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# A Review on the Role of EPA and DHA Through Goat Nutrition to Human Health: Could they be Effective both to Animals and Humans?

**Abstract**

Goat's milk is high digestible and useful for a number of human health problems, but this product could be further increased in its beneficial effects on immune response both at animal and human level through the use of dietary n-3 polyunsaturated fatty acids (PUFAs) sources in goat diet.

In the last decade, the interest on n-3 PUFAs, in particular eicosapentaenoic acid (EPA, C20:5) and docosahexaenoic acid (DHA, C22:6), has increased due to their proved positive effects on human health. These two PUFAs must be provided with the diet both in human and animals. Anyway EPA and DHA can be successfully transferred into milk and thus can be available to human consumption with positive related effects on immune response. But what is the best dietary source of EPA and DHA in goat diet? The most promising and investigated raw material seems to be fish oil, but the interest in alternative n-3 PUFAs-rich sources, such as algae, is increasing. The EPA and DHA mechanisms of action, affecting metabolic and pathological pathways, must be still elucidated in goats, especially for what concern the use of dietary algae. The inclusion of fish oil and algae in the diet of dairy goat can increase the healthy properties of milk with consequential health benefit in human.

**Keywords:** Polyunsaturated fatty acids; Fish oil; Algae; Goat; Human; Immune response

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**Farina G[1], Cattaneo D[1], Lecchi C[2], Invernizzi G[1], Savoini G[1] and Agazzi A[1]**

[1]*Dipartimento di Scienze veterinarie per la salute, Università degli Studi di Milano, Italy*

[2]*Dipartimento di Patologia Animale, Igiene e Sanità Pubblica Veterinaria, Università di Milano, Italy*

***Corresponding author:** Alessandro Agazzi, Dipartimento di Scienze veterinarie per la salute, la produzione animale e la sicurezza alimentare, Università degli Studi di Milano, Milano, Italy. via Celoria 10, 20133 Milan, Italy, Tel: +39 0250318038; Fax: +39 0250317819; E-mail: alessandro.agazzi@unimi.it

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**Abbreviations:** UFA: Unsaturated Fatty Acids; PUFA: Polyunsaturated Fatty Acids; NEFA: Non Esterified Fatty Acids; EPA: Eicosapentaenoic Acid; DHA: Docosahexaenoic Acid; CLA: Conjugated Linoleic Acid; DMI: Dry Matter Intake; NRC: National Research Council; $PGE_2$: prostaglandin $E_2$; $TXB_2$: Thromboxane $B_2$; $LTB_4$: Leukotriene $B_4$; $LTE_4$: Leukotriene $E_4$ ; MUFAs: Monounsaturated Fatty Acids; ROS: Reactive Oxygen Species; PMN: PolyMorphoNuclear; LPS: LipoPolySaccharide; CLA: Conjugated Linoleic Acid

## Introduction

High nutritional value of goat's milk and its higher digestibility than cow's milk due to the smaller fat globules size are evident since years [1] and represent a good challenge in resolving a number of human health problems [2,3] such as malabsorption syndrome [4] or high total and LDL cholesterol levels in human blood. Goat's milk has a low content of polyunsaturated fatty acids (PUFAs; 2-6% of fatty acid composition) and a relatively high amount of saturated fatty acids (SFAs; 53-72%); the remaining part is represented by monounsaturated fatty acids (MUFAs) [5].

The rising question is: how can we further improve this unique animal product and contribute to human health beside goat's milk natural properties?

One of the major chances we have is to increase n-3 PUFAs milk content since there is evidence that these fatty acids have the potential to improve long-term human health [6]. Moreover the western diet is very high in n-6 fatty acids relative to n-3 fatty acids [7,8]. The ratio n-6/n-3 PUFAs recommended is less than 5 [9,10] and today this ratio in animal products is between 10 and 15 [11]. This is the point, but what kind of n-3 PUFAs do we want to increase? And more: what is the most interesting source of these fatty acids?

