



UNIVERSITI PUTRA MALAYSIA

***IN VITRO ANTIOXIDANT AND ANTI-ATHEROSCLEROTIC ACTIVITIES
OF PHENOLIC SAPONIN RICH FRACTION OF COSMOS CAUDATUS
KUNTH***

SAID FAROQ SAIYID MOSHAWIH

FPSK(m) 2016 70



***In Vitro ANTIOXIDANT AND ANTI-ATHEROSCLEROTIC ACTIVITIES
OF PHENOLIC SAPONIN RICH FRACTION OF *Cosmos caudatus*
KUNTH***



**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of
Master of Science**

November 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfillment of the requirement for the Degree of Master of Science

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OF PHENOLIC SAPONIN RICH FRACTION OF *Cosmos caudatus*
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By

SAID FAROOQ SAIYID MOSHAWIH

November 2016

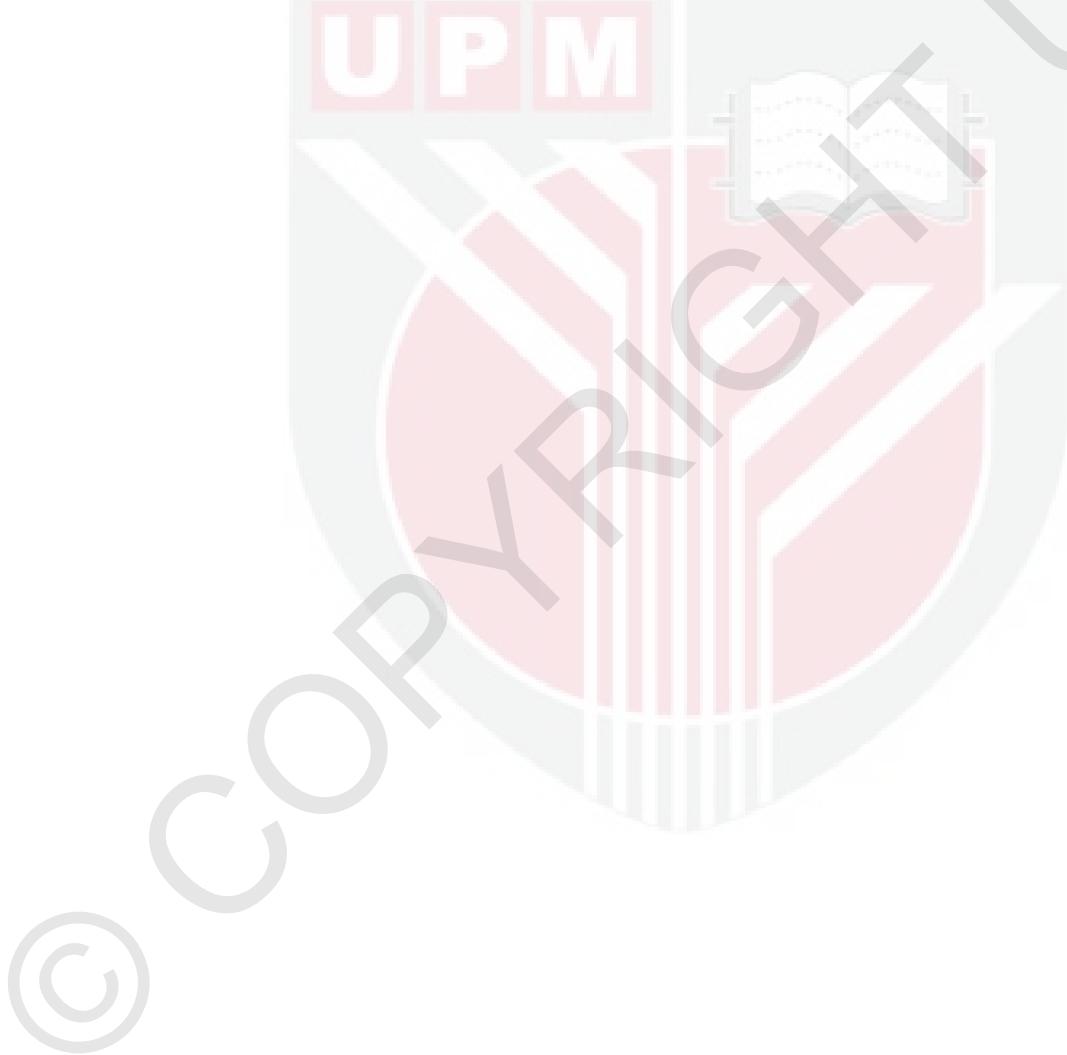
**Chairman : Professor Muhammad Nazrul Hakim Bin Abdullah, PhD
Faculty : Medicine and Health Sciences**

Cosmos caudatus or Ulam raja as it is known in Malaysia, is a tropical plant that has been known in South East Asia for many traditional uses, some were proved by pharmacological studies, while others yet to be validated. In this study, we are inspecting the atheroprotective effects for ulam raja extract/fractions through *in vitro*. The shoot samples were collected fresh from the plant stem, oven dried and grinded. A 100 gm sample was macerated in 50% (v/v) ethanol, twice for 48hours each time with intermittent shaking, after extract pooling, solvent was removed under reduced pressure and lyophilized to produce the crude extract. Subsequently, the crude extract was fractionated to produce the butanol and the aqueous fractions. In order to determine the phenolic, saponin and steroidal saponin contents, we employed folin-ciocalteu, vanillin-sulfuric acid and anisaldehyde-sulfuric acid methods respectively. Butanol fraction was the phenolic-saponin rich fraction (PSRF) due to the fact that it possesses the richest total phenolic, saponin and steroidal saponin contents (88.45 mg GAE/g DW, 64.23 and 23.3 mg DE/g DW) respectively ($P<0.05$), compared to the other *C. caudatus* derived fractions.

Similarly, butanol fraction showed the highest antioxidant activity in four assays namely, DPPH, ABTS, iron chelating and BCB activity assays. Antioxidant activities for butanol fraction were (72.65 mg TEAC/ g DW), (94.47 mg TEAC/ g DW), (16.03 mg EDTA/ g DW) and (19.43 mg TEAC/ g DW) respectively. Cytotoxic effects of crude, butanol and aqueous fractions were determined by MTT assay in rat aortic smooth muscle cell line (A-10) for 24 hours, and IC_{50} were as follows 460.78, 313.96 and 949.2 μ g/mL respectively. IC_{50} for the three fractions was going in the same trend of the previous assays, as butanol was the most potent, followed by crude, and lastly the aqueous fraction. Three concentrations (IC_{10} , IC_{25} and IC_{50}) of the three fractions were used to treat A-10 cell line to assess their activities in inhibiting migration and invasion of cells using Boyden chamber assay. Butanol fraction Intermediate

concentration (IC_{25}) demonstrated the strongest activity to inhibit smooth muscle cells from migration and invasion in *in vitro* assays, as they were reduced by 53.93% and 59.94% respectively compared to untreated control cells. However, the invasion inhibition index referred to that butanol and crude mild concentrations (IC_{10}) showed the highest index value ~68%, which means the best cell invasion to cell migration inhibition *in vitro*.

Taken together, results strongly support that PSRF from *C. caudatus* can be used to reduce atherosclerotic plaque formation by reducing its key mechanism represented by SMCs migration and invasion, and its subsequent events such as myocardial infarction and strokes. This can be attributed to the high contents of phenolics, saponins and vitamins, and their ability to reduce the oxidative stress and free radicals, in addition to the other mechanism of action of other flavonoids.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

AKTIVITI ANTIOKSIDAN *In Vitro* DAN ANTI-ATHEROSCLEROTIC DALAM *Cosmos caudatus* YANG KAYA DENGAN BAHAGIAN FENOLIK-SAPONIN

Oleh

SAID FAROOQ SAIYID MOSHAWIH

November 2016

Pengerusi : Profesor Muhammad Nazrul Hakim Bin Abdullah, PhD
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Cosmos caudatus (*C.Caudatus*) atau Ulam raja seperti yang dikenali di Malaysia adalah sejenis tumbuhan tropika yang terkenal di Asia Tenggara untuk pelbagai kegunaan tradisional. Walaubagaimanpun, beberapa penggunaannya telah disahkan melalui kajian-kajian farmakologi namun yang selainnya yang masih belum disahkan. Kajian ini ekstrak/pecahan ulam raja untuk mengkaji kesan pencegahan *atherosclerosis* melalui kajian *in vitro*. Pucuk segar yang telah dipetik dari batang tumbuhan, dikeringkan di dalam ketuhar dan dikisar untuk dijadikan sampel. Sampel sebanyak 100g dihancurkan sambil direndam di dalam larutan (ip/ip) ethanol 50% selama 48 jam. Larutan sampel tersebut digoncang secara berkala. Proses penghancuran sampel di dalam larutan ethanol ini diulang sekali lagi. Supernatan dibuang dan ekstrak tersebut dikumpulkan serta di *lyophilize* untuk menghasilkan ekstrak mentah. Kemudian, ekstrak mentah tersebut dipisahkan untuk mendapatkan ekstrak butanol dan akueus. Kaedah *folin-ciocalteu*, asid *vanillin-sulfuric* and asid *anisaldehyde-sulfuric* masing-masing digunakan untuk mengenalpasti kandungan *phenolic*, *saponin* dan steroid *saponin*. Ekstrak butanol telah dikenalpasti sebagai *phenolic-saponin rich fraction (PSRF)* kerana ia menunjukkan jumlah kandungan jumlah *phenolic* paling tinggi, jumlah *saponin* dan jumlah steroid *saponin* masing masing adalah (88.45 ± 2.7 mg GAE/g DW, 64.23 ± 2.5 dan 23.3 ± 0.5 mg DE/g DW) pada ($P<0.05$) berbanding dengan ekstrak lain dari *C.caudatus*.

Bersamaan dengan itu, ekstrak butanol menunjukkan aktiviti anti-oksida paling tinggi di dalam empat assay iaitu DPPH, ABTS, *iron chelating* dan assay aktiviti BCB. Aktiviti antioksidan bagi ekstrak butanol di dalam setiap assay masing-masing adalah (72.65 mg TEAC/ g DW), (94.47 mg TEAC/ g DW), (16.03 mg EDTA/ g DW) dan (19.43 mg TEAC/ g DW). Kesan sitotoksik untuk 24 jam bagi ekstrak mentah, ekstrak butanol dan ekstrak akueus terhadap sel otot licin vaskular Aorta tikus (A-10) ditentukan melalui assay

MTT dan nilai IC₅₀ masing-masing adalah seperti berikut 460.78, 313.96 dan 949.2 µg/ml. Nilai IC₅₀ bagi ketiga-tiga ekstrak menunjukkan corak yang sama dengan assay sebelum ini, dimana ekstrak butanol mempunyai kekuatan paling tinggi diikuti dengan ekstrak mentah dan ekstrak akueus. Tiga konsentrasi (IC₁₀, IC₂₅ dan IC₅₀) untuk setiap ekstrak digunakan untuk merawat sel A-10 bagi menilai aktiviti-aktiviti setiap ekstrak dalam menghalang migrasi dan invasi cell A-10 menggunakan *Boyden chamber assay*. Dalam assay *in vitro*, konsentrasi pertengahan (IC₂₅) bagi ekstrak butanol menunjukkan aktiviti paling tinggi dalam menghalang migrasi dan invasi sel otot licin, di mana aktiviti migrasi dan invasi masing-masing mengalami pengurangan sebanyak 53.93% dan 59.94% berbanding dengan sel kawalan yang tidak dirawat. Bagaimanapun, indeks penghalang invasi yang merujuk kepada butanol dan konsentrasi sederhana mentah (IC₁₀) menunjukkan nilai indeks tertinggi ~68%, yang membawa maksud invasi sel terbaik kepada penghalangan migrasi sel secara *in vitro*.

Secara keseluruhan, hasil keputusan menyokong dengan kukuh bahawa PSRF dari *C. caudatus* boleh digunakan untuk mengurangkan pembentukkan plak *atherosclerosis* dengan mengurangkan mekanisma utamanya iaitu migrasi SMC dan invasi serta mampu mengurangkan kejadian berkaitan seperti infarksi myokardia dan angin ahmar. Ini mungkin disebabkan oleh kandungan *phenolics*, *saponins* dan vitamin yang tinggi serta kebolehannya untuk mengurangkan keadaan tekanan oksidatif dan radikal bebas, termasuk mekanisma tindakan yang lain bagi flavonoid yang berbeza.

ACKNOWLEDGEMENTS

Glory and praise be to Allah, the lord of the world, the omnipotent, omniscient and omnipresent, with his blessing and bounties, I was able to finish this thesis as required. My gratitude for ALLAH for blessing me with strength, courage and patience to complete the journey of my research and thesis.

