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Staphylococcus haemolyticus superinfection of legionella pneumonia during infliximab therapy

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ABSTRACT

We present the case of a 42-year-old man affected by psoriasis with Staphylococcus Haemolyticus superinfection of Legionella pneumonia during infliximab therapy. The introduction of compounds that block TNF- α has yielded great benefits for patients affected by selected autoimmune diseases that fail to respond to classic anti-inflammatory agents, but, on the other hand, has led to an increased susceptibility to infections, in particular of those caused by intracellular pathogens, such as L. Pneumophila. Emerging evidence suggests that legionellosis can be complicated by superinfection with other agents, including saprophytic microorganisms, among which coagulasenegative staphylococci. To our knowledge, this is the first report of systemic legionellosis with superinfection by S. Haemolyticus, an emerging nosocomial multi-resistant pathogen that commonly causes septicemia, osteomyelitis or endocarditis, but has not so far been associated with necrotizing pneumonia. Despite the optimal antimicrobial therapy for Staphylococcus spp. Pneumonia is still controversial, evidence suggests that in patients with confirmed positivity for methicillin resistant strains, particularly if sensitivity to vancomycin is suboptimal, linezolid should be the first choice therapy, being superior to vancomycin and teicoplanin.

Keywords: Infliximab; Legionella Pneumophila; Linezolid; Necrotizing Pneumonia; Psoriasis; Staphylococcus Haemolyticus

1. INTRODUCTION

On September 2010, a 42-year-old man presented to the emergency department because of fever, productive cough, and chest pain. He had a history of psoriasis, pre-

viously treated with acitretine 25 mg, which was discontinued because of dyslipidemia. Nine months before the current admission he required hospitalization for erythrodermic psoriasis. Since then, he was treated with infliximab, and received the eighth infusion one week before presentation. Due to the incomplete control of disease activity, he was concurrently started on methotrexate (10 mg s.c.). Five days before admission he reported the abrupt onset of fever which reached 39°C, dry cough, asthenia, and burning pain in the right subscapular region, which were unresponsive to acetaminophen and amoxicillin-clavulanate.

On admission the temperature was 39.5°C, blood pressure 125/80 mmHg, pulse 115, respiratory rate 24, peripheral oxygen saturation 98% while he was breathing ambient air. Clinical examination revealed bilateral latero-cervical lymph nodes (soft, non-tender, and mobile, < 2 cm), regular heart activity, slight rales at the left lower lobe, and splenomegaly. An electrocardiogram showed sinus tachycardia without alterations of the ST tract; biochemical test are shown in Table 1. Chest Rx showed a right middle lobe infiltrate (Figure 1(a)). Abdominal ultrasonography was only notable for splenomegaly (153 mm). Oxygen supplementation and hydration were provided.

Microbiological analyses of blood were repeatedly negative and sputum was negative for M. tuberculosis and other microorganisms; L. Pneumophila serotype 1 antigen was detected in urine samples. Intravenous levofloxacin 750 mg/day was started with improvement in systemic symptoms, and correction within 5 days of the alterations of creatine-phosphokinase levels, and hyponatremia, (Table 1). Because of the persistence of unremitting fever (> 38°C), and increased inflammatory markers, azithromycin 500 mg/day was added on day 5. Blood cultures remained negative, as well as autoimmunity markers.



One week after admission, the patient underwent a chest computed tomography (CT) scan showing upper and middle right lobe extensive infiltrate with an aerial bronchogram and patchy ground-glass infiltrates in the upper left lobe, minimal pleural and pericardial effusions and multiple bilateral supraclavicular, paratracheal, undercarinal, and axillary lymphoadenopathies (up to 1.5 cm). On day 8 Staphylococcus Haemolyticus was isolated from the sputum, but initially interpreted as a contaminant (supplementary Table 1). A bronchoscopy was performed on day 10: the airways appeared normal and cytologic examination of bronchoalveolar-lavage (BAL) specimens of the right lobes revealed acute inflammation, but not malignant cells. Tests were negative for M. Tubercolosis, Aspergillus spp, P. Jirovecii, Epstein-Barr Virus, Cytomegalovirus, Respiratory Syncythial virus multiplex PCR, whereas a nested PCR-based assay confirmed the positivity for L. Pneumophila.

Broad-spectrum empirical antimicrobical therapy was started including piperacillin-tazobactam and imipenem (day 10); while clindamycin 600 mg t.i.d. i.v. was added on day 14 according to the *S. Haemolyticus* antibiogram (supplementary table), without benefit.

On day 15 a CT scan was repeated, which showed the progression of the right pulmonary infiltrate with areas of colliquation and excavation (**Figure 1(b)**). Rifampicin replaced previous empirical therapy on day 17 because of the reported synergic effect with fluoroquinolones, and macrolides in the treatment of resistant legionellosis in immunosuppressed patients [1].

To exclude an immunosuppression related lymphomas, laterocervical lymph node biopsy was performed, which showed a reactive inflammatory pattern. A positron emission tomography CT scan showed inflammation in the right upper and lower pulmonary lobes and in the mediastinal lymph nodes.

Several reports highlited an increased risk of infection by atypical pathogens associated with infliximab, including *M. Tubercolosis*, *Listeria*, *Pneumococcus*, *Legionella*, *Salmonella*, *and Bartonella spp.*), fungi, and viruses [2]. Although the patient chest Rx and protein purified derivative intradermoreaction were negative before starting infliximab and BAL was PCR negative for M.Tubercolosis, a quantiferon test for Mycobacterium was performed, but it confirmed negativity for M.Tubercolosis. An echocardiogram was carried out on day 21, with no evidence of endocarditis.

