

Hindawi Publishing Corporation  
Journal of Pregnancy  
Volume 2012, Article ID 638476, 8 pages  
doi:10.1155/2012/638476

## Review Article

# The Long and Short of It: The Role of Telomeres in Fetal Origins of Adult Disease

Stephanie E. Hallows,<sup>1</sup> Timothy R. H. Regnault,<sup>1,2,3</sup> and Dean H. Betts<sup>1,3</sup>

<sup>1</sup>Department of Physiology and Pharmacology, University of Western Ontario, Ontario, London, ON, Canada N6A 5C1

<sup>2</sup>Department of Obstetrics and Gynaecology, University of Western Ontario, Ontario, London, ON, Canada N6H 5W9

<sup>3</sup>Children's Health Research Institute, Lawson Health Research Institute, London, ON, Canada N6C 2V5

Correspondence should be addressed to Dean H. Betts, dean.betts@schulich.uwo.ca

Received 7 April 2012; Accepted 24 August 2012

Academic Editor: Mark Nijland

Copyright © 2012 Stephanie E. Hallows et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Placental insufficiency, maternal malnutrition, and other causes of intrauterine growth restriction (IUGR) can significantly affect short-term growth and long-term health. Following IUGR, there is an increased risk for cardiovascular disease and Type 2 Diabetes. The etiology of these diseases is beginning to be elucidated, and premature aging or cellular senescence through increased oxidative stress and DNA damage to telomeric ends may be initiators of these disease processes. This paper will explore the areas where telomere and telomerase biology can have significant effects on various tissues in the body in IUGR outcomes.

## 1. Intrauterine Growth Restriction and Placental Insufficiency

The World Health Organization (WHO) estimates that the incidence of infants born low birth weight (LBW) in North America is approximately 7% [1] and is commonly characterized as a birth weight under 2500 g. A further sub classification within the LBW classification is that of intrauterine growth restriction (IUGR) where a fetus fails to reach its genetic growth potential, as a result of a compromised intrauterine environment and is generally defined as being less than the 3rd percentile of normal birth weight. IUGR can be caused by a heterogeneous mix of fetal, placental and maternal factors. Fetal factors include genetic abnormalities, multiple gestation, and infections [2], while maternal contributing factors for IUGR include malnutrition, drug intake, hypertension, Type I or gestational diabetes, and persistent hypoxia due to cardiovascular disease or high altitude [2]. Placental insufficiency is a common cause of IUGR, accounting for ~60% of IUGR and includes reduced placental development, abnormal trophoblast invasion into the maternal decidua, placenta previa, and placental infarcts [3, 4]. Human studies and animal models of placental insufficiency have demonstrated a decreased rate of nutrient

transfer across the placenta. Specifically, IUGR fetuses are characterized by alterations in oxygen supply [5–7], glucose and amino acid supply [8–10], and with increased fetal triglycerides [11, 12]. Due to the lack of oxygen and altered nutrient balance, the fetus redirects these scarce resources to the brain, heart, and adrenal glands, leaving other tissues in the body more severely growth restricted, resulting in asymmetric IUGR [13, 14]. This redistribution of nutritional supplies leads to a decrease in muscularity and an increase in the percentage of body fat in these infants that persists throughout childhood and adult life [15, 16] and is commonly associated with changes in insulin sensitivity and other markers of the metabolic syndrome [17, 18]. These observations and others set the stage for the idea that changes in growth during *in utero* life may predispose offspring to increased risk of disease in later life, or the concept of the developmental origins of health and disease (DOHaD).

## 2. Developmental Origins of Health and Disease

LBW infants primarily present an increased risk for perinatal morbidity and mortality [19]. However, through the work of David Barker and colleagues, the concept that there further

exists a relationship between birth weight and an increased risk for developing diseases including coronary heart disease, Type 2 Diabetes, and hypertension in later life has been generally accepted as a secondary concern for LBW infants [20, 21]. Since the early observations, this relationship between low birth weights, followed by a rapid catch up growth leading to increased risk of adult disease has been reported in a number of human population studies and in many animal models of IUGR [22, 23].

Barker and colleagues theorized that there are critical periods during development when the fetus adapts and is programmed to its *in utero* surroundings, and after which the fetuses phenotype is established [24]. This is the basis of the “thrifty phenotype” hypothesis, where there is a mismatch between the intrauterine environment the fetuses encounters, and the exuterine environment an individual grows up in [25]. This can cause a relative over compensation in glucose and insulin pathways promoted by an affluent adult environment which makes the offspring more susceptible to adult disease [25].

To study this phenomenon, several IUGR animal models have been developed, most commonly carried out in the monkey, pig, sheep, and rodents [26]. The animal models use different intervention strategies to cause IUGR and some of the most widely used methods include nutritional models with decreased caloric or protein intakes; surgical or utero-placental blood flow alterations such as uterine artery ligations; glucocorticoid treatment; and increased maternal stressors such as high heat [26]. These animal models have shown offspring to be IUGR, but do not exhibit the same adult disease manifestations, which may depend on the particular IUGR model utilized [27]. While these models have given insight into disease progression correlated with LBW, there is still much to be understood about the molecular pathways that can lead to adult disease. Recent studies have indicated telomere length and telomerase activity to be affected in various tissues, and this may affect disease progression and accelerated cellular aging in post-natal life.

### 3. Telomere Biology and Cellular Senescence

Cellular senescence can be triggered by “critically” short or uncapped telomere(s) [28]. Telomeres are comprised of tandem DNA repeats (TTAGGG)<sup>n</sup> found at the ends of chromosomes to protect them from inappropriate DNA fusions, DNA breaks and to prevent DNA shortening into coding DNA [29]. They protect DNA ends by forming a protective cap with a single strand telomere overhang and telomere binding proteins including TRF1, TRF2, and shelterin among others [30]. Telomeres are maintained in cells by the holoenzyme, telomerase. It consists of an enzymatic protein component, telomerase reverse transcriptase (TERT) and an RNA template component, telomerase RNA component (TERC). TERC is widely expressed, but TERT expression is tightly regulated and is only found highly expressed in germ cells, stem cells, and ~90% of cancer cell lines contain a functional telomerase [31]. Without a functional telomerase,

a cell undergoing cell division will have progressive telomere shortening, resulting in telomere-dependent replicative senescence and an inability to divide further when a “critically” short telomere length is reached [32, 33]. Premature senescence can occur without critically short telomeres when cells encounter stressors including oxidative stress that can cause telomere dysfunction, telomere uncapping or other DNA damage. These stressors can trigger DNA damage response or senescent pathways, including p53-p21 ARF pathways and p16<sup>INK4a</sup>-Rb pathways [34, 35]. Average telomere length of circulating leukocytes has been shown to be a good indicator of aging and age-related diseases [36], but only one critically short telomere can initiate a DNA damage response to become senescent or apoptotic [37].

