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Session V: Getting Vaccines to the Market: Case studies

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Time to Apply the 3 R's to Virus Testing?

Rebecca Sheets, Grimalkin Partners

ECI VTI, Albufeira, Portugal, June 13, 2016

Disclaimer

- This presentation represents solely the opinions of the speaker and does not reflect any U.S. government policies or the opinions of the National Institutes of Health or the Food & Drug Administration.

Context

- 3 R's – reduce, refine, or replace the use of animals in product testing
 - Lack of regulatory convergence
- EU has directive (mandatory law) requiring that 3 R's be applied; whenever *in vitro* methods are available, *in vivo* tests should not be used.
 - Illegal for European companies to not comply, but if they sell to US or other countries that still require *in vivo* tests, they are in catch 22.
 - EDQM is changing EP in consideration of data from study I will present
- US has public law to REVIEW their regulations to see where 3 R's could be applied, but only policy to actually apply 3 R's
 - Despite data I will present, FDA has not changed requirements (policy is not translated into action)

Introduction

- Routine Tests for Adventitious Viruses
 - *in vivo* with mortality or morbidity read-outs
 - In tissue culture with CPE & HAd read-outs
 - Transmission Electron Microscopy
 - Specific PCRs for selected viruses
 - PCR-based reverse transcriptase assay
 - (infectivity for retroviruses)
 - Bovine, porcine viruses (9 CFR tests in cell culture with CPE, HAd, and IFA read-outs)
 - (MAP, RAP, HAP – *in vivo* or PCR)

“Routine” Adventitious Virus Tests

Purpose of Work

- Provide regulators and manufacturers with information needed for decision-making
 - Such info normally comes from assay validation
- Provide baseline data to serve as basis of comparison for new methods
- Provide protocols and viral stocks to permit “direct” comparisons by developers of new methods
- Determine “value added” by *in vivo* methods in consideration of 3 R’s policy

“Routine” Adventitious Virus Tests Breadth & Sensitivity

- These tests were developed for clinical diagnostics in mid-20th century
- Initially used to detect SPECIFIC adventitious agents
- Use expanded to broad general screening assays
- Breadth/sensitivity had not been systematically assessed & published
- Not validated in the manner currently developed assays would be required
- No regulatory requirements to do so and costly to do

“Routine” Adventitious Virus Tests Implementation Phase

- The prime contractor, Advanced BioScience Laboratories, awarded task to Charles River Labs to implement this project
 - Compliant with Good Laboratory/Manufacturing Practices
 - Experienced with routine adventitious agent testing
 - *In vivo* and *in vitro* capabilities
 - Virology expertise to prepare and characterize viral stocks required



“Routine” Adventitious Virus Tests Implementation Phase (2)

- Viral stocks prepared in cell culture
 - Titrated in production cell line or positive control cell line
 - Characterized for purity & identity
- *In vivo* testing
 - Test at highest concentration for breadth
 - If positive, sensitivity determined by titration (dilutions)
- *In vitro* testing
 - Breadth and sensitivity assessed simultaneously

“Routine” Adventitious Virus Tests

Results and Conclusions

The results can answer questions
such as:

- Is using two human cell lines useful?
- Is a 14-day *in vitro* test sufficient or are 28 days needed?
- Is sub-passage useful for suckling mouse test sensitivity?
- Which is more sensitive – *in vitro* or *in vivo*?
- Yes, MRC-5 & HeLa had different sensitivities, sometimes one was better, sometimes the other
- 28 days more sensitive in some cases
- No, for the viruses tested
- With the exception of flu & VSV, the *in vitro* tests were always more sensitive, generally by logs, sometimes the difference between detecting & not detecting

“Routine” Adventitious Virus Tests Results and Conclusion

Table 4. *In Vitro* Limit of Detection of Research Virus Stocks

Virus	Vero		MRC-5		HeLa		Other	
	CPE	HA/HAD/IF ^a	CPE	HA/HAD/IF ^a	CPE	HA/HAD/IF ^a	CPE	HA/HAD/IF ^a
Adenovirus 5								
Adenovirus 41								
BVDV								
BoPIV-3							nd ^b	nd
Coxsackie A16								
Coxsackie B3						nd	nd	nd
Echovirus 11							nd	nd
Influenza A							nd	nd
HSV-1							nd	nd
Measles							nd	nd
Mumps							nd	nd
Rhinovirus 2		nd		nd		nd	nd	nd
Rubella								
Simian CMV								
SV-40							nd	nd
VSV		nd		nd		nd	nd	nd

