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# The Journal of Membrane Biology

## Stereospecific Interactions of Cholesterol in a Model Cell Membrane: Implications for the Membrane Dipole Potential --Manuscript Draft--

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<b>Abstract:</b>	<p>Cholesterol is a major constituent of the plasma membrane in higher-order eukaryote organisms. The effect of cholesterol on the structure and organisation of cell membranes has been studied extensively by both experimental and computational means. In recent years, a wealth of data has been accumulated illustrating how subtle differences in the structure of cholesterol equate to considerable changes in the physical properties of the membrane. The effect of cholesterol stereoisomers, in particular, has been established, identifying a direct link with the activity of specific membrane proteins. In this study, we perform extensive molecular dynamics simulations of phospholipid bilayers containing three isomers of cholesterol, the native form (nat-cholesterol), the enantiomer of the native form (ent-cholesterol), and an epimer of cholesterol that differs by the orientation of the polar hydroxyl group (epi-cholesterol). Based on these simulations, an atomic-level description of the stereospecific cholesterol-phospholipid interactions is provided, establishing a potential mechanism for the perturbation of membrane properties, specifically the membrane dipole potential.</p>	
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# Stereospecific Interactions of Cholesterol in a Model Cell Membrane: Implications for the Membrane Dipole Potential

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**Keywords:** computer simulations; molecular dynamics; cholesterol; ent-cholesterol; epi-cholesterol; GPCRs

## ABSTRACT

Cholesterol is a major constituent of the plasma membrane in higher-order eukaryote organisms. The effect of cholesterol on the structure and organisation of cell membranes has been studied extensively by both experimental and computational means. In recent years, a wealth of data has been accumulated illustrating how subtle differences in the structure of cholesterol equate to considerable changes in the physical properties of the membrane. The effect of cholesterol stereoisomers, in particular, has been established, identifying a direct link with the activity of specific membrane proteins. In this study, we perform extensive molecular dynamics simulations of phospholipid bilayers containing three isomers of cholesterol, the native form (*nat*-cholesterol), the enantiomer of the native form (*ent*-cholesterol), and an epimer of cholesterol that differs by the orientation of the polar hydroxyl group (*epi*-cholesterol). Based on these simulations, an atomic-level description of the stereospecific cholesterol-phospholipid interactions is provided, establishing a potential mechanism for the perturbation of membrane properties, specifically the membrane dipole potential.

## INTRODUCTION

1 Phospholipid bilayers serve as the fundamental scaffold of all biological membranes,  
2 forming a barrier between the cell interior and the surrounding environment. Integral  
3 membrane proteins are responsible for the controlled passage of substances across this  
4 barrier. Cholesterol is particularly abundant in the plasma membrane of higher-order  
5 eukaryotes, and is an important requirement for the functioning of a diverse range of  
6 membrane proteins, whilst acting as a crucial regulator of membrane fluidity.<sup>1</sup> As a  
7 result, the specific effect of cholesterol on the structure of a phospholipid bilayer has  
8 been under discussion for much of the last century.<sup>2</sup> Arguably the most striking  
9 response to membrane cholesterol is the ‘condensing effect’, whereby the area per  
10 lipid molecule is noticeably lower than would be observed during ideal mixing, and  
11 the membrane thickness is increased concordantly, indicating changes in lipid  
12 organisation.<sup>3</sup>

13 In recent years, significant interest has been focused toward characterising the  
14 physical effect of naturally occurring cholesterol analogues, as well as synthetic  
15 cholesterol isomers and derivatives on lipid membranes. The native form of  
16 cholesterol (*nat*-cholesterol) is an asymmetric amphiphilic molecule, composed of a  
17 hydroxyl group attached to a tetracyclic sterane backbone, exhibiting a smooth  $\alpha$  face  
18 and rough  $\beta$  face (Fig. 1A). The enantiomer of *nat*-cholesterol (*ent*-cholesterol) is the  
19 non-superimposable mirror image (Fig. 1B), exhibiting physiochemical properties  
20 indistinguishable to native cholesterol, except for the rotation of plane-polarised  
21 light.<sup>4-7</sup> Subtle changes in cholesterol membrane structure have been shown to exert  
22 profound effects on innate physiochemical properties. Certain attributes are known to  
23 significantly diverge in membranes containing related cholesterol molecules,  
24 including but not limited to *epi*-cholesterol, a stereoisomer of cholesterol, which  
25 differs in the orientation of the hydroxyl group (Fig. 1C), and 7-dehydrocholesterol  
26 and desmosterol, cholesterol precursors that differ by a double bond.<sup>8-13</sup> Membrane  
27 dipole potential, the potential difference within the membrane bilayer, exemplifies  
28 this phenomenon. A significant increase in dipole potential is observed upon  
29 cholesterol replenishment of both model and natural membranes, a lesser increase  
30 when 7-dehydrocholesterol and desmosterol are present, whilst replacement with *epi*-  
31 cholesterol causes the dipole potential to decrease, thus demonstrating that this  
32 property is highly dependent on the exact cholesterol structure.<sup>14-17</sup> The molecular

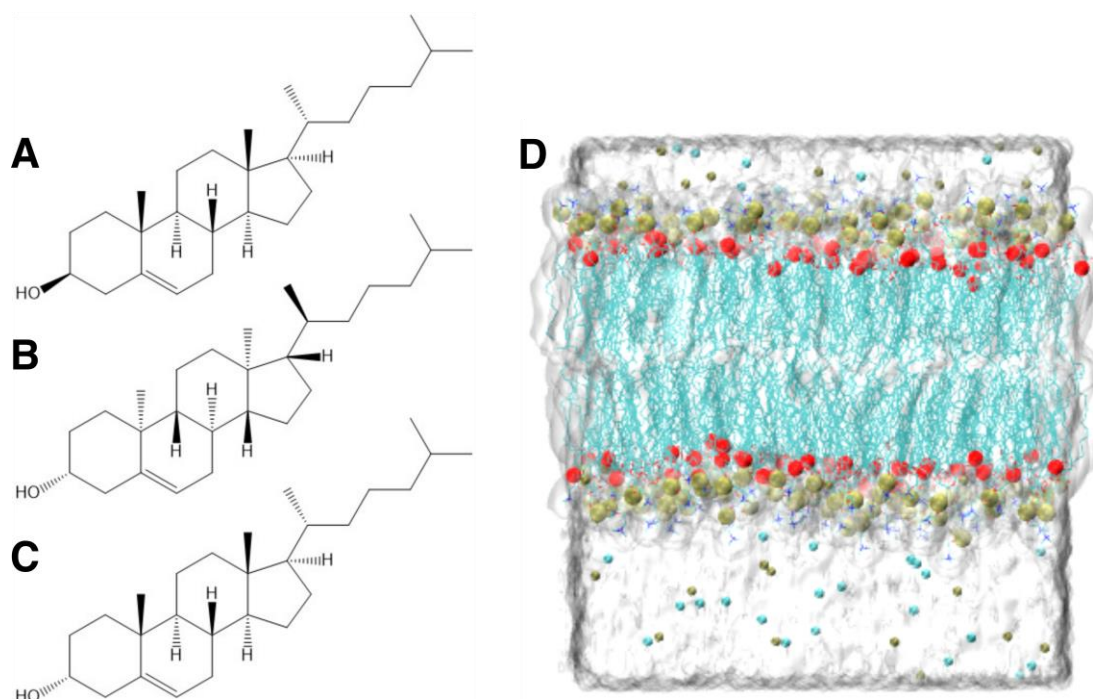
1 mechanism underlying the regulation of membrane dipole potential via cholesterol  
2 and related molecules is currently unclear, however.

