximum value of 23. This was used as the response variable in separate linear
187 mixed models estimated for the total study population and secondly for the subgroups of
188 strictly dated spontaneous and IVF/ICSI pregnancies. This analysis allows the linear
189 modelling of longitudinal measurements, taking into account the existing correlation
190 between serial measurements within the same pregnancy and potential confounders for
191 adjustment (parity, alcohol use, smoking, folic acid/multivitamin supplement use, fetal
192 gender, maternal age, BMI and comorbidity). Firstly, we performed a crude analysis with
193 adjustment for gestational age only (model 1) and secondly we adjusted for additional
194 confounders (model 2). Finally, the estimates of embryonic developmental change
195 expressed in days were determined comparing women with high (+2 standard deviation
196 (SD)) and low (-2 SD) concentrations of the biomarkers that were significantly associated
197 with the Carnegie stages in model 2. Due to the exclusion of pregnancies with uncertain
198 dating and the possibility of selection bias, we additionally performed a sensitivity analysis
199 including pregnancies with discordant CRL (n=15). P-values ≤ 0.05 were considered
200 significant. All analyses were performed using IBM SPSS version 21.0 (Armonk, NY: IBM
201 Corp) and R version 3.2.1 (The R Foundation for Statistical Computing).

202 **Ethical approval**

203 The protocol has been approved by the local medical ethics committee and all women
204 signed a written informed consent before participation.

205 **RESULTS**

206 We included 234 pregnancies with a median of three scans per pregnancy (range 1-5),
207 counting for a total of 745 3D US scans. The Carnegie stage annotation was feasible in
208 600 good quality datasets (success rate 80.5%). Carnegie stage distribution in the total
209 study population ranged from stage 13 to 23 (6^{+0} – 10^{+2} weeks of gestation). **Table I**
210 **shows maternal characteristics and biomarker concentrations at baseline with**
211 **comparisons between included and excluded ongoing pregnancies.** The prevalence of
212 hyperhomocysteinemia in the total study population was 1.3% ($> 13 \mu\text{mol/l}$). Vitamin B12
213 was significantly associated with maternal age ($\beta=0.15 \text{ pmol/l}$, (95%CI: 0.13; 0.16),
214 $p<0.05$), RBC folate ($\beta=0.20 \text{ pmol/l}$, (95% CI: 0.19; 0.20), $p<0.01$) and tHcy
215 concentrations ($\beta= -0.34 \text{ pmol/l}$, (95% CI: -0.37; -0.32), $p<0.001$). RBC folate was
216 significantly associated with maternal age ($\beta=0.20 \text{ nmol/l}$, (95% CI: 0.19; 0.21), $p<0.01$),
217 smoking ($\beta= -0.16 \text{ nmol/l}$, (95% CI: -0.25; -0.07), $p<0.05$), folic acid supplement use
218 ($\beta=0.23 \text{ nmol/l}$, (95% CI: 0.04; 0.43), $p<0.001$), comorbidity ($\beta= -0.14 \text{ nmol/l}$, (95% CI: -
219 0.24; -0.04), $p<0.05$) and tHcy concentrations ($\beta= -0.25 \text{ nmol/l}$, (95% CI: -0.27; -0.24),
220 $p<0.001$).

221 **Embryonic development**

222 Embryonic development according to the Carnegie stages was comparable between the
223 subgroups of strictly dated spontaneous and IVF/ICSI pregnancies (model 2, group effect:
224 $\beta= -0.20$, (95% CI: -0.46; 0.05), $p=0.12$). Table II shows the estimates from linear mixed
225 models. In model 2, vitamin B12 concentrations were positively associated with
226 embryonic development in the total study population and in strictly dated spontaneous
227 pregnancies, resulting in small, albeit significant estimates. In the total study population,
228 low vitamin B12 concentrations (-2 SD , corresponding to 73.4 pmol/l) were associated

