

Research Article

Comparison of Ligation-Mediated PCR Methods in Differentiation of *Mycobacterium tuberculosis* Strains

Anna Zaczek,¹ Anna Brzostek,² Arkadiusz Wojtasik,²
Anna Sajduda,³ and Jaroslaw Dziadek^{1,2}

¹ Department of Biochemistry and Cell Biology, University of Rzeszow, 35-601 Rzeszow, Poland

² Institute of Medical Biology, Polish Academy of Science, 93-232 Lodz, Poland

³ Department of Microbial Genetics, Faculty of Biology and Environmental Protection, University of Lodz, 90-237 Lodz, Poland

Correspondence should be addressed to Jaroslaw Dziadek; jdziadek@cbm.pan.pl

Received 20 November 2013; Accepted 8 January 2014; Published 16 February 2014

Academic Editor: Tomasz Jagielski

Copyright © 2014 Anna Zaczek et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Fast and inexpensive identification of epidemiological links between limited number of *Mycobacterium tuberculosis* strains is required to initially evaluate hospital outbreaks, laboratory crosscontaminations, and family or small community transmissions. The ligation-mediated PCR methods (LM-PCR) appear sufficiently discriminative and reproducible to be considered as a good candidate for such initial, epidemiological analysis. Here, we compared the discriminative power of the recently developed in our laboratory fast ligation amplification polymorphism (FLAP) method with fast ligation-mediated PCR (FLiP). Verification of the results was based on analyzing a set of reference strains and RFLP-IS6110 typing. The HGDI value was very similar for both LM-PCR methods and RFLP-IS6110 typing. However, only 52% of strains were correspondingly grouped by both FLiP and FLAP methods. Differentiation by FLAP method demonstrated a limited similarity to IS6110-RFLP (37,7%). As much as 78,7% of strains were grouped identically when differentiated by FLiP and IS6110-RFLP methods. The analysis differentiated 31, 35, and 36 groups when using FLAP, FLiP, and RFLP-IS6110 methods, respectively.

1. Introduction

Recent development of molecular methods has substantially improved the identification of many bacterial pathogens, both at the species and strain levels. *M. tuberculosis*, the causative agent of tuberculosis, is still one of the most dangerous human pathogens causing high morbidity and mortality worldwide. The genetic typing of mycobacteria has greatly improved knowledge about tuberculosis epidemiology and enabled a molecular-guided control of the disease. Various genetic markers are used in molecular epidemiology of tuberculosis. In particular, identification of repeated sequences in mycobacterial genome and their analysis at molecular level allowed to develop the intraspecies discrimination methods for mycobacteria [1].

The current international standard for epidemiological typing of *M. tuberculosis* is restriction fragment length polymorphism (RFLP) based on the detection of variability in the number of copies and chromosomal locations of IS6110

insertion sequences [2–5]. The second widely used method is mycobacterial interspersed repetitive unit-variable number of tandem repeats typing (MIRU-VNTR) based on variable number of tandem repeats [6]. Finally spoligotyping (spacer oligonucleotide typing) based on polymorphism in the chromosomal direct repeat (DR) locus is often used as a fast screening method [7]. Interesting alternative for the methods mentioned above are those based on ligation-mediated PCR (LM-PCR), which have proven useful in epidemiological analysis of a number of bacterial species [8–10]. Such methods can be adapted to mycobacterial typing when they are based on variability in IS6110 flanking regions [11–15].

Here, we assess the usefulness of a recently described in our group LM-PCR method, termed fast ligation amplification polymorphism (FLAP), for differentiation of *M. tuberculosis* strains [15]. We present the results of its application in context of published results of reference set [14, 16] and compare its discriminatory power to that of IS6110-RFLP and FLiP (fast ligation-mediated PCR) methods.

2. Materials and Methods

2.1. Bacterial Strains. The 61 strains used in this analysis were obtained in 2006-2007 from patients hospitalized in the Center for Lung Diseases Treatment and Rehabilitation in Lodz, Poland. All strains were tested for susceptibility to isoniazid, rifampicin, pyrazinamide, streptomycin, and ethambutol using the Bactec 460 TB system (BD Diagnostic Systems, Sparks, MD, USA), as described previously [16]. This set of strains was previously characterized by IS6110-RFLP analysis, 15 locus MIRU-VNTR typing, and spoligotyping [16].

2.2. DNA Preparation. Genomic DNA was extracted and purified from all the isolates using the protocol by van Embden et al. [2]. The concentration of DNA was measured with NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies, Montchanin, DE, USA).

