

# 1 ***Morcl* knockout evokes a depression-like phenotype in mice**

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47 **Abstract**

48

49 *Morc1* gene has recently been identified by a DNA methylation and genome-wide association  
50 study as a candidate gene for major depressive disorder related to early life stress in rodents,  
51 primates and humans. So far, no transgenic animal model has been established to validate  
52 these findings on a behavioral level. In the present study, we examined the effects of a *Morc1*  
53 loss of function mutation in female C57BL/6N mice on behavioral correlates of mood  
54 disorders like the Forced Swim Test, the Learned Helplessness Paradigm, O-Maze and Dark-  
55 Light-Box. We could show that *Morc1*<sup>-/-</sup> mice display increased depressive-like behavior  
56 whereas no behavioral abnormalities regarding locomotor activity or anxiety-like behavior  
57 were detectable. The baseline CORT plasma levels did not differ significantly between  
58 *Morc1*<sup>-/-</sup> mice and their wildtype littermates, yet – surprisingly - total BDNF mRNA-levels in  
59 the hippocampus were up-regulated in *Morc1*<sup>-/-</sup> animals. Although further work would be  
60 clarifying, *Morc1*<sup>-/-</sup> mice seem to be a promising epigenetically validated mouse model for  
61 depression associated with early life stress.

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80 **Keywords:** depression; *Morc1*; transgenic mice; early life stress; epigenetics; BDNF.

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## 83 **1 Introduction**

84

85 Clinical studies show that early life stress has profound and persistent effects on brain  
86 functions and is one of the major risk factors for developing a depressive disorder later in life  
87 [1], [2], [3]. The fundamental role of epigenetics mediating this process became clear in the  
88 groundbreaking animal study of Weaver et al. [4]: life-long epigenetic modifications of the  
89 glucocorticoid receptor gene in the rat hippocampus induced by early life stress were followed  
90 by a stable enhanced HPA response to stress. In the meantime, several studies identified  
91 effects of early life stress on DNA methylation for further genes in rodents [5], [6], [7], [8],  
92 [9]. For some of these genes, like BDNF, the serotonin transporter and the glucocorticoid  
93 receptor, altered DNA methylation was also revealed in humans after experiencing early life  
94 stress [10], [11], [12]. All of the above-cited studies focused on a candidate gene approach.  
95 Transgenic mice models with altered expression of the examined genes only partially show a  
96 consistent depressive-like phenotype [13], [14], [15], [16], [17]. Two more recent studies used  
97 a genome-wide methylation analysis and reported a wide range of epigenetic alterations as a  
98 result of early life stress [18], [19], but none of them led to a behaviorally validated mouse  
99 model for depression.

100 In a novel systematic translational genome-wide epigenetic approach, we recently succeeded  
101 in detecting an epigenetic marker of early life stress that is present in blood cell progenitors at  
102 birth in humans and monkeys, and also detectable in the prefrontal cortex of adult rats:  
103 Microorchidia (MORC) 1 – alias MORC family CW-type zinc finger 1 [20]. Moreover, we  
104 were able to verify an association between *Morc1* and major depressive disorder in a gene-set  
105 based analysis of an already available genome-wide association study [21].

106

107 MORC was first described in mammals by Watson et al. [22], who found that it is mainly  
108 expressed in male germ cells where it regulates mammalian germ cell development and  
109 meiosis. Male *Morc1*<sup>-/-</sup> mice are infertile due to a disrupted spermatogenesis, whereas male

