



Inhibitory effect of hydroalcoholic and flavonoids extracts of *Dracocephalum kotschyi*, and its components luteolin, apigenin and apigenin-4'-galactoside on intestinal transit in mice

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ABSTRACT

Introduction: *Dracocephalum kotschyi* is an Iranian indigenous herbal plant which has been reported to have antispasmodic activity *in vitro*. The antispasmodic activity of hydroalcoholic extract of *D. kotschyi* is reported to be due to its flavonoids constituents including apigenin and luteolin. The objective of this research was to compare antispasmodic activities of hydroalcoholic and flavonoids extracts of *D. kotschyi* on ileum contractions *in vivo*. In addition, spasmolytic activity of apigenin, luteolin and apigenin-4'-galactoside were compared.

Methods: The hydroalcoholic extract was prepared by percolation method. Flavonoids extract was obtained by solvent-solvent extraction. Antispasmodic effect of the test compounds was assessed by measurement of percent small intestine transit following oral administration of a charcoal meal and compared with control group and standard drug loperamide.

Results: Biochemical assessment of flavonoids content of the extracts showed that ethyl-acetate fraction contained higher quantity of flavonoids. Loperamide (2.5 mg/kg) reduced charcoal meal movement by 58% in comparison to control group. Hydroalcoholic extract of *D. kotschyi* (20 mg/kg) and its ethyl-acetate fraction (20 mg/kg) reduced the intestinal charcoal meal transit by 32% and 90%, respectively. Apigenin, luteolin and apigenin-4'-galactoside with oral dose of 20 mg/kg inhibited intestinal movement of the charcoal meal 93%, 89% and 45%, respectively in comparison with the vehicle treated control groups.

Conclusion: This study confirms that both the hydroalcoholic and flavonoids extracts of *D. kotschyi* have antispasmodic properties *in vivo*. Antimotility of apigenin-4'-galactoside in mice is probably due to release of apigenin in the gastrointestinal tract.

Implication for health policy/practice/research/medical education:

This paper provides pharmacological evidence for antimotility of apigenin and luteolin two component of *Dracocephalum kotschyi* *in vivo*. In addition it was revealed that inactive apigenin-4'-galactoside in the gastrointestinal tract is converted to active apigenin.

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Introduction

Dracocephalum kotschyi Boiss. is a traditional Iranian medicinal plant belonging to Labiatae family (1). *D. kotschyi* is an aromatic plant enriched in essential oil including carveol, α -pinene, geraniol, α -citral, cyclononadiene, terpinene-4-ol, linalool, neral, myrcene, germacrene-D, isopinocarveol, limonene and α -terpineol (2-4). In recent years several reports have been published

on pharmacological activities of *D. kotschyi*. For instance, the essential oil of *D. kotschyi* reported to have anti-inflammatory and antinociceptive activities (5). Hydroalcoholic extract of *D. kotschyi* was reported to have antihyperlipidemic effect in an animal model (6) and inhibit tumor proliferation (7-9). Hydroalcoholic extract of *D. Kotschyi* has spasmolytic activities on isolated ileum and uterus (10,11). In addition, *D. kotschyi*

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extract has also been shown to have immunomodulatory, anti-inflammatory and anti-colitis properties (12,13). It is believed that pharmacological effect of the extract is mainly due to presence of flavonoids constituents (14). Flavonoids are widely distributed in the plant kingdom and occur in many medicinal plants (13,14). The flavonoids constituents of *D. kotschyi* extract include, apigenin, apigenin 4'-O-beta-D-glucopyranoside, luteolin, luteolin 7-O-beta-D-glucopyranoside, luteolin 3'-O-beta-D-glucuronide, calycopterin, xanthomicrol, isokaempferide, acacetin 7-O-beta-D-glucopyranoside and rosmarinic acid (15, 16). It has been reported that calycopterin is responsible for immunomodulatory effect of *D. kotschyi* (17), while apigenin was reported to have anti-inflammatory action (18). Among flavonoids constituents of *D. kotschyi* apigenin and luteolin have potent antispasmodic activities *in vitro* (19). However, apigenin-4'-galactoside shows no significant antispasmodic activity on isolated rat ileum (20). Therefore, it seems that aglycones forms of the flavonoids existed in the plant are pharmacological active constituents. Nevertheless, it is likely that in the gastrointestinal tract glycoside molecule can be removed by gut enzymes and aglycones forms of flavonoids could be released. Therefore, first objective of this research was to investigate the antimotility effect of *D. kotschyi* flavonoids rich fraction with hydroalcoholic extract *in vivo*. The second objective was to compare antimotility of luteolin and apigenin with that of apigenin-4'-galactose as well as the total extracts to find out which have stronger antimotility activities.

