

A novel alpha1-antitrypsin null variant (*PiQ0Milano*)

Raffaella Rametta, Gabriella Nebbia, Paola Dongiovanni, Marcello Farallo, Silvia Fargion, Luca Valenti

Raffaella Rametta, Silvia Fargion, Luca Valenti, Department of Pathophysiology and Transplantation, Università degli Studi di Milano, 20122 Milano, Italy

Gabriella Nebbia, Paola Dongiovanni, Silvia Fargion, Luca Valenti, Internal Medicine, Fondazione IRCCS Ca' Granda Ospedale Policlinico Milano, 20122 Milano, Italy

Marcello Farallo, Pediatric Clinic, Fondazione IRCCS Ca' Granda Ospedale Policlinico, 20122 Milano, Italy

Author contributions: Rametta R performed the genetic analysis and wrote the first manuscript draft; Dongiovanni P supervised genetic analysis; Nebbia G and Farallo M were responsible for the clinical care of the patients; Fargion S and Valenti L supervised the study; Valenti L edited the manuscript; all authors contributed to data interpretation, read and approved the final manuscript draft.

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Correspondence to: Luca Valenti, MD, Department of Pathophysiology and Transplantation, Università degli Studi di Milano, Fondazione IRCCS Ca' Granda Ospedale Policlinico, Pad. Granelli, via F Sforza 25, 20122 Milano, Italy. luca.valenti@unimi.it

Telephone: +39-2-50320278 Fax: +39-2-50320296

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Abstract

Alpha1-antitrypsin deficiency is an autosomal recessive disease characterized by reduced serum levels of alpha1-antitrypsin (AAT) due to mutations in the *SERPINA1* gene causing early onset pulmonary emphysema and, occasionally, chronic liver disease. We report an incidental finding of a novel null *AAT* allele, *Q0Milano*, consisting of a 17 nucleotides deletion in exon 3 of *SERPINA1* gene, in an Italian child with persistently increased liver enzymes, a mild decrease in circulating AAT levels and without any pulmonary disease. *Q0Milano* variant results in an unfunctional protein lacking of AAT active site, as the resultant protein is truncated near PiS locus involved in AAT protein stability.

Key words: Alpha1-antitrypsin deficiency; Rare variant; Alpha1-antitrypsin null mutation; Liver disease

Core tip: We report an incidental finding of a novel null alpha1-antitrypsin (*AAT*) allele, *Q0Milano*, consisting of a 17 nucleotides deletion in exon 3 of *SERPINA1* gene, in an Italian child with persistently increased in liver enzymes and a mild decrease in circulating AAT levels. *Q0Milano* variant results in an unfunctional protein lacking of AAT active site, as the resultant protein is truncated near PiS locus involved in AAT protein stability.

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INTRODUCTION

Alpha1-antitrypsin deficiency (AATD) is an autosomal recessive disease characterized by reduced serum levels of alpha1-antitrypsin (AAT, *SERPINA1*), a 52 kDa glycoprotein functioning as the main extracellular protease inhibitor. AAT is mainly produced by liver, which releases about 2 g of AAT daily into the circulation under physiological conditions. The normal serum concentration may range between 1.5-3.5 g/L (or 20-48 $\mu\text{mol/L}$)^[1]. AATD is associated with early onset pulmonary emphysema and, occasionally, with chronic liver disease in childhood, hepatocellular carcinoma and/or cirrhosis in adulthood^[2]. AAT functions as neutrophil elastase inhibitor, playing a key role in the protection of the lower respiratory tract. AAT serum levels below 11 $\mu\text{mol/L}$ are not sufficient to inhibit elastase *in vivo*, permitting progressive destruction of alveoli culminating in emphysema^[3-5]. The pathophysiology of liver disease related to AATD is less well understood, but some deficient variants accumulate in endoplasmic reticulum of hepatocytes and are inefficiently secreted, leading to protein aggregation and culminating

Table 1 Sequence of primers used in Alpha1-antitrypsin coding sequence amplification and sequencing

| | Primers forward 5'→3' | Primers reverse 5'→3' |
|----------|------------------------|-----------------------|
| AAT Ex2A | CCCCCATCTCTGTCTTGC | GAGGAGTTCCTGGAAGCCTT |
| AAT Ex2B | ATGAAATCCITGGAGGGCCIG | CAGGCTGGTTGAGCAACCTT |
| AAT Ex3 | CCCACCTCCCTCTCTCC | CACCCTCAGGTTGGGGAATC |
| AAT Ex4 | CTTGAATTTCTTTCTGCACGAC | AAGGTCGTCAGGGTGATCTC |
| AAT Ex5 | GTCTCTGCTCTCTCCCTC | AGGGACCAGCTCAACCTTC |

AAT: Alpha1-antitrypsin; Ex: Exon.

in hepatocytes injury and liver disease^[6].