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4 It has also been demonstrated that variations in cholesterol structure can modulate the  
5 functioning of specific membrane proteins, including but not limited to inwardly  
6 rectifying potassium channel Kir2.1,<sup>18</sup> ligand-gated ion channel receptor GABA<sub>A</sub><sup>19</sup>  
7 and G-protein coupled receptor 5-HT<sub>1A</sub>.<sup>20</sup> Membrane cholesterol has been suspected  
8 to regulate membrane proteins by a direct mechanism, where direct coordination of  
9 cholesterol provokes conformational changes in the receptor, or an indirect  
10 mechanism, by modifying the membrane properties. Unnatural isomers, *ent*- and *epi*-  
11 cholesterol, have proven to be useful experimental probes to determine the exactitude  
12 of the interaction of cholesterol with such membrane proteins, and consequently the  
13 mechanism of cholesterol action.<sup>6, 18-21</sup> *Ent*-cholesterol, in particular, is a useful  
14 candidate to elucidate specific cholesterol effects, as the membrane properties are  
15 unchanged.

16  
17 To elucidate how cholesterol and related molecules evoke differences in the physical  
18 properties of the membrane and the dynamics of membrane proteins, a high-  
19 resolution description of the specific interactions is required. As a consequence,  
20 molecular dynamics (MD) simulations have become an established method to typify  
21 the impact of cholesterol on membrane lipids and proteins, in atomic detail.<sup>22-24</sup> In this  
22 study, we have performed all-atom MD simulations of phospholipid bilayers  
23 containing 30% *nat*-, *ent*- and *epi*-cholesterol, as well as a 100% phospholipid bilayer,  
24 in order to determine the atomic behaviour of the cholesterol isomers. The simulations  
25 were 500 ns in length, significantly improving the timescale of prior MD simulations  
26 examining the dynamics of *epi*-cholesterol, which were executed for 2 ns.<sup>25</sup> Key  
27 biophysical properties, including area per lipid, cholesterol tilt-angle and solvent-  
28 accessible-surface area, as well as the identity of crucial cholesterol-phospholipid  
29 interactions, were derived from the simulations. The results indicate potential  
30 mechanisms by which *epi*-cholesterol influences the membrane dipole potential and  
31 interfere with the functioning of membrane proteins. This is particularly relevant for  
32 the functioning of 5-HT<sub>1A</sub> receptor, as ligand binding is abated in the presence of the  
33 *epi*-cholesterol.<sup>20</sup>

34  
35 Figure 1. Chemical structure of (A) *nat*-cholesterol, (B) *ent*-cholesterol and (C) *epi*-  
36 cholesterol molecules. (D) Representative snapshot of a simulation system: a lipid  
37 bilayer is solvated in a water box with a final concentration of 150 mM final of KCl.  
38 Phosphate atoms of the phospholipid molecules, and oxygen atoms of the cholesterol

1 molecules are represented by a brown and red spheres, respectively, to indicate the  
2 position of the headgroups. Carbon atoms are shown in cyan.



## 28 METHODS

### 31 System Setup

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Membranes containing 1- palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine (POPC) molecules and *nat*-cholesterol molecules were generated using the CHARMM-GUI Membrane Builder.<sup>26 27 28</sup> The POPC membrane contained 244 POPC molecules, and the mixed bilayer contained 196 POPC molecules and 84 cholesterol molecules which represents ~30% cholesterol content to mimic known experimental conditions. The mixed POPC/*nat*-cholesterol membrane was then modified to generate additional membranes containing the epimer and enantiomer of cholesterol. The combined system was then solvated to produce a rectangular water box of dimensions (92 x 92 x 80) Å<sup>3</sup>. K<sup>+</sup> and Cl<sup>-</sup> ions were added using the Autoionise Plugin of VMD, to neutralise the system and obtain a 150 mM concentration.<sup>29</sup> The final systems contained approximately 60,000 atoms, and can be seen in Fig. 1D.

### 54 Molecular Dynamics Simulations

NAMD2.9 was employed to perform molecular dynamics (MD) simulations of the systems throughout.<sup>30</sup> CHARMM36 parameters were used for phospholipids<sup>31</sup>, with standard parameters for ions<sup>32</sup> and the TIP3P<sup>33</sup> model for water. The modified

CHARMM36 parameters by Lim et al were used for *nat*-cholesterol, and manually modified to accommodate the *ent*- and *epi*-cholesterol molecules.<sup>34</sup> All systems were equilibrated using the documented CHARMM-GUI protocol.<sup>27</sup> Unrestrained dynamics for each system was then performed in the NPT ensemble for 500 ns. The Particle Mesh Ewald method was used for the treatment of full-system periodic electrostatic interactions; interactions were evaluated every second timestep with a value of 1 Å to determine grid spacing.<sup>35</sup> Electrostatic and van der Waals forces were calculated every timestep and up to a cut-off distance of 12 Å. A switching distance of 10 Å was chosen to smoothly truncate the non-bonded interactions. Only atoms in a Verlet pair list within a cut-off distance of 13.5 Å were considered, with the list reassigned every 20 steps.<sup>36</sup> The SETTLE algorithm was used to constrain all bonds involving hydrogen atoms to allow the use of a 2 fs timestep.<sup>37</sup> The Nose-Hoover-Langevin piston was employed to control the pressure with a 200 fs period, 50 fs damping constant and a desired value of 1 atmosphere.<sup>38 39</sup> The systems were coupled to a Langevin thermostat to sustain a temperature of 298 K throughout, in line with the experimental conditions used for radiolabel binding assays by Jafurulla et al.<sup>20</sup>

### Trajectory Analysis

The atom names of the cholesterol and POPC molecules referred to in the following text are defined Fig. S1 in the Supplementary Material. MEMBPLUGIN was used to perform analyses of (1) membrane thickness, (2) area per lipid, (3) the carbon-deuterium (SCD) order parameter, (4) lipid tilt angle, and (5) lipid interdigitation.<sup>40</sup> To begin with, the membrane thickness was calculated as the distance between the projected mass distributions of the P atoms in the phospholipid headgroups in the upper and lower leaflets. To calculate the area per lipid, the simulation box was divided into a Voronoi diagram, based on the coordinates of atom O3 in cholesterol molecules and atoms C2, C21 and C31 in POPC molecules (see Supplementary Fig. S1 for notation), and the resulting area of the polygons was evaluated, for each bilayer leaflet. The SCD order parameter was calculated as  $S_{CD} = -\frac{1}{2}\langle 3 \cos^2 \theta - 1 \rangle$ , to gauge the motility of the C-H bonds on the phospholipid acyl chains.<sup>41</sup> The tilt angle was calculated as the angle between the vector defined by the C10 and C13 cholesterol atoms and the bilayer normal, which is aligned to the z-axis throughout the simulation trajectories. The degree of acyl chain interdigitation was measured via a coordination-based fraction, obtained by evaluating the number of heavy atoms