Telomerase may also exhibit other extra-telomeric functions involved in gene expression, DNA damage response, apoptosis regulation and a possible role in mitochondria and oxidative stress protection [38–40]. The role of telomerase in the mitochondria is an emerging topic, as recent studies have shown telomerase to translocate out of the nucleus to the mitochondria under oxidative stress conditions, leaving the genome vulnerable to telomeric DNA shortening and/or damage that could lead to premature cellular senescence [40]. Due to its role in apoptosis and senescence, telomerase/telomere biology has been explored for its roles in reproduction, placental development, and premature aging following IUGR [41–47]. These topics will be explored further in this paper.

### 4. Functions of Telomeres and Telomerase in Germ Cells

Germ cells are important as they determine the telomere length set for all the cells in our bodies and for the next generation [48]. Ovarian development is also greatly influenced by maternal nutrition and early growth, and it has been shown that undernutrition can affect ovarian size, follicular development and onset of puberty [49–52]. In the oocyte, high telomerase activity is present in the developing oocyte but significantly diminishes upon maturation [53, 54]. Once fertilization takes place telomerase activity dramatically rises, but is downregulated when cells go through differentiation [53–55]. To demonstrate the importance of telomerase activity in the oocyte, mice that are mTert<sup>-/-</sup> have been used to show that it is imperative to have a functional telomerase enzyme for proper germ cell formation and fertility. Tert<sup>-/-</sup> mice display infertility due to numerous causes that are present in the human populations including impaired meiotic synapsis and recombination, with more germ cells preferentially arresting in early meiosis [56, 57]. Similar problems can occur in human infertility, which may show a link between telomerase activity, telomeres, and germ cell health [58]. In fact, there appears to be a direct link between high telomerase activity in luteinised granulosa cells, higher rates of embryo implantation, and pregnancy during *in vitro* fertilization [42].

Today, women are delaying becoming pregnant and may also therefore encounter difficulties trying to conceive [59].

This decrease in fertility could be due, in part, to telomere dysfunction and DNA damage, as the germ cells have been exposed to a lifelong accumulation of possible reactive oxygen species (ROS)- induced damage upon the telomeric ends. Indeed, ROS-induced telomere shortening/uncapping has been observed in oocytes and/or preimplantation embryos leading to apoptosis or senescence at relatively long telomere lengths [60, 61]. Furthermore, oogenesis begins during early fetal life, where it can be exposed to an aged maternal environment. A recent paper has shown that maternal undernutrition can significantly affect primary, secondary, and antral follicle numbers in the adult rat offspring, with decreased factors involved in folliculogenesis, ovarian steroidogenesis, and ovulation [52]. Furthermore, female rats that have been exposed to undernutrition during development have been shown to go through reproductive senescence at an earlier age compared to the control counterparts [62]. It has been suggested these effects could be due to increased oxidative stress, as there was increased ovarian protein carbonyl content and hyperoxidized Prx3 [52]. Treatments with an antioxidant N-acetyl-L-cysteine (NAC) has been shown to delay oocyte aging, and increase oocyte quality and litter sizes [63]. Additionally, telomere length was elongated and telomerase activity greater in ovaries with this short term NAC treatment [63].

Stem cells are also susceptible to aging and ROS damage. In human and mice, telomeres shorten in stem cell populations as an organism ages [63, 64]. Over a lifespan, telomere shortening and increased oxidative stress can significantly reduce the self-renewal and proliferative capacity of a stem cell compartment [65, 66]. This has recently been shown to occur early in development, as stem cell populations from LBW infants show altered abilities compared to normal stem cells. Low birth weight endothelial colony forming progenitor cells (LBW-ECFCs) taken from cord blood have been shown to form fewer colonies compared to controls and take longer to appear, and show alterations in their ability to proliferate, migrate, and form sprouts and tubes *in vitro* [67]. LBW-ECFCs also displayed reduced capillary networks *in vivo* when injected in matrigel plugs in mice and was likely the result of an increase in gene expression of antiangiogenic genes including thrombospondin 1 (THBS1) compared to controls [67]. In addition, studies have found that there is a relationship between birth weight and the concentrations of endothelial progenitor cells in the cord blood, where LBW infants have the lowest concentrations compared to higher birth weight infants [68].

These studies suggest that telomere health may begin during germ cell development, when a fetus is still developing. These tantalizing outcomes show that an adverse intrauterine environment can negatively affect telomere health for future generations.

## 5. Telomeres and Telomerase in the Placenta

The placenta is an organ that is designed to be invasive into the maternal tissue to allow for an adequate supply of maternal blood for delivery of oxygen and nutrients to the

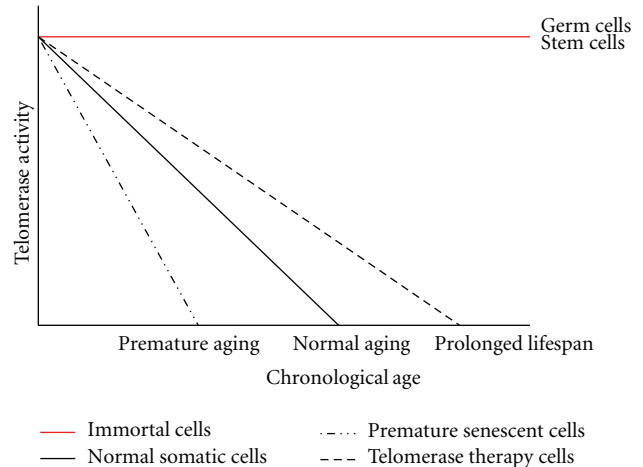


FIGURE 1: Telomere length and stability decrease throughout a lifespan. *In utero* stress can cause premature telomere shortening and destabilization that will cause cells to age and become senescent compared to normal individuals. This can be further exacerbated with poor post-natal nutrition. With new molecular modifiers of telomerase becoming available, it may soon be possible to rescue cells and reduce premature aging in a tissue. Cells that have a functional telomerase can maintain telomere integrity and be immortalized, such as stem cells and germ cells.