To exclude a pulmonary superinfection, a CT-guided needle-aspiration of the pulmonary lesion was performed, with isolation of rare colonies of S. Haemolyticus (supplementary **Table 1**). Vancomycin (minimal inhibitory concentration (MIC): 1) was started i.v. (day 22) without response. In view of the sub-optimal MIC,

incomplete pulmonary diffusion of vancomycin, and the relatively low plasma concentration achieved after treatment titration (12.5 μ g/mL), linezolid (600 mg td i.v.) was introduced on day 33, with immediate remission of fever and normalization of inflammatory indices (**Table 1**). The patient was discharged on day 45. A chest CT scan after 4 weeks demonstrated resolution of the inflammatory infiltrates and a reduction of the excavated areas.

This case is notable for several aspects. The introduction into clinical practice of compounds that block TNF- α has yielded great benefits for patients affected by selected autoimmune diseases that fail to respond to classic anti-inflammatory agents, but, on the other hand, has led to an increased susceptibility to infections [3], in particular of those caused by intracellular pathogens [4], and L. Pneumophila pneumonia has been reported as a life-threatening infection that frequently complicates anti-TNF- α therapy [3]. In line with our experience, evidence is accumulating that legionellosis, particularly in patients under immunosuppressive therapy, can be complicated by superinfection with other agents, including saprophytic microorganisms. Furthermore, positivity for Legionella DNA may persist despite appropriate treatment in immunocompromised patients, and lung colliquative necrosis supports a diagnosis of coinfection with other bacterial agents. To our knowledge, this is the first report of systemic legionellosis with superinfection by S. Haemolyticus, an emerging nosocomial multiresistant pathogen that commonly causes septicemia, osteomyelitis or endocarditis, but has not so far been associated with necrotizing pneumonia [5].

Finally, despite the optimal antimicrobial therapy for Staphylococcus spp. pneumonia is still controversial, evidence suggests that in cases with confirmed positivity

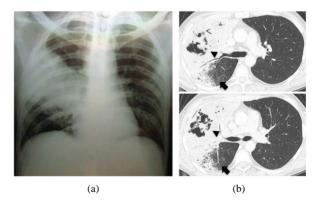


Figure 1. (a) Chest X-ray on admission: right middle lobe infiltrate and left mild perihilar infiltrate; (b) Chest CT scan (day 15): upper right lobe extensive infiltrate with areas of colliquation and excavation (arrowheads) and patchy ground-glass infiltrates in the upper right lobe (arrows).

Table 1. Laboratory data.

	Reference	Admission	Day 5	Day 22	Day 33	Discharge (day 45)
Hb (g/dl)	13.5 - 17.5	14.8	11.3	9.5	9.1	9.6
WBCs (µl)	4.8 - 10.8	15250	12720	8090	4560	6390
Neutrophils %	-	91	77	68	73	59
CRP (mg/dl)	< 0.5	67.05	16.89	4.80	8.14	0.81
Procalcitonin (ng/mL)	0.02 - 0.06	17.16	na	0.06	0.18	0.08
AST (UI/ml)	5 - 38	80	81	24	17	21
ALT (UI/ml)	5 - 41	33	68	25	14	13
CPK (UI/ml)	38 - 174	1806	1065	16	15	16
LDH (UI/ml)	240 - 480	1076	691	307	341	256
PT ratio	0.89 - 1.16	1.3	1.24	1.23	1.2	1.07
aPTT ratio	0.85 - 1.18	1.38	1.06	1.12	1.15	1.10
Creatinine (mg/dL)	0.50 - 1.20	0.82	0.48	0.5	0.60	0.73
Na ⁺ (mEq/L)	135 - 145	125	140	140	141	142
K^{+} (mEq/L)	3.30 - 5.10	3.6	4.1	4	3.1	4.3
Cl ⁻ (mEq/L)	96 - 108	90	103	101	101	104
TSH (mIU/mL)	0.280 - 4.300	4.07	na	na	na	na
pН	7.35 - 7.45	7.51	na	7.54	na	7.53
pO ₂ (mmHg)	83 - 108	72	na	78	na	102
pCO ₂ (mmHg)	32 - 48	26	na	28	na	36
HCO_3^- (mEq/L)	24 - 26	24	na	23.9	na	30
Lactate mEq/L	0.5 - 2.0	1.5	na	1.2	na	1.4
BE	-2 - +2	-2.3	na	1.9	na	7.4

Hb: hemoglobin; WBC: white blood cells; CRP: C reactive protein; AST: aspartate aminotransferase; ALT: alanine aminotransferase; CPK: creatine phosphokinase; LDH: lactate dehydrogenase; PT: prothrombin time; aPTT: activated partial thromboplastin time; BE: base excess, na: not available.

for methicillin resistant strains, particularly if sensitivity to vancomycin is suboptimal as in our case, linezolid might be superior to vancomycin and teicoplanin because of the better penetration into lung allowing to reach sustained concentrations above the MIC.

In conclusion, clinicians should be aware of the increased risk of severe and difficult-to-treat infections by atypical and saprophytic pathogens in the increasing number of patients treated anti-TNF- α agents. Among them, S. Haemolyticus is an emergent nosocomial pathogen, which may be associated with necrotizing pneumonia. Long-term follow-up and systematic assessment of infective episodes may help to better define the clinical risk carried by anti-TNF- α agents in everyday clinical practice, and possibly to identify risk factors for these complications.

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