110 *Morc1*<sup>+/-</sup> and female *Morc1*<sup>-/-</sup> mice do not show deficits in their reproductive system, *Morc1*  
111 knockout mice were seen exclusively as a model for male fertility defects. The human MORC  
112 protein consists of 984 amino acids and is 66% identical to the mouse MORC. Mutations in  
113 humans might also cause male infertility and be involved in testicular germ cell tumors [23].  
114 More recently Pastor et al. [24] reported that the male infertility in *Morc1* mutant mice is  
115 caused by defects of DNA methylation of specific classes of transposons, resulting in failed  
116 transposon silencing at these sites. Further analysis revealed that the MORC family is not only  
117 decisive for male reproduction, but is also involved in the pathophysiology of numerous  
118 forms of cancer: *Morc1* e.g. is frequently expressed in multiple myeloma [25] and mutated in  
119 estrogen receptor-positive lobular breast cancer [26]. Additionally, *Morc1* has been related to  
120 diabetes traits in a genome-wide complex trait analysis [27], which could play a role in the  
121 association of major depressive disorder with type 2 diabetes mellitus. In plants, *Morc1*  
122 influences immunological processes by different gene silencing mechanisms and  
123 heterochromatin condensation [28], [29], [30]. Recent evidence suggests that *Morc1* plays a  
124 more general biological role as part of a highly conserved nuclear protein superfamily that  
125 serves as epigenetic regulators in diverse nuclear processes that are not yet fully understood  
126 [31], [32], [33]. Hence *Morc1* appears to be primarily involved in gene silencing and changes  
127 of the chromatin structure [34], [35]. We were the first to report a different methylation of  
128 *Morc1* in brain tissue – the prefrontal cortex of adult rats – and thus strongly support the  
129 hypothesis of an epigenetic influence of this gene in the brain [20].

130

131 Animal models are still valuable tools in preclinical research, yet no rodent model of *Morc1*  
132 regarding depressive-like behavior has been established. Thus, in the present study we aimed  
133 to implement a transgenic mouse model to validate the role of *Morc1* expression in affective  
134 disorders. For this purpose we used *Morc*<sup>Tg(Tyr)1Az/J</sup> mice, in which exons 2-4 of *Morc1* gene  
135 on chromosome 16 had been deleted with the help of a transgenic insert [36]. We

136 characterized  $MORC^{Tg(Tyr)1Az/J}$  mice behaviorally in a test battery for locomotion and  
137 exploratory, anxiety-like and despair behavior. Plasma corticosterone levels were analyzed as  
138 a possible indicator of a depression-like HPA-system dysfunction. Furthermore, we  
139 determined total BDNF mRNA-levels known to be decreased in hippocampus in depressive  
140 patients and animal models of depression [37], [38], [39] as well as in closely connected and  
141 presumably affected structures, such as prefrontal cortex and amygdala [40], [41]. Due to  
142 difficult breeding and an increased mortality of male *Morc1*<sup>-/-</sup> mice, we only used female  
143 animals for our experiments, which is also the sex more predisposed to develop depression in  
144 humans.

145

## 146 **2 Materials and Methods**

147

### 148 **2.1 Animals**

149

150 *Morc*<sup>Tg(Tyr)1Az/J</sup> mice had been generated by introducing a transgenic construct containing a  
151 tyrosinase gene under the control of an RNA polymerase II 1 promoter into FVB/N fertilized  
152 mouse eggs. Sequences adjacent to the transgenic insert are disrupted resulting in the deletion  
153 of *Morc1* exons 2-4 [36]. Heterozygous *Morc1*<sup>+/-</sup> on the original FVB/N background were  
154 purchased from the Jackson Labs (Bar Harbour, Maine, USA) and crossed with C57BL/6N  
155 (Charles River, Sulzfeld, Germany) mice to obtain an F1 generation. Heterozygous F1  
156 offspring were then inter-crossed to generate an F2 generation, which was used for behavioral  
157 and molecular analyses. Male *Morc1*<sup>-/-</sup> mice were infertile and had smaller testes than  
158 wildtypes, whereas male *Morc1*<sup>+/-</sup> mice and female *Morc1*<sup>-/-</sup> mice showed no reproductive  
159 deficits. Animals were genotyped by PCR as recommended by the Jackson Labs (Bar Harbour,  
160 Maine, USA). Female *Morc1*<sup>-/-</sup> mice and their wild-type littermates were housed individually  
161 2 weeks before the first behavioural test started in macrolon type II cages with nesting  
162 material under a reversed day-night cycle (lights on from 19.00-07:00 hrs with 12h dark and  
163 12-h light phase) and supplied with food and water *ad libitum*. All procedures complied with  
164 the regulations covering animal experimentation within the EU (European Communities  
165 Council Directive 2010/63/EU). They were conducted in accordance with the institutions'  
166 animal care and use guidelines and approved by the national and local authorities  
167 (Regierungspräsidium Karlsruhe).