Among all n-3 PUFAs, eicosapentaenoic acid (EPA, C20:5) and docosahexaenoic acid (DHA, C22:6), have been shown to be essential in mammals species leading to nutritional and health beneficial actions [12], often with anti-inflammatory properties [13].

If we account for human health, DHA leads to higher neuronal development in foetal and early infants [14], but the endogenous production of this fatty acid by the mother from α-linoleic acid (ALA), as example, is very poor [15] due to the competition with linoleic acid (LA, 18:2 n-6) for the same metabolizing enzymes, particularly at the first Δ6-desaturation step [16]. In human both EPA and DHA must then be introduced with the diet and the consumption of milk with increased EPA and DHA content can be a good chance. Unfortunately the content of these two fatty acids is minimal in traditional ruminant diets and consequently milk has a very low amount of them (<0.1% of total fatty acids) [17] but improving EPA and DHA content in the diet of dairies could lead to an higher available content of these fatty acids in milk for human consumption.

What is the best source of EPA and DHA in animal nutrition? During years a big interest has been pointed out on fish oil that seems the most promising raw material for this purpose due to the fact that it mostly contains EPA and DHA among all the other n-3 PUFAs. Anyway, fish oil is a costly supplementation of lipids compared to other more conventional fatty acid sources and there is increasing concern on the sustainability of the use of fish products in animal diet [18], which leads to consider alternative sources of long-chain n-3 PUFAs [19].

One of these alternatives is the use of a source of α-linoleic

acid (ALA) as a precursor for EPA and DHA such as linseed, that anyway is affected by two main disadvantages:

a. The low digestibility of linseed due to the fact that the digestive enzymes cannot penetrate the seed coat, usually solved with treatments like extrusion and expansion;

b. The presence of anti nutritional factors, which however can be destroyed by thermo-extrusion in some extent [11].

Beside linseed, algae have high content of PUFAs and can be considered as an alternative to fish oil [20]. They have several advantages: easy to cultivate, relatively low cost, high yielding and fast in growth. Moreover these compounds were found to have some interesting bioactive components such as antioxidants, sulfated polysaccharides, phlorotannins, diterpenes, minerals and vitamins, high protein content, valuable nutrients [5,21]. Thus, the use of algae as a dietary additive in livestock diets could be of great interest [22] especially with the aim of decrease SFAs content through an increased content of conjugated linoleic acid (CLA) and other PUFAs in the milk of ruminants [23].

It is recognized that algal lipids have a higher PUFAs content than terrestrial plants [24] although some differences in the fatty-acids composition among species and intra-species variations occurs also related to environmental and geographical factors [25,26], season [27], and algal parts [28].

The use of algae as PUFAs source is a very new topic and the scientific investigations on algae in dairy goat nutrition are very few, but the interest is continuously increasing thanks to several health benefits found in other species through their prebiotic functions, antimicrobial activities, improved digestibility, antioxidant role and anti-inflammatory and immunomodulatory properties [21].

## Dietary n-3 PUFAs: Milk yield, Milk fatty acid composition, EPA and DHA Transfer Efficiency

During years, goat milk yield and composition responses, when adding n-3 PUFAs in the diet, often produced contradictory results. Some studies [12,29,30] found no effects on milk production when feeding fish oil, but the supplementation significantly reduced milk fat. Differently Kitessa et al. [31] evidenced a significant milk yield decrease without changes in milk fat content in response to dietary tuna oil. To the author's knowledge very few literature is available on the effect of dietary algae on production and quality of goat milk [5], but generally no differences in milk yield and a significant decrease in milk fat content in cow [32] and sheep [23,33] was observed.