2.3. The FLiP Method. The FLiP analysis was performed as originally described by Reisig et al. [13]. Briefly, the method is based on the ligation of oligonucleotide adaptors. Following restriction digestion, genomic DNA is ligated with an adaptor composed of two oligonucleotides, one of which is complementary to the end created by restrictase, while the other contains uracil instead of thymine. A pair of starters is used for amplification; one of them is specific to the IS6110 sequence and the other is complementary to the oligonucleotide ligated with restricted genomic DNA fragments. Amplification products are analyzed using electrophoresis; the obtained band patterns are strain specific.

2.4. The FLAP Method. The FLAP method was performed as we previously described [15]. Briefly, genomic DNA of *M. tuberculosis* strains was digested with *PvuII* and *SalI* restriction enzymes. The *PvuII* endonuclease recognizes a single nucleotide sequence within IS6110 and generates blunt ends. After the digestion step, oligonucleotide adaptors (36 and 40 nucleotides in length) are ligated to *SalI* cohesive ends. All restriction fragments are used as templates for PCR amplification, with one primer complementary to adaptor sequence and the second primer complementary to the inner fragment of IS6110. The PCR products were separated on acrylamide gels and visualized by UV light illumination to generate the FLAP patterns.

2.5. Clustering and Computer Analysis. The fingerprint patterns obtained by both methods were analysed by using BioNumerics software, version 5.0 (Applied Maths, Sint-Martens-Latem, Belgium). Dendrograms were generated based on the Unweighted Pair Group Method with Arithmetic Averages (UPGMA) algorithm for clustering and Dice similarity coefficient. The Hunter-Gaston discriminatory index (HGDI) was calculated as previously described and used to evaluate the discriminatory power of the typing methods [17].

3. Results and Discussion

The “gold standard” method for epidemiological typing of *M. tuberculosis* is IS6110-RFLP analysis providing the best resolution at the population level [2–5]. It seems obvious that appropriate epidemiological analyses of *M. tuberculosis* clinical strains should be based on more than one molecular method. Recently developed PCR-based genotyping methods are rapid, do not require a large quantity of purified DNA, and provide reproducible digital results. In particular, the relatively novel ligation-mediated PCR FLiP and FLAP typing methods seem to be promising alternatives for genotyping of *M. tuberculosis*, as well as for the detection of genotypic heterogeneity, mixed infection, and crosscontamination of mycobacterial samples [11–15].

In this study, we compared the results obtained by FLAP for 61 strains of *M. tuberculosis*, isolated from TB patients in 2006-2007, with both FLiP and IS6110 RFLP methods, and estimated their discriminatory power by HGDI. All the methods are based on detection of insertion sequence IS6110.

The FLAP analysis subdivided the 61 analyzed strains into 31 clusters; 13 of which demonstrated unique patterns. The remaining 48 strains were grouped into two clusters of 5 strains each, three clusters of 4 strains each, and thirteen clusters of 2 strains each (Table 1).

The total of 35 FLiP patterns were detected and distributed in 14 clusters within 40 strains (65,6%) and 21 unique patterns (34,4%). One cluster consisted of 8 strains with identical FLiP pattern, three clusters contained 5, 4, and 3 strains, respectively, while eleven clusters comprised 2 strains each (Table 1).

The discriminatory power of the FLAP typing for the 61 *M. tuberculosis* isolates, calculated as HGDI, was 0.9757 compared to 0.9713 for FLiP method.

Previously performed analysis by reference methods, IS6110-RFLP and MIRU-VNTR typing, grouped this set of strains into 36, for IS6110-RFLP, and 27 patterns, for MIRU-VNTR, and gave the resolving power, 0.9743 and 0.9697, respectively [16].

We observed differences in the number of copies of IS6110 when determined by reference method IS6110 RFLP and LM-PCR methods (FLiP and FLAP). Therefore minimal and maximal number of bands in particular patterns were calculated. The copy number of IS6110, as determined by the reference method in each of 61 strains ranged from 6 to 14. The majority, 48 (78,7%) of the strains, contained 8–12 copies. FLAP patterns obtained for the same set of strains varied from 4 to 12, with 51 (83,6%) of strains containing 7–9 bands in pattern. In contrast, the number of IS6110 in FLiP patterns varied from 3 to 8, and 49 (80,3%) of strains possessed 6–8 bands in pattern.

We found that FLAP patterns of 46 strains (75,4%) possessed more bands in particular than when analyzed by FLiP, 5 strains (8,2%) consisted of less bands, and 10 (16,4%) of strains contained the same number of bands in both methods. Next, we compared band patterns between FLAP and IS6110 RFLP methods. 41 strains (67,2%) possessed less bands than when analyzed by IS6110 RFLP typing; in eight patterns (13,1%) we observed more bands while 12 (19,7%) strains contained identical number of band patterns. Comparison of

TABLE 1: The comparison of 61 clustered strains by FLAP, IS6110-RFLP, and FLiP analysis.

