UNIVERSITY^{OF} BIRMINGHAM

Research at Birmingham

Optical control of a receptor-linked guanylyl cyclase using a photoswitchable peptidic hormone

Podewin, Tom; Broichhagen, Johannes; Frost, Christina; Groneberg, Dieter; Ast, Julia; Meyer-Berg, Helena; Fine, Nicholas; Friebe, Andreas; Zacharias, Martin; Hodson, David; Trauner, Dirk; Hoffmann-Röder, Anja

DOI:

10.1039/C6SC05044A

License

Creative Commons: Attribution (CC BY)

Document Version

Publisher's PDF, also known as Version of record

Citation for published version (Harvard):

Podewin, T, Broichhagen, J, Frost, C, Groneberg, D, Ast, J, Meyer-Berg, H, Fine, NHF, Friebe, A, Zacharias, M, Hodson, DJ, Trauner, D & Hoffmann-Röder, A 2017, 'Optical control of a receptor-linked guanylyl cyclase using a photoswitchable peptidic hormone', Chemical Science, vol. 8, no. 6, pp. 4644-4653. https://doi.org/10.1039/C6SC05044A

Link to publication on Research at Birmingham portal

General rights

Unless a licence is specified above, all rights (including copyright and moral rights) in this document are retained by the authors and/or the copyright holders. The express permission of the copyright holder must be obtained for any use of this material other than for purposes permitted by law.

- Users may freely distribute the URL that is used to identify this publication.
- Users may download and/or print one copy of the publication from the University of Birmingham research portal for the purpose of private study or non-commercial research.
- User may use extracts from the document in line with the concept of 'fair dealing' under the Copyright, Designs and Patents Act 1988 (?)
- Users may not further distribute the material nor use it for the purposes of commercial gain.

Where a licence is displayed above, please note the terms and conditions of the licence govern your use of this document.

When citing, please reference the published version.

Take down policy

While the University of Birmingham exercises care and attention in making items available there are rare occasions when an item has been uploaded in error or has been deemed to be commercially or otherwise sensitive.

If you believe that this is the case for this document, please contact UBIRA@lists.bham.ac.uk providing details and we will remove access to the work immediately and investigate.

Download date: 01. Feb. 2019

Chemical Science



EDGE ARTICLE

View Article Online
View Journal | View Issue



Cite this: Chem. Sci., 2017, 8, 4644

Optical control of a receptor-linked guanylyl cyclase using a photoswitchable peptidic hormone;

Tom Podewin, D‡8ª Johannes Broichhagen, D‡8ª Christina Frost, Db Dieter Groneberg, Dc Julia Ast, Dde Helena Meyer-Berg, Da Nicholas H. F. Fine, Dde Andreas Friebe, Dc Martin Zacharias, Db David J. Hodson, Dde Dirk Trauner D*a and Anja Hoffmann-Röder D*a

The optical control over biological function with small photoswitchable molecules has gathered significant attention in the last decade. Herein, we describe the design and synthesis of a small library of photoswitchable peptidomimetics based upon human atrial natriuretic peptide (ANP), in which the photochromic amino acid [3-(3-aminomethyl)phenylazo]phenylacetic acid (AMPP) is incorporated into the peptide backbone. The endogeneous hormone ANP signals *via* the natriuretic peptide receptor A (NPR-A) through raising intracellular cGMP concentrations, and is involved in blood pressure regulation and sodium homeostasis, as well as lipid metabolism and pancreatic function. The *cis*- and *trans*-isomers of one of our peptidomimetics, termed TOP271, exhibit a four-fold difference in NPR-A mediated cGMP synthesis *in vitro*. Despite this seemingly small difference, TOP271 enables large, optically-induced conformational changes *ex vivo* and transforms the NPR-A into an endogenous photoswitch. Thus, application of TOP271 allows the reversible generation of cGMP using light and remote control can be afforded over vasoactivity in explanted murine aortic rings, as well as pancreatic beta cell function in islets of Langerhans. This study demonstrates the broad applicability of TOP271 to enzyme-dependent signalling processes, extends the toolbox of photoswitchable molecules to all classes of transmembrane receptors and utilizes photopharmacology to deduce receptor activation on a molecular level.