229 with a 1.4-day delay (95% CI: 1.3-1.4) in embryonic development compared to high
230 concentrations (+2SD, corresponding to 563.1 pmol/l) (figure 2A). After full adjustment,
231 RBC folate was positively associated with the Carnegie stages only in the IVF/ICSI
232 subgroup, and low concentrations (-2SD, corresponding to 875.4 nmol/l) were associated
233 with a 1.8-day delay (95% CI: 1.7-1.8) in embryonic development compared to high
234 concentrations (+2SD, corresponding to 2119.9 nmol/l). Finally, tHcy was strongly and
235 negatively associated with the Carnegie stages in the total study population and in the
236 IVF/ICSI subgroup. In the total study population, high tHcy concentrations (+2SD,
237 corresponding to 10.4 μ mol/l) were associated with a 1.6-day delay (95% CI: 1.5-1.7) in
238 embryonic development compared to low concentrations (-2SD, corresponding to 3.0
239 μ mol/l) (figure 2B). The sensitivity analysis including pregnancies with discordant CRL
240 (n=15) did not modify the resulting associations (model 2, vitamin B12: β = 0.001, (95%
241 CI: 0.0001 – 0.002), p=0.03; RBC folate: β = 0.000, (95% CI: -0.000 - 0.001), p=0.06; tHcy:
242 β = -0.08, (95% CI: -0.15 - -0.02;), p=0.01).

243 DISCUSSION

244 This study shows significant associations between periconceptual maternal biomarkers
245 of I-C metabolism and embryonic morphological development according to the Carnegie
246 classification in ongoing non-malformed pregnancies. Moreover, IVF/ICSI conception did
247 not affect embryonic morphological development compared to spontaneous conception
248 in strictly dated pregnancies. The inclusion of pregnancies with discordant CRL revealed
249 the same associations.

250 Our results are in line with previous data showing associations between maternal I-C
251 metabolism and several reproductive, pregnancy and health outcomes (Solé-Navais et

252 al., 2016; Yajnik and Deshmukh, 2012). Recently, maternal early pregnancy high tHcy
253 ($\geq 8.31 \mu\text{mol/L}$) and low folate concentrations ($\leq 9.10 \text{ nmol/L}$) have been negatively
254 associated with fetal growth parameters, finally affecting birth weight (Bergen et al.,
255 2016). We also showed that an optimal periconceptional RBC folate level is associated
256 with increased first trimester longitudinal CRL measurements compared to the lowest ($\beta =$
257 $0.24 \sqrt{\text{mm}}$ (95%CI: 0.04; 0.44), $p = 0.02$) and highest quartile of concentrations ($\beta = 0.29$
258 $\sqrt{\text{mm}}$ (95%CI: 0.09; 0.49), $p < 0.01$) (van Uiter et al., 2014). This result emphasizes that
259 CRL accuracy in pregnancy dating is impacted by maternal I-C metabolism, as well as by
260 several maternal characteristics and exposures (van Uiter et al., 2013b). Moreover,
261 embryonic volume (EV) has been described as a more sensitive marker of first trimester
262 growth restriction compared to CRL (Baken et al., 2013). We focused on the Carnegie
263 stages as a century old classification that, together with embryonic size measurements,
264 could implement first trimester investigation and better define a proper embryonic
265 development. Since we excluded all pregnancies with congenital anomalies detected both
266 *in utero* and after birth, our results indicate that even the developmental events of normal
267 ongoing pregnancies are impacted by maternal I-C metabolism. This and previous
268 findings indicate that first trimester growth and development are important embryonic
269 outcomes affected by maternal environment. Nevertheless, CRL, EV and Carnegie
270 stages also represent non-invasive reproducible markers with predictable associations
271 with gestational age, leading to their potential use for pregnancy dating and raising the
272 question which biomarker should be the best candidate (Robinson and Fleming, 1975;
273 O'Rahilly and Müller, 2010). **Due to the lack of an optimal pregnancy dating strategy and**
274 **to unavoidable systematic errors related to the recall of the LMP, imprecise**

275 ovulation/implantation dates and parental characteristics impacting embryonic ultrasound
276 measurements, we defined gestational age based on a known LMP, regular cycle and
277 concordant CRL. In this way, all ultrasound measurements could be read as response
278 variables and outcome measurements. In order to reduce selection bias, we compared
279 maternal baseline characteristics, showing that excluded women had a higher BMI, lower
280 age and RBC folate concentrations. This may be mainly explained by the inclusion of a
281 large population of subfertile women and pregnancies achieved after IVF/ICSI treatment
282 (higher age, lower BMI, higher use of folic acid supplements). We also compared the
283 subgroup of included and excluded spontaneous pregnancies showing indeed no
284 significant results (data not shown). Moreover, the sensitivity analysis including
285 pregnancies with discordant CRL confirmed the detected associations, reducing the
286 possibility of selection bias.

