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1	Soil carbon stocks in tropical forests regulated by base cation effects on fine roots
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1	Tropical forests are the most carbon (C)-rich ecosystems on Earth, containing 25-
2	40% of global terrestrial C stocks (1, 2). While large-scale quantification of aboveground
3	biomass in tropical forests has improved recently (3), soil C dynamics remain one of the
4	largest sources of uncertainty in Earth system models (4), inhibiting our ability to predict
5	future climate (5). Globally, soil texture and climate predict < 30% of the variation in soil
6	C stocks (1), so ecosystem models often predict soil C stocks using measures of
7	aboveground plant growth (4). However, this approach underestimates tropical soil C
8	stocks, and is inaccurate compared with estimates for data-rich northern ecosystems (6).
9	By quantifying soil organic C stocks to 1 m depth for 48 humid tropical forest plots across
10	gradients of rainfall and soil fertility in Panama, we show that soil C across these diverse
11	tropical forests does not correlate with common predictors used in models, such as plant
12	biomass or litter production. Instead, a structural equation model including base cations,
13	soil clay content, and rainfall as exogenous factors and root biomass as an endogenous
14	factor predicted nearly 50% of the variation in tropical soil C stocks, indicating a strong
15	indirect effect of base cation availability on tropical soil C. The importance of cations
16	shown here expands on long-standing work showing that phosphorus (P) commonly
17	controls aboveground plant growth in tropical forests (7, 8). Including soil cations in C
18	cycle models, and thus emphasizing mechanistic links among nutrients, root biomass, and
19	soil C stocks, will improve prediction of climate-soil feedbacks in tropical forests.
20	

21 Significance Statement

Tropical forest soils contain some of the largest stocks of carbon on Earth, making them of broad importance in the global carbon cycle. Nonetheless, our understanding of what drives variation in soil carbon stocks across tropical landscapes is limited, inhibiting our ability to predict large-scale responses to global change. This study revealed a strong effect of soil base cations on soil C stocks via changes in root biomass. In contrast, aboveground plant growth was not related to soil C stocks, contrary to ecosystem model assumptions. The 48 Panamanian forests included here covered a two-fold range in rainfall, five soil orders, 25 geological formations, and a 20-fold range nutrient availability, likely making our results applicable to a much broader geographic range.

8

1 Introduction

2 Tropical forest soils contain some of the largest stocks of carbon on Earth, making them of broad importance in the global carbon (C) cycle (1, 2). Aboveground C stocks and litterfall in 3 4 closed-canopy tropical forests can be measured with precision at large scales using remote 5 sensing (3), so extrapolating soil C stocks from aboveground measures with few *in situ* soil data has formed the basis for many modeling efforts (4). However, aboveground plant metrics can be 6 poor predictors of soil C stocks globally, although there are few examples for lowland tropical 7 forests (6). While major increases in plant litterfall have been shown to influence C stocks at the 8 9 soil surface in tropical forests (9), most long-term C storage within mineral soils probably originates from root biomass (10). Particularly in humid tropical forests, decomposition of leaf 10 litter on the forest floor is rapid (<3 years) (11), and a large proportion of surface litter is likely 11 respired back to the atmosphere as CO₂ rather than transported downward and stored in mineral 12 soils. Root C, in contrast, is transferred directly into mineral soils, where tough, lignin-rich 13 tissues, saturated conditions, and low oxygen levels can slow decomposition, leading to greater C 14 retention of root tissues relative to leaf litter (12, 13). Thus, estimates of root biomass, rather than 15 aboveground C stocks, are likely to be more useful for predicting tropical soil C stocks. 16 However, tropical forest root biomass is notoriously difficult to quantify, spatially variable, and 17 is not easily predictable as a proportion of aboveground biomass (14). 18 An alternative and widely-used approach for predicting soil C stocks at broad scales 19 20 relies on commonly measured abiotic properties, particularly climate and soil texture (1, 15).

For example, global-scale data show that soil C pools are generally greater in wetter forests (15).
A relationship between rainfall and soil C storage within tropical forests would be of particular
interest in the context of regional drying that is expected with climate change (16). Among soil

characteristics, clay content has been identified as particularly linked to soil C stocks, likely
 because of the importance of organo-mineral associations for soil C storage (17). An advantage
 of using abiotic properties like clay content to predict soil C is that data for soil texture are
 widely available, whereas root biomass data are relatively scarce (18).