AATD is caused by mutations in *SERPINA1*, a highly polymorphic genetic locus located on the distal long arm of chromosome 14. More than 100 alleles have been identified. They can be classified according to AAT serum levels and protein functionality: (1) normal variants, all common M types, accounting for 95% of those found in Caucasian individuals, and characterized by normal plasma levels (more than 20 $\mu\text{mol/L}$); (2) deficient variants associated with reduced AAT serum levels, lower than 20 $\mu\text{mol/L}$; (3) null variants determining undetectable serum levels; and (4) dysfunctional variants characterized by normal serum levels of dysfunctional AAT protein^[7].

The firstly described, and most common cause of AATD, associated with very low serum concentration of the protein, is homozygosity for the PiZ mutation, the most severe AAT deficient variant known with plasma levels among homozygotes of about 5-6 $\mu\text{mol/L}$, resulting in the development of lung and liver disease^[8]. It became later clear that AATD is a heterogeneous disease, caused by several gene defects expressed codominantly, mostly determining reduced serum AAT levels.

The most common deficient alleles are PiS and PiZ, with an allelic frequency of 2%-4% and 1%-2% respectively in Caucasian population, and are both caused by missense mutations responsible for intracellular protein accumulation and degradation. The PiZ mutation leads to a conformational change of AAT reactive site into a β -sheet polymer which forms characteristic periodic acid-Schiff-positive inclusions and can be isolated for liver of AAT PiZZ subjects. Several studies have shown that PiMZ in heterozygous state may lead to chronic liver disease, cryptogenic cirrhosis, and chronic active hepatitis^[9-11], while the PiS variant is associated to liver disease only if carried in compound heterozygosity with the PiZ allele^[12]. Null alleles are very rare (frequency < 0.001, 13% of AATD subjects in Italy^[13]) and derived from nucleotide deletion, insertion, or non-sense mutations, causing premature stop codons and producing structurally unstable and truncated protein. Individuals with null-null AAT phenotype are not affected by liver disease, because of the lack of aggregation of mutant proteins in the endoplasmic reticulum, but are associated with an increased risk of emphysema.

In this study we report a novel AAT allele in a child with reduced protein levels.

CASE REPORT

An 11-years-old male child was referred to our centre for a persistent increase in liver enzymes (aspartate aminotransferase and alanine aminotransferase spanning from 58-239 UI/L, and 98-114 UI/L respectively). All common causes of liver disease were excluded, but mildly decreased AAT serum levels were detected (76 mg/dL). The subject did not show abnormalities in pulmonary function.

Sequencing of the proband revealed a novel null mutation in *AAT* gene (Table 1), g.9752-9768del (PiQ_{0Milano}) (gene ID: 5265, official name SERPINA1, genomic sequence number: NC_000014.8; NCBI Reference Sequence: NG_008290.1). This variant, localized in exon 3 near PiS locus (p.Glu264Val), consists in a 17 bp deletion (AAA CTA CAG CAC CTG GA), resulting in a frameshift causing a new stop codon downstream the deletion site (Figure 1), which leads to a premature termination of protein translation at amino acid 259. The truncated protein lacks of AAT active site centred around Met³⁵⁸-Ser³⁵⁹.

The mutation arose in PiM3 *AAT* allele, which the proband inherited from his mother, whose genotype was PiM1A/Q0Milano and had normal liver and pulmonary function, whereas his father, who had normal genotype PiM1V/PiM3, showed nonalcoholic steatohepatitis associated with hyperferritinemia (Figure 2). Liver biopsy of the proband showed aspecific findings, unrelated to AATD.

DISCUSSION

The g.9752-9768del mutation (Q0Milano) occurs in a key functional region of AAT gene where several other deficiency variants (PiLowell/PiDuarte at codon 256, Q0Cairo at codon 259, T/S at codon 264) have been reported^[14-18].

