

SCIENTIFIC REPORTS



OPEN

Migration phenology and breeding success are predicted by methylation of a photoperiodic gene in the barn swallow

Received: 10 January 2017

Accepted: 16 February 2017

Published: 31 March 2017

Nicola Saino¹, Roberto Ambrosini², Benedetta Albetti³, Manuela Caprioli¹, Barbara De Giorgio³, Emanuele Gatti², Felix Liechti⁴, Marco Parolini¹, Andrea Romano¹, Maria Romano¹, Chiara Scandolaro¹, Luca Gianfranceschi¹, Valentina Bollati³ & Diego Rubolini¹

Individuals often considerably differ in the timing of their life-cycle events, with major consequences for individual fitness, and, ultimately, for population dynamics. Phenological variation can arise from genetic effects but also from epigenetic modifications in DNA expression and translation. Here, we tested if CpG methylation at the poly-Q and 5'-UTR loci of the photoperiodic *Clock* gene predicted migration and breeding phenology of long-distance migratory barn swallows (*Hirundo rustica*) that were tracked year-round using light-level geolocators. Increasing methylation at *Clock* poly-Q was associated with earlier spring departure from the African wintering area, arrival date at the European breeding site, and breeding date. Higher methylation levels also predicted increased breeding success. Thus, we showed for the first time in any species that CpG methylation at a candidate gene may affect phenology and breeding performance. Methylation at *Clock* may be a candidate mechanism mediating phenological responses of migratory birds to ongoing climate change.

Organisms fine-tune the timing of their life-cycle events (i.e. their phenology) according to circannual variation in ecological conditions¹. Temporal orchestration of life-cycle events such as budding and flowering in plants, and breeding and migration in animals, is a major target of natural selection, which sets a fitness premium on temporal matching of these activities with the ecological conditions when they are best performed^{1–3}. Variation in the time schedule of major life-history events among individuals, however, can be large, and our understanding of the mechanisms that generate such variation is lagging behind the appreciation of its pervasive occurrence. In particular, no studies to date have addressed the role that DNA epigenetic modifications may have in shaping the phenology of animals. However, recent studies have started to unveil the role of epigenetic processes in causing variation in behavioural traits in the wild, like novelty seeking⁴, personality⁵, or migration propensity⁶.

Perception of day length (photoperiod) variation is a core mechanism that dictates the timing of seasonal behavioural and physiological changes of several organisms^{7,8}. 'Circadian clock' systems sensing seasonal changes in light/dark cycles are conserved in organisms as taxonomically diverse as plants and vertebrates^{9,10}. *Clock* genes, in particular, code for elements of a transcriptional-translational feedback loop ('core circadian oscillator'; CCO)⁹. Within CCO, *Clock* and *Bmal1* products heterodimerize to produce a transcription factor, which acts both as a positive driver of the molecular oscillations and as an 'output' signal from the circadian oscillator¹¹. The central pacemakers in the avian pineal gland, retinae and SCN dynamically interact to maintain stable phase relationships and then influence downstream rhythms through entrainment of peripheral oscillators in the brain controlling behavior¹². An important role in controlling circadian rhythms is played by the *Clock* carboxyl-terminal polyglutamine repeat (poly-Q) domain, which serves as a transcriptional trans-activation domain¹³ and appears to be highly evolutionarily conserved¹⁴.

¹Department of Biosciences, University of Milan, via Celoria 26, I-20133 Milan, Italy. ²Department of Earth and Environmental Sciences (DISAT), University of Milano Bicocca, Piazza della Scienza, 1, I-20126 Milan, Italy. ³EPIGET - Epidemiology, Epigenetics and Toxicology Lab - Department of Clinical Sciences and Community Health, University of Milan, via San Barnaba 8, I-20122 Milan, Italy. ⁴Swiss Ornithological Institute, Seerose 1, CH-6204, Sempach, Switzerland. Correspondence and requests for materials should be addressed to N.S. (email: nicola.saino@unimi.it) or V.B. (email: valentina.bollati@unimi.it)

Clock polymorphism predicts among-individual variation in breeding, migration and molt time schedules in some species^{15–18}, although evidence is inconsistent across species and phenological traits^{18–21}. In the focal species of the present study, the barn swallow (*Hirundo rustica*), *Clock* exhibits low polymorphism at the poly-Q region^{16,22}. Albeit low, however, polymorphism at poly-Q predicts timing of spring migration and breeding, and of plumage molt in the sub-Saharan wintering quarters, with the individuals carrying alleles with larger *Clock* poly-Q region showing delayed phenology^{16,22,23}.

Strong genetic determinism of phenological traits may evolve because of the advantages that strict endogenous control of circannual rhythmicity can afford in terms of appropriate timing of life history events^{24,25}. ‘Hardwired’ mechanisms of phenological control relying on photoperiodic information may be favoured because seasonal changes of environmental conditions are predictably associated with change in photoperiod. Yet, genetic variation at the few candidate phenological genes that have been investigated so far, like *Clock* and other photoperiodic genes, can be small even in populations showing large phenological variation^{15,16}, rather suggesting low environmental canalization of phenological traits. In fact, environmental conditions can be a major source of phenological variation via their effects on individual general state, because major life-history events like reproduction and migration are energy and time demanding, and individual physical conditions can thus constrain individual phenological decisions²⁶.

Epigenetic processes can reconcile small genetic variation in phenological genes with often large among-individuals variation in phenology, and thus also with the observed phenological responses of populations to climate change²⁷. Environmental experience can orchestrate the way in which genetic information is expressed and translated, and epigenetic DNA alterations can thus mediate the effects of environmental conditions on phenotypic expression²⁸. Methylation at CpG dinucleotides located in CpG islands, for example, has been shown to correlate with a stable shutdown of the associated promoter, and consequently a block of gene transcription²⁹.

