

Review

ALK in Neuroblastoma: Biological and Therapeutic Implications

Ricky M. Trigg  and Suzanne D. Turner * 

Division of Cellular and Molecular Pathology, Department of Pathology, University of Cambridge, Cambridge CB2 0QQ, UK; rt473@cam.ac.uk

* Correspondence: sdt36@cam.ac.uk; Tel.: +44-(0)-122-376-2655

Received: 21 March 2018; Accepted: 6 April 2018; Published: 10 April 2018



Abstract: Neuroblastoma (NB) is the most common and deadly solid tumour in children. Despite the development of new treatment options for high-risk NB, over half of patients relapse and five-year survival remains at 40–50%. Therefore, novel treatment strategies aimed at providing long-term disease remission are urgently sought. *ALK*, encoding the anaplastic lymphoma kinase receptor, is altered by gain-of-function point mutations in around 14% of high-risk NB and represents an ideal therapeutic target given its low or absent expression in healthy tissue postnatally. Small-molecule inhibitors of Anaplastic Lymphoma Kinase (ALK) approved in ALK fusion-positive lung cancer are currently undergoing clinical assessment in patients with ALK-mutant NB. Parallel pre-clinical studies are demonstrating the efficacy of ALK inhibitors against common ALK variants in NB; however, a complex picture of therapeutic resistance is emerging. It is anticipated that long-term use of these compounds will require combinatorial targeting of pathways downstream of ALK, functionally-related ‘bypass’ mechanisms and concomitant oncogenic pathways.

Keywords: neuroblastoma; ALK; kinase inhibitors

1. Introduction

Neuroblastoma (NB) is the most common solid extracranial malignancy in children, representing 7–10% of paediatric cancers and accounting for 15% of all paediatric cancer deaths [1–3]. Deriving from the sympathoadrenal lineage of the neural crest, the disease presents at various sites along the sympathoadrenal axis, most commonly in the adrenal medulla or the paraspinal sympathetic ganglia between the neck and pelvis [4]. The median age of diagnosis is around 2 years, with 90% of cases diagnosed by 5 years of age. Foetal and neonatal cases are also well documented [5]. Often termed a ‘clinical enigma’, NB is a complex disease due to its heterogeneous biology and clinical behaviour, ranging from spontaneous regression to treatment-resistant progression, metastasis and death [4]. As such, in addition to a common staging system, NB is further classified into risk categories, each having therapeutic and prognostic significance. Whereas low- and intermediate-risk disease is highly curable, with five-year survival exceeding 90%, high-risk disease, which accounts for around half of all cases at diagnosis, is associated with frequent relapse and a markedly poorer five-year survival of 40–50% [6].

2. Brief Overview of the Genomics of Neuroblastoma

Over the past decade, our understanding of the genomic events underlying the development of NB has significantly improved. However, recent whole-genome sequencing studies have identified a relative scarcity of recurrent somatic alterations, and this has hampered efforts to develop molecularly targeted therapeutics [7–10].

2.1. Familial Neuroblastoma

Around 1–2% of NB cases are inherited in an autosomal-dominant manner within families. Half of familial NB cases are associated with germline, gain-of-function mutations in anaplastic lymphoma kinase (*ALK*) [11,12]. These single-base substitutions cluster in the kinase domain and permit constitutive signaling [13]. A subset of familial cases co-segregate with other neural crest disorders, namely Hirschsprung disease and central congenital hypoventilation syndrome and are associated with loss-of-function mutations in *PHOX2B* [11,14]. This gene encodes a transcription factor, designated a master regulator of neural crest development. Genome-wide sequencing efforts are ongoing to identify the remaining high-penetrance genetic drivers of familial NB.

2.2. Sporadic Neuroblastoma

For sporadic NB, it is believed that several common germline variations, each with low relative risk, act in combination to increase the chances of disease occurrence [15]. A large genome-wide association study (GWAS) employing 720 NB cases and 2128 paediatric healthy controls, identified predisposing single nucleotide polymorphisms (SNPs) in several predicted genes and the gene set was validated and expanded with further studies employing both low- and high-risk NB cases [16]. Loss- or gain-of-function SNPs were identified in the following genes: *BARD1*, *CASC15*, *DDX4*, *DUSP12*, *KIF15*, *HACE1*, *HSD17B12*, *IL31RA*, *LIN28B*, *LMO1*, *NEFL*, and *TP53* [17–28]. In addition, a heritable copy number variation (CNV) at chromosome 1q21.1 encompassing *NBPF53* is associated with NB [17].

The first discovered somatic genomic alteration in NB is high-level amplification (≥ 10 copies per diploid genome) of the transcription factor *MYCN* on chromosome 2p24. *MYCN* amplification occurs in 20% of cases overall, rising to 50% in high-risk tumours [18,19]. Amplified *MYCN* is strongly associated with advanced, aggressive tumours and frequent disease relapse [18,19]. N-MYC is expressed in the developing neural crest and overexpression of N-MYC in neural crest progenitor cells of transgenic mice and zebrafish is sufficient to induce NB development, thus confirming *MYCN* as a driver oncogene [20,21]. *ATRX*, encoding a SWI/SNF chromatin-remodeling ATP-dependent helicase, is mutated in NB at a frequency significantly correlated with age at diagnosis [8–10]. In a Whole Genome Sequencing (WGS) study of 104 patients with metastatic disease, no mutations were found in patients <18 months, whereas point mutations and small in-frame deletions occurred in 17% of patients between 18 months and 12 years and 44% of adolescent and adult patients [10]. Despite its association with poor prognosis, *ATRX* mutations appear to be mutually exclusive of *MYCN* amplification [8,10]. Interestingly, *ATRX* mutations in NB and several other cancer types are associated with a telomerase-independent telomere maintenance mechanism known as alternative lengthening of telomeres (ALT), which is believed to be suppressed by wild-type *ATRX* in ALT-negative tumours [22,23]. *ALK* is the most common somatically mutated gene in NB, with mutations present in around 9% of primary NB tumours and approximately 14% in the high-risk setting [11,12,24–27]. Mutations are often identical to those identified in familial NB and are distributed with even frequency among clinical stages [7–11,24,25,27,28]. Focal amplification of *ALK* is reported in 1–2% of NB cases and is mutually exclusive of point mutation [8,11,12,25,27,29]. *ALK* amplification is almost exclusively associated with co-amplification of *MYCN* [12,29]. *ALK* alterations confer poorer prognosis for tumours in the intermediate- and high-risk categories [24].

3. Anaplastic Lymphoma Kinase (ALK)

3.1. Structure, Function, and Signaling

First discovered as a partner to Nucleophosmin 1 (NPM1) in the NPM-ALK fusion oncoprotein of Anaplastic Large Cell Lymphoma (ALCL), Anaplastic Lymphoma Kinase (ALK) is a receptor tyrosine kinase and a member of the insulin receptor superfamily. Wild-type (full-length) ALK is a 177 kDa, 1620 amino acid protein with an extracellular region containing MAM (meprin, A-5 protein and receptor protein- tyrosine phosphatase mu), low-density lipoprotein class A (LDLa), and glycine-rich

domains, in addition to a transmembrane region and intracellular region containing the kinase domain (Figure 1a). ALK undergoes N-glycosylation to produce a single-chain glycoprotein of 200 kDa at full-length and 140 kDa when truncated by extracellular cleavage, though the functional relevance of this modification is unknown [30,31].

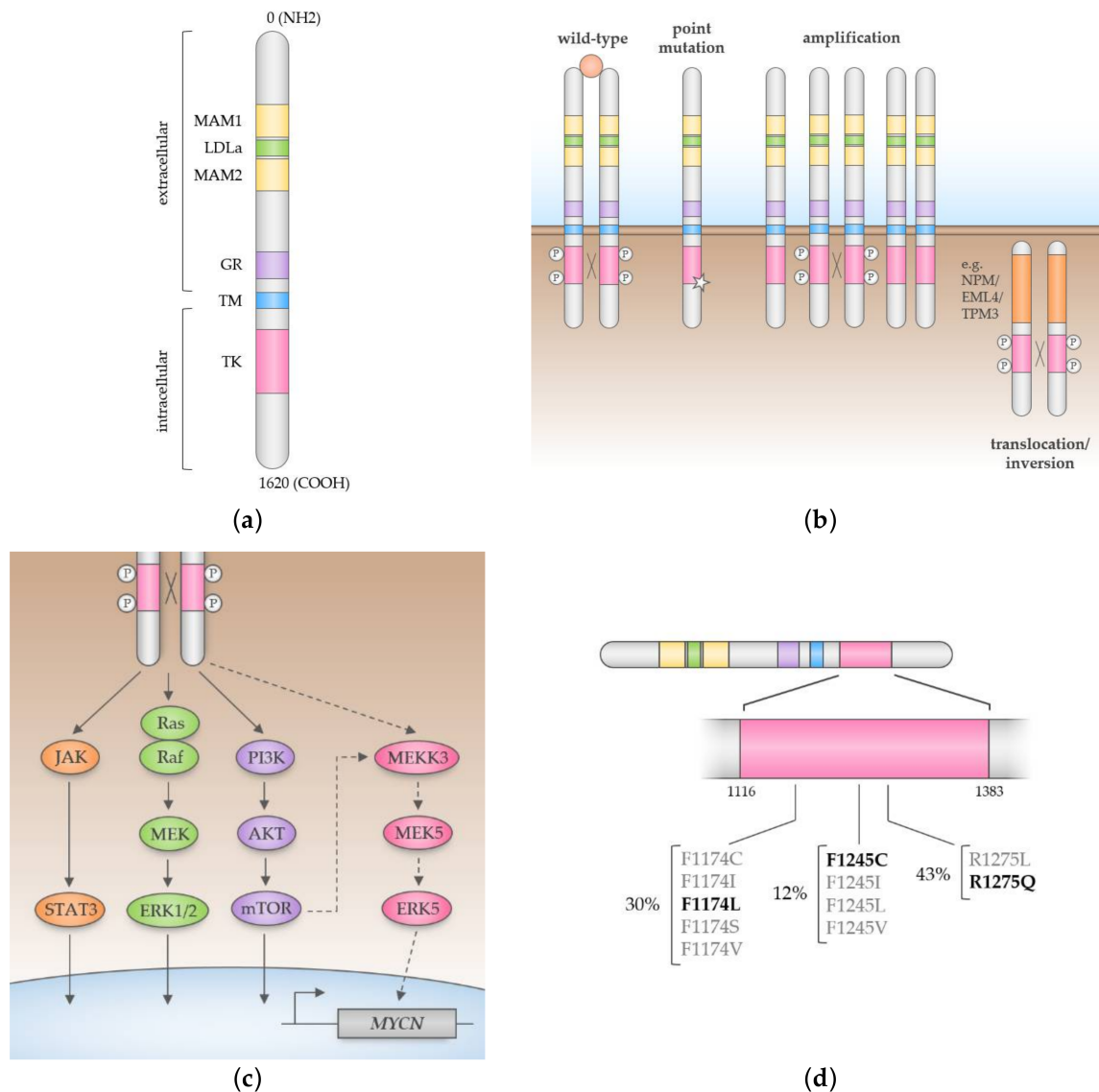


Figure 1. (a) Domain structure of Anaplastic Lymphoma Kinase (ALK). The N-terminal extracellular domain comprises two MAM domains flanked by a low-density lipoprotein class A (LDLa) domain, and a glycine-rich (GR) domain. The C-terminal intracellular region comprises the tyrosine kinase (TK) domain; (b) In the wild-type receptor, ligand-induced dimerisation of the extracellular region permits auto- and transphosphorylation of the kinase domain and subsequent recruitment of signal transducers. Aberrant forms of ALK expressed in cancer are ligand-independent and are caused by point mutations in the kinase domain, gene amplification, or gene fusion; (c) Full-length ALK signals through the Ras/MAPK, PI3K/AKT and JAK/STAT pathways. In neuroblastoma, MYCN expression is activated in a pathway mediated by ALK, PI3K/AKT, MEKK3, MEK5 and ERK5 (dashed lines); (d) In neuroblastoma, gain-of-function mutations cluster in the kinase domain of ALK. Mutations in three key positions—F1174, F1245, and R1275—account for around 85% of ALK mutations in neuroblastoma. The wild-type forms of these residues maintain the kinase in an auto-inhibited conformation. The diagram shows (in bold) the most common mutation at each position.