Several mechanisms are responsible for AAT deficiency, including: gene deletion, mRNA degradation, intracellular protein accumulation and degradation, and production of dysfunctional proteins. Only mutations causing intracellular protein accumulation and polymerization of the newly synthesized protein are associated with increased risk of liver disease, as in PiZ and in PiM-Malton variants^[19]. Many previously described null mutations (PiDuarte, PiHong Kong, PiGranite Falls) have been associated with intra-reticular accumulation of unfunctional AAT proteins, which are immediately degraded without any

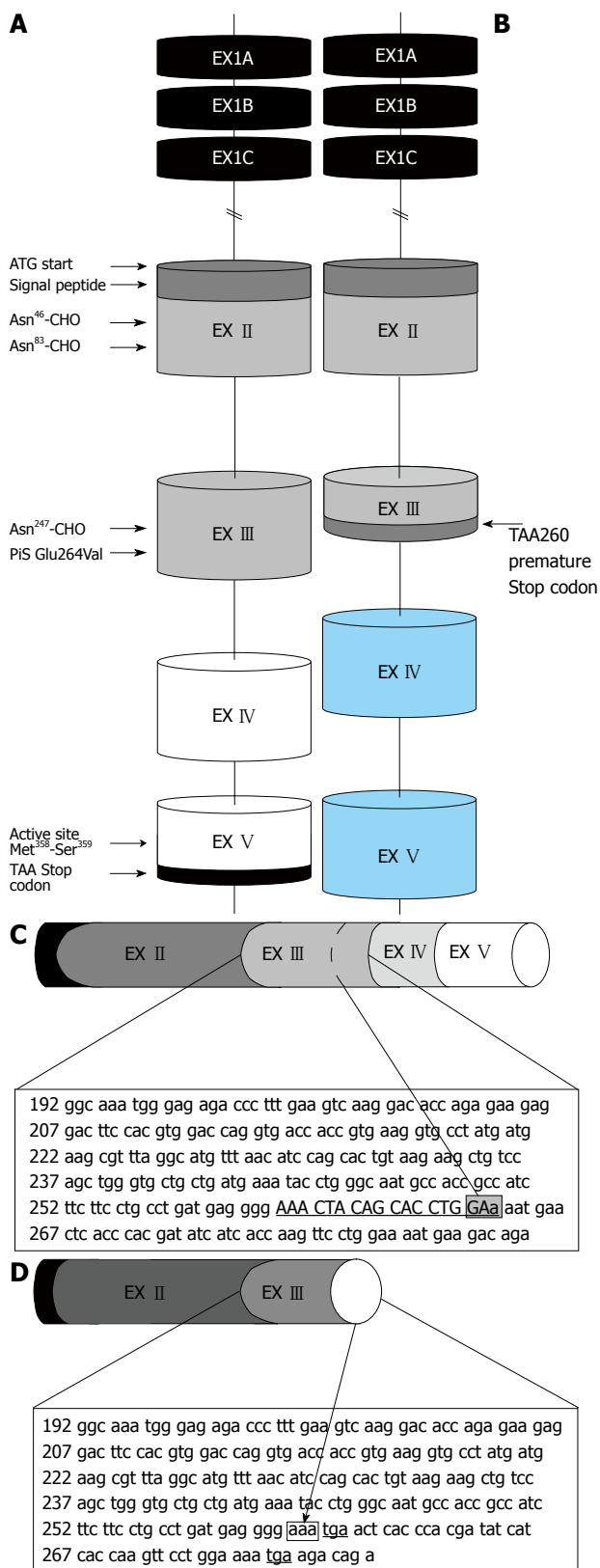


Figure 1 Schematic representation of wild type and mutant Alpha1-antitrypsin gene and protein. A: Alpha1-antitrypsin (AAT) wild type gene; B: AAT mutated gene: Blue region represents out of frame sequence. Untranslated regions are shown in black; C: AAT wild type protein: Exon 3 wild-type sequence is indicated in square; the deleted nucleotides are underlined; PIS locus is highlighted; D: AAT truncated unfunctional protein; Exon 3 deleted sequence is indicated below; premature stops codons resulting from frameshift were underlined.

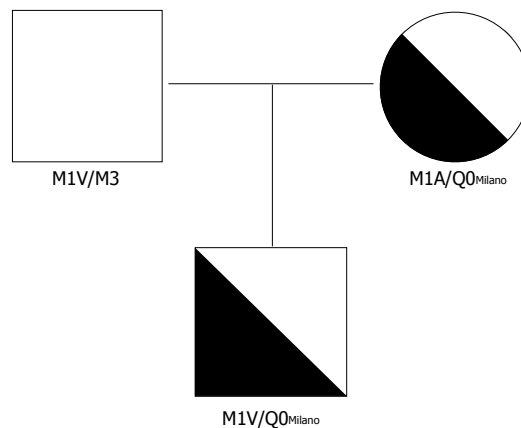


Figure 2 Pedigree of the Q0Milano proband's family. Alpha1-antitrypsin genotypes are listed below.

liver damage^[20-22]. Heterozygosity for this novel null mutation is consistent with the lower AAT serum levels (76 mg/dL) measured in the proband, which collocates the patient in intermediate deficiency condition comparable to those of individuals carrying PiMZ genotype^[23].