	Strain	FLAP	IS6110-RFLP	FLiP
1	50/8	F.1	R.1	1
2	674/7		R.2	
3	149/8	F.2	R.3	2
4	146/7	F.3	R.4	3
5	319/7		R.5	
6	19/7	F.4		4
7	118/7		R.6	5
8	126/7		R.7	6
9	147/8	F.5	R.8	7
10	54/8		R.9	
11	102/7	F.6	R.10	
12	176/7		R.11	8
13	171/8	F.7	R.12	
14	412/7		R.13	9
15	129/7	F.8	R.14	
16	41/7		R.15	10
17	216/8	F.9	R.16	
18	307/7		R.17	11
19	165	F.10	R.18	12
20	218/8	F.11	R.19	13
21	230	F.12	R.20	
22	9/7		R.21	14
23	84	F.13	R.22	15
24	321	F.14		
25	65/7	F.15		
26	632			
27	179/8			16
28	386/7			
29	391/7	F.16		
30	611			
31	564			
32	152/7		R.18	17
33	690/7		R.19	18
34	34/7	F.17		
35	549/7			
36	550		R.20	19
37	567/7	F.18		
38	696			
39	232/8	F.19	R.21	20
40	571/7		R.22	
41	565	F.20		21

TABLE 1: Continued.

	Strain	FLAP	IS6110-RFLP	FLiP
42	80/7	F.21	R.23	22
43	490/7	F.22	R.24	23
44	704	F.23		24
45	222		R.25	25
46	237/8	F.24	R.26	26
47	253			
48	1/7			
49	10/7	F.25	R.27	27
50	601			
51	120/7		R.28	
52	723	F.26	R.29	28
53	91/8		R.30	29
54	459	F.27	R.31	30
55	306/7		R.32	
56	725	F.28		31
57	724		R.33	
58	108/8		R.34	32
59	671	F.29	R.35	33
60	305/7	F.30	R.36	34
61	37/7	F.31		35

The identical clusters determined in both FLiP and FLAP methods are marked in bold.

patterns obtained by FLiP and IS6110 RFLP typing revealed lower number of bands in 56 strains (91,8%) analyzed by FLiP when compared to IS6110 RFLP, and 2 strains (3,3%) possessed higher number of bands, and 3 (4,9%) strains presented identical number of bands in these two methods. The intrinsically lower band numbers of FLiP patterns might contribute to slightly lower discriminatory power of this method in comparison to IS6110 RFLP. The results indicate that the different number of bands in FLiP and FLAP DNA fingerprints may not necessarily reflect the number of IS6110 copies in the strains tested. Moreover, the observed differences could be caused by technical difficulties, including mistakes in interpretation of visualized bands and insufficient separation of PCR fragments in polyacrylamide gel, or by difficulties of the PCR amplifying large products itself. It has been shown that PCR-based DNA fingerprinting patterns could result from nonspecific amplification of some of the products [13]. However, the analysis of identically clustered strains clearly showed the highest concordance for the FLiP and IS6110 RFLP methods (48 strains, 78,7%). Both FLAP and FLiP methods allowed to group 32 strains (52,4%), whereas the lowest clustering identity was observed for FLAP and IS6110 RFLP methods for 23 strains (37,7%).

4. Conclusions

In summary, the LM-PCR methods proved to be effective and reproducible for the differentiation of *M. tuberculosis* strains, showing high discriminatory power comparable to that of the IS6110 RFLP. Based on the previous and present results, the LM-PCR methods seem to be a valuable alternative (highly discriminating, inexpensive, and very fast) to the widely applied and standardized IS6110 RFLP method. However, the main limitation of the PCR-based methods, including LM-PCR, is the incapability in construction of reference database, especially containing strain patterns of isolates from different laboratories. Therefore, these methods might be rather used as a second-line test for verification of epidemiological links and could be valuable molecular epidemiology tools for analyzing collections with a limited number of strains. Nevertheless, based on our results, it is clear that it is necessary to apply more than one PCR-based method simultaneously.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This work was supported by a Grant from the National Science Centre (NCN; Project no. N N302 111338) and by a Grant from the European Regional Development Fund (POIG.01.01.02-10-107/09) under the Operational Programme Innovative Economy. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the paper.

