1	Effect of time and storage temperature on anthocyanin decay and antioxidant activity in wild
2	blueberry (Vaccinium angustifolium) powder
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15 <u>Abstract</u>

We investigated the decay kinetics of total and single anthocyanins (ACNs), and Total Antioxidant 16 Activity (TAA) of freeze-dried wild blueberry (WB) powder stored at 25°C, 42°C, 60°C and 80°C 17 18 for 49 days utilizing the Arrhenius equation. At storage time-intervals of 3-4 days, ACNs and TAA 19 were determined. Moreover, the Arrhenius equation was used to predict the shelf-life of ACNs and 20 TAA at 4°C. Results demonstrated that the degradation of ACNs followed a first-order kinetic. 21 Total and single ACN decay occurred at all the temperatures but was slower at 25°C compared to 22 60°C and 80°C. On the contrary, TAA was unaffected after storage at 42°C for 49 days. 23 In conclusion, WB powder maintains the content of ACNs and TAA longer (up to 130 days) at 24 25°C; however, storage at 4°C represents the best way to preserve the nutritional quality of the 25 product and delay decay.

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<u>KEYWORDS</u>: wild blueberry powder, anthocyanins, total antioxidant activity, storage temperature.
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30 Introduction

In recent years, several studies documented the beneficial effects of berries (i.e. cranberries, 31 32 raspberries, and blueberries) on human health. Wild blueberries (Vaccinium angustifolium) have 33 been reported to have a protective effect against chronic diseases, especially cardiovascular disease; ¹⁻³ this has generally been attributed to their polyphenol content, anthocyanins (ACNs) in particular. 34 These compounds are responsible for the blue and purple colour of the berries, and for these 35 reasons are also used as natural food colorants in the food industry.⁴ However, ACNs are labile in 36 nature and susceptible to deterioration during processing and storage.⁵ Blueberries are often quick 37 frozen at very low temperatures (-80°C) for long-term preservation with minimal effects on 38 quality.⁶⁻⁷ The majority of berries, including blueberries, are consumed as processed foods ie. 39 40 juices, purees, jams, syrups, jellies and various ready-to drink beverages to ensure extension of shelf-life and consumption independent of the growing season.⁸⁻¹¹ Another common system to 41 42 preserve blueberries is through the freeze-drying process which has several advantages for the food 43 industry such as a reduction of storage space, size and cost. Moreover the freeze-drying process 44 permits the standardization content of nutrient and phytochemicals useful for human health.

Several mechanisms of degradation during processing and storage have been documented. In freezing and cold storage, the retention of ACNs depends on the rate of freezing, temperature, and the presence/absence of oxygen, and the food matrix.^{5,12} Studies verified the stability or at least a slight increase in ACN content in berries/blueberries during cold storage¹³⁻¹⁴ or storage at highoxygen atmospheres.¹⁵ On the contrary, a reduction was observed for extruded products such as cereal blueberry-rich products¹⁶ and for thermally processed foods such as juices,^{8-9,17} jams,^{10,18} and purées.¹¹

Anthocyanin degradation is high when these products are treated at higher temperature (up to 121°C) and then refrigerated.¹⁹⁻²⁰ Concerning dry storage, the major parameters determining the stability of ACNs are water content, water activity (a_w), temperature, presence/absence of oxygen, light, and relative humidity.^{11,19} However, no data is available concerning the effect of storage on ACN content in freeze-dried wild blueberry powder. This is very important since the food industry uses the freeze-dried products as ingredients in many food formulations, such as jams, jellies, sauces, purées, toppings, syrups, juices, and bakery and dairy products. Moreover, in past studies the ACN concentration was commonly quantified as total ACNs and no information was reported on the fate of the single compounds contained in the blueberries.^{8-11,17-20}

For these reasons, the objective of this study was to investigate for the first time, the degradation kinetics of single ACNs contained in freeze-dried wild blueberry (WB) powder samples stored at different temperatures (25°C, 42°C, 60°C and 80°C) for 49 days. Total ACN content and total antioxidant activity (TAA) were investigated as well, under the above conditions.