However, it is unlikely that this genetic variant explained liver disease in the proband, as it was carried in heterozygous state, and it does not affected liver function tests of the mother. Moreover, the absence of periodic acid-Schiff-positive inclusions revealed in liver biopsy excluded hepatic AAT protein accumulation. Thus, the novel null *AAT* variant was not responsible for liver damage because of the lack of hepatic protein polymerization.

It is likely that other hepatotoxic insults, as non alcoholic steatohepatitis associated with hyperferritinemia, a strongly heritable condition reported in the father of proband^[24,25], were involved in the development of liver disease.

In conclusion, in this study we report a novel *AAT* null variant (*Q0Milano*) generated by a 17 nucleotides deletion in exon 3 of *AAT*, which leads to a premature stop codon.

REFERENCES

- 1 **Fagerhol MK**, Laurell CB. The polymorphism of "prealbumins" and alpha-1-antitrypsin in human sera. *Clin Chim Acta* 1967; **16**: 199-203 [PMID: 4166396]
- 2 **Crystal RG**. Alpha 1-antitrypsin deficiency, emphysema, and liver disease. Genetic basis and strategies for therapy. *J Clin Invest* 1990; **85**: 1343-1352 [PMID: 2185272]
- 3 **Gadek JE**, Fells GA, Zimmerman RL, Rennard SI, Crystal RG. Antielastases of the human alveolar structures. Implications for the protease-antiprotease theory of emphysema. *J Clin Invest* 1981; **68**: 889-898 [PMID: 6169740]
- 4 **Wewers MD**, Casolaro MA, Sellers SE, Swayze SC, McPhaul KM, Wittes JT, Crystal RG. Replacement therapy for alpha 1-antitrypsin deficiency associated with emphysema. *N Engl J Med* 1987; **316**: 1055-1062 [PMID: 3494198]
- 5 **Ferrarotti I**, Thun GA, Zorzetto M, Ottaviani S, Imboden M, Schindler C, von Eckardstein A, Rohrer L, Rochat T, Russi EW, Probst-Hensch NM, Luisetti M. Serum levels and genotype distribution of α 1-antitrypsin in the general