287 The mechanisms linking maternal I-C metabolism and embryonic development are not
288 fully understood. Animal data showed that abnormal activations of I-C metabolism were
289 associated with hypermethylation of mitochondrial DNA, mitochondrial malfunction and
290 decreased oocyte quality (Jia et al., 2016). Recently, a suppression of the inflammatory
291 and upregulation of the high-density lipoprotein pathways have been demonstrated in
292 human follicular fluid of preconception folic acid supplement users (Twigt et al., 2015).
293 Cellular apoptosis and protein homocysteinylation, both dependent on tHcy
294 concentrations, have been suggested as contributors to neural tube, orofacial and cardiac
295 defects (Jakubowski, 2006; Taparia et al., 2007). Finally, periconceptual I-C biomarker
296 mediated epigenetic modifications could modify subsequent gene expression in the

297 embryo (Steeegers-Theunissen et al., 2013). All these events may finally lead to impaired
298 first trimester development, thereby supporting our results.

299 Our findings also reveal that conception mode seems to modify the associations between
300 blood biomarkers and Carnegie stages, despite the fact that no differences in embryonic
301 development have been detected between the two subgroups. As expected, biomarker
302 concentrations differed between spontaneous and IVF/ICSI pregnancies. Besides higher
303 and longer preconceptional folic acid supplement use in the IVF/ICSI subgroup, also the
304 ovarian stimulation treatment may affect I-C blood biomarker concentrations (Boxmeer et
305 al., 2008). Moreover, the IVF/ICSI technique has been associated with different
306 epigenetic patterns, gene expression and preimplantation embryo phenotype compared
307 to spontaneous conception, possibly affecting embryonic responses to maternal I-C
308 biomarkers and explaining different associations detected in our results (Kroener et al.,
309 2016; Zandstra et al., 2015; Giritharan et al., 2007; Song et al., 2015).

310 The major strength of our study is the longitudinal evaluation of embryonic development
311 using a median of three scans per patient, the use of 3D US with VR visualization and the
312 consequent high success rate of the Carnegie stage assessment. This gives an accurate
313 and precise picture of the course of first trimester development. Confounding by
314 gestational age is minimized by including women with strict pregnancy dating only. The
315 high rate of folic acid supplement use, resulting in an extremely low rate of
316 hyperhomocysteinemia and high RBC folate concentrations, strongly underlines the
317 importance of our results, since even clinically normal values of tHcy and a non-deranged
318 I-C metabolism could impact embryonic development of non-malformed ongoing
319 pregnancies. The most relevant limitation of this study is related to the tertiary care

320 setting, resulting in expected high rates of folic acid supplement use, chronic comorbidity
321 and pregnancy complications. This may reduce the external validity of our findings.
322 Despite it is reassuring that significant associations were confirmed in IVF/ICSI
323 pregnancies where conception date is known by definition, the implantation date is not
324 known and systematic errors in pregnancy dating are expected. Therefore, it is also
325 possible that the small differences detected in embryonic development reflect an impact
326 on the timing of implantation.

327 Inadequacies in dietary B vitamins and lifestyle (i.e. smoke, alcohol and coffee
328 consumption) have led to increased dangerous plasma tHcy concentrations in the last
329 decades (Stegers-Theunissen et al., 2013). Our results suggest that this may negatively
330 impact first trimester embryonic development resulting in the highest effect estimates in
331 line with previous findings (Blanco et al., 2016; Steegers-Theunissen et al., 2013). Since
332 plasma tHcy is an overall stable marker within the same individual and in uncomplicated
333 pregnancies, a random periconceptional tHcy measurement is reflective of an individual's
334 status and therefore could represent a potential useful predictor of embryonic
335 development in a clinical setting (McKinley et al., 2001; López-Alarcón et al., 2015).
336 Conversely, the small estimates detected for vitamin B12 and RBC folate may not
337 address for their clinical use as embryonic development predictors. Nevertheless, while
338 reduced CRL measurements have been associated with adverse pregnancy and health
339 outcomes in the offspring (Mook-Kanamori et al., 2010; van Uiter et al., 2013c; Jaddoe
340 et al., 2014), nothing is known about the clinical implications of first trimester
341 developmental delay in ongoing pregnancies.

