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ORGANOSOLV PRETREATMENT AS A MAJOR STEP OF LIGNOCELLULOSIC BIOMASS REFINING

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ABSTRACT

Five solvents (ethanol, methanol, diethylene glycol, acetone, butanol) were evaluated for organosolv pretreatment of wheat straw. The biorefinery process was catalyzed by H_2SO_4 at 160 °C for 20 min. Maximum cellulose concentration was 72% w/w using acetone while delignification was 59% w/w and 95% w/w of xylan hydrolyzed to xylose.

INTRODUCTION

The conversion of biomass within biorefineries is seen as a potential alternative to current reliance on non-renewable resources. The transition from a traditional "oil-refinery" to a "bio-refinery", based on renewable lignocellulosic biomass, is crucial if we are to move to a more environmentally friendly economy ($\underline{1}$). Lignocellulosic biomass receives more attention because it does not compete as a food resource, and it can reduce carbon dioxide in the atmosphere by up to 75–100% ($\underline{2}$).

The key components of lignocellulosic biomass, i.e., cellulose, hemicelluloses and lignin, are closely associated with each other at the plant cell level. This close association, together with the partly crystalline nature of cellulose, reduces cellulose reactivity towards acid and enzymatic hydrolysis in native biomass $(\underline{3})$. Thus, organosolv pretreatment is necessary to render the carbohydrate fraction to acid, enzymatic and microbial action $(\underline{4})$.

Biorefining is the sustainable processing of biomass into a spectrum of marketable products (food and feed, materials, and chemicals) and energy (fuels, power, and heat). In biorefinery appropriate fractionation of the complex lignocellulose material, into its constituents, is of most importance. Fig. 1 depicts a possible lignocellulosic biorefinery scheme (5-6).

From structure study to pulping and currently to energy usage organosolv fractionation has a long history. The earliest study applying organic solvents to treat lignocellulosic material was back in 1893, when Klason (7-8) used ethanol and hydrochloric acid to separate wood into its components to study the structure of lignin and carbohydrates. Nowadays organosolv pretreatment has been used for lignin and other potentially valuable co-products (e.g. acetone, butanol, biogas) production (9-14).

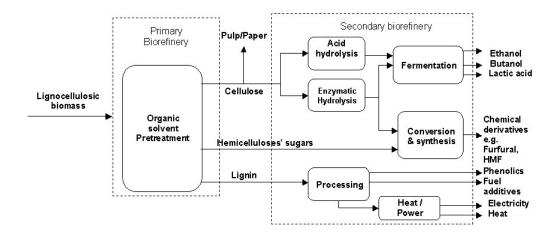


Fig.1: Organosolv-based lignocellulosic biorefinery

Treatment with organic solvents involves the use of an organic liquid and water, with or without addition of a catalyst agent (acid or base). This mixture partially hydrolyzes lignin bonds and lignin–carbohydrate bonds, resulting in a solid residue composed mainly by cellulose and some hemicellulose. Organosolv pretreatment efficiently removes lignin from lignocellulosic materials but most of the hemicellulose sugars are also solubilized by this process.

As catalysts, organic or inorganic acids, bases, and so on have been tested for lignocellulosic biomasses ($\underline{15}$). Sulfuric acid, a strong mineral acid, is one of the most-used reagents for the pretreatment process due to its strong reactivity and high efficiency ($\underline{16}$). Although ethanol seems to be the most popular organic solvent over the years ($\underline{17-20}$) other solvents like methanol ($\underline{21}$), acetone ($\underline{22}$) formic acid ($\underline{23-26}$), acetic acid have been also studied ($\underline{27-30}$).

This study focused on the organosolv pretreatment process of wheat straw which facilitates hydrolysis and fermentation processes. Ethanol (CH $_3$ CH $_2$ OH), Methanol (CH $_3$ OH), Diethylene glycol (C $_4$ H $_1$ OO $_3$), Acetone (C $_3$ H $_6$ O) and Butanol (C $_4$ H $_9$ OH) were evaluated as solvents. Sulfuric acid was the pretreatments' catalyst. The effect of the five organic solvents on pretreatment results was analyzed.