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169

### 170 **2.2 Behavioral tests**

171

172 At the age of 3, 5 and 6 months, 11 female *Morc1*<sup>-/-</sup> mice and 12 of their wild-type littermates  
173 were tested behaviorally in the dark phase, i.e., in the animals' active phase. Mice were  
174 acclimatized to the experimental room for at least 30 minutes prior to each test and tested by  
175 an investigator, who was blinded regarding their genotype. The order of the tests followed

176 earlier recommendations ranking the tests from least stressful to more stressful. Mice were  
177 sacrificed 2-3 weeks after the last experiment at an age of 21 - 25 weeks.

178  
179 *2.2.1 Novel Cage Test*

180  
181 Explorative behaviour was measured by counting the number of rearings within the first 5  
182 minutes after placing the animal into a fresh standard type II macrolon cage as described  
183 earlier [42].

184  
185 *2.2.2 Open Field Test*

186  
187 To evaluate locomotor and exploratory behavior, mice were individually placed into an open  
188 arena measuring 50x50 cm<sup>2</sup> under dimmed light conditions (25 Lux). Activity monitoring was  
189 conducted 15 min via a Video camera (Sony CCD IRIS). The resulting data were analyzed  
190 using the image processing system EthoVison 1.96 (Noldus Information Technology,  
191 Wageningen, the Netherlands) as described earlier [43].

192  
193 *2.2.3 Dark-Light-Box*

194  
195 Anxiety-related behavior was tested in the Dark-Light Box consisting of two plastic chambers  
196 connected by a small tunnel. Mice were placed into the dark chamber, which was covered by  
197 a lid and measured 20x15 cm<sup>2</sup>. Latency to first exit, number of exits and total time in the  
198 aversive light compartment (30x15 cm<sup>2</sup>, illuminated with 600 Lux) was recorded for 5 min as  
199 described earlier [44].

200  
201 *2.2.4 O-Maze*

202  
203 The Elevated Zero-Maze analyzes anxiety-related behavior by assessing avoidance of the  
204 aversive unsheltered compartment of the arena. A grey plastic annular runway (width 6 cm,  
205 outer diameter 46 cm, 50 cm above ground level, illuminated with 25 Lux) was covered with  
206 black cardboard paper to prevent animals slipping off the maze. Two opposing sectors were  
207 protected by inner and outer walls with a height of 10 cm. Animals were placed in one of the

208 protected sectors and latency to first exit, number of exits and total time spent in the open  
209 compartments was measured for 5 minutes as described earlier [45].

210  
211 *2.2.5 Forced Swim Test*

212  
213 The Forced Swim Test was applied to measure depressive-like behavior. For this purpose  
214 mice were placed into a glass cylinder (23 cm height, 13 cm diameter) filled with water (22°C)  
215 up to a height of 8 cm. Within a period of 6 min the onset and the percentage of floating was  
216 determined as described earlier [46]. 24 hours later the animals were tested again under the  
217 same conditions as before.

218  
219 *2.2.6 Hot Plate Test*

220 To exclude altered pain sensitivity as a confounding factor for Learned Helplessness, the  
221 Hotplate Test (ATLab, Vendargues, France) was applied. Temperature was set at 53 °C  
222 ( $\pm 0.3$  °C) and a 45s cut-off was determined to prevent injury. Latency to first reaction, i.e.  
223 licking hind paws or jumping, was assessed as described earlier [47].