I am heartily thankful to Professor Dr. Muhammad Nazrul Hakim Bin Abdullah as the chairperson of my supervisory committee for his patience, helpful advices, inspiring guidance and support throughout the duration of my project. I would like to forward special appreciations and thanks to Dr. Manraj Singah Cheema, my co-supervisor for the constructive criticism, understanding and advices.

I also wish to express my thanks to the staff of faculty of medicine and health sciences, especially pharmacology and toxicology lab and physiology lab staff, and all friends who participated directly and indirectly in my success. My grateful will be mainly for my father and mother who endured a lot of sufferings and pains to grow me up and struggle to reach this success.

I certify that a Thesis Examination Committee has met on 11 November 2016 to conduct the final examination of Said Faroq Saiyid Moshawih on his thesis entitled "In Vitro Antioxidant and Anti-Atherosclerotic Activities of Phenolic Saponin Rich Fraction of *Cosmos caudatus* Kunth" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

μl	Microliter
$^1\text{O}_2$	Singlet oxygen
ABC	ATP-binding cassette transporter
ABTS	2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid)
AI	Atherogenic index
AIDS	Acquired immunodeficiency syndrome
Akt	Protein kinase B
AP-1	Activation protein -1
AqF/Aq	Aqueous fraction
ATS	Atherosclerosis
BCB	B-carotene bleaching assay
BF	Butanol fraction
BHA	Butylated hydroxyanisole
BHT	Butylated hydroxytoluene
CD36	Cluster of differentiation 36
CEE/Cr	Crude ethanolic extract
COX	Cyclooxygenase enzyme
Cu-ZnSOD	Copper-Zinc superoxide dismutase
DE	Diosgenin equivalent
DMEM	Dulbecco's Modified Eagle's medium
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DPPH	2,2-diphenyl-1-picrylhydrazyl
ECM	Extracellular matrix

EDTA	Ethylenediaminetetraacetic acid
eNOS	Endothelial nitric oxide synthase
ERK	Extracellular signal regulated kinases
FBS	Fetal bovine serum
FC	Folin–Ciocalteu
FH	Familial hyperlipidemia
FRAP	Ferric reducing ability of plasma
GAE	Gallic acid equivalent
GM-CSF	Granulocyte macrophages colony stimulating factor
GPx	Glutathione peroxidase
GSH	Glutathione
H ₂ O ₂	Hydrogen peroxide
HDL	High density lipoprotein
HPLC	High performance liquid chromatography
IC	Inhibitory concentration
ICAM-1	Intracellular adhesion molecule -1
IFN-γ	Interferon gamma
IKB	Inhibitor of kappa B
IL	Interleukin
iNOS	Inducible nitric oxide synthase
JNK	c-Jun N-terminal kinases
LDL	Low density lipoprotein
LDLR	Low density lipoprotein receptor
LPL	Lipoprotien lipase
MAPK	Mitogen-activated protein kinase

mBar	Milli bar
MCP-1	Monocyte chemotactic protein-1
MHC	Major histocompatibility complex
MI	Myocardial infarction
mM	Millimolar
MMLDL	Minimally modified low density lipoprotein
MMP	Matrix metalloproteinase enzyme
MnSOD	Manganese superoxide dismutase
MTT	Microculture tetrazolium assay
NADP	Nicotinamide adenine dinucleotide phosphate
NF- κ B	Nuclear factor kappa B
NO $^{\cdot}$	Nitric oxide
O ₂ $^{\cdot\cdot}$	Superoxide
OD	Optical density
OH $^{\cdot}$	Hydroxyl radical
ONOO $^{\cdot}$	Peroxynitrite
ORAC	Oxygen radical absorbance capacity
oxLDL	Oxidized low density lipoprotein
PAF	Platelet activating factor
PBS	Phosphate buffered saline
PCSK9	Proprotein convertase subtilisin/kexin type 9
PGC1	Peroxisome proliferator-activated receptor gamma coactivator 1
PI3K	Phosphoinositide 3-kinases
PPAR	Peroxisome proliferator-activated receptor
PSRF	Phenolic-saponin rich fraction

R	Pearson correlation
R.S.	Reactive species
RNS	Reactive nitrogen species
RO [·] ₂	Peroxyl radical
ROS	Reactive oxygen species
RPM	Round per minute
SMC	Smooth muscle cells
SO ₂	Sulfur dioxide
SOD	Superoxide dismutase
STAT-1	Signal transducers and activators of transcription -1
TC	Total cholesterol
TCM	Traditional Chinese medicine
TEAC	Trolox equivalent antioxidant capacity
TG	Triglyceride
T _h	T-helper lymphocytic cells
TIMP	Tissue inhibitor of metalloproteinases
TLC	Thin layer chromatography
TNF	Tumor necrosis factor
TPC	Total phenolic content
TSC	Total saponin content
TSSC	Total steroidal saponin content
USDA	United state department of agriculture
VCAM-1	Vascular cell adhesion molecule -1
VSMC	Vascular smooth muscle cells

CHAPTER 1

INTRODUCTION

Patients had become more dependent on herbal medicines in the second half of the 20th century than before, this was proceeded by fascination with patent medicines in the 19th century, when the scientific methods were advanced and preferred (Winslow et al., 1998). Nowadays, and due to the increasing use of natural resources by pharmaceutical industry and medicine synthesis, people realize that returning back to the nature is the best way to decrease side effects and high costs of synthetic medicines (Alsarhan et al., 2014). This is more appreciated in tropical and subtropical regions which contain 60% of the total world plant species (Goh et al., 1993). Due to the wide variety of flora and fauna in Malaysia, people are aware of its benefits as a rich source of antioxidants and vitamins (Bunawan et al., 2014). Eighty percent of plant species recorded in the Malaysian state of Sabah are being used by locals for traditional treatments and as a flavoring agents in food (Kulip et al., 2010).

Among this enormous legacy of plant species, *Cosmos caudatus* is within the most prominent and beneficial ones. *C. caudatus* in Malaysia is locally called "Ulam Raja" which means King's salad. (Figure 1.1). It is being used in food for its pungent flavor and distinct aroma, cooked or raw in salads (Rasdi et al., 2010). *Cosmos caudatus* is a short-lived annual plant, with aromatic leaves. It belongs to Asteraceae – formerly known as Compositae – family, only two species are available in Malaysia and around 26 worldwide (Abas, 2005). It grows up to 1.7 meters tall, and flowers at the 10th week of seeding, with a purple to pink and rarely white florets. The leaves are finely dissected, between 10 to 20 cm in length (Mediani, 2012). (Figure. 1.1). Spaniards brought *C. caudatus* along from tropical America to South East Asia during their travelling for its benefits as it was toning up their bodies for the long journey and poor nutrition conditions (Bodeker et al., 2009).



Figure 1.1 : *Cosmos caudatus* Kunth leaves and flowers.

Traditionally, *C. caudatus* is known for many medicinal uses, some of them were studied and others are being investigated and documented. Local people have used it for claims such as cleansing blood and bone reduced mineral density (Abas et al., 2006; Mediani et al., 2012). *Cosmos caudatus* Pharmacological reports proved its benefits as a bone healing herb in post-fracture (Rufus et al., 2015), and as a natural supplement to stop osteoporosis in ovariectomized rats (Mohamed et al., 2012; Mohamed et al., 2013). *Cosmos caudatus* was also found to possess a comparable efficacy with a currently used anti-hypertensive and diuretic medicines such as captopril and hydrochlorothiazide respectively (Amalia et al., 2012). Antibacterial, antifungal in addition to antidiabetic properties were documented in *in vitro* studies for *C. caudatus* (Fuzzati et al., 1995; Javadi et al., 2014; Loh et al., 2011; Ragasa et al., 1997; Rasdi et al., 2010).

Cosmos caudatus has also antihyperlipidemic properties, as described in animal model-induced hyperlipidemia (Perumal et al., 2014). Antimutagenic benefits, were tested by Ragasa et al. (1997) and Lee et al. (2011). Antioxidant activity has been described extensively by most of studies done on *C. caudatus* (Abas et al., 2003; Huda-Faujan et al., 2009; Mustafa et al., 2010; Shui et al., 2005). Furthermore, *in vivo* antioxidant and chemoprotective attributes have been recently documented by Abdullah et al. (2015).

Many flavonoids, phenols, phenylpropanoids, ascorbic acid, carotenoids, and terpenes were found in *C. caudatus* (Andarwulan et al., 2012). In addition to that, vitamins, carbohydrates, amino acids, minerals, organic acids and

essential oils were also detected (Javadi et al., 2014; Lee et al., 2011). The presence of other phytochemicals such as saponins and alkaloids were characterized by a preliminary tests in *C. caudatus* extracts by using bubble testing and TLC respectively (Musa et al., 2011; Rasdi et al., 2010).

Atherosclerosis is a chronic inflammatory disease for blood arterial walls, which allows for lipid accumulation and the formation atherosclerotic plaque. Beside genetic and environmental factors, inflammatory causes have been studied as an interpretation for the pathogenesis of atherosclerosis (Marín-García, 2014). Accumulation of lipids underneath blood vessels epithelium is the main feature of atherosclerosis (Djekic et al., 2015). This happens due to a sequence of events, starts from endothelial injury, and is followed by lipoproteins accumulation and blood monocytes mobilization to the site of action. Elevated levels of oxidized low-density lipoprotein (oxLDL) increase its engulfment by macrophages, which had differentiated from monocytes, and consequently, the formation of foam cells grows. Thereafter, these activated macrophages release factors to indicate the inflammatory status, and finally, the smooth muscle cells migrate to the intima layer and the atherosclerotic plaque developed (Palozza et al., 2010).

1.1 Study Justification

Currently, atherosclerosis is treated by three categories of medicines; antiplatelet aggregation, vasodilators and antihyperlipidemics (Bays, 2012). Up to date, there are no specific anti-atherosclerotic drugs. Even though atherosclerosis is defined as a chronic inflammatory disease, but anti-inflammatory drugs are not used for this purpose, except for acetylsalicylic acid, which is used for its antiplatelet aggregation properties (Kraus et al., 2014). On the other hand, since that the inflammatory cytokines play a significant role during the formation of atherosclerosis, therefore, anti-cytokine therapies have been studied on early stages of the subclinical atherosclerosis. They may form a promising trend for preventing atherosclerosis, but up to date, they still under investigation (Kirichenko et al., 2016).

The widely used medicines for both atherosclerosis and hyperlipidemia are statins. They have many beneficial activities in reducing LDL, TG, total cholesterol as well as their antiatherogenic properties. On the other flip, statins induce many side effects such as myopathies, hepatotoxicity and rhabdomyolysis. On top of this, upon long-term use, resistance may appear for statins due to genetic polymorphism (Reiner, 2014). Collectively, we found that it is imperative to seek a treatment for atherosclerosis from natural resources. *Cosmos caudatus* extract has been reported to contain a wide range of flavonoids and phenolics, in addition to saponins, alkaloids, and vitamins. Thus, this richness in bioactive constituents is promising to inhibiting atherosclerosis development.

1.2 Research objectives

1.2.1 General Objective

The general objective of this study is to evaluate the atheroprotective and antioxidant activities of *C. caudatus* extract and derived fractions *in vitro*.

1.2.2 Specific objectives

The specific objectives of this study are as follows:

1. To extract *C. caudatus* shoots to produce the crude ethanolic extract (CEE). Then, to fractionate the CEE into two more fractions; butanol fraction (BF) and aqueous fraction (AqF), and to determine the total phenolic contents (TPC), total saponin contents (TSC) and total steroidal saponin content (TSSC) for the produced extract/fractions.
2. To determine the antioxidant and free radical scavenging activities for *C. caudatus* extract/fractions using DPPH, ABTS, iron chelating and β -carotene bleaching assays.
3. To evaluate the cytotoxic and anti-atherosclerotic actions of CEE, BF and AqF on rat aortic vascular smooth muscle cell line, by employing microculture tetrazolium (MTT), and migration and invasion assays respectively.