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67 Materials and Methods

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69 Chemicals and Materials

70 Standard of cyanidin (Cy)-, delphinidin (Dp)-, petunidin (Pt)-, peonidin (Pe) and malvidin (Mv)-3-71 O-glucoside (glc), Cy-, Pt-, Pe-, and Mv-3-O-galactoside (gal) were purchased from Polyphenols 72 Laboratory (Sandes, Norway). Potassium chloride, hydrochloric acid, methanol, acetonitrile, 73 phosphoric acid, and trifluoroacetic acid (TFA) were from Merck (Darmstadt, Germany). 2,2-74 azinobis (3-ethyl-benzothiazoline-6-sulfonic acid) diammonium salt (ABTS), 6-hydroxy-2,5,7,8-75 tetramethyl-chroman- 2-carboxylic acid (Trolox) were purchased from Sigma (St. Louis, MO, 76 USA). Water was obtained from Milli-Q apparatus (Millipore, Milford, MA). Freeze-dried WB 77 powder was provided by FutureCeuticals Company (Momence, IL, USA).

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80 Sample preparation

81 Wild blueberry powder was stored at -80°C until analysis. Sixty samples of one gram each, were 82 placed in zip-lock plastic bags used for commercial products, sealed under vacuum and stored in 83 four controlled temperatures (25°C, 42°C, 60°C, 80°C).

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85 **Degradation studies**

The thermal degradation of ACNs as well as the TAA of the WB powder was investigated at 25°C, 42°C, 60°C and 80°C for 49 days. Two samples of WB powder (1 g each) were taken, based on the Accelerated Shelf Life Testing method at appropriate time intervals (3-4 days) for analyses. They were rapidly cooled and ACN extraction was performed for the determination of total and single ACNs concentration and TAA. All analyses were done in duplicate.

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92 Extraction of ACNs from wild blueberry powder

93 Anthocyanin extraction was performed as follows: 50 mg of WB powder was dissolved in 5 mL of 94 methanol acidified with 1% of TFA. The suspension was sonicated for 10 minutes, centrifuged at 95 3000xg for 15 min and the supernatant was recovered and the volume adjusted to 10 mL by 96 methanol acidified with 1% of TFA.

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98 **Determination of total anthocyanins**

99 The total content of ACNs was determined spectrophotometrically (Perkin Elmer Lambda 20, 100 Waltham, MA) as described by Lee et al.²¹ Briefly, two aliquots of the extracted ACNs were diluted 101 1:10 in KCl 0.025 M at pH 1 and in CH₃COONa 0.4 M at pH 4.5. The absorbance was measured 102 twice for each sample and buffer at the following wavelengths: 520 nm and 700 nm. The 103 absorbance *A* was calculated as follows:

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$$A = (A_{520nm} - A_{700nm}) at pH1 - (A_{520nm} - A_{700nm}) at pH4.5$$

105 The total ACN content was calculated as follows:

106 $mg ACNs/100g = A * \varepsilon^{-1} * MW * W/V * DF$

107 Where ε is the Cy-glc molar extinction coefficient (26900 mol L⁻¹ cm⁻¹), *MW* is the molecular 108 weight (449.2 Da), *W* is the sample weight, *V* is the volume (mL) and *DF* is the dilution factor.

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110 **Determination of single ACNs**

The liquid chromatography (LC) system was an Alliance mod. 2695 (Waters, Milford, MA) 111 112 equipped with a photodiode array detector (mod. 2998, Waters). The separation was carried out by a C₁₈ Kinetex column (150 x 4.6 mm, 2.6 µm, Phenomenex, Torrence, CA) maintained at 45°C. 113 The flow-rate was 1.7 mL min⁻¹ and the eluents were (A) 1% H₃PO₄ and (B) acetonitrile/water 114 115 (35:65, v/v). The elution gradient was linear as follows: 0-15 min 14% B; 15-25 min from 14 to 20% B; 25-35 min from 20 to 32% B; 35-45 min from 32 to 50% B; 45-48 min from 50 to 90% B; 116 117 90 % for 3 minutes. Chromatographic data were acquired from 200 nm to 700 nm and integrated at 118 520 nm.