224  
225 *2.2.7 Learned Helplessness*

226  
227 In the Learned Helplessness Paradigm mice were placed in a transparent plexiglas shock  
228 chamber (18×18×30cm<sup>3</sup>) with a stainless steel grid floor (Coulborn Instruments, Düsseldorf,  
229 Germany) through which they received 360 unpredictable and unavoidable footshocks  
230 (0.150mA) on 2 consecutive days. The footshocks applied varied regarding shock-duration  
231 (1–3s) and interval-episodes (1–15s) and lasted approximately 52 min in total. 24 hours after  
232 the second shock procedure, learned helplessness was assessed by testing shuttle box  
233 performance (Graphic State Notation, Coulborn Instruments, Düsseldorf, Germany) as  
234 described earlier [48]. Spontaneous initial shuttles from one compartment to the other were  
235 counted during the first 2min. Performance during 30 shuttle escape trials each starting with a



236 light stimulus of 5s, announcing a subsequent footshock (intensity 0.15mA) of maximum 10s  
237 duration was analyzed. Inter-trial interval was 30s and total testing time about 20min.

238

### 239 **2.3 CORT levels**

240 All animals were sacrificed at the age of 21-25 weeks by decapitation between 8.00 and 11.00  
241 hrs in the morning, and trunk blood was collected within 30s after the animal's removal from  
242 the cage. **Baseline** plasma corticosterone levels – **without applying any acute stress or**  
243 **intervention before taking the samples** – were determined using commercially available  
244 radioimmunoassay kits (ICN Biomedicals, Eschwege, Germany) as described earlier [49].

245

### 246 **2.4 RNA Preparation and Gene Expression Analysis by Quantitative Real-Time PCR**

247 After decapitation of the animals, the brains were immediately extracted from the skull. The  
248 hippocampus, prefrontal cortex and amygdala were rapidly dissected from the whole brain,  
249 frozen on dry ice and stored for later analyses.

250 Total RNA was isolated by single step of guanidinium isothiocyanate/phenol extraction using  
251 PureZol RNA isolation reagent (Bio-Rad Laboratories) according to manufacturer's  
252 instructions and quantified by spectrophotometric analysis. Following total RNA extraction,  
253 the samples were processed for real-time PCR (RT-PCR) to assess total BDNF mRNA levels.  
254 An aliquot of each sample was treated with DNase to avoid DNA contamination. RNA was  
255 analyzed by TaqMan qRT-PCR instrument (CFX384 real time system; Bio-Rad Laboratories)  
256 using the iScript™ one-step RT-PCR kit for probes (Bio-Rad Laboratories). Samples were  
257 run in 384 well formats in triplicate as multiplexed reactions with a normalizing internal  
258 control (36B4). Primers and probes sequences of total BDNF (forward primer:  
259 AAGTCTGCATTACATTCCTCGA, reverse primer:  
260 GTTTTCTGAAAGAGGGACAGTTTAT and probe: TGTGGTTTGTGCCGTTGCCAAG)  
261 and 36B4 (forward primer: AGATGCAGCAGATCCGCAT, reverse primer:

262 GTTCTTGCCCATCAGCACC and probe: CGCTCCGAGGGAAGGCCG) were purchased  
263 from Eurofins Genomics (Vimodrone, Italy), while probe and primer sequences for *Morc1*  
264 (Assay ID: Mm00501711\_m1) were purchased from Life Technologies (Monza, Italy) and  
265 are available on request.

266

## 267 **2.5 Statistical analyses**

268 Intergroup comparisons were calculated by one-sided t-tests assuming that knockout-mice  
269 display higher levels of depressive and anxiety-like behavior as well as higher CORT-levels,  
270 less body weight, less locomotor and exploratory behavior and decreased total BDNF mRNA-  
271 levels in hippocampus, prefrontal cortex and amygdala than wild-types. A one-way repeated  
272 measurements ANOVA was used to analyze the open field test. Calculations regarding  
273 mRNA levels were run with fold change values. The correlation between Learned  
274 Helplessness Escape Latency and Failures was analyzed applying spearman rho correlation. A  
275 p-value  $\leq 0.05$  was seen as the level of statistical significance in all tests. The statistical  
276 analyses were performed using the SPSS 21.0 software package for Windows.