5 An improved approach for predicting tropical soil C stocks may be to incorporate 6 metrics that are strongly correlated to root biomass into the existing abiotic framework. In particular, soil phosphorus (P) and cations availability are commonly shown to limit plant growth 7 in highly weathered tropical soils, which are leached of rock-derived nutrients and rich in 8 9 nitrogen (N) (7, 8, 19). Root biomass in particular has been inversely related to soil P and/or base cation availability across a range of distinct tropical forests (20, 21). Similarly, root biomass and 10 root growth in highly weathered tropical soils have shown consistent declines in response to 11 elevated potassium (K) over fifteen years of factorial nutrient fertilizations in Panama (22, 23). 12 These landscape-scale and experimental declines in root biomass in response to the availability 13 of rock-derived nutrients likely indicate decreased plant C allocation to root biomass when 14 nutrients are plentiful (24), which in turn reduces C inputs to soils. Root exudates are also 15 increasingly recognized as an important contribution of root C to belowground stocks. Root 16 17 exudates are produced as a plant strategy to promote the release of mineral nutrients via stimulated microbial decomposition and/or direct solubilization (25). Increases in rock-derived 18 nutrient availability in tropical soils appear to suppress root exudates (26), similar to trends for 19 20 root biomass, further decreasing root inputs to belowground C stocks. Thus, rock-derived nutrient concentrations might be a useful indicator of plant allocation to root activity in tropical 21 22 forests, so incorporating base cations into predictive models could improve predictions of soil C 23 stocks.

1	We studied 48 lowland tropical forest sites across the Isthmus of Panama to assess factors
2	driving soil C stocks in tropical forests. The forests have a tropical monsoon climate and mean
3	annual temperature of 26° C. The wetter Caribbean coast has a shorter dry season (~115 days),
4	compared with the drier Pacific coast (~150 days). The sites cover extensive geological variation
5	(Figure S1, Table S2) across a steep rainfall gradient (1756–3280 mm mean annual precipitation,
6	MAP). Variation in soil nutrient availability across these sites corresponds primarily to shifts in
7	geology, and soil properties are weakly or not at all correlated with rainfall (27), providing an
8	opportunity to isolate the influence of rainfall from soil properties. We quantified total soil C, N,
9	and P, resin-extractable P, extractable base cations, pH, soil texture and bulk density to 1 m
10	depth in 1-ha plots at each site. Fine root biomass (<2 mm diameter) was sorted by hand. Soils
11	were classified according to Soil Taxonomy (28), which provides an indication of soil
12	weathering status. Above ground biomass for all trees > 10 cm diameter at breast height was
13	calculated using published allometry (29). Litterfall biomass was collected at a subset of 8 sites
14	biweekly for one year (details in Methods and Materials).
15	We hypothesized that soil C stocks in tropical forests correspond to belowground
16	dynamics, not aboveground plant growth. Specifically, we hypothesized that root biomass, soil P
17	and cation availability, and rainfall predict soil C stocks across tropical forests.
18	
19	Results
20	Overall, soil C to 1 m depth ranged from $7.3 - 20.3$ kg C m ⁻² across the sites, with an
21	overall average of 13.3 ± 0.5 kg C m ⁻² . This average value for soil C stocks is between published
22	averages to 1 m depths for similar forest types of 15.8 kg C m ⁻² (ref. ⁴), and 11.5 kg C m ⁻² (ref.
23	(15)). The top 50 cm of soil contained an average of $75 \pm 1\%$ of soil C across sites, with a greater

concentration of C near the surface in less-weathered soil orders (Table 1). There was an even
greater concentration of root biomass in the top 50 cm of soil (93 ± 0.6%), with no variation
among soil orders (Table 1), suggesting that leaching of dissolved organic C from the soil
surface and rooting zone contributed to deeper soil C stocks.

5 In addition to the nearly three-fold change in soil C stock, the sites used here covered five soil orders (Table 1), over 25 geological formations (Figure S1), a two-fold range in rainfall, a 6 20-fold range in base cations, and a >100-fold range in available P (Table S2). Thus, although 7 the data come from a relatively restricted geographic region, the diversity of environmental 8 9 conditions means that the results are likely to be broadly applicable over much larger geographical ranges. The more strongly weathered soils were poorer in rock-derived nutrients, 10 more acidic, and contained greater fine root biomass and soil C stocks. However, most soil 11 characteristics were not clearly distinct among the orders (Table 1), indicating the importance of 12 using soil metrics like base cation availability rather than soil order in predictive efforts. 13 Across the 48 diverse tropical forest sites, aboveground biomass and litterfall were not 14 significant factors for predicting soil C stocks using a stepwise general linear model selection 15 process, which initially included all soil, aboveground, and climate data. Neither aboveground 16 biomass nor litterfall had a significant univariate correlation with soil C (Figures 1a & S3a). 17 Instead, soil clay content, root biomass, and precipitation were significant factors in the 18 minimum adequate general linear model for predicting soil C stocks to 1m (Table S4a). Soil clay 19 20 content (Figure 1b) and fine root biomass (Figure 1c) had the strongest univariate relationships with soil C to 1 m, and also explained the majority of the variation in the minimum adequate 21 general linear model (Table S4a). Mean annual precipitation alone was only a weak positive 22

predictor of soil C ($R^2 = 0.10$, n = 45, p < 0.05, Figure S5), despite the nearly two fold difference