Until recently, ALK was considered an orphan receptor; despite the identification of the ALK ligands Jeb and HEN-1 in invertebrates, homologous ligands in vertebrates remained elusive. However, several recent studies have identified and confirmed ALKAL1 and ALKAL2 as bona fide ligands of ALK and the related receptor, leukocyte tyrosine kinase (LTK), in mammalian cells and zebrafish [32–36]. Studies in mice have shown that expression of ALK is restricted to discrete tissues of the developing central and peripheral nervous systems and at markedly lower levels in specific cell types of the adult brain [30,37,38]. Of note, high expression of ALK is a frequent observation in paediatric neuroectodermal tumours and neuroblastoma [39]. Thus, aside from its implication in malignancy, ALK likely has a primary role in normal neuronal development.

The biology of ALK is best characterised in lymphoma cells expressing the NPM-ALK fusion oncoprotein. Fusion of the C-terminal region of ALK to the N-terminal region of Nucleophosmin 1 (NPM1) permits constitutive signaling from the kinase domain of ALK, leading to cell cycle progression, migration and evasion of apoptosis [40,41]. The constitutive kinase activity of NPM-ALK and indeed other ALK fusions such as echinoderm microtubule-associated protein-like 4 (EML4-ALK in Non-Small Cell Lung Cancer; NSCLC) is thought to arise from self-dimerisation through N-terminal oligomerisation domains, leading to auto- and transphosphorylation of ALK [41,42]. Other oncogenic mechanisms leading to aberrant ALK activity include point mutations in the kinase domain and gene amplification, as observed in neuroblastoma and NSCLC amongst other cancers (Figure 1b) [11,12,25,43,44]. However, not all mutations in *ALK* lead to ligand-independent or even kinase-active protein and may merely represent passenger mutations [45–47].

In ALK-positive ALCL, NPM-ALK has been shown to activate multiple signaling pathways, including PI3K/AKT, PLC γ , STAT3/STAT5 and RAS/MAPK [43,48–52]. However, signaling through full-length ALK is comparatively less understood since its activating ligands were only identified recently [32–36]. Efforts to delineate signaling through membrane-associated ALK have employed activating monoclonal antibodies or chimeric proteins consisting of the intracellular domain of ALK fused to ligand- or antibody-binding domains [53–59]. More recent studies have used RNA interference and small-molecule inhibitors to block ALK signaling in ALK-mutant NB cell lines and mouse models. These studies have consistently shown that full-length ALK signals through the PI3K/AKT and Ras/MAPK pathways [12,27,59–67] (Figure 1c). In addition, activation of STAT3 has been demonstrated in heterologous cells and ALK-mutant NB cell lines treated with activating ALK antibodies [46,59,68,69].

3.2. ALK in Neuroblastoma

Consistent with other tumours of neural origin, most NB tumours express full-length ALK [39,70]. However, reports of a potential association between levels of ALK expression and prognostic factors such as age, tumour stage, *MYCN* status and DNA ploidy are conflicting and variable in methodology [39,43,71–76]. In NB, single-base missense mutations cluster in key regulatory regions of the kinase domain of ALK and largely promote ligand-independent signaling through disruption of the auto-inhibited conformation of the kinase [13]. These mutations are found in both familial and sporadic NB as previously discussed [11,12,24]. Mutations in three positions—R1275, F1174, and F1245—account for around 85% of ALK mutations in NB and are hotspots for several lower frequency mutations (Figure 1d) [24]. R1275Q is the most common mutation, present in 45% of familial cases and a third of sporadic cases, whereas F1174 and F1245 mutants are exclusively found in sporadic disease at frequencies of around 30% and 12%, respectively [24,77]. ALK mutations are distributed with even frequency amongst clinical stages [7–11,24,25,27,28]. However, a recent study of 1596 diagnostic NB samples found that ALK mutations were associated with poorer survival in high- and intermediate-risk disease [24]. ALK mutations are correlated with *MYCN* amplification and studies in mouse and zebrafish models have demonstrated the cooperative activity of these two oncogenes in driving the development of NB [21,78]. Interestingly, wild-type and mutant forms of ALK have

been shown to induce transcription of *MYC* in NB and NSCLC cell lines; this may explain the poor prognosis of NB patients harbouring both *ALK* mutations and *MYCN* amplification [64,79].

A less common mechanism of *ALK* activation in NB, found in 2–3% cases, involves gene amplification, leading to increased protein expression and constitutive kinase activity [43,61,80]. *ALK* is almost exclusively co-amplified with *MYCN*, consistent with the proximity of these genes at 2p23-24 and therefore tumours harbouring *ALK* amplification tend to afford a poor prognosis [24,27,29]. In contrast, mutation and amplification of *ALK* within the same tumour is rare in NB [81]. Other infrequent mechanisms of ligand-independent *ALK* signaling include translocations and large deletions resulting in truncation of the extracellular region of *ALK* [69,82,83]. Interestingly, activation of downstream targets of *ALK* has also been observed in NB tumours without *ALK* mutation or amplification; it appears that the wild-type receptor requires a critical level of expression to permit constitutive signaling [26,43,76].

Recent genome sequencing analyses of matched primary and relapsed NB tumours have shown an increased frequency of *ALK* mutations at relapse [84–86]. Some mutations were found to be present at very low allele frequencies in primary tumours before undergoing clonal expansion at relapse, whereas others were confirmed to arise *de novo*. In a deep sequencing study of primary NB, *ALK* F1174 and R1275 mutations were identified in 10% (27/277) of patients at allele frequencies ranging from 0.56% to 40.41%, half of which would have been undetectable by Sanger sequencing [81]. These observations highlight the role of *ALK* as a driver oncogene in both primary and relapsed NB.

4. Therapeutic Targeting of *ALK* in NB

4.1. Preclinical Studies

ALK is an ideal tumour antigen for targeted inhibition, given its restricted distribution in normal tissue and frequent expression in NB [39,87]. Indeed, *ALK* is amenable to pharmacological inhibition with small-molecule, ATP-competitive inhibitors of the tyrosine kinase domain. The efficacy of *ALK* inhibitors in cells harbouring point mutations in *ALK* was first demonstrated in a screen of 602 cancer cell lines with TAE684, a highly selective compound previously shown to inhibit the NPM-*ALK* fusion oncoprotein in ALCL [62,88]. TAE684 suppressed growth of cells expressing both *ALK* fusions and point mutations, with the latter enriched in NB cell lines. Importantly, among the most TAE684-sensitive NB cell lines were KELLY and NB-1, harbouring the F1174L *ALK* mutation and *ALK* amplification, respectively [62]. Later studies characterised the differential sensitivity of *ALK* mutations, predominantly F1174L and R1275Q, to TAE684 and the *ALK*/MET/ROS1 inhibitor crizotinib (PF-02341066) [24,27,63,68,89]. A common finding among these studies was the relative resistance of *ALK* F1174L to crizotinib. In one study, crizotinib induced complete and sustained tumour regression in a panel of R1275Q-mutant xenografts while providing only partial growth inhibition to those driven by F1174L [60]. Though structurally similar to the wild-type residue, biochemical analysis showed the leucine substitution to increase ATP-binding affinity of the kinase domain [24,90]. Reflecting the high frequency of *ALK* F1174L in primary NB, several combinatorial approaches have been taken to overcome crizotinib resistance or enhance its anti-tumour activity in cell lines and xenografts driven by this *ALK* mutant. These include the use of chemotherapeutic agents; small-molecule inhibitors of signaling pathways downstream of *ALK*, such as mTOR and ERK5; and therapeutic antibodies against *ALK* [66,67,78,87,91,92].

Recent studies have evaluated the efficacy of structurally distinct second-generation *ALK* inhibitors, such as ceritinib (LDK-378), alectinib (CH5424802) and brigatinib (AP26113). Although these drugs were developed primarily to target the EML4-*ALK* fusion in NSCLC patients with crizotinib resistance, studies in *ALK*-driven NB cell lines and mouse models have demonstrated their superior potency over crizotinib. Ceritinib is a structural derivative of TAE684, inhibiting *ALK* and to a lesser extent the Insulin-like Growth Factor 1 Receptor (IGFR1). While ceritinib shows efficacy in crizotinib-naïve and resistant NSCLC and displays a 20-fold increased potency over crizotinib in

enzymatic assays, it is only partially active against the F1174L mutant [93]. Several studies have employed combinatorial targeting strategies to enhance the efficacy of ceritinib and overcome resistance in ALK F1174L-mutant NB. For example, two independent studies associated ceritinib and TAE684 resistance with activation of AXL, a receptor tyrosine kinase previously implicated in the metastatic potential of NB [94–96]. Small-molecule inhibitors of AXL were shown to dramatically increase the activity of ALK inhibitors against F1174L [94]. In another study, a drug screen was conducted to identify synergistic partners of ceritinib. The combination of ceritinib and the CDK4/6 inhibitor ribociclib was found to promote greater cytotoxicity in NB cells with mutant ALK relative to wild-type cells. In ALK F1174L and F1245C-mutant cell lines and patient-derived xenografts, whereas treatment with either agent alone delayed tumour growth, the combination of ceritinib and ribociclib led to a complete and sustained tumour regression [97]. Similarly, the combination of ceritinib and an MDM2 inhibitor was found to be synergistic in vitro and in NB xenografts driven by ALK mutations or gene amplification [98]. Alectinib and brigatinib have both shown efficacy in ALK F1174L-mutant NB cell lines and xenografts and may therefore overcome de novo resistance to crizotinib [99–101]. In Ba/F3 cell lines engineered to express ALK, brigatinib has shown activity against a wide range of ALK mutations, including many secondary mutations that mediate resistance to other ALK inhibitors and showed overall superior potency to crizotinib, ceritinib, and alectinib [102]. However, the relative activity of brigatinib and alectinib in NB driven by different ALK mutants and other aberrations remains to be assessed.