Materials and Methods

Dracocephalum kotschyi aerial parts were collected from a cultivated farm in Fereydun-Shahr mountain base (in Isfahan province, Iran) and identified at the Botany Department of the Faculty of Sciences, University of Isfahan. A voucher specimen was deposited at the herbarium of the School of Pharmacy and Pharmaceutical Sciences of Isfahan University of Medical Sciences (No. 1519).

The plant materials were dried in shade and ground to powder using electrical miller (Keep, Korea). The powdered materials were subjected to extraction by 70% ethanol in a percolator apparatus. The ratio of plant powder to solvent for hydroalcoholic extract was 1:8. The solvent was evaporated under vacuum using rotary (Heidolph, Germany) at 70°C and the yield of hydroalcoholic extract was determined. The hydroalcoholic extract (100 g) then was subjected to solvent-solvent extraction using chloroform/water mixture (1:1) and shaken thoroughly and the chloroform layer was decanted. Fresh chloroform was added and the process was repeated twice. The aqueous layer was then mixed with ethyl-acetate (2:1) and shaken as before. The ethyl-acetate layer was decanted and the fresh ethyl-acetate was added and the process was

repeated five times. These fractions were concentrated under vacuum using rotary at 30°C for ethyl-acetate and at 90°C for the aqueous fraction (21-23). The yield of dried extract was calculated for each fraction.

Biochemical standardization of the extract

Total flavonoids content of the hydroalcoholic extract and other fractions were determined by aluminum chloride colorimetric method (24). Fifty milligrams of quercetin was dissolved in 50 mL methanol and then diluted to 4, 20, 100 and 500 µg/mL. The diluted standard solutions (0.1 mL) were separately mixed with 0.1 mL of 20% aluminum chloride, 0.1 mL of glacial acetic acid and 2.7 mL of methanol. After incubation at room temperature for 40 minutes in the dark, the absorbance of the reaction mixture was measured at 415 nm with a Jenway 5105 U.V/V spectrophotometer (England). The spectrophotometer was initially calibrated with blank solution. In the blank solution, aluminum chloride was substituted by the same amount of distilled water. The assessment was repeated 3 times. Similarly, 0.1 mL of hydroalcoholic extract or other fraction solutions were reacted with aluminum chloride for determination of their total flavonoids content.

Drugs and solutions

The following drugs and solutions were used in this research: *D. kotschyi* hydroalcoholic extract, aqueous and ethyl-acetate fractions, loperamide, apigenin, luteolin and apigenin-4'-galactoside. The hydroalcoholic extract was made up as 10 and 40 mg/mL stock solution in 50% ethanol and diluted in distilled water to obtain concentrations of 1, 4 and 10 mg/mL. Aqueous and ethyl-acetate fractions were made up as 10 and 50 mg/mL stock solution in ethanol and further serial dilution was made in distilled water (1 and 5 mg/mL). Loperamide was dissolved in ethanol as 1 mg/mL stock solution and was further diluted with distilled water (100 µg/mL). Apigenin, luteolin (Aktin Chemical Ins. China) and apigenin-4'-galactoside was made up as 4 and 20 mg/mL stock suspension or solution in DMSO. Further dilution was made in distilled water (0.2 and 1 mg/mL in 5% DMSO, respectively). Charchol (3%) plus tragacanth powder (5%) suspension was prepared in distilled water. Unless stated, all the chemicals were purchased from Merck (Germany).

Pharmacological studies

Male albino mice (20-30 g), bred in School of Pharmacy and Pharmaceutical Sciences (Isfahan University of Medical Sciences, Iran) animal house, were kept at room temperature. The animals were fasted overnight prior to the experiments with free access to water. All animals were handled in accordance with the internationally accepted principles for laboratory animal use and care (25). In this study effect of hydroalcoholic and aqueous and ethyl-acetate fractions, loperamide (2.5 mg/kg), apigenin

and luteolin (2.5, 5, 10 and 20 mg/kg) and apigenin-4'-galactoside (5, 10, 20 and 40 mg/kg) were examined on gut motility using charcoal meal transit test. All experiments were conducted in parallel with control groups treated with equivalent volume of the vehicle. Each dose of drug was examined on 10 separate mice. Stock solution was adjusted in such way that each mouse was given 0.5 mL of test drugs or extracts.

