Simulated seawater flooding reduces oilseed rape growth, yield, and progeny performance

Mick E Hanley, Francesca C Hartley, Louise Hayes & Miguel Franco

School of Biological and Marine Sciences, University of Plymouth, Plymouth PL4 8AA, England.

Author for correspondence: Dr Mick Hanley Tel: +44 (0) 1752 584631 E-mail: mehanley@plymouth.ac.uk

Running Head: Seawater flooding reduces oilseed rape yield and seedling

performance

Background and Aims Of the many threats to global food security, one of the most
 pressing is the increased incidence of extreme weather events. In addition to extreme
 rainfall, a combination of global sea level rise and storm surge is likely to result in
 frequent episodes of seawater flooding in arable systems along low-lying coasts. Our
 aim was to elucidate the effects of simulated seawater and freshwater flooding on the
 survival, growth, and reproductive potential of four cultivars of the important seed
 crop, *Brassica napus* (canola, or oilseed rape [OSR]).

Methods Established plants were exposed to 24- or 96-hour freshwater or seawater
root zone immersion (with a no immersion 'control'). Initial post-treatment
performance over 7-weeks was quantified using dry weight biomass. A second group
of plants, cultivated until maturity, were used to quantify reproductive yield (siliqua
and seed number, and seed size) and subsequent progeny performance (germination
and seedling growth).

Results OSR growth and reproductive responses were unaffected by freshwater, but
 seawater negatively affected growth and siliqua number for all cultivars, and seed mass
 for two (Agatha & Cubic). In addition to impacts on crop yield, the growth of seedlings
 cultivated from seed collected from maternal plants subjected to seawater immersion
 was also reduced.

Conclusions Our results demonstrate the potential impact of seawater inundation on coastal cropping systems; although OSR may survive acute saline flooding, there are longer-term impacts on growth and yield for some cultivars. The threat may necessitate changes in land-use practice and/or the development of salt-tolerant cultivars to maintain economically viable yields. In addition, by evidencing a hitherto unknown effect on reproductive performance (i.e. reduced seed yield) and subsequent seedling

- 25 growth our study highlights an important potential impact of coastal flooding on plant
- 26 community dynamics for (semi) natural habitats.
- Key Words Flooding, Food security, Osmotic Stress; Salinity, Sea-level rise; Storm
 surge
- 29
- 30
- 31

32

INTRODUCTION

33 Food security and climate change are two of the most pressing environmental issues facing 34 the planet (Godfray et al., 2010; IPCC, 2014). When combined however, the increased 35 food demand imposed by a growing human population coupled with global-scale shifts in temperatures, precipitation, and sea-level rise will inevitably impose significant demands 36 37 on agriculture (Parry et al., 2005; Lobell et al., 2008). Much of the environmental threat 38 from climate change stems from acute extreme events rather than simply longer-term 39 chronic change alone (Rahmstorf & Coumou, 2011; Vasseur et al., 2014; Parmesan & Hanley, 2015), and of the former, flooding represents one of the greatest challenges. Future 40 41 climate scenarios consistently predict major regional shifts in the intensity of extreme 42 precipitation episodes across the globe and concomitant increases in large-scale regional flooding along river catchments (Li et al., 2013; Singh et al., 2013; IPCC, 2014). In 43 addition to fluvial flooding however, a combination of changes in sea-surface 44 temperatures, weather patterns, and sea-level rise are predicted to increase the frequency 45 and severity of oceanic storm surges (Vousdoukas et al., 2016; Vitousek et al., 2017). The 46 47 result will be that many low-lying coastal areas face an increased risk of seawater inundation over coming decades (Nicholls & Cazenave, 2010; Hoggart et al., 2014). When 48 taken together, extreme precipitation, storm surge, and sea-level rise are likely to cause 49 50 significant flood risk to global agriculture and as a result understanding crop response to, and tolerance of, coastal flooding has become a research priority (Jackson & Ismail, 2015). 51

Although flooding can cause significant losses at any time of the crop production cycle,
yields are most likely to suffer if flooding occurs during critical development stages
(Rosenzweig *et al.*, 2001; Parry *et al.*, 2005). For the United Kingdom for example,

55 extreme winter and spring precipitation is projected to increase (Fowler et al., 2010) at a 56 time when many arable crops are establishing ahead of summer harvest. This is one reason why the severe, but localised, 2014 winter floods that affected only a small portion of 57 southern England (14,000 Ha), caused economic losses estimated at £6.9 million in arable 58 systems alone (ADAS, 2014). Globally, severe freshwater and seawater flooding has long 59 60 been identified as a significant economic problem for agriculture, and one that poses an 61 additional regional threat to human nutrition (Page & Williams, 1926; Mirza, 2002; Chau 62 et al., 2015).