MATERIALS AND METHODS

2.1 Raw material

The wheat straw used was obtained from Kapareli Village of Thebes, Greece, as a suitable source for full-scale industrial applications. The moisture content of the material when received was 9% w/w; after screening, the fraction with particle sizes between 10 and 20 mm was isolated. The composition of the raw material was as follows (expressed in % w/w on a dry weight basis): 33.7% cellulose, hemicelluloses

24.1% (xylan 18.9 %, arabinose 2.7% and acetic groups 2.5%), 17% Klason lignin (acid insoluble) and 14.4% others.

2.2 Experimental equipment and procedures

A 3.75-L batch reactor PARR 4843 was used for the organosolv fractionation. Reaction ending temperature was 160 $^{\circ}$ C whereas the reaction time was 20 min (not including the preheating time). The reaction was catalyzed by H₂SO₄, 0.045 N, in a 50% v/v aqueous/organic solvent solution; the liquid/solid ratio was 20/1. The organic solvents used were: ethanol, methanol, diethylene glycol, acetone and butanol.

2.3. Analytical techniques

The Saeman et al. (1945) technique was used for the quantitative saccharification of the original lignocellulosic material as well as the autohydrolysis solid residues. The filtrates from the quantitative saccharification, as well as those from the autoclave (before and after the secondary hydrolysis, i.e. post-hydrolysis with 0.9 N H_2SO_4 at 100 °C for 4.5 h), were analyzed for their content of glucose and xylose enzymatically (Megazyme D-Glucose HK Assay kit K-GLUHK-220A and D-xylose Assay kit K-xylose). Glucose and xylose were produced via quantitative saccharification of the solid residue in each one of the experiments. Cellulose was estimated as glucan, and hemicelluloses were estimated as xylan. Finally, the acidinsoluble lignin (Klason lignin) was determined according to the Tappi T222 om-88 method.

RESULTS AND DISCUSSION

Due to the different molecular structure of the examined organic solvents the temperature and pressure profiles are presented in Fig. 2. As shown in Fig.2b the maximum pretreatments' pressures varie among the different organic solvents between 4.9 (Diethylene glycol) and 13.6 bar (Acetone). More specifically, were: 12.2 bar for Methanol, 11.2 bar for Ethanol and 8.6 bar for Butanol acid catalysed organosolv pretreatment.

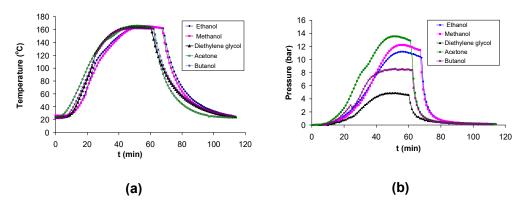


Fig.2: Organosolv pretreatment's (a) temperature and (b) pressure profile vs. time. Sulfuric acid 0.045N; temperature 160°C; time 20 min; liquid: solid ratio = 20:1.

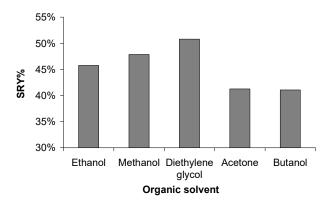


Fig.3: Effect of organic solvent on Solid Residue Yield. Sulfuric acid 0.045N; temperature 160°C; time 20 min; liquid: solid ratio = 20:1.

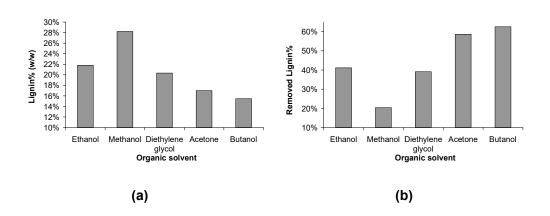


Fig.4: Effect of organic solvent on (a) lignin composition and (b) removed lignin percentage. Sulfuric acid 0.045N; 160°C; 20 min; liquid: solid ratio = 20:1.