342 In conclusion, we have shown significant associations between periconceptional maternal
343 biomarkers of I-C metabolism and Carnegie stages of embryonic development. Further
344 research is needed to investigate associations between Carnegie stages and birth
345 outcomes and to evaluate the validity of our results in the general population.

346 **AUTHORS' ROLES**

347 FP contributed to data collection, analysis and interpretation, she wrote the first draft and
348 revised all versions of the manuscript; MR performed embryonic measurements; AHJK
349 provided essential materials (V-scope software); SPW analyzed data and contributed to
350 the interpretation of results; IC supervised the writing of the manuscript; RPMST had
351 primary responsibility for final content, initiated the study and research questions and
352 supervised and contributed to all aspects of the study. All authors read and approved the
353 final manuscript.

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360 **CONFLICT OF INTEREST**

361 No conflict of interest has to be declared by any of the authors regarding the material
362 discussed in the manuscript. RPMST is CSO of the startup company Slimmere Zorg and
363 CEO of eHealth Care Solutions.

364 **REFERENCES**

- 365 Baken L, van Heesch PN, Wildschut HI, Koning AH, van der Spek PJ, Steegers EA,
366 Exalto N. First-trimester crown-rump length and embryonic volume of aneuploid fetuses
367 measured in virtual reality. *Ultrasound Obstet Gynecol* 2013;**41**:521-525.
- 368 Baken L, Rousian M, Koning AH, Bonsel GJ, Eggink AJ, Cornette JM, Schoonderwaldt
369 EM, Husen-Ebbinge M, Teunissen KK, van der Spek PJ, et al. First-Trimester Detection
370 of Surface Abnormalities: A Comparison of 2- and 3-Dimensional Ultrasound and 3-
371 Dimensional Virtual Reality Ultrasound. *Reprod Sci* 2014;**21**:993-999.
- 372 Bergen NE, Jaddoe VW, Timmermans S, Hofman A, Lindemans J, Russcher H, Raat H,
373 Steegers-Theunissen RP, Steegers EA. Homocysteine and folate concentrations in early
374 pregnancy and the risk of adverse pregnancy outcomes: the Generation R Study. *BJOG*
375 2012;**119**:739-751.
- 376 Bergen NE, Schalekamp-Timmermans S, Jaddoe VW, Hofman A, Lindemans J,
377 Russcher H, Tiemeier H, Steegers-Theunissen RP, Steegers EA. Maternal and Neonatal
378 Markers of the Homocysteine Pathway and Fetal Growth: The Generation R Study.
379 *Paediatr Perinat Epidemiol* 2016;**30**:386-396.
- 380 Blaas HG, Eik-Nes SH, Berg S, Torp H. In-vivo three-dimensional ultrasound
381 reconstructions of embryos and early fetuses. *Lancet* 1998;**352**:1182–1186.
- 382 Blanco R, Colombo A, Pardo R, Suazo J. Maternal biomarkers of methylation status and
383 non-syndromic orofacial cleft risk: a meta-analysis. *Int J Oral Maxillofac Surg* 2016. In
384 press.

