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Murein Lytic Enzyme TgaA of *Bifidobacterium bifidum* MIMBb75 Modulates Dendritic Cell Maturation through Its Cysteine- and Histidine-Dependent Amidohydrolase/Peptidase (CHAP) Amidase Domain

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Bifidobacteria are Gram-positive inhabitants of the human gastrointestinal tract that have evolved close interaction with their host and especially with the host's immune system. The molecular mechanisms underlying such interactions, however, are largely unidentified. In this study, we investigated the immunomodulatory potential of *Bifidobacterium bifidum* MIMBb75, a bacterium of human intestinal origin commercially used as a probiotic. Particularly, we focused our attention on TgaA, a protein expressed on the outer surface of MIMBb75's cells and homologous to other known bacterial immunoactive proteins. TgaA is a peptidoglycan lytic enzyme containing two active domains: lytic murein transglycosylase (LT) and cysteine- and histidine-dependent amidohydrolase/peptidase (CHAP). We ran immunological experiments stimulating dendritic cells (DCs) with the *B. bifidum* MIMBb75 and TgaA, with the result that both the bacterium and the protein activated DCs and triggered interleukin-2 (IL-2) production. In addition, we observed that the heterologous expression of TgaA in *Bifidobacterium longum* transferred to the bacterium the ability to induce IL-2. Subsequently, immunological experiments performed using two purified recombinant proteins corresponding to the single domains LT and CHAP demonstrated that the CHAP domain is the immune-reactive region of TgaA. Finally, we also showed that TgaA-dependent activation of DCs requires the protein CD14, marginally involves TRIF, and is independent of Toll-like receptor 4 (TLR4) and MyD88. In conclusion, our study suggests that the bacterial CHAP domain is a novel microbe-associated molecular pattern actively participating in the cross talk mechanisms between bifidobacteria and the host's immune system.

The human intestinal microbiota comprises more than 1,000 microbial taxa, which through evolution have adopted different strategies to interact with the host, from commensalism or symbiosis to parasitism (1). A well-established and rapidly growing body of literature demonstrates how deeply the intestinal microbiota is involved in several host physiologic dysfunctions such as obesity, diabetes, autoimmune diseases, and cancer (1, 2). Furthermore, there is scientific evidence that certain members of the microbiota more than others play a crucial role in maintaining a physiological homeostasis in the host (3, 4). For instance, bifidobacteria are Gram-positive inhabitants of the human gastrointestinal tract that have evolved a deep interaction with the host (5, 6, 7, 8). In particular, bifidobacteria, which colonize the human intestine immediately after birth, are the predominant taxon of the microbiota of breast-fed infants and affect the maturation of the host's immune system during the neonatal period, especially the T_H1/T_H2 balance (9, 10). Accordingly, a relation has been established between allergic diseases and bifidobacterial colonization (11, 12, 13, 14). In fact, allergic patients have exhibited lower *Bifidobacterium* counts than healthy control subjects. In addition, the species *Bifidobacterium adolescentis* and *Bifidobacterium longum* subsp. *longum* have been isolated from allergic infants as the predominant bifidobacteria, whereas the predominant ones isolated from age-matched healthy infants have been *Bifidobacterium breve*, *Bifidobacterium longum* subsp. *infantis*, and *Bifidobacterium bifidum* (13). These findings testify to a plausible link be-

tween bifidobacteria and atopy/tolerance balance (15, 16), with certain species such as *B. bifidum* being potentially crucial in limiting the development of a long-term T_H2 -skewed immunological memory in infants and, therefore, in preventing allergies. The ability of *B. bifidum* strains to interact with the host immune system has been reported in several studies (17, 18), but very little information is available concerning the molecular components that support the host-*B. bifidum* cross talk (8, 19).

Whereas adaptive immunity recognizes a microorganism by its specific microbial molecular components known as antigens, innate immunity relies on recognizing microbe-associated molecular patterns (MAMPs), that is, conserved structures present within a class of microorganisms but not in the host tissue (20). Several MAMPs have been identified, such as lipopolysaccharide (LPS, also referred to as endotoxin), peptidoglycan, lipoteichoic acid,

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and bacterial CpG DNA motifs. However, many other yet uncharacterized MAMPs govern the intricate mechanisms of host-microbiota interactions and either help maintain or compromise immunological homeostasis. Consequently, it is critically important to study the immunological role of microbial molecular cell components to decipher the operating principles of the immune system. In this regard, our use of a reductionist molecular approach allowed us to define the immunological role of an intramolecular region belonging to an outer surface enzyme, TgaA, identified in an accompanying paper (21) by comparative genomic analysis in strain *B. bifidum* MIMBb75.

MATERIALS AND METHODS

Bifidobacterial culture conditions. Bifidobacteria were grown under anaerobic conditions (Anaerocult A System; Merck, Darmstadt, Germany) at 37°C in prerduced DeMan-Rogosa-Sharpe (MRS) broth (Difco Laboratories, Inc., Detroit, MI) supplemented with 0.05% L-cysteine hydrochloride (cMRS).