Charcoal meal transit test

In this experiment, movement of charcoal meal in the small intestine was assessed. For this purpose *D. kotschy* hydroalcoholic extract (5, 20, 80 and 120 mg/kg), flavonoids extract (5 and 20 mg/kg), apigenin and luteolin (2.5, 5, 10 and 20 mg/kg) and apigenin-4'-galactoside (5, 10, 20 and 40 mg/kg) or loperamide (2.5 mg/kg) was given orally to mice and 45 minutes later 0.5 mL of charcoal meal containing 3% charcoal plus 5% tragacanth suspension was administered orally. Thirty minutes after charcoal meal administration, each animal was sacrificed and distance of charcoal movement in the small intestine was measured.

Measurement and statistical analysis

Ileum transit was expressed as percentage of charcoal moved from pylorus to the caecum relative to the whole length of the ileum. All results were expressed as mean \pm standard error of mean (SEM) and compared with corresponding vehicle-treated control group using unpaired Student's *t* test. *P* values less than 0.05 was considered statistically significant. SigmaPlot computer program (version 11) was used for statistical analysis and plotting the graphs.

Results

The yield of hydroalcoholic extract was 23% (W/W). Amounts of total flavonoids content were 0.36%, 0.003%, 0.1% and 1.12% for hydroalcoholic, chloroform, aqueous and ethyl-acetate fractions, respectively.

Charcoal meal transit test

In the control group treated with distilled water, charcoal meal moved up to 97% of small intestine within 30 minutes of oral administration of charcoal meal. Loperamide (2.5 mg/kg), tested as a standard drug, reduced charcoal movement by 58% relative to the vehicle treated control group. There was no statistically significant difference between distilled water treated control and the control group treated with vehicle (ethanol) (Figure 1). Hydroalcoholic extract of *D. kotschy* with dose of 5 mg/kg had no significant inhibitory effect on intestinal charcoal meal transit but with dose of 20 mg/kg significantly reduced meal transit by 32%. When the dose of hydroalcoholic extract was increased to 80 mg/kg intestinal transit was inhibited by 70% compared to

vehicle treated control group (Figure 2). Further increase in dose of *D. kotschy* hydroalcoholic extract (120 mg/kg) had no more inhibitory effect on intestinal meal transit.

Ethyl-acetate fraction prepared from *D. kotschy* extract had more pronounced inhibitory effect on charcoal meal transit compared to hydroalcoholic extract. Oral dose of 5 mg/kg ethyl-acetate fraction reduced intestinal transit by 45% relative to vehicle treated control group. Ethyl-acetate fraction of *D. kotschy* with oral dose of 20 mg/kg inhibited the charcoal meal transit by 90% (Figure 1).

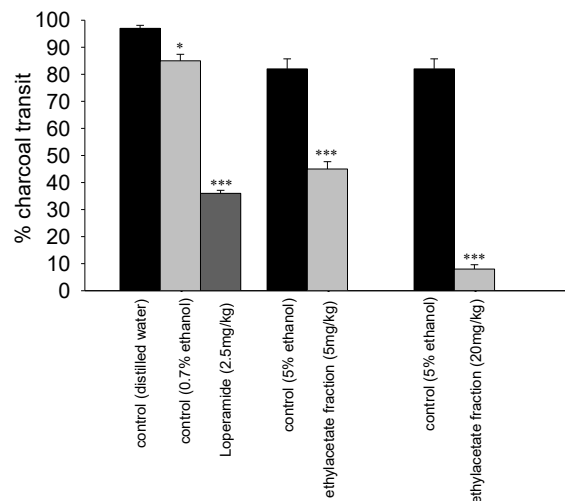


Figure 1. Effect of loperamide and ethyl-acetate fraction of *D. kotschy* on intestinal distance moved by charcoal meal (0.5 mL; per orum) in comparison with the vehicle treated control group in mice. Data are expressed as mean \pm SEM (n=10). Stars show degree of statistically significant difference in comparison with its appropriate control group (**P*<0.05, ****P*<0.001, Student's *t* test).