fetus. To achieve this, extravillous trophoblast cells migrate and invade the maternal spiral artery walls in decidua and myometrium [69]. In the first weeks of pregnancy, there is rapid growth of the cells in a relatively hypoxic environment. Telomerase activity is the highest in placental trophoblast in the first trimester, and activity decreases through the duration of pregnancy to term [41, 70, 71]. Normal telomerase activity is correlated with HIF-1 $\alpha$  expression, and it was shown to induce hTERT expression by binding to two HIF-1 $\alpha$  consensus-binding sites in the hTERT promoter [72]. As the maternal arterial circulation is initiated, there is three-fold rise in intraplacental oxygen concentration [73], which could explain the normal decrease in telomerase activity through HIF-1 $\alpha$  from 93.5% positive telomerase activity in first trimester chorionic villi versus 62.5% from second and third trimester chorionic villi [71]. Interestingly, in pregnancies complicated with IUGR and/or preeclampsia there is decreased telomerase activity and hTERT staining in placental trophoblasts compared to controls between 26 and 39 weeks [71, 74]. Additionally, decreased or absent telomerase activity can also be found in association with unfavourable outcomes of pregnancy such as spontaneous abortions and intrauterine fetal death [41]. Conversely, in hydatidiform molar pregnancies there is an increase in telomerase activity and hTERT staining that accompanies uncontrolled cell growth and division in the formation of a mass [70, 75].

To further demonstrate a relationship between placental telomerase activity and low birth weights, Kim and colleagues carried out a discordant twin study, where the smaller twin was found to have significantly decreased placental telomerase activity in relation to the normal birth weight twin [43]. Normal pregnancies were found to have long

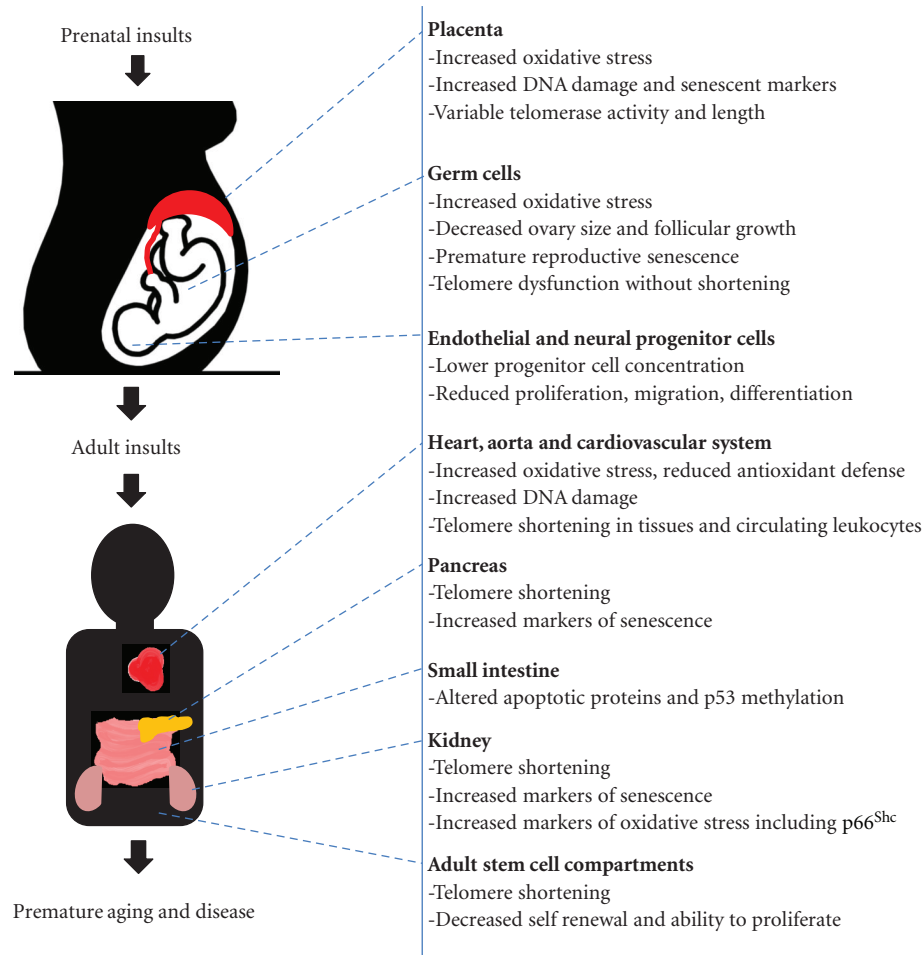


FIGURE 2: Various tissues in the body are affected by IUGR and have shown measurable changes in oxidative stress, telomerase activity, telomere shortening, and increased markers of senescence. These tissues can have a role in the development of adult diseases such as type 2 diabetes, cardiovascular disease, and metabolic syndrome.

telomeres [76], and pregnancies complicated with IUGR and/or preeclampsia were found to have shortened placental telomeres compared to controls, but this shortening was not apparent in the cord blood [44, 74]. In association with telomere shortening, studies have found decreases in antiapoptotic protein Bcl-2 and upregulation of senescent markers p16 and p21 in the IUGR placenta [44]. These effects may be due to increases in oxidative stress markers in the cord blood and DNA damage in the IUGR placenta [77].