The highest percentage (50.8%) of Solid residue yield (SRY) was achieved, as shown in Fig.3, using diethylene glycol as organic solvent. Regarding lignin removal (Fig.4) methanol use appears to have the poorest results just 20.4% while butanol has the best 62.6%. When examined the correlation between removed lignin % and SRY % (Fig.5) a simple linear regression fits the data. It must be noted that diethylene glycol was not included in the graph due to its disproportionally high delignification compared to SRY %. Diethylene glycol acid pretreatment combines high delignification percentage with low cellulose removal (Fig.6). All five solvents show low cellulose removal (See Fig. 6b).

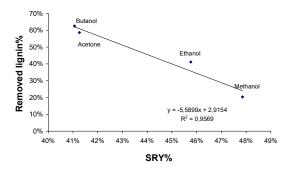


Fig.5: Removed lignin vs. Solid Residue Yield. Sulfuric acid 0.045N; temperature 160°C; time 20 min; liquid: solid ratio = 20:1.

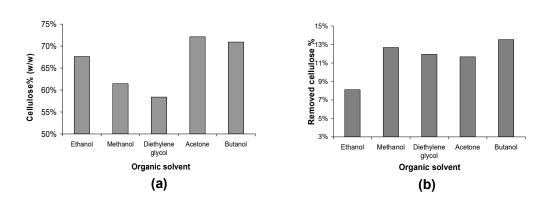


Fig.6: Effect of organic solvent on (a) cellulose composition and (b) removed cellulose percentage. Sulfuric acid 0.045N; 160°C; 20 min; liquid: solid ratio = 20:1.

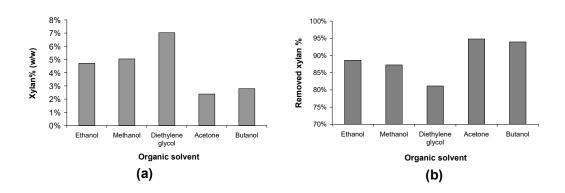


Fig.7: Effect of organic solvent on (a) xylan composition and (b) removed xylan percentage. H_2SO_4 0.045N; 160°C; 20 min; liquid: solid ratio = 20:1.

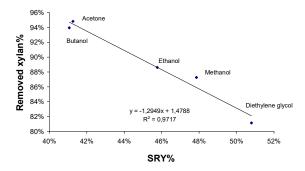


Fig.8: Removed xylan vs. Solid Residue Yield. Sulfuric acid 0.045N; temperature 160°C; time 20 min; liquid: solid ratio = 20:1.

In Fig. 7 acetone and butanol stand out among the other organic solvents due to their high xylan removal percentages 94.81% and 93.94% respectively. The correlation between removed xylan % and SRY% was also examined (Fig.8). In this case the linear regression gave also good results (R²=0.97) without exception. Unlike low glycose concentration before and after post hydrolysis, xylose concentration appears to be promising. As shown in Fig. 9 xylan of the initial straw in the case of acetone was hydrolyzed to 88% w/w xylose (expressed as % w/w on initial xylan).

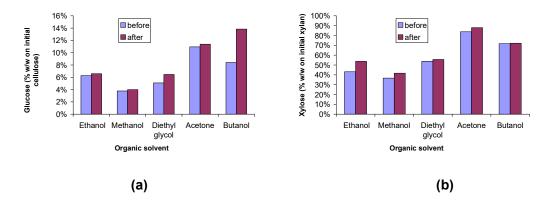


Fig.9: Effect of organic solvent on (a) glucose and (b) xylose concentration, before and after post-hydrolysis. Sulfuric acid 0.045N; temperature 160°C; time 20 min; liquid: solid ratio = 20:1.