Received 15th November 2016 Accepted 9th April 2017

DOI: 10.1039/c6sc05044a

rsc.li/chemical-science

Introduction

Controlling biological function with light has been achieved using two general approaches, *viz.* optogenetics¹ and photopharmacology.^{2,3} While the first relies on the genetic introduction

of light-responsive proteins, the latter describes the exogenous use of small photochromic molecules that interact with a specific target. The advantage of photopharmacology is the precise control of cell signalling through native receptors, without necessarily introducing foreign genes. While optogenetics has successfully targeted the receptor-linked enzyme (RLE) class,⁴ in particular receptor tyrosine kinases,^{5,6} photopharmacology has not kept pace. One reason is that RLE ligands are usually large peptides with few known small molecule activators, making it a challenge to find a suitable "azologable" pharmacophore. However, we and others recently reported the optical control of cell function with photoswitchable peptides,^{7,8} an approach that is highly applicable to RLEs.

Accordingly, we focused on the natriuretic peptide receptor A (NPR-A), with its endogenous agonist atrial natriuretic peptide (ANP), as a suitable target for RLE photocontrol (Fig. 1a). The physiological actions of ANP are widespread and range from blood pressure regulation and sodium homeostasis to effects on fat metabolism and pancreatic beta cell function/survival.⁹⁻¹¹ ANP is mainly expressed and stored as inactive proANP in atrial cardiac myocytes, with lesser concentrations found in the ventricles and kidneys. Upon secretion, primarily controlled by mechanical stimulation following atrial wall stretching, ^{12,13}

[&]quot;Department of Chemistry and Center for Integrated Protein Science, LMU Munich, Butenandtstr. 5-13, 81377 Munich, Germany. E-mail: anja.hoffmann-roeder@cup. lmu.de; dirk.trauner@cup.lmu.de

^bDepartment of Physics, Technical University of Munich, James-Franck-Str. 1, 85748 Garching, Germany

Julius-Maximilian-University Würzburg, Institute of Physiology, Röntgenring 9, 97070 Würzburg, Germany

^dInstitute of Metabolism and Systems Research (IMSR) and Centre of Membrane Proteins and Receptors (COMPARE), University of Birmingham, Edgbaston, B15 2TT, UK

^cCentre for Endocrinology, Diabetes and Metabolism, Birmingham Health Partners, Birmingham, B15 2TH, UK

 $[\]dagger$ Electronic supplementary information (ESI) available: Details on solid phase peptide synthesis and characterisation of all peptides can be found here, as well as experimental details on cGMP assays, aortic tensometry, islet treatment, statistics and a detailed description of modelling and simulations. See DOI: 10.1039/c6sc05044a

[‡] These authors contributed equally.

[§] Present address: Max Planck Institute for medical research (MPImF), Jahnstraβe 29, 69120 Heidelberg, Germany.

Edge Article

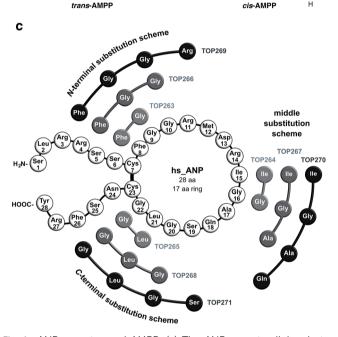


Fig. 1 ANP receptor and AMPP. (a) The ANP receptor links photoswitchable peptide binding to activation of guanylyl cyclase, that is cGMP generation and subsequent vasodilation in its UV-light adapted *cis*-state. (b) The photoswitch AMPP can be toggled between its *trans*- and *cis*-isomer with blue (460 nm) and UV-light (365 nm), respectively. (c) Initial assessment of AzoANP peptides TOP263-271 potency towards cGMP synthesis focuses on TOP271, where four amino acids are replaced on the C-terminal side of the macrocycle.

proANP is rapidly cleaved by the cardiac serine protease corin to release the active 28 amino acid ANP. The active form comprises a central 17 amino acid macrocycle linked by a disulfide bridge between Cys7 and Cys23. Following ligand activation of NPR-A, the membrane-proximal regions of the monomeric receptor units undergo a global conformational change, triggering guanylyl cyclase activity. This leads to generation of cGMP, a major player in intracellular cell signalling.