1	in rainfall across the sites. Predictors of soil C stocks to 0.5 m were similar, except that soil clay
2	content was not a significant factor, and root biomass explained an even larger portion of the
3	variation in the minimum adequate general linear model (Table S4b). Rainfall was a much
4	stronger predictor of cumulative annual litterfall (Figure S3b), and especially of dry season
5	litterfall (Figure S3c), showing significant negative relationships. Soil nutrients were not
6	significant factors in the general linear model for predicting soil C stocks. However, a similar
7	model selection process indicated that root biomass to 1 m was most strongly predicted by total
8	extractable base cations, showing a logarithmic decline (Figure 1d).
9	Structural equation modeling was then used to infer mechanistic and directional
10	relationships among all of the factors, and particularly to explore indirect linkages between soil
11	nutrients and soil C stocks. Soil base cations, clay content, and rainfall as exogenous factors,
12	with root biomass included as an endogenous factor linking base cations to soil C stocks,
13	comprised the most parsimonious structural equation model for the data (Figure 2). Thus, the
14	influence of base cations on soil C was mediated via effects on root biomass. This model
15	explained nearly 50% of the variation in soil C stocks across the 48 sites.
16	

17 Discussion

It was surprising that soil C was only weakly correlated with precipitation across these sites, despite a nearly two-fold change in rainfall over the gradient. This suggests that rainfall may not be an important factor driving differences in soil C stocks among tropical forests once soil fertility is accounted for, even when leaf litterfall is strongly responsive to rainfall. Similarly, five years of irrigation at a site near the middle of the Panama rainfall gradient did not result in changing root biomass or root turnover rates (30). However, a study in seasonal Amazonian

forests across soil fertility gradients showed decreased root biomass with declining soil moisture,
both within sites due to seasonality, and between sites due to experimental drying (31). These
contrasting results across observational and manipulative studies indicate that the potential
effects of climatic drying on root activity, and links to soil C storage, merit further mechanistic
research in tropical forests.

Base cations emerged here as a major driver of tropical forest root dynamics, with greater 6 investment in fine root biomass in cation-poor soils to maximize acquisition of these scarce 7 nutrients (24). This result highlights the benefit of including rock-derived nutrients as a driver of 8 9 plant and soil dynamics in tropical forest ecosystem models. Data on soil cations, as well as texture, are available for many international sites (18), and can be estimated from geological 10 information and rainfall where data is unavailable (27, 32). Interestingly, no single rock-derived 11 nutrient was strongly correlated with root biomass or soil C across the sites. This contrasts with a 12 strong effect of K fertilization on plant growth and root dynamics been observed at sites near the 13 middle of this rainfall gradient (22, 23, 33). Also, it was surprising that P did not emerge as an 14 important driver of belowground dynamics across these sites, particularly since P varied so 15 strongly among these sites, and P has been shown to limit aboveground plant growth in other 16 17 tropical forests on highly weathered soils (8, 19). The general importance of base cations shown here suggests that different rock-derived nutrients may be important among sites that vary in 18 geological substrate and soil type. 19

Overall, this work illustrates a decoupling between aboveground biomass and soil C stocks in tropical forests, suggesting that roots, rather than aboveground plant growth or litter, provide the primary source of C to tropical soils. Predicting how tropical soil C stocks will

respond to global change thus requires a mechanistic representation of links among soil cations
 and root dynamics in highly-weathered soils.

3

4 Materials and Methods

5 Study Sites

Soils were sampled from a series of 1 ha forest census plots located in the Panama Canal 6 watershed, central Panama (Figure S1). Details of the plots and sampling of vegetation and soils 7 have been reported previously (27, 34-36). Briefly all plots were in lowland tropical forests 8 9 (elevation 10 - 410 meters above sea level) and included secondary, mature secondary, and old growth primary forests (34). The plots span a distinct rainfall gradient (annual rainfall ~1800 mm 10 y^{-1} on the Pacific coast to ~4000 mm y^{-1} on the Caribbean coast) (34, 36). The mean annual 11 temperature is 26° C and mean monthly temperature varies by $< 1^{\circ}$ C during the year (37). 12 The soils have developed on a range of geological substrates (Figure S1) (27, 34, 38, 39), 13 including volcanic (basalt, andesite, agglomerate, rhyolitic tuff) and marine sedimentary 14 (limestone, calcareous sandstone, siltstone, mudstone) lithologies. As a result, soils have marked 15 variation in fertility (27, 35), which spans a range of taxonomic orders (27). Soils were classified 16 by digging a profile pit adjacent to the southwest corner of each plot. Profiles were described by 17 genetic horizon and samples of horizons were collected for chemical and physical analysis. 18

19

20 Soil Sampling and Analyses

Each 1 ha plot is marked on a 20 m x 20 m grid. Soil cores were taken to 100 cm in
quadrats in the four corners and the center of each 1 ha plot (subsample n=5), in increments of 10
- 20 cm, 20 - 50 cm, and 50 - 100 cm. The 10 - 20 cm samples were taken with a constant