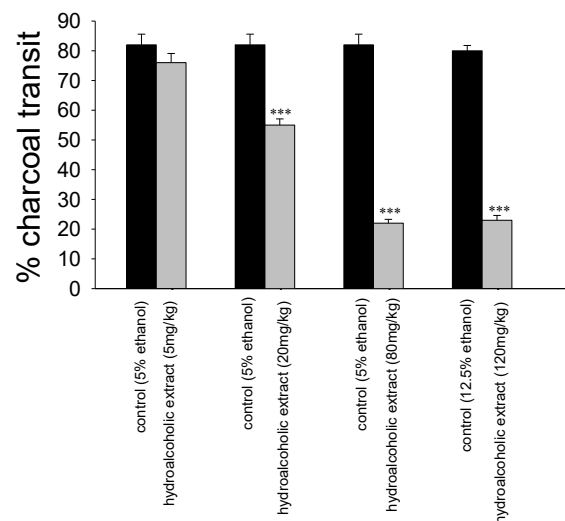


Figure 2. Effect of hydroalcoholic extract of *D. kotschy* on intestinal distance moved by charcoal meal (0.5 mL; per orum) in comparison with the vehicle treated control group in mice. Data are expressed as mean \pm SEM (n=10). Stars show degree of statistically significant difference in comparison with its appropriate control group (****P*<0.001, Student's *t* test).

Remaining aqueous fraction up to dose of 120 mg/kg exhibited no inhibitory effect on intestinal charcoal meal transit (Figure 3).

Luteolin and apigenin are two known flavonoids which have been identified in *D. kotschy* extract. Both these flavonoids components in a dose dependent manner inhibited mice intestinal motility. Luteolin and apigenin at oral dose of 2.5 mg/kg reduced intestinal charcoal meal transit by 40% and 42%, respectively compared to vehicle treated control group (Figures 4 & 5). With oral dose of 20 mg/kg, movement of transit meal was inhibited by 89% and 93%, respectively. Apigenin-4'-galactoside with oral dose of 5 mg/kg had no significant effect on charcoal meal transit. However, with higher doses (10, 20 and 40 mg/kg) significantly inhibited intestinal transit by 38%, 45% and

55%, respectively (Figure 6). In the control groups treated with vehicle (DMSO) there were no statistically significant differences compared with distilled water treated group.

Discussion

Hydroalcoholic extract of *D. kotschy* possessed spasmolytic activity on isolated rat ileum and uterus (10,11). In addition, *D. kotschy* extract has been shown to reduce diarrhea induced in mice (26). Furthermore, it has been shown to have anti-colitis activity in animal model of colitis (12). Antispasmodic activity of *D. kotschy* extract has been attributed to its flavonoids contents including luteolin and apigenin (20). In this research we have focused on antispasmodic action of hydroalcoholic and flavonoids extracts of *D. kotschy* in whole animal to

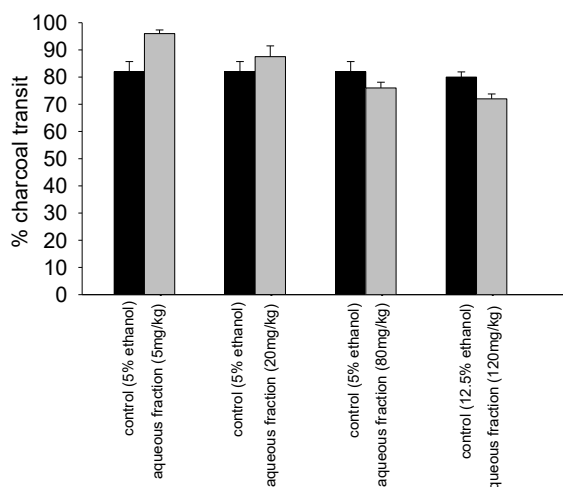


Figure 3. Effect of aqueous fraction of *D. kotschy* on intestinal distance moved by charcoal meal (0.5 mL; per orum) in comparison with the vehicle treated control group in mice. Data are expressed as mean±SEM (n=10). There is no statistically significant difference between test and control groups.