Overproduction and purification of TgaA-derived recombinant proteins. All enzymes and reagents for molecular biology reactions were from Fermentas (Vilnius, Lithuania) or New England BioLabs (EuroClone S.p.A., Pero, Italy). Commercial kits for the extraction and purification of nucleic acids were from MoBio Laboratories (Cabru s.a.s., Arcore, Italy) or Qiagen s.r.l. (Milan, Italy). Vectors pET-Tga, pET- Δ CHAP (where CHAP is cysteine- and histidine-dependent amidohydrolase/peptidase) and pET- Δ LT (where LT is lytic murein transglycosylase) (prepared as described in the accompanying paper [21]) were used for the isopropyl- β -D-thiogalactopyranoside (IPTG)-dependent overexpression in *Escherichia coli* of the recombinant proteins Δ SP-TgaA-His (containing both domains of TgaA protein; also named here recombinant TgaA, or rTgaA, and SP_{PEIB}-TgaA in the accompanying paper [21]), protein Δ SP-TgaA- Δ CHAP-His (containing only the LT domain; rLT), and Δ SP-TgaA- Δ LT-His (containing only the CHAP domain; rCHAP), respectively. In brief, mutant *E. coli* strains were grown in 2 \times YT (yeast extract, tryptone) broth at 37°C for 2 h before 1 mM IPTG was added, and incubation was prolonged for 4 h. Afterwards, proteins were extracted under denaturing conditions with PerfectPro Ni-nitrilotriacetic acid (NTA) agarose (5 Prime; Eppendorf Italia, s.r.l., Milan, Italy) according to the manufacturer's instructions. The molecular mass of recombinant Δ SP-TgaA-His, Δ SP-TgaA- Δ CHAP-His, and Δ SP-TgaA- Δ LT-His proteins was confirmed by reverse-phase high-pressure liquid chromatography (RP-HPLC)/electrospray ionization mass spectrometry (ESI-MS) analysis under conditions reported by Taverniti et al. (22). A further purification and removal of residue LPS from recombinant proteins was carried out by analytical or preparative RP-HPLC. After acidification to pH 2 by addition of 0.1% of trifluoroacetic acid (TFA), aliquots of 0.1 ml of Ni-NTA-purified protein were injected into a Waters 600 HPLC system fitted with a VYDAC C₄ (4.6 by 250 mm; 300-Å pore size; 5- μ m particle size) column (Grace, Deerfield, IL). The eluents used for the separation were solvent A, consisting of 0.1% trifluoroacetic acid (TFA) in MilliQ-treated water, and solvent B, consisting of 0.1% TFA in acetonitrile. The protein separation was performed at room temperature by using a linear elution gradient (60% to 30% of solvent A in 25 min). Proteins were eluted at a flow rate of 0.8 ml min⁻¹ and monitored at 220 and 280 nm. Protein-containing fractions were collected, and the degree of purification was checked by SDS-PAGE. Finally, eluted fractions were lyophilized and stored at -20°C.

Generation of Δ SP-TgaA- Δ LT-His-specific antibodies and immunogold labeling. The preparation of an antibody against the recombinant protein Δ SP-TgaA- Δ LT-His (rCHAP) was raised in rabbits by Primm s.r.l. (Milan, Italy). Immunogold labeling was performed as described in the companion paper (21).

Expression of TgaA in *Bifidobacterium longum* NCC2705. All cloning steps for the preparation of the vectors used to transform *B. longum*

NCC2705 were carried out in *E. coli* XL1-Blue (see Fig. S1 in the supplemental material). In detail, the promoter region from phage T5 (PT5) was obtained from vector pGBL8b (23) through digestion with restriction enzymes BamHI and HindIII. The resulting DNA fragment was introduced in vector pUC19 in the same restriction sites, yielding vector pUC-T5. At the same time, the DNA region containing the gene coding for the chloramphenicol acetyltransferase of pC194 (a natural plasmid of *Staphylococcus aureus*) with its original promoter was obtained through PCR with primers catBC-f (5'-CATGGATCCATCGATCTGCA-3'; restriction sites BamHI and ClaI are underlined) and catS-r 5'-TACCCGGAGCTCCTCTAGA-3'; the restriction site SalI is underlined) from vector pGBL8b. The resulting amplicon was digested with BamHI and SalI and cloned in pUC-T5 digested with the same restriction enzymes, yielding vector pUT5cat. Subsequently, the gene *tgaA* was amplified from the chromosomal DNA of *B. bifidum* MIMBb75 with primers tgaDie-f (5'-A ACTTCGTGAGATCTGCCGCGTCCCGCCAT-3'; restriction site BglII is underlined) and tgaC-r (5'-GCTCATCGATTTTACTTTCCTT-3'; restriction site ClaI is underlined); the amplicon was digested with enzymes BglII and ClaI and cloned in sites BamHI and ClaI of vector pUT5cat, yielding vector pUTgaCat. Plasmid pUTgaCat was then digested with HindIII and XbaI; then, the restriction fragment containing the gene *tgaA* was extracted from the agarose gel, purified, and cloned in the same restriction sites of vector pGOSBif33 (24), yielding the final vector pTgaBif5, which was employed for the expression of *tgaA* in *B. longum*. The control vector pT5CatBif21 (empty vector) was prepared according to the same protocol, with the exception of the cloning step for the introduction of gene *tgaA*, which was not carried out. Finally, vectors pTgaBif5 and pT5CatBif21 were introduced in *B. longum* NCC2705 (courteously provided by Nestlé Research Center, Lausanne, Switzerland) as previously described (23, 24).