119 Calibration curves ranged from 2 to 50 μ g mL⁻¹; the working solution was obtained diluting the 120 stock solution (1 mg mL⁻¹ in methanol acidified with 0.1% TFA) with 0.1% TFA. Each analysis 121 was carried out in duplicate.

The concentration of the five ACNs not commercially available (Dp-gal, Dp-ara, Cy-ara, Mv-ara, and Pt-ara) was estimated using the calibration curve equation of the same anthocyanidin with different glycosylation. The acetylated ACNs were determined by the Cy-glc curve and the resulting data was corrected by their corresponding molecular weight ratios. The identification of single ACNs was confirmed by LC ESI/MS according to method previously published.²²

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128 Determination of TAA

The Total Antioxidant Activity (TAA) was determined by the Trolox Equivalent Antioxidant
Capacity (TEAC) assay as described by Pellegrini et al.²³

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132 **Degradation kinetic studies**

133 The thermal degradation of ACNs was performed according to the method reported by Kechinski et

134 al.²⁴ Degradation is a temperature-dependent process, as described by the Arrhenius equation:

$$k = k_0 * e^{-Ea/RT}$$

Where k_0 is the frequency factor (per min), E_a the activation energy (J mol⁻¹), *R* the universal gas constant (8.314 J mol⁻¹ K⁻¹), and *T* the absolute temperature (K).

138 The coefficient Q_{10} expresses ACN degradation when the temperature is increased to 10°C and it is 139 calculated as follows:

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$$Q_{10} = (k_{at,T2}/k_{at,T1})^{(10/(T2-T1))}$$

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- 142

143 **Statistical analysis**

Statistical analysis was performed by means of STATISTICA software (Statsoft Inc., Tulsa, OK, US). Analysis of variance (ANOVA) with type of treatment as the dependent factor was used to evaluate the variations of ACNs and TAA. One-way ANOVA was performed to determine the variation among the samples stored at different temperatures. Differences between means were evaluated by the Least Significant Difference (LSD) test. Differences were considered significant at $P \le 0.05$.

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151 **Results and Discussion**

This is the first study that focuses on ACN degradation in a freeze-dried WB powder and its shelflife. The ACN profile of the WB powder before the storage treatment is reported in **Figure 1**. The HPLC method used, allowed for the separation of 21 ACNs, 15 glycosylated anthocyanidins and 6 acetylated forms, which identities were confirmed by LCMS and MS/MS as previously reported.²² The mean relative standard deviation (RSD) was 6.1 % for concentrations from 0.5 to 20 µg mL⁻¹. The main ACNs detected in the WB powder were: Pe-glc, Mv-glc, Dp-glc, and Dp-gal; these four compounds represented about 35% of the total amount of ACNs. The decay of total ACNs evaluated at four different temperatures (25°C, 42°C, 60°C and 80°C) is reported in **Figure 2**. In general, a significant difference ($P \le 0.0001$) on ACN content was detected for each temperature studied. Predictably, time and degree of ACN decay was dependent on temperature. In fact, we observed that the ACN decay occurred slowly up to 3% at day 14 at 25°C and 42°C while it was faster, achieving about 60% and 85% decay at day 3 at 60°C and 80°C, respectively.

The quantification of the single ACNs allowed for the calculation of the decay slope (mean 165 166 + SD) in the WB powder (Figure 3). The reduction in ACN content at 80°C was higher than 90% 167 after 3 days only, thus the data of single ACNs at this temperature were not used to evaluate their 168 degradation rate. The slopes calculated at 25°C, 42°C and 60°C showed that the degradation rate 169 followed a first-kinetic order. This trend was in accordance to that observed by several researchers on different juices, such as blood orange, blackberry, and blueberry juices and red wine.²⁴⁻²⁷ Each 170 compound displayed its own specific decay, related to the sugar binding and the storage 171 172 temperature. Moreover, it seems that the ACNs bound to glucose, exhibited a faster degradation 173 rate than those bound to galactose (data not shown). For all the ACNs, the correlation indices (R^2) 174 were higher than 0.90, demonstrating a direct correlation between ACN concentration decrease and storage time. Good correlation indices were also found for the acetylated forms ($R^2 > 0.81$), which 175 176 seems more stable that the correspondent glycosides.