CONCLUSIONS

Within the Biorefinery concept, from the five solvents (ethanol, methanol, diethylene glycol, acetone, butanol) examined herein as regards organosolv pretreatment (sulfuric acid 0.045 N, 160 °C, 20 min) of wheat straw, butanol gave the highest delignification effect, i.e., 63% lignin removal. Maximum cellulose concentration was 72% w/w (on pretreatment's solid residue) using acetone while delignification was 59% and 95% of xylan of the initial straw was hydrolyzed to 88% w/w xylose.

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REFERENCES

- 1. Rubin, E.M. 2008. Genomics of cellulosic biofuels. Nature 454, 841–845.H. Yu, Y. Xing, F. Lei, Z. Liu, Z. Liu, J. Jiang. Improvement of the enzymatic hydrolysis of furfural residues by pretreatment with combined green liquor and ethanol organosolv. Bioresource Technology, 167, 2014, 46-52
- 2. Fulton, L., Howes, T., Hardy, J., 2004. Biofuels for Transport: An International Perspective. OECD, International Energy Agency.
- 3. Zhang, Y.-H. P.; Lynd, L. R., 2004. Biotechnol. Bioeng., 88, 797–824
- 4. Bing, Y.; Wyman, C. 2008. Biofuels, Bioprod. Biorefin. 2, 26–40
- 5. De Wild, P.J., Huijgen, W.J.J., Heeres, H.J., 2012. Pyrolysis of wheat-straw derived organosolv lignin. J. Anal. Appl. Pyrolysis 93, 95–103.
- 6. Zhang K., Pei Z., Wang D., 2015. Organic solvent pretreatment of lignocellulosic biomass for biofuels and biochemicals: A review. Biosource Technology. In press.
- 7. Klason P.,1893 Bidrag till kannedomen om sammansattningen af granens ved samt de kemiska processerna vid framstallning af cellulosa darur. Teknisk Tidskrift, Afdelningen for Kemi och Metallurgi 23(2):17–22
- 8. Klason P., 1893. Framstallning af rent lignin ur granved och denna sednares kemiska sammansattning. Teknisk Tidskrift, Afdelningen for Kemi och Metallurgi 23(2):55–56W.J.J.
- 9. Pan X., Gilkes N., Kadla J., Pye K., Saka S., Gregg D., Ehara K., Xie D., Lam D., Saddler J., 2006. Bioconversion of hybrid poplar to ethanol and coproducts using an organosolv fractionation process: optimization of process yields. Biotechno.l Bioeng. 94:851–861
- 10. Kim DE, Pan XJ (2010) Preliminary study on converting hybrid poplar to high-value chemicals and lignin using organosolv ethanol process. Ind Eng Chem Res 49(23):12156–12163. doi:10.1021/le101671r
- 11. Arato C, Pye EK, Gjennestad G (2005) The lignol approach to biorefining of woody biomass to produce ethanol and chemicals. Appl. Biochem Biotech 121:871–882. doi:10.1385/ABAB:123:1-3:0871
- 12. Fernando E.F., Vallejos E.M., Area MC, 2010 Lignin recovery from spent liquors from ethanol-water fractionation of sugar cane bagasse. Cell Chem Technol 44(9):311–318
- 13. D. Sidiras, E. Koukios, 2004. Simulation of acid catalysed organosolv fractionation of wheat straw, Bioresource Technology, 94(1),1-98.
- 14. Araque E., Parra C, Freer J., Contreras D., Rodriguez J., Mendonca R., Baeza J., 2008 Evaluation of organosolv pretreatment for the conversion of Pinus radiata D. Don to ethanol. Enzyme Microb. Technol. 43(2):214–219. doi:10.1016/j.enzmictec.2007.08.006
- 15. Zhao, X., Cheng, K., Liu, D., 2009. Organosolv pretreatment of lignocellulosic biomass for enzymatic hydrolysis. Applied Microbiology and Biotechnology 82, 815–827.
- 16. Pan, X., Xie, D., Yu, R., Lam, D., Saddler, J., 2007. Pretreatment of lodgepole pine killed by mountain pine beetle using the ethanol organosolv