Here, for the first time we analyzed individual variation in major phenological events (i.e. start and end dates of spring migration, and breeding date) in relation to methylation at two regions (poly-Q and 5'-UTR) of the photoperiodic *Clock* gene³⁰. We focused on long-distance migratory barn swallows that were equipped with light-level geolocators to record the timing of spring migration from sub-Saharan Africa to the breeding sites in southern Europe. We expected methylation at both loci to result in earlier spring migration and breeding phenology, because methylation is usually associated with a reduced transcription and thus an expected weaker activation signal from the CLOCK/BMAL1 transcription^{31–35}. Importantly, epigenetic components of phenotypic variation may not be independent of underlying genetic variation. In the analyses, we therefore controlled for *Clock* genotype, expressed as number of the poly-Q repeats at the *Clock* region, because genetic polymorphism at poly-Q has been shown to predict phenological variation in barn swallows^{16,23}. The methodological and technical information on genotyping of the *Clock* poly-Q region and on the relationships between genotype and migration phenology have been reported in a previous study²³.

In addition, we analyzed the consequences of methylation at *Clock* for seasonal breeding success. The link between methylation and breeding success could be an indirect one, being mediated by the cascading ‘carry-over’ effects³⁶ of timing of migration on timing of breeding and thus number of breeding events per season. Under this scenario larger methylation was expected to result in larger seasonal reproductive success because larger methylation was predicted to advance migration and breeding dates and thus to increase the seasonal number of broods. In an attempt to discern between pattern and process we therefore partitioned the direct and indirect (mediated by phenology) contributions of methylation to breeding success by means of path analysis.

Results

Methylation at both loci did not significantly differ between males and females (poly-Q: males: 74.82% (0.99, n = 65), females: 77.11% (1.11, n = 29); $t_{92} = 1.37$, $P = 0.173$; 5'-UTR: males: 80.95% (0.43, n = 66), females: 81.27% (0.79, n = 28), $t_{92} = 0.39$, $P = 0.696$).

Methylation levels at poly-Q recorded in two consecutive years (i.e. the years when the geocator was deployed or, respectively, when it was recovered) were positively correlated ($r = 0.470$, $n = 25$, $P = 0.018$), whereas there was no significant correlation between years in methylation at the 5'-UTR ($r = 0.141$, $n = 23$, $P = 0.521$). Hence, methylation levels at poly-Q at one life stage reflect methylation also at other life stages.

Methylation and spring phenology. Departure date from the wintering area was significantly earlier in females but not in males with higher levels of poly-Q methylation (Table 1 and Fig. 1a). Arrival date to the breeding site was significantly earlier in both males and females with increased poly-Q methylation (Table 1 and Fig. 1b). In addition, breeding date was significantly earlier in females but not in males with relatively high poly-Q methylation (Table 1 and Fig. 1c). For all phenological variables, the relationship with poly-Q methylation was significantly steeper and negative among females, as implied by the significant sex by methylation interaction effect (Table 1 and Fig. 1). The proportion of variance in phenology of females that was explained by methylation was as large as 50.0% for departure date from the wintering area, 42.3% for arrival date to the breeding site and 29.9% for breeding date. The corresponding values for males were considerably smaller: 2.8%, 6.8% and 0.10%, respectively. Thus, the size of the effect of methylation on migration and breeding phenology was larger in females as compared to males.

In barn swallows, phenological traits including timing of spring migration and breeding, and molt schedules during wintering are predicted by genetic polymorphism at the poly-Q *Clock* regions^{16,22,23}. We therefore re-ran the models reported in Table 1 for poly-Q while also including the effect of mean within-individual number of poly-Q repeats or, respectively, the number of poly-Q repeats of the ‘longer’ allele as covariates (see also refs 16,18). The results of these models were qualitatively consistent with those in Table 1, meaning that all the significant interaction effects and also the within-sex relationships in Table 1 remained statistically significant (details

	poly-Q					5'-UTR				
	χ^2_1	F	df	P	Coefficient (SE)	χ^2_1	F	df	P	Coefficient (SE)
Departure date from wintering area										
Year	0.00					0.00 ^a				
Area	0.00					0.00 ^a				
Sex		10.79	1,62	0.002			0.81 ^a	1,61	0.371	
Methylation		17.43	1,62	<0.001			0.13 ^a	1,61	0.717	
Sex × Methylation		10.20	1,62	0.002			1.54	1,60	0.219	
Males					−0.29 (0.24)					0.57 (0.56)
Females					−2.15 (0.53) ^{***}					−0.58 (0.75)
Arrival date to breeding site										
Year	9.30 ^{**}					5.40 ^{**}				
Area	3.00					3.70 ^a				
Sex		4.38	1,57	0.041			2.16 ^a	1,57	0.147	
Methylation		17.64	1,57	<0.001			0.60 ^a	1,57	0.440	
Sex × Methylation		3.74	1,57	0.058			0.00	1,56	0.997	
Males					−0.59 (0.22) ^{**}					−0.31 (0.52)
Females					−1.61 (0.50) ^{**}					−0.31 (0.67)
Breeding date										
Year	7.70 ^{**}					7.70 ^{***}				
Area	4.30 [*]					4.30 ^{**}				
Sex		3.67	1,77	0.059			6.39 ^a	1,78	0.014	
Methylation		4.97	1,77	0.029			2.65 ^a	1,78	0.108	
Sex × Methylation		4.60	1,77	0.035			1.45	1,77	0.232	
Males					−0.02 (0.19)					−0.95 (0.47)
Females					−0.97 (0.40) [*]					0.14 (0.77)

Table 1. Linear mixed models of phenological variables in relation to sex and methylation at *Clock* poly-Q or 5'-UTR. *0.05 > P > 0.01; **0.01 ≥ P ≥ 0.001; ***P < 0.001. ^aEstimated from a model excluding the interaction term. χ^2_1 : likelihood ratio test for the random effect of year or area. Methylation (percentage of 5-methyl-cytosine) is included as a covariate. Year and area are included as random factors; sex as a fixed-effect factor. Sample sizes for the analyses of poly-Q were as follows (males, females): departure date from wintering area: 49, 20; arrival date to breeding site: 45, 19; breeding date: 58, 26. The corresponding values for the analyses of 5'-UTR were: 48, 19; 45, 18; 59, 25; 61, 25.

not shown). Hence, the effect of methylation at *Clock* poly-Q on phenology were independent of the underlying individual genetic constitution at *Clock* poly-Q.

Methylation at 5'-UTR, however, did not predict any of the phenological variables, either *per se* or in combination with the effect of sex (Table 1).