Lorlatinib (PF-6463922) is a third-generation inhibitor of ALK and ROS1, effective against all known ALK inhibitor-resistant mutants in NSCLC [103,104]. It exhibits remarkably improved potency over crizotinib towards three of the most common ALK mutants in NB—F1174L, F1245C and R1275Q, among others, as shown by in vitro kinase activity [104,105]. The compound was found to have low ED₅₀ values relative to crizotinib across NB cell lines of differing ALK status [104]. Most encouraging was a ~30-fold decrease in ED₅₀ for F1174L- and F1245C-mutant cells, thus demonstrating its ability to overcome de novo crizotinib and ceritinib resistance conferred by these mutations [104]. In the same study, lorlatinib induced rapid and complete regression of NB cell line and patient-derived xenografts driven by all three ALK mutants, which was sustained for the nine-week duration of treatment [104]. Moreover, despite withdrawal of the compound after complete response, tumours did not return for at least four weeks. In another study, lorlatinib was found to significantly reduce growth of tumours in the ALK F1174L/MYCN-driven genetically engineered mouse model of aggressive NB, which did not respond to crizotinib [105]. Although these studies have clearly demonstrated the superior potency of lorlatinib to previous ALK inhibitors in both inhibitor-naïve and -resistant NB, acquired resistance to lorlatinib has been reported in the context of ALK fusion-positive NSCLC [106]. A recent case study reported a patient who relapsed on crizotinib therapy due to a mutation in the ALK kinase domain (C1156Y) of the EML4-ALK fusion. Though the patient did not respond to second-generation inhibitors, she responded to lorlatinib for eight months, after which she relapsed and was found to have an additional mutation (L1198F) on the same EML4-ALK allele. Paradoxically, L1198F enhanced binding to crizotinib and negated the effect of the resistance-conferring C1156Y mutation and the patient was resensitized to crizotinib [106]. This scenario exemplifies the complex relationships between the modified structures imparted by mutations of the ALK kinase domain and the differential binding of ALK inhibitors to such structures, as highlighted by crystallographic studies [13,90].

4.2. Clinical Trials

Crizotinib was the first ALK inhibitor to undergo clinical assessment and showed encouraging results in early trials for EML4-ALK-positive NSCLC (NCT00585195; NCT00932451) [107–109]. Two phase III trials (NCT00932893; NCT01154140) confirmed the sustained, superior efficacy of crizotinib over standard first-line chemotherapy, leading to its accelerated FDA approval in 2011 [110,111]. With the identification of ALK as a common driver of NB in 2008, the results of early crizotinib trials in NSCLC provided the rationale for further evaluation in other ALK-driven cancers such as ALCL and ALK-positive NB. In 2009, the Children's Oncology Group (COG) initiated the ADVL0912 phase I–II trial of crizotinib in paediatric

patients with relapsed or refractory solid tumours or ALCL (NCT00939770) (Table 1) [112]. Recently published results confirmed the pharmacokinetic profile of crizotinib in children and adults to be similar and efforts to delineate the response of crizotinib to specific ALK alterations are ongoing [113]. However, early results from NB patients in this trial were discouraging; of eleven patients with known ALK mutations, only one had a complete response and two had stable disease [112]. This is consistent with the differential sensitivity of ALK mutants to crizotinib, since 3/7 tumours from patients with progressive disease harboured the resistance-conferring ALK F1174L mutation. A Pfizer phase Ib basket trial of crizotinib was launched in 2011 for patients aged ≥ 15 years with any ALK-positive malignancy other than NSCLC (NCT01121588). To enable safe and tolerable integration of crizotinib into chemotherapeutic regimens, a COG phase I trial of crizotinib with combination chemotherapy (cyclophosphamide and topotecan or doxorubicin, dexrazoxane and vincristine) was initiated in 2013 for patients with high-risk NB and ALCL (NCT01606878). Encouraging results from this trial provided the rationale for a COG phase III trial evaluating the addition of crizotinib to standard therapy in high-risk NB patients with ALK mutations (NCT03126916).

Table 1. Clinical trials evaluating ALK inhibitors in neuroblastoma.

Trial Identifier	Sponsor	Phase	Disease Eligibility	Study Drug(s)	Start Date	References
NCT00939770	COG	I/II	r/r solid tumours and ALCL	crizotinib	September 2009	[112–114]
NCT01121588	Pfizer	I	all ALK-positive tumours except NSCLC; ≥ 15 years	crizotinib	March 2011	[115]
NCT01606878	COG	I	r/r solid tumours and ALCL	crizotinib + chemotherapy	March 2013	[116]
NCT01742286	Novartis	I	all r/r ALK-positive tumours	ceritinib	August 2013	[117]
NCT02650401	Ignitya	I/Ib	r/r solid tumours	entrectinib	December 2015	[118]
NCT02780128	Y.P. Mossé	I	r/r NB	ceritinib + ribociclib	July 2016	[119]
NCT03213652	NCI	II	r/r ALK/ROS1-positive solid tumours, NHL, histiocytic disorders	ensartinib	July 2017	[120,121]
NCT03107988	NANT Consortium	I	high-risk NB	lorlatinib +/- chemotherapy	September 2017	[122]
NCT03126916	COG	III	high-risk NB and ganglioneuroblastoma	crizotinib + standard therapy	December 2017	[123]

ALCL: anaplastic large cell lymphoma; COG: Children's Oncology Group; NANT: New Approaches to Neuroblastoma Therapy; NB: neuroblastoma; NCI: National Cancer Institute; NHL: non-Hodgkin's lymphoma; r/r: relapsed or refractory.

Other trials are evaluating the safety and efficacy of second- and third-generation ALK inhibitors alone or in combination with chemotherapy and molecularly targeted compounds in patients with NB and other ALK-driven cancers (Table 1). A phase I study of ceritinib showed an overall response rate of 56% in ALK fusion-positive NSCLC patients who had progressed on crizotinib (NCT01283516) and the compound subsequently received FDA approval for this indication [124,125]. Ceritinib is currently undergoing phase I assessment as a monotherapy in relapsed or refractory ALK-positive paediatric cancers including NB (NCT01742286). In addition, the NEPENTHE (Next Generation Personalized Neuroblastoma Therapy) clinical study (NCT02780128) initiated in 2016 is recruiting patients with relapsed or refractory NB into treatment arms based on actionable genetic alterations determined by deep sequencing. Patients with ALK mutations are treated with the synergistic combination of ceritinib and ribociclib. Similarly, the National Cancer Institute (NCI)-COG Pediatric MATCH (Molecular Analysis for Therapy Choice) is a large phase II study that involves the stratification of patients into molecularly targeted treatments based on genetic profiling (NCT03155620) [120]. In this basket trial, the ALK inhibitor ensartinib is under assessment in patients with relapsed or refractory NB among other solid tumours, non-Hodgkin's lymphoma and histiocytic disorders with genetic alteration of ALK or ROS1 (NCT03213652). A phase I trial is assessing the pan-TRK, ROS1 and ALK inhibitor entrectinib in paediatric patients with relapsed or refractory solid and Central Nervous

System (CNS) tumours with and without TRK, ROS1 and ALK fusions (NCT02650401). Recently, the NANT (New Approaches to Neuroblastoma Therapy) consortium opened a phase I trial of lorlatinib in combination with chemotherapy (cyclophosphamide and topotecan) in patients with high-risk NB (NCT03107988).

5. ALK Inhibitor Resistance: Lessons from ALK Fusion-Positive NSCLC

Studies in ALK fusion-positive NSCLC have shown that resistance to ALK inhibition is largely acquired during therapy and involves both on-target mechanisms, such as secondary mutation and amplification of ALK and off-target mechanisms such as activation of 'bypass' signaling pathways [126]. For example, resistance to crizotinib has been associated with mutation of EGFR and KRAS, activation of the IGFR1 pathway, amplification of KIT, upregulation of EGFR ligands, activation of ErbB family RTKs by phosphorylation and autophagy [127–131]. Despite the ability of second- and third-generation ALK inhibitors to overcome crizotinib resistance in ALK fusion-positive NSCLC, resistance to these compounds has been shown to arise through secondary point mutations in the ALK kinase domain [93,132–135]. Thus, it is becoming apparent that therapeutic resistance to ALK inhibition is inevitable for all current compounds.

In NB, our understanding of resistance to ALK inhibition is limited to pre-clinical studies of short treatment duration and have therefore focused primarily on the de novo resistance of ALK F1174L to crizotinib and ceritinib [60,93,94,97,98]. As previously discussed, targeting of bypass signaling pathways such as AXL and oncoproteins such as cyclin-dependent kinases and MDM2 has been shown to enhance the efficacy of ALK inhibition and overcome de novo resistance [94,97,98]. However, proactive investigation into acquired resistance mechanisms to ALK inhibition in NB is warranted and may lead to the identification of targets that are druggable with existing therapeutic strategies. Ultimately, the landscape of ALK inhibitor resistance mechanisms in NB will only become clear following the completion of current clinical trials, but is likely to involve both on- and off-target mechanisms as observed in NSCLC.

6. Conclusions

Preclinical studies with ALK-driven NB cell lines and mouse models have clearly demonstrated the potential of ALK inhibition with existing small-molecule inhibitors. However, therapeutic resistance is an innate concern with molecularly targeted approaches and has been shown to arise with all ALK inhibitors undergoing clinical assessment in NB, irrespective of on-target potency. The preclinical studies described in this review highlight how sensitivity to ALK inhibitors is defined by complex relationships between mutation-induced changes in the structure of the ALK tyrosine kinase domain and the differential binding profiles of ALK inhibitors. The true potential of ALK inhibitors in NB will be determined over the next few years after completion of current clinical studies, though it is anticipated that long-term use of these compounds will require combinatorial targeting of pathways downstream of ALK, functionally-related 'bypass' mechanisms and concomitant oncogenic pathways. Further work should focus on investigation of putative resistance mechanisms to the more potent ALK inhibitors and novel strategies to target such mechanisms.

Acknowledgments: Ricky M. Trigg and Suzanne D. Turner are in receipt of a project grant from Children with Cancer UK and are members of the European Research Initiative for ALK-Related Malignancies (www.erialcl.net). Suzanne D. Turner is in receipt of an EU Horizon 2020 Marie Curie Innovative Training Network Grant (#675712).