REFERENCES

- Abas, F. (2005). Phytochemical and Biological Activity Studies of Cosmos Caudatus and Curcuma Mangga and the Online Characterization of Bioactive Fractions from Melicope ptelefolia. (PhD Thesis), Universiti Putra Malaysia, Malaysia.
- Abas, F., Lajis, N., & Kalsom, Y. U. (2003). Antioxidative and radical scavenging properties of the constituents isolated from Cosmos caudatus Kunth. *Natural Product Sciences*, 9(4), 245-248.
- Abas, F., Lajis, N. H., Israf, D., Khozirah, S., & Kalsom, Y. U. (2006). Antioxidant and nitric oxide inhibition activities of selected Malay traditional vegetables. *Food Chemistry*, 95(4), 566-573.
- Abdel-Hamid, A. A. M., & Firgany, A. E.-D. L. (2015). Vitamin E supplementation ameliorates aflatoxin B1-induced nephrotoxicity in rats. *Acta Histochemica*, 117(8), 767-779.
- Abdullah, A., Dhaliwal, K. K., Roslan, N. N. F., Lee, C. H., Kalaiselvam, M., Radman, H. M., Saad, Q. H. M., Yusof, K., & Jaarin, K. (2015). The effects of Cosmos caudatus (Ulam Raja) on detoxifying enzymes in extrahepatic organs in mice. *Journal of Applied Pharmaceutical Science*, 5(1), 082-088.
- Akishima, Y., Akasaka, Y., Ishikawa, Y., Zhang, L., Kiguchi, H., Ito, K., Itabe, H., & Ishii, T. (2005). Role of macrophage and smooth muscle cell apoptosis in association with oxidized low-density lipoprotein in the atherosclerotic development. *Modern Pathology*, 18(3), 365-373.
- Albini, A., & Benelli, R. (2007). The chemoinvasion assay: a method to assess tumor and endothelial cell invasion and its modulation. *Nature Protocols*, 2(3), 504-511.
- Albini, A., Iwamoto, Y., Kleinman, H., Martin, G., Aaronson, S., Kozlowski, J., & McEwan, R. (1987). A rapid in vitro assay for quantitating the invasive potential of tumor cells. *Cancer Research*, 47(12), 3239-3245.
- Alsarhan, A., Sultana, N., Al-Khatib, A., & Mohammed Rafiq Abdul, K. (2014). Review on Some Malaysian Traditional Medicinal Plants with Therapeutic Properties. *Journal of Basic & Applied Sciences*, 10(14), 214-226.
- Amalia, L., Anggadiredja, K., Fidrianny, I., & Inggriani, R. (2012). Antihypertensive Potency of Wild Cosmos (Cosmos caudatus Kunth, Asteraceae) Leaf Extract. *Journal of Pharmacology & Toxicology*, 7(8), 359-368.

- Andarwulan, N., Batari, R., Sandrasari, D. A., Bolling, B., & Wijaya, H. (2010). Flavonoid content and antioxidant activity of vegetables from Indonesia. *Food Chemistry*, 121(4), 1231-1235.
- Andarwulan, N., Kurniasih, D., Apriady, R. A., Rahmat, H., Roto, A. V., & Bolling, B. W. (2012). Polyphenols, carotenoids, and ascorbic acid in underutilized medicinal vegetables. *Journal of Functional Foods*, 4(1), 339-347.
- Arumugam, B., Palanisamy, U. D., Chua, K. H., & Kuppusamy, U. R. (2016). Potential antihyperglycaemic effect of myricetin derivatives from *Syzygium malaccense*. *Journal of Functional Foods*, 22, 325-336.
- Baccou, J., Lambert, F., & Sauvaire, Y. (1977). Spectrophotometric method for the determination of total steroidal sapogenin. *Analyst*, 102(1215), 458-465.
- Balasubramanian, D. (2005). Photodynamics of Cataract: An Update on Endogenous Chromophores and Antioxidants. *Photochemistry and Photobiology*, 81(3), 498-501.
- Banerjee, A., Basu, K., & Sengupta, P. K. (2007). Effect of β -cyclodextrin nanocavity confinement on the photophysics of robinetin. *Journal of Photochemistry and Photobiology B: Biology*, 89(2–3), 88-97.
- Baranski, M., Srednicka-Tober, D., Volakakis, N., Seal, C., Sanderson, R., Stewart, G. B., Benbrook, C., Biavati, B., Markellou, E., Giotis, C., Gromadzka-Ostrowska, J., Rembialkowska, E., Skwarlo-Sonta, K., Tahvonen, R., Janovská, D., Niggli, U., Nicot, P., & Leifert, C. (2014). Higher antioxidant and lower cadmium concentrations and lower incidence of pesticide residues in organically grown crops: a systematic literature review and meta-analyses. *The British Journal of Nutrition*, 112(5), 794-811.
- Bays, H. E. (2012). Adiposopathy, Diabetes Mellitus, and Primary Prevention of Atherosclerotic Coronary Artery Disease: Treating “Sick Fat” Through Improving Fat Function with Antidiabetes Therapies. *The American Journal of Cardiology*, 110(9, Supplement), 4B-12B.
- Bhullar, K. S., Lassalle-Claux, G., Touaibia, M., & Rupasinghe, H. P. V. (2014). Antihypertensive effect of caffeic acid and its analogs through dual renin–angiotensin–aldosterone system inhibition. *European Journal of Pharmacology*, 730, 125-132.
- Bjelakovic, G., Nikolova, D., Simonetti, R., & Gluud, C. (2008). Systematic review: primary and secondary prevention of gastrointestinal cancers with antioxidant supplements. *Alimentary Pharmacology & Therapeutics*, 28(6), 689-703.

- Blanco-Colio, L. M., Tuñón, J., Martín-Ventura, J. L., & Egido, J. (2003). Anti-inflammatory and immunomodulatory effects of statins. *Kidney International*, 63(1), 12-23.
- Bodduluru, L. N., Kasala, E. R., Madhana, R. M., Barua, C. C., Hussain, M. I., Haloi, P., & Borah, P. (2016). Naringenin ameliorates inflammation and cell proliferation in benzo(a)pyrene induced pulmonary carcinogenesis by modulating CYP1A1, NFkB and PCNA expression. *International Immunopharmacology*, 30, 102-110.
- Bodeker, G., Shekar, S., & Salleh, H. (2009). *Health and beauty from the rainforest: Malaysian traditions of ramuan*: Biotropics Ramuan.
- Boeing, J. S., Barizão, É. O., e Silva, B. C., Montanher, P. F., de Cinque Almeida, V., & Visentainer, J. V. (2014). Evaluation of solvent effect on the extraction of phenolic compounds and antioxidant capacities from the berries: application of principal component analysis. *Chemistry Central Journal*, 8(11), 321-333.
- Boyd, M. R. (1997). The NCI in vitro anticancer drug discovery screen *Anticancer Drug Development Guide* (pp. 23-42): Springer.
- Boyer, R. F., & McCleary, C. J. (1987). Superoxide ion as a primary reductant in ascorbate-mediated ferretin iron release. *Free Radical Biology and Medicine*, 3(6), 389-395.
- Bridgeman, B. B., Wang, P., Ye, B., Pelling, J. C., Volpert, O. V., & Tong, X. (2016). Inhibition of mTOR by apigenin in UVB-irradiated keratinocytes: A new implication of skin cancer prevention. *Cellular Signalling*, 28(5), 460-468.
- Bruneton, J. (2008). *Pharmacognosy: Phytochemistry, Medicinal Plants* (Vol. 6). France: Lavoisier.
- Buckley, M. L., & Ramji, D. P. (2015). The influence of dysfunctional signaling and lipid homeostasis in mediating the inflammatory responses during atherosclerosis. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 1852(7), 1498-1510.
- Bunawan, H., Baharum, S. N., Bunawan, S. N., Amin, N. M., & Noor, N. M. (2014). Cosmos Caudatus Kunth: A Traditional Medicinal Herb. *Global Journal of Pharmacology*, 8(3), 420-426.
- Burkill, I. H. (1966). A dictionary of the economic products of the Malay Peninsula. *A Dictionary of the Economic Products of the Malay Peninsula*, 2(2nd edition).
- Cao, G., & Prior, R. L. (2000). Postprandial increases in serum antioxidant capacity in older women. *Journal of Applied Physiology*, 89(3), 877-883.

- Carlotto, J., da Silva, L. M., Dartora, N., Maria-Ferreira, D., Sabry, D. d. A., Filho, A. P. S., de Paula Werner, M. F., Sassaki, G. L., Gorin, P. A. J., Iacomini, M., Cipriani, T. R., & de Souza, L. M. (2015). Identification of a dicaffeoylquinic acid isomer from *Arctium lappa* with a potent anti-ulcer activity. *Talanta*, 135, 50-57.
- Castellano, G., González-Santander, J. L., Lara, A., & Torrens, F. (2013). Classification of flavonoid compounds by using entropy of information theory. *Phytochemistry*, 93, 182-191.
- Chagas-Paula, D. A., Oliveira, R. B. d., da Silva, V. C., Gobbo-Neto, L., Gasparoto, T. H., Campanelli, A. P., Faccioli, L. H., & Da Costa, F. B. (2011). Chlorogenic acids from *Tithonia diversifolia* demonstrate better anti-inflammatory effect than indomethacin and its sesquiterpene lactones. *Journal of Ethnopharmacology*, 136(2), 355-362.
- Chan, K. T. (2010). The roles of FAK and calpain 2 in regulating adhesion dynamics during cell migration and invasion. (3437194 Ph.D.), The University of Wisconsin - Madison, Ann Arbor.
- Chan, K. W., Iqbal, S., Khong, N. M., Ooi, D.-J., & Ismail, M. (2014). Antioxidant activity of phenolics-saponins rich fraction prepared from defatted kenaf seed meal. *LWT-Food Science and Technology*, 56(1), 181-186.
- Chan, K. W., Khong, N. M., Iqbal, S., & Ismail, M. (2013). Isolation and antioxidative properties of phenolics-saponins rich fraction from defatted rice bran. *Journal of Cereal Science*, 57(3), 480-485.
- Chan, K. W., Khong, N. M., Iqbal, S., Mansor, S. M., & Ismail, M. (2013). Defatted kenaf seed meal (DKSM): Prospective edible flour from agricultural waste with high antioxidant activity. *LWT-Food Science and Technology*, 53(1), 308-313.
- Chang, W.-C., Kuo, P.-L., Chen, C.-W., Wu, J. S.-B., & Shen, S.-C. (2015). Caffeic acid improves memory impairment and brain glucose metabolism via ameliorating cerebral insulin and leptin signalling pathways in high-fat diet-induced hyperinsulinemic rats. *Food Research International*, 77, Part 1, 24-33.
- Chen, A. Y., & Chen, Y. C. (2013). A review of the dietary flavonoid, kaempferol on human health and cancer chemoprevention. *Food Chemistry*, 138(4), 2099-2107.
- Chen, G.-d., Zhu, Y.-Y., Cao, Y., Liu, J., Shi, W.-q., Liu, Z.-m., & Chen, Y.-m. (2015). Association of dietary consumption and serum levels of vitamin A and β-carotene with bone mineral density in Chinese adults. *Bone*, 79, 110-115.

- Cheok, C. Y., Salman, H. A. K., & Sulaiman, R. (2014). Extraction and quantification of saponins: A review. *Food Research International*, 59(0), 16-40.
- Cheung, L. W., Leung, P. C., & Wong, A. S. (2006). Gonadotropin-releasing hormone promotes ovarian cancer cell invasiveness through c-Jun NH₂-terminal kinase-mediated activation of matrix metalloproteinase (MMP)-2 and MMP-9. *Cancer Research*, 66(22), 10902-10910.
- Chittenden, F. (1965). The Royal Horticultural Society dictionary of gardening: a practical and scientific encyclopaedia of horticulture, vol. II, 2nd edn. Clarendon: Oxford.
- Choi, H. J., Song, J. H., & Kwon, D. H. (2012). Quercetin 3-rhamnoside exerts antiinfluenza A virus activity in mice. *Phytotherapy Research*, 26(3), 462-464.
- Chu, H., Tang, Q., Huang, H., Hao, W., & Wei, X. (2016). Grape-seed proanthocyanidins inhibit the lipopolysaccharide-induced inflammatory mediator expression in RAW264.7 macrophages by suppressing MAPK and NF- κ b signal pathways. *Environmental Toxicology and Pharmacology*, 41, 159-166.
- Chukwumah, Y. C., Walker, L. T., Verghese, M., & Ogutu, S. (2009). Effect of frequency and duration of ultrasonication on the extraction efficiency of selected isoflavones and trans-resveratrol from peanuts (*Arachis hypogaea*). *Ultrasonics Sonochemistry*, 16(2), 293-299.
- Chun, J., Kang, M., & Kim, Y. S. (2014). A triterpenoid saponin from *Adenophora triphylla* var. *japonica* suppresses the growth of human gastric cancer cells via regulation of apoptosis and autophagy. *Tumor Biology*, 35(12), 12021-12030.
- Corns, C. M. (2003). Herbal remedies and clinical biochemistry. *Annals of Clinical Biochemistry*, 40, 489-507.
- D'Mello, J. F., Duffus, C. M., & Duffus, J. H. (1991). *Toxic substances in crop plants*: Elsevier.
- Das, A., Raychaudhuri, U., & Chakraborty, R. (2012). Cereal based functional food of Indian subcontinent: a review. *Journal of Food Science and Technology*, 49(6), 665-672.
- Davies, M. J., & Thomas, A. C. (1985). Plaque fissuring--the cause of acute myocardial infarction, sudden ischaemic death, and crescendo angina. *British Heart Journal*, 53(4), 363.