Methylation and seasonal breeding success. Methylation at poly-Q positively predicted seasonal breeding success (total number of eggs), with no statistically significant evidence for sex-dependent effects (Table 2). This result implies that the slopes of the relationships between breeding success and methylation did not differ between the sexes (Table 2 and Fig. 2). However, the correlation coefficient between breeding success and methylation at poly-Q was statistically significant for females ($r = 0.552$, $R^2 = 0.305$; $P = 0.003$, $n = 26$) but not for males ($r = 0.226$, $R^2 = 0.051$, $P = 0.082$, $n = 60$). Methylation at 5'UTR did not predict breeding success (Table 2).

Path analysis of the effect of methylation on breeding success. Phenological events occur in ordered sequences and each of them is therefore potentially influenced by timing of the preceding events³⁶. Such carry-over effects can therefore have cascading effects through the annual cycle, generating correlations between dates of phenological events as well as correlations with fitness traits (see ref. 37). In female barn swallows, departure date from the wintering area positively predicted arrival date at the breeding site, which in turn positively predicted breeding date (Fig. 3). Moreover, females that started breeding early had larger seasonal reproductive success (Fig. 3). In males, there were also positive relationships between the dates of consecutive phenological stages, and a negative relationship between breeding date and seasonal breeding success, although the size of the effect was smaller than in females (Fig. 3). Hence, any association between methylation and breeding success could result either from a direct effect or through an indirect effect mediated by phenology. Path analysis suggested that in females methylation at poly-Q affected breeding success mostly indirectly, through the relationships with date of departure from the wintering area and date of arrival to the breeding site, whereas the indirect effect via breeding date and the direct effect on breeding success were smaller (Fig. 3). In contrast, in males most of the effect of methylation was a direct one (Fig. 3). The overall compound path coefficients for the effect of methylation at poly-Q on breeding success was somewhat larger for females (0.339) as compared to males (0.252).

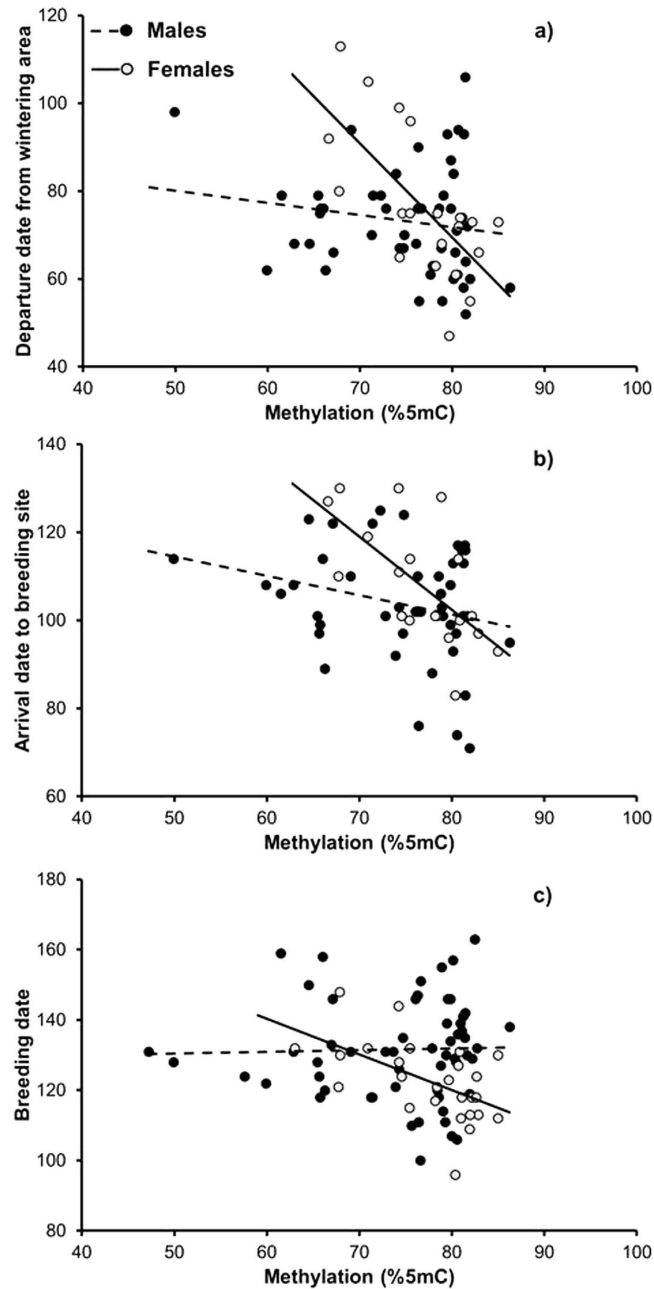


Figure 1. Phenological variation in departure date from the wintering area (a), arrival date at the breeding site (b), and breeding date (c) of male (full symbols) and female (open symbols) barn swallows in relation to methylation at the poly-Q locus. Linear regression lines for males (dashed) and females (continuous) are shown. Dates are expressed as Julian day (1 = 1 January). See Table 1 for sample sizes.

Discussion

Variation in ecologically important traits can be generated by epigenetic modifications of the expression and function of genes even when genetic variation at those traits is small. Epigenetic processes may therefore be pivotal to our understanding of how organisms respond to environmental variation via phenotypic plasticity²⁷. Ecological and evolutionary studies have long sought for the proximate mechanisms that control individual variation in the timing of major life-history events, because such variation has consequences for individual fitness and for the dynamics and evolution of populations. Here, we showed, for the first time in any species, that epigenetic regulation via methylation of a *Clock* poly-Q region can have a major impact in regulating timing of migration and breeding. Methylation at *Clock* poly-Q explained a very large fraction (>30%) of the variance in spring migration phenology, arrival dates at the breeding site and thus in seasonal breeding success, particularly in females. In males, the effects of methylation on spring migration phenology were considerably weaker than in females, and the consequences of methylation for seasonal breeding success seemed to be independent of timing of spring migration and breeding date. The sign of the relationship between methylation levels and dates

	poly-Q					5'-UTR				
	χ^2_1	F	df	P	Coefficient (SE)	χ^2_1	F	df	P	Coefficient (SE)
Seasonal fecundity										
Year	0.00 ^a					0.00 ^a				
Area	0.50 ^a					0.70 ^a				
Sex		5.86 ^a	1,80	0.018			6.92 ^a	1,80	0.010	
Methylation		7.01 ^a	1,80	0.010			0.00 ^a	1,80	0.969	
Sex × Methylation		1.02	1,79	0.316			0.01	1,79	0.924	
Males					0.07 (0.03)					-0.00 (0.09)
Females					0.15 (0.07)*					0.01 (0.14)