## 6. Telomeres and Telomerase in the Developmental Origins of Health and Disease

Placental insufficiency resulting in LBW and IUGR can have a wide range of detrimental effects on a developing fetus, in addition to just being born small. Many studies are currently focusing on the “programming” or epigenetic modifications that can occur in utero and persist long term [78] but other disturbances are also taking place including

increased oxidative stress which can have widespread effects. Interestingly, recent studies have implicated ROS in altering chromatin remodelling enzymes and histone modifications that control gene expression [79]. In humans, pregnancy complications including IUGR and preeclampsia have been shown to increase markers of oxidative stress and antioxidant defences in the mother and in neonates [80–83]. Oxidative stress can lead to lipid peroxidation, protein damage and DNA damage and can cause apoptosis if severe, or cells can exit the cell cycle and enter into premature senescence. There has been increasing evidence that cellular senescence contributes to the onset of disease, and is perhaps beginning *in utero*. Low birth weight studies in animals have indicated an increase in cell cycle inhibitors p16 and p21 along with alterations in the methylation and expression of p53 in various tissues including the kidney and pancreatic  $\beta$ -cells [45, 47, 84, 85]. Higher levels of ROS have been measured in these tissues, including an increase in the stress adaptor protein p66<sup>Shc</sup>, which has been shown to have significant impact on decreasing longevity [45, 86]. ROS in these tissues could contribute to DNA damage, and a study has



shown a greater amount of DNA single strand breaks, shortened telomeres and increased oxidative stress in aortic tissues of maternal low protein recuperated pups [46]. This type of DNA damage can activate DNA damage responses, including the p16-Rb pathway and p53-p21 pathway to remove damaged cells from the replicating population to deter mutations from arising and being replicated. Telomere shortening or breaks can also turn on these responses in a cell, and studies have found an association between adverse intrauterine environment and shorter telomere length later in life in both human and animal studies [46, 87, 88]. Telomere attrition is also associated with cardiovascular disease and atherosclerosis [89–93] and type 2 diabetes [94–96].

As a person ages, they acquire more senescent cells and accompanied by various aging pathologies including atherosclerosis [89]. If this accumulation begins at a younger age these senescent cells could not only intrinsically age organs more quickly, but they may affect neighbouring cells by secreting altered cytokines, extracellular matrix proteins, and growth factors that lead to further changes within aging organs and tissues [97]. Baker and colleagues designed an inducible transgenic system in a progeroid mouse model that eliminated p16 expressing senescent cells *in vivo*. The loss of senescent cells was sufficient to delay the onset of age-related pathologies in adipose, skeletal muscle, and eye tissue of these mice [98]. Furthermore, when this clearance of p16 expressing senescent cells occurred later in life, the progression of various aging diseases was attenuated [98]. This study demonstrates that the accumulation of senescent cells in tissues is involved in generating age-related organ pathologies and that their removal can prevent or delay tissue dysfunction and extend healthspan. This could be achieved with the maintenance of telomeres and telomerase activity to aid and repair oxidative stress and DNA damage, which could decrease senescent cells in a tissue (Figure 1).

## 7. Perspectives

There is accumulating evidence that points to premature cellular aging as being a contributor to the poor postnatal outcomes of IUGR patients that can lead to increased risk for adult disease. Questions may arise whether early senescence onset is a cause or effect of IUGR? Nevertheless, telomere length and integrity is becoming a strong molecular marker of cellular aging in many studies including cardiovascular disease, type 2 diabetes, and overall health and longevity [95, 96, 99] (Figure 2). The shortened telomere lengths observed in young adults after intrauterine stress exposure [88] may predispose these individuals to early onset of various age-related diseases. Increased shortening of telomeres may be a result of the rapid postnatal catch up growth in tissues that have downregulated telomerase activity levels or due to the redistribution telomerase out of the nucleus in stressed tissues. Altered cellular metabolism is a hallmark of IUGR [100] and these metabolic shifts that elevate ROS production can increase telomere shortening or induce telomere dysfunction leading to premature senescence, [101] that may

remain permanently damaged in a cell [102]. Conversely, telomere dysfunction itself induces altered metabolic and mitochondrial functions that may in turn cause further metabolic and oxidative stress deregulations in various tissues [103]. The induced metabolic syndrome observed in various IUGR models may be due to the early and increased accumulation of senescent cells within certain tissues. The pro/antioxidant balance is significantly affected in IUGR pregnancies [81, 83]. Postnatal antioxidant treatment with resveratrol prevents diet-induced metabolic syndrome in IUGR rats [104]. Future studies characterizing the role of the uterine environment on telomere and telomerase dynamics with premature senescence will lead to new knowledge on the etiology behind the developmental origins of health and disease leading to novel treatments such as telomerase therapy as a possible means to ameliorate the premature onset of adult disease in IUGR outcomes.

## References

- [1] T. Wardlaw, A. Blanc, J. Zupan, and E. Ahman, *Low Birth Weight: Country, Regional and Global Estimates*, World Health Organization, 2004.
- [2] D. Brodsky and H. Christou, "Current concepts in intrauterine growth restriction," *Journal of Intensive Care Medicine*, vol. 19, no. 6, pp. 307–319, 2004.
- [3] R. Pijnenborg, L. Vercruyse, and M. Hanssens, "The uterine spiral arteries in human pregnancy: facts and controversies," *Placenta*, vol. 27, no. 9–10, pp. 939–958, 2006.
- [4] T. Y. Khong, F. De Wolf, W. B. Robertson, and I. Brosens, "Inadequate maternal vascular response to placentation in pregnancies complicated by pre-eclampsia and by small-for-gestational age infants," *BJOG*, vol. 93, no. 10, pp. 1049–1059, 1986.
- [5] T. R. H. Regnault, B. de Vrijer, H. L. Galan, R. B. Wilkening, F. C. Battaglia, and G. Meschia, "Development and mechanisms of fetal hypoxia in severe fetal growth restriction," *Placenta*, vol. 28, no. 7, pp. 714–723, 2007.
- [6] T. M. Mayhew, R. Manwani, C. Ohadike, J. Wijesekara, and P. N. Baker, "The placenta in pre-eclampsia and intrauterine growth restriction: studies on exchange surface areas, diffusion distances and villous membrane diffusive conductances," *Placenta*, vol. 28, no. 2–3, pp. 233–238, 2007.
- [7] R. Resnik, "Intrauterine growth restriction," *Obstetrics and Gynecology*, vol. 99, no. 3, pp. 490–496, 2002.
- [8] A. M. Marconi, C. L. Paolini, L. Stramare et al., "Steady state maternal-fetal leucine enrichments in normal and intrauterine growth-restricted pregnancies," *Pediatric Research*, vol. 46, no. 1, pp. 114–119, 1999.
- [9] C. L. Paolini, A. M. Marconi, S. Ronzoni et al., "Placental transport of leucine, phenylalanine, glycine, and proline in intrauterine growth-restricted pregnancies," *Journal of Clinical Endocrinology and Metabolism*, vol. 86, no. 11, pp. 5427–5432, 2001.
- [10] P. J. Thureen, K. A. Trembler, G. Meschia, E. L. Makowski, and R. B. Wilkening, "Placental glucose transport in heat-induced fetal growth retardation," *American Journal of Physiology*, vol. 263, no. 3, pp. R578–R585, 1992.
- [11] G. Alvino, V. Cozzi, T. Radaelli, H. Ortega, E. Herrera, and I. Cetin, "Maternal and fetal fatty acid profile in normal and intrauterine growth restriction pregnancies with and without