- population. *Thorax* 2012; **67**: 669-674 [PMID: 22426792 DOI: 10.1136/thoraxjnl-2011-201321]
- 6 **Carrell RW**, Lomas DA. Alpha1-antitrypsin deficiency--a model for conformational diseases. *N Engl J Med* 2002; **346**: 45-53 [PMID: 11778003]
 - 7 **Zaimidou S**, van Baal S, Smith TD, Mitropoulos K, Ljujic M, Radojkovic D, Cotton RG, Patrinos GP. A1ATVar: a relational database of human SERPINA1 gene variants leading to alpha1-antitrypsin deficiency and application of the Vari-Vis software. *Hum Mutat* 2009; **30**: 308-313 [PMID: 19021233 DOI: 10.1002/humu.20857]
 - 8 **Laurell CB**, Eriksson S. The electrophoretic α 1-globulin pattern of serum in α 1-antitrypsin deficiency. *COPD* 2013; **10** Suppl 1: 3-8 [PMID: 23527532 DOI: 10.1080/00365516309051324]
 - 9 **Hodges JR**, Millward-Sadler GH, Barbatis C, Wright R. Heterozygous MZ alpha 1-antitrypsin deficiency in adults with chronic active hepatitis and cryptogenic cirrhosis. *N Engl J Med* 1981; **304**: 557-560 [PMID: 6969850]
 - 10 **Graziadei IW**, Joseph JJ, Wiesner RH, Therneau TM, Batts KP, Porayko MK. Increased risk of chronic liver failure in adults with heterozygous alpha1-antitrypsin deficiency. *Hepatology* 1998; **28**: 1058-1063 [PMID: 9755243]
 - 11 **Fischer HP**, Ortiz-Pallardó ME, Ko Y, Esch C, Zhou H. Chronic liver disease in heterozygous alpha1-antitrypsin deficiency PiZ. *J Hepatol* 2000; **33**: 883-892 [PMID: 11131449]
 - 12 **Chan CH**, Steer CJ, Vergalla J, Jones EA. Alpha1-antitrypsin deficiency with cirrhosis associated with the protease inhibitor phenotype SZ. *Am J Med* 1978; **65**: 978-986 [PMID: 217266]
 - 13 **Ferrarotti I**, Baccheschi J, Zorzetto M, Tinelli C, Corda L, Balbi B, Campo I, Pozzi E, Faa G, Coni P, Massi G, Stella G, Luisetti M. Prevalence and phenotype of subjects carrying rare variants in the Italian registry for alpha1-antitrypsin deficiency. *J Med Genet* 2005; **42**: 282-287 [PMID: 15744045]
 - 14 **Faber JP**, Weidinger S, Goedde HW, Ole K. The deficient alpha-I-antitrypsin phenotype PI P is associated with an A-to-T transversion in exon III of the gene. *Am J Hum Genet* 1989; **45**: 161-163 [PMID: 2787118]
 - 15 **Graham A**, Kalsheker NA, Newton CR, Bamforth FJ, Powell SJ, Markham AF. Molecular characterisation of three alpha-1-antitrypsin deficiency variants: proteinase inhibitor (Pi) nullcardiff (Asp256----Val); PiMmalton (Phe51----deletion) and PiI (Arg39----Cys). *Hum Genet* 1989; **84**: 55-58 [PMID: 2606478]
 - 16 **Hildesheim J**, Kinsley G, Bissell M, Pierce J, Brantly M. Genetic diversity from a limited repertoire of mutations on different common allelic backgrounds: alpha 1-antitrypsin deficiency variant Pduarte. *Hum Mutat* 1993; **2**: 221-228 [PMID: 8364590]
 - 17 **Zorzetto M**, Ferrarotti I, Campo I, Balestrino A, Nava S, Gorrini M, Scabini R, Mazzola P, Luisetti M. Identification of a novel alpha1-antitrypsin null variant (Q0Cairo). *Diagn Mol Pathol* 2005; **14**: 121-124 [PMID: 15905697]
 - 18 **Curiel DT**, Chytil A, Courtney M, Crystal RG. Serum alpha 1-antitrypsin deficiency associated with the common S-type (Glu264----Val) mutation results from intracellular degradation of alpha 1-antitrypsin prior to secretion. *J Biol Chem* 1989; **264**: 10477-10486 [PMID: 2567291]
 - 19 **Eriksson S**, Carlson J, Velez R. Risk of cirrhosis and primary liver cancer in alpha 1-antitrypsin deficiency. *N Engl J Med* 1986; **314**: 736-739 [PMID: 3485248]
 - 20 **Lieberman J**, Gaidulis L, Klotz SD. A new deficient variant of alpha1-antitrypsin (MDUARTE). Inability to detect the heterozygous state by antitrypsin phenotyping. *Am Rev Respir Dis* 1976; **113**: 31-36 [PMID: 1082282]
 - 21 **Nukiwa T**, Takahashi H, Brantly M, Courtney M, Crystal RG. alpha 1-Antitrypsin nullGranite Falls, a nonexpressing alpha 1-antitrypsin gene associated with a frameshift to stop mutation in a coding exon. *J Biol Chem* 1987; **262**: 11999-12004 [PMID: 3040726]
 - 22 **Sifers RN**, Brashears-Macatee S, Kidd VJ, Muensch H, Woo SL. A frameshift mutation results in a truncated alpha 1-antitrypsin that is retained within the rough endoplasmic reticulum. *J Biol Chem* 1988; **263**: 7330-7335 [PMID: 3259232]
 - 23 **Zorzetto M**, Russi E, Senn O, Imboden M, Ferrarotti I, Tinelli C, Campo I, Ottaviani S, Scabini R, von Eckardstein A, Berger W, Brändli O, Rochat T, Luisetti M, Probst-Hensch N. SERPINA1 gene variants in individuals from the general population with reduced alpha1-antitrypsin concentrations. *Clin Chem* 2008; **54**: 1331-1338 [PMID: 18515255 DOI: 10.1373/clinchem.2007.102798]
 - 24 **Dongiovanni P**, Anstee QM, Valenti L. Genetic Predisposition in NAFLD and NASH: Impact on Severity of Liver Disease and Response to Treatment. *Curr Pharm Des* 2013; **19**: 5219-5238 [PMID: 23394097]
 - 25 **Dongiovanni P**, Fracanzani AL, Fargion S, Valenti L. Iron in fatty liver and in the metabolic syndrome: a promising therapeutic target. *J Hepatol* 2011; **55**: 920-932 [PMID: 21718726 DOI: 10.1016/j.jhep.2011.05.008]

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