| 1 | Proteomics and metabolomics characterizing the pathophysiology of adaptive reactions to the |
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| 2 | metabolic challenges during the transition from late pregnancy to early lactation in dairy |
| 3 | cows |
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| 8 | List of abbreviations |
| 9 | AGP, α_1 -acid glycoprotein; APP, acute phase protein; ATP, adenosine triphosphate; BHB, β - |
| 10 | hydroxybutyrate; CoA, Coenzyme A; DIM, days in milk; GPC, glycerophosphocholine; NADP, |
| 11 | nicotinamide adenine dinucleotide phosphate; NAFLD, non-alcoholic fatty liver disease; NEFA, |
| 12 | non-esterified fatty acids; NNB, negative nutrient balance; PC, phosphocholine; PI, physiological |
| 13 | imbalance; SAA, serum amyloid A; SARA, subacute ruminal acidosis; TCA cycle, tricarboxylic |
| 14 | acid cycle; TG, triglyceride; VLDL, very low density lipoprotein. |
| 15 | Abstract |

The transition from late pregnancy to early lactation is a critical period in a dairy cow's life due to 16 the rapidly increasing drain of nutrients from the maternal organism towards the foetus and into 17 colostrum and milk. In order to cope with the challenges of parturition and lactation, comprehensive 18 adaptive reactions comprising the endocrine and the immune system need to be accomplished. 19 There is high variation in this coping ability and both metabolic and infectious diseases, 20 21 summarized as "production diseases", such as hypocalcaemia (milk fever), fatty liver syndrome, laminitis and ketosis, may occur and impact welfare, productive lifespan and economic outcomes. 22 Proteomics and metabolomics have emerged as valuable techniques to characterize proteins and 23

metabolite assets from tissue and biological fluids, such as milk, blood and urine. In this review we provide an overview on metabolic status and physiological changes during the transition period and the related production diseases in dairy cows, and summarize the state of art on proteomics and metabolomics of biological fluids and tissues involved in metabolic stress during the peripartum period. We also provide a current and prospective view of the application of the recent achievements generated by omics for biomarker discovery and their potential in diagnosis.

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31 Significance

For high-yielding dairy cows there are several "occupational diseases" that occur mainly during the 32 33 metabolic challenges related to the transition from pregnancy to lactation. Such diseases and their sequelae form a major concern for dairy production, and often lead to early culling of animals. 34 Beside the economical perspective, metabolic stress may severely influence animal welfare. There 35 36 is a multitude of studies about the metabolic backgrounds of such so called production diseases like ketosis, fatty liver, or hypocalcaemia, although the investigations aiming to assess the complexity of 37 the pathophysiological reactions are largely focused on gene expression, i.e. transcriptomics. For 38 extending the knowledge towards the proteome and the metabolome, the respective technologies are 39 of increasing importance and can provide an overall view of how dairy cows react to metabolic 40 41 stress, which is needed for an in-depth understanding of the molecular mechanisms of the related diseases. We herein review the current findings from studies applying proteomics and 42 metabolomics to transition-related diseases, including fatty liver, ketosis, endometritis, 43 44 hypocalcaemia and laminitis. For each disease, a brief overview of the up to date knowledge about its pathogenesis is provided, followed by an insight into the most recent achievements on the 45 46 proteome and metabolome of tissues and biological fluids, such as blood serum and urine, highlighting potential biomarkers. We believe that this review would help readers to be become 47 more familiar with the recent progresses of molecular background of transition-related diseases thus 48 encouraging research in this field. 49

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51 Introduction

The transition period from late pregnancy to early lactation is a critical period in a dairy cow's life 52 53 due to the rapidly increasing drain of nutrients from the maternal organism towards the foetus and 54 into colostrum and milk. During this transition period, fetal growth reaches its exponential course during the last weeks of pregnancy and concomitantly the mammary gland parenchyma mass 55 56 markedly grows [1]. After calving, the output of nutrients with milk exceeds the input by voluntary feed intake. The negative nutrient balance (NNB) resulting therefrom requires a massive 57 mobilization of body reserves, mainly body fat but also protein. Albeit NNB is a common 58 59 phenomenon in mammals, both the duration and the extent observed in modern high yielding dairy cows represent a biological extreme. To be able to cope with the challenges of parturition and 60 lactation, comprehensive adaptive mechanisms comprising the endocrine and the immune system 61 need to be accomplished. There is high variation in this coping ability and both metabolic and 62 infectious diseases, summarized as "production diseases", may occur and have an impact on 63 64 welfare, productive lifespan and economic outcomes. The incidence of such diseases is greatest during early lactation with hypocalcaemia (milk fever), fatty liver syndrome and ketosis (or 65 acetonaemia) being the most common metabolic diseases. In case of infectious diseases, metritis 66 67 and mastitis, attributable to the immune-compromised situation during the metabolic challenge, are most frequent. Fig.1 presents the relationship between metabolic stress and disease development. 68 Several studies have attempted to identify the causes and risk factors associated with the high 69 70 incidence of health problems observed during the periparturient period [2–5], and systems biology approaches addressing the issue of the regulatory mechanisms of nutrient metabolism in lactation 71 72 are published [6–8]. Beside environmental factors, production diseases also have a genetic component; e.g. for subclinical macromineral disorders and major clinical diseases the heritability 73 74 reported were low to moderate [9].

75 To further investigate the complexity of these diseases, omics approaches which include multivariate, and large-scale analyses, may be applied. Such studies gather information either at the level 76 of the DNA, RNA, miRNA, protein or the metabolites, and provide a snapshot of the current 77 78 condition in cells, tissues or body fluids (Fig 2). However, variation in sampling times between different studies can yield different results and it is important to take this into account when 79 interpreting omics results or planning further studies. The multivariate results from omics 80 81 approaches require extensive bioinformatics resources that are mostly available online. Both proteomics and metabolomics have evolved as the functional continuation of transcriptomics in less 82 than two decades, and have developed rapidly, due to improvements in technology and 83 84 bioinformatics tools. General reviews about the application of proteomics and metabolomics to livestock science were published earlier [10–12]. Both proteomics and metabolomics involve the 85 resolution of a complex mixture of compounds into components that can then be identified and 86 87 characterized. For what concerns proteins, their identification always involves matching each the amino acid sequence to the respective encoding gene and thus depends of the sequence information 88 89 available for the target species, but can also include the description of posttranslational protein 90 modifications.

Two major mass spectrometry (MS) platforms are available for proteomics, following the 91 92 mechanism through which ions are generated: these ion sources are termed matrix-assisted laser 93 desorption/ionization (MALDI) and electrospray ionization (ESI). Before analysis, proteins are fractionated either by electrophoretic, for intact proteins, or chromatographic, for peptides generated 94 after protein cleavage, techniques. Protein fractions are then digested, to generate peptides that can 95 be further fractionate by chromatographic techniques and then characterized by MS, which can 96 record the mass of analytes to generate information about their structure. The resulting data are 97 further analysed with search engines, such as Mascot (Matrix Science Ltd), to generate in silico MS 98 data for the specified genome sequence database. 99

Absolute protein quantification is difficult to achieve with proteomics techniques. On the contrary, relative quantitation can be achieved by gel-based methods, such as 2DE using semiquantitative protein stains, or protein labeling strategies, such as difference gel electrophoresis (DIGE) [13]. As an example, DIGE was applied to blood serum samples to identify potential biomarkers related to hypocalcaemia [14].

At the peptide level, relative quantification can be achieved by stable isotope-labeling approaches
(iTRAQ) or by label-free comparison. Isolated proteins or tryptic peptides can be chemically
labeled before separation (iTRAQ) [15]. Quantification of proteins by means of iTRAQ was used,
among the others, to identify liver proteins related to physiological imbalance [16].

109 For metabolites, species-specificity is not an issue albeit relative quantities may differ. The major analytical approaches used in metabolomics rely on two techniques: MS and NMR techniques [17]. 110 For MS-based techniques, samples are fractionated through chromatographic techniques such as 111 112 Gas Chromatography (GC) of High Pressure Liquid Chromatography (HPLC), or capillary electrophoresis. The GC is used to fractionate volatile metabolites, such as for example BHB of 113 other organic acids, or fatty acids, whereas HPLC is used for lipophilic metabolites, such as acyl 114 carnitines for example. Fractionated metabolites are then ionized and identified following mass 115 spectrometry analysis. NMR techniques can identify simultaneously all the analytes, and do not 116 117 require any prefractionation of the sample. Successful identification of individual metabolites depends of high quality mass spectra, powerful spectral *matching* algorithms and comprehensive 118 and reliable spectral libraries [18]. 119

120 Metabolomics strategies are commonly divided into targeted and non-targeted metabolomics.

121 Targeted metabolomics aims to quantify defined groups of metabolites, such as for example, in case

122 of transition period-related diseases, acylcarnitines, carbohydrates, amino acids or organic acids.