Author Contributions: Ricky M. Trigg and Suzanne D. Turner contributed equally to the writing of this review.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Brodeur, G.M. Neuroblastoma: Biological insights into a clinical enigma. *Nat. Rev. Cancer* **2003**, *3*, 203–216. [[CrossRef](#)] [[PubMed](#)]

2. Smith, M.A.; Seibel, N.L.; Altekruze, S.F.; Ries, L.A.; Melbert, D.L.; O'Leary, M.; Smith, F.O.; Reaman, G.H. Outcomes for children and adolescents with cancer: Challenges for the twenty-first century. *J. Clin. Oncol.* **2010**, *28*, 2625–2634. [[CrossRef](#)] [[PubMed](#)]
3. Spix, C.; Pastore, G.; Sankila, R.; Stiller, C.A.; Steliarova-Foucher, E. Neuroblastoma incidence and survival in european children (1978–1997): Report from the automated childhood cancer information system project. *Eur. J. Cancer* **2006**, *42*, 2081–2091. [[CrossRef](#)] [[PubMed](#)]
4. Maris, J.M.; Hogarty, M.D.; Bagatell, R.; Cohn, S.L. Neuroblastoma. *Lancet* **2007**, *369*, 2106–2120. [[CrossRef](#)]
5. Isaacs, H., Jr. Fetal and neonatal neuroblastoma: Retrospective review of 271 cases. *Fetal. Pediatr. Pathol.* **2007**, *26*, 177–184. [[CrossRef](#)] [[PubMed](#)]
6. Park, J.R.; Bagatell, R.; London, W.B.; Maris, J.M.; Cohn, S.L.; Mattay, K.K.; Hogarty, M.; COG Brain Tumor Committee. Children's oncology group's 2013 blueprint for research: Neuroblastoma. *Pediatr. Blood Cancer* **2013**, *60*, 985–993. [[CrossRef](#)] [[PubMed](#)]
7. Sausen, M.; Leary, R.J.; Jones, S.; Wu, J.; Reynolds, C.P.; Liu, X.; Blackford, A.; Parmigiani, G.; Diaz, L.A., Jr.; Papadopoulos, N.; et al. Integrated genomic analyses identify arid1a and arid1b alterations in the childhood cancer neuroblastoma. *Nat. Genet.* **2013**, *45*, 12–17. [[CrossRef](#)] [[PubMed](#)]
8. Pugh, T.J.; Morozova, O.; Attiyeh, E.F.; Asgharzadeh, S.; Wei, J.S.; Auclair, D.; Carter, S.L.; Cibulskis, K.; Hanna, M.; Kiezun, A.; et al. The genetic landscape of high-risk neuroblastoma. *Nat. Genet.* **2013**, *45*, 279–284. [[CrossRef](#)] [[PubMed](#)]
9. Molenaar, J.J.; Koster, J.; Zwijnenburg, D.A.; van Sluis, P.; Valentijn, L.J.; van der Ploeg, I.; Hamdi, M.; van Nes, J.; Westerman, B.A.; van Arkel, J.; et al. Sequencing of neuroblastoma identifies chromothripsis and defects in neurogenesis genes. *Nature* **2012**, *483*, 589–593. [[CrossRef](#)] [[PubMed](#)]
10. Cheung, N.K.; Zhang, J.; Lu, C.; Parker, M.; Bahrami, A.; Tickoo, S.K.; Heguy, A.; Pappo, A.S.; Federico, S.; Dalton, J.; et al. Association of age at diagnosis and genetic mutations in patients with neuroblastoma. *JAMA* **2012**, *307*, 1062–1071. [[CrossRef](#)] [[PubMed](#)]
11. Mosse, Y.P.; Laudenslager, M.; Longo, L.; Cole, K.A.; Wood, A.; Attiyeh, E.F.; Laquaglia, M.J.; Sennett, R.; Lynch, J.E.; Perri, P.; et al. Identification of ALK as a major familial neuroblastoma predisposition gene. *Nature* **2008**, *455*, 930–935. [[CrossRef](#)] [[PubMed](#)]
12. Janoueix-Lerosey, I.; Lequin, D.; Brugieres, L.; Ribeiro, A.; de Pontual, L.; Combaret, V.; Raynal, V.; Puisieux, A.; Schleiermacher, G.; Pierron, G.; et al. Somatic and germline activating mutations of the ALK kinase receptor in neuroblastoma. *Nature* **2008**, *455*, 967–970. [[CrossRef](#)] [[PubMed](#)]
13. Lee, C.C.; Jia, Y.; Li, N.; Sun, X.; Ng, K.; Ambing, E.; Gao, M.Y.; Hua, S.; Chen, C.; Kim, S.; et al. Crystal structure of the ALK (anaplastic lymphoma kinase) catalytic domain. *Biochem. J.* **2010**, *430*, 425–437. [[CrossRef](#)] [[PubMed](#)]
14. Trochet, D.; Bourdeaut, F.; Janoueix-Lerosey, I.; Deville, A.; de Pontual, L.; Schleiermacher, G.; Coze, C.; Philip, N.; Frebourg, T.; Munnich, A.; et al. Germline mutations of the paired-like homeobox 2b (phox2b) gene in neuroblastoma. *Am. J. Hum. Genet.* **2004**, *74*, 761–764. [[CrossRef](#)] [[PubMed](#)]
15. Tolbert, V.P.; Coggins, G.E.; Maris, J.M. Genetic susceptibility to neuroblastoma. *Curr. Opin. Genet. Dev.* **2017**, *42*, 81–90. [[CrossRef](#)] [[PubMed](#)]
16. Maris, J.M.; Mosse, Y.P.; Bradfield, J.P.; Hou, C.; Monni, S.; Scott, R.H.; Asgharzadeh, S.; Attiyeh, E.F.; Diskin, S.J.; Laudenslager, M.; et al. Chromosome 6p22 locus associated with clinically aggressive neuroblastoma. *N. Engl. J. Med.* **2008**, *358*, 2585–2593. [[CrossRef](#)] [[PubMed](#)]
17. Diskin, S.J.; Hou, C.; Glessner, J.T.; Attiyeh, E.F.; Laudenslager, M.; Bosse, K.; Cole, K.; Mosse, Y.P.; Wood, A.; Lynch, J.E.; et al. Copy number variation at 1q21.1 associated with neuroblastoma. *Nature* **2009**, *459*, 987–991. [[CrossRef](#)] [[PubMed](#)]
18. Brodeur, G.M.; Seeger, R.C.; Schwab, M.; Varmus, H.E.; Bishop, J.M. Amplification of N-myc in untreated human neuroblastomas correlates with advanced disease stage. *Science* **1984**, *224*, 1121–1124. [[CrossRef](#)] [[PubMed](#)]
19. Seeger, R.C.; Brodeur, G.M.; Sather, H.; Dalton, A.; Siegel, S.E.; Wong, K.Y.; Hammond, D. Association of multiple copies of the N-myc oncogene with rapid progression of neuroblastomas. *N. Engl. J. Med.* **1985**, *313*, 1111–1116. [[CrossRef](#)] [[PubMed](#)]
20. Weiss, W.A.; Aldape, K.; Mohapatra, G.; Feuerstein, B.G.; Bishop, J.M. Targeted expression of MYCN causes neuroblastoma in transgenic mice. *EMBO J.* **1997**, *16*, 2985–2995. [[CrossRef](#)] [[PubMed](#)]

21. Zhu, S.; Lee, J.S.; Guo, F.; Shin, J.; Perez-Atayde, A.R.; Kutok, J.L.; Rodig, S.J.; Neuberg, D.S.; Helman, D.; Feng, H.; et al. Activated ALK collaborates with MYCN in neuroblastoma pathogenesis. *Cancer Cell* **2012**, *21*, 362–373. [[CrossRef](#)] [[PubMed](#)]
22. Bower, K.; Napier, C.E.; Cole, S.L.; Dagg, R.A.; Lau, L.M.; Duncan, E.L.; Moy, E.L.; Reddel, R.R. Loss of wild-type ATRX expression in somatic cell hybrids segregates with activation of alternative lengthening of telomeres. *PLoS ONE* **2012**, *7*, e50062. [[CrossRef](#)] [[PubMed](#)]
23. Clynes, D.; Jelinska, C.; Xella, B.; Ayyub, H.; Scott, C.; Mitson, M.; Taylor, S.; Higgs, D.R.; Gibbons, R.J. Suppression of the alternative lengthening of telomere pathway by the chromatin remodelling factor ATRX. *Nat. Commun.* **2015**, *6*, 7538. [[CrossRef](#)] [[PubMed](#)]
24. Bresler, S.C.; Weiser, D.A.; Huwe, P.J.; Park, J.H.; Krytska, K.; Ryles, H.; Laudenslager, M.; Rappaport, E.F.; Wood, A.C.; McGrady, P.W.; et al. ALK mutations confer differential oncogenic activation and sensitivity to ALK inhibition therapy in neuroblastoma. *Cancer Cell* **2014**, *26*, 682–694. [[CrossRef](#)] [[PubMed](#)]
25. Chen, Y.; Takita, J.; Choi, Y.L.; Kato, M.; Ohira, M.; Sanada, M.; Wang, L.; Soda, M.; Kikuchi, A.; Igarashi, T.; et al. Oncogenic mutations of ALK kinase in neuroblastoma. *Nature* **2008**, *455*, 971–974. [[CrossRef](#)] [[PubMed](#)]
26. Caren, H.; Abel, F.; Kogner, P.; Martinsson, T. High incidence of DNA mutations and gene amplifications of the ALK gene in advanced sporadic neuroblastoma tumours. *Biochem. J.* **2008**, *416*, 153–159. [[CrossRef](#)] [[PubMed](#)]
27. George, R.E.; Sanda, T.; Hanna, M.; Frohling, S.; Luther, W., 2nd; Zhang, J.; Ahn, Y.; Zhou, W.; London, W.B.; McGrady, P.; et al. Activating mutations in ALK provide a therapeutic target in neuroblastoma. *Nature* **2008**, *455*, 975–978. [[CrossRef](#)] [[PubMed](#)]
28. Carpenter, E.L.; Mosse, Y.P. Targeting ALK in neuroblastoma—Preclinical and clinical advancements. *Nat. Rev. Clin. Oncol.* **2012**, *9*, 391–399. [[CrossRef](#)] [[PubMed](#)]
29. De Brouwer, S.; De Preter, K.; Kumps, C.; Zabrocki, P.; Porcu, M.; Westerhout, E.M.; Lakeman, A.; Vandesompele, J.; Hoebeek, J.; Van Maerken, T.; et al. Meta-analysis of neuroblastomas reveals a skewed ALK mutation spectrum in tumors with MYCN amplification. *Clin. Cancer Res.* **2010**, *16*, 4353–4362. [[CrossRef](#)] [[PubMed](#)]
30. Iwahara, T.; Fujimoto, J.; Wen, D.; Cupples, R.; Bucay, N.; Arakawa, T.; Mori, S.; Ratzkin, B.; Yamamoto, T. Molecular characterization of ALK, a receptor tyrosine kinase expressed specifically in the nervous system. *Oncogene* **1997**, *14*, 439–449. [[CrossRef](#)] [[PubMed](#)]
31. Degoutin, J.; Brunet-de Carvalho, N.; Cifuentes-Diaz, C.; Vigny, M. ALK (anaplastic lymphoma kinase) expression in DRG neurons and its involvement in neuron-Schwann cells interaction. *Eur. J. Neurosci.* **2009**, *29*, 275–286. [[CrossRef](#)] [[PubMed](#)]
32. Guan, J.; Umapathy, G.; Yamazaki, Y.; Wolfstetter, G.; Mendoza, P.; Pfeifer, K.; Mohammed, A.; Hugosson, F.; Zhang, H.; Hsu, A.W.; et al. Fam150a and fam150b are activating ligands for anaplastic lymphoma kinase. *Elife* **2015**, *4*, e09811. [[CrossRef](#)] [[PubMed](#)]
33. Reshetnyak, A.V.; Murray, P.B.; Shi, X.; Mo, E.S.; Mohanty, J.; Tome, F.; Bai, H.; Gunel, M.; Lax, I.; Schlessinger, J. Augmentor α and β (FAM150) are ligands of the receptor tyrosine kinases ALK and LTK: Hierarchy and specificity of ligand-receptor interactions. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 15862–15867. [[CrossRef](#)] [[PubMed](#)]
34. Zhang, H.; Pao, L.I.; Zhou, A.; Brace, A.D.; Halenbeck, R.; Hsu, A.W.; Bray, T.L.; Hestir, K.; Bosch, E.; Lee, E.; et al. Deorphanization of the human leukocyte tyrosine kinase (LTK) receptor by a signaling screen of the extracellular proteome. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 15741–15745. [[CrossRef](#)] [[PubMed](#)]
35. Mo, E.S.; Cheng, Q.; Reshetnyak, A.V.; Schlessinger, J.; Nicoli, S. ALK and LTK ligands are essential for iridophore development in zebrafish mediated by the receptor tyrosine kinase LTK. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 12027–12032. [[CrossRef](#)] [[PubMed](#)]
36. Fadeev, A.; Mendoza-Garcia, P.; Irion, U.; Guan, J.; Pfeifer, K.; Wiessner, S.; Serluca, F.; Singh, A.P.; Nusslein-Volhard, C.; Palmer, R.H. ALKals are in vivo ligands for ALK family receptor tyrosine kinases in the neural crest and derived cells. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E630–E638. [[CrossRef](#)] [[PubMed](#)]
37. Vernersson, E.; Khoo, N.K.; Henriksson, M.L.; Roos, G.; Palmer, R.H.; Hallberg, B. Characterization of the expression of the ALK receptor tyrosine kinase in mice. *Gene Expr. Patterns* **2006**, *6*, 448–461. [[CrossRef](#)] [[PubMed](#)]