- De Vita, D., Friggeri, L., D'Auria, F. D., Pandolfi, F., Piccoli, F., Panella, S., Palamara, A. T., Simonetti, G., Scipione, L., Di Santo, R., Costi, R., & Tortorella, S. (2014). Activity of caffeic acid derivatives against *Candida albicans* biofilm. *Bioorganic Medicinal Chemistry Letters*, 24(6), 1502-1505.
- Decker, E. A., & Welch, B. (1990). Role of ferritin as a lipid oxidation catalyst in muscle food. *Journal of Agricultural and Food Chemistry*, 38(3), 674-677.
- del Valle, P., García-Armesto, M. R., de Arriaga, D., González-Donquiles, C., Rodríguez-Fernández, P., & Rúa, J. (2016). Antimicrobial activity of kaempferol and resveratrol in binary combinations with parabens or propyl gallate against *Enterococcus faecalis*. *Food Control*, 61, 213-220.
- Devi, K. P., Malar, D. S., Nabavi, S. F., Sureda, A., Xiao, J., Nabavi, S. M., & Daglia, M. (2015). Kaempferol and inflammation: From chemistry to medicine. *Pharmacological Research*, 99, 1-10.
- Devi, K. P., Rajavel, T., Habtemariam, S., Nabavi, S. F., & Nabavi, S. M. (2015). Molecular mechanisms underlying anticancer effects of myricetin. *Life Sciences*, 142, 19-25.
- Dinis, T. C. P., Madeira, V. M. C., & Almeida, L. M. (1994). Action of Phenolic Derivatives (Acetaminophen, Salicylate, and 5-Aminosalicylate) as Inhibitors of Membrane Lipid Peroxidation and as Peroxyl Radical Scavengers. *Archives of Biochemistry and Biophysics*, 315(1), 161-169.
- Dixit, V., Tewari, J., & Obendorf, S. K. (2010). Fungal Growth Inhibition of Regenerated Cellulose Nanofibrous Membranes Containing Quillaja Saponin. *Archives of Environmental Contamination and Toxicology*, 59(3), 417-423.
- Djekic, D., Nicoll, R., Novo, M., & Henein, M. (2015). Metabolomics in Atherosclerosis. *IJC Metabolic & Endocrine*(0), 1-11.
- Doss, H. M., Dey, C., Sudandiradoss, C., & Rasool, M. K. (2016). Targeting inflammatory mediators with ferulic acid, a dietary polyphenol, for the suppression of monosodium urate crystal-induced inflammation in rats. *Life Sciences*, 148, 201-210.
- Dudonne, S., Vitrac, X., Coutiere, P., Woillez, M., & Mérillon, J.-M. (2009). Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *Journal of Agricultural and Food Chemistry*, 57(5), 1768-1774.

- Eid, H. M., Ouchfoun, M., Saleem, A., Guerrero-Analco, J. A., Walshe-Roussel, B., Musallam, L., Rapinski, M., Cuerrier, A., Martineau, L. C., Arnason, J. T., & Haddad, P. S. (2016). A combination of (+)-catechin and (-)-epicatechin underlies the in vitro adipogenic action of Labrador tea (*Rhododendron groenlandicum*), an antidiabetic medicinal plant of the Eastern James Bay Cree pharmacopeia. *Journal of Ethnopharmacology*, 178, 251-257.
- Eliza, J., Daisy, P., Ignacimuthu, S., & Duraipandian, V. (2009). Normoglycemic and hypolipidemic effect of costunolide isolated from *Costus speciosus* (Koen ex. Retz.)Sm. in streptozotocin-induced diabetic rats. *Chemico-Biological Interactions*, 179(2–3), 329-334.
- Fahrioglu, U., Dodurga, Y., Elmas, L., & Seçme, M. (2016). Ferulic acid decreases cell viability and colony formation while inhibiting migration of MIA PaCa-2 human pancreatic cancer cells in vitro. *Gene*, 576(1, Part 3), 476-482.
- Fang, S.-Q., Wang, Y.-T., Wei, J.-X., Shu, Y.-H., Xiao, L., & Lu, X.-M. (2016). Beneficial effects of chlorogenic acid on alcohol-induced damage in PC12 cells. *Biomedicine & Pharmacotherapy*, 79, 254-262.
- Fay, W. P. (2004). Plasminogen activator inhibitor 1, fibrin, and the vascular response to injury. *Trends in Cardiovascular Medicine*, 14(5), 196-202.
- Ferrazzano, G. F., Amato, I., Ingenito, A., Zarrelli, A., Pinto, G., & Pollio, A. (2011). Plant polyphenols and their anti-cariogenic properties: a review. *Molecules*, 16(2), 1486-1507.
- Filipović, M., Marković, Z., Đorović, J., Marković, J. D., Lučić, B., & Amić, D. (2015). QSAR of the free radical scavenging potency of selected hydroxybenzoic acids and simple phenolics. *Comptes Rendus Chimie*, 18(5), 492-498.
- Floegel, A., Kim, D.-O., Chung, S.-J., Koo, S. I., & Chun, O. K. (2011). Comparison of ABTS/DPPH assays to measure antioxidant capacity in popular antioxidant-rich US foods. *Journal of Food Composition and Analysis*, 24(7), 1043-1048.
- Formica, J., & Regelson, W. (1995). Review of the biology of quercetin and related bioflavonoids. *Food and Chemical Toxicology*, 33(12), 1061-1080.
- Foti, M. C., Daquino, C., Mackie, I. D., DiLabio, G. A., & Ingold, K. (2008). Reaction of phenols with the 2, 2-diphenyl-1-picrylhydrazyl radical. Kinetics and DFT calculations applied to determine ArO-H bond dissociation enthalpies and reaction mechanism. *The Journal of Organic Chemistry*, 73(23), 9270-9282.

- Fuzzati, N., Dyatmiko, W., Rahman, A., & Hostettmann, K. (1995). Phenylpropane derivatives from roots of *Cosmos caudatus*. *Phytochemistry*, 39(2), 409-412.
- Gabay, O., Sanchez, C., Salvat, C., Chevy, F., Breton, M., Nourissat, G., Wolf, C., Jacques, C., & Berenbaum, F. (2010). Stigmasterol: a phytosterol with potential anti-osteoarthritic properties. *Osteoarthritis and Cartilage*, 18(1), 106-116.
- Galli, R. L., SHUKITT-HALE, B., Youdim, K. A., & Joseph, J. A. (2002). Fruit polyphenolics and brain aging. *Annals of the New York Academy of Sciences*, 959(1), 128-132.
- Gao, Y.-T., Zhang, W.-B., Yang, L.-R., Wang, X.-M., & Yang, Y.-L. (2008). Effects of *Panax notoginseng* saponins on antioxidation and preventing DNA damage caused by hydroxyl radical. *Journal of Chinese Medicinal Materials*, 31(9), 1399-1402.
- Gao, Y., Wang, X., & He, C. (2016). An isoflavanoid-enriched extract from *Pueraria lobata* (kudzu) root protects human umbilical vein endothelial cells against oxidative stress induced apoptosis. *Journal of Ethnopharmacology*, 193, 524-530.
- Garbisa, S., Biggin, S., Cavallarin, N., Sartor, L., Benelli, R., & Albini, A. (1999). Tumor invasion: molecular shears blunted by green tea. *Nature Medicine*, 5(11), 1216-1216.
- Garcia-Salas, P., Morales-Soto, A., Segura-Carretero, A., & Fernández-Gutiérrez, A. (2010). Phenolic-compound-extraction systems for fruit and vegetable samples. *Molecules*, 15(12), 8813-8826.
- Geng, Y.-J., & Libby, P. (2002). Progression of atheroma a struggle between death and procreation. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 22(9), 1370-1380.
- Gifkins, D., Olson, S. H., Demissie, K., Lu, S.-e., Kong, A.-n. T., & Bandera, E. V. (2012). Total and individual antioxidant intake and endometrial cancer risk: results from a population-based case-control study in New Jersey. *Cancer Causes & Control*, 23(6), 887-895.
- Goh, S. H., Soepadmo, E., & Chuah, C. H. (1993). *Phytochemical guide to Malaysian flora*: Institute of Advanced Studies, University of Malaya.
- Gong, Y.-S., Guo, J., Hu, K., Gao, Y.-Q., Xie, B.-J., Sun, Z.-D., Yang, E.-N., & Hou, F.-L. (2016). Ameliorative effect of lotus seedpod proanthocyanidins on cognitive impairment and brain aging induced by d-galactose. *Experimental Gerontology*, 74, 21-28.

- Gopiesh Khanna, V., & Kannabiran, K. (2008). Antimicrobial activity of saponin fractions of the leaves of *Gymnema sylvestre* and *Eclipta prostrata*. *World Journal of Microbiology and Biotechnology*, 24(11), 2737-2740.
- Griep, L. M. O., Geleijnse, J. M., Kromhout, D., Ocké, M. C., & Verschuren, W. M. (2010). Raw and processed fruit and vegetable consumption and 10-year coronary heart disease incidence in a population-based cohort study in the Netherlands. *European Journal of Clinical Nutrition*, 5(10), 136-150.
- Guardia, T., Rotelli, A. E., Juarez, A. O., & Pelzer, L. E. (2001). Anti-inflammatory properties of plant flavonoids. Effects of rutin, quercetin and hesperidin on adjuvant arthritis in rat. *Farmaco*, 56(9), 683-687.
- Güçlü-Üstündağ, Ö., & Mazza, G. (2007). Saponins: properties, applications and processing. *Critical Reviews in Food Science and Nutrition*, 47(3), 231-258.
- Halliwell, B. (1994). Free radicals, antioxidants, and human disease: curiosity, cause, or consequence? *The Lancet*, 344(8924), 721-724.
- Halliwell, B. (2006). Reactive species and antioxidants. Redox biology is a fundamental theme of aerobic life. *Plant Physiology*, 141(2), 312-322.
- Halliwell, B. (2011). Free radicals and antioxidants – quo vadis? *Trends in Pharmacological Sciences*, 32(3), 125-130.
- Halliwell, B. (2012). Free radicals and antioxidants: updating a personal view. *Nutrition Reviews*, 70(5), 257-265.
- Halliwell, B., & Gutteridge, J. M. (2006). *Free radicals in biology and medicine* (Vol. 15): Oxford University press.
- Han, X., Shen, T., & Lou, H. (2007). Dietary polyphenols and their biological significance. *International Journal of Molecular Sciences*, 8(9), 950-988.
- Hanelt, P. (2001). *Mansfeld's encyclopedia of agricultural and horticultural crops*: Springer Science & Business Media.
- Hao, S., Xu, R., Li, D., Zhu, Z., Wang, T., & Liu, K. (2015). Attenuation of Streptozotocin-Induced Lipid Profile Anomalies in the Heart, Brain, and mRNA Expression of HMG-CoA Reductase by Diosgenin in Rats. *Cell Biochemistry and Biophysics*, 72(3), 741-749.
- Hassan, W. W., & Mahmood, M. (2007). *Healing herbs of Malaysia*: Biotropics Malaysia Berhad.