1 within 4 Å (default value) of atoms in the opposing leaflets.<sup>40</sup> Density profiles were  
2 calculated using the Density Profile Tool of VMD.<sup>42</sup> The solvent-accessible surface  
3 area and radial distribution functions were also calculated using built-in VMD tools.  
4 Further analysis was performed using in-house TCL scripts. A 4 Å distance cut-off  
5 was used for the calculation of intermolecular charge pairs and atomic interaction  
6 pairs between heavy atoms. A 3.5 Å distance and 35° angle cut-off was used for the  
7 calculation of H-bonding. The lifetime of *epi*-cholesterol dimers was calculated by  
8 considering the period between H-bond formation and disruption of the H-bond for a  
9 minimum of 0.5 ns. Reformation after this interval is considered as a new dimer, and  
10 hence lifetime. The initial 200 ns of the simulation trajectories were considered as an  
11 equilibration period, following assessment of the fluctuations of the membrane  
12 thickness and area per lipid (Table S1). These properties were regarded as converged  
13 by 500 ns, thus all analysis is performed on the 200-500 ns time interval.  
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## 24 RESULTS

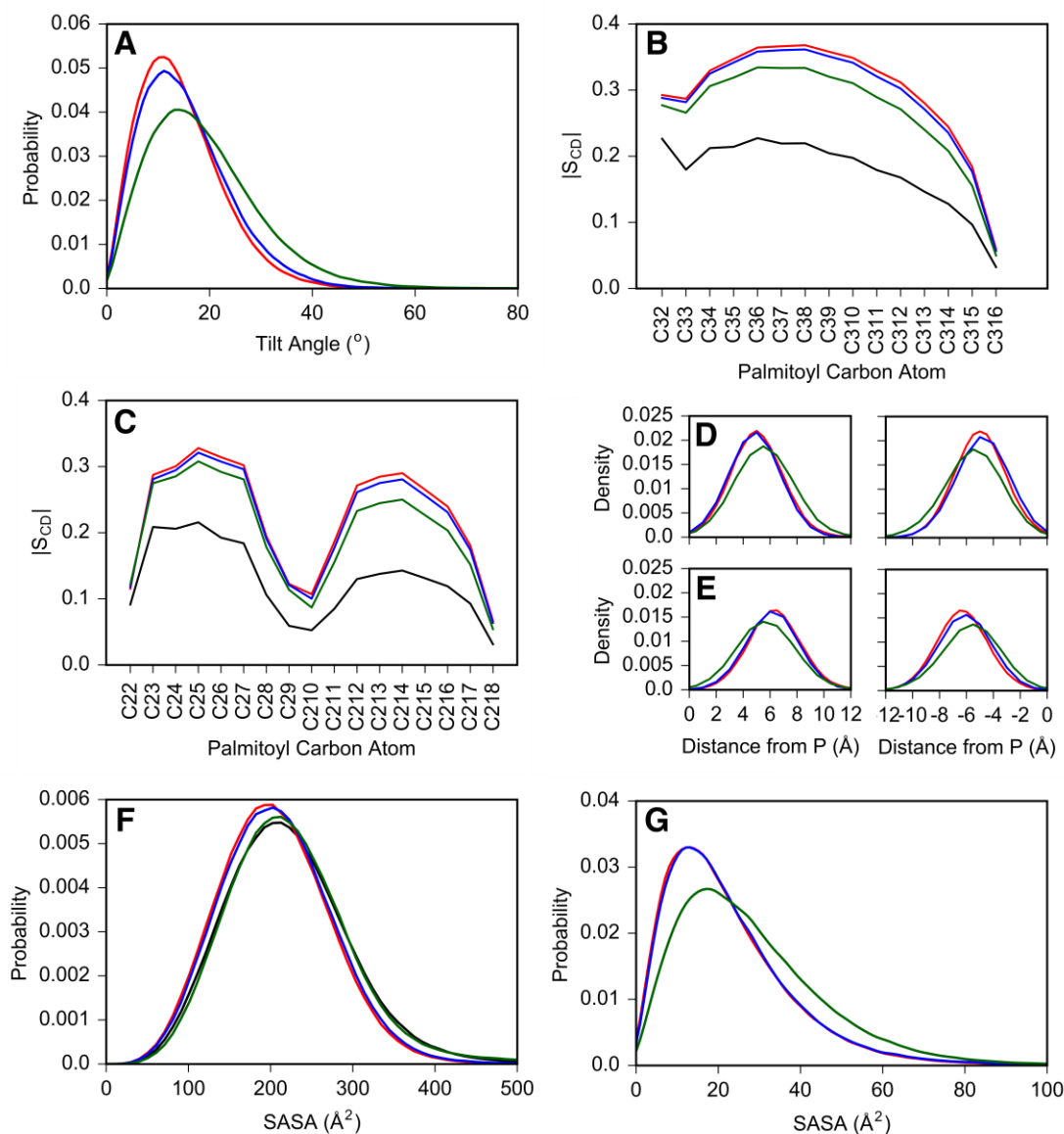
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26 In the first instance, global membrane properties have been calculated to elucidate the  
27 differences detectable using the MD simulation protocol described in this study, and  
28 the conformance of such results with known experimental data. In the POPC  
29 simulation, the membrane thickness and the area per lipid are calculated as  $40 \pm 1$  Å  
30 and  $63 \pm 1$  Å<sup>3</sup> respectively (Table 1), in close agreement with accepted experimental  
31 values.<sup>43</sup> The addition of cholesterol causes a net reduction of total bilayer area, with  
32 the overall average area per lipid in the system reducing to  $45 \pm 1$  Å<sup>3</sup> in CHL and  $46 \pm$   
33  $1$  Å<sup>3</sup> in ENT, characteristic of the well-known condensing effect of cholesterol.<sup>3</sup> This  
34 effect is slightly offset in EPI, with an overall average area per lipid of  $48 \pm 1$  Å<sup>3</sup>,  
35 primarily as a result of an increased area of POPC molecules (Table 1).  
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45 Correspondingly, an increase in membrane thickness is observed in CHL ( $46 \pm 1$  Å),  
46 ENT ( $46 \pm 1$  Å) and to a lesser degree in EPI ( $45 \pm 1$  Å). The extent to which the  
47 components of the upper and lower leaflets overlap, known as lipid interdigitation, are  
48 expectantly inversely correlated, displaying an increase in interdigitation alongside a  
49 decrease in thickness (Table 1).  
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Table 1. Summary of key membrane physical properties ( $\pm$  SD) obtained from MD simulations on the 200-500 ns time interval.

		POPC	CHL	ENT	EPI
Thickness ( $\text{\AA}$ )	POPC	$39.8 \pm 0.9$	$46.3 \pm 0.9$	$45.9 \pm 0.9$	$44.8 \pm 1.0$
	Sterol	-	$35.7 \pm 0.8$	$35.6 \pm 0.7$	$33.2 \pm 0.8$
Area per Lipid ( $\text{\AA}^3$ )	POPC+Sterol	$62.5 \pm 1.2$	$45.4 \pm 0.7$	$45.9 \pm 0.7$	$47.4 \pm 0.9$
	POPC	$62.5 \pm 0.9$	$53.3 \pm 0.7$	$53.9 \pm 0.6$	$55.7 \pm 0.8$
	Sterol	-	$27.2 \pm 0.8$	$27.0 \pm 0.8$	$28.1 \pm 1.1$
Tilt Angle ( $^\circ$ )	Sterol	-	$14.4 \pm 1.2$	$15.3 \pm 1.2$	$19.0 \pm 1.5$
Interdigitation (%)		$5.6 \pm 0.4$	$3.4 \pm 0.2$	$3.5 \pm 0.2$	$3.8 \pm 0.2$