38. Morris, S.W.; Naeve, C.; Mathew, P.; James, P.L.; Kirstein, M.N.; Cui, X.; Witte, D.P. ALK, the chromosome 2 gene locus altered by the t(2;5) in non-hodgkin's lymphoma, encodes a novel neural receptor tyrosine kinase that is highly related to leukocyte tyrosine kinase (LTK). *Oncogene* **1997**, *14*, 2175–2188. [[CrossRef](#)] [[PubMed](#)]
39. Lamant, L.; Pulford, K.; Bischof, D.; Morris, S.W.; Mason, D.Y.; Delsol, G.; Mariame, B. Expression of the ALK tyrosine kinase gene in neuroblastoma. *Am. J. Pathol.* **2000**, *156*, 1711–1721. [[CrossRef](#)]
40. Fujimoto, J.; Shiota, M.; Iwahara, T.; Seki, N.; Satoh, H.; Mori, S.; Yamamoto, T. Characterization of the transforming activity of p80, a hyperphosphorylated protein in a ki-1 lymphoma cell line with chromosomal translocation t(2;5). *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 4181–4186. [[CrossRef](#)] [[PubMed](#)]
41. Bischof, D.; Pulford, K.; Mason, D.Y.; Morris, S.W. Role of the nucleophosmin (NPM) portion of the non-hodgkin's lymphoma-associated NPM-anaplastic lymphoma kinase fusion protein in oncogenesis. *Mol. Cell. Biol.* **1997**, *17*, 2312–2325. [[CrossRef](#)] [[PubMed](#)]
42. Soda, M.; Choi, Y.L.; Enomoto, M.; Takada, S.; Yamashita, Y.; Ishikawa, S.; Fujiwara, S.; Watanabe, H.; Kurashina, K.; Hatanaka, H.; et al. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature* **2007**, *448*, 561–566. [[CrossRef](#)] [[PubMed](#)]
43. Osajima-Hakomori, Y.; Miyake, I.; Ohira, M.; Nakagawara, A.; Nakagawa, A.; Sakai, R. Biological role of anaplastic lymphoma kinase in neuroblastoma. *Am. J. Pathol.* **2005**, *167*, 213–222. [[CrossRef](#)]
44. Salido, M.; Pijuan, L.; Martinez-Aviles, L.; Galvan, A.B.; Canadas, I.; Rovira, A.; Zanui, M.; Martinez, A.; Longaron, R.; Sole, F.; et al. Increased ALK gene copy number and amplification are frequent in non-small cell lung cancer. *J. Thorac. Oncol.* **2011**, *6*, 21–27. [[CrossRef](#)] [[PubMed](#)]
45. McDuff, F.K.; Lim, S.V.; Dalbay, M.; Turner, S.D. Assessment of the transforming potential of novel anaplastic lymphoma kinase point mutants. *Mol. Carcinog.* **2013**, *52*, 79–83. [[CrossRef](#)] [[PubMed](#)]
46. Chand, D.; Yamazaki, Y.; Ruuth, K.; Schonherr, C.; Martinsson, T.; Kogner, P.; Attiyeh, E.F.; Maris, J.; Morozova, O.; Marra, M.A.; et al. Cell culture and drosophila model systems define three classes of anaplastic lymphoma kinase mutations in neuroblastoma. *Dis. Model. Mech.* **2013**, *6*, 373–382. [[CrossRef](#)] [[PubMed](#)]
47. Schonherr, C.; Ruuth, K.; Eriksson, T.; Yamazaki, Y.; Ottmann, C.; Combaret, V.; Vigny, M.; Kamaraj, S.; Palmer, R.H.; Hallberg, B. The neuroblastoma ALK(i1250t) mutation is a kinase-dead RTK in vitro and in vivo. *Transl. Oncol.* **2011**, *4*, 258–265. [[CrossRef](#)] [[PubMed](#)]
48. Bai, R.Y.; Dieter, P.; Peschel, C.; Morris, S.W.; Duyster, J. Nucleophosmin-anaplastic lymphoma kinase of large-cell anaplastic lymphoma is a constitutively active tyrosine kinase that utilizes phospholipase c- γ to mediate its mitogenicity. *Mol. Cell. Biol.* **1998**, *18*, 6951–6961. [[CrossRef](#)] [[PubMed](#)]
49. Bai, R.Y.; Ouyang, T.; Miething, C.; Morris, S.W.; Peschel, C.; Duyster, J. Nucleophosmin-anaplastic lymphoma kinase associated with anaplastic large-cell lymphoma activates the phosphatidylinositol 3-kinase/Akt antiapoptotic signaling pathway. *Blood* **2000**, *96*, 4319–4327. [[PubMed](#)]
50. Nieborowska-Skorska, M.; Slupianek, A.; Xue, L.; Zhang, Q.; Raghunath, P.N.; Hoser, G.; Wasik, M.A.; Morris, S.W.; Skorski, T. Role of signal transducer and activator of transcription 5 in nucleophosmin/anaplastic lymphoma kinase-mediated malignant transformation of lymphoid cells. *Cancer Res.* **2001**, *61*, 6517–6523. [[PubMed](#)]
51. Slupianek, A.; Nieborowska-Skorska, M.; Hoser, G.; Morrione, A.; Majewski, M.; Xue, L.; Morris, S.W.; Wasik, M.A.; Skorski, T. Role of phosphatidylinositol 3-kinase-Akt pathway in nucleophosmin/anaplastic lymphoma kinase-mediated lymphomagenesis. *Cancer Res.* **2001**, *61*, 2194–2199. [[PubMed](#)]
52. Amin, H.M.; McDonnell, T.J.; Ma, Y.; Lin, Q.; Fujio, Y.; Kunisada, K.; Leventaki, V.; Das, P.; Rassidakis, G.Z.; Cutler, C.; et al. Selective inhibition of stat3 induces apoptosis and g(1) cell cycle arrest in ALK-positive anaplastic large cell lymphoma. *Oncogene* **2004**, *23*, 5426–5434. [[CrossRef](#)] [[PubMed](#)]
53. Motegi, A.; Fujimoto, J.; Kotani, M.; Sakuraba, H.; Yamamoto, T. ALK receptor tyrosine kinase promotes cell growth and neurite outgrowth. *J. Cell Sci.* **2004**, *117*, 3319–3329. [[CrossRef](#)] [[PubMed](#)]
54. Moog-Lutz, C.; Degoutin, J.; Gouzi, J.Y.; Frobert, Y.; Brunet-de Carvalho, N.; Bureau, J.; Creminon, C.; Vigny, M. Activation and inhibition of anaplastic lymphoma kinase receptor tyrosine kinase by monoclonal antibodies and absence of agonist activity of pleiotrophin. *J. Biol. Chem.* **2005**, *280*, 26039–26048. [[CrossRef](#)] [[PubMed](#)]