- preeclampsia," *Pediatric Research*, vol. 64, no. 6, pp. 615–620, 2008.
- [12] V. A. Rodie, M. J. Caslake, F. Stewart et al., "Fetal cord plasma lipoprotein status in uncomplicated human pregnancies and in pregnancies complicated by pre-eclampsia and intrauterine growth restriction," *Atherosclerosis*, vol. 176, no. 1, pp. 181–187, 2004.
- [13] R. Verkauskiene, J. Beltrand, O. Claris et al., "Impact of fetal growth restriction on body composition and hormonal status at birth in infants of small and appropriate weight for gestational age," *European Journal of Endocrinology*, vol. 157, no. 5, pp. 605–612, 2007.
- [14] E. M. Widdowson, D. E. Crabb, and R. D. Milner, "Cellular development of some human organs before birth," *Archives of Disease in Childhood*, vol. 47, no. 254, pp. 652–655, 1972.
- [15] M. L. Hediger, M. D. Overpeck, R. J. Kuczumski, A. McGlynn, K. R. Maurer, and W. W. Davis, "Muscularity and fatness of infants and young children born small- or large-for-gestational-age," *Pediatrics*, vol. 102, no. 5, article E60, 1998.
- [16] L. Ibáñez, K. Ong, D. B. Dunger, and F. De Zegher, "Early development of adiposity and insulin resistance after catch-up weight gain in small-for-gestational-age children," *Journal of Clinical Endocrinology and Metabolism*, vol. 91, no. 6, pp. 2153–2158, 2006.
- [17] A. C. J. Ravelli, J. H. P. Van Der Meulen, R. P. J. Michels et al., "Glucose tolerance in adults after prenatal exposure to famine," *The Lancet*, vol. 351, no. 9097, pp. 173–177, 1998.
- [18] D. J. P. Barker, "Adult consequences of fetal growth restriction," *Clinical Obstetrics and Gynecology*, vol. 49, no. 2, pp. 270–283, 2006.
- [19] D. D. Mcintire, S. L. Bloom, B. M. Casey, and K. J. Leveno, "Birth weight in relation to morbidity and mortality among newborn infants," *The New England Journal of Medicine*, vol. 340, no. 16, pp. 1234–1238, 1999.
- [20] D. J. P. Barker, C. Osmond, J. Golding, D. Kuh, and M. E. J. Wadsworth, "Growth in utero, blood pressure in childhood and adult life, and mortality from cardiovascular disease," *BMJ*, vol. 298, no. 6673, pp. 564–567, 1989.
- [21] D. J. P. Barker, A. R. Bull, C. Osmond, and S. J. Simmonds, "Fetal and placental size and risk of hypertension in adult life," *BMJ*, vol. 301, no. 6746, pp. 259–262, 1990.
- [22] S. E. Ozanne, "Metabolic programming in animals," *British Medical Bulletin*, vol. 60, pp. 143–152, 2001.
- [23] S. R. Thorn, P. J. Rozance, L. D. Brown, and W. W. Hay, "The intrauterine growth restriction phenotype: fetal adaptations and potential implications for later life insulin resistance and diabetes," *Seminars in Reproductive Medicine*, vol. 29, no. 3, pp. 225–236, 2011.
- [24] D. J. P. Barker, "Fetal origins of coronary heart disease," *BMJ*, vol. 311, no. 6998, pp. 171–174, 1995.
- [25] C. N. Hales and D. J. P. Barker, "The thrifty phenotype hypothesis," *British Medical Bulletin*, vol. 60, pp. 5–20, 2001.
- [26] P. M. Vuguin, "Animal models for small for gestational age and fetal programming of adult disease," *Hormone Research*, vol. 68, no. 3, pp. 113–123, 2007.
- [27] K. D. Nüsken, H. Schneider, C. Plank et al., "Fetal programming of gene expression in growth-restricted rats depends on the cause of low birth weight," *Endocrinology*, vol. 152, no. 4, pp. 1327–1335, 2011.
- [28] G. Hewitt, D. Jurk, F. D. M. Marques et al., "2012Telomeres are favoured targets of a persistent DNA damage response in ageing and stress-induced senescence," *Nature Communications*, vol. 3, article 708, 2012.
