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Automation of Assays: Improving Precision and Throughput

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Introduction

- Manual immunoassays, especially functional assays involving living organisms are inherently variable.
 - Assay variability is a consequence of the assay complexity and high sensitivity of live organisms to external factors.
 - Multiple external factors affecting the assay, such as variability of individual techniques between the analysts, are often hard to control.
 - Manual assays usually require extensive technical training of analysts.



Potential Benefits of Assay Automation

- Numerous potential benefits are considered when the possibility of automation in the laboratory are evaluated, including:
 - improvement of the quality and reliability of test results;
 - increased workplace ergonomics, employee productivity and safety;
 - enhancement of the assay efficiency/throughput;
 - standardization of lab techniques;
- Complete automation presents operational and financial challenges and is not always economically justifiable; partial automation of the process workflow may be beneficial.



Example 1. Automation of the Meningococcal Serum Bactericidal Assay (Mn SBA) for Serogroups A, C, Y and W135.



Principle of the Serum Bactericidal Assay









Step 2: Adding complement ()



Step 3: Complement-mediated killing (defined as reduction in colony counts)





Manual Mn SBA ACYW135 assay

- Classic Meningococcal Serum Bactericidal Assay.
 - Four individual manual SBAs are performed to measure bactericidal antibody titers to Neisseria meningitidis serogroup A, C, Y and W135.
 - The SBA is a complex assay involving multiple manual dilutions performed by analysts.
- Qualification studies were conducted to objectively evaluate the assay parameters and performance.
 - The statistical analysis revealed that total variability was higher than expected.
- Further analysis demonstrated that among the ruggedness factors, the variability associated with the analyst showed the highest impact on the assay.



Manual Mn SBA ACYW135 Assay Variability

Variance estimates (%RSD)				
Serogroup	Analyst to Analyst	Run within Analyst	Total	
А	91%	90%	175%	
С	15%	74%	100%	
W135	29%	39%	65%	
Y	19%	34%	74%	

- The analysts were producing substantially different titers
 - Within analyst variability changed from analyst to analyst.
 - High variability was observed for both assay controls and test samples.



Automation of the sample processing for the Mn SBA ACYW135: Hamilton Microlab STAR





Summary of The Mn SBA ACYW135 Automation Effort

- Assay scripts were developed and validated. Scripts were created based on the manual process described in the SOP.
- The control and sample processing was performed by Hamilton.
- The use of the sample master dilution plate enabled simultaneous testing of the four serogroups improving assay throughput.



Standardize Sample Processing: Sample Assay Plate Transfer



Deck is reloaded 1-2 times dependent on the number of Master plates.

PPD[°]

Improvement of Overall Assay Precision After the Implementation of the Automation Step, per Serogroup

	Total Assay Variability (%RSD)			
Serogroup	Before Automation	After Automation	Automation	
A	175%	41%		
С	100%	36%		
Y	65%	27%		
W135	74%	35%		







Example 2. Automation of Seasonal Influenza HAI



Principle of the Hemagglutination Inhibition Assay

Hemagglutination







Hemagglutination

Inhibition of hemagglutination





Red blood cells

Antiviral antibodies from serum

Viruses



Viruses neutralized and hemagglutination inhibited



Seasonal Flu HAI

- The HAI assay is used to test various seasonal strains of Influenza virus
- The assay procedure consists of two analyst serially titrating a sample replicate with a 2-fold serial dilution for up to 12 microtiter assay plates per analyst, per strain
- Variability is increased because two analysts are generating replicate plates (and independent results)

Analyst 1: Sample Replicate 1



Analyst 2: Sample Replicate 2



Seasonal FLU HAI: Sample Processing Applications

- Similar to the MnSBA sample processing standardization
- Hamilton automation methods were designed to facilitate the testing of multiple virus strains against the same sample dilution and reduce sample repeat rates





Seasonal Flu HAI: Replicate Repeat Rate Results



Improved Biofunctional Assay Variability Conclusions

- Utilizing automation for high throughput sample processing applications has improved two of our biofunctional assays by reducing:
 - Variability and sample retest rate for Mn SBA ACYW135
 - Variability and sample retest rate for Seasonal Flu
 HAI



Example 3. Multiplexing and Automation of PCR Assays.





Each of these steps can be automated



Nucleic Acid Extraction: QIAGEN BioRobot MDx (DNA) and BioMèrieux NucliSens easyMAG (RNA)

- The Qiagen BioRobot MDx Workstation
 - Fully automated
 - High-throughput
 - Accurate liquid handling
 - filtered tips
 - On-board vacuum processing
 - Locked down once program begins
- The NucliSens easyMAG
 - Automated system
 - Magnetic silica beads
 - On-board magnet
 - Requires little hands-on interaction during a run







Increased Efficiencies – DNA and RNA Extraction

- Manual DNA Extraction (QIAGEN Blood Kit)
 - 6 hours touch time = 384 samples

- Automated Extraction (Qiagen BioRobot MDx)
 - 3 hours touch time = **1152** samples

- QIAGEN Manual Method
 - 4.5 hours touch time = 72 samples
 - OR
- Boom Silica Manual Method
 - 5 hours touch time = 48 samples

- BioMèrieux NucliSens easyMAG
 - 3 hours touch time = **144** samples



Automation of Sample Processing by TECAN Freedom Evo

- The TECAN Freedom Evo
 - Automated liquid handling system
 - Highly accurate pipetting
 - Decreased cross-contamination
 - Can be customized
 - Different arm configurations
 - Integration of other lab equipment.