RT-PCR. The expression of the gene *tgaA* from plasmid pTgaBif5 was verified in *B. longum* NCC2705 by reverse transcription-PCR (RT-PCR). For the extraction of RNA, 20 ml of exponentially growing cultures of *B. bifidum* MIMBb75 were centrifuged for 10 min at 4,000 \times g at 4°C in the presence of RNA-Later (Ambion). The pellet was then immediately frozen in liquid nitrogen and subjected to RNA extraction using a previously described method, which includes a DNase treatment (25). The quality and integrity of the RNA were checked by Experion (Bio-Rad) analysis. cDNA was synthesized using an iScript cDNA synthesis kit (Bio-Rad), according to the supplier's instructions. PCR was performed with primers B-7x (5'-CACAGTTCATACCGTCCACA-3') and Bext-f-II (5'-GTAGTGGTGGTCTCCGTGA-3') according to the following PCR protocol: 1 cycle of 95°C for 3 min, 39 cycles consisting of 95°C for 30 s, 58°C for 40 s and 72°C for 30 s, and a final elongation step of 72°C for 7 min. The obtainment of an amplicon with the expected size of 260 bp demonstrated the presence of *tgaA* mRNA in the recombinant NCC2705 strain.

Preparation and analysis of immunofluorescent *B. bifidum* MIMBb75. *B. bifidum* MIMBb75 cells were cultivated until early stationary growth phase, harvested by centrifugation, washed once with deionized sterile water, and incubated at room temperature for 10 min with Δ SP-TgaA- Δ LT-His antiserum (1:100 diluted in deionized water). Subsequently, bacterial cells were centrifuged, resuspended in a solution of Cy5-conjugated goat anti-rabbit secondary antibody (Molecular Probes, Eugene, OR) (1:10 diluted in deionized water), and incubated for 10 min at room temperature in the dark. Afterwards, stained cells were visualized with a fluorescence optical digital microscope (Leica DM1000; Leica Microsystems, Wetzlar, Germany) at a magnification of \times 1,000.

Study of BMDC activation. Dendritic cells (DCs) were obtained *in vitro* from bone marrow hematopoietic precursors isolated from C57BL/6 mouse femurs as described previously (26). In brief, hematopoietic precursors were recovered from mouse femoral bone marrow and resuspended in conditioned medium (complete medium supplemented with 10% of the growth supernatant of granulocyte-macrophage colony-stimulating factor [GM-CSF]-transduced B16 cells). About 7×10^6 cells were plated in 100-mm suspension plates. The proportion of CD11c⁺ cells