Dysregulated ANP secretion has been linked to different cardiovascular diseases, *i.e.* atrial fibrillation, ¹⁵ hypertension ^{16,17} and heart failure. ^{18,19} Moreover, genetic variants in or close to the ANP gene (NPPA) locus, which lead to increased circulating levels of plasma ANP, were shown to lower blood pressure and

the risk of hypertension in healthy individuals.^{20,21} Furthermore, individuals harbouring one copy of the G allele of rs5068 have lower likelihood of diabetes,²² and ANP has been shown to increase muscle insulin sensitivity,²³ although whether insulin release itself is stimulated is more debated.^{10,11,24} Such fundamental and pleiotropic actions of ANP have made its receptors an important pharmacological target, resulting in recently introduced therapies for the treatment of cardiovascular diseases.^{25,26} Despite this, many facets of ANP function and action remain elusive. Thus, the development of novel tools for unravelling and controlling ANP/NPR-A-stimulated signalling processes would be a valuable asset.

To address this, we report the synthesis of a photochromic ligand based on human ANP that enables the photocontrol of RLE activity (Fig. 1a). The NPR-A was endowed with lightsensitivity by incorporation of the photoswitchable amino acid [3-(3-aminomethyl)phenylazo]phenylacetic acid (AMPP)^{27,28} into ANP, which along with related derivatives, 8,29-31 has proven to be a valuable building block for photocontrol of peptide conformation and activity (Fig. 1b). One out of nine of our synthesised photochromic ANPs (AzoANPs), termed TOP271, allowed optical control to be exerted over NPR-A activity, intracellular cGMP levels, and downstream processes using UV and blue light.

Results and discussion

Our initial design approach was based on the incorporation of AMPP into the peptidic backbone of ANP, to induce maximal structural changes upon photoisomerisation. Nine different photochromic AzoANP peptides (dubbed TOP263-271) were designed and synthesised to obtain a small library (Fig. 1c), whereby AMPP replaced two, three or four amino acids in ANP. These numbers were based on our experience with other peptides, in which AMPP displaced two amino acids and the fact that AMPP covers up to four amino acids in length. Replaced that AMPP covers up to four amino acids in length. See The substitutions, following a circular permutational fashion, were located either near the N- or C-terminus or facing the Cys7-Cys23 disulfide bridge in the native 17 amino acid cycle of ANP (see ESI† for details on synthesis and characterization).

For the incorporation of azobenzenes into cyclic peptides several approaches have been developed in recent years. They range from synthesis and screening of small peptide libraries to combinatorial approaches utilising phage selection for the in vitro evolution of photoswitchable ligands.32-35 We selected a more rational design approach, in which the substitution sites were selected based on perceptions of ANP binding to NPR-A from mutational and structural studies.36-38 Upon ANP binding, the NPR-A receptor forms a homodimer, where the ligand is buried between the two extracellular domains of the respective monomers. The binding of ANP is asymmetric and one domain interacts mainly with the N-terminal, while the other interacts with the C-terminal part. Important binding interactions involve Phe8, Arg14 and Asn24 of ANP. Phe8 extends to and interacts with a hydrophobic binding pocket, which is critically important for hormonal activity. Arg14 forms hydrogen bonds with both monomers (Asp 62 and Glu119),

stabilizing the partially open dimer interface. The C-terminal

Chemical Science

located Asn24 forms two hydrogen bonds that are important for receptor binding and hormone activity.38

Thus, the substitution schemes either involved the direct replacement of the important binding residue, as in the case of N-terminal substitution, or located AMPP close to it, as for the C-terminal and middle substitution schemes. By substituting Phe8 with AMPP, it was envisaged that in these peptides the hydrophobic photoswitch would differentially engage the hydrophobic pocket in its cis- or trans-form, modifying receptor binding and hormone activity. We dismissed the design of peptides in which AMPP was placed adjacent to Phe8, i.e. through substitution of residues Gly9 - Met12, due to the possibility of aromatic stacking. In the peptides with the middle substitution scheme, AMPP was placed adjacent to Arg14, to control the hydrophilic interactions of this residue with the receptor. In the N-terminal substitution scheme, AMPP was placed in the ring alongside Cys23 and the disulfide bridge. In these peptides, isomerisation should not only shift the ring structure, but also alter the binding interactions of Asn24 to domain B of the dimerised NPR-A receptor.