In this view, one possible mechanism for diet-induced milk fat depression in dairy ruminants involves direct inhibition of lipid synthesis in the mammary gland in response to fatty acid intermediates such as trans-C18:1 and related metabolites, formed during partial biohydrogenation of unsaturated fatty acids in the rumen [33,34]. The fatty acid composition of milk can be seen as the result of both the tissue fatty acid biosynthesis and the dietary fatty acid composition of the diet [35] but, if in monogastrics animals the relationship between dietary fatty acid profile and milk fatty acid content is quite stronger and linear, this is not the same for ruminants because rumen microorganisms are responsible for the biohydrogenation process of dietary fatty acids that leave the rumen and pass in the small intestine, where they can be absorbed [20,36,37].

The administration of both fish oil and algae in ruminant are reported to strongly affect the fatty acid profile of milk where the observed results are a decrease in SFAs with increased PUFAs content. More in details, the fatty acid profile of mature goat milk was influenced by unprotected fish oil administration with a lower average concentration of C18:0 and higher content of C16:1, C18:3, and very long-chain n-3 PUFAs EPA and DHA [12].

The use of algae in the diet of dairy goat are reported to decrease palmitic acid content in the milk with increased levels of oleic, linoleic and linolenic acid and a positive change in SFA:MUFA:PUFA with a reduced proportion of SFA and a tendency to a positive shift in n-6:n-3PUFA, with increasing proportion of n-3 [5]. The observed increase in PUFAs milk content [23,33] in dairy sheep when feeding algae is relative to an higher content of very-long chain PUFAs and particularly EPA and DHA, according with what previously observed in cow [32].

Thus, EPA and DHA content in milk can be significantly enhanced through goat nutrition, but what is the extent to which these fatty acids are transferred from the diet into small ruminants' milk fat? The question is still debated because it mainly accounts for the source of n-3 PUFAs and if a rumen protection is applied over these compounds.

Beside the different dietary sources, the observed discrepancies in transfer efficiencies have been explained by the dosage applied, the duration of administration, and level/technological methods for rumen protection of the dietary fatty acid sources. With regard to this last point the transfer efficiency of EPA and DHA from diet into milk is however generally low when not rumen protected because of the high amount of rumen biohydrogenation together with the preferential incorporation into plasma phospholipids and cholesterol esters [31].

Anyway, the alternative to introduce "inert" fats in the diet rather than the native forms of supplementation to escape or bypass fat rumen biohydrogenation is preferable because a more stable form can preserve fatty acids form oxidation and the inclusion in the diet of even poorly protected fats can reduce the negative effects of high levels of unprotected dietary fats on feed intake, ruminal fermentation, and milk fat content [38].

At the moment it seems that the transfer efficiency to mature goat milk ranges from 7.0% to 14.0% for EPA and from 7.0% to 8.1% for DHA when a rumen unprotected form of dietary fish oil is used [39], although some works found lower EPA transfer efficiency ranging from 2.8% to 7.9% [30,31,40,41] and a greater variability for DHA rate of passage from 2.8% [30] to 20.3% [40].

Considering algae dietary supplementation, DHA transfer efficiency in dairy cow was 8.4% when using an unprotected form of algae, and 16.7% when a rumen-protected form was fed [32]. Similarly, the DHA transfer rate into sheep' milk from unprotected algae compound was between 5.0% [33] and 8.0%, while 30% to 60% of dietary EPA was incorporated [23].

## Improving Immune Response with EPA and DHA

Fatty acids metabolism plays a significant role in immune cells functions, both in humans and in animals [42]. The most powerful immunomodulatory effect is achieved by the n-3 PUFAs and, among them, by EPA, C20:5(n-3) and DHA C22:6(n-3).

Long chain n-3 PUFAs modulate immune functions in several ways by replacing, for example, arachidonic acid during the eicosanoid signalling cascade [43], thus decreasing the production of inflammatory eicosanoids such as of PGE2 [44], TXB2 [45], LTB4 [46], 5-hydroxyeicosatetraenoic acid [47] and LTE4 [48]. Long chains n-3 PUFAs can also directly interfere with cytokine gene expression [49]. Further regulatory pathways include regulation of cell surface expression of adhesion molecules [50], membrane fluidity and apoptosis rates [51]. In addition, both EPA and DHA give rise to family of anti-inflammatory mediators called resolvins [52]. Most of these activities directly target leukocyte function [53].