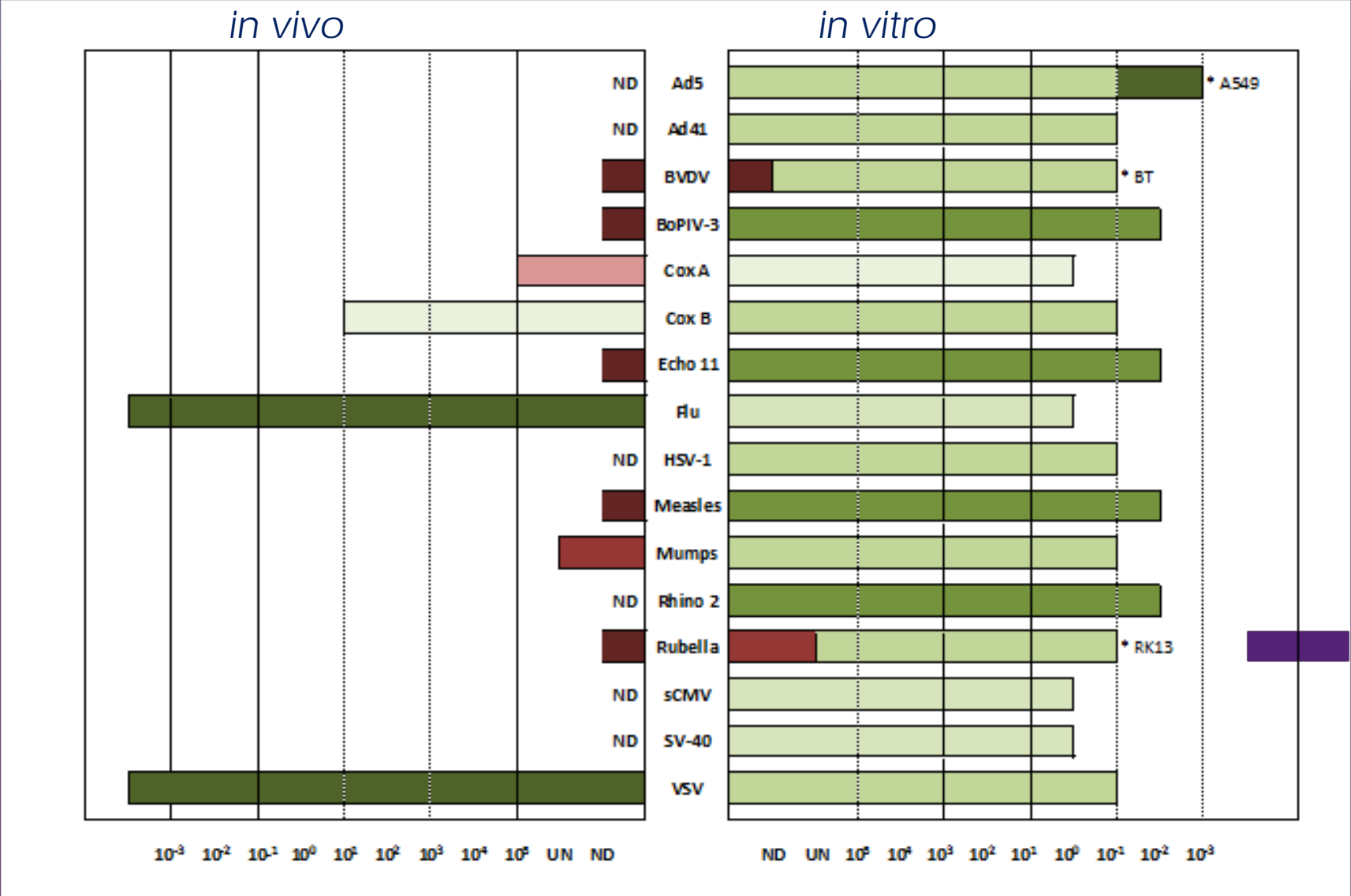
^aAll virus-infected cultures were tested for hemadsorption activity except BVDV (immunofluorescence), influenza A and rubella (hemagglutination) and rhinovirus 2 and VSV (CPE only)

^bNot done

^b Detection by immunofluorescence

0.001 ID	
0.01 ID	
0.1 ID	
1.0 ID	
10.0 ID	
100.0 ID	
1,000.0 ID	
100,000.0 ID	
Undiluted	
Undetected	

“Routine” Adventitious Virus Tests Results



“Routine” Adventitious Virus Tests Outcomes/Deliverables

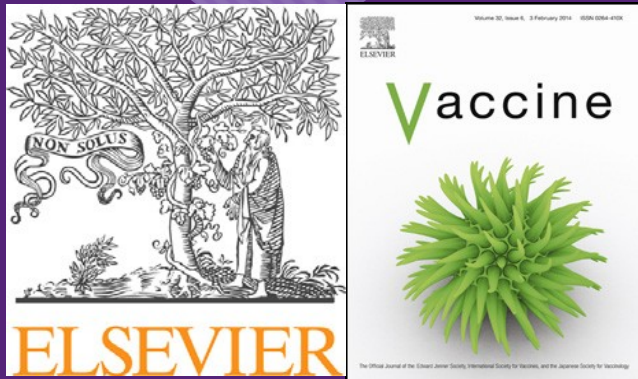
- Viral stocks will be made available through the NIAID/DAIDS *Reagent Resource Support Program for AIDS Vaccine Development*
 - <http://www.niaid.nih.gov/topics/hivaids/research/vaccines/resources/reagent/pages/default.aspx>
 - A research repository, not a regulatory authority control lab reagent repository
 - Not international reference materials, but research reagents
- Protocols for virus preparation, titration, and for *in vivo* and *in vitro* test methods will also be made available

“Routine” Adventitious Virus Tests

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Publication



Systematic evaluation of *in vitro* and *in vivo*
adventitious virus assays for the detection of viral
contamination of cell banks and biological products

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Time to Apply 3 R's to Virus Testing?

YES