- Haytowitz, D. B., & Bhagwat, S. (2010). USDA database for the oxygen radical absorbance capacity (ORAC) of selected foods, Release 2. US Department of Agriculture.
- Hazwani, Z. (2012). Optimization of extraction parameters of total phenolic compound from *Cosmos caudatus*. Universiti Malaysia Pahang.
- Heinrich, M. (2014). Ethnopharmacology: quo vadis? Challenges for the future. *Revista Brasileira de Farmacognosia*, 24(2), 99-102.
- Helaly, F., Soliman, H., Soheir, A., & Ahmed, A. (2001). Controlled release of migration of molluscicidal saponin from different types of polymers containing *Calendula officinalis*. *Advances in Polymer Technology*, 20(4), 305-311.
- Herrmann, F., & Wink, M. (2011). Synergistic interactions of saponins and monoterpenes in HeLa cells, Cos7 cells and in erythrocytes. *Phytomedicine*, 18(13), 1191-1196.
- Hiai, S., Oura, H., & Nakajima, T. (1976). Color reaction of some sapogenins and saponins with vanillin and sulfuric acid. *Planta Medica*(29), 116-122.
- Higuchi, K., Saito, I., Maruyama, K., Eguchi, E., Mori, H., Tanno, S., Sakurai, S., Kishida, T., Nishida, W., Osawa, H., & Tanigawa, T. (2015). Associations of serum β -carotene and retinol concentrations with insulin resistance: The Toon Health Study. *Nutrition*, 31(7–8), 975-980.
- Hong, Q., Ma, Z.-C., Huang, H., Wang, Y.-G., Tan, H.-L., Xiao, C.-R., Liang, Q.-D., Zhang, H.-T., & Gao, Y. (2016). Antithrombotic activities of ferulic acid via intracellular cyclic nucleotide signaling. *European Journal of Pharmacology*.
- Hovland, A., Jonasson, L., Garred, P., Yndestad, A., Aukrust, P., Lappegård, K. T., Espenvik, T., & Mollnes, T. E. (2015). The complement system and toll-like receptors as integrated players in the pathophysiology of atherosclerosis. *Atherosclerosis*, 241(2), 480-494.
- Huang, D., Ou, B., & Prior, R. L. (2005). The chemistry behind antioxidant capacity assays. *Journal of Agricultural and Food Chemistry*, 53(6), 1841-1856.
- Huang, L.-I., Pan, C., Wang, L., Ding, L., Guo, K., Wang, H.-z., Xu, A. m., & Gao, S. (2015). Protective effects of grape seed proanthocyanidins on cardiovascular remodeling in DOCA-salt hypertension rats. *The Journal of Nutritional Biochemistry*, 26(8), 841-849.

- Huang, Q., Gao, B., Wang, L., Hu, Y.-Q., Lu, W.-G., Yang, L., Luo, Z.-J., & Liu, J. (2014). Protective effects of myricitrin against osteoporosis via reducing reactive oxygen species and bone-resorbing cytokines. *Toxicology and Applied Pharmacology*, 280(3), 550-560.
- Huda-Faujan, N., Noriham, A., Norrakiah, A., & Babji, A. (2009). Antioxidant activity of plants methanolic extracts containing phenolic compounds. *African Journal of Biotechnology*, 8(3), 547-555.
- Imam, M. U., Ismail, M., Ooi, D. J., Azmi, N. H., Sarega, N., Chan, K. W., & Bhanger, M. I. (2015). Are bioactive-rich fractions functionally richer? *Critical Reviews in Biotechnology*(0), 1-9.
- Ionita, M. G., Arslan, F., De Kleijn, D. P., & Pasterkamp, G. (2010). Endogenous inflammatory molecules engage Toll-like receptors in cardiovascular disease. *Journal of Innate Immunity*, 2(4), 307-315.
- Jakopič, J., Veberič, R., & Štampar, F. (2009). Extraction of phenolic compounds from green walnut fruits in different solvents. *Acta Agriculturae Slovenica*, 93(1), 11-15.
- Javadi, N., Abas, F., Hamid, A. A., Simoh, S., Shaari, K., Ismail, I. S., Medianı, A., & Khatib, A. (2014). GC-MS-Based Metabolite Profiling of Cosmos caudatus Leaves Possessing Alpha-Glucosidase Inhibitory Activity. *Journal of Food Science*, 79(6), C1130-C1136.
- Jeong, Y. M., Ha, J. H., & Park, S. N. (2016). Cytoprotective effects against UVA and physical properties of luteolin-loaded cationic solid lipid nanoparticle. *Journal of Industrial and Engineering Chemistry*, 35, 54-62.
- Jiang, W., Cen, Y., Song, Y., Li, P., Qin, R., Liu, C., Zhao, Y., Zheng, J., & Zhou, H. (2016). Artesunate attenuated progression of atherosclerosis lesion formation alone or combined with rosuvastatin through inhibition of pro-inflammatory cytokines and pro-inflammatory chemokines. *Phytomedicine*, 23(11), 1259-1266.
- Jin, U.-H., Lee, J.-Y., Kang, S.-K., Kim, J.-K., Park, W.-H., Kim, J.-G., Moon, S.-K., & Kim, C.-H. (2005). A phenolic compound, 5-caffeoylequinic acid (chlorogenic acid), is a new type and strong matrix metalloproteinase-9 inhibitor: Isolation and identification from methanol extract of Euonymus alatus. *Life Sciences*, 77(22), 2760-2769.
- Joung, D.-K., Lee, Y.-S., Han, S.-H., Lee, S.-W., Cha, S.-W., Mun, S.-H., Kong, R., Kang, O.-H., Song, H.-J., Shin, D.-W., & Kwon, D.-Y. (2016). Potentiating activity of luteolin on membrane permeabilizing agent and ATPase inhibitor against methicillin-resistant *Staphylococcus aureus*. *Asian Pacific Journal of Tropical Medicine*, 9(1), 19-22.

- Jovanovski, E., Jenkins, A., Dias, A. G., Peeva, V., Sievenpiper, J., Arnason, J. T., Rahelic, D., Josse, R. G., & Vuksan, V. (2010). Effects of Korean Red Ginseng (*Panax ginseng* C.A. Mayer) and Its Isolated Ginsenosides and Polysaccharides on Arterial Stiffness in Healthy Individuals. *American Journal of Hypertension*, 23(5), 469-472.
- Juturu, V. (2015). Chapter 4 - Lutein, Brain, and Neurological Functions A2 - Preedy, Ronald Ross Watson Victor R *Bioactive Nutraceuticals and Dietary Supplements in Neurological and Brain Disease* (pp. 41-47). San Diego: Academic Press.
- Kang, J. S., Yoon, Y. D., Lee, K. H., Park, S.-K., & Kim, H. M. (2004). Costunolide inhibits interleukin-1 β expression by down-regulation of AP-1 and MAPK activity in LPS-stimulated RAW 264.7 cells. *Biochemical and Biophysical Research Communications*, 313(1), 171-177.
- Kataja-tuomola, M. K., Kontto, J. P., Männistö, S., Albanes, D., & Virtamo, J. (2011). Intake of antioxidants and risk of type 2 diabetes in a cohort of male smokers. *European Journal of Clinical Nutrition*, 65(5), 590-597.
- Kaur, C., & Kapoor, H. C. (2002). Anti-oxidant activity and total phenolic content of some Asian vegetables. *International Journal of Food Science & Technology*, 37(2), 153-161.
- Kerem, Z., German-Shashoua, H., & Yarden, O. (2005). Microwave-assisted extraction of bioactive saponins from chickpea (*Cicer arietinum* L.). *Journal of the Science of Food and Agriculture*, 85(3), 406-412.
- Khedgikar, V., Kushwaha, P., Gautam, J., Sharma, S., Verma, A., Choudhary, D., Mishra, P. R., & Trivedi, R. (2016). Kaempferol targets Krt-14 and induces cytoskeletal mineralization in osteoblasts: A mechanistic approach. *Life Sciences* 21(12), 145-159.
- Khokhar, S., & Apenten, R. K. O. (2003). Iron binding characteristics of phenolic compounds: some tentative structure-activity relations. *Food Chemistry*, 81(1), 133-140.
- Kim-Park, W. K., Allam, E. S., Palasuk, J., Kowolik, M., Park, K. K., & Windsor, L. J. (2016). Green tea catechin inhibits the activity and neutrophil release of Matrix Metalloproteinase-9. *Journal of Traditional and Complementary Medicine* 4(3), 456-465.
- Kim, M., Choi, S.-Y., Lee, P., & Hur, J. (2015). Neochlorogenic acid inhibits lipopolysaccharide-induced activation and pro-inflammatory responses in BV2 microglial cells. *Neurochemical Research*, 40(9), 1792-1798.
- Kim, N.-R., Kim, H.-Y., Kim, M.-H., Kim, H.-M., & Jeong, H.-J. (2016). Improvement of depressive behavior by Sweetme Sweet Pumpkin™ and its active compound, β -carotene. *Life Sciences*, 147, 39-45.

- Kim, Y., Narayanan, S., & Chang, K.-O. (2010). Inhibition of influenza virus replication by plant-derived isoquercetin. *Antiviral Research*, 88(2), 227-235.
- Kirichenko, T. V., Sobenin, I. A., Nikolic, D., Rizzo, M., & Orekhov, A. N. (2016). Anti-cytokine therapy for prevention of atherosclerosis. *Phytomedicine*, 23(11), 1198-1210.
- Kite, G. C., Porter, E. A., & Simmonds, M. S. J. (2007). Chromatographic behaviour of steroid saponins studied by high-performance liquid chromatography-mass spectrometry. *Journal of Chromatography A*, 1148(2), 177-183.
- Kraus, S., Naumov, I., Shapira, S., Kazanov, D., Aroch, I., Afek, A., Eisenberg, O., George, J., Arber, N., & Finkelstein, A. (2014). Aspirin but not meloxicam attenuates early atherosclerosis in apolipoprotein E knockout mice. *The Israel Medical Association Journal: IMAJ*, 16(4), 233-238.
- Kulip, J., Fan, L. N., Manshoor, N., Julius, A., Said, I. M., Gisil, J., Joseph, J. A., & Tukin, W. F. (2010). Medicinal plants in Maliau Basin, Sabah, Malaysia. *Journal of Tropical Biology and Conservation*, 6, 21-33.
- Lapidot, T., Walker, M. D., & Kanner, J. (2002). Can apple antioxidants inhibit tumor cell proliferation? Generation of H₂O₂ during interaction of phenolic compounds with cell culture media. *Journal of Agricultural and Food Chemistry*, 50(11), 3156-3160.
- Lee, J., Lim, S., Kang, S.-M., Min, S., Son, K., Lee, H. S., Park, E. M., Ngo, H. T. T., Tran, H. T. L., Lim, Y.-S., & Hwang, S. B. (2012). Saponin Inhibits Hepatitis C Virus Propagation by Up-regulating Suppressor of Cytokine Signaling 2. *PLoS One*, 7(6), 1245-1255.
- Lee, T. K., & Vairappan, C. S. (2011). Antioxidant, antibacterial and cytotoxic activities of essential oils and ethanol extracts of selected South East Asian herbs. *Journal of Medicinal Plant Research*, 5(1), 5284-5290.
- Lee, Y. S., & Choi, E. M. (2011). Costunolide stimulates the function of osteoblastic MC3T3-E1 cells. *International Immunopharmacology*, 11(6), 712-718.
- leyman, H. S., Mshvildadze, V., Gepdiremen, A., & Elias, R. (2003). Acute and chronic antiinflammatory profile of the ivy plant, Hedera helix, in rats. *Phytomedicine*, 10(5), 370-374.
- Li, H., Horke, S., & Förstermann, U. (2014). Vascular oxidative stress, nitric oxide and atherosclerosis. *Atherosclerosis*, 237(1), 208-219.

- Lin, W.-L., Liang, W.-H., Lee, Y.-J., Chuang, S.-K., & Tseng, T.-H. (2010). Antitumor progression potential of caffeic acid phenethyl ester involving p75NTR in C6 glioma cells. *Chemico-Biological Interactions*, 188(3), 607-615.
- Liu, F. F., Wu, X., Zhang, Y., Wang, Y., & Jiang, F. (2014). TRAIL/DR5 Signaling Promotes Macrophage Foam Cell Formation by Modulating Scavenger Receptor Expression. *PLoS One*, 9(1), 870-879.
- Liu, J., & Henkel, T. (2002). Traditional Chinese medicine (TCM): are polyphenols and saponins the key ingredients triggering biological activities? *Current Medicinal Chemistry*, 9(15), 1483-1485.
- Liu, J., Lu, J.-f., Wen, X.-y., Kan, J., & Jin, C.-h. (2015). Antioxidant and protective effect of inulin and catechin grafted inulin against CCl₄-induced liver injury. *International Journal of Biological Macromolecules*, 72, 1479-1484.
- Liu, R., Hu, Y., Li, J., & Lin, Z. (2007). Production of soybean isoflavone genistein in non-legume plants via genetically modified secondary metabolism pathway. *Metabolic Engineering*, 9(1), 1-7.
- Lo, H.-M., Tsai, Y.-J., Du, W.-Y., Tsou, C.-J., & Wu, W.-B. (2012). A naturally occurring carotenoid, lutein, reduces PDGF and H₂O₂ signaling and compromised migration in cultured vascular smooth muscle cells. *Journal of Biomedical Science*, 19, 18-28.
- Loh, S. P., & Hadira, O. (2011). In vitro inhibitory potential of selected Malaysian plants against key enzymes involved in hyperglycemia and hypertension. *Malaysian Journal of Nutrition*, 17(1), 77-86.
- Long, L. H., Clement, M. V., & Halliwell, B. (2000). Artifacts in Cell Culture: Rapid Generation of Hydrogen Peroxide on Addition of (-)-Epigallocatechin, (-)-Epigallocatechin Gallate, (+)-Catechin, and Quercetin to Commonly Used Cell Culture Media. *Biochemical and Biophysical Research Communications*, 273(1), 50-53.
- Long, L. H., Hoi, A., & Halliwell, B. (2010). Instability of, and generation of hydrogen peroxide by, phenolic compounds in cell culture media. *Archives of Biochemistry and Biophysics*, 501(1), 162-169.
- Lotito, S. B., & Frei, B. (2006). Consumption of flavonoid-rich foods and increased plasma antioxidant capacity in humans: Cause, consequence, or epiphenomenon? *Free Radical Biology and Medicine*, 41(12), 1727-1746.
- Louis, S. F., & Zahradka, P. (2010). Vascular smooth muscle cell motility: From migration to invasion. *Experimental & Clinical Cardiology*, 15(4), e75-e85.