123 Internal standards, such as stabile isotope (¹H and ¹³C) labelled metabolites are used to measure

analytes in a quantitative or semi-quantitative ways [19]. Non-targeted metabolomics allows for the

detection of all the metabolites in a sample, in theory. Indeed, non-targeted metabolomics is able to

detect up to 10,000 independent spectral features in biological samples [20], but only a fraction of 126 127 them can be actually identified. Targeted metabolomics was already applied in several dairy cow studies e.g. to characterize the respective changes in blood throughout the transition phase [21–24] 128 and non-targeted metabolomics for testing a supplement aiming to ameliorate metabolic stress [25], 129 respectively. The aim of this review is to provide an overview of the state of the art applications of 130 proteomics and metabolomics to address production diseases of dairy cows, whose prevention 131 132 represents a priority for intensive milk production. We reconcile and summarize the information currently available on metabolic status and physiological changes during the transition period, and 133 discuss the most recent achievements in proteomics and metabolomics applications for biomarker 134 135 discovery, their potential as diagnostic tools, but also for comprehension of complex (patho) physiological contexts. However, the review will be largely limited to metabolic diseases and to 136 metritis. For mastitis, the reader is referred to the specific "mastitomics" literature [26–28] and also 137 138 to the latest considerations of meta-proteomics and meta-metabolomics which include the milk microbiome as recently reviewed by Addis and coworkers [29]. 139 140 Metabolomics by means of near infrared spectroscopy is used since many years in routine milk recordings to assess gross milk composition (mainly the content of fat, protein and lactose), but is 141 also increasingly considered for minor milk constituents to provide information about the 142 143 physiological status of the animals [30]. Such information could be used for herd management decisions and for phenotyping in animal breeding as well. However, to be able to relate the spectra 144

obtained to certain metabolites and, in consequence, to identify health disturbances, circumspect
validations and algorithms are necessary [31–33]. Those applications are beyond the scope of this
review that will be limited to the identification of metabolites related to production diseases in
tissues, mainly liver and adipose tissue, and biological fluids, such as blood, but also urine and milk
where appropriate.

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151 Metritis and endometritis

The uterine mucosal environment is protected, in principle, from invading pathogens by physical 152 153 anatomical barriers and molecular mechanisms. However, dairy cows may develop endometritis or metritis (an inflammation of the inner mucosal layer of the uterus, or of the entire uterine wall, 154 respectively), when these mechanisms are compromised. The opening of the cervix for giving birth 155 as well as tissue lesions related to labour and expulsion of the calf and placental membranes violate 156 the anatomical barriers, concomitantly the innate immune defense is suppressed thus facilitating 157 158 bacterial infections that cause metritis and endometritis [34]. These diseases have a high incidence postpartum [35–37] and often result in decreased fertility as shown by reduced conception rates and 159 increased calving-to-conception intervals [38]. 160

161 A pioneering proteomic study provided first clues about the alterations in blood serum in pathological (endometritis) versus physiological states [39]. Patterns indicative for an inflammatory 162 reaction were also found in healthy animals around calving as demonstrated by increasing 163 164 concentrations of positive acute phase proteins (APP), such as \Box_1 -acid glycoprotein (AGP) and haptoglobin, and a correspondent decrease of negative APP, such as \Box_2 HS glycoprotein (fetuin-A) 165 suggesting that the approaching calving resembles an acute-phase reaction. This is in line with the 166 common observation of an inflammatory reaction peripartum that is considered as physiological and 167 even necessary for the successful adaptation to the metabolic challenge as recently reviewed [40]. 168 169 In the proteome study [39], the concentrations of AGP two weeks before calving were lower in animals that developed endometritis postpartum, but were higher two weeks after calving, 170 suggesting AGP not only as a potential prepartum bioindicator for an early detection of an increased 171 172 risk for endometritis but also indicating a complex functional role for AGP in the context of endometritis. 173 In a 2-DGE analysis followed by MALDI-TOF, endometritis-associated proteins were identified in 174

endometrial samples [41]. Several proteins, including desmin, α -actin-2, heat-shock protein (HSP)

176 27, peroxiredoxin-6, luteinizing hormone receptor isoform 1, collectin-43 precursor,

177 deoxyribonuclease-I (DNase-I), and MHC class I heavy chain (MHC-Ih) were up-regulated in

endometritis, whereas, transferrin, interleukin-2 precursor, hemoglobin β subunit, and potassium 178 179 channel tetramerisation domain containing 11 (KCTD11) were down-regulated as compared to normal endometrium. Desmin and α -actin-2 identified in this proteomic study to be related with 180 endometritis are common in mammalian cells, but being also up-regulated during other diseases, 181 such as cancer [42], they would not qualify as specific biomarkers for endometritis. However, truly 182 specific associations between the proteins found to be divergently regulated are unlikely anyway, in 183 184 view of the pleiotropic defense reactions. Fig. 3 summarizes the differences in proteomes of blood and endometrial samples associated with the development of metritis. When comparing the uterine 185 proteome of cows infected with a specific bacterium (Trueperella pyogenes - formerly 186 187 Arcanobacterium), with that of uninfected cows, using 2-DGE, annexins A1 and A2 (ANXA1 and ANXA2), apolipoprotein A-1, calprotectin (S100A9), cathelicidin, enolase 1 (ENO1), 188 peptidoglycan recognition protein 1 (PGLYRP1), phosphoglycerate mutase 1 (PGAM1), serine 189 190 dehydratase (SDS) and serine protease inhibitors (SERPIN) B1, B3 and B4 proteins were found to be differentially regulated [43]. In the second part of the study ten of these proteins were monitored 191 192 in uterine samples from dairy cows at 15 and 42 days post-partum, and strong positive correlations 193 between the cytology scores (percentage of polymorphonuclear neutrophils) and cathelicidin, PGLYRP1, SERPINB1 and S100A9 levels at day 15 were found. 194 195 Retention of the placenta e.g. the failure to expel the placenta within 12 - 24 h after calving, is known as a major predisposing factor for the development of endometritis or metritis and thus 196 impairs fertility but also animal health in general [44]. Beside infectious diseases, non-infectious 197 risk factors like dystocia, but also nutritional deficiencies, are listed as causes for the placental 198 retention albeit the aetiology is not completely understood [44]. Few investigations on proteomic 199 differences between retained and normal placenta were carried out so far. A review describing the 200 involvement of extracellular matrix proteins in placenta release was recently published [45]. The 2-201

DGE reference map for bovine placenta during late pregnancy identified 273 proteins, providing the

background for studies on molecular mechanism of placenta modification and diseases during latepregnancy [46].

Protein differences between retained and normally delivered placentae were studied starting from 205 tissue obtained from both the fetal and the maternal side of the placenta (i.e. cotyledon villi and 206 caruncle crypts). Using 1-D and 2-DE, differences between the protein profiles in the two groups 207 were assessed by means of computer-aided analysis but the identification of specific proteins 208 209 remained undone [47]. In a follow-up study, the protein patterns in normal and in retained placentae were investigated by 2-DIGE [48]. In this study, differentially regulated proteins were identified by 210 means of MALDI-TOF analysis. Comparisons between fetal healthy/retained and maternal 211 212 healthy/retained placentae yielded only five differentially regulated proteins. Albeit preliminary, the results point to an involvement of RabGTPases, which are known as master regulators of 213 intracellular trafficking: Ras-related protein Rab-7b was up-regulated only in healthy maternal 214 215 placenta, whereas Rab GDP dissociation inhibitor beta was up-regulated in the cotyledons of both retained and healthy placentae. In addition, short transient receptor potential channel 5 was 216 identified in caruncles of both retained and healthy placentae, and transforming growth factor $\Box 2$ 217 was highly abundant in both maternal and fetal parts of retained placenta. The proteins identified in 218 placental tissues indeed aggrandize the spectrum of relevant pathways to consider in the context of 219 220 placental maturation. However, these results were limited to tissue analyses and thus realistic predictive approaches for retained placentae using body fluids are not coming into reach so far. 221 The application of metabolomics techniques to metritis and endometritis is still lacking, in both 222 223 human and veterinary medicine.

Further studies have to be conducted to distinguish between physiological and pathological protein and metabolite patterns due to the challenges of parturition and thus to unravel the complexity of the underlying processes; identifying early indicators for the cow's ability to cope with the situation for eventually providing metaphylactic measures is a further goal.