References

- [1] B. Mathema, N. E. Kurepina, P. J. Bifani, and B. N. Kreiswirth, "Molecular epidemiology of tuberculosis: current insights," *Clinical Microbiology Reviews*, vol. 19, no. 4, pp. 658–685, 2006.
- [2] J. D. A. van Embden, M. D. Cave, J. T. Crawford et al., "Strain identification of *Mycobacterium tuberculosis* by DNA fingerprinting: recommendations for a standardized methodology," *Journal of Clinical Microbiology*, vol. 31, no. 2, pp. 406–409, 1993.
- [3] P. F. Barnes and M. D. Cave, "Molecular epidemiology of tuberculosis," *New England Journal of Medicine*, vol. 349, no. 12, pp. 1149–1156, 2003.
- [4] J. T. Crawford, "Genotyping in contact investigations: a CDC perspective," *International Journal of Tuberculosis and Lung Disease*, vol. 7, no. 12, pp. S453–S457, 2003.
- [5] K. Kremer, C. Arnold, A. Cataldi et al., "Discriminatory power and reproducibility of novel DNA typing methods for *Mycobacterium tuberculosis* complex strains," *Journal of Clinical Microbiology*, vol. 43, no. 11, pp. 5628–5638, 2005.
- [6] P. Supply, C. Allix, S. Lesjean et al., "Proposal for standardization of optimized mycobacterial interspersed repetitive unit-variable-number tandem repeat typing of *Mycobacterium tuberculosis*," *Journal of Clinical Microbiology*, vol. 44, no. 12, pp. 4498–4510, 2006.
- [7] L. S. Cowan, L. Diem, T. Monson et al., "Evaluation of a two-step approach for large-scale, prospective genotyping of *Mycobacterium tuberculosis* isolates in the United States," *Journal of Clinical Microbiology*, vol. 43, no. 2, pp. 688–695, 2005.
- [8] A. Masny and A. Plucienniczak, "Ligation mediated PCR performed at low denaturation temperatures—PCR melting profiles," *Nucleic Acids Research*, vol. 31, no. 18, article e114, 2003.
- [9] B. Krawczyk, A. Samet, J. Leibner, A. Śledzińska, and J. Kur, "Evaluation of a PCR melting profile technique for bacterial strain differentiation," *Journal of Clinical Microbiology*, vol. 44, no. 7, pp. 2327–2332, 2006.
- [10] B. Krawczyk, J. Leibner-Ciszak, K. Stojowska, and J. Kur, "The new LM-PCR/Shifter method for the genotyping of microorganisms based on the use of a class IIS restriction enzyme and ligation-mediated PCR," *Journal of Microbiology and Biotechnology*, vol. 21, no. 12, pp. 1336–1344, 2011.
- [11] G. Prod'Hom, C. Guillhot, M. C. Gutierrez, A. Varnerot, B. Gicquel, and V. Vincent, "Rapid discrimination of *Mycobacterium tuberculosis* complex strains by ligation-mediated PCR fingerprint analysis," *Journal of Clinical Microbiology*, vol. 35, no. 12, pp. 3331–3334, 1997.
- [12] W. H. Haas, W. R. Butler, C. L. Woodley, and J. T. Crawford, "Mixed-linker polymerase chain reaction: a new method for rapid fingerprinting of isolates of the *Mycobacterium tuberculosis* complex," *Journal of Clinical Microbiology*, vol. 31, no. 5, pp. 1293–1298, 1993.
- [13] F. Reisig, K. Kremer, B. Amthor, D. Van Soolingen, and W. H. Haas, "Fast ligation-mediated PCR, a fast and reliable method for IS6110-based typing of *Mycobacterium tuberculosis* complex," *Journal of Clinical Microbiology*, vol. 43, no. 11, pp. 5622–5627, 2005.
- [14] A. Zaczek, M. Ziolkiewicz, A. Wojtasik, J. Dziadek, and A. Sajduda, "IS6110-based differentiation of *Mycobacterium tuberculosis* strains," *Polish Journal of Microbiology*, vol. 62, no. 2, pp. 201–204, 2013.
- [15] A. Zaczek, A. Brzostek, and A. Kuron, "Development of a new ligation-mediated PCR method for the differentiation of *Mycobacterium tuberculosis* strains," *International Journal of Tuberculosis and Lung Disease*, vol. 18, no. 3, pp. 302–309, 2014.
- [16] M. Krawczyk, A. Brzostek, A. Gorna et al., "Epidemiological analysis of *Mycobacterium tuberculosis* strains isolated in Lodz, Poland," *International Journal of Tuberculosis and Lung Disease*, vol. 15, no. 9, pp. 1252–1258, 2011.
- [17] P. R. Hunter and M. A. Gaston, "Numerical index of the discriminatory ability of typing systems: an application of Simpson's index of diversity," *Journal of Clinical Microbiology*, vol. 26, no. 11, pp. 2465–2466, 1988.



Hindawi

Submit your manuscripts at
<http://www.hindawi.com>