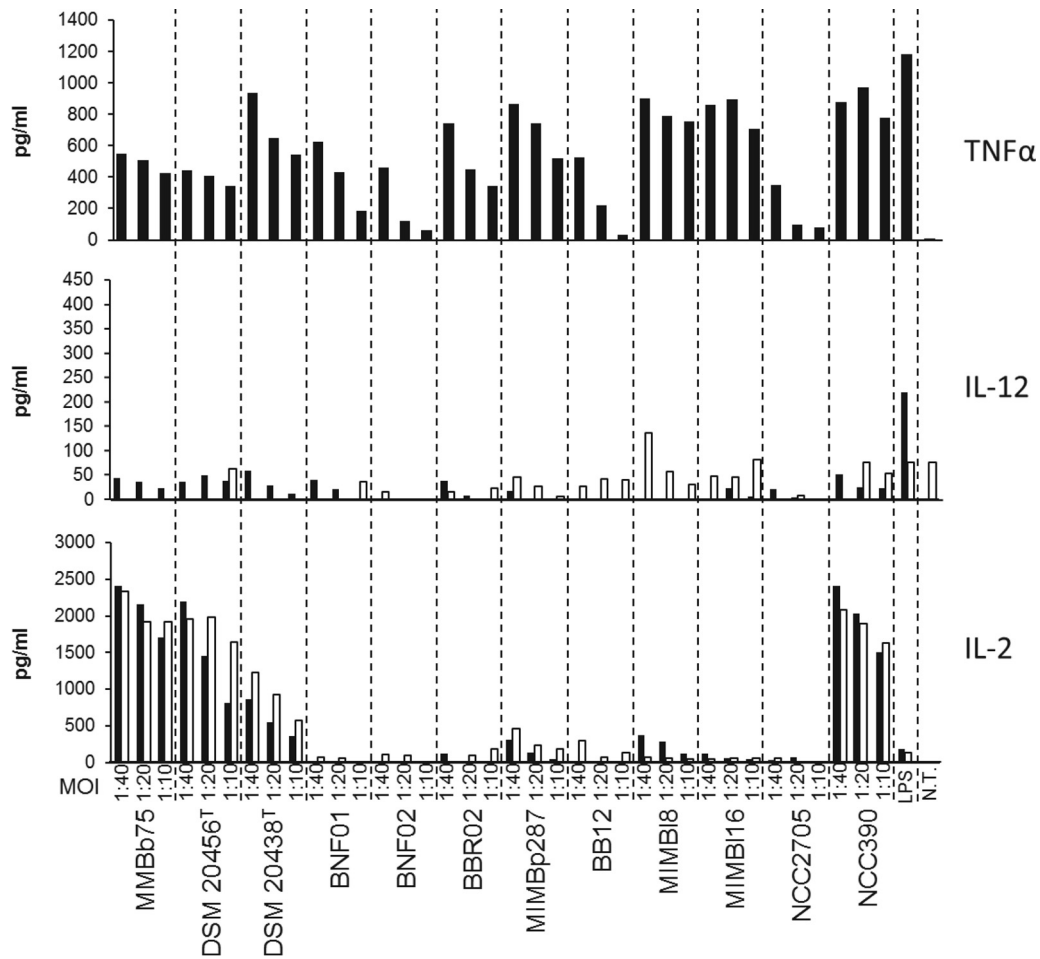


FIG 1 ELISA quantification of cytokines produced by dendritic cells after stimulation with 12 different strains belonging to the genus *Bifidobacterium* and 10 $\mu\text{g/ml}$ of lipopolysaccharides (LPS) from *E. coli*. NT, sample not treated. One representative experiment out of three performed is shown. Black bars, bone marrow-derived dendritic cells (BMDCs); white bars, D1 cell line. All strains were tested at four different multiplicities of induction (MOI; DC/bacteria ratio). Strains belong to the following bifidobacterial species: *B. bifidum* (MIMBb75, DMS 20456^T, NCC390); *B. longum* subsp. *infantis* (BNF01, BNF02); *B. breve* (BBR02); *B. pseudocatenulatum* (DSM20438^T, MIMBp287); *B. longum* subsp. *longum* (MIMB8, MIMB16, NCC2705); *B. animalis* subsp. *lactis* (BB12).

(corresponding to dendritic cells) was monitored periodically by flow cytometry until it reached 90% (ca. 8 days). The bone marrow-derived dendritic cells (BMDCs) were then used for bacterial activation assays. On the day of bacterial infection, BMDCs were plated at a concentration of 0.5 million per ml in 96-well plates (10^5 cells/well). After 1 h, BMDCs were incubated with four different concentrations of bacterial cells for 2 h, washed with saline, resuspended in a culture medium containing penicillin G, streptomycin, tetracycline, and gentamicin, and incubated overnight. Finally, interleukin-2 (IL-2) and tumor necrosis factor alpha (TNF- α) in the supernatant were quantified by enzyme-linked immunosorbent assay (ELISA) using DuoSet kits (R&D Systems, Minneapolis, MN). The same procedure was followed to prepare BMDCs from four mutant mouse lines. Wild-type animals were supplied by Harlan Italy. Ticam1Lps2 (Trif^{-/-}) mice were purchased from The Jackson Laboratory. Myd88^{-/-} and Tlr4^{-/-} mice were provided by S. Akira (IFReC, Japan). Cd14^{-/-} mice were from CNRS d'Orléans (Orléans, France). D1 cell line was cultured in Iscove's modified Dulbecco's medium (IMDM; Sigma, St. Louis, MO) containing 10% heat-inactivated fetal bovine serum (Gibco-BRL, Gaithersburg, MD), 100 IU of penicillin, 100 $\mu\text{g ml}^{-1}$ of streptomycin, 2 mM L-glutamine (all from Sigma), and 50 μM β -mercaptoethanol (in complete IMDM) with 30% supernatant from R1 medium (supernatant from NIH 3T3 fibroblasts transfected with GM-CSF).

Unpaired Student's *t* tests were run to determine statistically significant differences.

RESULTS AND DISCUSSION

Recently, we generated a draft genome sequence of *Bifidobacterium bifidum* MIMBb75, a probiotic strain with demonstrated ability to interact with the host (19, 27–29). Comparative genomics revealed in MIMBb75's genome the presence of a gene encoding TgaA, a peptidoglycan-lytic enzyme which contains two conserved domains: lytic murein transglycosylase (LT; cd00254.3) and cysteine- and histidine-dependent amidohydrolase/peptidase (CHAP; pfam05257.4) (21).

The biomolecular composition of a bacterium's cell wall mainly determines the cross talk processes with the host. In fact, most known bacterial molecules governing bacterial adhesion to epithelia or interaction with immune cells are cell wall constituents (lipopolysaccharides, teichoic and lipoteichoic acids, murein, and proteinaceous adhesins) or cell wall appendages (flagella, pili, and fimbriae) (6, 8, 19, 22). Interestingly, TgaA protein was found to be abundantly expressed and located on the outer surface of