The linear regression approach allows also for the calculation of the reaction rate constant (*k*). A direct relationship between *k* values and temperature was found (**Figure 4**), confirming the major effect of temperature on ACN degradation.

The values of E_a and half-life of total and single ACNs are reported in **Table 1**. The value of E_a for the total ACNs was about 58 kJ mol⁻¹. This data is lower than that reported by Kechinski et al.²⁴ which found a value of about 80 kJ mol⁻¹ in blueberry juice. The difference may be due to the different type of tested product, suggesting that ACNs contained in the WB powder are more susceptible to temperature than that in the juice. This could be attributable to a matrix effect and/or a different pH (pH 6 or lower in case of juice) that maintains ACN stability. Considering single ACNs, as already observed from the slope values (**Table 1**), the ACNs linked to galactose such as Cy-gal, Mv-gal, Pt-gal, and Pe-gal have values of E_a higher than 70 kJ mol⁻¹. This implies that in the WB powder, the galactosylated ACNs are more heat-stable. These data are in accordance with those reported by Scibsz et al,²⁸ that hypothesized a possible protective effect of galactose compared to glucose.

191 Contrarily the data reported for blueberry juice, delphinidin glycosides were not the compounds decaying faster with increased temperature.¹⁹ Indeed, in our product the most temperature labile 192 compounds were Pt-glc ($E_a = 18.14 \text{ kJ mol}^{-1}$) and Cy-ara ($E_a = 38.98 \text{ kJ mol}^{-1}$) as reported in **Table** 193 194 1. The possible relation between their chemical structure, such as the number of the hydroxyl 195 groups or the glycosylation degree or the acylated form and heat stability was studied by several researchers.²⁹⁻³¹ Unfortunately, the data reported in literature are often contradictory.⁵ For example, 196 Trost et al.³² reported that ACN stability in a blueberry-aronia nectar stored for over 207 days at 197 198 30°C, was higher for Cy- and Pe-, and lower for Pt-, Mv- and Dp-glycosides. In regard to conjugated sugars, the ranking order was glucoside > galactoside > arabinoside from the most to the 199 least stable.³² The greater stability of ACNs bound to glucose and galactose compared to arabinose 200 201 was proposed to be due to steric hindrance which results larger for the hexose sugars. On the contrary, Ichiyangi et al.³³ documented that the ranking order was arabinoside > galactoside >202 203 glucoside from the most to the least stable.

From our observations, the relative amount of a single ACN did not affect its heat stability. Indeed, Pe-gal is one of the compounds present in lower amount in the WB powder but with the highest E_a (84.15 kJ mol⁻¹) (**Table 1**). Among the acetylated forms, Pe-glc-Ac is the most heat sensitive ($E_a =$ 7.76 kJ mol⁻¹), while Cy-glc-Ac is the compound most heat resistant ($E_a = 84.28$ kJ mol⁻¹).

In addition to the degradation rate, the half-life time $(t_{1/2})$ was calculated by the Arrhenius equation for the single and total ACNs in relation to the investigated temperatures (**Table 1**). The $t_{1/2}$ values obtained for the total ACNs decay were 139, 39, 12, and 4 days at 25°C, 42°C, 60°C and 80 °C, 211 respectively. Large differences in the $t_{1/2}$ value existed among the single compounds stored at same 212 temperatures (**Table 1**), as well as at different temperatures. The $t_{1/2}$ value ranged from 86 to 611 213 days at 25°C, from 48 to 199 days at 42°C, and from 15 to 69 days at 60°C. Thus, storage at room 214 temperature (25 °C) can induce important loss of some ACNs such as Pe-glc, Cy-ara and Dp-glc 215 even though for most of them the $t_{1/2}$ value is much higher than 150 days (**Table 1**). Moreover, the 216 acetylated forms were more resistant than only glycosylated compounds for all the temperatures 217 considered. Additionally, our study found considerable changes for the different acetylated ACNs, 218 whose $t_{1/2}$ values were from few days to 1948, 519, and 296 days at 25°C, 42°C and 60°C, 219 respectively (Table 1).