- Ma, X. H., Zheng, C. J., Han, L. Y., Xie, B., Jia, J., Cao, Z. W., Li, Y. X., & Chen, Y. Z. (2009). Synergistic therapeutic actions of herbal ingredients and their mechanisms from molecular interaction and network perspectives. *Drug Discovery Today*, 14(11–12), 579-588.
- Malaviya, T., & Sharma, P. K. (2014). Phytochemical, Pharmacological Profile and Commercial Utility of Tropically Distributed Plant *Bauhinia variegata*. *Global Journal of Pharmacology*, 8(2), 196-205.
- Mancuso, C., & Santangelo, R. (2014). Ferulic acid: Pharmacological and toxicological aspects. *Food and Chemical Toxicology*, 65(0), 185-195.
- Mandel, S., & Youdim, M. B. H. (2004). Catechin polyphenols: neurodegeneration and neuroprotection in neurodegenerative diseases. *Free Radical Biology and Medicine*, 37(3), 304-317.
- Marín-García, J. (2014). Chapter 6 - Molecular Determinants of Atherosclerosis. In J. Marín-García (Ed.), *Post-Genomic Cardiology (Second Edition)* (pp. 183-215). Boston: Academic Press.
- Marinelli, J. (2005). Herbaceous Plants. In J. Marinelli (Ed.), *Plant*. UK: DK.
- Martin, A., & Clynes, M. (1993). Comparison of 5 microplate colorimetric assays for in vitro cytotoxicity testing and cell proliferation assays. *Cytotechnology*, 11(1), 49-58.
- Maxwell, C. A., Restrepo-Hartwig, M. A., Hession, A. O., & McGonigle, B. (2004). Chapter eight Metabolic engineering of soybean for improved flavor and health benefits. In T. R. John (Ed.), *Recent Advances in Phytochemistry* (Vol. 38, pp. 153-176): Elsevier.
- McLaren, J. E., Michael, D. R., Ashlin, T. G., & Ramji, D. P. (2011). Cytokines, macrophage lipid metabolism and foam cells: Implications for cardiovascular disease therapy. *Progress in Lipid Research*, 50(4), 331-347.
- Mediani, A. (2012). Influence of growth stages and harvesting season, and drying methods on phytochemical content and antioxidant activity of *Cosmos Caudatus Kunth* leaves. (Master of Science), Univirsiti Putra Malaysia, Malaysia.
- Mediani, A., Abas, F., Khatib, A., Maulidiani, H., Shaari, K., Choi, Y. H., & Lajis, N. (2012). 1 H-NMR-based metabolomics approach to understanding the drying effects on the phytochemicals in *Cosmos caudatus*. *Food Research International*, 49(2), 763-770.
- Mediani, A., Abas, F., Khatib, A., & Tan, C. P. (2013). *Cosmos caudatus* as a potential source of polyphenolic compounds: Optimisation of oven drying conditions and characterisation of its functional properties. *Molecules*, 18(9), 10452-10464.

- Mediani, A., Abas, F., Ping, T. C., Khatib, A., & Lajis, N. H. (2012). Influence of growth stage and season on the antioxidant constituents of *Cosmos caudatus*. *Plant foods for human nutrition*, 67(4), 344-350.
- Menendez, C., Jimenez, R., Moreno, L., Galindo, P., Cogolludo, A., Duarte, J., & Perez-Vizcaino, F. (2011). Lack of synergistic interaction between quercetin and catechin in systemic and pulmonary vascular smooth muscle. *The British Journal of Nutrition*, 105(9), 1287-1293.
- Michikawa, T. M. D., Ishida, S. M. D., Nishiwaki, Y. M. D., Kikuchi, Y. P., Tsuboi, T. M., Hosoda, K. B., Ishigami, A. P., Iwasawa, S. M. D., Nakano, M. M. D., & Takebayashi, T. M. D. (2009). Serum antioxidants and age-related macular degeneration among older Japanese. *Asia Pacific Journal of Clinical Nutrition*, 18(1), 1-7.
- Miean, K. H., & Mohamed, S. (2001). Flavonoid (myricetin, quercetin, kaempferol, luteolin, and apigenin) content of edible tropical plants. *Journal of agricultural and food chemistry*, 49(6), 3106-3112.
- Mika, M., Kostogrys, R. B., Franczyk-Żarów, M., Wikiera, A., & Maślak, E. (2015). Anti-atherosclerotic activity of catechins depends on their stereoisomerism. *Atherosclerosis*, 240(1), 125-130.
- Mishra, K., Ojha, H., & Chaudhury, N. K. (2012). Estimation of antiradical properties of antioxidants using DPPH assay: A critical review and results. *Food Chemistry*, 130(4), 1036-1043.
- Mohamed, N., Gwee Sian Khee, S., Shuid, A. N., Muhammad, N., Suhaimi, F., Othman, F., Babji, A. S., & Soelaiman, I.-N. (2012). The effects of *Cosmos caudatus* on structural bone histomorphometry in ovariectomized rats. *Evidence-Based Complementary and Alternative Medicine*, 2012.
- Mohamed, N., Sahhugi, Z., Ramli, E. S. M., & Muhammad, N. (2013). The effects of *Cosmos caudatus* (ulam raja) on dynamic and cellular bone histomorphometry in ovariectomized rats. *BMC Research Notes*, 6(1), 239.
- Müller, P., & Downard, K. M. (2015). Catechin inhibition of influenza neuraminidase and its molecular basis with mass spectrometry. *Journal of Pharmaceutical and Biomedical Analysis*, 111, 222-230.
- Musa, N. L. W., Zain, W. Z. W. M., Kassim, J., & Karim, S. A. (2011). Preliminary studies on phytochemical screening of ulam and fruit from malaysian. *Journal of Chemistry*, 8(S1), S285-S288.
- Mustafa, R., Hamid, A. A., Mohamed, S., & Bakar, F. A. (2010). Total phenolic compounds, flavonoids, and radical scavenging activity of 21 selected tropical plants. *Journal of food science*, 75(1), C28-C35.

- Naczk, M., & Shahidi, F. (2004). Extraction and analysis of phenolics in food. *Journal of Chromatography A*, 1054(1–2), 95-111.
- Naczk, M., & Shahidi, F. (2006). Phenolics in cereals, fruits and vegetables: occurrence, extraction and analysis. *Journal of Pharmaceutical and Biomedical Analysis*, 41(5), 1523-1542.
- Narasimhan, A., Chinnaiyan, M., & Karundevi, B. (2015). Ferulic acid regulates hepatic GLUT2 gene expression in high fat and fructose-induced type-2 diabetic adult male rat. *European Journal of Pharmacology*, 761, 391-397.
- Nardini, M., D'Aquino, M., Tomassi, G., Gentili, V., Di Felice, M., & Scaccini, C. (1995). Inhibition of human low-density lipoprotein oxidation by caffeic acid and other hydroxycinnamic acid derivatives. *Free Radical Biology and Medicine*, 19(5), 541-552.
- Nasr Bouzaiene, N., Kilani Jaziri, S., Kovacic, H., Chekir-Ghedira, L., Ghedira, K., & Luis, J. (2015). The effects of caffeic, coumaric and ferulic acids on proliferation, superoxide production, adhesion and migration of human tumor cells in vitro. *European Journal of Pharmacology*, 766, 99-105.
- Neubig, R. R., Spedding, M., Kenakin, T., & Christopoulos, A. (2003). International Union of Pharmacology Committee on Receptor Nomenclature and Drug Classification. XXXVIII. Update on terms and symbols in quantitative pharmacology. *Pharmacological Reviews*, 55(4), 597-606.
- Nichenametla, S. N., Taruscio, T. G., Barney, D. L., & Exon, J. H. (2006). A review of the effects and mechanisms of polyphenolics in cancer. *Critical Reviews in Food Science and Nutrition*, 46(2), 161-183.
- Noda, Y., Kaneyuki, T., Mori, A., & Packer, L. (2002). Antioxidant activities of pomegranate fruit extract and its anthocyanidins: delphinidin, cyanidin, and pelargonidin. *Journal of agricultural and food chemistry*, 50(1), 166-171.
- Norouzi, L., Shirpoor, A., Khadem Ansari, M.-H., & Ilkhanizadeh, B. (2015). Vitamin E attenuates alcohol-induced aortic wall damage in rats. *Artery Research*, 10, 20-26.
- Nowakowska, Z. (2007). A review of anti-infective and anti-inflammatory chalcones. *European Journal of Medicinal Chemistry*, 42(2), 125-137.
- Nwachukwu, I. D., Udenigwe, C. C., & Aluko, R. E. (2016). Lutein and zeaxanthin: Production technology, bioavailability, mechanisms of action, visual function, and health claim status. *Trends in Food Science & Technology*, 49, 74-84.

- Ohuchi, M., Kaku, T., & Matsuyama, T. (1996). Characterization of Human Osteosarcoma Cell Lines Established from Primary and Secondary Lesions with Prominent Osteoid Formation. *Oral Medicine & Pathology*, 1(1), 11-22.
- Oleszek, W., & Hamed, A. (2010). Saponin-based surfactants. *Surfactants from Renewable Resources*, 239.
- Oleszek, W., & Marston, A. (2000). *Saponins in food, feedstuffs and medicinal plants* (Vol. 45): Kluwer Academic Publisher.
- Orekhov, A. N., & Ivanova, E. A. (2016). Cellular models of atherosclerosis and their implication for testing natural substances with anti-atherosclerotic potential. *Phytomedicine*, 23(11), 1190-1197.
- Oroian, M., & Escriche, I. (2015). Antioxidants: Characterization, natural sources, extraction and analysis. *Food Research International*, 74(0), 10-36.
- Palozza, P., Parrone, N., Simone, R. E., & Catalano, A. (2010). Lycopene in atherosclerosis prevention: An integrated scheme of the potential mechanisms of action from cell culture studies. *Archives of Biochemistry and Biophysics*, 504(1), 26-33.
- Pan, S. Y., Zhou, J., Gibbons, L., Morrison, H., & Wen, S. W. (2011). Antioxidants and breast cancer risk- a population-based case-control study in Canada. *BMC Cancer*, 11, 372.
- Panda, S., Jafri, M., Kar, A., & Meheta, B. K. (2009). Thyroid inhibitory, antiperoxidative and hypoglycemic effects of stigmasterol isolated from *Butea monosperma*. *Fitoterapia*, 80(2), 123-126.
- Pang, C., Zheng, Z., Shi, L., Sheng, Y., Wei, H., Wang, Z., & Ji, L. (2016). Caffeic acid prevents acetaminophen-induced liver injury by activating the Keap1-Nrf2 antioxidative defense system. *Free Radical Biology and Medicine*, 91, 236-246.
- Pannala, A., Razaq, R., Halliwell, B., Singh, S., & Rice-Evans, C. A. (1998). Inhibition of Peroxynitrite Dependent Tyrosine Nitration by Hydroxycinnamates: Nitration or Electron Donation? *Free Radical Biology and Medicine*, 24(4), 594-606.
- Park, S. J., Kim, D. H., Jung, J. M., Kim, J. M., Cai, M., Liu, X., Hong, J. G., Lee, C. H., Lee, K. R., & Ryu, J. H. (2012). The ameliorating effects of stigmasterol on scopolamine-induced memory impairments in mice. *European Journal of Pharmacology*, 676(1–3), 64-70.