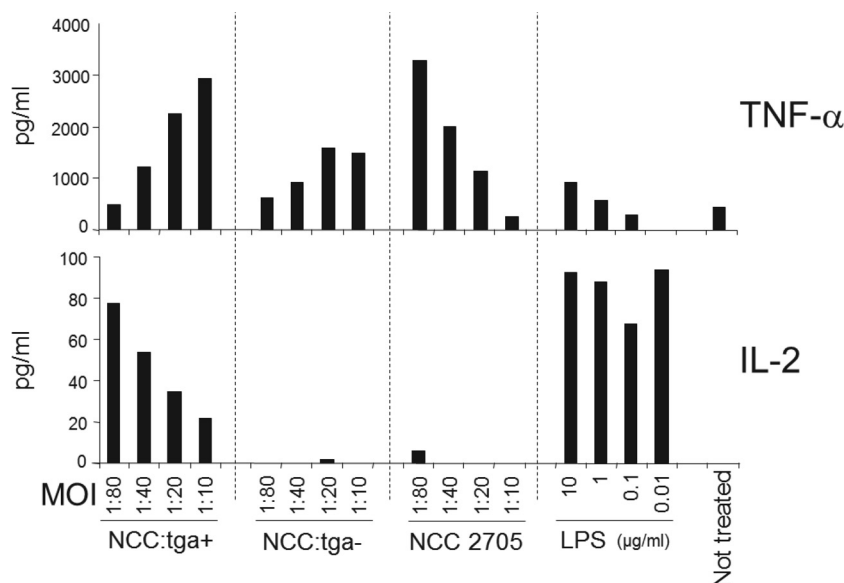


FIG 2 ELISA quantification of cytokine production by BMDCs after stimulation with *E. coli* lipopolysaccharides (LPS), *Bifidobacterium longum* NCC2705, and its recombinant strains NCC:tga+ (expressing TgaA protein) and NCC:tga- (harboring the empty vector). All samples were tested at four different multiplicities of infection (MOI; BMDC/bacteria ratio). One representative experiment out of two performed is shown.

MIMBb75 bacterial cells (21). Furthermore, notably, a search of the Conserved Domain Database (30) for the TgaA protein produced a significant match (E value of $8.30e-23$) between the CHAP module and the conserved domain COG3942, which has been annotated as a surface antigen. The description of COG3942 originated from two studies that identified the CHAP domain-containing protein as an immune-reactive molecule of *Listeria monocytogenes* (31) and *Streptococcus pyogenes* (32). Stimulated by the above data, we evaluated the potential contribution of the TgaA protein to the immunomodulatory activity of the strain MIMBb75.

We assessed the immunological properties of *B. bifidum* MIMBb75 in parallel and independently of the TgaA protein study and discovered through our immunological model a peculiar immunomodulatory capacity of this strain. In particular, we focused on dendritic cells (DCs), the sentinels of the immune system present at various mucosal sites and especially in the intestinal mucosa. At this site, they may sense the intestinal lumen content (including bacteria) via transepithelial processes by responding to the ligation of the DCs' specific microbe recognition receptors (MRRs) to MAMPs (such as LPS, peptidoglycan, and bacterial and viral nucleic acids) to trigger immune responses (33, 34). Particularly, upon activation and migration to secondary lymphoid organs, DCs induce immunocompetence in lymphocytes through antigen presentation and cytokine production. Hence, DCs form a link between innate and adaptive immunity and are considered arbiters of immunological tolerance (34).

In DCs, a major outcome of the ligation of pattern recognition receptors (PRRs) may be the production of interleukin-2 (IL-2), which has several effects. In fact, DC-derived IL-2 increases gamma interferon (IFN- γ) production by the natural killer (NK) cells (35), enhances T cell responses in both mice (26) and humans (36), regulates thymic development (37), and promotes regulatory T cell (Treg) expansion and function (38). Furthermore, it has been linked with T_H1 -skewing stimuli (35). Inducing IL-2

production in DCs can thus steer the adaptive immune system toward preventing T_H2 cell-dominated immune responses, such as those associated with allergy (39, 40).

Our experiments showed that out of 12 bifidobacterial strains belonging to six different species and subspecies, the strain *B. bifidum* MIMBb75 was among the strongest inducers of IL-2 production by murine bone marrow-derived DCs (BMDCs) (Fig. 1).

Subsequently, we studied the potential immunomodulatory activity of TgaA with the same immunological model we used for bifidobacterial cells. Initially, we cloned a complete *tgaA* gene (including the original signal sequence for exporting the protein at a cell wall level) in *Bifidobacterium longum* NCC2705 using a shuttle vector previously developed and used for successful expression of heterologous genes in *B. longum* (23, 24). We followed this strategy because all of our previous efforts to knock out *tgaA* had been unsuccessful since this species has been proved to be recalcitrant to genetic manipulation (41, 42). We selected *B. longum* NCC2705 as the host for *tgaA* expression after analysis of its whole genome revealed that the *tgaA* gene as absent in this human intestinal bacterium. Furthermore, our immunological data (Fig. 1) showed that the strain NCC2705, unlike MIMBb75, is incapable of inducing IL-2 expression by BMDCs. We named the recombinant strain we obtained NCC:tga+. After verifying by reverse transcription-PCR (RT-PCR) and immunogold labeling (see Fig. S2A and B in the supplemental material) that NCC:tga+ expressed the recombinant *tgaA* gene, we incubated BMDCs with supernatant-free wild-type live cells and recombinant NCC2705 strains and then measured by ELISA the levels of the secreted tumor necrosis factor alpha (TNF- α) and IL-2. Secretion by BMDCs of the TNF- α cytokine, the principal marker of DC activation, was induced by all bacterial preparations (Fig. 2). In contrast, our experiments showed a significant secretion of IL-2 only when DCs were stimulated with the recombinant strain NCC:tga+ (i.e., NCC2705 transformed with the vector containing the gene *tgaA*) and not with control preparations (wild-type NCC2705 and NCC2705