The Q_{10} values for the total and single ACNs at the temperatures investigated are presented 220 221 in **Table 2**. The Q_{10} values for total ACN and for each ACN decreased as temperature increased. In 222 particular, for the single ACNs the highest values were observed at temperatures from 25 to 35°C, 223 ranging from 1.11 and 3.02. Since higher Q_{10} value means higher ACN degradation, under the 224 present conditions adopted, Q_{10} is mainly affected by temperatures in the range 25-35°C. Moreover, 225 since most of the Q_{10} values were about 2.0, the increase of temperature by 10°C approximately doubled the decay rate (**Table 2**). In contrast to our results, Kechinski et al.²⁴ observed a higher Q_{10} 226 227 value (4.27 at the range from 40 to 50°C) in highbush blueberry juice, probably due to the high 228 content of water in juice with respect to the powder.

The values of activation energy (E_a) , half-life $(t_{1/2})$ and Q_{10} were calculated for TAA of the WB powder stored at different temperatures (**Table 3**). The TAA showed values of E_a (52.31 kJ mol⁻¹) and Q_{10} comparable to that obtained for the total ACNs, while the value of $t_{1/2}$ was higher, ranging from 130 days at 60°C to 1200 days at 25°C.

Additionally, the Arrhenius equation was used to predict the shelf-life of total ACNs when stored at 4 °C. Under these experimental conditions, the half-life time for total ACNs is up to 829 days and for TAA more than 10 years. 236 The logarithmic reduction kinetics of total ACNs (A) and TAA (B) of the WB powder stored at different temperatures are reported in Figure 5. The TAA and the content of ACNs 237 238 decreased with increasing temperature but the reduction of the TAA does not seem directly 239 correlated to that of the ACNs. Indeed, no significant difference (p=0.89) was observed in TAA 240 values at 25°C and 42°C. Moreover, the logarithmic decrease of TAA at 80°C and 60°C (1.5 and 241 0.3) was lower in comparison to the logarithmic reduction of total ACN content (2.5 and 1.5). This 242 result is not surprising since it has been reported that at high temperatures (i.e. 60°C and 80°C) 243 Maillard and caramelization reactions occur and the generated products show an increase of TAA.³⁴ These reactions can also occur in presence of hexoses and in absence of the aminic group.³⁴ 244

The maintenance of TAA related to loss of total ACNs was described for several processed blueberry products.^{11,35-36} This is probably due to the formation of antioxidant polymers, such as low molecular weight procyanidins, which balance the reduction of monomer ACNs during storage and maintain the TAA, as well.^{5,37}

In our study, the initial TAA value of the investigated wild blueberry powder was 58.5 mmol Trolox eq TE/100 g DW of product, similar to the data (52.9 mmol Trolox eq TE /100 g DW) obtained from fresh wild blueberry by Kalt et al.⁸ These results further confirm the importance of freeze drying process to preserve TAA. In fact, after storage for 50 days, the TAA was 48.7, 49, 41.9 and 22.5 eq TE/100 g of product, stored at 25°C, 42°C, 60°C and 80°C, respectively.

254 In summary, the degradation of ACNs in freeze-dried WB powder followed a first-order kinetic, 255 thus its storage at room temperature (25°C) reduced ACN content less in comparison to other 256 temperatures. The decrease of single ACN monomers may be attributed to the formation of ACN polymers through a mechanism which is not well understood. The TAA of the WB powder was 257 almost unchanged after storage at 42°C for 50 days, suggesting that other compounds (e.g. fiber, 258 259 polymers, Maillard reaction products) affect its antioxidant power. The use of this freeze-dried WB 260 powder for food ingredients may be important since the content of ACNs and the TAA are maintained longer, up to 130 days at 25°C, in comparison to other blueberry products. 261