- The Molecular Testing Laboratory at PPD VBL utilizes two separate Freedom Evo configurations to maximize the sample preparation potential of our high throughout PCR process.
- TECAN Freedom Evo can process 50 assay plates/FTE/Day with 1.5-2 h of touch time, resulting in a throughput of
 - Up to 200 plates per day
 - Or approximately 6 times the throughput of a single analyst



Principle of a Singleplex PCR assay



Throughput calculation, Example for 90 samples:

- 1 plate
- 15 different strains/organisms
- 3 Open Reading Frames
- 45 Assay Plates
- 1.5 FTE



Principle of a Multiplex PCR assay



Throughput calculation Example for 90 samples:

- 1 plate
- 15 different strains/organisms
- 3 Open Reading Frames measured simultaneously
- 15 Assay Plates
- 0.5 FTE
- Note: Target genes may come from the same or a different organism.



Reaction Efficiency and Linearity of a Multiplexed PCR Assay



- Typical characteristics of the assay:
 - The assay is linear over 6 logarithms of concentration
 - Each target is amplified equivalently
 - Each target has similar sensitivity



Benefits of Automating and Multiplexing of the Assays

- Increases quality of the data compared to manual assays (reduces error rate, including sample dilutions, placement, cross-contamination, etc.)
- Increases assay throughput
- Increases the assay development flexibility
 - Combination of different targets (i.e. antigens) can be tested simultaneously
- Improves assay parameters
 - Simultaneous detection of different targets to enhance specificity and sensitivity of detection
 - Reduced variability









Considerations to be taken into account when automating an assay

- Why is automation required? What is the reason for automating a process? What is this going to achieve for the laboratory?
 - What is the testing volume?
 - What are the potential benefits (throughput, retest rate, precision)?
- How much automation is required? Is a fully automated system needed or a semi-automated system?
- What systems are available?



Potential drawbacks of automation

- Once that step is taken it is very hard to remove automation from a laboratory. It becomes a part of the validated process.
- Logistical issues. In partial automation (automation of individual steps) balancing the throughput of different steps and eliminating bottlenecks may be challenging.
- Re-training of personnel is needed.
- Significant upfront investment may be required.



Principle of the Serum Bactericidal Assay









Step 2: Adding complement ()



Step 3: Complement-mediated killing (defined as reduction in colony counts)





Variability: Summary for SBAs

		Variance Estimates (% RSD)					
	_	Before automation		After automation			
Sero group	Analyst	Run Within Analyst	Plate Within Run	Total	Run Within Analyst	Plate Within Run	Total
А	1	10%	0%	44%	0%	5%	31%
С		86%	19%	103%	18%	20%	44%
W		0%	18%	35%	2%	4%	19%
Y		14%	13%	34%	0%	16%	23%
Α	2	256%	20%	337%	49%	0%	59%
С		98%	29%	140%	17%	12%	33%
W		78%	9%	97%	0%	11%	20%
Y		41%	28%	83%	0%	0%	16%
Α	3	20%	0%	74%	2%	10%	20%
С		6%	19%	41%	9%	13%	29%
W		26%	13%	41%	44%	8%	52%
Y		38%	65%	145%	0%	0%	23%



Standardize Sample Processing: Sample Master Dilution Plates





Standardize Sample Processing: Sample Assay Plate Transfer



Deck is reloaded 1-2 times dependent on the number of Master plates.

PPD[°]

Improvement of Overall Assay Precision After the Implementation of the Automation step (Serogroups C and Y).









Improvement of Overall Assay Precision After the Implementation of the Automation Step



Serogroup A

Serotype W135



Applied Biosystem 3730xl DNA Analyzer-Sequencing

- The ABI 3730xl DNA Analyzer
 - Fully automated sequence analyzer
 - 16-plate stacker
 - Monitoring software
 - will alert an analyst of any errors





Increased Efficiencies - Sequencing

- GE MegaBACE 1000
 - Assay 1 = 3.0 hour run / 96-well plate
 - Plus 30 minutes touch time / plate
 - Max three 96-well plates / day
 - 288 samples
 - 1 FTE, 1.5 hours touch time
- 1.5 hours touch time = 288 samples
 - Assay 2 = 5.0 hour run / 96-well plate
 Plus 30 minutes touch time / plate
 - Max two 96-well plates / day
 - 192 samples
 - 1 FTE, 1.0 hours touch time
- 1.0 hours touch time = 192 samples

- ABI 3730xl
 - Weekday
 - Assay 1 and 2 = 2 hour run / 96-well plate
 - Miniscule amount of touch time to load
 run settings
 - Max 12 plates in 24 hours
 - 1, 152 samples
 - 1 FTE, 30 mins touch time
- 30 minutes touch time = 1, 152 samples
 - Friday Night
 - Max 16 plates / stacker
 - 1, 536 samples
 - 1 FTE, 30 mins touch time
- 30 minutes touch time = 1, 536 samples



Improvement of Analyst-related Variability After Implementation of the Automation Step

	Variability (%RSD) Attibuted to Analyst		
Serogroup	Before Automation	After Automation	
А	91%	5%	
С	15%	0%	
Y	29%	0%	
W135	19%	0%	

_	Fold-Difference in Titer Between Analysts		
Serogroup	Before Automation	After Automation	
А	5.7	1.4	
С	2.1	1.3	
Y	2.4	1.5	
W135	1.5	1.1	



Benefits of Automating Immunoassays

- A contemporary immunoassay used for clinical trials is typically a complex multistep process which is qualified and validated.
- Automation of the most labor intensive and manual steps of immunoassays often improves assay precision and reduces the possibility of costly human errors.
- Automated liquid handling technology should be incorporated in the assay workflow as early as possible, ideally, at the development stage.