- Peng, B. J., Zhu, Q., Zhong, Y. L., Xu, S. H., & Wang, Z. (2015). Chlorogenic Acid Maintains Glucose Homeostasis through Modulating the Expression of SGLT-1, GLUT-2, and PLG in Different Intestinal Segments of Sprague-Dawley Rats Fed a High-Fat Diet. *Biomedical and Environmental Sciences*, 28(12), 894-903.
- Perumal, V., Hamid, A. A., Ismail, A., Saari, K., Abas, F., Ismail, I. S., Lajis, N. H., & Khatib, A. (2014). Effect of cosmos caudatus kunth leaves on the lipid profile of a hyperlipidemia-induced animal model. *Journal of Food Chemistry and Nutrition*, 2(1), 43-51.
- Pietta, P.-G. (2000). Flavonoids as antioxidants. *Journal of natural products*, 63(7), 1035-1042.
- Pietta, P., Mauri, P., Simonetti, P., & Testolin, G. (1995). HPLC and MEKC determination of major flavonoids in selected food pools. *Fresenius' Journal of Analytical Chemistry*, 352(7-8), 788-792.
- Pinent, M., González-Abuín, N., Blay, M., & Ardévol, A. (2016). Chapter 16 - Dietary Proanthocyanidin Modulation of Pancreatic β Cells: Molecular Aspects A2 - Mauricio, Didac *Molecular Nutrition and Diabetes* (pp. 197-210). San Diego: Academic Press.
- Pisoschi, A. M., & Pop, A. (2015). The role of antioxidants in the chemistry of oxidative stress: A review. *European Journal of Medicinal Chemistry*, 97(0), 55-74.
- Poljsak, B., Šuput, D., & Milisav, I. (2013). Achieving the balance between ROS and antioxidants: when to use the synthetic antioxidants. *Oxidative Medicine and Cellular Longevity*, 2013, 214-227.
- Ponnuswamy, P., Schröttle, A., Ostermeier, E., Grüner, S., Huang, P. L., Ertl, G., Hoffmann, U., Nieswandt, B., & Kuhlencordt, P. J. (2012). eNOS Protects from Atherosclerosis Despite Relevant Superoxide Production by the Enzyme in apoE-/- Mice. *PLoS One*, 7(1), 478-492.
- Pradhananga, S., & Shim, W.-S. (2015). Caffeic acid exhibits anti-pruritic effects by inhibition of multiple itch transmission pathways in mice. *European Journal of Pharmacology*, 762, 313-321.
- Quattrocchi, U. (1999). *CRC World Dictionary of Plant Names: Common Names, Scientific Names, Eponyms, Synonyms, and Etymology* (Vol. 3): CRC Press.
- Quattrocchi, U. (2012). *CRC World Dictionary of Medicinal and Poisonous Plants: Common Names, Scientific Names, Eponyms, Synonyms, and Etymology* (5 Volume Set): CRC Press.

- Ragasa, C., Nacpil, Z., Penalosa, B., Coll, J., & Rideout, J. (1997). Antimutagen and antifungal compounds from *Cosmos caudatus*. *Philippine Journal of Science (Philippines)*, 115(5), 215-301.
- Raines, E. W., & Ferri, N. (2005). Thematic review series: The immune system and atherogenesis. Cytokines affecting endothelial and smooth muscle cells in vascular disease. *Journal of Lipid Research*, 46(6), 1081-1092.
- Rajalingam, K., Sivakumar, K., & Suresh, K. (2012). Modulatory effects of diosgenin and farnesol on cell surface integrity during 7,12-dimethylbenz(a) anthracene induced oral carcinogenesis. *Journal of Cell and Tissue Research*, 12(1), 2991-2997.
- Raju, J., & Bird, R. P. (2007). Diosgenin, a naturally occurring furostanol saponin suppresses 3-hydroxy-3-methylglutaryl CoA reductase expression and induces apoptosis in HCT-116 human colon carcinoma cells. *Cancer Letters*, 255(2), 194-204.
- Ramasamy, S., Wahab, N. A., Abidin, N. Z., Manickam, S., & Zakaria, Z. (2012). Growth inhibition of human gynecologic and colon cancer cells by *Phyllanthus watsonii* through apoptosis induction. *PLoS One*, 7(4), 347-353.
- Ramsay, A., Williams, A. R., Thamsborg, S. M., & Mueller-Harvey, I. (2016). Galloylated proanthocyanidins from shea (*Vitellaria paradoxa*) meal have potent anthelmintic activity against *Ascaris suum*. *Phytochemistry*, 122, 146-153.
- Rasdi, M., Hafipah, N., Othman, A. S., Sule, A., & Ahmed, Q. U. (2010). Antimicrobial studies of *cosmos caudatus* kunth. *Journal of Medicinal Plants Research*, 4(8), 669-673.
- Rasoanaivo, P., Wright, C. W., Willcox, M. L., & Gilbert, B. (2011). Whole plant extracts versus single compounds for the treatment of malaria: synergy and positive interactions. *Malaria Journal*, 10(Suppl 1), S4.
- Reiner, Ž. (2014). Resistance and intolerance to statins. *Nutrition, Metabolism and Cardiovascular Diseases*, 24(10), 1057-1066.
- Ren, B., Qin, W., Wu, F., Wang, S., Pan, C., Wang, L., Zeng, B., Ma, S., & Liang, J. (2016). Apigenin and naringenin regulate glucose and lipid metabolism, and ameliorate vascular dysfunction in type 2 diabetic rats. *European Journal of Pharmacology*, 773, 13-23.
- Rice-Evans, C., Miller, N., & Paganga, G. (1997). Antioxidant properties of phenolic compounds. *Trends in Plant Science*, 2(4), 152-159.
- Rice-Evans, C. A., Miller, N. J., & Paganga, G. (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology and Medicine*, 20(7), 933-956.

- Ristow, M., Zarse, K., Oberbach, A., Klöting, N., Birringer, M., Kiehntopf, M., Stumvoll, M., Kahn, C. R., & Blüher, M. (2009). Antioxidants prevent health-promoting effects of physical exercise in humans. *Proceedings of the National Academy of Sciences*, 106(21), 8665-8670.
- Rohit, S., Gulab, T. S., Bhagwan, S. S., Mukeshwar, P., & Prakash, B. S. (2012). Saponin: A Wonder Drug from Chlorophytum Species. *Global Journal of Research on Medicinal Plants & Indigenous Medicine*, 1(10), 503-515.
- Rufus, P. G., Mohamed, N., & Shuid, A. N. (2015). Cosmos caudatus enhances fracture healing in ovariectomised rats: A preliminary biomechanical evaluation. *International Journal of Applied Research in Natural Products*, 8(1), 11-19.
- Salehan, N. M., Meon, S., & Ismail, I. S. (2013). Antifungal activity of Cosmos caudatus extracts against seven economically important plant pathogens. *International Journal of Agriculture & Biology*, 15, 864-870.
- Samsonowicz, M., Kamińska, I., Kalinowska, M., & Lewandowski, W. (2015). Alkali metal salts of rutin – Synthesis, spectroscopic (FT-IR, FT-Raman, UV-VIS), antioxidant and antimicrobial studies. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 151, 926-938.
- Sargent, J., & Taylor, C. (1989). Appraisal of the MTT assay as a rapid test of chemosensitivity in acute myeloid leukaemia. *British Journal of Cancer*, 60(2), 206-211.
- Sato, K., Gosho, M., Yamamoto, T., Kobayashi, Y., Ishii, N., Ohashi, T., Nakade, Y., Ito, K., Fukuzawa, Y., & Yoneda, M. (2015). Vitamin E has a beneficial effect on nonalcoholic fatty liver disease: A meta-analysis of randomized controlled trials. *Nutrition*, 31(7–8), 923-930.
- Scalbert, A., Manach, C., Morand, C., Rémesy, C., & Jiménez, L. (2005). Dietary Polyphenols and the Prevention of Diseases. *Critical Reviews in Food Science and Nutrition*, 45(4), 287-306.
- Schaich, K. M., Tian, X., & Xie, J. (2015). Hurdles and pitfalls in measuring antioxidant efficacy: A critical evaluation of ABTS, DPPH, and ORAC assays. *Journal of Functional Foods*, 14(0), 111-125.
- Seo, J., Lee, S., Elam, M. L., Johnson, S. A., Kang, J., & Arjmandi, B. H. (2014). Study to find the best extraction solvent for use with guava leaves (*Psidium guajava* L.) for high antioxidant efficacy. *Food Science & Nutrition*, 2(2), 174-180.
- Sepehri, H., & Ganji, F. (2016). The protective role of ascorbic acid on hippocampal CA1 pyramidal neurons in a rat model of maternal lead exposure. *Journal of Chemical Neuroanatomy*, 74, 5-10.

- Shabbir, M., Syed, D. N., Lall, R. K., Khan, M. R., & Mukhtar, H. (2015). Potent Anti-Proliferative, Pro-Apoptotic Activity of the Maytenus Royleanus Extract against Prostate Cancer Cells: Evidence in In-Vitro and In-Vivo Models. *PLoS One*, 10(3), 119-139.
- Shahidi, F., & Naczk, M. (2004). Antioxidant Properties of Food Phenolics. *Phenolics in food and nutraceuticals* (pp. 403-437). USA: CRC press.
- Shimoyamada, M., Osugi, Y., Shiraiwa, M., & Okubo, K., and Watanabe, K. (1993). Solubilities of Soybean Saponins and Their Solubilization with a Bisdesmoside Saponin. *Japanese Society for Food Science and Technology*, 40(3), 210-213.
- Shin, J.-C., Jung, H.-Y., Harikishore, A., Kwon, O.-D., Yoon, H. S., Kim, K.-T., & Choi, B.-H. (2013). The flavonoid myricetin reduces nocturnal melatonin levels in the blood through the inhibition of serotonin N-acetyltransferase. *Biochemical and Biophysical Research Communications*, 440(2), 312-316.
- Shirpoor, A., Barmaki, H., Khadem Ansari, M., Ikhanezadeh, B., & Barmaki, H. (2016). Protective effect of vitamin E against ethanol-induced small intestine damage in rats. *Biomedicine & Pharmacotherapy*, 78, 150-155.
- Shui, G., Leong, L. P., & Wong, S. P. (2005). Rapid screening and characterisation of antioxidants of *Cosmos caudatus* using liquid chromatography coupled with mass spectrometry. *Journal of Chromatography B*, 827(1), 127-138.
- Silvestre-Roig, C., de Winther, M. P., Weber, C., Daemen, M. J., Lutgens, E., & Soehnlein, O. (2014). Atherosclerotic Plaque Destabilization Mechanisms, Models, and Therapeutic Strategies. *Circulation Research*, 114(1), 214-226.
- Song, W. O., & Chun, O. K. (2008). Tea is the major source of flavan-3-ol and flavonol in the US diet. *The Journal of Nutrition*, 138(8), 1543S-1547S.
- Sparg, S. G., Light, M. E., & van Staden, J. (2004). Biological activities and distribution of plant saponins. *Journal of Ethnopharmacology*, 94(2-3), 219-243. doi:
- Sriraman, S., Ramanujam, G. M., Ramasamy, M., & Dubey, G. P. (2015). Identification of beta-sitosterol and stigmasterol in *Bambusa bambos* (L.) Voss leaf extract using HPLC and its estrogenic effect in vitro. *Journal of Pharmaceutical and Biomedical Analysis*, 115, 55-61.
- Staels, B. (2005). PPAR[gamma] and atherosclerosis. *Current Medical Research and Opinion*, 21, S13-20.

- Stocker, R., & Keaney, J. F. (2004). Role of oxidative modifications in atherosclerosis. *Physiological Reviews*, 84(4), 1381-1478.
- Suganthy, N., & Devi, K. P. (2016). Protective effect of catechin rich extract of Rhizophora mucronata against β -amyloid-induced toxicity in PC12 cells. *Journal of Applied Biomedicine* 10(54), 654-669.
- Sukrasno, S., Fidriany, I., Anggadiredja, K., Handayani, W. A., & Anam, K. (2011). Influence of drying method on flavonoid content of *Cosmos caudatus* (Kunth) leaves. *Research Journal of Medicinal Plant*, 5(2), 189-195.
- Sultana, B., Anwar, F., & Ashraf, M. (2009). Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts. *Molecules*, 14(6), 2167-2180.
- Sun, P., Dwyer, K. M., Merz, C. N. B., Sun, W., Johnson, C. A., Shircore, A. M., & Dwyer, J. H. (2000). Blood Pressure, LDL Cholesterol, and Intima-Media Thickness A Test of the "Response to Injury" Hypothesis of Atherosclerosis. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 20(8), 2005-2010.
- Sun, T., & Ho, C.-T. (2005). Antioxidant activities of buckwheat extracts. *Food Chemistry*, 90(4), 743-749.
- Sung, C.-C., Hsu, Y.-C., Chen, C.-C., Lin, Y.-F., & Wu, C.-C. (2013). Oxidative stress and nucleic acid oxidation in patients with chronic kidney disease. *Oxidative Medicine and Cellular Longevity*, 2013(8), 219-234.
- Tabas, I. (2010). Macrophage death and defective inflammation resolution in atherosclerosis. *Nature Reviews Immunology*, 10(1), 36-46.
- Tadera, K., Minami, Y., Takamatsu, K., & MATSUOKA, T. (2006). Inhibition of ALPHA.-Glucosidase and ALPHA.-Amylase by Flavonoids. *Journal of Nutritional Science and Vitaminology*, 52(2), 149-153.
- Tannock, L. R., & King, V. L. (2008). Proteoglycan mediated lipoprotein retention: A mechanism of diabetic atherosclerosis. *Reviews in Endocrine & Metabolic Disorders*, 9(4), 289-300.
- Taylor, N. (1961). *Taylor's encyclopedia of gardening* (t. U. o. Michigan Ed.). USA: Houghton Mifflin.
- Teng, Y.-N., Sheu, M.-J., Hsieh, Y.-W., Wang, R.-Y., Chiang, Y.-C., & Hung, C.-C. (2016). β -carotene reverses multidrug resistant cancer cells by selectively modulating human P-glycoprotein function. *Phytomedicine*, 23(3), 316-323.