229 Hypocalcaemia (milk fever)

The requirements for calcium (Ca) increase dramatically towards the end of pregnancy and the 230 onset of lactation. The Ca content of milk is about 1.2 g/L and a modern dairy cow may produce up 231 to 60 L per day during peak lactation, resulting in a daily Ca loss of more than 70 g/d. Thus, 232 metabolic adaptations need to be activated, otherwise the blood concentration of Ca falls below a 233 critical threshold and clinical and subclinical hypocalcaemia can result [49]. Hypocalcaemia 234 235 impacts health, future milk production, and reproductive performance and has been demonstrated to be linked with compromised immune function; cows with lower blood Ca concentrations within the 236 first day after calving were more likely to have retained placenta and resulting metritis, and mastitis 237 238 [50]. In addition, hypocalcaemia is also associated with metabolic diseases such as left displaced abomasum, ketosis and fatty liver [51,52]. 239

Both proteomics and metabolomics provided some molecular insight into pathogenesis of 240 241 hypocalcaemia. Proteomic comparisons of plasma samples from dairy cows with or without milk fever were performed by 2-DIGE, followed by in-gel digestion and MALDI-TOF-MS analysis for 242 peptide mass fingerprinting of selected protein spots [53]. Out of 23 protein spots found to be 243 different between the groups, eight were isolated and identified representing five unique proteins: 244 serpin peptidase inhibitor (angiotensin) and endopin 2B were increased in hypocalcaemic animals, 245 246 whereas albumin, fibrinogen beta chain, and IgG heavy-chain C-region (IgG-C(H)) were downregulated. Interestingly, the study demonstrated also a shift in the electrophoretic mobility of 247 albumin and angiotensin, suggesting that milk fever not only changes their concentration, but 248 249 possibly also their post-translational modification. In another study using weak cationic exchange protein chips for plasma protein profiling by SELDI-TOF-MS, six proteins were identified in 250 251 animals with subclinical hypocalcaemia (average milk yield 30 kg/day) differing from the healthy controls (average milk yield 28 kg/day) [54]: albumin, fibrinogen alpha chain, amyloid beta A4 252 proteins and VGF were increased, and apolipoprotein A-II and serum amyloid A proteins were 253 decreased. In a very recent study [14], serum samples from cows were collected on days -3, 0 and 254

+3 relative to calving. According to the Ca serum concentrations the animals were classified as 255 256 either healthy or having clinical or subclinical hypocalcaemia. Using samples from day -3, DIGE and MALDI-TOF MS were used to search for proteins suitable as predictors for postpartum 257 hypocalcaemia. Five proteins were differentially regulated when comparing cows that developed 258 clinical milk fever or stayed healthy: Vitamin-D binding protein precursor, paraoxonase, 259 apolipoprotein A-IV precursor and alpha-1-antitrypsin were decreased, and A2M protein was more 260 261 abundant in cows with clinical hypocalcaemia post partum. There was no overlap in these proteins when comparing healthy cows with those that developed subclinical hypocalcaemia later: compared 262 to healthy animals, complement C4 precursor, A2M protein, endopin 1 and haptoglobin were 263 264 decreased in cows with subclinical hypocalcaemia, and no protein was found to be increased [14]. The issue of hypocalcaemia was also addressed by assessing the metabolome in serum samples 265 from hypocalcaemic versus normocalcaemic cows yielding around 30kg milk/day: using a 500-266 267 MHz digital (1)H-NMR spectrometer, nine metabolites with differing concentrations between the groups were found [55]. Glucose, alanine, glycerol, phosphocreatine, and γ -aminobutyrate (GABA) 268 were decreased, and β -hydroxybutyrate (BHB), acetone, pyruvate, and lysine were increased in 269 270 cows with milk fever. The increase of pyruvate is probably also related to the decrease of phosphocreatinine observed in cows with milk fever, since elevated pyruvate decreases the 271 272 production of phosphocreatinine, by inhibiting creatinine-pyruvate kinase at least in humans [56]. The decrease of phosphocreatine also reduces ATP (adenosine triphosphate) production in muscles, 273 may partially explain the paresis, ataxia and paralysis that are associated with milk fever. In 274 275 addition, the decrease of the inhibitory neurotransmitter GABA may also account for the neurological symptoms of the disease, e.g. depression and coma. However, the importance of 276 277 GABA in the circulation is unknown.

Results provided by proteomics applied to hypocalcaemia are somehow contradictory: for example,
in one study albumin abundance is increased [53], whereas in another the albumin abundance is
decreased [54]. This apparent inconsistencies might be related to the fact that results were obtained

following two different proteomics techniques (2D-DIGE separation followed by identification with 281 MALDI-TOF-MS and SELDI-TOF, respectively), rather than to different sampling times (in both 282 studies sample collection was close to delivery (6 and 24 h, respectively). In view of positive APP, 283 such as SAA, haptoglobin, complement C4 precursor and alpha-1-antitrypsin being decreased in 284 hypocalcaemic cows, hypocalcemia might also be related to systemic inflammation. Metabolomics 285 results are probably more interesting: taken together, the findings reported by metabolomic analyses 286 287 indicate a relationship between hypocalcaemia and energy metabolism, rather than a specific association with Ca metabolism. Although limited to one study [57], metabolomics confirmed in 288 cows what has been already reported in humans, i.e. that calcium plays a pivotal role in regulating 289 energy homeostasis. One important finding in this context is that by increasing the Ca²⁺ 290 concentration in adipocytes, adipogenesis and a coordinated inhibition of lipolysis were stimulated, 291 292 thus demonstrating that Ca is capable of regulating adiposity [58,59]. In dairy cows in which 293 hypocalcaemia was experimentally induced by intravenous infusion of ethylene glycol tetraacetic acid (EGTA), a selective Ca-chelator, reduced blood concentrations of insulin and increased levels 294 295 of glucose and NEFA were reported, together with reduced phagocytotic and oxidative burst 296 activity of neutrophils [60]. Even though some of the observed effects in this study might have partly been caused by the reduced feed intake during EGTA infusion, the findings are in line with a 297 298 role of Ca in energy metabolism. Fig. 4 summarizes the differences in metabolome and proteome between healthy and hypocalcaemic cows. Future omics studies could allow to find possible 299 explanations for the contradictory results stated above and deepen the insight into the relationships 300 301 between hypocalcaemia and energy metabolism.

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303 Metabolic diseases related to energy metabolism:

304 During the transition period, nutrients need to be directed towards the growing foetus and the
305 mammary gland even though feed intake is often depressed around calving and does not increase as
306 does milk yield. To accomplish an adequate supply of nutrients to foetus and mother, several

adaptive mechanisms are activated. The main ones are: increasing gluconeogenesis, reducing 307 308 peripheral insulin sensitivity and increasing lipolysis. As outlined below, these reactions may also overshoot and result in the most common metabolic production diseases, i.e. in ketosis and fatty 309 liver. Ruminants almost entirely depend on gluconeogenesis since glucose from plant carbohydrates 310 hardly reaches the small intestine due to fermentation in the forestomaches which yields propionate 311 as the main gluconeogenetic substrate. In particular the mammary gland has a high demand for 312 313 glucose to produce lactose, the major osmole in milk. In contrast to other organs, glucose uptake of the mammary gland is insulin-independent and by decreasing the insulin-sensitivity in skeletal 314 muscle and adipose tissue, glucose can be drained towards the mammary gland [61]. The increase 315 316 in lipolysis provides fatty acids as energy substrates but also for milk fat synthesis; albeit the contribution of fatty acids from the mobilization of body fat is normally less than 10% of the milk 317 fatty acids, this share increases proportionally in early lactation with the extent of the energy deficit 318 319 [62]. When the rate of lipolysis exceeds the capacity of the liver, fatty liver and ketosis (or acetonaemia) can occur. With the importance of lipolysis, the central role of adipose tissue comes 320 321 into play and indeed many studies including proteomics and metabolomics investigated adipose tissue in context with peripartum diseases. Oxidation of fatty acids provides acetyl-CoA which is 322 than condensed with oxaloacetate to form citrate for entering the TCA cycle. However, when 323 324 glucose requirements are high, oxaloacetate is increasingly used for gluconeogenesis and thus acetyl-CoA cannot be completely oxidized but is converted into ketone bodies, mainly acetone, 325 acetoacetate and BHB. Ketosis, in particular subclinical ketosis, is a common disease in dairy cows 326 327 and often concurs with other peripartum diseases such as retained placenta and metritis [63]. Excess fatty acids can also be re-esterified in the liver and deposited as triglycerides; however, their export 328 329 into the circulation is limited based on the low intrinsic capacity for mainly VLDL (very low density lipoprotein) in ruminants [57,64]. Fatty liver is thus another production disease affecting 330 many animals at least in mild forms. Taken together the main adaptations to accomplish partitioning 331

of nutrients towards foetus and milk are basically known, including the temporal patterns of someproteins and metabolites during the transition period [65].

However, what makes these physiological adaptive mechanisms shift towards pathological conditions is largely unknown. Using proteomics and metabolomics provides new explanatory approaches and also potentially also predictors for unfavourable conditions which might be mitigated if diagnosed early enough. For the latter applied aspects, the use of body fluids, in particular those that can be collected non-invasively like milk, is certainly preferable; however, including those tissues that are the major players in energy metabolism, i.e. liver and adipose tissue, is necessary for clarifying the pathways included and their complex interrelationships.