277

## 278 **3 Results**

### 279 *3.1 Morc1 knockout mice display regular locomotor and exploratory behavior*

280 *Morc1*<sup>-/-</sup> mice showed normal vertical locomotor activity (rearing) in the Novel Cage Test  
281 (T(21)=0.874; p=0.196; data not shown). The horizontal locomotor activity and exploratory  
282 behavior as measured by the Open Field Test also did not differ significantly from control  
283 animals: e.g. the total distance moved by *Morc1*<sup>-/-</sup> mice was similar to those of the control  
284 group (time\*genotype: F<sub>1,21</sub>=1.125, p=0.301; between subject-factor genotype F<sub>1,21</sub>=0.032;  
285 p=0.859). Both genotypes moved significantly less in the second half of the test, showing that  
286 habituation had taken place as expected (Time: F<sub>1,21</sub>=8.792; p=0.007; data not shown).

287

### 288 *3.2 Morc1 knockout mice show unaltered anxiety-related behavior*

289 *Morc1*<sup>-/-</sup> mice did not display increased anxiety-like behavior as monitored in the Dark-Light  
290 Box (latency: T(21)=1.023; p=0.159; end exploration: T(14.011)=0.914; p=0.188; exits:  
291 T(21)=-1.289; p=0.106; light time: T(21)=-0.074; p=0.471; data not shown). In the Elevated  
292 O-Maze, again *Morc1*<sup>-/-</sup> mice exhibited similar latencies to enter the aversive compartment as  
293 their littermate controls (T(21)=0.314; p=0.3785; data not shown). They also did not exit  
294 (T(21)=-0.128; p=0.4495; data not shown) or fully cross (T(21)=0.095; p=0.4625; data not  
295 shown) the maze less than controls and spent about the same amount of time in the open arms  
296 of the maze (T(21)=-0.862; p=0.199; data not shown).

297

### 298 *3.3 Morc1 knockout mice demonstrate increased depressive-like behavior*

299 Depressive-like behavior as measured by the Forced Swim Test showed that *Morc1*<sup>-/-</sup> mice  
300 display a significant lower latency to float (T(21) =2.346; P=0.015) on day 1 than wild-types  
301 (s. Fig 1A). Additionally, on day 1 immobility times were increased in *Morc1*<sup>-/-</sup> mice, which  
302 resulted in a statistical tendency from minute 4 to 6 of the test (minute 0 to 2: T18.051=-0.819;

303 p=0.216; minute 2 to 4: T21=-0,567; p=0.289; minute 4 to 6: T21=-1.343; p=0.097; s. figure  
304 1B).

305 24 hours later (on day 2), the difference in the latency to start floating was not detectable any  
306 more (T21=0.113; p=0.456; s. fig. 1C.) Furthermore, *Morc1*<sup>-/-</sup> mice showed a significant  
307 increase in immobility on day 2 from minute 2-4 compared to wild-types (T(21)=-2.009;  
308 p=0.029) and a statistical tendency also from minute 4-6 (T21=-1.431; p=0.084) as depicted  
309 in fig. 1D. From minute 0 to 2 there was no significant difference in immobility time between  
310 the two groups (T21=-0.508; p=0.308).

311  
312 In the Learned Helplessness Paradigm of depression, *Morc1*<sup>-/-</sup> mice displayed significantly  
313 more failures to escape in comparison to littermate controls (T(12.573)=-1.844; p=0.045) (s.  
314 Fig 2A). As a tendency, *Morc1*<sup>-/-</sup> mice had a higher latency to escape than controls (T  
315 (16.093)=-1.539; p=0.072) as shown in fig. 2B. A clear correlation between escape latency  
316 and number of failures with the *Morc1*<sup>-/-</sup> mice showing the highest values (spearman  
317 rho=0.961, p<0.000) is depicted in fig. 2 C.