1 volume corer (5 cm diameter), and deeper samples taken with a 6.25 cm diameter auger. To account for greater variation in soil roots and C at the surface, 8 additional soil samples from 0 -2 10 cm depths were collected from alternate 20 x 20 m quadrats using the constant volume. 3 Stocks were calculated as the sum of the average values at each depth. This yielded 28 individual 4 5 samples per 1 ha plot, which were each analyzed for soil properties separately. In the two larger plots (BCI – 50 ha, Fort Sherman crane site – 5.96 ha) additional surface samples were collected, 6 yielding a total of 50 samples on BCI (including samples to 100 cm in eight locations) and 44 7 samples at Fort Sherman (including samples to 100 cm in five locations), and a total of 8 8 9 locations were also sampled to 1m depth at BCI. Within-plot variation for root biomass and soil chemistry is reported as the standard error of the 5 locations to 1m with plots (Table S2). The 10 within-plot relative standard error was 19% for root biomass to 1 m, and 13% for soil C stocks to 11 1m (Table S2). Including the addition surface soil samples for 0 - 10 cm depths decreased these 12 relative standard errors to 14% for surface roots, and 6% for surface soil C stocks. These 13 additional measures were included in plot-scale averages for surface soils prior to summing by 14 depth. 15

Samples were all taken during the eight-month wet season, to avoid seasonal changes in 16 17 soil carbon concentrations (40), which could confound comparison among sites. The majority of the plots were sampled during 2008 and 2009, with additional plots sampled between 2010 and 18 2014. Samples were returned to the laboratory within two hours of sampling and initially 19 20 screened (< 9 mm) to break up large aggregates, and stones and roots were removed by hand. Soils were not sieved ≤ 2 mm when fresh, because this is impractical for high clay soils. Fresh 21 soils were then extracted immediately using resin exchange beads for phosphorus, and within 24 22 23 h for exchangeable cations. Subsamples were air-dried for 10 days at ambient laboratory

1	temperature, sieved < 2 mm to isolate the fine earth fraction, and ground in a ball mill for total C
2	and N determination. For soil profile pits, samples were air-dried and sieved as above.

For both plot and pit samples, total C and N were determined by combustion and gas 3 chromatography using a Thermo Flash NC1112 Soil Analyzer (CE Elantech, Lakewood, NJ) and 4 5 soil pH was determined in a 1:2 soil to deionized water ratio using a glass electrode. Total phosphorus was determined by ignition (550°C, 1 h) and extraction in 1 M H₂SO₄ (16 h, 1:50 6 soil to solution ratio), with phosphate detection by automated neutralization and molybdate 7 8 colorimetry on a Lachat Quickchem 8500 (Hach Ltd, Loveland, CO, USA). Base cations were 9 extracted in Mehlich-3 solution (41), with detection by inductively coupled plasma optical emission spectrometry (Optima 7300 DV, Perkin Elmer, Inc., Shelton, CT). Total exchangeable 10 bases (TEB) was calculated as the sum of Ca, K, and Mg. 11 For profile pits, the concentrations of sand (0.053–2.0 mm), silt (0.002–0.053 mm), and 12 clay (< 0.002 mm) sized particles were determined by the pipette method after pretreatment to 13 remove salts (sodium acetate extraction), organic matter (H₂O₂ oxidation), and iron oxides 14

(dithionite reduction) (42). In profile pits, bulk density was determined by the excavation method 15

(43), removing ~1 L of soil and measuring the excavated volume of the plastic-lined hole with 16

17 water. For the upper 20 cm of the soil, bulk density was calculated by weighing the total fresh

soil mass from the constant volume core samples, and converting this to dry mass using oven dry 18

weight (calculated by drying a subsample for 24 h at 105°C) and stone content of the samples. 19

20 Roots were exhaustively removed from each sample by hand (i.e. until no more visible roots remained), washed on a 250 micron sieve, oven-dried for 3 d at 60°C, and weighed. Root 21 biomass was then calculated on an area basis using the core volume and depth for each sample, 22 23 averaging depths across the plot and summing the depths to 50 cm and to 100 cm.

Total carbon stocks were calculated in kg/m^2 using bulk density and depth increments, 1 averaging depths across the plot and summing the depths to 50 cm and to 100 cm. Bulk densities 2 were for the fine earth fraction as a proportion of the total soil volume -i.e. accounting for the 3 4 stone fraction (in both core samples and profile pits). Where samples were obtained to < 100 cm 5 due to bedrock, the plot values were corrected based on the proportion of cores in the plot reaching a particular depth (i.e. average soil depth for the plot). However, plot averages were not 6 corrected for floating boulders, where augering was prevented in some locations in the plot but 7 soil continued below. 8

9

10 *Litterfall*

Fifty 1 m x 1 m litter traps were established within each of 8 1ha forest plots. This subset of the 48 plots represented both high and low fertility sites within a given rainfall range. Litter traps were distributed evenly across each plot, with a pair of traps in the center of each of the twenty-five 20 x 20 m quadrats. Litter from each trap was collected biweekly and dried at 65°C until mass stabilized (~ 48 hours), after which dry mass was measured.

16

17 Statistical Analysis

To identify significant predictors of soil C stocks we used preliminary backward stepwise linear regression analyses including all soil data for order, roots, nutrients, and texture, as well as aboveground biomass, and rainfall factors. We then included only potentially significant factors identified by these stepwise regressions in final linear models. Similar general linear model selection was used for predicting fine root biomass. We then ran post-hoc means separation Fisher's Least Significant Difference (LSD) tests to compare soil C, nutrients, texture, and root biomass among soil orders. Where necessary, values were log transformed to meet assumptions
 of normality (e.g. total P and resin-extractable P).