341 Fatty liver: The fatty liver observed in dairy cows has many similarities with non-alcoholic fatty liver disease (NAFLD) in humans. Several proteomic and metabolomics studies were 342 published about NAFLD [66–68]. Yet the respective literature on fatty liver in dairy animals is 343 344 surprisingly limited. To the best of our knowledge there is only one review about fatty liver proteomics in farm animals, but it mainly focused on poultry [69]. In dairy cows a proteomic 345 346 analysis was carried out in liver obtained from animals (1st lactation, 16-201 DIM) fed ad libitum as compared with feed-deprived cows [70]. Proteins were separated by 2-DE, and those that were 347 differently regulated were identified by MALDI-TOF. Several pathways related to lipid and to 348 349 carbohydrate metabolism were found to be dysregulated. Acyl-CoA dehydrogenase and Acyl-CoA acetyltransferase 2 were both down-regulated in feed-deprived animals, suggesting decreased fatty 350 acid degradation and contributing to explain the insurgency of liver disease. The fatty acid binding 351 352 protein 1 was also found to be decreased. Other enzymes involved in fatty acid degradation that were decreased in fed-deprived cows include aldehyde dehydrogenase, which converts fatty acids to 353 their corresponding aldehydes. On the contrary, sterol carrier protein 2, which catalyzes the 354 transfer/exchange of cholesterol and phospholipids between membranes, was more abundant in 355 feed-restricted cows suggesting an increase in lipid trafficking. The decrease of peroxiredoxin-6, 356 whose main role is to protect against oxidative damages, suggests a possible increase in oxidative 357

stress in the ruminant liver during feed restriction. With regard to carbohydrate metabolism, several 358 359 enzymes, including 6-phosphofructokinase, enolase1, and triosephosphate isomerase, fructosebisphosphate aldolase B, sorbitol dehydrogenase and aldehyde dehydrogenase 2 were less abundant 360 in feed-deprived versus ad libitum fed cows. The corresponding up-regulation of parathymosin, an 361 inhibitor for glycolytic enzymes, confirms a reduction of glycolysis [71]. Besides, the results 362 indicated that protein metabolism was also affected by feed deprivation: proteins involved in 363 364 protein degradation, such as ubiquitin carboxyl-terminal esterase L3, proteasome 26S subunit, and protein disulfide-isomerase-related protein 5 were decreased. This result was unexpected, since feed 365 restriction is believed to result in increased protein degradation. The authors suggested that 366 367 downregulating proteins involved in protein degradation might help to protect the liver form excessive autophagy. In addition, skeletal muscle rather than liver is the greatest labile source of 368 amino acids for energy needs [72]. However, other "protecting" proteins, such as heat-shock 70 369 370 kDa protein 5 (HSPA5), a chaperon, were less abundant in liver of feed-restricted cows. The urea cycle in particular was dysregulated: proteins, such as arginase-1 and argininosuccinate synthetase, 371 372 were up-regulated in feed restriction. Conversely, L-arginine:glycine amidinotransferase and glutamate dehydrogenase 1, were decreased. Finally, proteins involved in calcium metabolism, such 373 as regucalcin, annexin IV and calcium binding protein SPEC 2D were increased in the liver of feed-374 375 restricted cows. In 2012, a comparison of the liver proteome of cows with either low or high liver triglyceride (TG) content in early lactation was published [57]. A high liver TG content was found 376 to be associated with increased oxidation of saturated fatty acids, oxidative stress, and urea 377 378 synthesis and decreased oxidation of unsaturated fatty acids, but not with impaired gluconeogenesis. Aiming to identify hepatic biomarkers for physiological imbalances (PI), the liver proteome of dairy 379 380 cows at early and mid lactation (49±22 DIM, average milk yield 42±7 kg/day versus 159±39 DIM, 29±7 kg/day) was determined by means of ITRAQ-based profiling; PI was calculated based on 381 plasma free fatty acids, BHB, and glucose concentrations and was compared between 6 cows with 382 greatest and least PI in early and mid lactation, respectively. PI was increased by a 4 day feed 383

restriction period and liver biopsies were collected one day before and on day 3 of the feed 384 restriction [16]. In early lactation, enzymes involved in gluconeogenesis and \Box -oxidation, such as 385 pyruvate carboxylase and very long chain specific acyl-CoA dehydrogenase, respectively, were 386 increased in cows with a higher PI indicating increased gluconeogenesis and fatty acid oxidation. In 387 addition, three enzymes involved in energy metabolism, such as mitochondrial isocitrate NADP+ 388 (nicotinamide adenine dinucleotide phosphate)-dependent dehydrogenase, glycine N-389 390 acyltransferase and UDP-glucose 6-dehydrogenase were decreased, partially explaining the molecular background of PI. By increasing nutrient restriction, thus aggravating the status of PI, the 391 increasing demand of energy coupled with a decrease of anti-oxidant defense is confirmed by an 392 393 upregulation of mitochondrial trifunctional protein, subunit α , enoyl-CoA hydratase and pyruvate carboxylase and downregulation of glutathione S-transferase Mu 1, manganese superoxide 394 dismutase, aldehyde oxidase, and glycine N-acyltransferase. Proteins related to apoptosis (14-3-3 395 396 protein β/α), and mobilization and targeting of fatty acid (liver fatty acid binding protein) were also decreased. 397

During mid lactation, before feed restriction, PI increased the amount of proteins involved in ketone 398 biosynthesis, such as alcohol dehydrogenase 4 and alcohol dehydrogenase NADP+, and in TCA 399 cycle, including dihydrolipoamide dehydrogenase 2 and methylmalonate-semialdehyde 400 401 dehydrogenase. Again, proteins involved in antioxidant defense and CO₂ transport, such as carbonic anhydrase 3 were decreased. After increasing PI by means of nutrient restriction, the upregulation 402 of proteins involved in fatty acid oxidation, such as acetyl-CoA oxidase 2, acyl-CoA-binding 403 404 protein and carnitine O-palmitoyltransferase 2 was even more pronounced. Proteins involved in ketone biosynthesis, such as acetyl-CoA acyltransferase were increased as well. Consistent with 405 changes related to PI during early lactation, the experiment amplification of PI decreased proteins 406 involved in antioxidant defenses, such as peptide methionine sulfoxide reductase, Glutathione S-407 transferase A1, and carbonic anhydrase 3. Proteins involved in amino acid metabolism, such as 408 peptide methionine sulphoxide reductase and ornithine carbamoyltransferase, and fatty acid 409

oxidation, such as short-chain specific acyl-CoA dehydrogenase and hydroxyacyl-CoA
dehydrogenase, were also downregulated. Beside the identification of potential biomarkers for
different degree of PI, the study of Moyes and coworkers [16] provided a better understanding of
the molecular background of liver diseases during PI, as shown by the downregulation of proteins
related to anti-oxidant defense, which might be responsible for the increasing cellular damage. For
aldehyde dehydrogenase and HSP70, relationships with NAFLD in rats and in humans,

416 respectively, were identified earlier [73,74].

Metabolomics approaches were also applied to study the pathogenesis of and the search for 417 biomarkers in fatty liver disease. A recent review highlighted the state of the art in human and 418 419 laboratory animal metabolomics investigations on fatty liver diseases [75]. Several studies addressed the modifications induced by the development of fatty liver disease in dairy ruminants by 420 means of metabolomic techniques. A serum metabolomic profile using a triple quadrupole 421 422 spectrometry identified a total of 29 metabolites which allowed to discriminate healthy cows from animals with hepatic lipidosis [23]. The experimental design included animals with different ranges 423 of hepatic lipidosis, ranging from low, medium to severe grade. Some animals also displayed, in 424 addition, other diseases, such as displaced abomasum, bronchopneumonia, retained placenta, and 425 mastitis. Six phosphatidylcholines were identified as promising predictive biomarkers of hepatic 426 427 lipidosis. The other discriminating metabolites included amino acids, such as glycine and glutamine, sphingomyelins and hydroxy-sphingomyelins and other phosphatidylcholines, but could 428 only discriminate the three unhealthy groups from the healthy animals. Beside their possible use as 429 430 biomarkers, the finding that the phosphatidylcholine asset is modified during lipidosis is interesting: phosphatidylcholines are precursors of hepatic triacylglycerols [76], and can decrease peripartum 431 due to an increased triacylglycerol production [77-79]. Assembly and secretion of lipoproteins 432 require the contribution of phosphatidylcholines. By limiting the hepatic synthesis of lipoproteins, 433 which export fatty acids from hepatocytes, the decrease of phosphatidylcholine content may 434 aggravate the accumulation of lipids in the liver [80]. 435

A parallel study focusing on the plasma lipidome also drove to similar conclusions [81]. The 436 437 investigation was carried out on animals with different grades of fatty liver, displaying weak, medium and severe disease. The lipid extracts were profiled by means of separation with ultra 438 performance LC and identification with LC-MS. The study confirmed that phosphatidylcholines are 439 reduced in animals with medium and severe fatty liver disease. Five bile acids were decreased as 440 related to increased severity of fatty liver. Remarkably, the insurgency of fatty liver diseases was 441 442 positively correlated to the presence of Resolvin E1 and palmytoil-ethanolamine. Resolvin E1 is synthesized from ω -3 polyunsaturated fatty acids (PUFAs) such as eicosapentaenoic acid (EPA) and 443 docosahexaenoic acid (DHA), and is an anti-inflammatory lipid mediator[82,83][82] [83]. Also 444 445 palmytoil-ethanolamine is an endogenous amide with anti-inflammatory activity [84]. It is known that metabolic diseases such as fatty liver also induce an inflammation [85]. The study of Gerspach 446 and coworkers [81], therefore, confirmed that anti-inflammatory pathways are activated during fatty 447 448 liver disease. One estrogen metabolite, not further specified, was found to discriminate between mild or strong and weak fatty liver in the same study. 449