- [29] E. H. Blackburn, "Structure and function of telomeres," *Nature*, vol. 350, no. 6319, pp. 569–573, 1991.
- [30] R. J. O'Sullivan and J. Karlseder, "Telomeres: protecting chromosomes against genome instability," *Nature Reviews Molecular Cell Biology*, vol. 11, no. 3, pp. 171–181, 2010.
- [31] C. Belgiovine, I. Chiodi, and C. Mondello, "Telomerase: cellular immortalization and neoplastic transformation. Multiple functions of a multifaceted complex," *Cytogenetic and Genome Research*, vol. 122, no. 3-4, pp. 255–262, 2009.
- [32] L. Hayflick and P. S. Moorhead, "The serial cultivation of human diploid cell strains," *Experimental Cell Research*, vol. 25, no. 3, pp. 585–621, 1961.
- [33] C. B. Harley, A. B. Futcher, and C. W. Greider, "Telomeres shorten during ageing of human fibroblasts," *Nature*, vol. 345, no. 6274, pp. 458–460, 1990.
- [34] F. D'Adda Di Fagagna, P. M. Reaper, L. Clay-Farrace et al., "A DNA damage checkpoint response in telomere-initiated senescence," *Nature*, vol. 426, no. 6963, pp. 194–198, 2003.
- [35] H. Takai, A. Smogorzewska, and T. De Lange, "DNA damage foci at dysfunctional telomeres," *Current Biology*, vol. 13, no. 17, pp. 1549–1556, 2003.
- [36] S. Bekaert, T. De Meyer, and P. Van Oostveldt, "Telomere attrition as ageing biomarker," *Anticancer Research*, vol. 25, no. 4, pp. 3011–3022, 2005.
- [37] M. T. Hemann, M. A. Strong, L. Y. Hao, and C. W. Greider, "The shortest telomere, not average telomere length, is critical for cell viability and chromosome stability," *Cell*, vol. 107, no. 1, pp. 67–77, 2001.
- [38] Y. Cong and J. W. Shay, "Actions of human telomerase beyond telomeres," *Cell Research*, vol. 18, no. 7, pp. 725–732, 2008.
- [39] D. M. Gordon and J. H. Santos, "The emerging role of telomerase reverse transcriptase in mitochondrial DNA metabolism," *Journal of Nucleic Acids*, vol. 2010, Article ID 390791, 7 pages, 2010.
- [40] S. Ahmed, J. F. Passos, M. J. Birket et al., "Telomerase does not counteract telomere shortening but protects mitochondrial function under oxidative stress," *Journal of Cell Science*, vol. 121, no. 7, pp. 1046–1053, 2008.
- [41] R. J. Chen, C. T. Chu, S. C. Huang, S. N. Chow, and C. Y. Hsieh, "Telomerase activity in gestational trophoblastic disease and placental tissue from early and late human pregnancies," *Human Reproduction*, vol. 17, no. 2, pp. 463–468, 2002.
- [42] H. Chen, W. Wang, Y. Mo et al., "Women with high telomerase activity in luteinised granulosa cells have a higher pregnancy rate during in vitro fertilisation treatment," *Journal of Assisted Reproduction and Genetics*, vol. 28, pp. 797–780, 2011.
- [43] S. Y. Kim, S. P. Lee, J. S. Lee, S. J. Yoon, G. Jun, and Y. J. Hwang, "Telomerase and apoptosis in the placental trophoblasts of growth discordant twins," *Yonsei Medical Journal*, vol. 47, no. 5, pp. 698–705, 2006.
- [44] P. Davy, M. Nagata, P. Bullard, N. S. Fogelson, and R. Allsopp, "Fetal growth restriction is associated with accelerated telomere shortening and increased expression of cell senescence markers in the placenta," *Placenta*, vol. 30, no. 6, pp. 539–542, 2009.
- [45] V. A. Luyckx, C. A. Compston, T. Simmen, and T. F. Mueller, "Accelerated senescence in kidneys of low-birth-weight rats after catch-up growth," *American Journal of Physiology*, vol. 297, no. 6, pp. F1697–F1705, 2009.
- [46] J. L. Tarry-Adkins, M. S. Martin-Gronert, J. H. Chen, R. L. Cripps, and S. E. Ozanne, "Maternal diet influences DNA damage, aortic telomere length, oxidative stress, and