Table 2. Linear mixed models of seasonal breeding success in relation to sex and methylation at *Clock* poly-Q or 5'-UTR. *0.05 > P > 0.01; ^aestimated from a model excluding the interaction term. χ^2_1 : likelihood ratio test for the random effect of year or area. Methylation (percentage of 5-methyl-cytosine) is included as a covariate. Year and area are included as random factors; sex as a fixed-effect factor. Sample sizes for the analyses of poly-Q were 60 males and 26 females. Sample sizes for analyses of 5'-UTR were 61 males and 25 females.

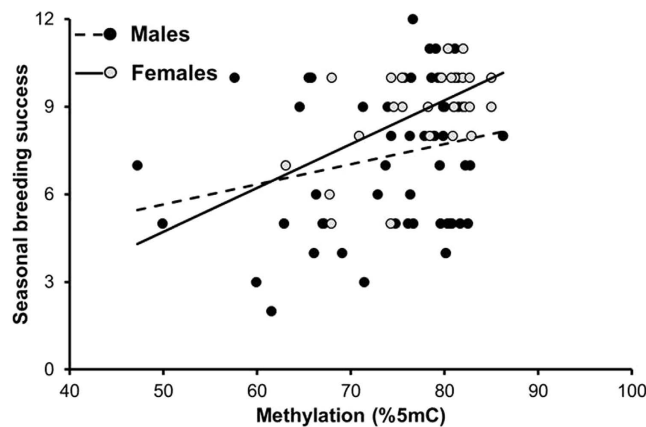


Figure 2. Seasonal breeding success of male (full symbols) and female (open symbols) barn swallows in relation to methylation at the poly-Q locus. Linear regression lines for males (dashed) and females (continuous) are shown. See Table 2 for sample sizes.

of phenological events, with larger methylation levels being associated with earlier phenology, was consistent with the expectation since hypermethylation is expected to reduce the overall quantity of the CLOCK/BMAL1 transcription factor, thus reducing the transcriptional activation of the clock genes. Such reduction may possibly mimic the condition of small length of the poly-Q stretch, which is associated with earlier phenology of breeding, migration and molt^{16,22,23}, although with a different mechanism, as the poly-Q region is directly involved in the transcriptional trans-activation of the CLOCK/BMAL1 regulated genes. The effect of methylation on phenology that we documented here, however, was independent of genetic polymorphism at *Clock* poly-Q, as demonstrated by analyses where we controlled statistically for the effect of length of the *Clock* poly-Q repeat. It should be noted that in the present study we measured *Clock* methylation in peripheral blood because for ethical reasons we could not sample other tissues (e.g. brain). While the present results should be considered under this caveat, methylation differences measured in blood have been suggested could have potential for being a good proxy of the changes occurring in other tissues, in particular at a central level³⁸.

As a photoperiodic gene, *Clock* has a major role in controlling endogenous circadian rhythmicity^{12,39}. However, by interacting with exogenous cues, circadian genes like *Clock* can be entrained by local environmental factors thereby participating in the regulation of seasonality in behavioural and physiological traits¹⁰, and genetic variation at *Clock* genotype has been shown to predict phenological variation in breeding, migration or plumage molt in several animals from diverse taxa^{15,18,22,23,40}. A causal link between methylation at *Clock* and phenological variation at photoperiod-dependent activities, like migration in birds, is suggested by the observation that DNA methylation can affect circadian rhythmicity^{41,42}. In addition, methylation at genes of the circadian rhythm system, including those that influence responsiveness to light can also impact on discrete behavioural traits like migratoriness (i.e. being migrant or not) and on related morphological and physiological syndromes, as observed for smoltification in rainbow trouts (*Onchorhynchus mykiss*)⁶. Hence, methylation of genes of the circadian system may not only influence timing of migration and of breeding, as suggested by the present study, but also cause discontinuous variation in migratoriness and related phenotypic syndromes.

The association between migration phenology and methylation at *Clock* poly-Q in our study was apparent only among females, while methylation levels did not differ between the sexes. This result may suggest differentially larger phenological control by methylation at *Clock* in females. Thus, in females methylation at *Clock*