Monocytes are recognized as major effector cells of the immune system, playing a central role in the initiation, development and outcome of the innate immune response. Long chain n-3 fatty acids may affect monocyte/macrophage defensive functions in several ways. For example, both EPA and DHA can increase phagocytosis [54] and decrease chemotaxis of human monocytes [55,56]. Cytokine expression can be modulated as well, since both EPA and DHA can down-regulate the *in vitro* production of IL-1β, IL-6 and TNF-α [49,57,58]. Monocytes/macrophages, together with neutrophils, produce high amount of reactive oxygen species (ROS), thus contributing to the oxidative stress, a prominent and common feature of many disease processes. Generation of ROS is of paramount importance for the killing of micro-organism. Yet, if not properly regulated, such as in chronic inflammatory or immune diseases, ROS become hazardous for the organism by causing damage to cellular lipids, proteins and DNA, eventually impairing cells function. Long chain PUFAs modulated cellular production of ROS in macrophages [59], neutrophils [60] and lymphocytes [61], but their activity is still debated, since some authors found an increase while others, on the contrary, found a decrease in ROS production [62].

Regulation of leukocytes apoptosis is another important homeostatic mechanism to control the activity of monocytes, by reducing or increasing their lifespan during inflammation, eventually determining the positive outcome of the inflammatory process. There is increasing evidence that PUFAs can cause cell death in normal and cancer cells [63,64]. Several cell types, including macrophages [62], neutrophils [60] and leukaemia cells [61], exhibit morphological features of apoptosis and necrosis after exposure to PUFAs at high doses. The mechanisms by which PUFAs cause apoptosis vary markedly with cell type and fatty acid species, and most of them are still debated.

As discussed previously, studies carried out in ruminants had the prominent aim to evaluate the possible higher EPA and DHA content in animal-derived products, but there is the increasing evidence from the few available investigations that feeding ruminants with n-3 PUFAs may affect fertility [65] and modulate immune response, including for example, lipopolysaccharide (LPS)-induced hyperthermia [66].

Dietary fish oil in transition dairy goats was found to be effective on cell-mediated immune response [67], with modified mononuclear and polymorphonuclear (PMN) cells ratio as result. Treating cells with DHA exerted [68] in increased PMN phagocytic activity and lower reactive oxygen species (ROS) production after *in vitro* challenged with EPA and DHA. A subsequent validation *in vivo* of the obtained results demonstrated that both EPA and DHA have beneficial effects on goats health by improving the defensive performances of neutrophils [69] avoiding cellular and tissue damages by ROS. EPA and DHA affected also goat monocytes activities by up-regulating phagocytic activity and ROS production [70] and by interfering with the formation of lipid droplets and by upregulating proteins belonging to PAT protein family [71].

## Conclusion

The transfer of health-promoting PUFAs such EPA and DHA can be achieved from dairy goat nutrition to human through the milk with positive effects also at animal level. The increased EPA and DHA milk content represent a future challenge to better characterize an already high-nutritional and health-providing product. Among different EPA and DHA sources, at the present moment fish oil still represents the preferable source, but algae can be an alternative as soon experimental results will be consolidated.