At this point, it is worth noting that initial screening of cGMP accumulation returned a single compound, i.e. TOP271, as being the most active and most isomer-dependent peptide. Thus, we focused on characterisation and investigation of this lead compound and further details on the cGMP assays can be found under "optical control of cGMP generation". Although our design approach was restricted to nine peptides, with many possible patterns being omitted, it led to the isolation of the functional compound TOP271. This not only validates our rational design strategy, but also confirms the targeting of ANP/ NPR-A interactions for the control of receptor binding and activation.

Photophysical properties of AzoANP peptides TOP263-271

The photochromic AzoANP peptides including native human ANP (hsANP) were synthesised through solid-phase peptide synthesis (SPPS), characterized by high resolution mass spectrometry, and their purity assessed by reverse-phase HPLC: all were <3.7 ppm of the calculated mass and RP-HPLC revealed high purity (see ESI† for MS and HPLC data). The switching kinetics of all "azologued" peptides were determined by UV/Vis spectroscopy (TOP271 Fig. 2a-c, TOP263-270 ESI Fig. 1-3 and ESI Tables 1 and 2†). Starting with compounds in the darkadapted state (vide infra), a decrease in the π - π * and an increase in the $n-\pi^*$ band was observed in response to UV light $(\lambda = 365 \text{ nm})$ (Fig. 2a), with reversion of this switching process in response to blue light ($\lambda = 460 \text{ nm}$).

As expected for an electron-poor azobenzene, all peptides were bistable (TOP271 Fig. 2c, TOP263-270 ESI Fig. 3†). This advantageous trait allows samples to be pre-illuminated prior to application rather than needing constant illumination. We determined the thermal $cis \rightarrow trans$ relaxation rate k_{obs} in PBS buffer. This was performed at room temperature, and because the compounds were bistable, the initial back-relaxation was obtained as a linear function from the first 30 min after ceasing

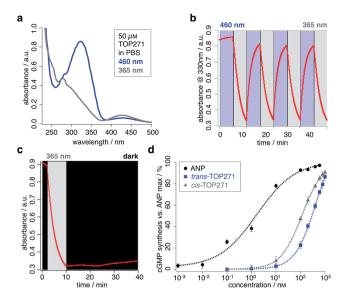


Fig. 2 TOP271 photodynamics and cGMP concentration-responses. (a) UV/Vis spectra of cis- and trans-TOP271. (b) Reversible switching of TOP271 with UV (365 nm) and blue (460 nm) light. (c) Bistability of TOP271 when switched to the cis-isomer by means of UV light (365 nm) and subsequently left in the dark. (d) Concentration-response curves of cis- and trans-TOP271 in NPR-A transfected HEK293T cells assessed by cGMP HTRF, 30 min incubation time. EC₅₀ values for ANP = 2.0 \pm 0.4 nM, cis-TOP271 = 127 \pm 11 nM and trans-TOP271 = 468 \pm 59 nM correspond to two independent assays. Values represent the mean \pm SEM.

illumination. Peptides TOP263-270 showed $k_{\rm obs} \times 10^{-4}$ a.u._{330 nm} min⁻¹ between 0.28-2.83, with the most potent compound TOP271 being 0.98 a.u. $_{330~\mathrm{nm}}$ min $^{-1}$ (see ESI Table 2,† compare with $k_{\rm obs} = 3.58 \times 10^{-4}$ a.u. min⁻¹ for unsubstituted azobenzene in benzene at 0 °C).39 To examine the structural relations between ANP and both isomers of TOP263-271, CD spectra were recorded in 40% buffered aqueous 2,2,2-trifluoroethanol (TFE) solutions. TFE is needed for the peptides to form stabilised secondary structures instead of random coils,40 and the optimal TFE concentration of 40% was determined with ANP in different aqueous buffered mixtures (ESI Fig. 4a†). The spectra of the peptides TOP263-271 showed differences between their cis/trans-isomers, but remained similar to that of native ANP, with no observable trend (ESI Fig. 4b and c†). TOP271 was subjected to further characterisation for both its cis- and trans-form by NMR spectroscopy alongside ANP. The spectra were recorded in 35% aqueous TFE-d₃ solutions to suppress signal broadening and aggregation (see ESI† for NMR data on ANP and cis/trans-TOP271). The cis-TOP271 NMR spectrum showed overlapping signals and thus could not be resolved. Nevertheless, the ¹H and ¹³C chemical shift values of the trans-TOP271 peptide could be unambiguously identified, showing the incorporation of AMPP into the backbone of the peptide and the overall correct structure.