- 385 Boxmeer JC, Steegers-Theunissen RP, Lindemans J, Wildhagen MF, Martini E, Steegers
386 EA, Macklon NS. Homocysteine metabolism in the pre-ovulatory follicle during ovarian
387 stimulation. *Hum Reprod* 2008;**23**:2570-2576.
- 388 Finkelstein JL, Layden AJ, Stover PJ. Vitamin B-12 and Perinatal Health. *Adv Nutr*
389 2015;**6**:552-563.
- 390 Giritharan G, Talbi S, Donjacour A, Di Sebastiano F, Dobson AT, Rinaudo PF. Effect of
391 in vitro fertilization on gene expression and development of mouse preimplantation
392 embryos. *Reproduction* 2007;**134**:63-72.
- 393 Hogeveen M, Blom HJ, den Heijer M. Maternal homocysteine and small-for-gestational-
394 age offspring: systematic review and meta-analysis. *Am J Clin Nutr* 2012;**1**:130-136.
- 395 Jakubowski H. Pathophysiological consequences of homocysteine excess. *J Nutr*
396 2006;**136**:1741S-1749S.
- 397 Jaddoe VW, de Jonge LL, Hofman A, Franco OH, Steegers EA, Gaillard R. First trimester
398 growth restriction and cardiovascular risk factors in childhood. *BMJ* 2014;**348**:g14.
- 399 Jia L, Li J, He B, Jia Y, Niu Y, Wang C, Zhao R. Abnormally activated one-carbon
400 metabolic pathway is associated with mtDNA hypermethylation and mitochondrial
401 malfunction in the oocytes of polycystic gilt ovaries. *Sci Rep* 2016;**6**:19436.
- 402 Kalhan SC. One carbon metabolism in pregnancy: Impact on maternal, fetal and neonatal
403 health. *Mol Cell Endocrinol* 2016. In press.
- 404 Koning AH, Rousian M, Verwoerd-Dikkeboom CM, Goedknecht L, Steegers EA, van der
405 Spek PJ. V-scope: design and implementation of an immersive and desktop virtual reality
406 volume visualization system. *Stud Health Technol Inform* 2009;**142**:136-138.

407 Kroener L, Wang ET, Pisarska MD. Predisposing Factors to Abnormal First Trimester
408 Placentation and the Impact on Fetal Outcomes. *Semin Reprod Med* 2016;**34**:27-35.

409 López-Alarcón M, Montalvo-Velarde I, Vital-Reyes VS, Hinojosa-Cruz JC, Leaños-
410 Miranda A, Martínez-Basila A. Serial determinations of asymmetric dimethylarginine and
411 homocysteine during pregnancy to predict pre-eclampsia: a longitudinal study. *BJOG*
412 2015;**122**:1586-1592.

413 McKinley MC, Strain JJ, McPartlin J, Scott JM, McNulty H. Plasma homocysteine is not
414 subject to seasonal variation. *Clin Chem* 2001;**47**:1430-1436.

415 Mook-Kanamori DO, Steegers EA, Eilers PH, Raat H, Hofman A, Jaddoe VW. Risk factors
416 and outcomes associated with first-trimester fetal growth restriction. *JAMA* 2010;**303**:527-
417 534.

418 O'Rahilly R. Human embryo. *Nature* 1987;**329**:385.

419 O'Rahilly R, Müller F. Developmental stages in human embryos: revised and new
420 measurements. *Cells Tissues Organs* 2010;**192**:73-84.

421 Robinson HP, Fleming JE. A critical evaluation of sonar 'crown-rump length'
422 measurements. *BJOG* 1975;**82**:702-710.

423 Ronnenberg AG, Venners SA, Xu X, Chen C, Wang L, Guang W, Huang A, Wang X.
424 Preconception B-vitamin and homocysteine status, conception, and early pregnancy loss.
425 *Am J Epidemiol* 2007;**166**:304-312.

426 Rousian M, Koning AH, van Oppenraaij RH, Hop WC, Verwoerd-Dikkeboom CM, van der
427 Spek PJ, Exalto N, Steegers EA. An innovative virtual reality technique for automated
428 human embryonic volume measurements. *Hum Reprod* 2010;**25**:2210-2216.

- 429 Solé-Navais P, Cavallé-Busquets P, Fernandez-Ballart JD, Murphy MM. Early pregnancy
430 B vitamin status, one carbon metabolism, pregnancy outcome and child development.
431 *Biochimie* 2016;**126**:91-96.
- 432 Song S, Ghosh J, Mainigi M, Turan N, Weinerman R, Truongcao M, Coutifaris C,
433 Sapienza C. DNA methylation differences between in vitro- and in vivo-conceived children
434 are associated with ART procedures rather than infertility. *Clin Epigenetics* 2015;**7**:41.
- 435 Steegers-Theunissen RP, Boers GH, Trijbels FJ, Eskes TK. Neural-tube defects and
436 derangement of homocysteine metabolism. *N Engl J Med* 1991;**324**:199-200.
- 437 Steegers-Theunissen RP, Twigt J, Pestinger V, Sinclair K. The periconceptional period,
438 reproduction and long-term health of offspring: the importance of one-carbon metabolism.
439 *Hum Reprod Update* 2013;**19**:640-655.
- 440 Steegers-Theunissen RP, Verheijden-Paulissen JJ, van Uitert EM, Wildhagen MF, Exalto
441 N, Koning AH, et al. Cohort Profile: The Rotterdam Periconceptional Cohort (Predict
442 Study). *Int J Epidemiol* 2016;**45**:374-381.
- 443 Taparia S, Gelineau-van Waes J, Rosenquist TH, Finnell RH. Importance of folate-
444 homocysteine homeostasis during early embryonic development. *Clin Chem Lab Med*
445 2007;**45**:1717-1727.
- 446 Twigt JM, Bezstarosti K, Demmers J, Lindemans J, Laven JS, Steegers-Theunissen RP.
447 Preconception folic acid use influences the follicle fluid proteome. *Eur J Clin Invest*
448 2015;**45**:833-841.
- 449 van Uitert EM, Steegers-Theunissen RP. Influence of maternal folate status on human
450 fetal growth parameters. *Mol Nutr Food Res* 2013a;**57**:582-595.