450 A novel approach relying on (1)H-NMR investigated the metabolome in fatty livers of 171 Holstein cows [86]. The main advantage of this technique in the metabolomic field is that it can provide a 451 profile of proton-containing, low-molecular-weight metabolites, starting from a limited amount of 452 453 sample [87]. Plasma and liver tissue samples from animals with fatty liver disease (16.31 ± 4.30) DIM, average milk yield 28.32±4.73 kg/day) and healthy animals (15.5±6.02 DIM, average milk 454 yield 30.75±3.7 kg/day) were included in the study. As expected, plasma from animals with fatty 455 456 liver disease had increased BHB, isobutyrate and acetone concentrations. The amino acids glycine, valine, trimethylamine-N-oxide, and citrulline were also increased. Conversely, other amino acids 457 458 such as alanine and asparagine were decreased. Glucose, GABA, glycerol, and creatinine were decreased as well. The decrease of alanine and asparagine is consistent with their role as 459 gluconeogenic amino acids, since both can enter the TCA cycle to generate glucose during an 460 energy-deficient status. The finding that trimethylamine-N-oxide and citrulline are increased what is 461

usually accompanied by oxidative stress and liver damage, corresponds to what has beendemonstrated in other species, such as mice [88] and humans [89].

Ketosis: to extend the understanding of the pathogenic effects of ketosis, protein 464 modifications were determined by using 2-DE coupled with MALDI-TOF for comparing the 465 proteomic profiles in liver from healthy and ketotic cows [90]. Several metabolic pathways were 466 found to be altered in ketosis. Structural proteins, such as myosin related proteins (myosin light 467 chain for example, and tropomyosin) and MGC128326 were significantly up-regulated in liver from 468 cows with ketosis. As shown by the increased number of isoforms, post translational modifications 469 related to ketosis were assumed. Myoglobin was also more abundant, suggesting an activation of 470 471 oxidative stress defense pathways, which is confirmed by the upregulation of proteins belonging to the peroxired oxin families, such as peroxired oxin -5 and -6, glutathione S-transferase alpha-1, 472 flavin reductase and sulfotransferases, all of them fulfilling anti-oxidant activities. As expected, 473 474 proteins related to gluconeogenesis, such as α -enolase, were also increased during ketosis. The list of proteins that were decreased in ketotic liver included also proteins involved in fatty acid 475 476 oxidation, such as acetyl-CoA acetyltransferase 2 and 3-hydroxyacyl-CoA dehydrogenase, suggesting a further possible relationship between ketosis and lipidosis: as a consequence of the 477 decreased ability to utilize them, fatty acids accumulate in liver cells, contributing to the 478 479 development of hepatic lipidosis.

Beside liver and plasma, the difference of proteomic profiles in animals with ketosis were also 480 explored in urine using SELDI-TOF techniques [91]. Samples were collected 7 - 28 DIM from two 481 482 groups of animals, one affected by clinical ketosis and the other from healthy cows, with a milk production of 9625 kg/year. The urinary profile of animals with ketosis showed a decrease for 11 483 proteins, most of them involved in the inflammatory response, such as fibrinogen, C1 inhibitor, 484 osteopontin, also hepcidin and human neutrophil peptides 1-3. Interestingly, also proteins associated 485 with the neuron function, such as VGF (non-acronymic) protein and amyloid precursor protein, 486 were decreased in the animals with ketosis as compared with the control group. Proteins related to 487

lipid metabolism, as well as inflammation, were also decreased during ketosis, namely SAA and 488 489 apolipoprotein C-III, indicating a change in lipid metabolism during ketosis. Two other proteins, i.e. transthyretin, a transport protein, and cystatin C, a protease inhibitor, were also decreased. 490 Fibrinogen, hepcidin and SAA are acute phase proteins [92] and their concentration is supposed to 491 increase during an inflammatory status. Targeted assessments of acute phase proteins in blood have 492 shown increased concentrations of inflammatory markers in ketotic cows. Thus the finding of 493 494 decreased concentrations of SAA in urine of ketotic cows is opposing the situation in blood and deserves further investigation. The proteins differentially regulated in liver and urine of cows with 495 ketosis as compared to healthy animals are presented in Fig. 5. 496

497 More than proteomics, it is metabolomics that contributed to address the changes in biological fluids during ketosis. A NMR-based metabolomic analysis on milk from animals with ketosis was 498 carried out during a time course study on 264 high yielding dairy cows with an average milk yield 499 500 of 32.8 ± 4.7 kg energy corrected milk/day [93]. Milk samples were collected weekly for 5 weeks and once again 6 months post partum. NMR spectroscopy was carried out, and presented evidence 501 that high milk glycerophosphocholine (GPC) levels and high ratios of GPC to phosphocholine (PC) 502 during the first four weeks of lactation, and GPC at mid lactation, could provide reliable biomarkers 503 for the development of ketosis. Although milk provides an ideal substrate for metabolomic analysis, 504 505 being collected routinely and non-invasively, more studies were carried out on plasma metabolome, which, conversely, yields a higher amount of potential biomarker metabolites. A metabolomic 506 approach using GC/MS techniques analysed the differential plasma metabolomes of dairy cows 507 508 with clinical (12 ± 5 DIM, 32.1 ± 7.8 kg milk/day) and subclinical ketosis (14 ± 6 DIM, 35.0 ± 7.2 kg milk/day), as compared with healthy animals (16 ± 6 DIM, 37.0 ± 6.2 kg milk/day) [94]. The 509 510 study revealed that the metabolomes of animals with subclinical and clinical ketosis were mostly identical, whereas 25 potential biomarkers were found between animals with ketosis, both clinical 511 and subclinical, when compared with healthy animals indicating that several biochemical pathways 512 were modified. The nine metabolites decreased during ketosis suggested a decrease in 513

gluconeogenesis in affected animals, and a parallel activation of the pentose-phosphate pathway. A 514 515 decrease of ribitol levels, related to riboflavin deficiency, and vitamin C, may increase the oxidative stress. The decrease of lactic acid and L-alanine, which are both gluconeogenetic precursors, 516 suggested a close relationship between ketosis and carbohydrate metabolism, as a consequence of 517 hypoglycaemia and lack of precursors of gluconeogenesis. Sixteen metabolites were increased in 518 animals with ketosis. As expected, this list included ketone bodies, and fatty acids, such as BHB, 519 520 palmitic acid, heptadecanoic acid, stearic acid, trans-9-octadecenoic acid, myristic acid, cis-9hexadecenoic acid, confirming also the mobilization of adipose tissue. Amino acids were also found 521 to be increased, namely L-isoleucine and a catabolic product of lysine, 2-piperidinecarboxylic acid, 522 523 two amino acids involved in ketogenesis, and glycine, suggesting an increase of proteolysis needed to fuel gluconeogenesis. The study confirmed BHB acid as gold biomarker for ketosis, and 524 suggested cis-9-hexadecenoic acid as novel biomarker for clinical ketosis, and an indicator of fat 525 526 mobilization. Plasma metabolic profiling from cows affected by clinical ketosis was determined with two other different techniques, namely LC/MS and (1)-NMR. In a study whose experimental 527 design was very similar to that of Zhang and coworkers [94] plasma metabolomics of cow with 528 clinical and subclinical ketosis was determined by means of 1H-NMR [95]. A total of 25 529 metabolites were found to be dysregulated as a consequence of various stages of the disease, and as 530 531 compared with healthy animals. Amino acids including histidine, glutamic acid, glutamine, lysine and phenylalanine, together with lactate and glucose, were decreased during ketosis. Amino acids 532 such as alanine, proline and tyrosine decreased only in clinical ketosis, as well as LDL and VLDL. 533 534 As expected, metabolites related to biosynthesis of ketone bodies, such as BHB, N-Acetylglucosamine, acetate, acetoacetate and acetone were increased in ketotic cows. Glycine, 535 536 leucine, isoleucine and valine were increased in clinical ketosis only. A last study, this time focused on the differences in metabolomes between Holstein Friesian cows, 12 - 16 DIM and 18 - 21 kg 537 milk /day, affected with clinical ketosis or being healthy, was carried out by means of LC/MS [96]: 538 aminoacyls such as valine, glycine, and lipids, such as glycocholic, tetradecenoic and palmitoleic 539

acid, were increased in clinical ketosis, whereas other amino acids, such as arginine, leucine,
isoleucine, tryptophan and lysine decreased. Aminobutyric acid, creatinine, undecanoic acid and
norcotinine were decreased as well. Consistent with previous reports, obtained with different
metabolomic techniques, the study from Li and coworkers [96] confirmed that amino acids are
affected during ketosis: e.g. glycine was increased whereas lysine was decreased.