## References

1. Jandal JM (1996) Comparative aspects of goat and sheep milk. Small Rumin Res 22(2): 177-185.
2. Haenlein GFW (2004) Goat milk in human nutrition. Small Rumin Res 51(2): 155-163.
3. Reynolds M (2009) The nutritional benefits of goat's milk. Goat J 87: 23-24.
4. Alférez MJ, Barrionuevo M, López Aliaga I, Sanz-Sampelayo MR, Lisbona F, et al. (2001) Digestive utilization of goat and cow milk fat in malabsorption syndrome. J Dairy Res 68(3): 451-461.
5. Kouřimská L, Vondráčková E, Fantová M, Nový P, Nohejlová L, et.al. (2014). Effect of feeding with algae on fatty acid profile of goat's milk. Sci agri bohemica 45(3): 162-169.
6. Shingfield KJ, Chilliard Y, Toivonen V, Kairenius P, Givens DI (2008) Trans fatty acids and bioactive lipids in ruminant milk. In: Bosze Z (Ed.), Bioactive Components of Milk. Springer, pp. 3-65.
7. Hargis PS, Van Elswyk ME (1993) Manipulating the fatty acid composition of poultry meat and eggs for the health conscious consumer. World's Poult Sci J 49(3): 251-264.
8. Enser M, Richardson RI, Wood JD, Gill BP, Sheard PR (2000) Feeding linseed to increase the n-3 PUFA of pork: fatty acid composition of muscle, adipose tissue, liver, and sausages. Meat Sci 55(2): 201-212.
9. Kouba M, Enser M, Whittington FM, Nute GR, Wood JD (2003) Effect of a high linolenic acid diet on lipogenic enzyme activities, fatty acid composition, and meat quality in the growing pig. J Anim Sci 81(8): 1967-1979.
10. Wood JD, Richardson I, Nute GR, Fisher AV, Campo MM, et al. (2003) Effects of fatty acids on meat quality: a review. Meat Sci 66(1): 21-32.
11. Kouba M, Mourot J (2011) A review of nutritional effects on fat composition of animal products with special emphasis on n-3 polyunsaturated fatty acids. Biochimie 93(1): 13-17.
12. Cattaneo D, Dell'Orto V, Varisco G, Agazzi A, Savoini G (2006) Enrichment in n-3 fatty acids of goat's colostrums and milk by maternal fish oil supplementation. Small Rumin Res 64(1-2): 22-29.
13. Calder PC (2001) The ratio of n-6 to n-3 fatty acids in the diet: impact on T lymphocyte function. Eur J Lipid Sci Technol 103(6): 390-398.
14. Hornstra G (2000) Essential fatty acids in mothers and their neonates. Am J Clin Nutr 71(5 Suppl): 1262S-1269S.
15. Simopoulos AP (2000) Human requirement for n-3 polyunsaturated fatty acids. Poultry Sci 79(7): 961-970.

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16. Kuratko CN, Salem N Jr (2013) Docosahexaenoic acid from algal oil. Eur J Lipid Sci Technol 115(9): 965-976.

17. Lock AL, Bauman DE (2004) Modifying Milk Fat Composition of Dairy Cows to Enhance Fatty Acids Beneficial to Human Health. Lipids 39(12): 1197-1206.

18. Botsford LW, Castilla JC, Peterson CH (1997) The Management of Fisheries and Marine Ecosystems. Sci 277(5325): 509-515.

19. Matthews KR, Horner DB, Thies F, Calder PC (2000) Effect of whole linseed (Linum usitatissimum) in the diet of finishing pigs on growth performance and on the quality and fatty acid composition of various tissues. Br J Nutr 83(6): 637-643.

20. Woods VB, Fearon AM (2009) Dietary sources of unsaturated fatty acids for animals and their transfer into meat, milk and eggs: A review Livestock Sci 126(1-3): 1-20.

21. Maghin F, Ratti S, Corino C (2014) Biological Functions and Health Promoting Effects of Brown Seaweeds in Swine Nutrition. J Dairy Vet Anim Res 1(1): 00005.

22. Rasoul-Amini S, Ghasemi Y, Morowvat MH, Mohagheghzadeh A (2009) PCR amplification of 18S rRNA, single cell protein production and fatty acid evaluation of some naturally isolated microalgae. Food Chem 116(1): 129-136.

23. Papadopoulos G, Goulas C, Apostolaki E, Abril R (2002) Effects of dietary supplements of algae, containing polyunsaturated fatty acids, on milk yield and the composition of milk products in dairy ewes. J Dairy Res 69(3): 357-365.