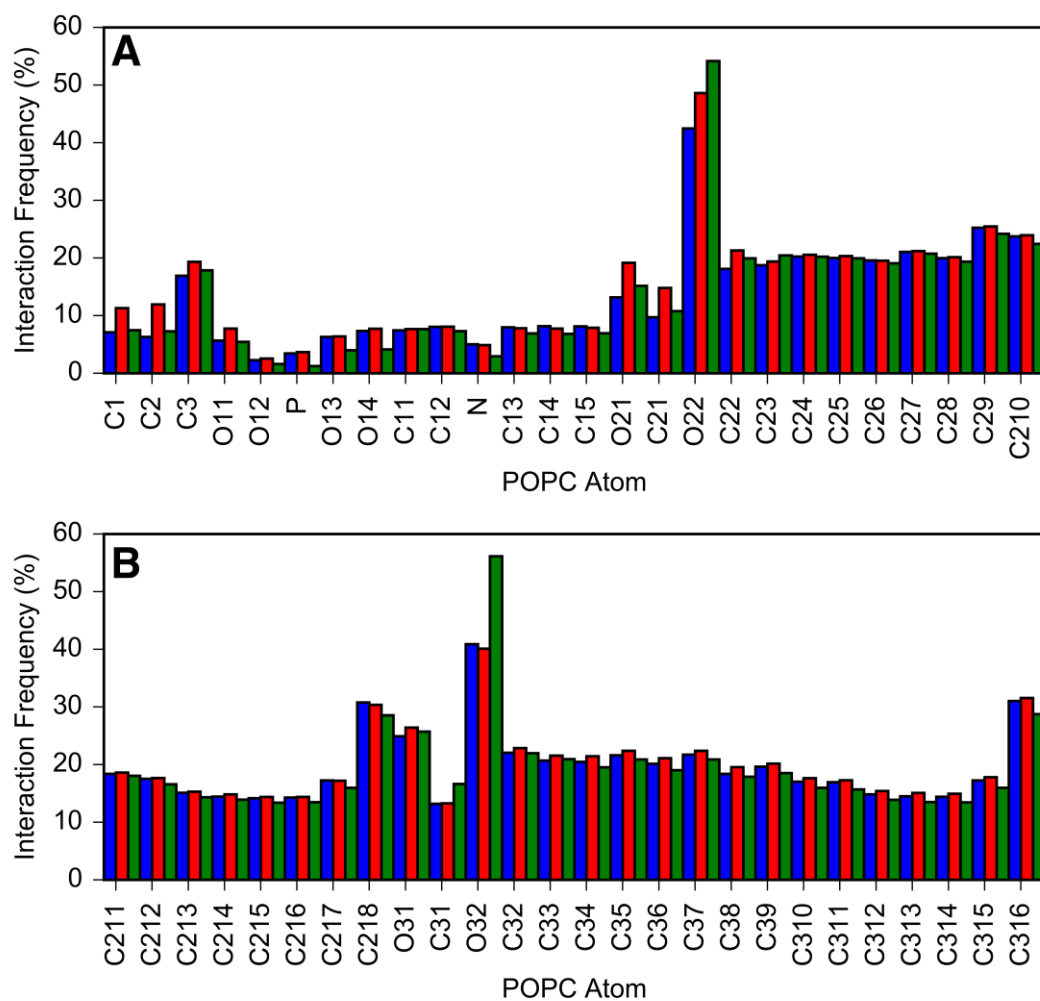
Figure 2. (A) Distribution of cholesterol tilt angles, normalized to 1 with a bin width of 1. SCD order parameter of (B) 16:0 saturated acyl chain at the glycerol *sn1* position (C3) and (C) 18:1 unsaturated acyl chain at the glycerol *sn2* position (C2). Mass density profiles of cholesterol (D) C3 and (E) O3 atoms, in the lower (left) and upper (right) leaflets, following alignment of the POPC phosphate atoms, normalized to 1 with a bin width of 0.5. SASA per (F) POPC and (G) cholesterol molecule, normalized to 1 with a bin width of 10 and 1 respectively. CHL, ENT, EPI and POPC simulations are displayed in red, blue, green and black respectively.



Such differences in the membrane physical properties arise from differences in the position and orientation of cholesterol in the bilayer, and the synergistic interaction with other bilayer components.<sup>44</sup> Cholesterol tilt angle is an important quantity, which is closely related to the ability of cholesterol to order its environment.<sup>45</sup> In this example, CHL and ENT display the narrowest distribution of tilt angles, with mean values of  $14 \pm 1^\circ$  and  $15 \pm 1^\circ$ , respectively, representing a conformation almost parallel to the bilayer normal (Fig. 2A). The distribution of tilt angles in EPI is noticeably more spread, with an increased mean of  $19 \pm 2^\circ$ . This effect is reflected in the calculated  $S_{CD}$  order parameter, which is commonly used as a measure of lipid tail organisation, and can be directly compared to experimentation.<sup>46-48</sup> Overall,  $S_{CD}$  values increase in the cholesterol-containing bilayers, relative to POPC alone (Fig. 2B-C), indicative of the ordering effect of cholesterol that is well established in the

1 literature.<sup>49</sup> This effect has been attributed to lipid packing around the rigid  
 2 cholesterol frame. EPI exerts a weaker ordering effect overall, relative to CHL and  
 3 ENT, with the greatest differences observed in the saturated regions of the acyl  
 4 chains. These results contribute to a wealth of previous literature, which note the  
 5 specificity of membrane ordering with regards the atomic structure of cholesterol.<sup>50</sup>  
 6 To directly compare the position of the cholesterol hydrophobic ring system and the  
 7 hydrophilic hydroxyl group in the membrane, the mass density profiles of the C3 and  
 8 O3 atoms (defined in the Supplementary Fig. S1), following alignment of PC  
 9 phosphate atoms, can be calculated (Fig. 2D-E). The C3-O3 maxima of EPI overlap at  
 10  $\sim 5.5$  Å resulting from the axial conformation of the hydroxyl group, suggesting that  
 11 whilst C3 adopts a shallower position in the membrane ( $\sim 6$  Å in CHL and ENT), O3  
 12 is more buried ( $\sim 5$  Å in CHL and ENT).

23 Figure 3. (A-B) Average interaction frequency between cholesterol and POPC atoms  
 24 (specified). Interactions are calculated using a 4 Å criterion between heavy atoms.



To assess how this altered position affects the organisation of the membrane on an atomic-level, the average interaction frequency between individual POPC and cholesterol atoms has been calculated (Fig. 3). In general, headgroup atoms exhibit an average interaction frequency of ~0-10% of POPC, whilst tail atoms and those in the linker region, display average interaction frequencies in the 10-30% range, with EPI consistently presenting lower values. The carbonyl groups of the ester linkages (O22 and O32) are the exceptions to this, with average interaction frequencies in excess of 40% and elevated average interaction frequencies in EPI (54%, 56%) are relative to CHL (42%, 41%) and ENT (49%, 40%). The observed net increases can be attributed to enhanced interactions with C1, C5, C6 and C7 atoms, offsetting the reduced contact interface with O3.

To assess the impact of such differences at the membrane-water interface, the solvent accessible surface area (SASA) per molecule has been calculated for POPC and each sterol species (Fig. 2F-G and Table 2). The SASA of POPC in a single component membrane is calculated as  $\sim 220 \text{ \AA}^2$ , which reduces by  $\sim 15 \text{ \AA}^2$  in CHL,  $12 \text{ \AA}^2$  in ENT and remarkably increases by  $\sim 5 \text{ \AA}^2$  in EPI, despite a decrease in the area per lipid for all cholesterol isomers relative to POPC-only. These observations provide evidence that SASA, and hence POPC-solvent interactions, are modulated by specific lipid-cholesterol interactions. It is interesting to note that the hydration of POPC headgroups decreases by  $\sim 4\%$  in CHL and ENT, and  $2\%$  in EPI, as calculated from the individual RDFs of all POPC atoms (Fig. S2).