545

Pathophysiological alterations in the proteome and metabolome of adipose tissue related to the transition period

Adipose tissue fulfills a dual role: it regulates energy storage by storing and releasing fatty acids, and is a major endocrine gland, capable of modulating metabolism by secreting hormones and adipokines [97]. As mentioned earlier, the transition from late pregnancy to early lactation is accompanied by an increased rate of lipolysis. Furthermore, adipose tissue can regulate, by means of modifying its adipokine asset, the development of major metabolic changes such as insulin resistance or sensitivity [7,61,98].

554 The proteome of subcutaneous adipose tissue from cows classified as either insulin-resistant or insulin-sensitive according to phosphorylation of protein kinase B (AKT) in this tissue was 555 characterized by quantitative shotgun proteomics (nanoLC-MS/MS) [99]. Adipose tissue biopsies 556 were collected 17 days before and 3-5 days after calving. From 586 proteins detected, 143 were 557 differentially regulated in prepartum versus postpartum tissue. Several functions, such as those 558 related to lipid metabolism, including fatty acid metabolism, the esterification of lipids and 559 560 oxidation of fatty acids, appeared as changed. The proteins whose abundance was decreased after calving included fatty acid synthase, complement C3, annexin-A1 and acyl-CoA desaturase. 561 562 Comparing insulin-resistant and insulin-sensitive subcutaneous adipose tissue yielded 111 proteins that were differentially regulated. Most of them (a total number of 106) were more abundant in the 563 insulin-resistant state, whereas only five were decreased. Insulin resistance was associated with a 564 dysregulation of pathways related to energy and lipid metabolism, including gluconeogenesis and 565

glycolysis, signaling mediated by 14–3–3, TCA cycle, ERK/MAPK signaling, lipid accumulation,
release and lipolysis. Inflammatory responses, such as leukocyte migration and proliferation of T
lymphocytes, were also activated in insulin-resistant adipose tissue.

During the transition period, adipose tissue may also react to stress related to environmental factors, 569 such as heat. A label-free, quantitative shotgun proteomics (nano-LC-MS/MS) approach 570 investigated the effects of seasonal heat stress on the adipose proteome, aiming to highlight 571 572 biomarkers of heat stress on late pregnant cows during summer heat stress (average milk yield 33.8 kg/day) as compared to the winter season (38.2 kg/day) [100,101]. The proteome in subcutaneous 573 adipose tissue biopsies obtained 14 day before calving yielded a total number of 107 out of 1495 574 575 proteins identified that were differentially abundant between summer and winter. The pathways that were found to be modified included the Keap1-Nrf2 pathway which is the major regulator of 576 cytoprotective responses to oxidative and electrophilic stress [102], such as STIP1 and ubiquitin-577 578 conjugating enzyme E2 K, which were increased in summer, and GSTM1, microsomal GST 1, (MGST1), GST Mu 3 (GSTM3), ferritin heavy chain and MAP2K1 which were decreased. The 579 580 acute phase response was also modified. In particular, albumin, hemopexin, serotransferrin, AGP, apolipoprotein A-II, α-2-HS-glycoprotein, C-reactive protein and MAP2K1 were decreased in 581 summer, whereas the abundance of the von Willebrand factor and fibrinogen α chain was increased. 582 583 Protein related to the farnesoid X receptor (FXR)[103], which is a member of the nuclear family of receptors in control of numerous metabolic pathways, and, jointly with retinoid X receptor (RXR), 584 plays a crucial role in linking bile acid productions with lipoprotein, lipid and glucose metabolism, 585 were also modified, as well as proteins belonging to Liver X receptor/RXR pathways [104], whose 586 function is to regulate cholesterol, fatty acid, and glucose homeostasis. The finding of this study 587 provided the evidence that heat stress has a local impact on adipose tissue in late pregnant cows, 588 highlighting meanwhile a list of possible biomarkers for heat stress related to transition period. 589

591 Pathophysiological alterations in the proteome and metabolome of skeletal muscle related to 592 the transition period

Several studies point to the importance of amino acid metabolism in context with the 593 pathophysiological changes related to the metabolic challenges of the transition period. An 594 increased need for amino acids results from fetal growth and milk protein synthesis but is also 595 related to the use of amino acids for generating energy by direct oxidation or as precursors for 596 gluconeogenesis. The dogma that amino acids are significant contributors to hepatic 597 gluconeogenesis has recently been revised based on quantitative data on the uptake of amino acids 598 by the liver; only alanine remains in the list of quantitatively important gluconeogenetic amino 599 600 acids [105]. However, there is also an increased need for amino acids for positive acute phase proteins [106–108] which show a distinct and typical peak around calving [39]. The biggest labile 601 source for amino acids in the body is skeletal muscle, but the number of proteomics or 602 603 metabolomics studies actually addressing this tissue in context with transition cow is very limited. Kuhla and coworkers [105] used 2-DE and MALDI-TOF-MS on muscle biopsies collected in week 604 -3, 0, +2 and +4 relative to calving. In total 43 differentially regulated muscle protein spots were 605 identified throughout the periparturient period. In early lactation, abundance of cytoskeletal proteins 606 and enzymes involved in glycogen synthesis and in the TCA cycle was decreased, whereas proteins 607 608 related to glycolysis, fatty acid degradation, lactate, and ATP production were increased. Metabolomic investigations of skeletal muscle in transition cows are only at the verge of being 609 published: in view of several abstracts presented at the International Animal Science and Dairy 610 611 Science meeting in 2016 and 2017, several publications on this topic can be expected [109–111]. 612

Associations of proteomes and metabolomes with productive life span, energy balance and feed regulation

Several studies focusing on the metabolic situation during the transition period in general ratherthan on specific production diseases also contributed to our understanding on the pathophysiology

of the underlying adaptive responses. Aiming to find biomarkers for any kind of production 617 618 diseases relevant for transition dairy cows, a quantitative targeted metabolomics approach was used [21]. Plasma collected at 4 time points from 4 weeks prepartum to 4 weeks post partum from 12 619 cows of which 6 developed multiple peripartum diseases, including laminitis, mastitis, metritis and 620 retained placenta whereas the other 6 cows remained healthy was compared. The study identified 621 five plasma metabolites that could be related to periparturient diseases: carnitine, valerylcarnitine, 622 623 propionyl carnitine, lysophosphatidylcholine acyl C18:2 and lysophosphatidylcholine acyl C14:0 were increased in animals that developed peripartum related diseases as compared with healthy 624 controls as early as 4 weeks before parturition. Two phosphatidylcholines, namely 625 626 phosphatidylcholine acyl-alkyl C42:4 and phosphatidylcholine diacyl C42:6, were increased 1 week before delivery. Carnitine, lysophosphatidylcholine acyl C18:2 and lysophosphatidylcholine acyl 627 C14:0 were increased 1 week postpartum as well, whereas carnitine was decreased after 4 weeks. 628 629 These results are remarkable, since they highlighted the possible use of three metabolites, namely carnitine, propionyl carnitine, and lysophosphatidylcholine acyl C14:0 as potentially predictive of 630 peripartum-related diseases up to 4 weeks before delivery [21,96]. 631 632 The occurrence of production diseases exerts profound effects on productive life span since continued disorders like decreased fertility may result which in turn give reason to premature 633 634 culling. Even in case of non-ouvert disease, a metabolic predisposition for the risk of leaving the herd prematurely might exist. Huber and colleagues [22] applied targeted metabolomics but also 635 "classical" variables (e.g. insulin, free fatty acids, BHB) to search for factors predisposing for 636 637 shorter or longer productive lifespan in 19 cows that remained apparently healthy during the first 100 days of lactation. Eight of these cows left productive life within the current lactation due to 638 various health and fertility problems and 11 cows finished the current lactation without any signs of 639 clinical illness. Long-chain acylcarnitines and biogenic amines were found to be associated with 640 extended productive life span. These metabolites are mainly secreted by the liver and depend on the 641 functionality of hepatic mitochondria. The concentrations of biogenic amines and some 642

acylcarnitines differed already before the onset of lactation thus indicating their predictive potentialfor continuation or early ending of productive life.

Using milk samples from cows with great differences in energy balance, achieved by varying the 645 length of the dry period, Lu and coworkers [112] applied untargeted metabolomics and proteomics 646 techniques, i.e. as NMR in milk serum and milk lipids as well as FASP Dimethyl Labeling-647 NanoLC-Orbitrap-MS/MS on milk fat globule membrane proteins. They found that a severely 648 649 negative energy balance was related to greater concentrations of acute phase response proteins, unsaturated fatty acids, and galactose-1-phosphate. In contrast, the concentrations of cholesterol, 650 cholesterol synthesis-related proteins, and stomatin were increased in improved energy balance. The 651 652 appropriateness of using milk proteomic and metabolomic data to draw conclusions not only about milk quality but about the individual cow's health situation is of outstanding importance. Using 653 654 NMR techniques in routine milk recordings is aimed at applying such information in herd health 655 management programs or for breeding purposes. Maher and co-workers [113] compared the metabolic profiles from blood and from milk samples obtained from Holstein cows via 1H NMR 656 methods and statistical heterospectroscopy. The authors summarized their results as being 657 confirmative for milk being a distinct metabolic compartment with a metabolite composition largely 658 not influenced by plasma composition under normal circumstances. Similarly, Ilves and coworkers 659 660 [114] reported that there is only little correlation between the composition of the metabolomes in milk and in blood. However, the group of Maher [113] found trimethylamine and dimethylsulfone, 661 both originating from rumen fermentation, being correlated across both body fluids, indicating that 662 663 measuring these substances in either body fluid might allow to evaluate rumen function. Taken together, omics techniques provide a detailed view on a great number of metabolites or proteins and 664 thus enable to also consider additional factors previously not considered. The knowledge of the 665 factors influencing or indicating the metabolic situation and productive lifespan of dairy cows can 666 thus be deepened. Nevertheless, extrapolating results from a given experimental design, body fluid 667

or tissue, and performance level of cows likely has its limitations when conditions e.g. timing ofsamples relative to physiological status, are different.