451 van Uitert EM, van der Elst-Otte N, Wilbers JJ, Exalto N, Willemsen SP, Eilers PH, Koning
452 AH, Steegers EA, Steegers-Theunissen RP. Periconception maternal characteristics and
453 embryonic growth trajectories: the Rotterdam Predict study. *Hum Reprod* 2013b;**28**:3188-
454 3196.

455 van Uitert EM, Exalto N, Burton GJ, Willemsen SP, Koning AH, Eilers PH, et al. Human
456 embryonic growth trajectories and associations with fetal growth and birthweight. *Hum*
457 *Reprod* 2013c;**28**:1753-1761.

458 van Uitert EM, van Ginkel S, Willemsen SP, Lindemans J, Koning AH, Eilers PH, et al.
459 An optimal periconception maternal folate status for embryonic size: the Rotterdam
460 Predict study. *BJOG* 2014;**121**:821-829.

461 Verwoerd-Dikkeboom CM, Koning AH, van der Spek PJ, Exalto N, Steegers EA.
462 Embryonic staging using a 3D virtual reality system. *Hum Reprod* 2008;**23**:1479-1484.

463 Verwoerd-Dikkeboom CM, Koning AH, Hop WC, van der Spek PJ, Exalto N, Steegers
464 EA. Innovative virtual reality measurements for embryonic growth and development. *Hum*
465 *Reprod* 2010;**25**:1404-1410.

466 Vollset SE, Refsum H, Irgens LM, Emblem BM, Tverdal A, Gjessing HK, Monsen AL,
467 Ueland PM. Plasma total homocysteine, pregnancy complications, and adverse
468 pregnancy outcomes: the Hordaland Homocysteine study. *Am J Clin Nutr* 2000;**71**:962-
469 968.

470 Yajnik CS, Deshmukh US. Fetal programming: maternal nutrition and role of one-carbon
471 metabolism. *Rev Endocr Metab Disord* 2012;**13**:121-127.

472 Yajnik CS, Chandak GR, Joglekar C, Katre P, Bhat DS, Singh SN, Janipalli CS, Refsum
473 H, Krishnaveni G, Veena S, et al. Maternal homocysteine in pregnancy and offspring

474 birthweight: epidemiological associations and Mendelian randomization analysis. *Int J*
475 *Epidemiol* 2014;**43**:1487-1497.

476 Zandstra H, Van Montfoort AP, Dumoulin JC. Does the type of culture medium used
477 influence birthweight of children born after IVF? *Hum Reprod* 2015;**30**:530-542.

478 **FIGURE LEGENDS**

479 **Figure 1. Flow chart of the study population.** IUFD: intrauterine fetal death, US:
480 ultrasound, LMP: last menstrual period, CRL: crown-rump length, IUI: intrauterine
481 insemination, IVF: in vitro fertilization, ICSI: intracytoplasmic sperm injection.

482 **Figure 2. Average regression lines for vitamin B12 (A) and total homocysteine**
483 **(tHcy) (B) concentrations in the total study population.** In model 2, a low vitamin B12
484 (-2 standard deviation (SD), corresponding to 73.4 pmol/l) delays embryonic development
485 by 1.4 days (95% CI: 1.3-1.4) compared to high concentrations (+2SD, 563.1 pmol/l).
486 Conversely, high tHcy concentrations (+2SD, 10.4 $\mu\text{mol/L}$) delay embryonic development
487 by 1.6 days (95% CI: 1.5-1.7) compared to low concentrations (-2SD, 3.0 $\mu\text{mol/l}$). GA:
488 gestational age.