The impact of flooding on crop plants is well known, although research has almost 63 exclusively focussed on the effects of freshwater inundation. Flooding typically causes soil 64 anoxia, as O_2 is consumed without replacement, and the accumulation of various ions 65 $(Mn^{2+}, Fe^{2+}, S^{2+})$ and compounds detrimental to plant growth, while submergence also 66 denies plant tissues access to O₂ and CO₂ (Flowers & Colmer, 2008; Perata et al., 2011). 67 All of these factors can limit significantly crop growth and productivity (Malik et al., 2001; 68 Perata et al., 2011; Ren et al., 2014; Mutava et al., 2015). Due its high salinity (typically 69 about 35 gL⁻¹ salt of which chloride and sodium contribute 19 gL⁻¹ and 11 gL⁻¹ 70 71 respectively), seawater flooding imposes additional osmotic and ionic stresses; the former 72 limits the plant's ability to absorb water and the latter can result in toxicity through the 73 accumulation of Na⁺ and Cl⁻ in tissues (Munns & Tester, 2008). As a response, plants have 74 adapted to avoid or lessen the impact of salt ions via the synthesis and accumulation of stress metabolites and the regulation of other tissue ions (i.e. K⁺) to exclude or 75 76 compartmentalize Na⁺ and Cl⁻ and re-establish homeostatic function (Maathuis & Amtmann, 1999; Munns & Tester, 2008). 77

78 From the perspective of agricultural production, even if crop plants survive freshwater or 79 seawater flooding, any impact on growth or reproductive potential is of concern. Our understanding of the effects of salinity on crop performance is, however, focussed largely 80 81 on soil salinization in arid cropping systems (Pitman & Läuchli, 2002; Albacete et al., 2008; Flowers et al., 2010), and only a handful of studies have examined how 82 83 agriculturally important species respond to short-duration immersion in seawater. White et 84 al. (2014) report how immersion in seawater for only 24-hr resulted in the accumulation of Na⁺ and Cl⁻ in leaves of the pasture crop white clover (*Trifolium repens*). A concomitant 85 increase in two key organic solutes (proline and sucrose) to re-establish osmotic balance 86 87 was also observed, but nonetheless flooded plants showed reduced growth and flowering 88 potential. Interestingly White et al. (2014) also found variation between the responses of three different ecotypes to seawater immersion, suggesting that the relative tolerance of 89 90 one ecotype could be of value in producing flood-tolerant cultivars for cultivation in low-91 lying coastal pastures. Sun et al. (2015) is the only study to look at effects of seawater 92 immersion on arable crops, their experiment focussing on the responses of 1 month old 93 plants of ten crop plants (six of which were Brassicaceae) to 24-hr seawater immersion. 94 All crops were negatively affected by immersion, with two (cucumber and Chinese greens) 95 suffering complete mortality and the remainder showing reduced growth. Since plants 96 were harvested only 2 weeks after immersion treatments were imposed, it is unclear however, what potential any of the surviving crops had for long-term recovery or whether 97 other key aspects of the crop production cycle (flowering & fruit/seed production) were 98 99 affected by flooding.

100 The aim of this study was to determine how exposure to simulated seawater and freshwater101 flooding affected both immediate growth and longer-term reproductive performance of the

102 widely cultivated crop canola, or oilseed rape (OSR - Brassica napus L.). Beyond gaining 103 an understanding of the short-term impact of flooding on mortality and growth of 104 established plants, we also test the hypothesis that impacts on reproductive potential and 105 subsequent seedling performance vary across four different cultivars. Elucidation of these 106 responses is important for several reasons; OSR is grown primarily for seed yield and thus 107 any impact of prolonged fluvial or seawater flooding on reproductive output has potential 108 economic repercussions. In addition, establishing variation in growth and yield responses 109 may help identify established genetic materials from which to develop flood/salinity 110 tolerant cultivars. A more general understanding of plant growth and reproductive 111 responses to acute seawater flooding is also lacking in the literature (see Hanley et al., 112 Submitted), and no study to date has tested the hypothesis that progeny performance is 113 affected by the exposure of parent plants to short-duration seawater immersion. 114 Consequently, our study offers an important insight into the longer-term impacts that this 115 increasing environmental issue may have on natural, as well as agricultural, plant species and communities. 116

117

MATERIALS AND METHODS

118 Study species

Oilseed rape is a hybrid of *B. rapa* and *B. oleracea* (Chalhoub *et al.*, 2014), wild forms of
the latter being a coastal species with some natural salt tolerance (Snogerup *et al.*, 1990).
OSR is grown globally and its seeds are harvested to produce food and bioenergy oils, and
for use as an animal feedstock. It accounts for 14% (by area) of agricultural land use in the
UK (Garthwaite *et al.*, 2018) and major producers globally include the EU, Canada, India
and China (USDA, 2015), regions likely to experience significant flooding events over

125 coming decades (IPCC, 2014). The OSR cultivars used in these experiments were Cracker

- 126 (LS Plant Breeding, Impington, UK), Agatha, Astrid, and Cubic (Grainseed Ltd., Eye,
- 127 Suffolk, UK). All are commonly cultivated in the UK.

128 Plant cultivation

129 One hundred seeds of each cultivar were germinated in 90 mm-diameter Petri dishes 130 containing two layers of Whatman No. 1 filter paper moistened with 5 ml of distilled water, maintained in an incubator at 16.5 °C on a 12:12 light:dark cycle. On germination (radicle 131 132 emergence), seventy seedlings per cultivar were transplanted individually into 50mm 133 diameter, 50mm deep plastic pots containing John Innes No.2 potting compost and grown 134 on in a naturally lit greenhouse with weekly watering with tap water, until early November 135 2014. At this time seedlings possessing at least the first true leaf (equivalent to OSR 136 growth-stage 1.1; Sylvester-Bradley, 1985), were re-potted into larger 110 x 110 x 120 mm plastic pots containing John Innes No. 2 potting compost. Greenhouse temperatures 137 138 during this cultivation period were: mean daily minimum = 10.4 °C (± 0.4 °C SE); mean 139 daily maximum = $25.7 \text{ °C} (\pm 1.0 \text{ °C})$.