670

671 **Proteomics during laminitis.**

Laminitis (Pododermatitis aseptica diffusa), also known as sole-haemorrhage, is an inflammation of 672 the laminar corium of the hoof. Cow laminitis accounts for 41% of cases of lameness [115] and is 673 674 most prevalent around day 50 to day 100 of lactation [116]. It is thus not a typical disease for early lactation as the other diseases included in this review, but the course of the preceding reactions 675 during the transition period might form predisposing elements for developing laminitis. Related to 676 677 laminitis are several claw horn lesions, such as white line disease and ulcers of the sole. Metabolic diseases, in particular acidosis, both clinical and subacute ruminal acidosis (SARA) are regarded as 678 underlying cause of laminitis [117], although the pathogenesis is not fully understood. The digestive 679 680 disorder SARA is found in up to 19% of early lactation dairy cows as well as 26% of mid-lactation cows, and is related to diets with high portions of concentrate. Increasing the portion of concentrate 681 is a common strategy to improve the energy supply for cows, but there is a risk for shifting the 682 rumen microbiota toward increased lactate production and thus acidification of rumen content 683 which may result in increased histamine secretion and thereby also affect the claw capillaries [118]. 684 685 However, the causes of laminitis and associated claw horn lesions are multi-factorial in nature [119,120]. 686

In equine medicine, laminitis can be induced by feeding and using such models several omics studies have been carried out to unravel the pathogenesis of the disease, or to look for predictive biomarkers [121,122]. As a preliminary step to unravel the changes in plasma of dairy cows affected from laminitis, a proteome analysis carried out by 2-DE coupled with MALDI-TOF identification of differentially regulated proteins between animals with spontaneously occurring clinical laminitis and healthy animals used as controls was carried out [123]. A semi-quantitative analysis of the 2-DE gels revealed that 16 proteins were differentially regulated between the two

groups of animals, of which 12 were more abundant in laminitis, and 4 were decreased, as 694 695 compared to healthy animals. Proteins involved in inflammation and in defensive mechanisms were increased during laminitis, namely complement component C9, haptoglobin and conglutinin, as 696 well as apolipoprotein A-IV, and apolipoprotein A-I, which are also involved in inflammatory 697 reaction, but also in lipid metabolism, together with 3-hydroxy-3-methylglutaryl-CoA reductase. 698 699 The group of more abundant proteins includes also zinc finger protein 300-like, transmembrane 700 protein TMP10, isocitrate dehydrogenase, and serum albumin. The upregulation of serum albumin 701 is remarkable, and apparently in contradiction with the behaviour of albumin during acute inflammation. Serum albumin is a negative acute phase protein [92,124] and, therefore, its serum 702 703 concentration decreases during acute inflammation, as it has been demonstrated also in laminitis [125]. It must be said that the upregulation of albumin is hardly demonstrable in the context of the 704 705 experimental design of the study, on the background that the samples were analyzed after depletion 706 of abundant proteins, such as albumin and immunoglobulins, to decrease the dynamic range of the samples. Besides, it has been demonstrated that albumin may be regarded as a local positive acute 707 708 phase protein during mastitis, being up-regulated by epithelial cells of the mammary gland [126]. 709 The proteins that were less abundant in plasma from animals affected by clinical laminitis include two members of complement pathways, namely C4BP, which is an inhibitor of complement, and 710 711 Complement C9 precursor, and Glycerol-3-phosphate dehydrogenase 1-like protein, and Ectodermneural cortex protein 1, which is an actin-binding protein playing a role in the oxidative stress 712 response. The number of proteomics studies on bovine laminitis is yet too limited to draw final 713 714 conclusions about the pathogenesis. Nevertheless, proteomics results confirmed the development of a pro-inflammatory loop, as demonstrated by the upregulation of inflammation-related pathways, 715 and a converse down-regulation of anti-inflammatory factors, as demonstrated by the decrease of 716 C4BP. The increase of complement C9 and the parallel decrease of complement C9 precursor 717 suggests that the complement pathway is activated, since the C9 precursor is less abundant, likely to 718

produce the active C9 protein. To the best of the knowledge of the authors, no metabolomics studieson cow laminitis were carried out to date.

721

Proteomics and metabolomics in transition period related diseases: gaps and perspectives 722 The main features of the metabolic derangements occurring in the typical production diseases of 723 724 dairy cows that are related to the transition from pregnancy to lactation are basically known since 725 several years. In particular for the metabolic diseases, overshooting lipolysis and ketogenesis were identified as aetiologic key elements. However, the knowledge of the molecular basis of successful 726 versus compromised adaptation to the metabolic challenge of early lactation is still incomplete. 727 728 Omics technologies such as proteomics and metabolomics provide important tools to close this gap in understanding the pathophysiology and also hold some promise for developing biomarkers. 729

Understanding the pathophysiology: beside transcriptomics, proteomics applied to adipose 730 731 tissue, for example, has evidently contributed to a huge advancement in the knowledge of the involvement of this tissue in the development of transition-related diseases [127]. Proteome maps 732 have been established for several biological fluids and tissues in cattle [128], and the amount of the 733 734 literature available on cow proteomics and metabolomics is steadily growing, as shown in Fig. 6. Nonetheless, although increasing exponentially during the last few years, the application of 735 736 proteomics and metabolomics in veterinary and animal science is lagging behind those in human medicine [11], and several gaps have yet to be covered. A wider application of proteomics 737 techniques in bovine peripartum-related diseases is hampered by the lack of tools to validate the MS 738 739 findings. The availability of the bovine genome [129] will allow for a very precise identification of selected proteins and is poised for closing this gap. Moreover, the complete annotation of the 740 741 genome provided a full application of bioinformatics tools to characterize the pathways where these proteins are involved in. Yet, for a biological validation of proteins identified at different abundance 742 in proteomics approaches, the options are limited to the use of antibodies in immunoassays (e.g. 743 ELISA) or in Western blotting. For both approaches, the specificity of the antibodies is limiting, 744

and species determined differences in the amino acid sequence might hamper the applicability of 745 746 antibodies developed against the target protein in different species. Besides, the performance of antibodies in different methodological set-ups can differ. The quantitative power of immunoassays 747 like ELISA is usually good, but developing a valid ELISA system is laborious and time consuming. 748 In Western blotting, the effort of setting up a system with a working antibody is often considered as 749 750 faster, but when aiming to work quantitatively, comprehensive validation is also required and 751 determining differences as low as 2 to 4-fold may remain impossible [130]. In view of these 752 limitations, assessing the matching mRNA concentration, preferably from identical sample for validating results from proteomics is often used as "biological validation". However, the 753 754 abundance of mRNA may not correspond to the abundance of the respective protein [131] and therefore relying on the direct, *absolute* correlation between protein and mRNA levels is hardly an 755 adequate validation measurement [132]. Nevertheless, albeit being far from perfect, in many cases 756 757 quantifying the mRNA abundance remains the only approach available [131], but is further limited to cells and tissues, where both mRNA and proteins are accessible. When proteomics is carried out 758 759 in biological fluids, such as saliva, urine and blood serum, only protein but not mRNA material is 760 available. Nonetheless, the growing economic interests in producing antibodies and also assays specific for various animal species, including cattle, might cover this gap in the near future. The 761 762 validation of metabolomics results is easier, there is no species issue and expectedly the comparison between results obtained via MS-based targeted metabolomics with classical assays yielded good 763 correlations. For example, in a study in which 17 free amino acids were measured in 54 dairy cows' 764 765 sera both via classical methods or via a target metabolomics approach [133] the concentrations obtained by the two methods were all correlated (P < 0.0001), with an average r-value of 0.82 ± 766 767 0.14 (mean ± SD; Dr Hassan Sadri, personal communication). In non-targeted metabolomics, the 768 ongoing improvement of the livestock metabolome database (LMBDB, available at http://www.lmdb.ca), will facilitate future untargeted metabolomics studies by increasing the 769 number of identified metabolites. The main gap to be covered includes metabolite coverage and 770

their quantification, however improving the data set and information included in the livestock 771 772 metabolome database (LMBDB, available at http://www.lmdb.ca), will facilitate future untargeted metabolomics studies. For NMR-based approaches, there is a large variability of reference methods 773 774 used for calibration, and thus standardizing the methods used within and across countries is still a major challenge [134]. Moreover, albeit the application in milk is attractive in terms of being non-775 776 invasive and combined with well-established routine assessments of macro nutrients in milk, it 777 should be kept in mind that there is only little correlation between the composition of the 778 metabolomes in milk and in blood [114].