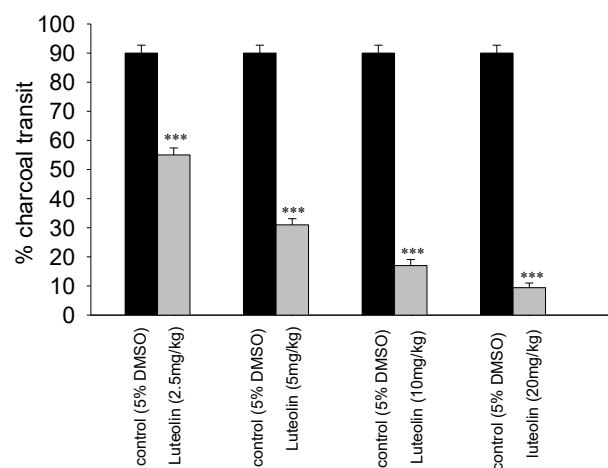


Figure 4. Effect of luteolin on intestinal distance moved by charcoal meal (0.5 mL; per orum) in comparison with the vehicle treated control group in mice. Data are expressed as mean±SEM (n=10). Stars show degree of statistically significant difference in comparison with its appropriate control group (*** P <0.001, Student's *t* test).

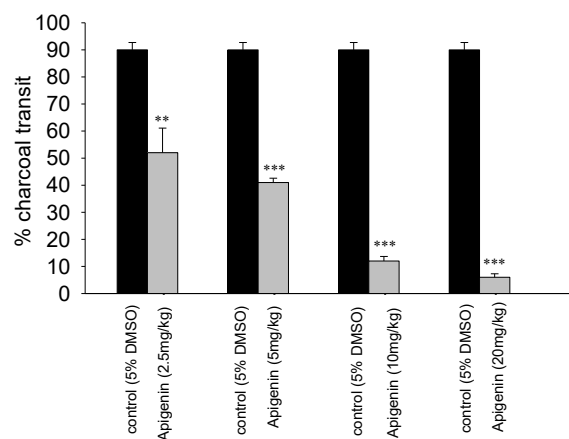


Figure 5. Effect of apigenin on intestinal distance moved by charcoal meal (0.5 mL; per orum) in comparison with the vehicle treated control group in mice. Data are expressed as mean±SEM (n=10). Stars show degree of statistically significant difference in comparison with its appropriate control group (** P <0.05, *** P <0.001, Student's *t* test).

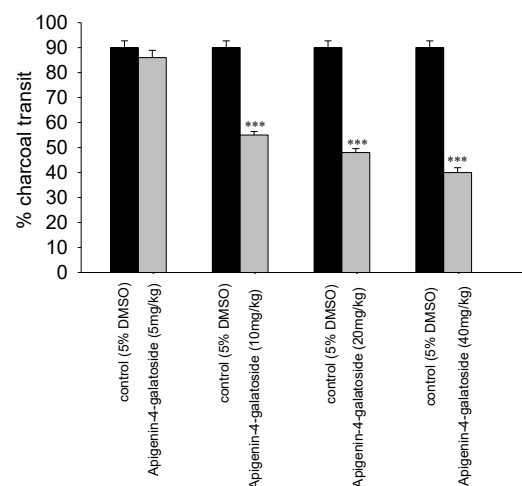


Figure 6. Effect of apigenin-4'-galactoside on intestinal distance moved by charcoal meal (0.5ml; per orum) in comparison with the vehicle treated control group in mice. Data are expressed as mean±SEM (n=10). Stars show degree of statistically significant difference in comparison with its appropriate control group (*** P <0.001, Student's *t* test).

find out which one would be more suitable remedy for gastrointestinal ailments. Furthermore, antispasmodic actions of three components of *D. kotschy* extract were also assessed *in vivo*.