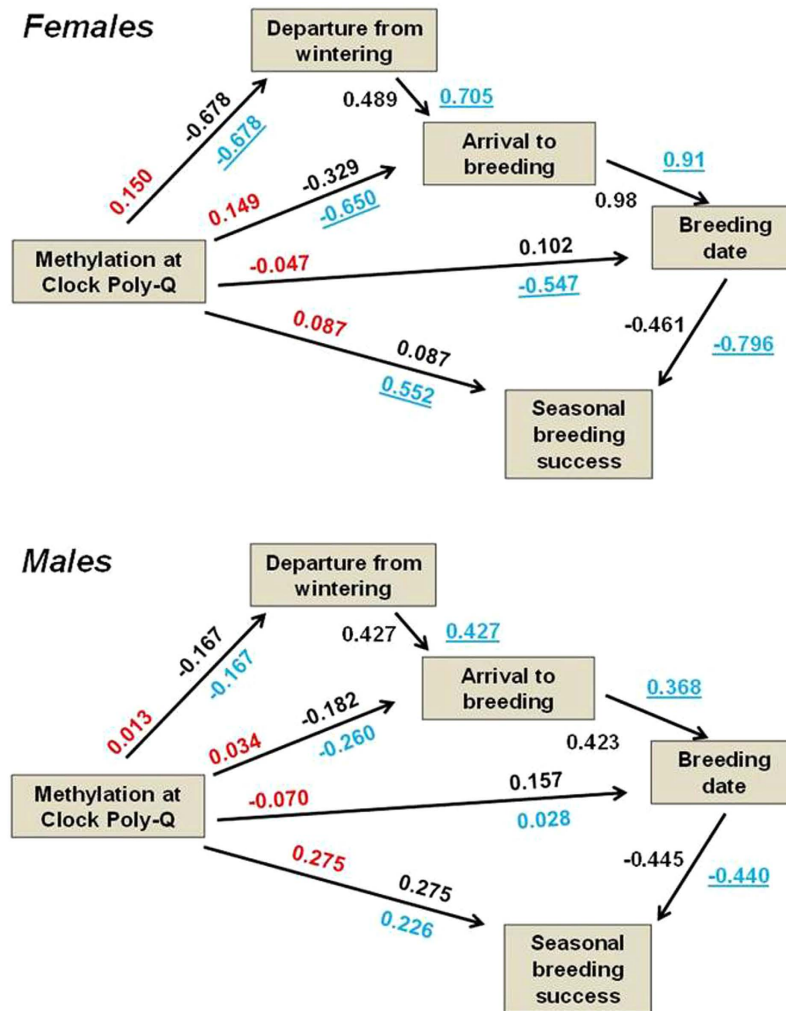


Figure 3. Path diagram of the direct or the indirect effect of methylation at the poly-Q mediated by spring migration phenological variables and breeding date on breeding success for females and males. Black numbers: path coefficients (i.e. the standard partial regression coefficient that estimates the strength of the relationship between each putative causative variable and the corresponding effect). Red numbers: products of the chains of path coefficients along all the paths connecting methylation at *Clock* poly-Q and seasonal breeding success; the sum of these products is used to compute the compound correlation between methylation at *Clock* poly-Q and seasonal breeding success (see Results). Blue numbers: simple correlation coefficients between pairs of variables. Underlining of correlation coefficients indicates statistical significance ($P < 0.05$). See Tables 1 and 2 for sample sizes.

could constrain early departure from the wintering grounds, thereby generating carry-over effects on timing of subsequent migration and arrival to the breeding site and, ultimately, affecting seasonal breeding success³⁷. From a different, functional perspective, however, variation in methylation could be a mechanism enforcing *adaptive* variation in timing of start of migration. Decisions on timing of migration may have to be tuned according to the ecological conditions experienced during wintering and their effects on individual physiological state or, for example, the progress of the single annual molt that barn swallows normally complete during wintering⁴³. Ecological conditions during wintering may directly affect methylation at *Clock* thereby allowing barn swallows to tune their migration phenology according to their physiological state. Hence, variation in methylation may serve as an adaptive, condition-dependent epigenetic control mechanism of individual timing of migration. Under the hypothesis that variation in methylation depends on ecological conditions during wintering, an ultimate, functional interpretation of the differentially larger effect of methylation at *Clock* poly-Q on phenology of females compared to males is that performance during spring migration and subsequent breeding performance of females is more dependent on the carry-over effects of condition during wintering as compared to males, thereby requiring stronger epigenetic control³⁷ (see also Fig. 3).

The proximate mechanisms that cause variation in methylation levels between barn swallows and sex-dependence of the effects of methylation on phenology are open to speculation. Methylation of DNA has been shown to be affected by a host of environmental factors, ranging from nutrition to psychological stress and exposure to environmental pollution^{30,44}, but our knowledge of which factors are effective on animals in the wild

is nihil. Yet, ample variation in nutritional conditions and exposure to diverse form of environmental stress may exist among individual barn swallows during the wintering period, making any potential effects of such exogenous factors on methylation levels most likely to operate.

Intriguingly, in both sexes seasonal breeding success was predicted by methylation levels at *Clock* poly-Q, but path analysis suggested partly different causative links between methylation, spring migration and breeding phenology, and seasonal breeding success in either sex. In females, positive carry-over effects of methylation levels on breeding success were largely mediated by timing of spring migration and arrival to the breeding site, which depends on methylation and strongly predicts seasonal reproductive success mostly via an effect on number of breeding events per season. Indeed, early breeding translates into larger number of broods and thus reproductive output per breeding season³⁷. In males, the relationship between methylation and seasonal breeding success was weaker compared to females, with a 6-fold difference in the variance in seasonal reproductive success being explained by methylation in either sex. Contrary to females, however, according to path analysis in males this relationship was mostly a direct one, being largely independent of any relationship between timing of spring migration and methylation. Hence, for the first time our study shows that methylation at *Clock* poly-Q have consistent implications for seasonal reproductive in both sexes, but the mechanisms underpinning this relationship are different in either sex.

Epigenetic processes may boost the evolutionary potential of populations when faced with environmental challenges²⁷. The present results therefore have a bearing on the interpretation of the consequences of ongoing environmental change, including rapid climate change, on animal populations. Long-term studies of widely diverse organisms, like plants, insects and vertebrates have demonstrated that average phenological traits are rapidly changing within populations^{45–47}. These temporal phenological trends have been largely interpreted as a response to climate change, and specifically to increase in temperatures at temperate and high boreal latitudes⁴⁸. The nature of such responses, however, is far from being ascertained. Micro-evolutionary change has often been invoked^{48,49}, although change in the genetic composition of populations has not been demonstrated, and has only been inferred indirectly. In addition, some estimates of the rate of evolutionary change in bird phenology would be inconsistent with a ‘sustainable evolution’ scenario, with selection for phenological change being not compatible with population persistence, according to theoretical models^{50,51}. Our results suggest that temporal phenological trends may arise as a consequence of epigenetic changes in the orchestration of photoperiodic genes that control phenology. Future ecological studies may therefore seek for the ultimate causes and mechanisms that can translate environmental experience into variation of epigenetic DNA modifications. In addition, they should strive to assess whether epigenetic processes have the potential to cause trans-generational effects by generating heritable variation phenology and other fitness-related traits. Finally, in the context of the study of climate change effects, they should search for empirical evidence that methylation at phenological genes can have consequences for long-term temporal variation in phenology of populations.

Methods

Model species. The barn swallow is a small (ca. 20 g), long-distance migratory passerine bird^{52,53}. Western Palearctic breeding populations almost exclusively winter south of the Sahara Desert, with some migratory connectivity⁵⁴. Autumn migration occurs in August–October whereas pre-breeding, spring migration occurs in February–May^{53,55}. Barn swallows are aerially insectivorous and socially monogamous, and mostly breed semi-colonially in rural buildings. Breeding pairs have up to 3 clutches (1–7 eggs each) per breeding season.