24. Balboa EM, Conde E, Moure A, Falque E, Dominguez H (2013) *In vitro* antioxidant properties of crude extract and compounds from brown algae. Food Chem 138(2-3): 1764-1785.

25. Li X, Fan X, Han L, Lou Q (2002) Fatty acids of some algae from the Bohai Sea. Phytochem 59(2): 157-161.

26. Narayan B, Miyashita K, Hosakawa M (2004) Comparative Evaluation of Fatty Acid Composition of Different Sargassum (Fucales, Phaeophyta) Species Harvested from Temperate and Tropical Waters. J Aquat Food Prod T 13(4): 53-70.

27. Terasaki M, Hirose A, Narayan B, Baba Y, Kawagoe C, et al. (2009) Evaluation of recoverable functional lipid components of several brown seaweeds (phaeophyta) from Japan with special reference to fucoxanthin and fucosterol contents. J Phy 45(4): 974-980.

28. Khotimchenko SV, Kulikova IV (2000) Lipids of different parts of the lamina of Laminaria japonica aresch. Botanica Marina 43(1): 87-91.

29. El-Nor SAHA, Khattab MSA (2012) Enrichment of milk with conjugated linoleic acid by supplementing diets with fish and sunflower oil. Pak J Biol Sci 15(14): 690-693.

30. Toral PG, Rouel J, Bernard L, Chilliard Y (2014) Interaction between fish oil and plant oils or starchy concentrates in the diet: Effects on dairy performance and milk fatty acid composition in goats. Anim Feed Sci Technol 198: 67-82.

31. Kitessa SM, Gulati SK, Ashes JR, Fleck E, Scott TW, et al. (2001) Utilization of fish oil in ruminants. II. Transfer of fish oil fatty acids into goats' milk. Anim Feed Sci Technol 89(3-4): 201-208.

32. Franklin ST, Martin KR, Baer RJ, Schingoethe DJ, Hippen AR (1999) Dietary marine algae (Schizochytrium sp.) increases concentrations of conjugated linoleic, docosahexaenoic and trans vaccenic acids in milk of dairy cows. J Nutr 129(11): 2048-2054.

33. Toral PG, Hervás G, Gómez-Cortés P, Frutos P, Juárez M, et al. (2010) Milk fatty acid profile and dairy sheep performance in response to diet supplementation with sunflower oil plus incremental levels of marine algae. J Dairy Sci 93(4): 1655-1667.

34. Bauman DE, Griinari JM (2001) Regulation and nutritional manipulation of milk fat: low-fat milk syndrome. Livest Prod Sci 70(1-2): 15-29.

35. Mourot J, Hermier D (2002) Lipids in monogastric animal meat. Reprod Nutr Dev 41(2): 109-118.

36. Jenkins TC (1993) Lipid metabolism in the rumen. J Dairy Sci 76(12): 3851-3863.

37. Doreau M, Chilliard Y (1997) Digestion and metabolism of dietary fat in farm animals. Br J Nutr 78 (Suppl 1): S15-S35.

38. Jenkins TC, Bridges WC (2007) Protection of fatty acids against ruminal biohydrogenation in cattle. Eur J Lipid Sci Technol 109(8): 778-789.

39. Cattaneo D, Ferroni M, Caprino F, Moretti VM, Agazzi A, et al. (2010) Dietary fats in transition dairy goats: effects on milk FA composition. Energy and protein metabolism and nutrition. 3rd EAAP International Symposium on Energy and Protein Metabolism and Nutrition, Parma, Italy, pp. 667-668.

40. Donovan DC, Schingoethe DJ, Baer RJ, Ryali J, Hippen AR, et al. (2000) Influence of dietary fish oil on conjugated linoleic acid and other fatty acids in milk fat from lactating dairy cows. J Dairy Sci 83(11): 2620-2628.