55. Martinsson, T.; Eriksson, T.; Abrahamsson, J.; Caren, H.; Hansson, M.; Kogner, P.; Kamaraj, S.; Schonherr, C.; Weinmar, J.; Ruuth, K.; et al. Appearance of the novel activating f1174s ALK mutation in neuroblastoma correlates with aggressive tumor progression and unresponsiveness to therapy. *Cancer Res.* **2011**, *71*, 98–105. [[CrossRef](#)] [[PubMed](#)]
56. Piccinini, G.; Bacchiocchi, R.; Serresi, M.; Viviani, C.; Rossetti, S.; Gennaretti, C.; Carbonari, D.; Fazioli, F. A ligand-inducible epidermal growth factor receptor/anaplastic lymphoma kinase chimera promotes mitogenesis and transforming properties in 3T3 cells. *J. Biol. Chem.* **2002**, *277*, 22231–22239. [[CrossRef](#)] [[PubMed](#)]
57. Souttou, B.; Carvalho, N.B.; Raulais, D.; Vigny, M. Activation of anaplastic lymphoma kinase receptor tyrosine kinase induces neuronal differentiation through the mitogen-activated protein kinase pathway. *J. Biol. Chem.* **2001**, *276*, 9526–9531. [[CrossRef](#)] [[PubMed](#)]
58. Gouzi, J.Y.; Moog-Lutz, C.; Vigny, M.; Brunet-de Carvalho, N. Role of the subcellular localization of ALK tyrosine kinase domain in neuronal differentiation of PC12 cells. *J. Cell Sci.* **2005**, *118*, 5811–5823. [[CrossRef](#)] [[PubMed](#)]
59. Sattu, K.; Hochgrafe, F.; Wu, J.; Umapathy, G.; Schonherr, C.; Ruuth, K.; Chand, D.; Witek, B.; Fuchs, J.; Li, P.K.; et al. Phosphoproteomic analysis of anaplastic lymphoma kinase (ALK) downstream signaling pathways identifies signal transducer and activator of transcription 3 as a functional target of activated ALK in neuroblastoma cells. *FEBS J.* **2013**, *280*, 5269–5282. [[CrossRef](#)] [[PubMed](#)]
60. Bresler, S.C.; Wood, A.C.; Haglund, E.A.; Courtright, J.; Belcastro, L.T.; Plegaria, J.S.; Cole, K.; Toporovskaya, Y.; Zhao, H.; Carpenter, E.L.; et al. Differential inhibitor sensitivity of anaplastic lymphoma kinase variants found in neuroblastoma. *Sci. Transl. Med.* **2011**, *3*, 108ra114. [[CrossRef](#)] [[PubMed](#)]
61. Miyake, I.; Hakomori, Y.; Shinohara, A.; Gamou, T.; Saito, M.; Iwamatsu, A.; Sakai, R. Activation of anaplastic lymphoma kinase is responsible for hyperphosphorylation of SHCC in neuroblastoma cell lines. *Oncogene* **2002**, *21*, 5823–5834. [[CrossRef](#)] [[PubMed](#)]
62. McDermott, U.; Iafrate, A.J.; Gray, N.S.; Shioda, T.; Classon, M.; Maheswaran, S.; Zhou, W.; Choi, H.G.; Smith, S.L.; Dowell, L.; et al. Genomic alterations of anaplastic lymphoma kinase may sensitize tumors to anaplastic lymphoma kinase inhibitors. *Cancer Res.* **2008**, *68*, 3389–3395. [[CrossRef](#)] [[PubMed](#)]
63. Duijkers, F.A.; Gaal, J.; Meijerink, J.P.; Admiraal, P.; Pieters, R.; de Krijger, R.R.; van Noesel, M.M. Anaplastic lymphoma kinase (ALK) inhibitor response in neuroblastoma is highly correlated with ALK mutation status, ALK mRNA and protein levels. *Cell. Oncol. (Dordr.)* **2011**, *34*, 409–417. [[CrossRef](#)] [[PubMed](#)]
64. Schonherr, C.; Ruuth, K.; Kamaraj, S.; Wang, C.L.; Yang, H.L.; Combaret, V.; Djos, A.; Martinsson, T.; Christensen, J.G.; Palmer, R.H.; et al. Anaplastic lymphoma kinase (ALK) regulates initiation of transcription of MYCN in neuroblastoma cells. *Oncogene* **2012**, *31*, 5193–5200. [[CrossRef](#)] [[PubMed](#)]
65. Heukamp, L.C.; Thor, T.; Schramm, A.; De Preter, K.; Kumps, C.; De Wilde, B.; Odersky, A.; Peifer, M.; Lindner, S.; Spruessel, A.; et al. Targeted expression of mutated ALK induces neuroblastoma in transgenic mice. *Sci. Transl. Med.* **2012**, *4*, 141ra191. [[CrossRef](#)] [[PubMed](#)]
66. Moore, N.F.; Azarova, A.M.; Bhatnagar, N.; Ross, K.N.; Drake, L.E.; Frumm, S.; Liu, Q.S.; Christie, A.L.; Sanda, T.; Chesler, L.; et al. Molecular rationale for the use of pi3k/Akt/mTOR pathway inhibitors in combination with crizotinib in ALK-mutated neuroblastoma. *Oncotarget* **2014**, *5*, 8737–8749. [[CrossRef](#)] [[PubMed](#)]
67. Umapathy, G.; El Wakil, A.; Witek, B.; Chesler, L.; Danielson, L.; Deng, X.; Gray, N.S.; Johansson, M.; Kvarnbrink, S.; Ruuth, K.; et al. The kinase ALK stimulates the kinase ERK5 to promote the expression of the oncogene MYCN in neuroblastoma. *Sci. Signal.* **2014**, *7*, ra102. [[CrossRef](#)] [[PubMed](#)]
68. Schonherr, C.; Ruuth, K.; Yamazaki, Y.; Eriksson, T.; Christensen, J.; Palmer, R.H.; Hallberg, B. Activating ALK mutations found in neuroblastoma are inhibited by crizotinib and NVP-TAE684. *Biochem. J.* **2011**, *440*, 405–413. [[CrossRef](#)] [[PubMed](#)]
69. Fransson, S.; Hansson, M.; Ruuth, K.; Djos, A.; Berbegall, A.; Javanmardi, N.; Abrahamsson, J.; Palmer, R.H.; Noguera, R.; Hallberg, B.; et al. Intragenic anaplastic lymphoma kinase (ALK) rearrangements: Translocations as a novel mechanism of ALK activation in neuroblastoma tumors. *Genes Chromosomes Cancer* **2015**, *54*, 99–109. [[CrossRef](#)] [[PubMed](#)]
70. Dirks, W.G.; Fahnrich, S.; Lis, Y.; Becker, E.; MacLeod, R.A.; Drexler, H.G. Expression and functional analysis of the anaplastic lymphoma kinase (ALK) gene in tumor cell lines. *Int. J. Cancer* **2002**, *100*, 49–56. [[CrossRef](#)] [[PubMed](#)]

71. Lee, J.W.; Park, S.H.; Kang, H.J.; Park, K.D.; Shin, H.Y.; Ahn, H.S. ALK protein expression is related to neuroblastoma aggressiveness, but is not independent prognostic factor. *Cancer Res. Treat.* **2017**. [[CrossRef](#)] [[PubMed](#)]
72. Chen, S.; Zhou, C.; Ma, X.; Gong, L. Abnormality of anaplastic lymphoma kinase gene and its expression in pediatric neuroblastoma. *Zhonghua Bing Li Xue Za Zhi* **2014**, *43*, 541–545. [[PubMed](#)]
73. Wang, M.; Zhou, C.; Sun, Q.; Cai, R.; Li, Y.; Wang, D.; Gong, L. ALK amplification and protein expression predict inferior prognosis in neuroblastomas. *Exp. Mol. Pathol.* **2013**, *95*, 124–130. [[CrossRef](#)] [[PubMed](#)]
74. Duijkers, F.A.; Gaal, J.; Meijerink, J.P.; Admiraal, P.; Pieters, R.; de Krijger, R.R.; van Noesel, M.M. High anaplastic lymphoma kinase immunohistochemical staining in neuroblastoma and ganglioneuroblastoma is an independent predictor of poor outcome. *Am. J. Pathol.* **2012**, *180*, 1223–1231. [[CrossRef](#)] [[PubMed](#)]
75. Schulte, J.H.; Bachmann, H.S.; Brockmeyer, B.; Depreter, K.; Oberthur, A.; Ackermann, S.; Kahlert, Y.; Pajtler, K.; Theissen, J.; Westermann, F.; et al. High ALK receptor tyrosine kinase expression supersedes ALK mutation as a determining factor of an unfavorable phenotype in primary neuroblastoma. *Clin. Cancer Res.* **2011**, *17*, 5082–5092. [[CrossRef](#)] [[PubMed](#)]
76. Passoni, L.; Longo, L.; Collini, P.; Coluccia, A.M.; Bozzi, F.; Podda, M.; Gregorio, A.; Gambini, C.; Garaventa, A.; Pistoia, V.; et al. Mutation-independent anaplastic lymphoma kinase overexpression in poor prognosis neuroblastoma patients. *Cancer Res.* **2009**, *69*, 7338–7346. [[CrossRef](#)] [[PubMed](#)]
77. Devoto, M.; Specchia, C.; Laudenslager, M.; Longo, L.; Hakonarson, H.; Maris, J.; Mosse, Y. Genome-wide linkage analysis to identify genetic modifiers of ALK mutation penetrance in familial neuroblastoma. *Hum. Hered.* **2011**, *71*, 135–139. [[CrossRef](#)] [[PubMed](#)]
78. Berry, T.; Luther, W.; Bhatnagar, N.; Jamin, Y.; Poon, E.; Sanda, T.; Pei, D.; Sharma, B.; Vetharoy, W.R.; Hallsworth, A.; et al. The ALK(f1174l) mutation potentiates the oncogenic activity of MYCN in neuroblastoma. *Cancer Cell* **2012**, *22*, 117–130. [[CrossRef](#)] [[PubMed](#)]
79. Pilling, A.B.; Kim, J.; Estrada-Bernal, A.; Zhou, Q.; Le, A.T.; Singleton, K.R.; Heasley, L.E.; Tan, A.C.; DeGregori, J.; Doebele, R.C. ALK is a critical regulator of the myc-signaling axis in ALK positive lung cancer. *Oncotarget* **2018**, *9*, 8823–8835. [[CrossRef](#)] [[PubMed](#)]
80. Weiser, D.; Laudenslager, M.; Rappaport, E.; Carpenter, E.; Attiyeh, E.F.; Diskin, S.; London, W.B.; Maris, J.M.; Mosse, Y.P. Stratification of patients with neuroblastoma for targeted ALK inhibitor therapy. *J. Clin. Oncol.* **2011**, *29*, 9514. [[CrossRef](#)]
81. Bellini, A.; Bernard, V.; Leroy, Q.; Rio Frio, T.; Pierron, G.; Combaret, V.; Lapouble, E.; Clement, N.; Rubie, H.; Thebaud, E.; et al. Deep sequencing reveals occurrence of subclonal ALK mutations in neuroblastoma at diagnosis. *Clin. Cancer Res.* **2015**, *21*, 4913–4921. [[CrossRef](#)] [[PubMed](#)]
82. Okubo, J.; Takita, J.; Chen, Y.; Oki, K.; Nishimura, R.; Kato, M.; Sanada, M.; Hiwatari, M.; Hayashi, Y.; Igarashi, T.; et al. Aberrant activation of ALK kinase by a novel truncated form ALK protein in neuroblastoma. *Oncogene* **2012**, *31*, 4667–4676. [[CrossRef](#)] [[PubMed](#)]
83. Cazes, A.; Louis-Brennetot, C.; Mazot, P.; Dingli, F.; Lombard, B.; Boeva, V.; Daveau, R.; Cappel, J.; Combaret, V.; Schleiermacher, G.; et al. Characterization of rearrangements involving the ALK gene reveals a novel truncated form associated with tumor aggressiveness in neuroblastoma. *Cancer Res.* **2013**, *73*, 195–204. [[CrossRef](#)] [[PubMed](#)]
84. Schleiermacher, G.; Javanmardi, N.; Bernard, V.; Leroy, Q.; Cappel, J.; Rio Frio, T.; Pierron, G.; Lapouble, E.; Combaret, V.; Speleman, F.; et al. Emergence of new ALK mutations at relapse of neuroblastoma. *J. Clin. Oncol.* **2014**, *32*, 2727–2734. [[CrossRef](#)] [[PubMed](#)]
85. Eleveld, T.F.; Oldridge, D.A.; Bernard, V.; Koster, J.; Daage, L.C.; Diskin, S.J.; Schild, L.; Bentahar, N.B.; Bellini, A.; Chicard, M.; et al. Relapsed neuroblastomas show frequent ras-MAPK pathway mutations. *Nat. Genet.* **2015**, *47*, 864–871. [[CrossRef](#)] [[PubMed](#)]
86. Padovan-Merhar, O.M.; Raman, P.; Ostrovskaya, I.; Kalletta, K.; Rubnitz, K.R.; Sanford, E.M.; Ali, S.M.; Miller, V.A.; Mosse, Y.P.; Granger, M.P.; et al. Enrichment of targetable mutations in the relapsed neuroblastoma genome. *PLoS Genet.* **2016**, *12*, e1006501. [[CrossRef](#)] [[PubMed](#)]
87. Carpenter, E.L.; Haglund, E.A.; Mace, E.M.; Deng, D.; Martinez, D.; Wood, A.C.; Chow, A.K.; Weiser, D.A.; Belcastro, L.T.; Winter, C.; et al. Antibody targeting of anaplastic lymphoma kinase induces cytotoxicity of human neuroblastoma. *Oncogene* **2012**, *31*, 4859–4867. [[CrossRef](#)] [[PubMed](#)]