140 Effect of immersion on growth

141 When the plants were 67-d-old (early December 2014), twelve individuals from each 142 cultivar (OSR growth stage 1.3-1.6; i.e. emergence of third-sixth true leaf) were allocated 143 at random to one of five treatment groups. These were 24-, or 96-hrs immersion in seawater 144 (*Sw*) (collected from Plymouth Sound - electrical conductivity = 45.51 mS cm⁻¹ at 16.2 145 $^{\circ}$ C); 24-, or 96-hrs immersion in deionised water (*Dw*); or a no-immersion control 146 treatment. Although river floodplains can experience much longer periods of immersion, 147 sometimes extending to several months (Van Eck *et al.*, 2004; Muchan *et al.*, 2015), in this way we simulated the average 1-d long seawater flooding event reported for low-lying UK
coastline habitats and extended the period to the maximum reported flood duration of 4-d
(Environment Agency, 2014).

151 We recognise that inundation following coastal storm-surge or fluvial flooding events 152 would likely result in full shoot submergence, but by immersing to pot-level (in large 153 plastic tubs) our approach allowed us to separate the effect of ionic imbalance in the root-154 zone rather than the impact of oxygen deficiency caused by full immersion that both 155 treatments would impose. Immediately after immersion, the pots were allowed to drain 156 fully before being arranged randomly on a wire mesh-topped bench inside the greenhouse; the wire mesh allowed free drainage and prevented cross-contamination between treatment 157 158 groups. The pots were watered to capacity (with tap water) 48-hr after seawater immersion.

Eight plants per treatment/cultivar combination were retained inside the greenhouse; the remainder were positioned outside, on adjacent elevated mesh. For both sets of plants, each individual was positioned at random, 20cm apart from its nearest neighbour in 1m long rows separated by 30cm to simulate the recommended field density for OSR cropping systems in the UK (HGCA, 2014).

Greenhouse plants were watered twice weekly for a further 56-d; temperatures during this phase of the experiment were 4.9 °C (\pm 0.5) min and 18.0 °C (\pm 0.6) max. Surviving plants (growth stages 2.0-2.3; i.e. plant has a rosette growth form and extended stem with internodes) from each treatment/cultivar group were harvested at 113-d-old (late January 2015), cleaned of any adhering compost and oven-dried at 50 °C for 24-hr. A Levene's test for homogeneity of variance across treatment and cultivar levels was negative ($F_{19,189} =$ 2.896, P < 0.001) and biomass data were log₁₀ transformed, resulting in a positive Levene's

test ($F_{19,189} = 1.335$, P = 0.165). Univariate GLM analyses were used to compare the results of experimental flooding on transformed biomass. Rather than classifying plants in different groups into two separate factors (water: Dw vs. Sw, and time: 24h vs. 96h), which would decrease the power of the tests, contrasts were employed to evaluate specific differences between treatment levels (control, Dw24, Dw96, Sw24 and Sw96). In addition to the *F* statistic and its probability, we report effect sizes (η_p^2) and power at P<0.05 (power_{0.05}).

178 Effect of immersion on crop yield and progeny performance

179 The four individual plants from each treatment/cultivar group positioned on elevated 180 outdoor standing were grown to reproductive maturity in order to assess the effects of 181 immersion on seed yield. Plants were exposed to natural weather conditions and watered 182 (to capacity with tap water) only following prolonged dry periods. Since individuals were 183 randomly arranged with respect to treatment and cultivar, they were able to cross-184 pollinate and so resulting progeny were most likely hybrids between cultivars. Three 185 plants (Agatha 24-hr Dw; Cracker 96-hr Dw; Agatha 96-hr Sw) died during this period. 186 The main stems of plants were harvested in mid-June when most siliqua were fully 187 ripened (growth stage 9.9). We quantified the total number of siliqua per plant, and from 188 six (randomly selected) siliqua per plant, quantified seed number and mean individual 189 seed mass per pod (averaging across all 6 replicate siliqua for each plant). 190 All seeds form each of the six sampled siliqua per plant were pooled. From these, 40 191 seeds were set to germinate in 90 mm-diameter Petri dishes containing two layers of

192 Whatman No. 1 filter paper moistened with 5 ml of deionised water, maintained in an

193 incubator at 18°C on a 12:12 light:dark cycle. In addition to the three plants that died

194	before harvest, a further two parent plants - Cracker 96-hr Dw & Agatha 96-hr Sw –
195	failed to yield sufficient seeds for the germination/seedling growth trials. Petri dishes
196	were checked daily for 14 days; on germination (appearance of the radicle), seedlings
197	were counted and removed. Six seedlings from each Petri dish were retained, and planted
198	into a 50-mm diameter pot containing John Innes No.2 potting compost. These seedlings
199	were grown in controlled conditions (15°C on a 12:12 light:dark cycle with daily
200	watering to capacity) until 14-d-old when harvested and oven dried (at 50 $^{\circ}$ C for 24-hr)
201	to determine dry weight biomass.
202	Levene's tests of homogeneity of variances were significant for siliqua number ($F_{19,57} =$
203	2.621, $P = 0.003$), seed number ($F_{19,56} = 2.962$, $P < 0.001$) and seedling biomass ($F_{19,498}$
204	= 2.291, $P = 0.002$), but did not show departure from homogeneity for seed mass ($F_{19,56} =$
205	1.264, $P=0.244$). Logarithmic transformation of the former three homogenised the
206	variance for siliqua number ($F_{19,57} = 1.304$, $P=0.217$), but not for seed number ($F_{19,56} =$
207	10.625, $P < 0.001$) and seedling mass ($F_{19,497} = 3.289$, $P < 0.001$). Consequently, we
208	present the results of GLM's hypothesis testing for; (i) seed mass where the
209	untransformed variable did not depart from homogeneous; (ii) seed number and seedling
210	mass where logarithmic transformation did not homogenise the variance; (iii) log siliqua
211	number where logarithmic transformation resulted in variance homogeneity.
212	RESULTS