Table 2. Average SASA ( $\pm$  SD) of membrane lipids

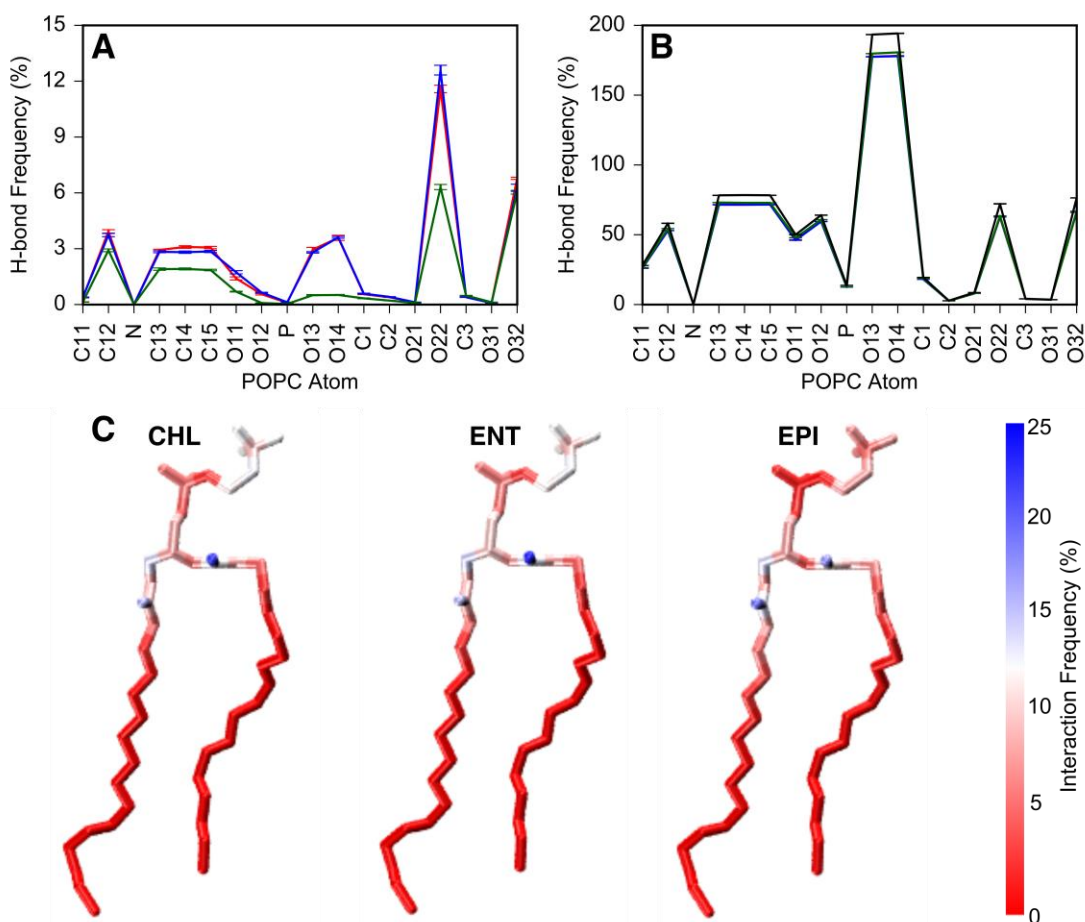
SASA ( $\text{\AA}^2$ )	POPC	CHL	ENT	EPI
POPC	$220 \pm 75$	$205 \pm 67$	$208 \pm 68$	$225 \pm 81$
Sterol	-	$22 \pm 16$	$23 \pm 16$	$28 \pm 18$

With regard to the SASA of cholesterol, CHL ( $22 \text{ \AA}^2$ ) and ENT ( $23 \text{ \AA}^2$ ) display similar values, increasing to  $28 \text{ \AA}^2$  in EPI (Table 2). This increase can be attributed to numerous effects, such as the increased area per cholesterol and tilt angle, the elevated position in the membrane and exposure of ring atoms as a result of the orientation of the hydroxyl group. The net increase in hydration in EPI is  $\sim 10\%$ , relative to CHL and ENT, as calculated from the individual RDFs of all cholesterol atoms (Fig. S3). It is worth noting that a decrease is observed for O3, signalling that

1 interactions with hydrophobic atoms are solely responsible for the observed increase.  
2 This will likely perturb the electrostatics of the cholesterol-water interfacial region.  
3 H-bonding is a further important feature of the membrane-water interface.  
4  
5 Approximately, all CHL and ENT molecules participate in at least one H-bond, via  
6 the cholesterol hydroxyl group, with 42% of molecules associated with PC  
7 headgroups and 60% of molecules H-bonding with water, in both cases. In contrast,  
8  
9 70% of EPI molecules are involved in H-bonding, as a result of diminished  
10 interactions with both PC headgroups (25%) and solvent (37%), and the evolution of  
11 cholesterol homo-dimers (8%). Dissection into the individual components of the PC  
12 headgroup (Fig. 4A, see Fig. S1 for atom names), identifies the largest reduction of  
13 H-bonding with atoms O13 (-3%) and O14 (-3%) in the phosphate group, and O22 (-  
14 6%) at the apex of the unsaturated acyl chain at the *sn2* acyl position, whilst H-  
15 bonding with O32 on the saturated acyl chain at the *sn1* position is generally  
16 conserved. The remaining reductions can be attributed to atoms C12, C13, C14, C15,  
17 O11 and O12). The total number of interactions with the cholesterol hydroxyl group  
18 displays an analogous pattern, as shown in Fig. 1C.

19  
20 With regards to the hydration of the PC headgroups, approximately 18% of POPC  
21 molecules H-bond with CHL and ENT, whilst 10% H-bond with EPI. The reduction  
22 in PC-sterol H-bonds in EPI is overcompensated for by an increase in solvent H-  
23 bonds, resulting in a total of 9.4, 9.4 and 9.5 H-bonds per POPC molecule, in the  
24 CHL, ENT and EPI simulations respectively. Thus it is effectively unchanged by the  
25 presence of different cholesterol isomers. This is noticeably less than in the pure  
26 POPC membrane simulation (~10.3 H-bonds per POPC molecules), which can largely  
27 be attributed to elevated H-bonding of phosphate oxygen atoms (O13 and O14; -  
28 32%), choline methyl groups (C13, C14, C15; -21%) and carbonyl atoms within the  
29 ester linkages (O22, O32; -20%) in the absence of cholesterol (Fig. 4B).