Optical control of cGMP generation

To assess the most suitable peptide for further analysis, cGMP generation was measured in HEK293T cells transiently Edge Article Chemical Science

transfected with NPR-A.⁴¹ cGMP is a major effector of cellular metabolism,⁴² with effects on adipose tissue,⁴³⁻⁴⁵ liver^{46,47} and the brain.⁴⁸ Alongside nitric oxide, the natriuretic peptides are the major potentiators of cGMP generation, with downstream signalling effects on phosphodiesterases (PDEs),⁴⁹ cGMP-dependent proteinkinases (PKGs)⁵⁰ and cyclic nucleotide-gated channels (CNGs).⁵¹ ANP induces smooth muscle relaxation through increases in intracellular cGMP levels and activation of PKGI, which subsequently leads to a decrease in cytosolic Ca²⁺ levels and reduced Ca²⁺-sensitivity of the contractile system.^{12,52,53} Furthermore, depleting cGMP levels leads to depolarization in rods of the retina, triggering action potentials that transduce signals to perceive light.⁵⁴

To test cGMP synthesis, each peptide was applied as the trans- or cis-isomer by keeping them either in the dark or preilluminating with UV light ($\lambda = 365$ nm) for 15 minutes, respectively. Using this approach, TOP271, i.e. the ANP analogue where AMPP replaces four amino acids at the Cterminal end of the ring, was identified as the most promising candidate due to its highest binding affinity. In addition, a trend in the activity of these azobenzene-containing peptides was revealed: activity towards cGMP synthesis was higher the more amino acids were replaced and the closer their substitution was located to the N-terminus (Fig. 1c). Besides TOP271, only three further peptides from our small AzoANP library, TOP264, -265 and -268, showed NPR-A activation in the μM range (ESI Fig. 5†). The low potency of these compounds and the inactivity of the remaining photochromic ANP peptides (TOP263, -266, -267, -269 and -270) likely stems from the substitution of residues crucial for receptor binding, such as Phe8.55,56

With TOP271 as the lead candidate, we attempted to access light-dependent NPR-A activity. The measured cGMP concentration-responses showed that both TOP271 isomers had similar potency to native ANP (EC $_{50} = 2.0 \pm 0.4$ nM) (Fig. 2d), but *cis*-TOP271 (EC $_{50} = 127 \pm 11$ nM) was roughly four times more potent than the *trans*-isomer (EC $_{50} = 468 \pm 59$ nM). Since these EC $_{50}$ values correspond to the maximum incubation time of 30 minutes, we wanted to assess the time-dependency of potency and isomer-biased receptor activation. Therefore, we collected further data with shorter incubation times, which showed fluctuations for the ANP/TOP271 potency difference (ESI Fig. 6, ESI Table 3†). Nevertheless, the difference in potency between *cis*- and *trans*-TOP271 was robust and reproducible.

cGMP competition assays between ANP and cis/trans-TOP271 showed right-shifted EC $_{50}$ values for ANP, as expected when ligands compete for the same binding site (ESI Fig. 6†). Although cGMP end point assays do not provide direct evidence for ligand-mediated NPR-A over intracellular GC-A activation, this screening shows clear competition. Given the fact that ANP is well characterised to activate NPR-A, we assume that extracellular activation is key to cGMP generation. However, further studies are required to conclusively understand whether TOP271 directly activates GC-A, for instance using FRET-based reporters.

It should also be noted that, although the increase in EC₅₀ seems small, signal integration and amplification of cGMP

leads to more pronounced responses *in cellulo.*⁵⁷ With this in mind, we decided to progress TOP271 through to more relevant studies *ex vivo*.