SULIS

Effect of immersion on plant growth 213

'Treatment' had a significant effect on adult plant biomass with a moderate effect size 214 $(F_{4,189} = 17.71, P < 0.001; \eta_p^2 = 0.273; power_{0.05} = 1; Fig 1)$, and with the contrasts between 215

216 control and each of the other four treatment levels significant only for Sw96 (Contrast = -0.339, SE = 0.066, P < 0.001). More specific comparison showed: (i) a negative contrast 217 218 between both Sw treatments vs. Control (Contrast = -0.219, SE = 0.058, P < 0.001); (ii) a 219 small, but significant, positive contrast between Dw treatments vs. Control (Contrast = 220 0.114, SE = 0.057, P = 0.048); and (iii) a then obvious positive contrast between Dw and 221 Sw (Contrast = 0.333, SE = 0.045, P < 0.001), highlighting the smaller size of plants 222 immersed in seawater. Plant biomass did not vary between cultivars but the cultivar effect size was small ($F_{3,189} = 1.85$, P = 0.140; $\eta_p^2 = 0.029$; power_{0.05}=0.475). A significant 223 224 'Treatment × Cultivar' interaction with moderate effect size ($F_{12,189} = 2.95$, P = 0.001; $\eta_p^2 =$ 0.158; power_{0.05}=0.989) emphasised several treatment- and cultivar-specific departures 225 226 from the general trends described above (Astrid and Agatha for example, were both 227 tolerant of 24-hr seawater immersion). No plants died during this first part of the 228 experiment.

229 Effect of immersion on crop yield and progeny performance

The number of siliqua (Table 1) varied with 'Treatment' with a small effect size ($F_{4,57}$ = 230 5.974, P < 0.001; $\eta_p^2 = 0.295$; power_{0.05}=0.978) but not by 'Cultivar' ($F_{3,57} = 2.004$, P =231 0.124; $\eta_p^2 = 0.095$; power_{0.05}=0.489) and there was no interaction ($F_{12,57} = 1.283$, P = 0.254; 232 $\eta_p^2 = 0.213$; power_{0.05}=0.643). The contrast analyses revealed a significant negative effect 233 234 of Dw and Sw treatments compared to the Control (Contrast = -0.103, SE = 0.050, P = 235 0.046); specifically highlighting lower siliqua numbers in Sw vs. Control (Contrast = -0.141, SE = 0.055, P = 0.014). There was no variation between Dw and Sw (Contrast = 236 237 0.76, SE = 0.046, P = 0.105) or Dw vs. Control (Contrast = -0.065, SE = 0.056, P = 0.250).

238 Seed number per pod (Table 1) was not influenced by 'Treatment' ($F_{4.57} = 2.206$, P =0.080; $\eta_p^2 = 0.134$; power_{0.05}=0.612), and while varying between cultivars ($F_{3,57} = 20.265$, 239 P < 0.001; $\eta_p^2 = 0.516$; power_{0.05}=1), there was no interaction with 'Treatment' ($F_{12,57} =$ 240 1.31, P = 0.239; $\eta_p^2 = 0.216$; power_{0.05}=0.655). Mean individual seed mass (Table 1) did 241 however, vary according to 'Treatment' ($F_{4,57} = 5.456$, P = 0.001; $\eta_p^2 = 0.277$; 242 243 power_{0.05}=0.965) and 'Cultivar' ($F_{3.57} = 19.658$, P < 0.001; $\eta_p^2 = 0.509$; power_{0.05}=1), although there was no significant interaction ($F_{12,57} = 1.799$, P = 0.070; $\eta_p^2 = 0.275$; 244 245 power_{0.05}=0.822). While contrasts highlighted that *Dw* and *Sw* had an overall difference 246 with Control (Contrast = -0.009, SE = 0.004, P = 0.036), this was driven primarily by 247 variation between Sw and Control (Contrast = -0.010, SE = 0.004, P = 0.023), and not by 248 the difference between Dw and Control plants (Contrast = -0.007, SE = 0.005, P = 0.122). 249 Interestingly, however, there was no variation between Dw and Sw (Contrast = 0.003, SE250 = 0.004, P = 0.373). These results emphasise the different intensity of effects on different 251 cultivars; i.e., Agatha and Cubic showed the most marked negative responses in the 96-252 hour Sw treatment (Table 1).

Germination was unaffected by any of the immersion treatments imposed on parent plants (data not shown); the lowest germination for any one cultivar/treatment group being the 76% recorded for seeds produced by cv Cracker 96-hr *Sw* (with 3 of 4 Petri dishes for this group nonetheless attaining >85% germination).