Hydroalcoholic extract contains both polar and non-polar constituents while the chloroform fraction mainly contains non-polar constituents including non-active substances such as chlorophyll. In the mixture of water and ethyl-acetate, polar substances would be dissolved in the aqueous part while less polar substances including flavonoids are dissolved in ethyl-acetate layer. Biochemical assay also confirms that ethyl-acetate fraction is rich in flavonoids. Inhibition of small intestine transit also confirms that more active substances are concentrated in the flavonoids rich fraction. Measurement of transit of meal in the small intestine has been used for assessment of gut motility in intact animal. It has been shown that hydroalcoholic extract of *D. kotschy* with oral doses of 20 and 80 mg/kg caused significant inhibition of intestinal motility. Furthermore, increase in dose had no further inhibitory effect indicating that it is possible that other components presence in the extract cause its inhibitory action. As mentioned above most of flavonoids components are concentrated in the ethyl-acetate fraction. Ethyl-acetate fraction with dose of 20 mg/kg reduced intestinal motility by 90% while hydroalcoholic extract reduced by 32%. This indicates that flavonoids extract is pharmacologically more active than the hydroalcoholic extract. Luteolin and apigenin are two flavonoids component of *D. kotschy* with antimotility activities. Therefore, it can be concluded that they have major contribution in the antispasmodic activity of *D. kotschy* extract. Comparison of antimotility of luteolin and apigenin with similar doses indicates that they have similar pharmacological activities on intestinal meal transit. On the other hand, apigenin-4'-galactoside was less active in comparison to apigenin. However, unlike rat isolated ileum (20), apigenin-4'-galactoside inhibited gut motility *in vivo* which is a good indication that in the gastrointestinal tract, the galactoside molecule is removed and aglycone apigenin is released. Therefore, it is very likely that the observed inhibitory effect of apigenin-4'-galactoside is due to action of apigenin.

Conclusion

In conclusion this study clearly shows the antimotility effect of *D. kotschy* extract in intact animal and indicates that antispasmodic effect of extract is mainly due to its flavonoids contents including apigenin and luteolin. As galactoside form of apigenin and luteolin lack antispasmodic activity *in vitro*, this study indicates that in the gastrointestinal tract, the sugar moiety is removed and active aglycones flavonoids are released.

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Moazeni as Pharm. D student at Isfahan University of Medical Sciences.

Authors' contributions

MG was responsible for preparation of extract while HS supervised the pharmacological studies. SM was responsible for performing the laboratory works. All contributed to preparation of the article and confirmed final edition for publication.

Conflict of interests

None.

Ethical considerations

Ethical issues have been approved by Isfahan University of Medical Sciences ethical committee (No. 397538).

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References

1. Naghibi F, Mosaddegh M, Mohammadi Motamed M, Ghorbani A. Labiatae family in folk medicine in Iran: from ethnobotany to pharmacology. *Iranian J Pharm Res.* 2010;4(2):63-79.
2. Yaghmai M, Taffazoli R. The essential oil of *Dracocephalum kotschy* Boiss. *Flavour Fragrance J.* 1988;3:33-36.
3. Saeidnia S, Gohari AR, Uchiyama N, Ito M, Honda G, Kiuchi F. Two new monoterpene glycosides and trypanocidal terpenoids from *Dracocephalum kotschy*. *Chem pharm bull.* 2004;52(10):1249-50.
4. Sadraei H, Asghari G, Kasiri F. Comparison of antispasmodic effects of *Dracocephalum kotschy* essential oil, limonene and alpha-terpineol. *Res pharm Sci.* 2015;10(2):109-16.
5. Golshani S, Karamkhani F, Monsef-Esfehani HR, Abdollahi M. Antinociceptive effects of the essential oil of *Dracocephalum kotschy* in the mouse writhing test. *J Pharm Pharm Sci.* 2004;7(1):76-9.
6. Sajjadi ES, Movahedian Atar A, Yektaian A. Antihyperlipidemic effect of hydroalcoholic extract, and polyphenolic fraction from *Dracocephalum kotschy* Boiss. *Pharm Acta Helv.* 1998;73(3):167-70.
7. Talari M, Seydi E, Salimi A, Mohsenifar Z, Kamalinejad M, Pourahmad J. *Dracocephalum*: novel anticancer plant acting on liver cancer cell mitochondria. *BioMed Res Int.* 2014; 40(5): 1-10. doi: 10.1155/2014/892170.
8. Moghaddam G, Ebrahimi SA, Rahbar-Roshandel N, Foroumadi A. Antiproliferative activity of flavonoids: influence of the sequential methoxylation state of the flavonoid structure. *Phytother Res.* 2012;26:1023-28.