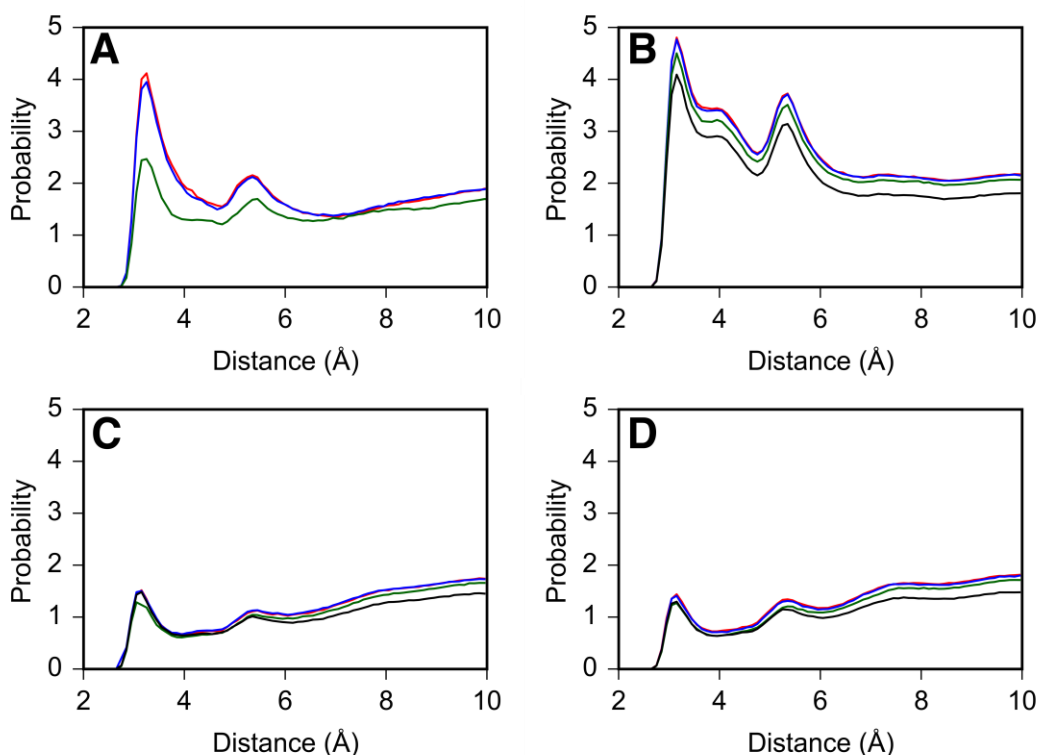
30  
31 Figure 4. (A) Frequency of PC-sterol H-bonds, per cholesterol molecule. (B)  
32 Frequency of PC-water H-bonds, per POPC molecule. CHL, ENT, EPI and POPC  
33 simulations are displayed in red, blue, green and black respectively. (C) Licorice  
34 representation of a POPC molecule coloured according to the percentage interaction  
35 with the cholesterol hydroxyl group. A distance criterion of 4 Å between heavy atoms  
36 is used in this case.



Charge-pairs also contribute significantly to the dynamics of the membrane headgroup region. Intermolecular charge-pairs can be formed between the methyl groups of the positively charged choline moiety of PC, and negatively charged groups in cholesterol and PC headgroups, including the cholesterol hydroxyl (O3), the non-ester phosphate oxygen atoms (O13, O14) and the carbonyl atoms within the ester linkages at the apex of the acyl chains (O22, O32). The frequency of CHL-PC charge pairs is significantly perturbed in EPI, likely as a direct result of the orientation of the O3 hydroxyl (Fig. 5A). Relative to the POPC-only membrane, the number of PC-PC charge-pairs is increased by approximately 15%, 14% and 3%, in CHL, ENT and EPI respectively. The number of charge-pairs involving O13 and O14 (POPC < EPI < CHL ≈ ENT) is inversely correlated in the observed trend for the values calculated for the area per lipid (POPC > EPI > CHL ≈ ENT) (Fig. 5B). It is apparent, however, that the distance between PC headgroups does not dictate the frequency of choline charge-pairs with O22 and O32 (Fig. 5C-D), and instead is likely directly influenced by the cholesterol headgroup. As a result, the number of charge-pairs for O22 displays the trend EPI ≈ POPC < CHL ≈ ENT, whilst the trend EPI < POPC ≈ CHL ≈ ENT is

observed for O32. Thus, the increased frequency of O13/O14 choline charge-pairs in EPI, relative to POPC, is offset by the reduction in O32 choline charge-pairs, resulting in an overall pattern of  $EPI \approx POPC < CHL \approx ENT$ .

Figure 5. Radial distribution function of (A) cholesterol O3, (B) lipid O13/O14, (C) lipid O32, and (D) O22 atoms with C12, C13 and C14 atoms of the lipid headgroup choline region, to represent the distribution of intermolecular charge pairs. CHL, ENT, EPI and POPC simulations are displayed in red, blue, green and black respectively.