262 Abbreviations

- 263 ACN(s), anthocyanin(s);
- 264 Dp-gal, delphinidin-galactoside;
- 265 Dp-glc, delphinidin-glucoside;
- 266 Dp-ara, delphinidin-arabinoside;
- 267 Cy-gal, cyanidin-galctoside;
- 268 Cy-glc, cyanidin-glucoside;
- 269 Cy-ara, cyanidin-arabinoside;
- 270 Pt-gal, petunidin-galactoside;
- 271 Pt-glc, petunidin-glucoside;
- 272 Pt-ara, petunidin-arabinoside;
- 273 Pe-gal, peonidin-galctoside;
- 274 Pe-glc, peonidin-glucoside;
- 275 Pe-ara, peonidin-arabinoside;
- 276 Mv-gal, malvidin-galctoside;
- 277 Mv-glc, malvidin-glucoside;
- 278 Mv-ara, malvidin-arabinoside;
- 279 Dp-glc-Ac, acetylated delphinidin-glucoside;
- 280 Cy-glc-Ac, acetylated cyanidin-glucoside;
- 281 Pt-glc-Ac, acetylated petunidin-glucoside;
- 282 Pe-glc-Ac, acetylated peonidin-glucoside;
- 283 Mv-gal-Ac, acetylated malvidin-galctoside;
- 284 Mv-glc-Ac, acetylated malvidin-glucoside;
- 285 DW, Dry weight;
- 286 TAA, total antioxidant activity;
- 287 TEAC, Trolox equivalent antioxidant capacity;

288	ASLT, accelerated shelf-life testing;
289	LC-ESI/MS, liquid chromatography coupled with electrospray ionization and mass spectrometry;
290	LSD, least significant difference;
291	RSD, relative standard deviation;
292	a _w , water activity;
293	<i>Ea</i> , activation energy;
294	$t_{1/2}$, half-life time;
295	Nd, not detectable .
296	
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301	
302	Notes

303 The authors declare no competing financial interest.

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404 FIGURE CAPTIONS

405 **Figure 1:** HPLC profile of the individual ACNs in WB (*Vaccinium angustifolium*) powder detected

- 406 at 520 nm. The compounds identified are: (1) Dp-gal, (2) Dp-glc, (3) Cy-gal, (4) Dp-ara, (5) Cy-glc,
- 407 (6) Pt-gal, (7) Cy-ara, (8) Pt-glc, (9) Pe-gal, (10) Pt-ara, (11) Pe-glc, (12) Mv-gal, (13) Pe-ara, (14)
- 408 Mv-glc, (15) Mv-ara, (16) Dp-glc-Ac, (17) Cy-glc-Ac, (18) Pt-glc-Ac, (19) Mv-gal-Ac, (20) Pe-glc409 Ac, and (21) Mv-glc-Ac.
- 410 Legend: HPLC, high performance liquid chromatography; ACNs, anthocyanins, WB, wild411 blueberry.
- 412
- 413 Figure 2: Decay (%) of total ACNs in the WB powder stored at (a) 25°C, (b) 42°C, (c) 60°C and

414 (d) 80 °C. Curves with different letters are significantly different at $P \le 0.05$

- 415 Legend: ACN (-): anthocyanin, ACN_Ac (\circ): Acetylated anthocyanin
- 416
- 417 **Figure 3**: Effect of temperature on slope (mean \pm SD) for glycosylated and acetylated ACN 418 degradation in WB powder stored at 25°C, 42°C, 60 °C.
- 419 *Data between curves (ACN vs ACN_Ac) at 42°C and 60°C are significantly different at $P \le 0.05$
- 420 ^{\pm,\dagger,\pm} Data between points (25°C, 42°C and 60°C) of the same curves are significantly different at P \leq 421 0.05.
- 422 Legend: ACN (-): anthocyanin, ACN_Ac (0): Acetylated anthocyanin
- 423

424 **Figure 4:** Effect of temperature on reaction rate constant (k) slope (mean \pm SD) for single ACN and

- 425 ACN_Ac degradation in WB powder stored at 25, 42 and 60 °C.
- 426 *Data between curves (ACN vs ACN_Ac) and within temperatures (25°C, 42°C and 60°C) of the
- 427 same curves are significantly different at $P \le 0.05$.
- 428 Legend: ACN (-): anthocyanin, ACN_Ac (0): Acetylated anthocyanin

- 430 **Figure 5:** Logarithmic reduction kinetics of total ACNs and TAA in WB powder stored at 25°C,
- 431 42°C, 60°C and 80 °C.
- 432 Legend: ACNs, anthocyanins; TAA, total antioxidant activity; WB, wild blueberry.