Finally, seedling mass (Figure 2) differed between treatments with a small effect size $(F_{4,498} = 6.192 \ P < 0.001; \ \eta_p^2 = 0.047; \ power_{0.05} = 0.988)$, but not among cultivars ($F_{3,498} = 1.874 \ P = 0.133; \ \eta_p^2 = 0.011; \ power_{0.05} = 0.486)$, and there was a significant 'Treatment × Cultivar' interaction also with a small effect size ($F_{12,498} = 2.295 \ P = 0.008; \ \eta_p^2 = 0.052;$ 261 power_{0.05}=0.959). Contrasts analysis revealed significant differences between Control and Dw & Sw together (Contrast = -0.004, SE = 0.001, P < 0.001), between Control and Dw262 (Contrast = -0.003, SE = 0.001, P = 0.004), and between Control and Sw (Contrast = -263 0.005, SE = 0.001, P < 0.001, but not between Dw and Sw (Contrast = 0.002, SE = 0.001, 264 P = 0.089). The significant interaction between the two main factors highlights differences 265 266 in the intensity and direction of cultivar responses. The most affected, Cubic, displayed 267 reduced seedling growth in both the 24-hr and 96-hr treatments, while progeny collected 268 from cv Cracker showed no response to Sw immersion (Fig 2). Seedlings grown from Astrid and Agatha parents exhibited reduced growth in the 96-hr Sw treatments only, 269 270 although given the low parental replication for Agatha (only two plants survived to 271 reproduction), the apparent negative response for seedlings collected from this cultivar 272 should be taken with caution.

273

DISCUSSION

274 Our results evidence substantial differences in OSR response to acute freshwater and 275 seawater inundation; while the former exerted some impact on some of the responses 276 examined here, the latter was more generally associated with reduced parent plant 277 growth, seed yield, and even progeny performance. As far as we are aware this is the first 278 time that the impacts of short-duration, acute, Sw flooding on several key growth and 279 reproductive responses have been demonstrated across stages of the life-cycle for any 280 plant species. Indeed the only previous study to examine this issue in arable crops, Sun et al., (2015) was limited to the immediate (2-week) post-immersion response, and while 281 282 White et al., (2014) did look at growth and flowering in white clover over a 70-d post-283 immersion period, they did not consider seed yield or progeny performance. While there

was some variation between cultivars, this study shows that even transitory immersion in
seawater has significant, consistent, and long-lasting impacts on OSR crop yield. Indeed,
the fact that reduced reproductive output (most notably siliqua number) was manifest 6
months after maternal plants were exposed to *Sw* immersion highlights the potential
long-term impacts of seawater flooding on arable cropping systems.

289 The consequences of reduced seed yields on agricultural crops like OSR are obvious, but 290 our results also raise the possibility that wild plant species could suffer reduced 291 reproductive success as a result of acute seawater flooding stress on seed development. It 292 is well established that maternal plant environment alters phenotypic expression in 293 progeny (Herman & Sultan, 2011) and as such the reduced growth of seedlings from 294 maternal plants subjected to Sw immersion reported here is unsurprising. Nonetheless, 295 and while there is evidence that salinity stress can induce similar shifts in maternal 296 provisioning and seedling performance (for example as shown in the grass Cenchrus 297 ciliaris; Ruiz & Taleisnik (2013)), the fact that parent immersion in seawater for only 4 days can negatively influence subsequent seedling growth, highlights more generally the 298 299 impact of coastal flooding on plant community dynamics. Put simply, even if parent 300 plants (of any native species) survive prolonged flooding, their later ability to contribute 301 to the recovering community could be compromised. We know of no study to have 302 demonstrated this response. For two of our maternal cultivars (Cubic and Agatha) where 303 96-hour Sw immersion reduced mean seed mass by 50% or more, the most plausible 304 explanation for the arrested growth of seedlings is that it resulted from reduced seed 305 provisioning by the parent (Zas et al., 2013). We cannot however, discount the 306 possibility that other responses to salinity stress, including epigenetic changes, impact 307 progeny performance and highlight this as a key area for future research.

308 From the perspective of arable plant species, progeny performance may be of minor 309 importance however, since crops are routinely cultivated from commercially sourced seeds. Nonetheless, we demonstrated a consistent and marked reduction in siliqua number 310 311 and seed mass for at least two cultivars (Cubic and Agatha) following 96-hr Sw immersion, 312 highlighting the significant economic impact that seawater flooding might have on coastal 313 OSR crop yields. While a number of studies have shown that prolonged (i.e. one to several 314 weeks) waterlogging reduces both OSR growth and yield (Cannell & Belford, 1980; Zhou 315 & Lin, 1995; Xu et al., 2015), these studies looked at freshwater effects only. Although 316 the relatively short-term Dw immersion treatments imposed here did not yield similar 317 responses (but noting that we did not consider seed oil yield or content (Cannell & Belford, 318 1980; Xu et al., 2015)), all four cultivars exhibited much reduced plant growth, and later 319 siliqua number in the longer duration Sw treatment (see also Hanley et al., 2013). This 320 highlights the challenge posed by salinity; even if a plant survives acute seawater 321 immersion it must prevent or alleviate damage caused by the accumulation of salt ions 322 (Na⁺ and Cl⁻) in tissues. It is interesting that even the most prolonged immersion times (4 323 days) imposed here failed to induce morality; a response that perhaps reflects an innate 324 salinity tolerance due to OSR's heritage in *B. oleracea* and this parent species' natural 325 affinity for maritime conditions (Snogerup et al., 1990). Salt tolerance is however, often 326 accomplished by the accumulation of stress metabolites and the regulation of tissue ions 327 to exclude or compartmentalize the potentially damaging Na⁺ and Cl⁻. Nonetheless even if 328 successfully achieved, as seems to be the case here, there are costs on subsequent plant 329 growth and reproductive performance (Munns & Tester, 2008; White et al., 2014).