3 We also analyzed the above relationships simultaneously using structural equation modeling (SEM) path analysis to assess mechanistic relationships among factors influencing soil 4 C stock sizes. We used soil chemical and physical factors, aboveground biomass, and rainfall as 5 6 exogenous factors, and soil C and root biomass as endogenous factors. We used SEM modification indices to add or remove specific pathways from the model. An initial model 7 included soil P as an exogenous factor, but this was removed because of poor model similarity to 8 the saturated model. The final default model (i.e. our model) was tested for overall similarity to 9 10 the saturated model (i.e., ideal model) using common metrics of comparison (i.e., no significant difference from saturated model, Chi-square p > 0.05, GFI > 0.9, NFI > 0.9, RMSEA < 0.05). 11 The model used 48 observations. 12

General linear models were performed using 13.0.0 JMP Pro software (SAS Institute Inc.,
2016), and SEM path analysis was performed using SPSS AMOS 23 software (IBM, 2015).

Statistical significance for all relationships and models was p < 0.05 unless otherwise noted, and
means are reported ± one standard error.

17

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Table 1. Soil chemical characteristics to 1m depth across tropical forest sites. Soil C, nutrients, clay content, pH and fine root

- 2 biomass are shown for tropical forest sites across gradients in rainfall and geological substrate. In general, chemical characteristics
- 3 were only weakly ($R^2 < 0.05$) or not significantly correlated with rainfall across the precipitation gradient, other than total C and total
- 4 N ($R^2 = 0.10$ and 0.12, respectively). Five soil orders were identified, including Inceptisols (Inc), Molisols (Mol), Alfisols (Alf),
- 5 Ultisols (Ult), and Oxisols (Ox). Inceptisols include modifiers because of the broad range of soil conditions found for this diverse
- 6 order. Modifiers include: aquic (Aq; periodic saturation at 40 75 cm depths), eutric (Eu; subsoil base saturation is high whether or
- 7 not the actual base concentrations are high or low), humic (Hu; high C), oxic (Ox; low effective cation exchange capacity). Means are
- 8 shown ± 1 standard error. Letters in columns show significant differences among soil orders using Fisher's LSD test (p < 0.05). Total
- 9 P and resin extractable P were log transformed for statistical tests to maintain assumptions of normality.

Soil Orde r (from least to most weat hered)	# of sit es (N)	Total C kg/m ²	% of C in the top 50 cm	Total N mg/m ²	Total P to 1 m g/m ²	Resin Extractabl e P g/m ²	Total Extractab le Bases kg/m ²	Profile Clay (%)	рН	Fine Roots g/m ^{2 ^}	% of Fine Roots in the top 50 cm
Inc Aq	2	$\begin{array}{c} 9.5\pm0.4\\ \textbf{b} \end{array}$	79 ± 1 ab	$813\pm35~\textbf{b}$	199 ± 67 ab	$\begin{array}{c} 1.2\pm0.2\\ \textbf{ab} \end{array}$	4.5 ± 1.5 a	$\begin{array}{c} 49\pm0.1\\ \textbf{ab} \end{array}$	5.5 ± 0.04 ab	172 ± 31 c	92 ± 0.5
Inc Eu	7	11.1 ± 1.1 b	75 ± 3 bc	$\begin{array}{c} 1058\pm86\\ \textbf{ab} \end{array}$	349 ± 90 ab	$\frac{1.8 \pm 1.3}{\mathbf{ab}}$	$\begin{array}{c} 2.5\pm0.6\\ \textbf{b} \end{array}$	51 ± 7 b	5.5 ± 0.3 ab	$\begin{array}{c} 207\pm27\\ \textbf{c} \end{array}$	92 ± 2
Inc Hu	4	$\begin{array}{c} 14.8\pm1.9\\ \textbf{ab} \end{array}$	80 ± 2 b	$\begin{array}{c} 1471 \pm 274 \\ \mathbf{a} \end{array}$	263 ± 48 ab	$\begin{array}{c} 0.3\pm0.1\\ \textbf{bc} \end{array}$	$\begin{array}{c} 1.7\pm0.5\\ \textbf{bcd} \end{array}$	65 ± 4 ab	5.2 ± 0.1 ab	$\begin{array}{c} 294\pm30\\ \textbf{bc} \end{array}$	92 ± 2
Inc Ox	3	$\begin{array}{c} 13.8\pm1.2\\ \textbf{ab} \end{array}$	71 ± 3 bc	$\begin{array}{c} 1173 \pm 120 \\ \textbf{ab} \end{array}$	184 ± 40 ab	$\begin{array}{c} 0.3\pm0.1\\ \textbf{bc} \end{array}$	$\begin{array}{c} 0.6\pm0.3\\ \textbf{cde} \end{array}$	66 ± 3 ab	$\begin{array}{c} 4.7\pm0.3\\ \textbf{bc} \end{array}$	273 ± 16 bc	96 ± 2
Mol	3	10.8 ± 1.7 b	90 ± 6 a	865 ± 88 b	248 ± 25 ab	2.0 ± 0.7 a	2.5 ± 0.7 abc	$\overline{47 \pm 12}$ b	5.6 ± 1.1 ab	$2\overline{09 \pm 47}$ bc	93 ± 5
Alf	13	$\frac{13.5\pm0.9}{ab}$	75 ± 1 b	1250 ± 82 ab	466 ± 129 a	2.9 ± 1.8 a	$\frac{2.8 \pm 0.4}{ab}$	51 ± 5 b	5.9 ± 0.1 a	294 ± 30 bc	95 ± 1