Optical control of smooth muscle tone and pancreatic beta cell function

We next sought to address whether TOP271 would allow the optical control of cGMP-dependent processes in a physiologically relevant system, i.e. the aortic ring preparation. The treatment of constricted aortic rings with ANP leads to a potent vasodilation, corresponding to the blood pressure reducing effect of ANP.58 Accordingly, we predicted that TOP271 would allow reversible, light-controlled vasoactive responses, with cis-TOP271 being the stronger effector at specific concentrations. Concentration-response curves were obtained for vasodilation in pre-constricted aortic rings following exposure to preilluminated cis- ($\lambda = 365$ nm) and trans-TOP271 ($\lambda = 460$ nm), and showed increased potency for the former isomer in the 100 nM to 1 μM range (Fig. 3a, ESI Fig. 7†). Although this concentration-response indicated a significant difference in receptor activation for 100 nM TOP271, we decided to use 400 nM TOP271 to trigger a stronger isomer-dependent vasodilation. Thus, the application of dark-adapted trans-TOP271 led to strong vasodilation, which was enhanced following UV ($\lambda = 365$ nm) illumination to induce *cis*-isomerisation, and again reversed after blue light ($\lambda = 460 \text{ nm}$) exposure to induce trans-accumulation (Fig. 3b and d, ESI Fig. 8†).

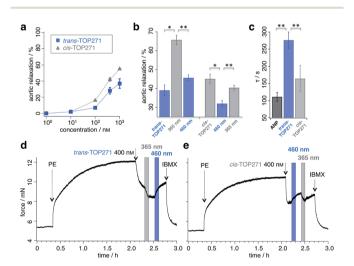


Fig. 3 TOP271 allows photocontrol of aortic tissue. (a) Concentration-response curves for trans- and cis-TOP271. (b) Aortic relaxation for cycles in (d) and (e). (c) τ of aortic relaxational responses for 17 nM ANP compared to 400 nM trans- and cis-TOP271. (d, e) Reversible photocontrol over aortic tension by application of trans-TOP271 (d) and cis-TOP271 (e) with UV-/blue and blue/UV-light (365 and 460 nm) cycles, respectively. All experiments were conducted in the presence of 200 μ M N- ω -nitro-L-arginine methyl ester (L-NAME) and 3 μ M diclofenac, which were used to block the vascular relaxants nitric oxide and prostacyclin, respectively. PE = phenylephrine was used for initial vasoconstriction, IBMX = 3-iso-butyl-1-methylxanthine was used to induce complete muscular relaxation as an end point control. (n = 6 aortic rings from 4 animals) (*P < 0.05, **P < 0.01, repeated measures Student's t-test). Values represent the mean \pm SEM.

Conversely, to examine the $cis \rightarrow trans \rightarrow cis$ isomerisation cycle, pre-illuminated ($\lambda = 365$ nm) cis-TOP271 was added to the organ bath, leading to a potent vasodilation (Fig. 3b and e). Subsequent trans-isomer accumulation by exposure to blue light ($\lambda = 460$ nm) elicited vasoconstriction, which again could be reversed by UV ($\lambda = 365$ nm) illumination. Notably, the speed of the initial vasodilation was $1.5\times$ higher for cis- compared to trans-TOP271, with the former being analogous to ANP (Fig. 3c). Although the experimental setting limited TOP271 switching to three iterations, isomerisation is robust and extended switching

Chemical Science

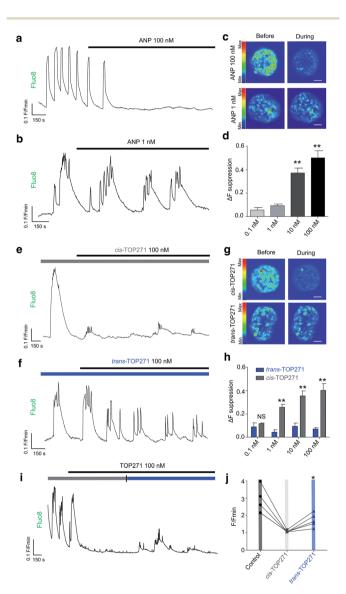


Fig. 4 TOP271 allows photocontrol of pancreatic beta cell function. (a–d) ANP concentration-dependently suppresses Ca²⁺ responses in beta cells (representative traces shown) (scale bar 25 μ M) (n=4-6 islets for each concentration) (**P<0.01, one-way ANOVA). (e–h) cis-TOP271 concentration-dependently inhibits Ca²⁺ spike amplitude, whereas trans-TOP271 is less effective at all concentrations tested (representative traces shown) (scale bar 25 μ M) (n=4-6 islets for each concentration) (**P<0.01, two-way ANOVA). (i and j) TOP271 allows reversible photocontrol of beta cell function, with blue (458–488 nm) light partially restoring Ca²⁺ responses (n=5 islets) (*P<0.05, repeated measures one-way ANOVA). Values represent the mean \pm SEM.

cycles are conceivable depending on the application, where degradation and clearance together with internalization and desensitization can play roles in repeated switching efficiency. Still, the multiple switching of TOP271 facilitates the implementation of this compound to induce desired vasoactive effects locally, within a spatially confined area. This clearly sets TOP271 apart from other agonists or inhibitors, which in general only allow for systemic application.