- antioxidant defense capacity in rats," *The FASEB Journal*, vol. 22, no. 6, pp. 2037–2044, 2008.
- [47] J. L. Tarry-Adkins, J. H. Chen, N. S. Smith, R. H. Jones, H. Cherif, and S. E. Ozanne, "Poor maternal nutrition followed by accelerated postnatal growth leads to telomere shortening and increased markers of cell senescence in rat islets," *The FASEB Journal*, vol. 23, no. 5, pp. 1521–1528, 2009.
- [48] L. Liu, M. A. Blasco, J. R. Trimarchi, and D. L. Keefe, "An essential role for functional telomeres in mouse germ cells during fertilization and early development," *Developmental Biology*, vol. 249, no. 1, pp. 74–84, 2002.
- [49] M. J. T. Engelbregt, M. M. van Weissenbruch, C. Popp-Snijders, and H. A. Delemarre-van de Waal, "Delayed first cycle in intrauterine growth-retarded and postnatally undernourished female rats: follicular growth and ovulation after stimulation with pregnant mare serum gonadotropin at first cycle," *Journal of Endocrinology*, vol. 173, no. 2, pp. 297–304, 2002.
- [50] P. Da Silva-Buttkus, R. van den Hurk, E. R. te Velde, and M. A. M. Taverne, "Ovarian development in intrauterine growth-retarded and normally developed piglets originating from the same litter," *Reproduction*, vol. 126, no. 2, pp. 249–258, 2003.
- [51] P. Da Silva, R. P. Aitken, S. M. Rhind, P. A. Racey, and J. M. Wallace, "Impact of maternal nutrition during pregnancy on pituitary gonadotrophin gene expression and ovarian development in growth-restricted and normally grown late gestation sheep fetuses," *Reproduction*, vol. 123, no. 6, pp. 769–777, 2002.
- [52] A. B. Bernal, M. H. Vickers, M. B. Hampton, R. A. Poynton, and D. M. Sloboda, "Maternal undernutrition significantly impacts ovarian follicle number and increases ovarian oxidative stress in adult rat offspring," *PLoS ONE*, vol. 5, no. 12, Article ID e15558, 2010.
- [53] C. A. Brenner, Y. M. Wolny, R. R. Adler, and J. Cohen, "Alternative splicing of the telomerase catalytic subunit in human oocytes and embryos," *Molecular Human Reproduction*, vol. 5, no. 9, pp. 845–850, 1999.
- [54] D. H. Betts and W. A. King, "Telomerase activity and telomere detection during early bovine development," *Developmental Genetics*, vol. 25, no. 4, pp. 397–403, 1999.
- [55] S. Bekaert, H. Derradji, and S. Baatout, "Telomere biology in mammalian germ cells and during development," *Developmental Biology*, vol. 274, no. 1, pp. 15–30, 2004.
- [56] L. Liu, S. Franco, B. Spyropoulos, P. B. Moens, M. A. Blasco, and D. L. Keefe, "Irregular telomeres impair meiotic synapsis and recombination in mice," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 17, pp. 6496–6501, 2004.
- [57] D. L. Keefe, L. Liu, and K. Marquard, "Telomeres and aging-related meiotic dysfunction in women," *Cellular and Molecular Life Sciences*, vol. 64, no. 2, pp. 139–143, 2007.
- [58] T. Hassold and P. Hunt, "To err (meiotically) is human: the genesis of human aneuploidy," *Nature Reviews Genetics*, vol. 2, no. 4, pp. 280–291, 2001.
- [59] J. A. Johnson and S. Tough, "Delayed child-bearing," *JOGC*, vol. 34, pp. 80–93, 2012.
- [60] L. Liu, J. R. Trimarchi, P. J. Smith, and D. L. Keefe, "Mitochondrial dysfunction leads to telomere attrition and genomic instability," *Aging Cell*, vol. 1, no. 1, pp. 40–46, 2002.
- [61] D. H. Betts and P. Madan, "Permanent embryo arrest: molecular and cellular concepts," *Molecular Human Reproduction*, vol. 14, no. 8, pp. 445–453, 2008.
- [62] N. Chernoff, M. I. Gage, T. E. Stoker, R. L. Cooper, M. E. Gilbert, and E. H. Rogers, "Reproductive effects of maternal and pre-weaning undernutrition in rat offspring: age at puberty, onset of female reproductive senescence and intergenerational pup growth and viability," *Reproductive Toxicology*, vol. 28, no. 4, pp. 489–494, 2009.
- [63] Y. Wang, N. Erdmann, R. J. Giannone, J. Wu, M. Gomez, and Y. Liu, "An increase in telomere sister chromatid exchange in murine embryonic stem cells possessing critically shortened telomeres," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 102, no. 29, pp. 10256–10260, 2005.
- [64] N. D. Allen and D. M. Baird, "Telomere length maintenance in stem cell populations," *Biochimica et Biophysica Acta*, vol. 1792, no. 4, pp. 324–328, 2009.
- [65] K. Ito, A. Hirao, F. Arai et al., "Reactive oxygen species act through p38 MAPK to limit the lifespan of hematopoietic stem cells," *Nature Medicine*, vol. 12, no. 4, pp. 446–451, 2006.
- [66] T. Yahata, T. Takanashi, Y. Muguruma et al., "Accumulation of oxidative DNA damage restricts the self-renewal capacity of human hematopoietic stem cells," *Blood*, vol. 118, no. 11, pp. 2941–2950, 2011.
- [67] I. Ligi, S. Simoncini, E. Tellier et al., "A switch toward angiostatic gene expression impairs the angiogenic properties of endothelial progenitor cells in low birth weight preterm infants," *Blood*, vol. 118, no. 6, pp. 1699–1709, 2011.
- [68] P. Aroviita, K. Teramo, V. Hiilesmaa, P. Westman, and R. Kekomäki, "Birthweight of full-term infants is associated with cord blood CD34+ cell concentration," *Acta Paediatrica, International Journal of Paediatrics*, vol. 93, no. 10, pp. 1323–1329, 2004.
- [69] J. W. Meekins, R. Pijnenborg, M. Hanssens, I. R. McFadyen, and A. Van Asshe, "A study of placental bed spiral arteries and trophoblast invasion in normal and severe pre-eclamptic pregnancies," *BJOG*, vol. 101, no. 8, pp. 669–674, 1994.
- [70] H. Nishi, N. Yahata, K. Ohyashiki et al., "Comparison of telomerase activity in normal chorionic villi to trophoblastic diseases," *International Journal of Oncology*, vol. 12, no. 1, pp. 81–85, 1998.
- [71] T. Kudo, T. Izutsu, and T. Sato, "Telomerase activity and apoptosis as indicators of ageing in placenta with and without intrauterine growth retardation," *Placenta*, vol. 21, no. 5-6, pp. 493–500, 2000.
- [72] H. Nishi, T. Nakada, S. Kyo, M. Inoue, J. W. Shay, and K. Isaka, "Hypoxia-inducible factor 1 mediates upregulation of telomerase (hTERT)," *Molecular and Cellular Biology*, vol. 24, no. 13, pp. 6076–6083, 2004.
- [73] G. J. Burton, E. Jauniaux, and D. S. Charnock-Jones, "The influence of the intrauterine environment on human placental development," *International Journal of Developmental Biology*, vol. 54, no. 2-3, pp. 303–311, 2010.
- [74] T. Biron-Shental, R. Sukenik-Halevy, Y. Sharon et al., "Short telomeres may play a role in placental dysfunction in preeclampsia and intrauterine growth restriction," *American Journal of Obstetrics and Gynecology*, vol. 202, no. 4, pp. 381.e1–381.e7, 2010.
- [75] R. Lehner, J. Bobak, N. W. Kim, A. L. Shroyer, and K. R. Shroyer, "Localization of telomerase hTERT protein and survivin in placenta: relation to placental development and hydatidiform mole," *Obstetrics and Gynecology*, vol. 97, no. 6, pp. 965–970, 2001.
- [76] R. Allsopp, J. Shimoda, D. Easa, and K. Ward, "Long telomeres in the mature human placenta," *Placenta*, vol. 28, no. 4, pp. 324–327, 2007.