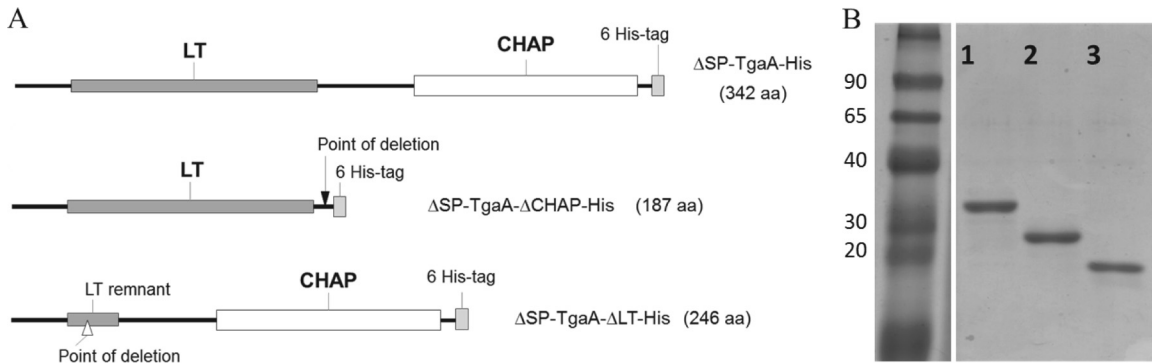


FIG 3 Preparation of TgaA-derived recombinant proteins. (A) Functional map of the recombinant proteins originated from a *tgaA* gene sequence and produced by overexpression in *E. coli* BL21(DE3) pLysS using the pET26b(+) expression vector. (B) SDS-PAGE of the purified six-His-tagged TgaA-derived recombinant proteins. Lane 1, Δ SP-TgaA-His (containing both TgaA domains; rTgaA); lane 2, Δ SP-TgaA- Δ LT-His (containing the CHAP domain; rCHAP), lane 3, Δ SP-TgaA- Δ CHAP-His (containing the LT domain; rLT). The relative molecular masses (in kDa) of standard proteins are indicated on the left. aa, amino acids.

transformed with the empty vector) (Fig. 2). Therefore, our data suggest that the TgaA protein may be directly involved in inducing the cytokine IL-2 in dendritic cells.

To further test our hypothesis, we thoroughly purified from *E. coli* cells the recombinant proteins rTgaA, rLT, and rCHAP by Ni-NTA affinity chromatography and subsequent RP-HPLC to remove any residual LPS and extraneous proteins (Fig. 3). Purified proteins were then used to stimulate two dendritic cell models: BMDCs and the D1 cell line, which is a long-term growth factor-dependent immature myeloid (CD11c⁺ CD8 α ⁻) DC line of splenic origin. Our results show clearly that the rTgaA protein can *in vitro* activate dendritic cells and induce dose-dependent secretion of IL-2 (Fig. 4). A similar stimulatory effect was found with

the recombinant protein rCHAP, whereas rLT activated both dendritic cell models only marginally but could not markedly trigger IL-2 expression (Fig. 4). Therefore, the TgaA protein activates DCs and triggers IL-2 secretion by DCs by means of the CHAP domain.

Finally, we studied the pathway induced by the protein TgaA in DCs. Since TgaA can induce TNF- α production by DCs and since the cluster of differentiation 14 (CD14) protein is essential in inducing such cytokines in LPS-stimulated DCs (43), we first tested if CD14 might have a role also in recognizing TgaA. As shown in Fig. 5, BMDC activation was totally abolished in the absence of CD14. Since CD14 is the best characterized as a coreceptor for Toll-like receptor 4 (TLR4), we then tested if BMDCs isolated