318

319 *3.4 Morc1 knockout mice reveal regular CORT levels, but increased total BDNF in*  
320 *hippocampus*

321 *Morc1*<sup>-/-</sup> mice showed similar plasma corticosterone levels as their control littermates  
322 (T(21)=-0.941; p=0.1785; s. fig. 3A). In the hippocampus however, total BDNF mRNA-levels  
323 of *Morc1*<sup>-/-</sup> mice were significantly increased compared to those of wild-types (T(21)= -3.538;  
324 p=0.001; s. Fig. 3B). As a tendency, the total BDNF mRNA-levels in the prefrontal cortex  
325 (T(14.968)=1.428; p=0.087; data not shown) and in the amygdala (T(18)=1,337; p=0.099;  
326 data not shown) were down-regulated.

327 As expected, *Morc1* mRNA was not expressed in *Morc1*<sup>-/-</sup> mice, whereas we found its full  
328 mRNA expression in the wildtype animals in hippocampus, prefrontal cortex and amygdala  
329 (data not shown).

330

## 331 **4 Discussion**

332 In the present study, we characterized for the first time *Morc1*<sup>-/-</sup> mice in a test battery for  
333 emotional behavior. *Morc1*<sup>-/-</sup> mice floated earlier in the Forced Swim Test and showed more  
334 immobility in general. In the Learned Helplessness Paradigm, they exhibited a higher latency  
335 to escape and more escape failures, two parameters that are seen as classical indicators for  
336 increased learned helplessness in rodents [50]. *Morc1*<sup>-/-</sup> mice showed a significant increase of  
337 immobility in the Forced Swim Test only on day 2 and just as a statistical tendency on day 1.  
338 Although day 1 is regarded as more relevant for mice than day 2, the consistently increased  
339 means of immobility on both days taken together with the results of the Learned Helplessness  
340 Test, allow to postulate a clear increase in depressive-like behavior of *Morc1*<sup>-/-</sup> mice.

341 Anxiety is often concomitant to depression in psychiatric patients. Up to now, the question if  
342 *Morc1* is also involved in anxiety disorders has not been examined in clinical trials. In our  
343 animal model we did not find any evidence of increased anxiety in *Morc1*<sup>-/-</sup> mice in the Dark-  
344 Light Box Test and Elevated O-Maze, indicating that *Morc1* may specifically be involved in  
345 depression but not anxiety. We were able to exclude reduced locomotion and exploratory  
346 behavior as possible confounding factors in our study, as *Morc1*<sup>-/-</sup> mice did not show any  
347 difference to their wild-type littermates in the Novel Cage and Open Field Test.

348 All in all, this new transgenic mutation seems to represent a promising model to further  
349 investigate the depressive phenotype and its underlying neurochemical, genetic and epigenetic  
350 pathophysiology.

351

352 Besides a deficient spermatogenesis, small testicular size and aberrant eye pigmentation,  
353 Watson et al. [22] described no phenotypic abnormalities in male *Morc1*-mutant mice. In our  
354 breeding, we found increased mortality in male *Morc1*<sup>-/-</sup> mice, and the small sample size of  
355 males in our cohort constrained us to focus only on females for our behavioral testing.  
356 Although we bred only for two generations, a random effect cannot be definitely dismissed,

357 yet this increased mortality might be another confirmation of *Morc1* – as already stated above  
358 – not only being involved in spermatogenesis but serving a more general biological function  
359 [31], [32], [33].