489

490 **Table I. Maternal baseline characteristics and biomarkers of I-C metabolism.**

491

Maternal characteristics	Total study population (n=234)	M	Excluded population (n=118)	M	p-value
Age, y median (range)	32 (22-42)	0	30 (21-44)	0	0.00
Geographical origin		1		5	0.30
Western, n(%)	206 (88.0)		104 (88.1)		
Non Western, n(%)	27 (11.5)		9 (7.6)		
Educational level		1		5	0.08
High, n(%)	135 (57.7)		65 (55.1)		
Intermediate, n(%)	93 (39.7)		45 (38.1)		
Low, n(%)	5 (2.1)		3 (2.5)		
BMI, kg/m ² median (range)	24.2 (17-42.3)	1	25.8 (17.8-45.0)	2	0.01
Nulliparous, n(%)	74 (31.8)	1	39 (33.9)	2	0.69
Alcohol use, n(%)	83 (35.8)	2	38 (34.2)	7	0.78
Periconception smoking, n(%)	32 (13.7)	1	21 (19.1)	8	0.20
Periconception folic acid/multivitamin use, n(%)	224 (97.4)	4	108 (93.9)	3	0.11
Chronic diseases, n(%)	25 (10.7)	0	22 (18.6)		0.05
Vitamin B12 (pmol/l) median (range)	297 (95-953)	0	295.5 (109-915)	20	0.76
RBC folate (nmol/l) median (range)	1408 (541-2811)	12	1294 (634-1942)	23	0.01
tHcy (µmol/l) median (range)	6.4 (3.7-17.6)	3	6.2 (3.4-13.6)	23	0.51

492

The total study population includes strictly dated pregnancies achieved after spontaneous conception (n=138) or IVF/ICSI (n=96). Excluded pregnancies include oocyte(s) donation (n=5), missing 3D US scans before 10⁺² weeks of gestation (n=7) and spontaneous pregnancies with discordant CRL measurements (≥ 7 days, n=15), unknown LMP (n=14) or self-reported irregular cycle (n=77). Chronic diseases include cardiovascular, autoimmune, endocrine and metabolic diseases. The comparison was performed using Chi-square or exact tests for ordinal variables and Mann-Whitney U test for continuous variables. M: missing values, BMI: body mass index, RBC: red blood cell, tHcy: total homocysteine.

494 **Table II. Maternal biomarker effect estimates for the Carnegie stages of**
 495 **embryonic development derived from linear mixed models.**

Biomarkers	EFFECT ESTIMATES CARNEGIE STAGES β (95%CI)	
	Model 1	Model 2
Total study population (n=234)		
Vitamin B12	0.001 (0.000; 0.002) *	0.001 (0.000; 0.002) *
RBC folate	0.0004 (0.0001; 0.0007) *	0.000 (0.000; 0.001)
tHcy	-0.09 (-0.15; -0.03) **	-0.08 (-0.14; -0.02) **
Strictly dated spontaneous pregnancies (n=138)		
Vitamin B12	0.002 (0.001; 0.003) *	0.002 (0.001; 0.003) *
RBC folate	0.000 (-0.000; 0.001)	0.003 (0.002; 0.004)
tHcy	-0.07 (-0.17; 0.03)	-0.07 (-0.10; 0.02)
IVF/ICSI pregnancies (n=96)		
Vitamin B12	-0.0004 (-0.002; 0.0008)	-0.000 (-0.002; 0.001)
RBC folate	0.000 (-0.000; 0.001)	0.001 (0.0005; 0.0015) *
tHcy	-0.09 (-0.16; -0.02) **	-0.08 (-0.15; -0.01) *

Effect estimates represent the change in Carnegie stage per unit of increase of biomarker concentration. Model 1 shows the crude model with adjustment for gestational age. Model 2 includes adjustment for potential confounders (parity, alcohol use, smoking habit, folic acid use, age, BMI, chronic diseases, fetal gender).