Standardization and comprehensive reporting of experimental conditions and animal characteristics 779 780 is a basic requirement but is often incomplete, albeit respective guidelines are available, at least in laboratory animals [135]. This is a general shortcoming that, albeit not specific for proteomics and 781 782 metabolomics studies, is of particular importance for omics techniques due to the snapshot character 783 of the results generated. In large animals as dairy cows, standardization of experimental conditions and animals is close to impossible but important information such as age, lactation number and 784 785 stage, body condition, diet composition and feeding regimen as well as detailed description of sampling procedures and timing relative to physiological state are sometimes missing. Given the 786 background that the development of metabolic diseases in ruminants is strongly related to the 787 788 nutrient requirements which in turn depend mainly on the level of milk production, the need for reporting milk yield and composition for an appropriate interpretation of data is obvious. 789 Applying proteomics and metabolomics to the complex of transition period-related diseases for 790 791 elucidating the underlying pathophysiological processes is beneficial and allows for developing a 792 holistic imagination but also a faster progress in research. Diseases apparently not related, such as 793 fatty liver, hypocalcaemia, ruminal acidosis and laminitis, could be identified as being associated not only from epidemiology data but also for what concerns their molecular backgrounds. Examples 794 for newly emerging disease associated pathways from proteomics and metabolomics comprise the 795 relationship with energy status in hypocalcaemia [57], the involvement of RabGTPases in retained 796

placenta [48] or the observation of decreased cystatin concentrations in urine from ketosis-affected 797 798 cows [43]. In addition, the concept of NEFA oxidation rather than lipolysis alone influences the adaptive capability to the metabolic challenge of early lactation, was substantially supported by the 799 acyl-carnitine data from metabolomics [22,96] but also from proteomics [16]. Applying omics 800 techniques distal from transcriptomics is also particularly promising in terms of quantitative aspects 801 since the relative importance of pathways can be evaluated by the flux changes and thus may allow 802 803 for identifying molecular targets most promising for prophylactic, metaphylactic or therapeutic interventions. For making more efficient use of the analytical techniques available for 804 understanding the adaptive responses in dairy cows, a systemic approach is required. Integration of 805 806 different omics techniques, namely metabolomics, proteomics and transcriptomics in the same study, will help to produce a holistic and comprehensive interpretation of multi-omics data. Hereby 807 production data and, if available, genotype information, should be combined for providing an 808 809 integrated network of the single elements, the knowledge of which could yield a level of information higher than the sum of individual parts. It must be said that this gap may be difficult to 810 bridge, given the shortage of funding available to livestock research as compared to medical studies. 811 Nevertheless when balancing the amount of information provided from metabolomics and 812 proteomics against the one from classical assays for the different targets, the omics approaches, in 813 814 particular those where no further biological validation is required as in case of metabolomics, might nevertheless work out superior in terms of costs. 815

Development of Biomarkers: Selecting biomarker candidates from proteomes and
metabolomes that show great differences when comparing diseases cows against healthy controls
and that can be assessed in body fluids may yield predictive and diagnostic tools. However, such
biomarkers will be based on quantitative results since none of the candidates will be found in
healthy or in diseased state only. Proteomics studies do mostly not provide truly quantitative data,
but refer to trends, e.g. an increase or a decrease as compared to internal standards. Applying
proteomic techniques to metabolic diseases has not provided any robust and consistent list of

biomarkers so far, although it has shed some light into the pathogenesis of many transition-period 823 824 related diseases. More than proteomics, metabolomics contributed more to discovering potential biomarkers. As mentioned above, carnitines emerged as potential biomarkers for metabolic diseases 825 related to transition period [21,22,96,109]. When considering the application of biomarker 826 candidates it is probably more realistic to assess patterns of different metabolites rather than single 827 components. Moreover, due to the common basis of many transition period-related diseases, 828 829 specific markers for individual diseases are improbable to emerge. In addition, comprehensive biological validation is needed before such measurements can indeed be considered as assessments 830 of biomarkers for predictive and diagnostic purposes. However, the integration of several 831 832 pathophysiological aspects e.g. lipolysis, ketogenesis and oxidative capacity in such patterns, by combining fatty acids, ketone bodies AND acylcarnitines, will likely yield more information than 833 the classical measurement of NEFA and BHB. Moreover, the expansion of the dynamic range of 834 835 detection of low-abundance proteins and metabolites is likely poised to pave the way for the detection of peripartum-specific biomarker patterns. Such (complex) biomarkers, will enable the 836 identification of phenotypes less sensitive to metabolic stress and thus the development and 837 implementation of strategies for early diagnosis, prognosis and prevention of transition-related 838 diseases. 839

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- 1314 Figure legends

1315 Fig. 1: The relationship between metabolic stress and disease development.

1316 The figure presents a schematic flow of the events during the pathogenesis of peripartum related1317 diseases

1318 Fig. 2: The application of OMICS technologies to production diseases in dairy cows

1319 Fig. 3: Changes in protein abundance during endometritis.

The figure presents the differences in proteomes of blood and endometrial samples associated with
the development of metritis. The figure was drawn following the papers of Cairoli et al. [39] and
Choe et al. [41]. As indicated by the arrows, the abundance of proteins listed on the left was

increased, whereas the ones on the right were decreased when compared to healthy animals.

Abbreviations: α1-acid glycoprotein (AGP), Heat-shock protein (HSP), deoxyribonuclease-I

(DNase-I), luteinizing hormone receptor isoform 1 (LH receptor isoform 1); MHC class I heavy

- the chain (MHC-Ih), Interleukin (IL), Hemoglobin (hb), potassium channel tetramerisation domain
- 1327 containing (KCTD).

Fig. 4: Changes in protein abundance and metabolites between healthy and hypocalcaemiccows.

1330 The figure highlights the differences in metabolome and proteome between healthy and

- 1331 hypocalcaemic cows. The figure was drawn following the papers of Xia et al. [53], Wang et al. [54]
- and Shu et al. [14], for what concerns proteomics, and Sun et al. [55] for what concerns
- 1333 metabolomics. The two arrows represent an increase (arrow up, left side) and a decrease (arrow
- down, right side) of the abundance of the respective proteins.

1335 Fig. 5: Hepatic proteins differentially expressed during ketosis.

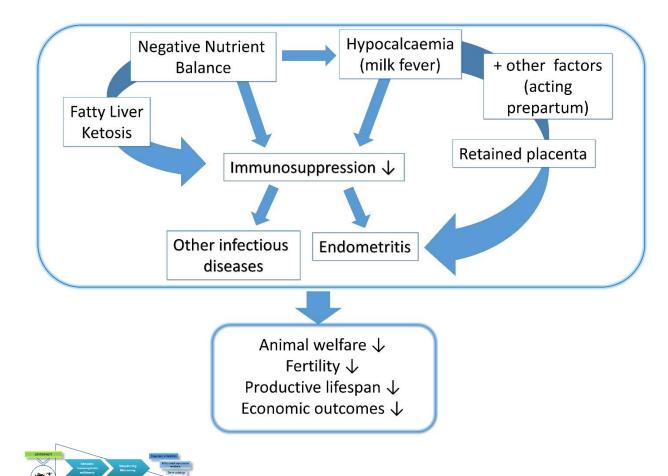
1336 The metabolic processes involved in increased formation of ketone bodies are schematically shown

1337 together with the list of proteins found to be differentially expressed in liver [90] of cows with

- 1338 ketosis as compared to healthy animals. The two arrows represent an increase (arrow up, left side)
- 1339 and a decrease (arrow down, right side) of the abundance of the respective proteins.

1340 Fig 6: Proteomics and metabolomics literature as related to bovine species.

Farm animal proteomics literature: number of manuscripts within the keyword cow proteomics andcow metabolomics from Medline up to July 2017.



1344

Blood and endometrial proteins associated with the development of endometritis



AGP (blood) Desmin α-actin-2 HSP 27 peroxiredoxin-6 LH receptor isoform 1 collectin-43 precursor DNase-I MHC-Ih



Transferrin IL-2 precursor hb β subunit KCTD11



Altered blood proteins in hypocalcaemia

141

len

TUN

- Serpin peptididase inhibitor
- Endopin 2B
- Albumin
- Fibrinogen Alpha Chain
- Amyloid beta A4 proteins
- VGF

•

A2M protein

- Fibrinogen Beta Chain
- lgG –C(H)
- Albumin

UND

- Apolipoprotein A-II
- Serum amyloid A
- Vit-D binding protein precursor
- Paraoxonase,
- Decreas

- β-hydroxybutyrate
- acetone
- pyruvate
- lysine

- Glucose
- alanine
- glycerol
- Phosphocreatine
- γ-aminobutyrate

Altered blood metabolites in hypocalcaemia

1346

Hepatic proteins differentially expressed during ketosis