Field procedures. We studied barn swallows at a total of 29 breeding colonies (= farms) in southern Switzerland (46°09′N, 8°55′E) and northern Italy (two neighbouring areas: 45°33′N, 8°44′E; 45°19′N, 9°40′E)^{55,56}, during spring-summer 2010–2012. In all study years, we captured the adults at the breeding colonies and individually marked them with numbered metal and color rings. We identified the composition of the breeding pairs by direct observation of individual markings at the nest, and monitored the breeding activities to record breeding date (= Julian date of laying of the first egg in the clutch), and clutch size (number of eggs at clutch completion). Laying date of the first clutch was chosen as the best proxy for seasonal timing of start of breeding (‘breeding date’). Total number of eggs laid by a female or, for males, by their social female mate was computed as the sum of the size of the clutches over the entire breeding season. Seasonal total number of eggs was chosen as the best proxy for seasonal fecundity, because nestling mortality is very low in our study areas (<5% of the nestlings in broods that produce any nestlings) and hatching failure rate is also low (7–10%) (N.S. and C.S. personal observation on the focal study population; see also ref. 52). Hence, clutch size strongly determines seasonal breeding success³⁷.

Full details of the geolocator deployment procedures have been reported in related studies^{55,56}. Briefly, in summer 2010 and 2011, light-level geolocators (SOI-GDL2.10; SOI-GDL2.11) were deployed using a leg-loop harness^{55,56}. Geolocators were then removed upon first capture of the individuals returning from migration in the year (2011 or 2012) following that of geolocator deployment. Thus, phenological information refers to wintering and breeding seasons preceding geolocator recovery. Because geolocators were deployed on adult birds in spring-summer, all the phenological information refers to individuals older than 1 year, implying that age effects, which are known to affect phenology during the first year after hatching⁵² should have not affected the results.

Phenological variables. Light data recorded by light-level geolocators were used to obtain information on phenological variables as reported in a previous study⁵⁵. For the purposes of the present study we estimated the departure date from the wintering area, as the Julian date of the end of the (last) ‘stationary period’ in the wintering range south of the Sahara Desert; and the arrival date to the breeding site, as the Julian date of arrival to the breeding colony in the year when the geolocator was recovered.

Analysis of methylation at Clock poly-Q and 5'-UTR. Upon capture we collected a blood sample from all individuals by puncturing the brachial vein for methylation analysis in blood cells. Genomic DNA from clotted whole blood, previously rehydrated with an aqueous solution containing 20% sodium dodecyl sulphate (SDS) and 1 M Tris (pH 8.0), was extracted using the QIAamp DNA Mini kit 250 (#51306). The analysis of methylation was conducted on two CG loci of the *Clock* gene: one close to the ATG start-codon in the 5'-UTR and the second in the last exon of the gene where the poly-Q region is present (hereafter 'poly-Q' for brevity). As the complete genomic sequence of *Clock* in the barn swallow is not available, the two loci were identified as previously described in details³⁰.

DNA methylation was quantified by bisulfite-PCR and pyrosequencing⁵⁷. Bisulfite treatment deaminates unmethylated cytosine to produce uracil in DNA, while methylated cytosines are protected from the conversion to uracil, allowing a quantification of DNA methylation⁵⁸. Briefly, the samples were bisulfite-treated using EZ DNA Methylation-Gold™ Kit (Zymo Research, Orange, CA, USA) and PCR-amplified using primers and conditions illustrated in Romano *et al.*³⁰. The degree of methylation was expressed as percentage of 5-methyl-cytosines (%5mC) over the sum of methylated and unmethylated cytosines. Every sample was tested in duplicate for each marker to confirm reproducibility of the degree of *Clock* methylation. Repeatability estimates (intra-class correlation coefficient) were 0.481 ($\chi^2_1 = 18.26$, $P < 0.001$) for *Clock* poly-Q exon and 0.413 ($\chi^2_1 = 10.71$, $P < 0.001$) for 5'-UTR.

Statistical analyses. We used Gaussian linear mixed models to analyze variation in phenological variables and seasonal number of eggs in relation to sex (fixed effect) and percentage methylation at either focal loci (continuous covariate). In the models, the interaction term between percentage methylation and sex was initially included, and then excluded when statistically non-significant. In all models, year (2 levels) and area (3 levels) were included as random factors. The effect of year or area were tested by likelihood ratio tests comparing the model including or, respectively, excluding each random effect at a time.

Path analysis⁵⁹ was used to describe the direct and the indirect effects, as mediated by phenological variables, of methylation on breeding success. In reporting on the results of path analysis we show: 1) the path coefficients (i.e. the standard partial regression coefficient that estimates the strength of the relationship between each putative causative variable and the corresponding effect; see numbers in black in Fig. 3); 2) the products of the chains of path coefficients along all the paths connecting methylation at *Clock* poly-Q and seasonal breeding success (numbers in red in Fig. 3). The sum of these products was used to compute the compound correlation between methylation at *Clock* poly-Q and seasonal breeding success; and 3) the simple correlation coefficients between pairs of variables (numbers in blue in Fig. 3).

Parameter estimates are reported with their associated standard error.

Sample sizes for the different analyses (see Tables 1 and 2) slightly differ because phenological/breeding success information was not available for some individuals, as did the information for methylation.

All methods were carried out in accordance with the relevant guidelines and regulations. All experimental protocols were approved by the Istituto Superiore per la Protezione dell'Ambiente, and by Regione Lombardia, Regione Piemonte and by the Swiss Federal Office for Environment.

Data Availability. The datasets generated during the current study are available from the corresponding author on reasonable request.