360 This leads to the more general question why *Morc1* might be involved in spermatogenesis as  
361 well as in mood disorders – two biological processes seemingly independent at first glance.  
362 Soumillon et al. [51] come to the conclusion that a considerable part of genes expressed in  
363 testes do not have testes-specific functions. Shen et al. [52] could demonstrate that testes-  
364 specific genes usually have a fast evolutionary rate and therefore are more likely to gain new  
365 functions. In their phylogenetic analysis they illustrate that some open reading frames were  
366 first expressed in testis and later in evolution got expressed in other tissues. The authors  
367 assume that testis may play a role in producing new genes and even in supporting testes-  
368 specific genes in gaining new functions for other organs/tissues. Blendy et al. [53] for  
369 example demonstrated that the *Crem* gene is involved in spermatogenesis. Aguado et al. [54]  
370 found the same gene being involved in hippocampal synaptogenesis. Wang et al. [55] proved  
371 another gene - *hsf-2* – to be involved in sperm production as well as central nervous  
372 development.

373

374 As women are more often affected by major depressive disorder than men [56], [57], we saw  
375 the restriction to female  $^{-/-}$  mice in the present study as a possibility for validly modeling  
376 depression. Nevertheless, sex differences with respect to the development of depressive-like  
377 behaviors in mice with targeted mutagenesis have also been reported for other genes such as  
378 BDNF [58], [59], [60]. Therefore another study on male *Morc1* $^{-/-}$  mice and their conceivable  
379 depressive-like behavior seems appealing. A possible sex effect in our animal model could  
380 also shed more light on the differing pathophysiology of depression in women and men.  
381 Moreover, additional behavioral tests concerning e.g. social behavior and cognition that are

382 often correlated to depressive-like behavior, are warranted to further characterize the novel  
383 *Morc1* mouse model of depression.

384

385 We did not detect any difference regarding baseline corticosterone levels in *Morc1*<sup>-/-</sup> mice  
386 suggesting that the HPA-axis in these animals is not disrupted under resting conditions. This  
387 does not exclude the possibility that the hormonal responsiveness is affected under  
388 challenging conditions.

389

390 We also investigated BDNF expression that represents a prototype marker of neuronal  
391 plasticity, which has often been associated with a depressive phenotype, its expression being  
392 reduced in the brain of depressed subjects as well as in different animal models of depression  
393 [61], [62] – summarized in the so-called neurotrophin hypothesis of depression. Contrary to  
394 these previously found changes, the hippocampal expression of BDNF was upregulated in  
395 *Morc1*<sup>-/-</sup> mice, whereas a trend toward a reduction was found in the prefrontal cortex. Despite  
396 the complex interactions between glucocorticoids and BDNF, this finding might be narrowed  
397 down to the undisturbed HPA-Axis in our animals, as BDNF is mostly suppressed by  
398 glucocorticoids [63], [64].

399 An alternative explanation for this unexpected result may be sex differences. Interestingly, the  
400 decrease of BDNF in depressed patients was found to be more pronounced in men than in  
401 women [65]. Hayley et al. [66] demonstrated that in female depressed patients who committed  
402 suicide BDNF protein levels were reduced in the frontopolar prefrontal cortex, but not in the  
403 hippocampus. Conversely, males displayed significantly decreased BDNF protein levels only  
404 in the hippocampus yet not in the prefrontal cortex. Jaworska et al. [67] showed a significant  
405 decrease of BDNF in the hippocampus of male, but not of female gerbils after early life stress.  
406 As we used only female mice for our experiment, this sex-specific regulation could be the  
407 reason for the lack of a BDNF down-regulation in our study. Furthermore, there is growing



408 evidence that estradiol can induce BDNF expression and vice versa that estradiol effects in  
409 the hippocampus are mediated by BDNF [68], [69], [70], [71].

410

411 In a recent study, Calabrese et al. [72] demonstrated that BDNF in serotonin transporter  
412 knock-out rats, which had been exposed to early life stress was down-regulated in the ventral  
413 hippocampus and the ventromedial prefrontal cortex, but was significantly increased in the  
414 dorsal hippocampus and the dorsomedial prefrontal cortex. A similar region-specific process  
415 might underlie the detected increase of BDNF in hippocampus in the present study with the  
416 amount of BDNF in the dorsal parts outweighing the downregulation in the ventral parts. We  
417 did not differentiate between the dorsal and ventral regions of hippocampus and prefrontal  
418 cortex, but this issue would be very interesting to address in future experiments.