At a time of human population growth and economic development there are increasingdemands on food supplies, but when coupled with unpredictable and likely ever more

332 extreme climate events, global food security is far from assured (Godfray et al., 2010; 333 IPCC, 2014). Flooding is widely recognised as one of the key threats to arable crops, but most research emphasis is placed on pluvial, freshwater flooding where the major negative 334 335 impact comes from soil anoxia. In the UK the threshold for crop viability under this scenario is 15 days (ADAS, 2014), but as we show here, by virtue of the added effect of 336 337 salinity, seawater flooding of only 4 days duration can impact OSR yield (while the same 338 duration under freshwater does not). Historically much of the global agricultural salinization problem stems from poor irrigation coupled with excessive evaporation and/or 339 340 deforestation in hot, dry climates (Vinod et al., 2015). Nonetheless, sea-level rise and the 341 expected increase in frequency and severity of storm surges (Vousdoukas et al., 2016; 342 Vitousek et al., 2017) are likely to increase the risk of seawater inundation to temperate coastal arable systems (Nicholls & Cazenave, 2010). Under these conditions, farmers face 343 344 a choice between changing land-use practise, or cultivation of flood-tolerant crops. There is a rich literature documenting salt-tolerance in crops grown in regions where the 345 346 problems of soil salinization are long established; indeed for one of the most important, 347 rice, crops grown near to coasts are frequently subjected to seawater intrusions and a 348 genetic capacity for salt-tolerance has been identified (Ganie et al., 2014). The impact of 349 seawater immersion demonstrated here for OSR, coupled with the increasing risk of 350 seawater flooding for coastal agriculture globally, underscores a new impetus for research into salt tolerance in a wider range of arable crop species (Jackson & Ismail, 2015). 351

352 Acknowledgements

We thank Jane Akerman and Tom Gove for technical assistance, and two anonymousreferees for their comments on an earlier draft of the MS.

355

356

LITERATURE CITED

- ADAS. 2014 Impact of 2014 Winter Floods on Agriculture in England. Agricultural
 Development and Advisory Service, Wolverhampton, UK.
- Albacete A, Ghanem ME, Martínez-Andújar C, Acosta M, Sanchez-Bravo J,
 Martinez V, Lutts S, Dodd IC, Perez-Alfocea F. 2008. Hormonal changes in
 relation to biomass partitioning and shoot growth impairment in salinized tomato
 (Solanum lycopersicum L.) plants. Journal of Experimental Botany 59: 4119-4131.
- 363 Cannell RQ, Belford RK. 1980. Effects of waterlogging at different stages of
 364 development on the growth and yield of winter oilseed rape (*Brassica napus* L.).
 365 *Journal of the Science of Food & Agriculture* 3: 963–965.
- 366 Chau VN, Cassells S, Holland J. 2015. Economic impact upon agricultural production
 367 from extreme flood events in Quang Nam, central Vietnam. *Natural Hazards* 7:
 368 1747-1765.
- 369 Chalhoub B, Denoeud F, Liu S, *et al.*, 2014. Early allopolyploid evolution in the post 370 Neolithic *Brassica napus* oilseed genome. *Science* 345: 950-953.
- 371 Environment Agency UK, 2014. DataShare. http://www.geostore.com/environment 372 agency/. Accessed 4th April 2014.
- Flowers TJ, Colmer TD. 2008. Salinity tolerance in halophytes. *New Phytologist* 179, 945-963.
- Flowers TJ, Gaur PM, Gowda CLL, Krishnamurthy L, Samineni S, Siddique KHM,
 Turner NC, Vadez V, Varshney RK, Colmer TD 2010. Salt sensitivity in
 chickpea. *Plant Cell Environment* 33: 490–509.
- Fowler HJ, Cooley D, Sain SR, Thurston M. 2010. Detecting change in UK extreme
 precipitation using results from the climateprediction.net BBC climate change
 experiment. *Extremes* 13: 241–267.
- Ganie S, Karmakar J, Roychowdhury R, Mondal T, Dey N. 2014. Assessment of
 genetic diversity in salt-tolerant rice and its wild relatives for ten SSR loci and one
 allele mining primer of salT gene located on 1st chromosome. *Plant Systematics & Evolution* 300: 1741–1747.
- 385 Garthwaite DG, Barker I, Ridley L, Mace A, Parrish GP, MacArthur R, Lu Y.2018.
 386 Pesticide Usage Survey Report 271: Arable Crops In The United Kingdom 2016.
 387 London, DEFRA.