References

- McNamara, J. M. & Houston, A. I. Optimal annual routines: behaviour in the context of physiology and ecology. *Phil. Trans. R. Soc. Lond. B* **363**, 301–319 (2008).
- Nussey, D. H., Wilson, A. J. & Brommer, J. E. The evolutionary ecology of individual phenotypic plasticity in wild populations. *J. Evol. Biol.* **20**, 831–844 (2007).
- Both, C., van Asch, M., Bijlsma, R. G., van den Burg, A. B. & Visser, M. E. Climate change and unequal phenological changes across four trophic levels: constraints or adaptations. *J. Anim. Ecol.* **78**, 73–83 (2009).
- Riyahi, S., Sánchez-Delgado, M., Calafell, F., Monk, D. & Senar, J. C. Combined epigenetic and intraspecific variation of the DRD4 and SERT genes influence novelty seeking behavior in great tit *Parus major*. *Epigenetics* **10**, 516–525 (2015).
- Verhulst, E. C. *et al.* Evidence from pyrosequencing indicates that natural variation in animal personality is associated with DRD4 DNA methylation. *Mol. Ecol.* **25**, 1801–1811 (2016).
- Baerwald, M. R. *et al.* Migration-related phenotypic divergence is associated with epigenetic modifications in rainbow trout. *Mol. Ecol.* **25**, 1785–1800 (2016).
- Vaz Nunes, M. & Saunders, D. Photoperiodic time measurement in insects: a review of clock models. *J. Biol. Rhythms*. **14**, 84–104 (1999).
- Sharp, P. J. Photoperiodic regulation of seasonal breeding in birds. *Ann. N. Y. Acad. Sci.* **1040**, 189–199 (2005).
- Darlington, T. K. *et al.* Closing the circadian loop: clock-induced transcription of its own inhibitors *per* and *tim*. *Science* **280**, 1599–1603 (1998).
- Kyriacou, C. P., Peixoto, A. A., Sandrelli, F. & Costa, R. Clines in clock genes: fine-tuning circadian rhythms to the environment. *Trends Genet.* **24**, 124–132 (2007).
- Panda, S., Hogenesch, J. B. & Kay, S. A. Circadian rhythms from flies to human. *Nature* **417**, 329–335 (2002).
- Cassone, V. M. Avian circadian organization: A chorus of clocks. *Front. Neuroendocrinol.* **35**, 76–88 (2014).
- Gekakis, N. *et al.* Role of the CLOCK Protein in the Mammalian Circadian Mechanism. *Science* **280**, 1564–1569 (1998).
- Saleem, Q., Anand, A., Jain, S. & Brahmachari, S. K. The polyglutamine motif is highly conserved at the Clock locus in various organisms and is not polymorphic in humans. *Hum. Genet.* **109**, 136–142 (2001).
- Liedvogel, M., Szulkin, M., Knowles, S. C., Wood, M. J. & Sheldon, B. C. Phenotypic correlates of Clock gene variation in a wild blue tit population: evidence for a role in seasonal timing of reproduction. *Mol. Ecol.* **18**, 2444–2456 (2009).
- Caprioli, M. *et al.* Clock gene variation is associated with breeding phenology and maybe under directional selection in the migratory barn swallow. *PLoS ONE* **7**, e35140 (2012).
- Bourret, A. & Garant, D. Candidate gene-environment interactions and their relationships with timing of breeding in a wild bird population. *Ecol. Evol.* **5**, 3628–3641 (2015).