434 **Table 1:** Activation energy (E_a) and half-life ($t_{1/2}$) of total and individual ACNs of the WB powder 435 stored at 25°C, 42°C and 60°C.

Compound	Ea	t1	t _{1/2} (days)			
Compound	(kJ mol ⁻¹)	25°C	42°C	60°C		
Total ACNs	58.26	139	39	12		
Individual ACNs						
Dp-gal	57.82	212	60	18		
Dp-glc	45.44	131	49	19		
Dp-ara	64.85	256	62	16		
Cy-gal	72.17	460	95	21		
Cy-glc	55.72	234	69	22		
Cy-ara	38.98	117	49	21		
Mv-gal	73.54	608	122	27		
Mv-glc	55.81	162	48	15		
Mv-ara	65.29	261	63	16		
Pt-gal	69.81	374	81	19		
Pt-glc	18.14	86	58	40		
Pt-ara	51.40	611	199	69		
Pe-gal	84.15	549	87	15		
Acetylated ACNs						
Dp-glc-Ac	62.00	625	161	45		
Cy-glc-Ac	84.28	1948	310	54		
Mv-gal-Ac	27.07	936	519	296		
Mv-glc-Ac	62.15	295	76	21		
Pt-glc-Ac	51.29	542	177	61		
Pe-glc-Ac	nd	nd	nd	nd		

437 Legend: *Ea*, activation energy; $t_{1/2}$ half-life; ACNs, anthocyanins; Ac, acetylated; nd, not detectable;

438 WB, wild blueberry.

C	mound	Te	mperature (°	C)
C	ompound	25 to 35	42 to 52	60 to 70
То	tal ACNs	2.15	1.98	1.85
Indiv	idual ACNs			
Dp-gal		2.14	1.97	1.84
Dp-glc		1.82	1.71	1.61
Dn-ara		2 34	2 1 5	1 98

440 **Table 2:** Q_{10} values for the total and individual ACNs of the WB powder stored at different 441 temperatures.

Dp-gal	2.14	1.97	1.84
Dp-glc	1.82	1.71	1.61
Dp-ara	2.34	2.15	1.98
Cy-gal	2.58	2.34	2.14
Cy-glc	2.08	1.93	1.80
Cy-ara	1.69	1.60	1.52
Mv-gal	2.63	2.38	2.17
Mv-glc	2.08	1.93	1.80
Mv-ara	2.36	2.16	1.99
Pt-gal	2.50	2.27	2.09
Pt-glc	1.27	1.24	1.21
Pt-ara	1.96	1.83	1.72
Pe-gal	3.02	2.69	2.43
Acetylated ACNs			
Dp-glc-Ac	2.26	2.07	1.92
Cy-glc-Ac	3.02	2.70	2.43
Mv-gal-Ac	1.43	1.38	1.33
Mv-glc-Ac	1.11	1.10	1.09
Pt-glc-Ac	1.96	1.83	1.72
Pe-glc-Ac	nd	nd	nd
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442 Legend: ACNs, anthocyanins; Ac, acetylated; nd, not detectable; WB, wild blueberry.

443

444 **Table 3:** Activation energy (E_a), half-life ($t_{1/2}$) and Q_{10} values of total antioxidant activity (TAA) of

445 WB powder stored at 25°C, 42°C and 60°C.

Ea		$t_{1/2}$ (days)		Q10		
(kJ mol ⁻¹)	25°C	42°C	60°C	25 to 35°C	42 to 50°C	60 to 70°C
52.31	1212	387	131	1.99	1.85	1.74

446 Legend: *Ea*, activation energy; $t_{1/2}$, half-life; WB, wild blueberry.