RBC: red blood cell, IVF: *in vitro* fertilization, ICSI: intracytoplasmic sperm injection, tHcy: total homocysteine; CI: confidence interval.

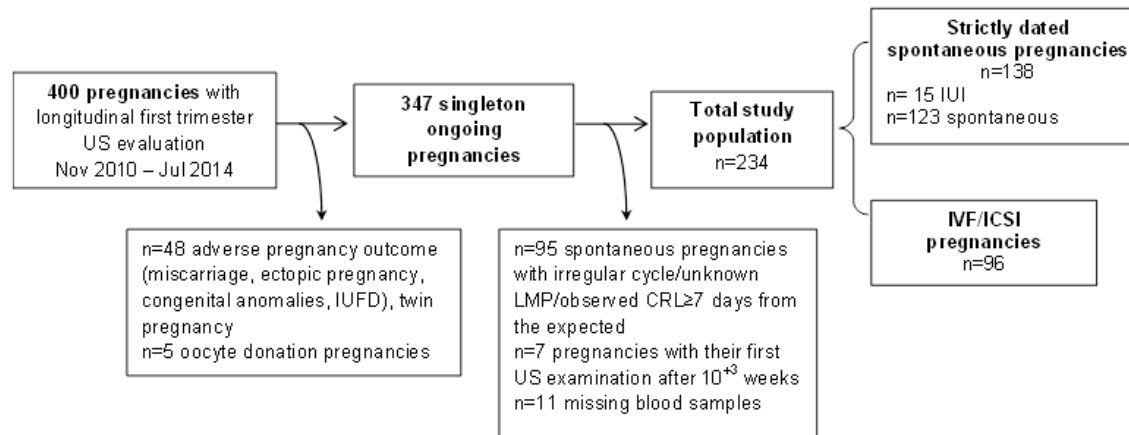
496 * $p < 0.05$, ** $p \leq 0.01$

497

498

499 FIGURE 2

500

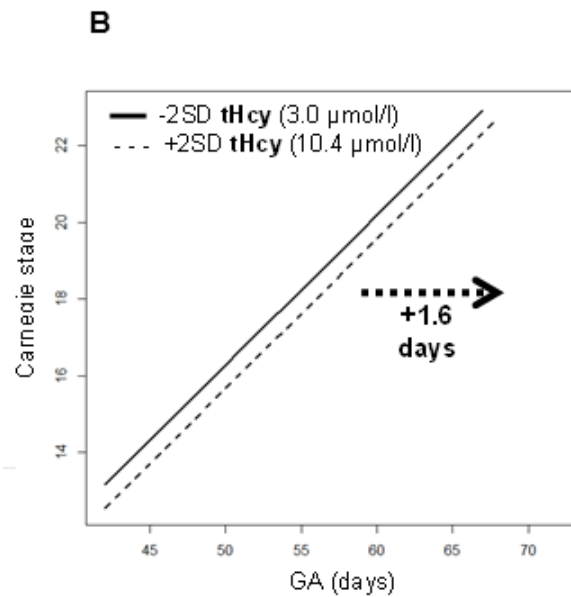
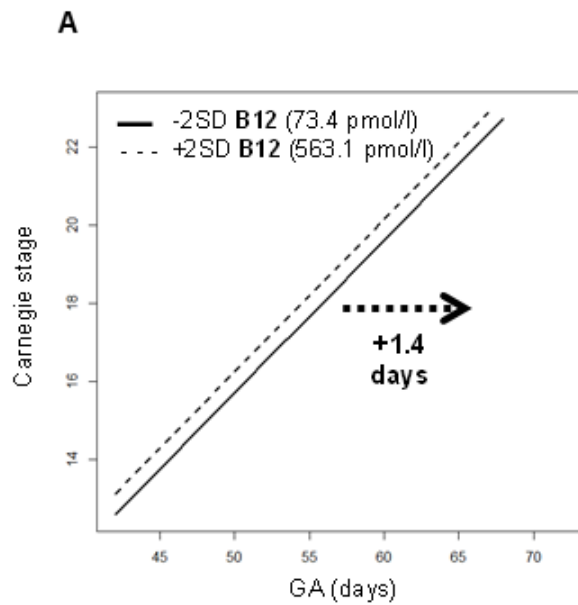


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503 FIGURE 2

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