18. Saino, N. *et al.* Polymorphism at the *Clock* gene predicts phenology of long-distance migration in birds. *Mol. Ecol.* **24**, 1758–1773 (2015).
19. Liedvogel, M. & Sheldon, B. C. Low variability and absence of phenotypic correlates of *Clock* gene variation in a great tit *Parus major* population. *J. Avian Biol.* **41**, 543–550 (2010).
20. Dor, R. *et al.* Low variation in the polymorphic *Clock* gene poly-Q region despite population genetic structure across barn swallow (*Hirundo rustica*) populations. *PLoS ONE* **6**, e28843 (2011).
21. Chakarov, N., Jonker, R. M., Boerner, M., Hoffman, J. I. & Kruger, O. Variation at phenological candidate genes correlates with timing of dispersal and plumage morph in a sedentary bird of prey. *Mol. Ecol.* **22**, 5430–5440 (2013).
22. Saino, N. *et al.* Timing of molt of barn swallows is delayed in a rare *Clock* genotype. *PeerJ* **1**, e17 (2013).
23. Bazzi, G. *et al.* *Clock* gene polymorphism and scheduling of migration: a geolocator study of the barn swallow *Hirundo rustica*. *Sci. Rep.* **5**, 12443 (2015).
24. Bradshaw, W. E. & Holzapfel, C. M. Phenotypic evolution and the genetic architecture underlying photo-periodic time measurement. *J. Insect Physiol.* **47**, 809–820 (2001).
25. Bradshaw, W. E. & Holzapfel, C. M. What season is it anyway? Circadian tracking vs photoperiodic anticipation in insects. *J. Biol. Rhythms* **25**, 155–165 (2010).
26. Zera, A. J. & Harshman, L. G. The physiology of life-history trade-offs in animals. *Ann. Rev. Ecol. Syst.* **32**, 95–126 (2001).
27. Bossdorf, O., Richards, C. L. & Pigliucci, M. Epigenetics for ecologists. *Ecol. Lett.* **11**, 106–115 (2008).
28. Duncan, E. J., Gluckman, P. D. & Dearden, P. K. Epigenetics, plasticity, and evolution: How do we link epigenetic change to phenotype? *J. Exp. Zool. B* **322**, 208–220 (2014).
29. Jones, P. A. Functions of DNA methylation: islands, start sites, gene bodies and beyond. *Nat. Rev. Genet.* **13**, 484–492 (2012).
30. Romano, A. *et al.* Methylation of the circadian *Clock* gene in the offspring of a free-living passerine bird increases with maternal and individual exposure to PM₁₀. *Environ Poll* (in press) <http://dx.doi.org/10.1016/j.envpol.2016.08.060> (2016).
31. Feng, D. & Lazar, M. A. Clocks, metabolism, and the epigenome. *Mol. Cell* **47**, 158–67 (2012).
32. Mazzi, E. A. & Soliman, K. F. A. Basic concepts of epigenetics. *Epigenetics* **7**, 119–130 (2012).
33. Aguilar-Arnal, L. & Sassone-Corsi, P. The circadian epigenome: how metabolism talks to chromatin remodeling. *Curr. Opin. Cell Biol.* **25**, 170–176 (2013).
34. Eckel-Mahan, K. & Sassone-Corsi, P. Epigenetic regulation of the molecular clockwork. *Prog. Mol. Biol. Transl. Sci.* **119**, 29–50 (2013).
35. Leenen, F. A. D., Muller, C. P. & Turner, J. D. DNA methylation: conducting the orchestra from exposure to phenotype? *Clin. Epigenetics* **8**, 92 (2016).
36. Harrison, X. A., Blount, J. D., Inger, R., Norris, D. R. & Bearhop, S. Carry-over effects as drivers of fitness differences in animals. *J. Anim. Ecol.* **80**, 4–18 (2011).
37. Saino, N. *et al.* Sex-dependent carry-over effects on timing of reproduction and fecundity of a migratory bird. *J. Anim. Ecol.* (in press) (2017).
38. Horvath S. *et al.* Aging effects on DNA methylation modules in human brain and blood tissue. *Genome Biol.* **13**, R97 (2012).
39. Kumar, V., Singh, B. P. & Rani, S. The bird clock: A complex, multi-oscillatory and highly diversified system. *Biol. Rhythm Res.* **35**, 121–144 (2004).
40. O'Malley, K. G. & Banks, M. A. A latitudinal cline in the Chinook salmon (*Oncorhynchus tshawytscha*) *Clock* gene: evidence for selection on PolyQ length variants. *Proc. R. Soc. Lond. B* **275**, 2813–2821 (2008).
41. DiTacchio, L. *et al.* Histone lysine demethylase JARID1a activates CLOCK-BMAL1 and influences the circadian clock. *Science* **333**, 1881–1885 (2011).
42. Azzi, A. *et al.* Circadian behavior is light-reprogrammed by plastic DNA methylation. *Nature Neurosci* **17**, 377–382 (2014).
43. Jenni, L. & Winkler, R. *Moult and ageing of European passerines*. (Academic Press, 1994).
44. Alegria-Torres, J. A., Baccarelli, A. & Bollati, V. Epigenetics and lifestyle. *Epigenomics* **3**, 267–277 (2011).
45. Parmesan, C. Ecological and evolutionary responses to recent climate change. *Annu. Rev. Ecol. Syst.* **37**, 637–669 (2006).
46. Lehikoinen, E. & Sparks, T. H. “Changes in migration” In *Effects of climate change on birds*, Møller, A. P., Fiedler, W., Berthold, P. Eds, pp 89–112 (Oxford University Press, 2010).
47. Dunn, P. O. & Winkler, D. W. “Effects of climate change on timing of breeding and reproductive success in birds” In *Effects of climate change on birds*, Møller, A. P., Fiedler, W., Berthold, P. Eds, pp 113–128 (Oxford University Press, 2010).
48. Bussièrè, E. M. S., Underhill, L. G. & Altwegg, R. Patterns of bird migration phenology in South Africa suggest northern hemisphere climate as the most consistent driver of change. *Global Change Biol.* **21**, 2179–2190 (2015).
49. Sheldon, B. C. “Genetic perspectives on the evolutionary consequences of climate change in birds” In *Effects of climate change on birds*, Møller, A. P., Fiedler, W., Berthold, P. Eds, pp 149–168 (Oxford University Press, 2010).
50. Lynch, M. (1996) “A quantitative genetic perspective on conservation issues” in *Conservation Genetics*, Avise, J. C., Hamric, J. L. Eds, pp. 471–501 (Chapman and Hall, New York, 2010).
51. Gienapp, P., Leimu, R. & Merilä, J. Responses to climate change in avian migration time – microevolution versus phenotypic plasticity. *Clim. Res.* **35**, 25–35 (2007).
52. Møller, A. P. *Sexual selection and the barn swallow*. (Oxford University Press, 1994).
53. Cramp, S. *The Complete Birds of the Western Palearctic on CD-ROM*. (Oxford University Press, 1998).
54. Ambrosini, R., Møller, A. P. & Saino, N. A quantitative measure of migratory connectivity. *J. Theor. Biol.* **257**, 203–211 (2009).
55. Liechti, F. *et al.* Timing of migration and residence areas during the non-breeding period of barn swallows *Hirundo rustica* in relation to sex and population. *J. Avian Biol.* **46**, 254–265 (2015).
56. Scandola, C. *et al.* Impact of miniaturized geolocators on barn swallows (*Hirundo rustica*) fitness traits. *J. Avian Biol.* **45**, 417–423 (2014).
57. Yang, A. S. *et al.* A simple method for estimating global DNA methylation using bisulfite PCR of repetitive DNA elements. *Nucleic Acid Res.* **32**, e38 (2004).
58. Frommer, M. *et al.* A genomic sequencing protocol that yields a positive display of 5-methylcytosine residues in individual DNA strands. *Proc. Natl. Acad. Sci. USA* **89**, 1827–1831 (1992).
59. Sokal, R. R. & Rohlf, F. J. *Biometry*. (W. H. Freeman and Company, New York, 2008).

Acknowledgements

We are grateful to farm owners and to many field assistants. We thank the Fondazione Bolle di Magadino for support to C.S., and the ‘Ente di gestione delle aree protette del Ticino e del Lago Maggiore’ for help with fieldwork. The study was funded by the EU INTERREG program (project ID 15 7624065), Fondazione Cariplo (grant UNIAGI 13357 to N.S.), Milan Univ. (grant 2009-ATE-0015 to DR), and Univ. of Milano-Bicocca (grant 2011-ATE-0272 to RA). The Swiss federal office for environment contributed financial support for the development of the data loggers (UTF-Nr. 254, 332, 363, 400).

Author Contributions

N.S.: field work, data analysis, paper writing; R.A., M.C., M.P., A.R., M.R., C.S., L.G., D.R.: field work; B.A., B.D.G., E.G., L.G., V.B.: methylation analysis; F.L.: data analysis.

Additional Information

Competing Interests: The authors declare no competing financial interests.

How to cite this article: Saino, N. *et al.* Migration phenology and breeding success are predicted by methylation of a photoperiodic gene in the barn swallow. *Sci. Rep.* 7, 45412; doi: 10.1038/srep45412 (2017).

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2017