- doi:10.1002/ptr.3678.
9. Jahaniani F, Ebrahimi SA, Rahbar-Roshandel N, Mahmoudian M. Xanthomicrol is the main cytotoxic component of *Dracocephalum kotschy* and a potential anti-cancer agent. *Phytochemistry*. 2005;66:1581-92. doi:10.1016/j.phytochem.2005.04.035.
 10. Sadraei H, Asghari G, Kasiri F. Antispasmodic effect of *Dracocephalum kotschy* hydroalcoholic extract on rat ileum contraction. *Res Pharm Sci*. 2015;10(5):446-52.
 11. Sadraei H, Asghari G, Alinejad M. Comparison of antispasmodic effect of hydroalcoholic extract of *Dracocephalum kotschy* Boiss. in rat uterus and ileum. *Res Pharm Sci*. 2016;11(4):284-92. doi: 10.4103/1735-5362.189295.
 12. Sadraei H, Asghari G, Khanabadi M, Minaiyan M. Anti-inflammatory effect of apigenin and hydroalcoholic extract of *Dracocephalum kotschy* on acetic acid-induced colitis in rats. *Res Pharm Sci*. 2017;12(4):322-9. doi: 10.4103/1735-5362.212050.
 13. Kilani-Jaziri S, Mustapha N, Mokdad-Bzeouich I, El Gueder D, Ghedira K, Ghedira-Chekir L. Flavones induce immunomodulatory and anti-inflammatory effects by activating cellular anti-oxidant activity: a structure-activity relationship study. *Tumor Biol*. 2016;37:6571-79. doi: 10.1007/s13277-015-4541-5.
 14. Havsteen B. Flavonoids, a class of natural products of high pharmacological potency. *Biochem Pharmacol*. 1983;32:1141-8.
 15. Gohari AR, Saeidnia S, Matsuo K, Uchiyama N, Yagura T, Ito M, Kiuchi F, Honda G. Flavonoid constituents of *Dracocephalum kotschy* growing in Iran and their trypanocidal activity. *Nat Med*. 2003;57(6):250-2.
 16. Fattahi M, Nazeri V, Torras-Claveria L, Sefidkon F, Cusido RM, Zamani Z, et al. Identification and quantification of leaf surface flavonoids in wild-growing populations of *Dracocephalum kotschy* by LC-DAD-ESI-MS. *Food chem*. 2013;141(1):139-46. doi: 10.1016/j.foodchem.2013.03.019.
 17. Faham N, Javidnia K, Bahmani M, Amirghofran Z. Calycopterin, an immunoinhibitory compound from the extract of *Dracocephalum kotschy*. *Phytother Res*. 2008;22(9):1154-8. doi: 10.1002/ptr.2382.
 18. Lee JH, Zhou HY, Cho SY, Kim YS, Lee YS, Jeong CS. Anti-inflammatory mechanism of apigenin: inhibition of cyclooxygenase expression, adhesion of monocytes to human umbilical vein endothelial cells and expression of cellular adhesion molecules. *Arch Pharm Res*. 2007;30(10):1318-27.
 19. Sadraei H, Ghanadian M, Asghari G, Sekhavati N. Antispasmodic activity of apigenin and luteolin, two components of *Dracocephalum kotschy* extract, on rat ileum contractions. *J Herbmed Pharmacol*. 2018;7(2):100-5. doi: 10.15171/jhp.2018.17
 20. Sadraei H, Ghanadian M, Asghari G, Sekhavati N. Bioactivity guided isolation of active fraction of *Dracocephalum kotschy* extract responsible for antispasmodic activity on rat ileum. [thesis]. Isfahan, Iran: School of Pharmacy, Isfahan University of Medical Sciences and Health Services; 2018.
 21. Samuelsson G. *Drugs of Natural Origin*. Stockholm: Swedish Pharmaceutical Press: Sweden; 1999:48-9.
 22. Ghasemi Dehkordi NA, Sajadi SE. *Iranian Herbal Pharmacopoeia*. Tehran: Ministry of Health Pub. 2002.
 23. Harborn JB, Mabary TJ, Mabary H. Physiological and functional of flavonoids. *Phytochemical Methods. Flavonoids*. 1975:970-1055.
 24. Chang CC, Yang MH, Wen HM, Chern JC. Estimation of Total Flavonoid Content in Propolis by Two Complementary Colorimetric Methods. *J Food Drug Anal*. 2002;10(3): 178-82.
 25. Committee for the update of the guide for the care and use of laboratory animals National Research Council. *Guide for the Care and use of Laboratory animals*. Washington DC: The National Academies Press; 2010. p. 11-37.
 26. Sadraei H, Asghari G, Shahverdi F. Antidiarrhoeal assessment of hydroalcoholic and hexane extracts of *Dracocephalum kotschy* Boiss. and apigenin in mice. *Res Pharm Sci*. 2016;11(3):200-9.