Ult	7	$\begin{array}{c} 14.1\pm1.0\\ \textbf{ab} \end{array}$	74 ± 4 bc	$\begin{array}{c} 1026\pm107\\ \textbf{b} \end{array}$	$226\pm56~\textbf{b}$	$\begin{array}{c} 0.3\pm0.1\\ \textbf{bc} \end{array}$	0.2 ± 0.1 e	47 ± 5 b	$\begin{array}{c} 4.4\pm0.2\\ \textbf{c}\end{array}$	$\begin{array}{c} 479\pm97\\ \textbf{a} \end{array}$	94 ± 2
Ox	9	$\begin{array}{c} 15.2\pm1.4\\ \textbf{a} \end{array}$	69 ± 1 c	$\frac{1381 \pm 140}{a}$	$224\pm52~\textbf{b}$	$\begin{array}{c} 0.2\pm0.04\\ \textbf{c} \end{array}$	$\begin{array}{c} 0.5\pm0.1\\ \textbf{de} \end{array}$	67 ± 4 a	$\begin{array}{c} 4.9\pm0.2\\ \textbf{bc} \end{array}$	$\begin{array}{c} 367\pm23\\ \textbf{ab} \end{array}$	92 ± 1

1 ^ Root biomass was available for only 44 of the 48 sites.

Figure 1. Comparing predictors of soil C stocks, (A) above ground biomass showed no correlation ($R^2 = 0.009$, n = 43, p = 0.52), whereas significant correlations were found for (B) soil clay content ($R^2 = 0.20$, n = 47, p < 0.05), and (C) fine root biomass ($R^2 = 0.27$, n = 44, p < 0.05). (D) Fine roots, in turn, showed a logarithmic decline with extractable base cations ($R^2 = 0.21$, n = 44, p < 0.05). (D) Fine roots taken from (29). Equations for the significant predictors are: (B) Carbon Stock = 8 + 0.09 *Clay%; (C)

5 Carbon Stock = 9 + 0.01 * Root Biomass; (D) Roots biomass = 307 - 47 * log(Total Extractable Bases).



Figure 2. Structural Equation Model path analysis for the relationships between exogenous 1 2 factors (soil base cations, rainfall, and clay content), and endogenous factors (fine roots and soil C stocks) is shown. Bold values in italics above each endogenous factor show the squared 3 4 multiple correlation for that factor, giving the amount of variation in that factor accounted for by all of its predictors (i.e., R^2). The sign below each arrow shows the direction of the relationship, 5 all of which are significant (p < 0.05). Correlations among exogenous factors were not significant 6 7 in this analysis. Factors in rectangles represent measured values, and circles represent unmeasured error terms (e1 and e2) for each endogenous factor. Our model was not significantly 8 different (p = 0.19) from the "ideal saturated model" (chi-square 8.7, df = 6), indicating a good 9 fit for the data. 10



1 Citations

2	1.	Jobbagy, E. G.Jackson, R. B. (2000) The vertical distribution of soil organic carbon and
3		its relation to climate and vegetation. <i>Ecological Applications</i> 10 : 423-436.
4	2.	Field, C. B., Behrenfeld, M. J., Randerson, J. T.Falkowski, P. (1998) Primary production
5		of the biosphere: Integrating terrestrial and oceanic components. Science 281: 237-240.
6	3.	Asner, G., Mascaro, J., Anderson, C., Knapp, D., Martin, R., Kennedy-Bowdoin, T., van
7		Breugel, M., Davies, S., Hall, J., Muller-Landau, H., Potvin, C., Sousa, W., Wright,
8		J.Bermingham, E. (2013) High-fidelity national carbon mapping for resource
9		management and REDD+. Carbon Balance and Management 8: doi: 10.1186/1750-0680-
10		8-7.
11	4.	Luo, Y. Q., Ahlstrom, A., Allison, S. D., Batjes, N. H., Brovkin, V., Carvalhais, N.,
12		Chappell, A., Ciais, P., Davidson, E. A., Finzi, A. C., Georgiou, K., Guenet, B., Hararuk,
13		O., Harden, J. W., He, Y. J., Hopkins, F., Jiang, L. F., Koven, C., Jackson, R. B., Jones,
14		C. D., Lara, M. J., Liang, J. Y., McGuire, A. D., Parton, W., Peng, C. H., Randerson, J.
15		T., Salazar, A., Sierra, C. A., Smith, M. J., Tian, H. Q., Todd-Brown, K. E. O., Torn, M.,
16		van Groenigen, K. J., Wang, Y. P., West, T. O., Wei, Y. X., Wieder, W. R., Xia, J. Y.,
17		Xu, X., Xu, X. F.Zhou, T. (2016) Toward more realistic projections of soil carbon
18		dynamics by Earth system models. Global Biogeochemical Cycles 30: 40-56.
19	5.	IPCC (2013), eds. Stocker, T. F., Qin, D., Plattner, GK., Tignor, M., Allen, S. K.,
20		Boschung, J., Nauels, A., Xia, Y., Bex, V. & Midgley, P. M. (Cambridge University
21		Press, Cambridge, UK), pp. http://www.climatechange2013.org/report/full-report/ipcc.
22	6.	Todd-Brown, K. E. O., Randerson, J. T., Post, W. M., Hoffman, F. M., Tarnocai, C.,
23		Schuur, E. A. G.Allison, S. D. (2013) Causes of variation in soil carbon simulations from