88. Galkin, A.V.; Melnick, J.S.; Kim, S.; Hood, T.L.; Li, N.; Li, L.; Xia, G.; Steensma, R.; Chopiuk, G.; Jiang, J.; et al. Identification of nvp-tae684, a potent, selective, and efficacious inhibitor of NPM-ALK. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 270–275. [[CrossRef](#)] [[PubMed](#)]
89. Heuckmann, J.M.; Holzel, M.; Sos, M.L.; Heynck, S.; Balke-Want, H.; Koker, M.; Peifer, M.; Weiss, J.; Lovly, C.M.; Grutter, C.; et al. ALK mutations conferring differential resistance to structurally diverse ALK inhibitors. *Clin. Cancer Res.* **2011**, *17*, 7394–7401. [[CrossRef](#)] [[PubMed](#)]
90. Epstein, L.F.; Chen, H.; Emkey, R.; Whittington, D.A. The r1275q neuroblastoma mutant and certain ATP-competitive inhibitors stabilize alternative activation loop conformations of anaplastic lymphoma kinase. *J. Biol. Chem.* **2012**, *287*, 37447–37457. [[CrossRef](#)] [[PubMed](#)]
91. Zhang, L.; Wu, B.; Baruchel, S. Oral metronomic topotecan sensitizes crizotinib antitumor activity in ALK(f1174l) drug-resistant neuroblastoma preclinical models. *Transl. Oncol.* **2017**, *10*, 604–611. [[CrossRef](#)] [[PubMed](#)]
92. Krytska, K.; Ryles, H.T.; Sano, R.; Raman, P.; Infarinato, N.R.; Hansel, T.D.; Makena, M.R.; Song, M.M.; Reynolds, C.P.; Mosse, Y.P. Crizotinib synergizes with chemotherapy in preclinical models of neuroblastoma. *Clin. Cancer Res.* **2016**, *22*, 948–960. [[CrossRef](#)] [[PubMed](#)]
93. Friboulet, L.; Li, N.; Katayama, R.; Lee, C.C.; Gainor, J.F.; Crystal, A.S.; Michellys, P.Y.; Awad, M.M.; Yanagitani, N.; Kim, S.; et al. The ALK inhibitor ceritinib overcomes crizotinib resistance in non-small cell lung cancer. *Cancer Discov.* **2014**, *4*, 662–673. [[CrossRef](#)] [[PubMed](#)]
94. Xu, F.; Li, H.; Sun, Y. Inhibition of axl improves the targeted therapy against ALK-mutated neuroblastoma. *Biochem. Biophys. Res. Commun.* **2014**, *454*, 566–571. [[CrossRef](#)] [[PubMed](#)]
95. Debruyne, D.N.; Bhatnagar, N.; Sharma, B.; Luther, W.; Moore, N.F.; Cheung, N.K.; Gray, N.S.; George, R.E. ALK inhibitor resistance in ALK(f1174l)-driven neuroblastoma is associated with axl activation and induction of emt. *Oncogene* **2016**, *35*, 3681–3691. [[CrossRef](#)] [[PubMed](#)]
96. Duijkers, F.A.; Meijerink, J.P.; Pieters, R.; van Noesel, M.M. Downregulation of axl in non-MYCN amplified neuroblastoma cell lines reduces migration. *Gene* **2013**, *521*, 62–68. [[CrossRef](#)] [[PubMed](#)]
97. Wood, A.C.; Krytska, K.; Ryles, H.T.; Infarinato, N.R.; Sano, R.; Hansel, T.D.; Hart, L.S.; King, F.J.; Smith, T.R.; Ainscow, E.; et al. Dual ALK and CDK4/6 inhibition demonstrates synergy against neuroblastoma. *Clin. Cancer Res.* **2017**, *23*, 2856–2868. [[CrossRef](#)] [[PubMed](#)]
98. Wang, H.Q.; Halilovic, E.; Li, X.; Liang, J.; Cao, Y.; Rakiec, D.P.; Ruddy, D.A.; Jeay, S.; Wuerthner, J.U.; Timple, N.; et al. Combined ALK and MDM2 inhibition increases antitumor activity and overcomes resistance in human ALK mutant neuroblastoma cell lines and xenograft models. *Elife* **2017**, *6*. [[CrossRef](#)] [[PubMed](#)]
99. Lu, J.; Guan, S.; Zhao, Y.; Yu, Y.; Woodfield, S.E.; Zhang, H.; Yang, K.L.; Bieberkehazhi, S.; Qi, L.; Li, X.; et al. The second-generation ALK inhibitor alectinib effectively induces apoptosis in human neuroblastoma cells and inhibits tumor growth in a th-MYCN transgenic neuroblastoma mouse model. *Cancer Lett.* **2017**, *400*, 61–68. [[CrossRef](#)] [[PubMed](#)]
100. Sakamoto, H.; Tsukaguchi, T.; Hiroshima, S.; Kodama, T.; Kobayashi, T.; Fukami, T.A.; Oikawa, N.; Tsukuda, T.; Ishii, N.; Aoki, Y. Ch5424802, a selective ALK inhibitor capable of blocking the resistant gatekeeper mutant. *Cancer Cell* **2011**, *19*, 679–690. [[CrossRef](#)] [[PubMed](#)]
101. Siaw, J.T.; Wan, H.; Pfeifer, K.; Rivera, V.M.; Guan, J.; Palmer, R.H.; Hallberg, B. Brigatinib, an anaplastic lymphoma kinase inhibitor, abrogates activity and growth in ALK-positive neuroblastoma cells, drosophila and mice. *Oncotarget* **2016**, *7*, 29011–29022. [[CrossRef](#)] [[PubMed](#)]
102. Zhang, S.; Anjum, R.; Squillace, R.; Nadworny, S.; Zhou, T.; Keats, J.; Ning, Y.; Wardwell, S.D.; Miller, D.; Song, Y.; et al. The potent ALK inhibitor brigatinib (ap26113) overcomes mechanisms of resistance to first- and second-generation ALK inhibitors in preclinical models. *Clin. Cancer Res.* **2016**, *22*, 5527–5538. [[CrossRef](#)] [[PubMed](#)]
103. Zou, H.Y.; Friboulet, L.; Kodack, D.P.; Engstrom, L.D.; Li, Q.; West, M.; Tang, R.W.; Wang, H.; Tsaparikos, K.; Wang, J.; et al. PF-06463922, an ALK/ROS1 inhibitor, overcomes resistance to first and second generation ALK inhibitors in preclinical models. *Cancer Cell* **2015**, *28*, 70–81. [[CrossRef](#)] [[PubMed](#)]
104. Infarinato, N.R.; Park, J.H.; Krytska, K.; Ryles, H.T.; Sano, R.; Szigety, K.M.; Li, Y.; Zou, H.Y.; Lee, N.V.; Smeal, T.; et al. The ALK/ROS1 inhibitor PF-06463922 overcomes primary resistance to crizotinib in ALK-driven neuroblastoma. *Cancer Discov.* **2016**, *6*, 96–107. [[CrossRef](#)] [[PubMed](#)]