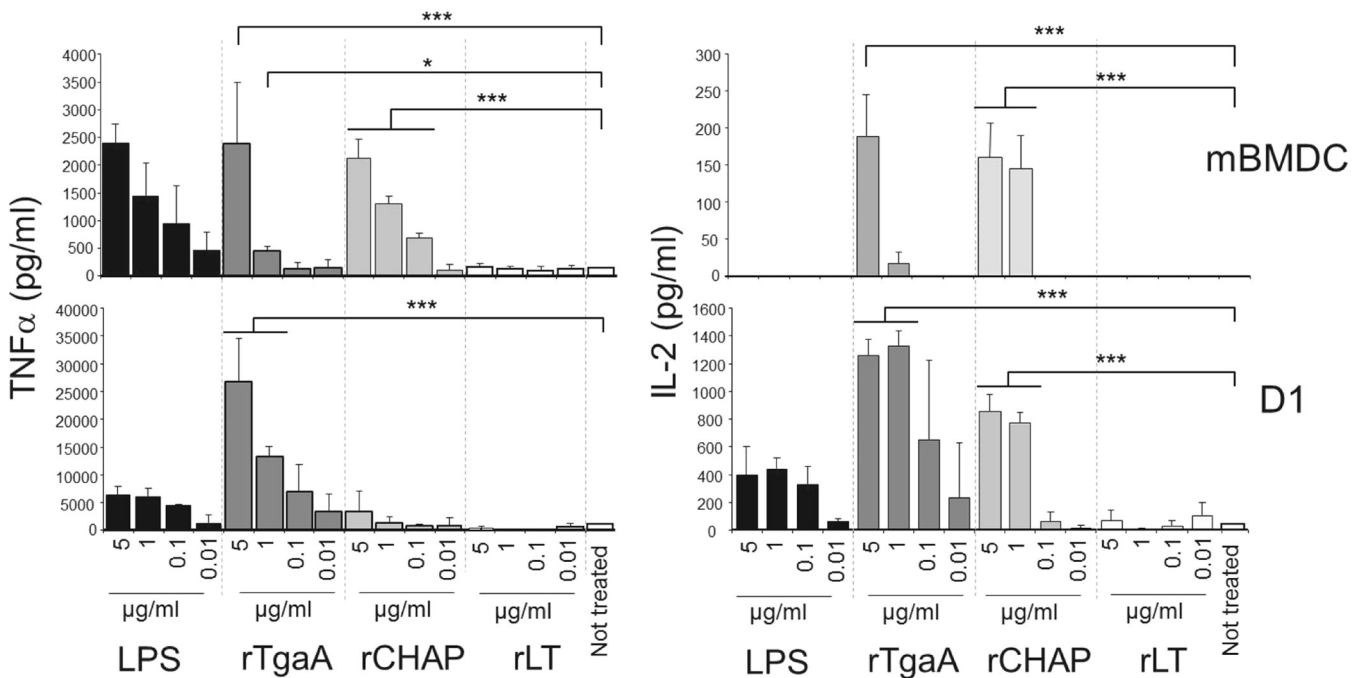


FIG 4 ELISA quantification of cytokine production by BMDCs and the D1 cell line stimulated with LPS or TgaA-derived recombinant proteins: rTgaA corresponds to Δ SP-TgaA-His, which contains both TgaA domains; rLT corresponds to Δ SP-TgaA- Δ CHAP-His, which contains only the LT domain; rCHAP corresponds to Δ SP-TgaA- Δ LT-His, which contains only the CHAP domain. Statistically significant differences were determined by an unpaired Student's *t* test (*, $P < 0.05$; ***, $P < 0.001$). mBMDC, murine BMDCs.

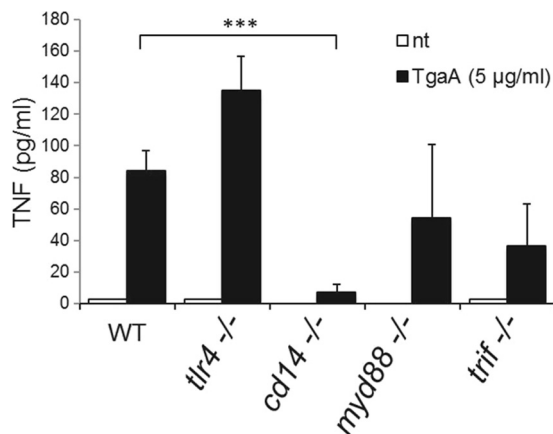


FIG 5 ELISA quantification of TNF- α produced by rTgaA-stimulated BMDCs isolated from wild-type (WT) and knockout mice. *tlr4*^{-/-}, Toll-like receptor 4 (TLR4) KO mutant mouse; *cd14*^{-/-}, CD14 protein KO mutant mouse; *myd88*^{-/-}, myeloid differentiation primary response gene 88 (MYD88) KO mutant mouse; *trif*^{-/-}, Toll IL-1 receptor (TIR) domain-containing adaptor-inducing IFN- β (TRIF) KO mutant mouse. Statistically significant differences were determined by an unpaired Student's *t* test (***, *P* < 0.001). nt, not treated.

from knockout mice lacking the expression of TLR4 can be activated by TgaA. According to our data, the receptor TLR4 was not necessary to induce TNF- α in BMDCs by TgaA (Fig. 5). Therefore, whereas LPS required TLR4 to induce TNF- α (43), the protein TgaA directly activated the downstream pathways through CD14 or involved CD14 for its presentation to a different unknown receptor, independently of TLR4.

To test if other TLRs might be important to recognizing TgaA by DCs, we used BMDCs derived from mice lacking the transducer of TLR signaling. In particular, TLR downstream signaling can occur via two pathways, which depend on two proteins: (i) the myeloid differentiation primary response gene 88 (MyD88) and (ii) the Toll IL-1 receptor (TIR) domain-containing adaptor-inducing IFN- β (TRIF) (44). Surprisingly, we found that without a functional TRIF or MyD88, production of TNF- α by TgaA-stimulated BMDCs was largely preserved (Fig. 5), suggesting that, in dendritic cells, the downstream signaling induced by TgaA could follow a CD14-dependent, TLR-independent signaling pathway.

Collectively, the above data show that TgaA is a cell wall protein of *B. bifidum* MIMBb75 and contains a C-terminal domain that initiates a CD14-dependent and TLR4-independent signaling pathway, which only partially involves the TRIF adapter protein.