419

420 However, all these aspects fail to entirely explain the highly significant increase of BDNF we  
421 found in hippocampus of *Morc1*<sup>-/-</sup> mice. In contrast, the trend of reduced BDNF in prefrontal  
422 cortex and amygdala of *Morc1*<sup>-/-</sup> mice is more in line with previous findings showing either  
423 no change of BDNF in these two regions [73] or a decrease in depressive-like animals [74]  
424 and depressive humans [75].

425

426 Last, increased BDNF levels in the hippocampus might also be the consequence of  
427 compensatory mechanisms set in motion in *Morc1* mutants. Thus, Faure et al. [76], Marais et  
428 al. [77] and Daniels et al. [78] found a significant increase of BDNF in (dorsal) rat  
429 hippocampus after maternal separation during early life. Hellweg et al. [79] also found a  
430 significant increase of BDNF in bulbectomized depressive-like mice. According to the  
431 authors this finding in addition to the fact that bulbectomized animals display different  
432 behavioral abnormalities than glucocorticoid receptor compromised mice as well as a  
433 serotonergic dysfunction possibly defines a new endophenotype of depression. Although

434 *Morc1*<sup>-/-</sup> mice show the same depressive-like phenotype with deficiencies in the FST and LH  
435 paradigm as described by the animals used for the neurotrophin hypothesis of depression,  
436 their serotonergic function has not been evaluated yet. Maybe *Morc1*<sup>-/-</sup> mice constitute an  
437 intermediate endophenotype situated between the two previously described animal models.  
438 Nonetheless, as BDNF-levels in the brain have never been analyzed before in *Morc1*<sup>-/-</sup> mice,  
439 the regulation of BDNF in regard to *Morc1* and depression remains a matter of pure  
440 speculation. Systematic investigation of this question is required.

441  
442 One of the most intriguing issues to address in the future will be the role of epigenetic  
443 modulation by *Morc1* in the brain. We were the first to detect a different methylation of  
444 *Morc1* in the prefrontal cortex of the rat brain [20]. In our cross-species and cross-tissues  
445 approach, we could prove the particular relevance of *Morc1* methylation after early life stress.  
446 However, to date the specific cerebral subtypes that are affected by *Morc1* and the concrete  
447 epigenetic function of *Morc1* in the brain is still completely unknown.

448 A further implication of this finding is the question if the methylation pattern will be  
449 replicable in the mouse brain or in other regions of the brain like e.g. the hippocampus. As  
450 epigenetic processes are always highly sex-specific, it would also be most interesting to  
451 examine in which way the epigenetic mark will be expressed differently in male individuals.

452  
453 One limitation of the presented study is certainly that only females have been used so that the  
454 model might be a sex-specific. Additionally, as we analyzed BDNF and CORT levels after  
455 applying stressful behavioral tests to the animals, we cannot exclude that e.g. the FST or the  
456 LH paradigm has had some influence on these parameters. It is thinkable that the increase of  
457 BDNF in the hippocampus of *Morc1*<sup>-/-</sup> mice is due to some compensatory mechanism  
458 induced by the higher amount of electric shocks these animals received in the LH paradigm  
459 due to their bad performance in this behavioral test.

460 Our study results are further limited by the fact that we only used *Morc1* homozygous  
461 knockout mice as heterozygous knockouts are usually regarded as a better model for the  
462 human condition. We are actually planning to also behaviorally analyze *Morc1* heterozygous  
463 - and male - mice after backcrossing our animals for a few more generations into the c57/bl6  
464 background.

465

466 In conclusion, although further work still has to be accomplished, the animal model of *Morc1*<sup>-/-</sup>  
467 mice will be useful in future studies on the role of *Morc1* in the pathophysiology and  
468 therapy of depression.

469

## 470 **5 Conflict of interest**

471 The authors declare no conflict of interest.

472

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