105. Guan, J.; Tucker, E.R.; Wan, H.; Chand, D.; Danielson, L.S.; Ruuth, K.; El Wakil, A.; Witek, B.; Jamin, Y.; Umopathy, G.; et al. The ALK inhibitor PF-06463922 is effective as a single agent in neuroblastoma driven by expression of ALK and MYCN. *Dis. Model. Mech.* **2016**, *9*, 941–952. [[CrossRef](#)] [[PubMed](#)]
106. Shaw, A.T.; Friboulet, L.; Leshchiner, I.; Gainor, J.F.; Bergqvist, S.; Brooun, A.; Burke, B.J.; Deng, Y.L.; Liu, W.; Dardaei, L.; et al. Resensitization to crizotinib by the lorlatinib ALK resistance mutation L1198F. *N. Engl. J. Med.* **2016**, *374*, 54–61. [[CrossRef](#)] [[PubMed](#)]
107. Kwak, E.L.; Bang, Y.J.; Camidge, D.R.; Shaw, A.T.; Solomon, B.; Maki, R.G.; Ou, S.H.; Dezube, B.J.; Janne, P.A.; Costa, D.B.; et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N. Engl. J. Med.* **2010**, *363*, 1693–1703. [[CrossRef](#)] [[PubMed](#)]
108. Shaw, A.T.; Yeap, B.Y.; Solomon, B.J.; Riely, G.J.; Gainor, J.; Engelman, J.A.; Shapiro, G.I.; Costa, D.B.; Ou, S.H.; Butaney, M.; et al. Effect of crizotinib on overall survival in patients with advanced non-small-cell lung cancer harbouring ALK gene rearrangement: A retrospective analysis. *Lancet Oncol.* **2011**, *12*, 1004–1012. [[CrossRef](#)]
109. Camidge, D.R.; Bang, Y.J.; Kwak, E.L.; Iafrate, A.J.; Varella-Garcia, M.; Fox, S.B.; Riely, G.J.; Solomon, B.; Ou, S.H.; Kim, D.W.; et al. Activity and safety of crizotinib in patients with ALK-positive non-small-cell lung cancer: Updated results from a phase 1 study. *Lancet Oncol.* **2012**, *13*, 1011–1019. [[CrossRef](#)]
110. Shaw, A.T.; Kim, D.W.; Nakagawa, K.; Seto, T.; Crino, L.; Ahn, M.J.; De Pas, T.; Besse, B.; Solomon, B.J.; Blackhall, F.; et al. Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. *N. Engl. J. Med.* **2013**, *368*, 2385–2394. [[CrossRef](#)] [[PubMed](#)]
111. Solomon, B.J.; Mok, T.; Kim, D.W.; Wu, Y.L.; Nakagawa, K.; Mekhail, T.; Felip, E.; Cappuzzo, F.; Paolini, J.; Usari, T.; et al. First-line crizotinib versus chemotherapy in ALK-positive lung cancer. *N. Engl. J. Med.* **2014**, *371*, 2167–2177. [[CrossRef](#)] [[PubMed](#)]
112. Mosse, Y.P.; Lim, M.S.; Voss, S.D.; Wilner, K.; Ruffner, K.; Laliberte, J.; Rolland, D.; Balis, F.M.; Maris, J.M.; Weigel, B.J.; et al. Safety and activity of crizotinib for paediatric patients with refractory solid tumours or anaplastic large-cell lymphoma: A children’s oncology group phase 1 consortium study. *Lancet Oncol.* **2013**, *14*, 472–480. [[CrossRef](#)]
113. Balis, F.M.; Thompson, P.A.; Mosse, Y.P.; Blaney, S.M.; Minard, C.G.; Weigel, B.J.; Fox, E. First-dose and steady-state pharmacokinetics of orally administered crizotinib in children with solid tumors: A report on advl0912 from the children’s oncology group phase 1/pilot consortium. *Cancer Chemother. Pharmacol.* **2017**, *79*, 181–187. [[CrossRef](#)] [[PubMed](#)]
114. ClinicalTrials.gov. National Library of Medicine. Crizotinib in Treating Young Patients with Relapsed or Refractory Solid Tumors or Anaplastic Large Cell Lymphoma. Identifier: NCT00939770. Available online: <https://clinicaltrials.gov/ct2/show/NCT00939770> (accessed on 9 April 2018).
115. ClinicalTrials.gov. National Library of Medicine. An Investigational Drug, Crizotinib (PF-02341066), Is Being Studied In Tumors, Except Non-Small Cell Lung Cancer, That Are Positive For Anaplastic Lymphoma Kinase (ALK). Identifier: NCT01121588. Available online: <https://clinicaltrials.gov/ct2/show/NCT01121588> (accessed on 9 April 2018).
116. ClinicalTrials.gov. National Library of Medicine. Crizotinib and Combination Chemotherapy in Treating Younger Patients with Relapsed or Refractory Solid Tumors or Anaplastic Large Cell Lymphoma. Identifier: NCT01606878. Available online: <https://clinicaltrials.gov/ct2/show/NCT01606878> (accessed on 9 April 2018).
117. ClinicalTrials.gov. National Library of Medicine. Phase I Study of LDK378 in Pediatric, Malignancies with a Genetic Alteration in Anaplastic Lymphoma Kinase (ALK). Identifier: NCT01742286. Available online: <https://clinicaltrials.gov/ct2/show/NCT01742286> (accessed on 9 April 2018).
118. ClinicalTrials.gov. National Library of Medicine. Study of RDX-101 in Children with Recurrent or Refractory Solid Tumors and Primary CNS Tumors, With or Without TRK, ROS1, or ALK Fusions. Identifier: NCT02650401. Available online: <https://clinicaltrials.gov/ct2/show/NCT02650401> (accessed on 9 April 2018).
119. ClinicalTrials.gov. National Library of Medicine. Next Generation Personalized Neuroblastoma Therapy (NEPENTHE). Identifier: NCT02780128. Available online: <https://clinicaltrials.gov/ct2/show/NCT02780128> (accessed on 9 April 2018).
120. Allen, C.E.; Laetsch, T.W.; Mody, R.; Irwin, M.S.; Lim, M.S.; Adamson, P.C.; Seibel, N.L.; Parsons, D.W.; Cho, Y.J.; Janeway, K.; et al. Target and agent prioritization for the children’s oncology group-national cancer institute pediatric match trial. *J. Natl. Cancer Inst.* **2017**, *109*. [[CrossRef](#)] [[PubMed](#)]

121. ClinicalTrials.gov. National Library of Medicine. Ensartinib in Treating Patients with Relapsed or Refractory Advanced Solid Tumors, Non-Hodgkin Lymphoma, or Histiocytic Disorders with ALK or ROS1 Genomic Alterations (A Pediatric MATCH Treatment Trial). Identifier: NCT03213652. Available online: <https://clinicaltrials.gov/ct2/show/NCT03213652> (accessed on 9 April 2018).
122. ClinicalTrials.gov. National Library of Medicine. Study of Lorlatinib (PF-06463922). Identifier: NCT03107988. Available online: <https://clinicaltrials.gov/ct2/show/NCT03107988> (accessed on 9 April 2018).
123. ClinicalTrials.gov. National Library of Medicine. Iobenguane I-131 or Crizotinib and Standard Therapy in Treating Younger Patients with Newly-Diagnosed High-Risk Neuroblastoma or Ganglioneuroblastoma. Identifier: NCT03126916. Available online: <https://clinicaltrials.gov/ct2/show/NCT03126916> (accessed on 9 April 2018).
124. Shaw, A.T.; Engelman, J.A. Ceritinib in ALK-rearranged non-small-cell lung cancer. *N. Engl. J. Med.* **2014**, *370*, 2537–2539. [[CrossRef](#)] [[PubMed](#)]
125. Khozin, S.; Blumenthal, G.M.; Zhang, L.; Tang, S.; Brower, M.; Fox, E.; Helms, W.; Leong, R.; Song, P.; Pan, Y.; et al. Fda approval: Ceritinib for the treatment of metastatic anaplastic lymphoma kinase-positive non-small cell lung cancer. *Clin. Cancer Res.* **2015**, *21*, 2436–2439. [[CrossRef](#)] [[PubMed](#)]
126. Camidge, D.R.; Doebele, R.C. Treating ALK-positive lung cancer—Early successes and future challenges. *Nat. Rev. Clin. Oncol.* **2012**, *9*, 268–277. [[CrossRef](#)] [[PubMed](#)]
127. Doebele, R.C.; Pilling, A.B.; Aisner, D.L.; Kutateladze, T.G.; Le, A.T.; Weickhardt, A.J.; Kondo, K.L.; Linderman, D.J.; Heasley, L.E.; Franklin, W.A.; et al. Mechanisms of resistance to crizotinib in patients with ALK gene rearranged non-small cell lung cancer. *Clin. Cancer Res.* **2012**, *18*, 1472–1482. [[CrossRef](#)] [[PubMed](#)]
128. Katayama, R.; Shaw, A.T.; Khan, T.M.; Mino-Kenudson, M.; Solomon, B.J.; Halmos, B.; Jessop, N.A.; Wain, J.C.; Yeo, A.T.; Benes, C.; et al. Mechanisms of acquired crizotinib resistance in ALK-rearranged lung cancers. *Sci. Transl. Med.* **2012**, *4*, 120ra117. [[CrossRef](#)] [[PubMed](#)]
129. Lovly, C.M.; McDonald, N.T.; Chen, H.; Ortiz-Cuaran, S.; Heukamp, L.C.; Yan, Y.; Florin, A.; Ozretic, L.; Lim, D.; Wang, L.; et al. Rationale for co-targeting IGF-1r and ALK in ALK fusion-positive lung cancer. *Nat. Med.* **2014**, *20*, 1027–1034. [[CrossRef](#)] [[PubMed](#)]
130. Yamada, T.; Takeuchi, S.; Nakade, J.; Kita, K.; Nakagawa, T.; Nanjo, S.; Nakamura, T.; Matsumoto, K.; Soda, M.; Mano, H.; et al. Paracrine receptor activation by microenvironment triggers bypass survival signals and ALK inhibitor resistance in EML4-ALK lung cancer cells. *Clin. Cancer Res.* **2012**, *18*, 3592–3602. [[CrossRef](#)] [[PubMed](#)]
131. Ji, C.; Zhang, L.; Cheng, Y.; Patel, R.; Wu, H.; Zhang, Y.; Wang, M.; Ji, S.; Belani, C.P.; Yang, J.M.; et al. Induction of autophagy contributes to crizotinib resistance in ALK-positive lung cancer. *Cancer Biol. Ther.* **2014**, *15*, 570–577. [[CrossRef](#)] [[PubMed](#)]
132. Wang, H.Y.; Ho, C.C.; Shih, J.Y. Multiple acquired resistance mutations of the ALK tyrosine kinase domain after sequential use of ALK inhibitors. *J. Thorac. Oncol.* **2017**, *12*, e49–e51. [[CrossRef](#)] [[PubMed](#)]
133. Katayama, R.; Friboulet, L.; Koike, S.; Lockerman, E.L.; Khan, T.M.; Gainor, J.F.; Iafrate, A.J.; Takeuchi, K.; Taiji, M.; Okuno, Y.; et al. Two novel ALK mutations mediate acquired resistance to the next-generation ALK inhibitor alectinib. *Clin. Cancer Res.* **2014**, *20*, 5686–5696. [[CrossRef](#)] [[PubMed](#)]
134. Ou, S.H.; Klempner, S.J.; Greenbowe, J.R.; Azada, M.; Schrock, A.B.; Ali, S.M.; Ross, J.S.; Stephens, P.J.; Miller, V.A. Identification of a novel hip1-ALK fusion variant in non-small-cell lung cancer (NSCLC) and discovery of ALK i1171 (i1171n/s) mutations in two ALK-rearranged nsclc patients with resistance to alectinib. *J. Thorac. Oncol.* **2014**, *9*, 1821–1825. [[CrossRef](#)] [[PubMed](#)]
135. Gainor, J.F.; Chi, A.S.; Logan, J.; Hu, R.; Oh, K.S.; Brastianos, P.K.; Shih, H.A.; Shaw, A.T. Alectinib dose escalation reinduces central nervous system responses in patients with anaplastic lymphoma kinase-positive non-small cell lung cancer relapsing on standard dose alectinib. *J. Thorac. Oncol.* **2016**, *11*, 256–260. [[CrossRef](#)] [[PubMed](#)]