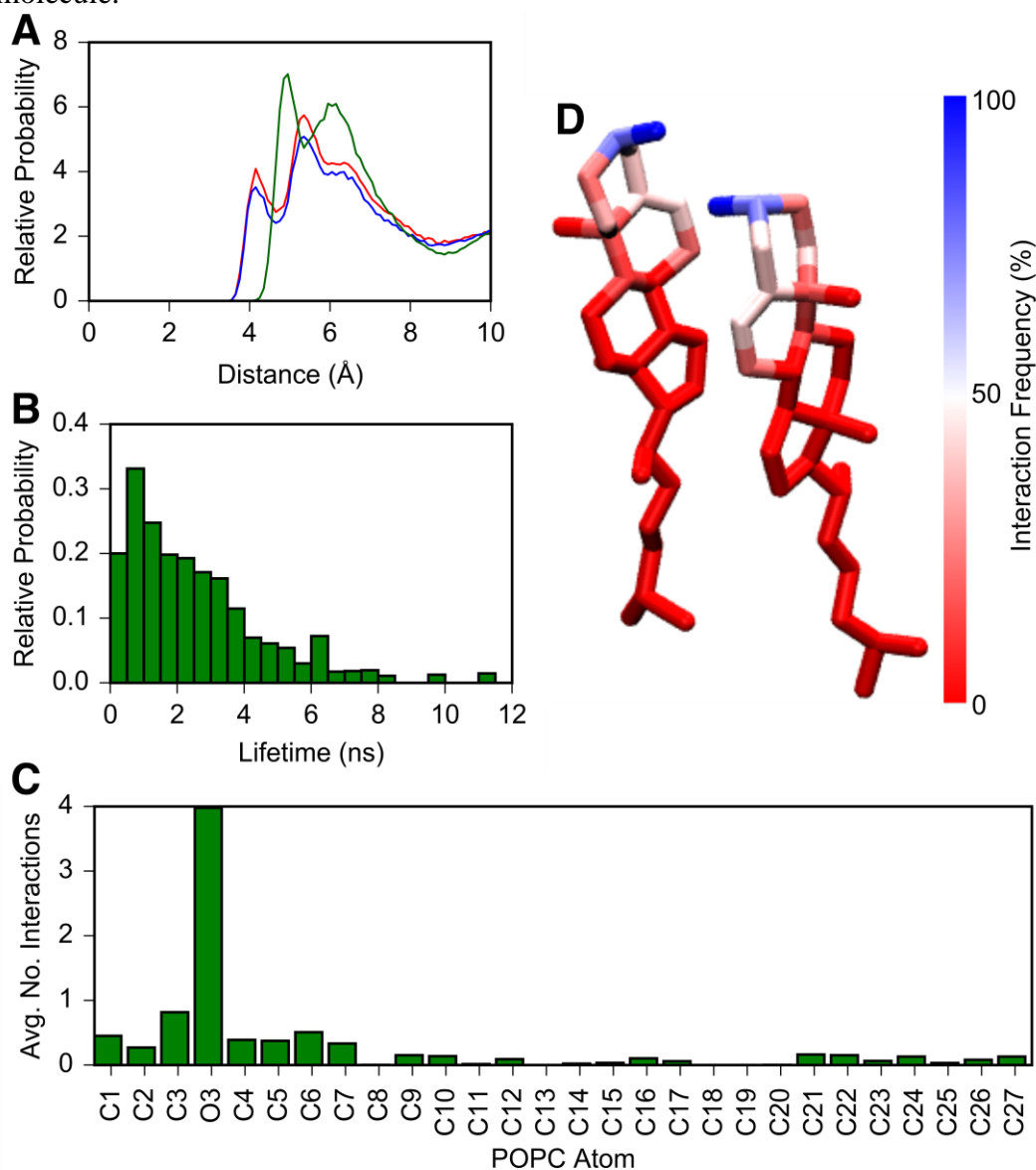


A further phenomenon worth considering is the formation of cholesterol dimers. The calculated RDF of the cholesterol C3 atoms, up to a distance of 10 Å, is shown in Fig. 6A. According to this data, all cholesterol isomers are guaranteed to be within ~6.5 Å of another cholesterol molecule, yet the nature in which such molecules interact is considerably different in EPI. The first RDF peak in CHL and ENT lies at 4 Å, whereas in EPI the first RDF peak ~5 Å and is approximately twice as probable, suggesting the alternative orientation of the hydroxyl group simultaneously acts as a spacer and a stabiliser. These results are consistent with the noticeable population of intermolecular cholesterol H-bonds identified in EPI by prior H-bond analysis. Using H-bond frequency as an indicator of dimer formation, the average lifetime of *epi*-cholesterol dimers has been calculated as  $0.9 \pm 1.1$  ns, with a maximum lifetime of ~11 ns (Fig. 6B). Characterisation of the interfacial region between two bound molecules (Fig. 6C) reveals the crucial interactions are localised in the hydroxyl



1 region, with the oxygen interacting with ~4 atoms on the adjacent molecule  
 2 simultaneously; the most probable interactions in descending order are O3, C3, C6  
 3 and C1 (Fig. 6D).

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 5 Figure 6. (A) RDF of cholesterol C3 atoms. CHL, ENT and EPI systems are shown in  
 6 red, blue and black respectively. (B) Weighted histogram of dimer lifetime, calculated  
 7 as the continuous occurrence of inter-cholesterol H-bonds. (C) Average number of  
 8 interaction pairs between *epi*-cholesterol molecules exclusively involved in dimers  
 9 (D) Interaction frequency between a hydroxyl O3 and an adjacent *epi*-cholesterol  
 10 molecule.  
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## 54 DISCUSSION

55 Of the cholesterol isomers in this study, *nat*-cholesterol exhibits the greatest influence  
 56 on the physical properties of the membrane, almost exactly replicated by its  
 57 enantiomer. *Epi*-cholesterol, on the other hand, exerts a weaker ordering effect as  
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1 represented by the  $S_{CD}$  order parameter, manifesting in a decrease in bilayer thickness.  
2 Such deviations can be attributed to adjustment of the position and tilt angle of *epi*-  
3 cholesterol in the phospholipid bilayer, and modification of the cholesterol surface  
4 available to interact with other cholesterol and phospholipid molecules.  
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6 The simulations presented here establish a direct link between the isomerisation of the  
7 cholesterol hydroxyl group and the occurrence of cholesterol dimers. Despite  
8  
9 significant interest in cholesterol aggregation and raft formation, experimental  
10 evidence concerning the existence of cholesterol dimers is negligible. Face-to-face  
11 dimers, stabilised by van der Waals interactions between the cholesterol  $\alpha$ -faces, were  
12 first proposed by Martin and Yeagle,<sup>51</sup> and have been examined in detail in recent  
13 years by computational means.<sup>52</sup> A study by Bandara et al, using unbiased MD  
14 simulations, calculated dimer lifetime at the order of hundreds of picoseconds, with  
15 instances of a nanosecond lifetime more prevalent at higher cholesterol and  
16 sphingomyelin concentrations.<sup>52</sup> Furthermore, Andoh and coworkers used  
17 thermodynamic integration to calculate the free-energy profile of cholesterol  
18 dimerization.<sup>53</sup> Cholesterol molecules separated by  $\sim 10$ - $15$  Å and interspersed by  
19 POPC acyl tails displayed an attractive interaction, with a well depth of  $-3.5$  kJ mol<sup>-1</sup>.  
20 This value approximates to a thermal energy of  $\sim 310$  K, highlighting the temperature  
21 dependence of the dimerization process.<sup>53</sup> Additional computational studies have  
22 noted the instability of artificial aggregates that rapidly dissociate to approximately  
23 monomeric units,<sup>54</sup> and the increased likelihood of cholesterol aggregates at elevated  
24 concentrations,<sup>55</sup> as well as the direct observation of raft formation in both all-atom  
25 and coarse-grained MD simulations.<sup>22, 56-58</sup> In this case, it is apparent the H-bonding  
26 properties of *epi*-cholesterol significantly enhance the stability of the dimer, resulting  
27 in an average lifetime at least one order of magnitudes greater than those previously  
28 calculated. The increased frequency of such assemblies inadvertently reduces the  
29 availability of the cholesterol  $\alpha$ -face, which forms the dimer interface, and can  
30 therefore modify the molecular interaction between other cholesterol and  
31 phospholipid molecules, as well as membrane proteins. A growing number of studies  
32 predicate a direct relationship between membrane cholesterol and GPCR  
33 oligomerisation,<sup>59-61</sup> with the involvement of the face-to-face dimer in the formation  
34 of a GPCR functional dimer postulated in earlier works.<sup>62</sup> It is therefore conceivable  
35 that evolved *epi*-cholesterol dimer species may directly interact with membrane  
36 proteins, translating as the observed functional changes. In the 5-HT<sub>1A</sub> receptor, for  
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1 example, the binding affinity of extracellular ligands is directly related to the presence  
2 of specific cholesterol isomers, *nat*- and *ent*- cholesterol in solubilized native  
3 hippocampal membranes.<sup>20</sup> Specific interactions between cholesterol and the 5-HT<sub>1A</sub>  
4 receptor will be explored in future computational studies.  
5

6 Bandari and coworkers have identified a close correlation between the activity of the  
7 5-HT<sub>1A</sub> receptor and the membrane dipole potential.<sup>17</sup> Dipole potential has been  
8 implicated in the modulation of channel gating in voltage-gated ion channels,<sup>63</sup> the  
9 formation of lipid rafts,<sup>64</sup> the clustering and ligand binding affinity of receptor  
10 tyrosine kinases<sup>64</sup> and the mechanism of anaesthetic action, and other small  
11 molecules.<sup>65</sup>  
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13 The membrane dipole potential emanates from the ordered orientation of lipid  
14 headgroups and associated water dipoles in the polar region separating the  
15 hydrocarbon interior from the solvent.<sup>66</sup> Regulation of membrane dipole potential by  
16 cholesterol has been ascribed to a number of factors, including (1) membrane  
17 condensation, resulting in increased charge density, (2) an ordered water network in  
18 the interfacial region, (3) reorientation of the P<sup>+</sup>-N<sup>-</sup> dipole in the phospholipid  
19 headgroup towards the membrane plane, and (4) the inherent dipole of cholesterol  
20 molecule itself.<sup>14, 63</sup> These hypotheses will be discussed in relation to the known  
21 experimental trend for the bilayer compositions under investigation: *epi*-cholesterol  
22 (~338 mV) < POPC-only (~369 mV) < *ent*-cholesterol (~480 mV) < *nat*-cholesterol  
23 (~500 mV).<sup>17</sup>  
24