1	CMIP5 Earth system models and comparison with observations. <i>Biogeosciences</i> 10:
2	1717-1736.

3	7.	Crews, T. E., Kitayama, K., Fownes, J. H., Riley, R. H., Herbert, D. A., Mueller-
4		Dombois, D.Vitousek, P. M. (1995) Changes in Soil Phosphorus Fractions and
5		Ecosystem Dynamics across a Long Chronosequence in Hawaii. Ecology 76: 1407.
6	8.	Vitousek, P. M.Sanford, R. L. (1986) Nutrient cycling in moist tropical forests. Annual
7		Review of Ecology and Systematics 17: 137-167.
8	9.	Tanner, E. V. J., Sheldrake, M.Turner, B. L. (2016) Changes in soil carbon and nutrients
9		following six years of litter removal and addition in a tropical semi-evergreen rain forest.
10		<i>Biogeosciences</i> 13: 6183-6190.
11	10.	Rasse, D. P., Rumpel, C.Dignac, M. F. (2005) Is soil carbon mostly root carbon?
12		Mechanisms for a specific stabilisation. Plant and Soil 269: 341-356.
13	11.	Cusack, D. F., Chou, W. W., Liu, W. H., Harmon, M. E., Silver, W. L.LIDET-Team
14		(2009) Controls on long-term root and leaf litter decomposition in Neotropical forests.
15		Global Change Biology 15: 1339-1355.
16	12.	Bird, J. A.Torn, M. S. (2006) Fine roots vs. needles: A comparison of C-13 and N-15
17		dynamics in a ponderosa pine forest soil. Biogeochemistry 79: 361-382.
18	13.	Guerrero-Ramirez, N. R., Craven, D., Messier, C., Potvin, C., Turner, B. L.Handa, I. T.
19		(2016) Root quality and decomposition environment, but not tree species richness, drive
20		root decomposition in tropical forests. Plant and Soil 404: 125-139.
21	14.	Waring, B. G.Powers, J. S. (2017) Overlooking what is underground: Root:shoot ratios
22		and coarse root allometric equations for tropical forests. Forest Ecology and Management
23		385 : 10-15.

1	15.	Post, W. M., Emanuel, W. R., Zinke, P. J.Stangenberger, A. G. (1982) Soil carbon pools
2		and world life zones. Nature 298: 156-159.
3	16.	Cusack, D., Karpman, J., Ashdown, D., Cao, Q., Ciochina, M., Halterman, S., Lydon,
4		S.Neupane, A. (2016) Global change effects on humid tropical forests: Evidence for
5		biogeochemical and biodiversity shifts at an ecosystem scale. Reviews of Geophysics
6		10.1002/2015RG000510.
7	17.	Sposito, G., Skipper, N. T., Sutton, R., Park, S. H., Soper, A. K.Greathouse, J. A. (1999)
8		Surface geochemistry of the clay minerals. Proceedings of the National Academy of
9		Sciences of the United States of America 96: 3358-3364.
10	18.	Batjes, N. H., Ribeiro, E., van Oostrum, A., Leenaars, J., Hengl, T.de Jesus, J. M. (2017)
11		WoSIS: providing standardised soil profile data for the world. Earth System Science Data
12		9 : 1-14.
13	19.	Harrington, R. A., Fownes, J. H.Vitousek, P. M. (2001) Production and resource use
14		efficiencies in N- and P-limited tropical forests: A comparison of responses to long-term
15		fertilization. Ecosystems 4: 646-657.
16	20.	Maycock, C. R.Congdon, R. A. (2000) Fine root biomass and soil N and P in north
17		Queensland rain forests. Biotropica 32: 185-190.
18	21.	Espeleta, J. F.Clark, D. A. (2007) Multi-scale variation in fine-root biomass in a tropical
19		rain forest: A seven-year study. Ecological Monographs 77: 377-404.
20	22.	Wurzburger, N.Wright, S. J. (2015) Fine-root responses to fertilization reveal multiple
21		nutrient limitation in a lowland tropical forest. Ecology 96: 2137-2146.