- Tian, Y., Kijlstra, A., Webers, C. A. B., & Berendschot, T. T. J. M. (2015). Lutein and Factor D: Two intriguing players in the field of age-related macular degeneration. *Archives of Biochemistry and Biophysics*, 572, 49-53.
- Traore, F., Faure, R., Ollivier, E., Gasquet, M., Azas, N., Debrauwer, L., Keita, A., Timon-David, P., & Balansard, G. (2000). Structure and antiprotozoal activity of triterpenoid saponins from *Glinus oppositifolius*. *Planta Medica*, 66(4), 368-371.
- Treutter, D. (2006). Significance of flavonoids in plant resistance: a review. *Environmental Chemistry Letters*, 4(3), 147-157.
- Ulatowski, L. M., & Manor, D. (2015). Vitamin E and neurodegeneration. *Neurobiology of Disease*, 84, 78-83.
- Van Remmen, H., Ikeno, Y., Hamilton, M., Pahlavani, M., Wolf, N., Thorpe, S. R., Alderson, N. L., Baynes, J. W., Epstein, C. J., & Huang, T.-T. (2003). Life-long reduction in MnSOD activity results in increased DNA damage and higher incidence of cancer but does not accelerate aging. *Physiological Genomics*, 16(1), 29-37.
- Vergeade, A., Mulder, P., Vendeville, C., Ventura-Clapier, R., Thuillez, C., & Monteil, C. (2012). Xanthine oxidase contributes to mitochondrial ROS generation in an experimental model of cocaine-induced diastolic dysfunction. *Journal of Cardiovascular Pharmacology*, 60(6), 538-543.
- Vermorken, A. J. M., Zhu, J., Van de Ven, W. J. M., & Andrès, E. (2012). Curcumin for monoclonal gammopathies. What can we hope for, what should we fear?. *Critical Reviews in Oncology/Hematology*, 84(3), 350-360.
- Verza, S. G., Silveira, F., Cibulski, S., Kaiser, S., Ferreira, F., Gosmann, G., Roehe, P. M., & Ortega, G. G. (2012). Immunoadjuvant activity, toxicity assays, and determination by UPLC/Q-TOF-MS of triterpenic saponins from *Chenopodium quinoa* seeds. *Journal of Agricultural and Food Chemistry*, 60(12), 3113-3118.
- Victor, V. M., & Rocha, M. (2007). Targeting Antioxidants to Mitochondria: A Potential New Therapeutic Strategy for Cardiovascular Diseases. *Current Pharmaceutical Design*, 13(8), 845-863.
- Vincken, J.-P., Heng, L., de Groot, A., & Gruppen, H. (2007). Saponins, classification and occurrence in the plant kingdom. *Phytochemistry*, 68(3), 275-297.
- Vogiatzi, G., Tousoulis, D., & Stefanadis, C. (2009). The role of oxidative stress in atherosclerosis. *Hellenic Journal of Cardiology*, 50(5), 402-409.

- Voutquenne, L., Guinot, P., Thoison, O., Sevenet, T., & Lavaud, C. (2003). Oleanolic glycosides from *Pometia ridleyi*. *Phytochemistry*, 64(3), 781-789.
- Vuong, Q. V., Hirun, S., Chuen, T. L. K., Goldsmith, C. D., Murchie, S., Bowyer, M. C., Phillips, P. A., & Scarlett, C. J. (2015). Antioxidant and anticancer capacity of saponin-enriched *Carica papaya* leaf extracts. *International Journal of Food Science and Technology*, 50(1), 169-177.
- Wagner, H. (1999). New approach in phytopharmacological research. *Pure and Applied Chemistry*, 71(9), 1649-1654.
- Wang, J., Xu, Q.-L., Zhou, Z.-Y., & Tan, J.-W. (2014). Caffeoylquinic acid derivatives from stems of *Akebia trifoliata*. *Zhong yao cai = Zhongyaocai = Journal of Chinese medicinal materials*, 37(7), 1190-1193.
- Weber, C., & Noels, H. (2011). Atherosclerosis: current pathogenesis and therapeutic options. *Nature Medicine*, 17(11), 1410-1422.
- Weng, L., Guo, X., Li, Y., Yang, X., & Han, Y. (2016). Apigenin reverses depression-like behavior induced by chronic corticosterone treatment in mice. *European Journal of Pharmacology*, 774, 50-54.
- Westmuckett, A. D., Thacker, K. M., & Moore, K. L. (2011). Tyrosine Sulfation of Native Mouse Psgl-1 Is Required for Optimal Leukocyte Rolling on P-Selectin In Vivo. *PLoS One*, 6(5), 204-216.
- Wettasinghe, M., & Shahidi, F. (1999). Antioxidant and free radical-scavenging properties of ethanolic extracts of defatted borage (*Borago officinalis* L.) seeds. *Food Chemistry*, 67(4), 399-414.
- Wiersema, J. H., & Leon, B. (2013). *World economic plants: a standard reference*: CRC press.
- Williamson, E. M. (2001). Synergy and other interactions in phytomedicines. *Phytomedicine*, 8(5), 401-409.
- Winslow, L. C., & Kroll, D. J. (1998). Herbs as medicines. *Archives of Internal Medicine*, 158(20), 2192-2199.
- Wong, S. P., Leong, L. P., & William Koh, J. H. (2006). Antioxidant activities of aqueous extracts of selected plants. *Food Chemistry*, 99(4), 775-783.
- Wu, J., Maoqiang, L., Fan, H., Zhenyu, B., Qifang, H., Xuepeng, W., & Liulong, Z. (2016). Rutin Attenuates Neuroinflammation in Spinal Cord Injury Rats. *Journal of Surgical Research* 9(25), 214-221.

- Xie, J.-H., Dong, C.-j., Nie, S.-P., Li, F., Wang, Z.-J., Shen, M.-Y., & Xie, M.-Y. (2015). Extraction, chemical composition and antioxidant activity of flavonoids from Cyclocarya paliurus (Batal.) Iljinckaja leaves. *Food Chemistry*, 186, 97-105.
- Xie, J., & Schaich, K. (2014). Re-evaluation of the 2, 2-Diphenyl-1-picrylhydrazyl Free Radical (DPPH) Assay for Antioxidant Activity. *Journal of Agricultural and Food Chemistry*, 62(19), 4251-4260.
- Xu, H., Yang, T., Liu, X., Tian, Y., Chen, X., Yuan, R., Su, S., Lin, X., & Du, G. (2016). Luteolin synergizes the antitumor effects of 5-fluorouracil against human hepatocellular carcinoma cells through apoptosis induction and metabolism. *Life Sciences*, 144, 138-147.
- Yalinkilic, O., & Enginar, H. (2008). Effect of X-Radiation on Lipid Peroxidation and Antioxidant Systems in Rats Treated with Saponin-containing Compounds. *Photochemistry and Photobiology*, 84(1), 236-242.
- Yang, Y.-I., Kim, J.-H., Lee, K.-T., & Choi, J.-H. (2011). Costunolide induces apoptosis in platinum-resistant human ovarian cancer cells by generating reactive oxygen species. *Gynecologic Oncology*, 123(3), 588-596.
- Yang, Z., Pan, A., Zuo, W., Guo, J., & Zhou, W. (2014). Relaxant effect of flavonoid naringenin on contractile activity of rat colonic smooth muscle. *Journal of Ethnopharmacology*, 155(2), 1177-1183.
- Yoshiki, Y., Takagi, S., Watanabe, M., & Okubo, K. (2005). Fractionation of soybean functional glycosides from soy-waste based on the chemical reaction of soyasaponin beta g. *Food Chemistry*, 93(4), 591-597.
- Yu, Y.-M., Lin, H.-C., & Chang, W.-C. (2008). Carnosic acid prevents the migration of human aortic smooth muscle cells by inhibiting the activation and expression of matrix metalloproteinase-9. *The British Journal of Nutrition*, 100(4), 731-738.
- Zampetaki, A., Dudek, K., & Mayr, M. (2013). Oxidative stress in atherosclerosis: The role of microRNAs in arterial remodeling. *Free Radical Biology and Medicine*, 64(0), 69-77.
- Zanariah, J., Rehan, A. N., Rosnah, O., & Noor-Rehan, A. (1986). Protein and amino acid compositions of Malaysian vegetables. *MARDI Research Bulletin*, 14(2), 140-147.
- Zhang, J., Jin, N., Liu, Y., & Rhoades, R. A. (1998). Hydrogen peroxide stimulates extracellular signal-regulated protein kinases in pulmonary arterial smooth muscle cells. *American Journal of Respiratory Cell and Molecular Biology*, 19(2), 324-332.

- Zhang, L., Nan, C., Chen, Y., Tian, J., Jean-Charles, P.-Y., Getfield, C., Wang, X., & Huang, X. (2015). Calcium desensitizer catechin reverses diastolic dysfunction in mice with restrictive cardiomyopathy. *Archives of Biochemistry and Biophysics*, 573, 69-76.
- Zhang, L. J., Quan, W., Wang, B. B., Shen, B. L., Zhang, T. T., & Kang, Y. (2011). Berberine inhibits HEp-2 cell invasion induced by Chlamydophila pneumoniae infection. *The Journal of Microbiology*, 49(5), 834-840.
- Zhang, R., Yao, Y., Wang, Y., & Ren, G. (2011). Antidiabetic activity of isoquercetin in diabetic KK -Ay mice. *Nutrition & Metabolism*, 8, 85-99.
- Zhang, T., Su, J., Guo, B., Wang, K., Li, X., & Liang, G. (2015). Apigenin protects blood-brain barrier and ameliorates early brain injury by inhibiting TLR4-mediated inflammatory pathway in subarachnoid hemorrhage rats. *International Immunopharmacology*, 28(1), 79-87.
- Zhou, J., Zhou, T., Chen, M., Jiang, M., Wang, X., Liu, Q., Zhan, Z., & Zhang, X. (2014). Research progress on synergistic anti-tumor mechanisms of compounds in traditional Chinese medicine. *Journal of Traditional Chinese Medicine*, 34(1), 100-105.
- Zhou, Y., Ruan, Z., Zhou, L., Shu, X., Sun, X., Mi, S., Yang, Y., & Yin, Y. (2016). Chlorogenic acid ameliorates endotoxin-induced liver injury by promoting mitochondrial oxidative phosphorylation. *Biochemical and Biophysical Research Communications*, 469(4), 1083-1089.
- Zuorro, A., & Lavecchia, R. (2013). Influence of Extraction Conditions on The Recovery of Phenolic Antioxidants From Spent Coffee Grounds. *American Journal of Applied Sciences*, 10(5), 478-486.

BIODATA OF STUDENT

Said Faroq Saiyid Moshawih was born in Syria, grew up in Amman, the capital city of Jordan. He has earned all of his school degrees in addition to the secondary education in the scientific stream from Jordan. His Bachelor degree was received from Al-Zaytoonah university of Jordan in 2005, in pharmacy, and his ranking was excellent. After graduation, he has tried to get scholarship to pursue his postgraduate studies, but all efforts were gone in vain. Therefore, he found himself obliged to work in the business sector as a pharmacist in a community pharmacy for five years, very far from his academic dreams, this was followed by another five years in a hospital pharmacy in Dubai – United Arab Emirates. However, because hope was still vivid in his mind, he left his family, work, salary and gossips and joined UPM to study Master of Science in pharmacology and toxicology.

His managerial experience in the hospital pharmacy was helpful for him to digest as much as possible from research tools and experimental studies within a short time, this experience was provided generously from UPM. His experience is especially in cell culture, antioxidation assays, animal studies and fluorescent microscopy. His research interests now and future intensions are mostly in stem cells, neurosciences and cancer research. Those are newly evolving topics can provide many medicinal benefits for patients afflicted with many diseases.



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