- process: fractionation and process optimization. Industrial and Engineering Chemistry Research 46, 2609–2617
- 17. Brosse N., Sannigrahi P., Ragauskas A., 2009. Pretreatment of Miscanthus x giganteus using the ethanol organosolv process for ethanol production. Ind. Eng. Chem. Res. 48(18):8328–8334
- 18. Mesa L, Gonzalez E, Ruiz E, Romero I, Cara C, Felissia F, Castro E (2010) Preliminary evaluation of organosolv pre-treatment of sugar cane bagasse for glucose production: application of 23 experimental design. Appl. Energ. 87(1):109–114. doi:10.1016/j.apenergy.2009.07.016
- 19. Teramoto Y., Lee S.H., Endo T., 2008. Pretreatment of woody and herbaceous biomass for enzymatic saccharification using sulfuric acid-free ethanol cooking. Bioresource Technology 99(18):8856–8863. doi:10.1016/j.biortech.2008.04.049
- 20. Lopez F., Perez A., Garcia JC., Feria M.J., Garcia M.M., Fernandez M., 2011. Cellulosic pulp from Leucaena diversifolia by soda-ethanol pulping process. Chem Eng J 166(1):22–29. doi:10.1016/j.cej.2010.08.039
- 21. Gilarranz M.A., Oliet M., Rodriguez F., Tijero J., 1999. Methanol-based pulping of Eucalyptus globulus. Can J. Chem. Eng 77(3):515–521. doi:10.1002/cjce.5450770312
- 22. Huijgen WJJ, Reith JH, den Uil H (2010) Pretreatment and fractionation of wheat straw by an acetone-based organosolv process. Ind Eng Chem Res 49(20):10132–10140. doi:10.1021/le101247w
- 23. Lam H.Q., Le Bigot Y., Delmas M., Avignon G., 2001. Formic acid pulping of rice straw. Ind Crop Prod 14(1):65–71
- 24. Jahan M.S., 2007 Formic acid pulping of bagasse. Bangladesh J Sci Ind Res 41(3):245–250
- Zhang M., Qi W., Liu R., Su R., Wu S., He Z., 2010. Fractionating lignocellulose by formic acid: characterization of major components. Biomass Bioenerg 34(4):525–532. doi:10.1016/j.biombioe.2009.12.018
- 26. Jahan M.S., Chowdhury DAN., Islam M.K., 2007. Atmospheric formic acid pulping and TCF bleaching of dhaincha (Sesbania aculeata), kash (Saccharum spontaneum) and banana stem (Musa cavendish). Ind Crop Prod 26(3):324–331. doi:10.1016/j.indcrop.2007.03.012
- 27. Pan XJ, Sano Y (2005) Fractionation of wheat straw by atmospheric acetic acid process. Bioresour. Technol. 96(11):12561263. doi:10.1016/j.biortech. 2004.10.018
- 28. Gan D.N., Xie Y.M., Aorigele Y.M., Wang P., Li S.L., Yang H.T., 2004. Acetic acid pulping of triploid clones of Polulus tomentosa Carr at atmospheric condition. Trans China Pulp Paper 19(1):15–18
- 29. Villaverde J.J., Ligero P., de Vega A., 2010. Formic and acetic acid as agents for a cleaner fractionation of Miscanthus x giganteus. J Clean Prod 18(4):395–401
- 30. Ligero P., Villauerde J.J., de Vega A., Bao M., 2008. Delignification of Eucalyptus globulus saplings in two organosolv systems (formic and acetic acid) preliminary analysis of dissolved lignins. Ind Crop Prod 27(1):110–117. doi:10.1016/j.indcrop.2007.08.008
- 31. Saeman J.F., J.F. Bubl, E.E. Harris Quantitative saccharification of wood and cellulose, Ind. Eng. Chem. Anal. Ed., 17 (1945), pp. 35–37.