- [77] Z. Hracsko, H. Orvos, Z. Novak, A. Pal, and I. S. Varga, "Evaluation of oxidative stress markers in neonates with intrauterine growth retardation," *Redox Report*, vol. 13, no. 1, pp. 11–16, 2008.
- [78] S. E. Pinney and R. A. Simmons, "Epigenetic mechanisms in the development of type 2 diabetes," *Trends in Endocrinology and Metabolism*, vol. 21, no. 4, pp. 223–229, 2010.
- [79] S. Sebert, D. Sharkey, H. Budge, and M. E. Symonds, "The early programming of metabolic health: is epigenetic setting the missing link?" *American Journal of Clinical Nutrition*, vol. 94, no. 6, pp. 1953S–1958S, 2011.
- [80] P. Gupta, M. Narang, B. D. Banerjee, and S. Basu, "Oxidative stress in term small for gestational age neonates born to undernourished mothers: a case control study," *BMC Pediatrics*, vol. 4, article 14, 2004.
- [81] M. Saker, N. Soulimane Mokhtari, S. A. Merzouk, H. Merzouk, B. Belarbi, and M. Narce, "Oxidant and antioxidant status in mothers and their newborns according to birthweight," *European Journal of Obstetrics Gynecology and Reproductive Biology*, vol. 141, no. 2, pp. 95–99, 2008.
- [82] M. Z. H. Howlader, S. Parveen, S. Tamanna, T. A. Khan, and F. Begum, "Oxidative stress and antioxidant status in neonates born to pre-eclamptic mother," *Journal of Tropical Pediatrics*, vol. 55, no. 6, pp. 363–367, 2009.
- [83] R. Negi, D. Pande, A. Kumar, R. S. Khanna, and H. D. Khanna, "Evaluation of biomarkers of oxidative stress and antioxidant capacity in the cord blood of preterm low birth weight neonates," *Journal of Maternal-Fetal and Neonatal Medicine*, vol. 25, no. 8, pp. 1338–1341, 2012.
- [84] M. Baserga, C. Bertolotto, N. K. MacLennan et al., "Uteroplacental insufficiency decreases small intestine growth and alters apoptotic homeostasis in term intrauterine growth retarded rats," *Early Human Development*, vol. 79, no. 2, pp. 93–105, 2004.
- [85] T. D. Pham, N. K. MacLennan, C. T. Chiu, G. S. Laksana, J. L. Hsu, and R. H. Lane, "Uteroplacental insufficiency increases apoptosis and alters p53 gene methylation in the full-term IUGR rat kidney," *American Journal of Physiology*, vol. 285, no. 5, pp. R962–R970, 2003.
- [86] E. Migliaccio, M. Gioglio, S. Mele et al., "The p66(shc) adaptor protein controls oxidative stress response and life span in mammals," *Nature*, vol. 402, no. 6759, pp. 309–313, 1999.
- [87] B. J. Jennings, S. E. Ozanne, M. W. Dorling, and C. N. Hales, "Early growth determines longevity in male rats and may be related to telomere shortening in the kidney," *FEBS Letters*, vol. 448, no. 1, pp. 4–8, 1999.
- [88] S. Entringer, E. S. Epel, R. Kumsta et al., "Stress exposure in intrauterine life is associated with shorter telomere length in young adulthood," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 33, pp. E513–E518, 2011.
- [89] E. Chang and C. B. Harley, "Telomere length and replicative aging in human vascular tissues," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 92, no. 24, pp. 11190–11194, 1995.
- [90] K. Okuda, M. Y. Khan, J. Skurnick, M. Kimura, H. Aviv, and A. Aviv, "Telomere attrition of the human abdominal aorta: relationships with age and atherosclerosis," *Atherosclerosis*, vol. 152, no. 2, pp. 391–398, 2000.
- [91] S. Demissie, D. Levy, E. J. Benjamin et al., "Insulin resistance, oxidative stress, hypertension, and leukocyte telomere length in men from the Framingham Heart Study," *Aging Cell*, vol. 5, no. 4, pp. 325–330, 2006.
- [92] A. L. Fitzpatrick, R. A. Kronmal, J. P. Gardner et al., "Leukocyte telomere length and cardiovascular disease in the cardiovascular health study," *American Journal of Epidemiology*, vol. 165, no. 1, pp. 14–21, 2007.
- [93] S. Bekaert, T. De Meyer, E. R. Rietzschel et al., "Telomere length and cardiovascular risk factors in a middle-aged population free of overt cardiovascular disease," *Aging Cell*, vol. 6, no. 5, pp. 639–647, 2007.
- [94] K. D. Salpea, P. J. Talmud, J. A. Cooper et al., "Association of telomere length with type 2 diabetes, oxidative stress and UCP2 gene variation," *Atherosclerosis*, vol. 209, no. 1, pp. 42–50, 2010.
- [95] R. Testa, F. Olivieri, C. Sirolla et al., "Leukocyte telomere length is associated with complications of Type 2 diabetes mellitus," *Diabetic Medicine*, vol. 28, no. 11, pp. 1388–1394, 2011.
- [96] F. Fyhrquist, K. Silventoinen, O. Saijonmaa et al., "Telomere length and cardiovascular risk in hypertensive patients with left ventricular hypertrophy: the LIFE study," *Journal of Human Hypertension*, 2011.
- [97] J. Campisi, "Senescent cells, tumor suppression, and organismal aging: good citizens, bad neighbors," *Cell*, vol. 120, no. 4, pp. 513–522, 2005.
- [98] D. J. Baker, T. Wijshake, T. Tchkonja et al., "Clearance of p16 Ink4a-positive senescent cells delays ageing-associated disorders," *Nature*, vol. 479, no. 7372, pp. 232–236, 2011.
- [99] B. J. Heidinger, J. D. Blount, W. Boner, K. Griffiths, N. B. Metcalfe, and P. Monaghan, "Telomere length in early life predicts lifespan," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 109, no. 5, pp. 1743–1748, 2012.
- [100] M. S. Martin-Gronert, J. L. Tarry-Adkins, R. L. Cripps, J. H. Chen, and S. E. Ozanne, "Maternal protein restriction leads to early life alterations in the expression of key molecules involved in the aging process in rat offspring," *American Journal of Physiology*, vol. 294, no. 2, pp. R494–R500, 2008.
- [101] Z. Song, G. von Figura, Y. Liu et al., "Lifestyle impacts on the aging-associated expression of biomarkers of DNA damage and telomere dysfunction in human blood," *Aging Cell*, vol. 9, no. 4, pp. 607–615, 2010.
- [102] M. Fumagalli, F. Rossiello, M. Clerici et al., "Telomeric DNA damage is irreparable and causes persistent DNA-damage-response activation," *Nature Cell Biology*, vol. 14, no. 4, pp. 355–365, 2012.
- [103] E. Sahin, S. Colla, M. Liesa et al., "Telomere dysfunction induces metabolic and mitochondrial compromise," *Nature*, vol. 470, no. 7334, pp. 359–365, 2011.
- [104] V. W. Dolinsky, C. F. Rueda-Clausen, J. S. Morton, S. T. Davidge, and J. R. B. Dyck, "Continued postnatal administration of resveratrol prevents diet-induced metabolic syndrome in rat offspring born growth restricted," *Diabetes*, vol. 60, no. 9, pp. 2274–2284, 2011.





# Hindawi

Submit your manuscripts at  
<http://www.hindawi.com>