25 Immediately, it can be proposed that membrane condensation has a minor effect on  
26 the dipole potential, as the calculated area per lipid is similar in the cholesterol  
27 isomers examined in this study. The relationship between headgroup hydration is  
28 more complex, however. The interaction of water with phospholipid bilayers has been  
29 investigated in detail in numerous studies;<sup>67-70</sup> revealing water molecules specifically  
30 orientate parallel to the bilayer normal, with hydrogen atoms directed towards the  
31 membrane, thus providing positive contributions to the dipole potential.<sup>63</sup> In theory,  
32 therefore, further hydration will increase the membrane dipole potential, whilst  
33 disruption will decrease it. Using MD simulations Villareal and coworkers attributed  
34 the observed changes in dipole potential of DMPC membranes in the presence of  
35 trehalose to replacement of lipid-water H-bonds with lipid-trehalose H-bonds, despite  
36 maintenance of the overall number of H-bonds.<sup>71</sup>  
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1 Here, we observe the loss of a single H-bond when cholesterol isomers are included in  
2 the bilayer systems. As a uniform decrease in dipole potential is not observed  
3 experimentally for the isomers we have examined, it seems unlikely that the  
4 organisation of ordered water molecules around the phospholipid headgroups  
5 underlies the experimental trend of the dipole potential, in contrast to the conclusions  
6 by Villareal et al concerning the DMPC-trehalose system.  
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10 Modification of the dipole potential has also been traced to conformational changes in  
11 the phospholipid headgroup, specifically the behaviour of the P<sup>+</sup>-N<sup>-</sup> dipole. The  
12 orientation of the P<sup>+</sup>-N<sup>-</sup> dipole has been proposed as a significant contributor to the  
13 difference in dipole potential observed between PC and PE headgroups, as well as the  
14 dynamics of the interfacial water.<sup>72</sup> The PE P<sup>+</sup>-N<sup>-</sup> dipole is thought to occupy a  
15 position more parallel to the membrane, relative to PC, maintain optimal contacts  
16 with the phospholipid acyl chains. Concurrently, the negative contribution to the  
17 dipole potential is reduced, resulting in a more positive overall value. This theory is  
18 supported by quadrupole splitting of NMR spectra of deuterated phospholipids.  
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20 In the context of this study, the calculated intermolecular charge-pair frequency can  
21 be directly correlated to the orientation of the P<sup>+</sup>-N<sup>-</sup> dipole. The increased number of  
22 charge-pairs involving phosphate oxygen atoms, for all isomers, can be qualitatively  
23 attributed to the decreased area per lipid. However, differential effects are observed  
24 for the carbonyl oxygen atoms involved in the ester linkages, suggesting a  
25 conformational change of the P<sup>+</sup>-N<sup>-</sup> dipole is responsible. Naturally, formation of  
26 charge pairs between the choline methyl groups and the carbonyl oxygens involved in  
27 the ester linkages cannot be formed when the P<sup>+</sup>-N<sup>-</sup> dipoles lie perpendicular to the  
28 membrane plane. Interestingly, the bilayer containing *epi*-cholesterol displays the  
29 lowest frequencies for O22 and O32 and therefore, the most perpendicular, with the  
30 latter value in close agreement with the POPC-only bilayer. The combined values are  
31 in overall agreement with the experimental trend for the dipole potential (*epi*-  
32 cholesterol < POPC-only < *ent*-cholesterol < *nat*-cholesterol).  
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34 Following this, the molecular mechanism underlying this phenomenon can be  
35 proposed. In terms of cholesterol-phospholipid atoms, we have demonstrated that the  
36 ester linkage of POPC forms the closest interactions with individual cholesterol  
37 molecules. However, the nature of the interactions differs significantly when  
38 comparing *nat*- and *ent*-cholesterol with *epi*-cholesterol. The number of interactions  
39 with the ester carbonyls is considerably increased when *epi*-cholesterol is present,  
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1 inadvertently obstructing potential interactions with other molecules in the system.  
2 Furthermore, polar interactions between the ester carbonyls and the hydroxyl group of  
3 *nat*- and *ent*-cholesterol are dominant, whereas the ester carbonyls primarily interact  
4 with the hydrophobic carbon backbone in *epi*-cholesterol. It is evident, therefore, that  
5 the formation of polar interactions between the ester carbonyls and other molecules in  
6 the system, in particular, will be less favourable when *epi*-cholesterol is present.  
7 These simple steric and electrostatic arguments can, therefore, be used to rationalize  
8 the diminished likelihood of charge-pair formation with the ester carbonyls observed  
9 when *epi*-cholesterol is present, which promotes the perpendicular orientation of the  
10 P<sup>+</sup>-N<sup>-</sup> dipole, relative to the bilayer surface. Overall, interpretation of the data  
11 presented in this study reveals the dynamics of P<sup>+</sup>-N<sup>-</sup> dipole in the phospholipid  
12 headgroup play a major role in modulation of the dipole potential of the membrane.  
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## 23 CONCLUSIONS

24 Using extensive MD simulations, totalling 2  $\mu$ s, the behaviour of *epi*-cholesterol in an  
25 unsaturated phospholipid bilayer has been characterised, and compared to *nat*-  
26 cholesterol and *ent*-cholesterol, as well as a cholesterol-free phospholipid bilayer.  
27 Slight differences in membrane thickness, area per lipid, interdigitation fraction and  
28 S<sub>CD</sub> order parameter are observed, relative to *nat*-cholesterol and *ent*-cholesterol  
29 which display identical properties throughout, in agreement with experimental  
30 studies. This can be attributed to minor deviations of the position and tilt angle of *epi*-  
31 cholesterol, as well as an observable population of long-lasting *epi*-cholesterol  
32 dimers, which are directly stabilised by H-bonding of the axial hydroxyl group on the  
33 cholesterol  $\alpha$ -face.  
34

35 Noteworthy variations are evident in the membrane-water interfacial region when *epi*-  
36 cholesterol is present, with the phospholipid SASA and interaction profile more  
37 closely resembling that of a cholesterol-free membrane. Following this, we have  
38 considered in detail how the divergent atomic-level interactions directly influence the  
39 dipole potential of the membrane, and provide a comprehensive mechanism for the  
40 action of cholesterol in this respect.  
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43 The membrane dipole potential is known to regulate transport processes, directly  
44 through the plasma membrane and also via embedded proteins. Consequently,  
45 understanding how the membrane dipole potential is modulated by various factors,  
46 such as the inclusion of small molecules, is of intense interest in the pharmaceutical  
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1 industry. Therefore, it is desired that the outcomes of this study provide crucial  
2 insights in this area, and contribute to our current understanding of the behaviour and  
3 role of the cholesterol in the cell membrane.  
4

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9

10 Conflict of Interest: Authors declares no conflict of interest.  
11

12 Ethical approval: This article does not contain any studies with human participants or  
13 animals performed by any of the authors.  
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5<sup>th</sup> October 2017

Dear Editorial Board,

Please find attached a manuscript to be considered for publication in The Journal of Membrane Biology, by Victoria Oakes and Carmen Domene. It is entitled: 'Stereospecific Interactions of Cholesterol in a Model Cell Membrane: Implications for the Membrane Dipole Potential', and it will be part of the special issue "Lipid membranes & reactions at lipid interfaces" organised by professors Ana-Nicoleta Bondar and Sandro Keller.

Yours faithfully,

A handwritten signature in red ink that reads "C. Domene".

Carmen Domene