41. Chilliard Y, Glasser F, Ferlay A, Bernard L, Rouel J, et al. (2007) Diet, rumen biohydrogenation and nutritional quality of cow and goat milk fat. Eur J Lipid Sci Technol 109(8): 828-855.

42. Yaqoob P, Calder PC (2007) Fatty acids and immune function: new insights into mechanisms. Br J Nutr 98 (Suppl 1): S41-S45.

43. Calder PC (2006) N-3 polyunsaturated fatty acids, inflammation, and inflammatory diseases. Am J Clin Nutr 83(6 Suppl): 1505S-1519S.

44. Rees D, Miles EA, Banerjee T, Wells SJ, Roynette CE, et al. (2006) Dose-related effects of eicosapentaenoic acid on innate immune function in healthy humans: A comparison of young and older men. Am J Clin Nutr 83(2): 331-342.

45. Caughey GE, Mantzioris E, Gibson RA, Cleland LG, James MJ (1996) The effect on human tumor necrosis factor and interleukin 1b production of diets enriched in n-3 fatty acids from vegetable oil or fish oil. Am J Clin Nutr 63(1): 116-122.

46. Kelley DS, Taylor PC, Nelson GJ, Schmidt PC, Ferretti A, et al. (1999) Docosahexaenoic acid ingestion inhibits natural killer cell activity and production of inflammatory mediators in young healthy men. Lipids 34(4): 317-324.

47. Endres S, Ghorbani R, Kelley VE, Georgilis K, Lonnemann G, et al. (1989) The effect of dietary supplementation with n-3 polyunsaturated fatty acids on the synthesis of interleukin-1 and tumor necrosis factor by mononuclear cells. N Eng J Med 320(5): 265-271.

48. Von Schacky C, Kiefl R, Jendraschak E, Kaminski WE (1993) n-3 fatty acids and cysteinyl-leukotriene formation in humans *in vitro*, *ex vivo* and *in vivo*. J Lab Clin Med 121(2): 302-309.

49. Weldon SM, Mullen AC, Loscher CE, Hurley LA, Roche HM (2007) Docosahexaenoic acid induces an anti-inflammatory profile in lipopolysaccharide-stimulated human THP-1 macrophages more effectively than eicosapentaenoic acid. J Nutr Biochem 18(4): 250-258.

50. De Caterina R, Libby P (1996) Control of endothelial leukocyte adhesion molecules by fatty acids. Lipids 31 Suppl: S57-S63.

51. Sweeney B, Puri P, Reen DJ (2001) Polyunsaturated fatty acids influence neonatal monocyte survival. Pediatr Surg Int 17(4): 254-258.

**Citation:** Farina G, Cattaneo D, Lecchi C, Invernizzi G, Savoini G, et al. (2015) A Review on the Role of EPA and DHA Through Goat Nutrition to Human Health: Could they be Effective both to Animals and Humans?. J Dairy Vet Anim Res 2(2): 00027. DOI: 10.15406/jdvar.2015.02.00027

52. Serhan CN, Hong S, Gronert K, Colgan SP, Devchand PR, et al. (2002) Resolvins: A family of bioactive products of omega-3 fatty acid transformation circuits initiated by aspirin treatment that counter pro-inflammation signals. J Exp Med 196(8): 1025-1037.

53. Sijben JW, Calder PC (2007) Differential immunomodulation with long-chain n-3 PUFA in health and chronic disease. Proc Nutr Soc 66(2): 237-259.

54. Halvorsen DS, Hansen JB, Grimsgaard S, Bønaa KH, Kierulf P, et al. (1997) The effect of highly purified eicosapentaenoic and docosahexaenoic acids on monocyte phagocytosis in man. Lipids 32(9): 935-942.

55. Schmidt EB, Varming K, Pedersen JO, Lervang HH, Grunnet N, et al. (1992) Long-term supplementation with n-3 fatty acids, II: Effect on neutrophil and monocyte chemotaxis. Scand J Clin Lab Invest 52(3): 229-236.