A putative CHAP coding sequence is present in many bacterial genomes and is included in a number of different deduced protein architectures (45). Interestingly, several proteins from Gram-positive bacteria that include the CHAP domain actively interact with the host's immune system. For instance, the *Streptococcus pyogenes* SibA protein binds all immunoglobulin G (IgG) subclasses, the Fc and Fab fragments, and also IgA and IgM (32). PcsB, another CHAP-containing protein of *Streptococcus* spp. and a known antigen of *Streptococcus pneumoniae*, is capable of activating T cells in humans by inducing strong IL-17A responses (46). Furthermore, P40, a CHAP-containing protein produced by *Lactobacillus rhamnosus* GG (perhaps the most studied probiotic strain so far), can inhibit cytokine-induced epithelial cell apoptosis, reduce TNF-induced colon epithelial damage, and promote cell growth

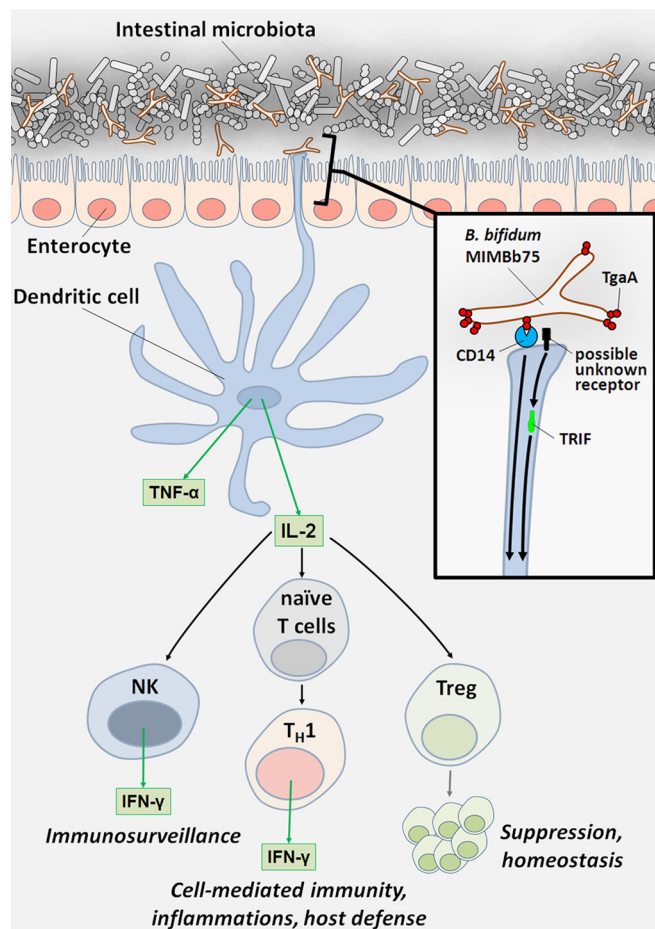


FIG 6 DC-mediated host immune response to TgaA protein from *Bifidobacterium bifidum* MIMBb75: possible consequences of IL-2 production. The contact between *B. bifidum* MIMBb75 and dendritic cells (DCs) determines DC activation and IL-2 secretion. Reportedly, stimuli able to induce IL-2 production by DCs are associated with a Th1-skewing of the immune responses, Treg lymphocyte proliferation, and natural killer (NK) cell activation (for a review, see reference 50).

in human and mouse colon epithelial cells and cultured mouse colon explants (47, 48). In addition, our study showed that TgaA from *B. bifidum* MIMBb75 is a cell surface-exposed molecule capable alone through its C-terminal CHAP domain of inducing DC activation and IL-2 production. Based on observed induction of IL-2 secretion, the TgaA stimulatory capacity here described may stimulate dendritic cells to acquire Th1 stimulatory capacity (35). Furthermore, the ability of *B. bifidum* MIMBb75 and its protein TgaA to induce IL-2 by DCs, together with the previous observation that *B. bifidum* strains induce an immune response affecting Treg/Th17 plasticity (49), supports the hypothesis that these commensal bacteria have a key role in mucosal tolerance. The presence of specific microbial stimuli, such as the CHAP-containing TgaA protein, may then have served as a way for commensals to co-evolve with the host and produce mechanisms for maintaining homeostasis and controlled reactions in the adaptive immune system (i.e., prevent an imbalance in T-helper cell subsets).

Interestingly, a BLASTP search revealed the presence of a gene closely similar to the CHAP domain of TgaA (with more than 80% identity in the amino acid sequence) not only in *B. bifidum* but

also in the genome of *B. breve* (e.g., GenBank accession number WP_016462813), which is another *Bifidobacterium* species typically associated with healthy breast-fed infants (13). Therefore, the presence in *B. bifidum* and other bifidobacteria of molecules similar to the TgaA protein and capable of stimulating the immune system could help explain the ability of these bacteria to protect the host and nurture the immune system in early life (15, 16).

Conclusions. The results of this study can be summarized as follows: (i) both the cells of *B. bifidum* MIMBb75 and its surface protein TgaA can activate dendritic cells and trigger IL-2 production; (ii) TgaA activates dendritic cells through its CHAP amidase domain; and (iii) the TgaA-dependent activation of dendritic cells requires the protein CD14, only marginally involves TRIF, and is independent of TLR4 and MyD88.

The bacterium *B. bifidum* MIMBb75, which is of human origin, has probiotic properties (28), and its ability to interact with the host may partly depend on the presence of the CHAP domain of its TgaA surface protein. In conclusion, the bacterial CHAP domain may well be considered a novel, potentially widespread MAMP that may participate in the cross talk mechanisms among Gram-positive bacteria and their mammalian host. The possible physiological consequences of the ability of *B. bifidum* MIMBb75 and its TgaA protein to activate DCs are schematically summarized in Fig. 6.

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