1	23.	Yavitt, J. B., Harms, K. E., Garcia, M. N., Mirabello, M. J.Wright, S. J. (2011) Soil
2		fertility and fine root dynamics in response to 4 years of nutrient (N, P, K) fertilization in
3		a lowland tropical moist forest, Panama. Austral Ecology 36: 433-445.
4	24.	Bloom, A. J., Chapin, F. S.Mooney, H. A. (1985) Resource limitation in plants - an
5		economic analogy. Annual Review of Ecology and Systematics 16: 363-392.
6	25.	Phillips, R. P., Finzi, A. C.Bernhardt, E. S. (2011) Enhanced root exudation induces
7		microbial feedbacks to N cycling in a pine forest under long-term CO2 fumigation.
8		<i>Ecology Letters</i> 14: 187-194.
9	26.	Nottingham, A. T., Turner, B. L., Chamberlain, P. M., Stott, A. W.Tanner, E. V. J. (2012)
10		Priming and microbial nutrient limitation in lowland tropical forest soils of contrasting
11		fertility. Biogeochemistry 111: 219-237.
12	27.	Turner, B. L.Engelbrecht, B. M. J. (2011) Soil organic phosphorus in lowland tropical
13		rain forests. Biogeochemistry 103: 297-315.
14	28.	Staff, S. S. (1999) Soil Taxonomy: A Basic System of Soil Classification for Making and
15		Interpreting Soil Surveys Lincoln, NE).
16	29.	Chave, J., Condit, R., Aguilar, S., Hernandez, A., Lao, S.Perez, R. (2004) Error
17		propagation and scaling for tropical forest biomass estimates. Philosophical Transactions
18		of the Royal Society of London Series B-Biological Sciences 359: 409-420.
19	30.	Yavitt, J. B.Wright, S. J. (2001) Drought and irrigation effects on fine root dynamics in a
20		tropical moist forest, Panama. Biotropica 33: 421-434.
21	31.	Metcalfe, D. B., Meir, P., Aragao, L., da Costa, A. C. L., Braga, A. P., Goncalves, P. H.
22		L., Silva, J. D., de Almeida, S. S., Dawson, L. A., Malhi, Y.Williams, M. (2008) The

1		effects of water availability on root growth and morphology in an Amazon rainforest.
2		<i>Plant and Soil</i> 311 : 189-199.
3	32.	Augusto, L., Achat, D., Jonard, M., Vidal, D.Ringeval, B. (in press) Soil parent material -
4		a major driver of plant nutrient limitations in terrestrial ecosystems Global Change
5		Biology doi: 10.1111/gcb.13691
6	33.	Wright, S. J., Yavitt, J. B., Wurzburger, N., Turner, B. L., Tanner, E. V. J., Sayer, E. J.,
7		Santiago, L. S., Kaspari, M., Hedin, L. O., Harms, K. E., Garcia, M. N.Corre, M. D.
8		(2011) Potassium, phosphorus, or nitrogen limit root allocation, tree growth, or litter
9		production in a lowland tropical forest. Ecology 92: 1616-1625.
10	34.	Pyke, C. R., Condit, R., Aguilar, S.Lao, S. (2001) Floristic composition across a climatic
11		gradient in a neotropical lowland forest. Journal of Vegetation Science 12: 553-566.
12	35.	Condit, R., Engelbrecht, B. M. J., Pino, D., Perez, R.Turner, B. L. (2013) Species
13		distributions in response to individual soil nutrients and seasonal drought across a
14		community of tropical trees. Proceedings of the National Academy of Sciences of the
15		United States of America 110: 5064-5068.
16	36.	Engelbrecht, B. M. J., Comita, L. S., Condit, R., Kursar, T. A., Tyree, M. T., Turner, B.
17		L.Hubbell, S. P. (2007) Drought sensitivity shapes species distribution patterns in tropical
18		forests. Nature 447: 80-U2.
19	37.	Windsor, D., Rand, A.Rand, W. (1990) Caracteristicas de la Precipitacion de la Isla de
20		Barro Colorado.
21	38.	Stewart, R., Stewart, J.Woodring, W. (1980), United States Geological Survey, Boulder,
22		CO).
23	39.	Woodring, W. (1958), ed. Smithsonian, Panama).

1	40.	Turner, B. L., Yavitt, J. B., Harms, K. E., Garcia, M. N.Wright, S. J. (2015) Seasonal
2		changes in soil organic matter after a decade of nutrient addition in a lowland tropical
3		forest. Biogeochemistry 123: 221-235.
4	41.	Mehlich, A. (1984) Mehlich 3 soil test extractant: a modification of Mehlich 2 extractant.
5		Communications in Soil Science and Plant Analysis 15: 1409-1416.
6	42.	Gee, G.Or, D. (2002) in Methods of Soil Analysis, Part 4 – Physical Methods, eds. Dane,
7		J. & Topp, C. (Soil Science Society of America, Wisconsin), pp. 255–293.
8	43.	Grossman, R.Reinsch, T. (2002) in Methods of Soil Analysis. Part 4. Physical Methods,
9		eds. Dane, J. & Topp, C. (Soil Society of America, Wisconsin), pp. 201-293.