56. Schmidt EB, Varming K, Møller JM, Bülow Pedersen I, Madsen P, et al. (1996) No effect of a very low dose of n-3 fatty acids on monocyte function in healthy humans. Scand J Clin Lab Invest 56(1): 87-92.

57. Babcock TA, Helton WS, Hong D, Espat NJ (2002) Omega-3 fatty acid lipid emulsion reduces LPS-stimulated macrophage TNF-alpha production. Surg Infect (Larchmt) 3(2): 145-149.

58. Zhao G, Etherton TD, Martin KR, Vanden Heuvel JP, Gillies PJ, et al. (2005) Anti-inflammatory effects of polyunsaturated fatty acids in THP-1 cells. Biochem Biophys Res Commun 336(3): 909-917.

59. Yaqoob P, Calder P (1995) Effects of dietary lipid manipulation upon inflammatory mediator production by murine macrophages. Cell Immunol 163(1): 120-128.

60. Healy DA, Watson RW, Newsholme P (2003) Polyunsaturated and monounsaturated fatty acids increase neutral lipid accumulation, caspase activation and apoptosis in a neutrophil-like, differentiated HL-60 cell line. Clin Sci (Lond) 104(2): 171-179.

61. Cury-Boaventura MF, Pompéia C, Curi R (2005) Comparative toxicity of oleic acid and linoleic acid on Raji cells. Nutrition 21(3): 395-405.

62. Martins de Lima T, Cury-Boaventura MF, Giannocco G, Nunes MT, Curi R (2006) Comparative toxicity of fatty acids on a macrophage cell line (J774). Clin Sci (Lond) 111(5): 307-317.

63. Sweeney B, Puri P, Reen DJ (2007) Induction and modulation of apoptosis in neonatal monocytes by polyunsaturated fatty acids. J Pediatr Surg 42(4): 620-628.

64. Heimli H, Giske C, Naderi S, Drevon CA, Hollung K (2002) Eicosapentaenoic acid promotes apoptosis in Ramos cells via activation of caspase-3 and -9. Lipids 37(8): 797-802.

65. Santos JE, Bilby TR, Thatcher WW, Staples CR, Silvestre FT (2008) Longchain fatty acids of diets as factors influencing re production in cattle. Reprod Domest Anima 43(Suppl 2): 23-30.

66. Farran TB, Reinhardt CD, Blasi DA, Minton JE, Elsasser TH, et al. (2008) Source of dietary lipid may modify the immune response in tressed feeder cattle. J Anim Sci 86(6): 1382-1394.

67. Agazzi A, Cattaneo D, Dell'Orto V, Moroni P, Bonizzi L, et al. (2004) Effect of administration of fish oil on aspects of cell-mediated immune response in periparturient dairy goats. Small Ruminant Research 55(1-3): 77-83.

68. Pisani LF, Lecchi C, Invernizzi G, Sartorelli P, Savoini G, et al. (2009) *In vitro* modulatory effect of omega-3 polyunsaturated fatty acid (EPA and DHA) on phagocytosis and ROS production of goat neutrophils. Vet Immunol Immunopathol 131(1-2): 79-85.

69. Bronzo V, Puricelli M, Agazzi A, Invernizzi G, Ferroni M, et al. (2010) Effects of protected fish oil in the diet of periparturient dairy goats on phenotypic variation in blood and milk leukocytes. Animal 4(9): 1510-1517.

70. Lecchi C, Invernizzi G, Agazzi A, Ferroni M, Pisani LF, et al. (2011) *In vitro* modulation of caprine monocyte immune functions by omega-3 polyunsaturated fatty acids. Vet J 189(3): 353-355.

71. Lecchi C, Invernizzi G Agazzi A, Modina S, Sartorelli P, et al. (2013) Effects of EPA and DHA on lipid droplet accumulation and mRNA abundance of PAT proteins in caprine monocytes. Res Vet Sci 94(2): 246-251.

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