388	Godfray HCJ, Beddington JR, Crute IR, Haddad L, Lawrence D, Muir JF, Pretty J,
389	Robinson S, Thomas SM, Toulmin C. 2010. Food security: The challenge of
390	feeding 9 billion people. Science 327: 812–818.
391	Hanley ME, Yip PYS, Hoggart S, Bilton DT, Rundle SD, Thompson RC. 2013. Riding
392	the storm: The response of Plantago lanceolata to simulated tidal flooding. Journal
393	of Coastal Conservation 17, 799-803.
394	Hanley ME, Sanders SKD, Stanton H-M, Billington RA, Boden R. (Submitted) A
395	pinch of salt: Response of coastal grassland plants to simulated seawater inundation
396	treatments. Annals of Botany
397	Herman JJ, Sultan SE 2011. Adaptive transgenerational plasticity in plants: case studies,
398	mechanisms, and implications for natural populations. Frontiers in Plant Science
399	2: 1-10.
400	HGCA. 2014. Oilseed Rape Guide Number 55. HGCA, Stoneleigh Park, Warwickshire,
401	UK, pp 32.
402	Hoggart SPG, Hanley ME, Parker DJ, et al., 2014. The consequences of doing nothing:
403	The effects of seawater flooding on coastal zones. Coastal Engineering 87: 169-
404	182.
405	IPCC. 2014. Climate Change 2014: Synthesis Report. Contribution of Working Groups
406	I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on
407	Climate Change [Core Writing Team, Pachauri RK, Meyer LA. (eds.)]. IPCC,
408	Geneva, Switzerland, 151 pp.
409	Jackson MB, Ismail AM. 2015. Introduction to the Special Issue: Electrons, water and
410	rice fields: plant response and adaptation to flooding and submergence stress. AoB
411	<i>PLANTS</i> 7: doi:10.1093/aobpla/plv078
412	Li JF, Zhang Q, Chen YD, Xu CY, Singh VP. 2013. Changing spatiotemporal patterns
413	of extreme precipitation regimes in China during 2071–2100 based on Earth system
414	models. Journal of Geophysical Research: Atmospheres 118, 12537–12555.
415	Lobell DB, Burke MB, Tebaldi C, Mastrandrea MD, Falcon WP, Naylor RL. 2008.
416	Prioritizing climate change adaptation needs for food security in 2030. Science 319:
417	607–610.
418	Maathuis FJM, Amtmann A. 1999. K ⁺ nutrition and Na ⁺ toxicity: the basis of cellular
419	K ⁺ /Na ⁺ ratios. Annals of Botany 84: 123–133.

420	Malik A, Colmer TD, Lambers H, Schortemeyer M. 2001. Changes in physiological							
421	and morphological traits of roots and shoots of wheat in response to different depths							
422	of waterlogging. Australian Journal of Plant Physiology 28: 1121-1131							
423	Mirza MQ. 2002. Global warming and changes in the probability of occurrence of floods							
424	in Bangladesh and implications. Global Environmental Change 12, 127–138.							
425	Muchan K, Lewis M, Hannaford J, Parry S. 2015. The winter storms of 2013/4 in the							
426	UK: hydrological responses and impacts. Weather 70: 55-61.							
427	Munns R, Tester M. 2008. Mechanisms of salt tolerance. Annual Reviews in Plant							
428	<i>Biology</i> 59 : 651-681.							
429	Mutava RN, Prince SJK, Syed NH, Song L, Valliyodan B, Chen W, Nguyen HT 2015.							
430	Understanding abiotic stress tolerance mechanisms in soybean: A comparative							
431	evaluation of soybean response to drought and flooding stress. Plant Physiology &							
432	Biochemistry 86: 109-120.							
433	Nicholls RJ, Cazenave A. 2010. Sea-level rise and its impact on coastal zones. Science							
434	328 : 1517–1520.							
435	Page HJ, Williams W. 1926. The effect of flooding with seawater on the fertility of the							
436	soil. Journal of Agricultural Science 16: 551-573.							
437	Parry M, Rosenzweig C, Livermore M. 2005. Climate change, global food supply and							
438	risk of hunger, Philosophical Transactions of the Royal Society London; Series B							
439	360 : 2125–2138.							
440	Perata P, Armstrong W, Laurentius AC, Voesenek J. 2011. Plants and flooding stress.							
441	New Phytologist 190: 269–273.							
442	Pitman MG, Läuchli A. 2002. Global impact of salinity and agricultural ecosystems. In							
443	Salinity: Environment – Plants – Molecules, pp. 3–20, Eds Läuchli A, Lüttg U.							
444	Kluwer, Dordrecht, the Netherlands.							
445	Rahmstorf S, Coumou D. 2011. Increase of extreme events in a warming world,							
446	Proceedings of the National Academy of Sciences, USA 108: 17905–17909.							
447	Ren BZ, Zhang JW, Li X, Fan X, Dong ST, Liu P, Zhao B. 2014. Effects of							
448	waterlogging on the yield and growth of summer maize under field conditions.							
449	Canadian Journal of Plant Science 94: 23-31.							

450	Rosenzweig C, Iglesias A, Yang X, Epstein PR, Chivian E. 2001. Climate change and
451	extreme weather events: Implications for food production, plant diseases, and pests.
452	Global Change & Human Health 2: 90–104.
453	Ruiz M, Taleisnik E. 2013. Field hydroponics assessment of salt tolerance in Cenchrus
454	ciliaris (L.): growth, yield, and maternal effect. Crop Pasture Science 64: 631–639.
455	Singh D, Tsiang M, Rajaratnam B, Diffenbaugh NS. 2013. Precipitation extremes over
456	the continental United States in a transient, high-resolution, ensemble climate
457	model experiment. Journal of Geophysical Research: Atmospheres 118: 7063-
458	7086
459	Snogerup S, Gustafsson M, Von Bothmer R. 1990. Brassica sect. Brassica (Brassicaceae)
460	I. Taxonomy and variation. Willdenowia 19: 271-365.
461	Sun YP, Masabni J, Niu GH. 2015. Simulated seawater flooding reduces the growth of
462	ten vegetables. Hortscience 50: 694-698.
463	Sylvester-Bradley R. 1985. Revision of a code for stages of development in oilseed rape
464	(Brassica napus L.). Annals of Applied Biology 10, 395-400.
465	USDA. 2015. Oilseeds: World Markets and Trade. Available at:
466	http://apps.fas.usda.gov/psdonline/circulars/oilseeds.pdf. [Accessed 9th September
467	2015
468	Van Eck WHJM, van de Steeg HM, Blom CWPM, de Kroon H. 2004. Is tolerance to
469	summer flooding correlated with distribution patterns in river floodplains? A
470	comparative study of 20 grassland species. Oikos 107: 393-405.
471	Vasseur DA, DeLong JP, Gilbert B, et al. 2014. Increased temperature variation poses a
472	greater risk to species than climate warming. Philosophical Transactions of the
473	Royal Society London; Series B 281: 1–8.
474	Vinod KK, Krishnana SG, Babu NN, Nagarajan M, Singh AK. 2013. Improving salt
475	tolerance in rice: looking beyond the conventional. In: Salt stress in plants:
476	signaling, omics and adaptations Ahmed P, Azooz MM, Prasad MNV (Eds),. New
477	York (NY): Springer.
478	Vitousek S, Barnard PL, Fletcher CH, Frazer N, Erikson L, Storlazzi CD. 2017.
479	Doubling of coastal flooding frequency within decades due to sea-level rise.
480	Scientific Reports 7: 1399.

481	Vousdoukas MI, Voukouvalas E, Annunziato A, Giardino A, Feyen L. 2016.
482	Projections of extreme storm surge levels along Europe. Climate Dynamics 47:
483	3171–3190.
484	White AC, Colmer TD, Cawthray GR, Hanley ME. 2014. Variable response of three
485	Trifolium repens ecotypes to soil flooding by seawater. Annals of Botany 114: 347-
486	356.
487	Xu MY, Ma HQ, Zeng L, Cheng Y, Lu GY, Xu JS, Zhang KK, Zou XL. 2015. The
488	effect of waterlogging on yield and seed quality at the early flowering stage in
489	Brassica napus L. Field Crop Research 180: 238-245.
490	Zas R, Cendan C, Sampredo L. 2013. Mediation of seed provisioning in the transmission
491	of environmental maternal effects in maritime pine (Pinus pinaster Aiton).
492	<i>Heredity</i> 111 : 248-255.
493	Zhou WJ, Lin XQ. 1995. Effects of waterlogging at different growth stages on
494	physiological characteristics and seed yield of winter rape (Brassica napus L.).
495	Field Crop Research 44: 103-110.
496	

497 Table 1: The effect of root-zone immersion in deionised (*Dw*) and seawater (*Sw*) on plant reproductive potential (crop yield) of four

498 oilseed rape (*Brassica napus*) cultivars (Cracker, Cubic, Agatha & Astrid) 7 months after plants (growth stage 1.3-1.6) were subject

to transient immersion (24, or 96 hours with a zero hour control). N=4 for all treatment/cultivar combinations except 'Cracker' 96h

500 Dw, and 'Agatha' 24h Dw & 96h Sw where N=3.

501

Treatment		Number of siliqua per plant				Seeds per siliqua				Seed mass (mg)			
		Cracker	Cubic	Astrid	Agatha	Cracker	Cubic	Astrid	Agatha	Cracker	Cubic	Astrid	Agatha
Control	Mean	35.5	29.8	22.0	35.8	13.2	22.4	22.3	20.3	0.04	0.08	0.07	0.06
	SE	8.6	2.8	2.4	4.8	2.7	1.2	1.0	1.2	0.009	0.009	0.004	0.005
24h <i>Dw</i>	Mean	30.3	18.3	22.8	21.7	16.5	21.6	22.5	17.7	0.04	0.06	0.07	0.04
	SE	3.4	2.7	6.2	3.3	0.6	1.0	2.0	1.0	0.005	0.003	0.01	0.004
96h <i>Dw</i>	Mean	40.7	28.0	28.8	27.5	11.4	25.0	20.9	19.9	0.03	0.07	0.06	0.05
	SE	13.2	3.6	2.7	5.0	4.9	0.5	1.1	1.4	0.02	0.004	0.005	0.004
24h Sw	Mean	25.8	38.5	25.5	43.8	13.8	22.8	23.0	20.2	0.04	0.08	0.07	0.07
	SE	6.0	11.1	0.9	5.9	1.6	0.8	0.7	1.5	0.004	0.006	0.008	0.006
96h <i>Sw</i>	Mean	24.3	11.5	17.5	14.0	13.6	19.4	22.5	9.2	0.03	0.04	0.07	0.02
	SE	6.2	3.2	3.0	5.1	3.2	2.0	1.2	4.7	0.01	0.006	0.006	0.01

502

503

504

506 Figure Legends

- 507 Figure 1: The effect of root-zone immersion in deionised (Dw) and seawater (Sw) on
- 508 mean (± SE) total above-ground dry mass of four oilseed rape (*Brassica napus*)
- 509 cultivars (Cracker, Cubic, Agatha & Astrid) 2 months after plants (67-d-old; growth
- stage 1.3-1.6) were subject to transient immersion (24, or 96 hours with a zero hour
- 511 control). N=8 for all treatment/cultivar combinations.
- 512
- 513 Figure 2: The effect of root-zone immersion in deionised (Dw) and seawater (Sw) on
- 514 mean (± SE) seedling dry mass of four oilseed rape (*Brassica napus*) cultivars
- 515 (Cracker, Cubic, Agatha & Astrid) of plants grown from seeds collected from parent
- 516 plants subject to transient immersion (24, or 96 hours with a zero hour control, at
- 517 growth stage 1.3-1.6). N=4 for all treatment/cultivar combinations except 'Agatha'
- 518 24h Dw where N =3 and 'Cracker' 96h Dw, & 'Agatha' 96h Sw where N=2.





523 Fig 2