Insulin-secreting pancreatic beta cells express NPR-A, and ANP action is thought to provide a potential explanation for the association between cardiovascular and metabolic dysregulation. 59,60 We observed that native ANP concentration-dependently (0.1-100 nM) suppressed beta cell function at physiologicallyelevated glucose (8 mM) levels, as shown by a reduction in the amplitude of intracellular Ca2+ fluxes in intact islets of Langerhans (Fig. 4a-d). These findings could be replicated using UV preilluminated ($\lambda = 365$ nm) cis-TOP271 (Fig. 4e-h), which also robustly suppressed Ca²⁺ rises from 1-100 nM. By contrast, darkadapted trans-TOP271 induced only a small decrease in beta cell Ca²⁺ spiking activity (Fig. 4e-h). Reversibility could be achieved by applying *cis*-TOP271 and then illuminating with blue light (λ = 458-482 nm) to induce trans-isomerisation (Fig. 4i and j). Restoration of beta cell function was only partial (Fig. 4j), however, possibly due to cGMP-mediated sequestration of Ca²⁺ into internal stores such as the endoplasmic reticulum.61

Pancreatic beta cells have been shown to express NPR-A, and links exist between ANP and diabetes risk.⁶² Indeed, ANP gene expression is increased in the ventricles of rats with reduced beta cell mass, and ANP levels are elevated during diabetes.⁵⁹ The effects of ANP on beta cell function are complex and controversial. While some studies have shown that ANP decreases Ca2+ levels and insulin secretion, 10,24 others have shown stimulatory effects. 11,63 This likely reflects differences in the time course of application, preparation under examination (i.e. dissociated vs. intact islets), stimulation state (i.e. low vs. high glucose) and concentration. With regards to the latter, we were able to show a bimodal relationship where low doses of ANP preferentially affect Ca²⁺ oscillation frequency without altering amplitude, whereas high doses do the opposite (ESI Fig. 9†). Thus, TOP271 may provide an important tool to allow ANP receptor conformation and signalling to be understood in the context of beta and other cell (dys)function.

Molecular dynamics simulations of ANP and cis/trans-TOP271

Atomic-level modelling in explicit solvent was conducted to better understand the structure-activity relationships of isomer-dependent NPR-A activation. We focused on the *in silico* structure of native ANP peptide and *cis/trans*-TOP271, both in aqueous solution and bound to the NPR-A receptor. The extracellular domains were modelled based on the NPR-A crystal structure (PDB: 1t34, in complex with rat ANP (rnANP)),⁵⁵ while the receptor-bound ANP peptide and the *cis/trans*-TOP271 isomers where based on the NPR-C crystal structure (PDB: 1yk0, in complex with human ANP).⁵⁶ We used the latter for our structural peptide modelling to account for the Met12/Ile12 difference between ANP and rnANP, respectively.

Edge Article

native ring structure (Fig. 5b).

In a first step, the *cis/trans*-TOP271 isomers were simulated for 200 ns in the absence of the NPR-A receptor. To compare the affinity of both isomers to adopt the bound ANP ring structure, distance restraints with respect to the ANP crystal structure were applied between all C α -atom pairs within the ring, neglecting the two terminal tails. The resulting restraint energy distributions show a clear difference between the isomers (Fig. 5a), with the energy of *trans*-TOP271 being on average 6.7 kcal mol $^{-1}$ higher. This likely derives from the rigid, extended *trans*-azobenzene structure, which sterically prevents adoption of the

To elucidate the effect of the isomeric conformational differences on receptor geometry, unrestrained simulations of 1 µs length were performed for NPR-A bound to cis/trans-TOP271, ANP and apo-NPR-A. The overall receptor RMSD significantly differs for cis- and trans-TOP271, with the former matching ANP and the latter being similar to apo-NPR-A (Fig. 5c). These isomerdependent differences in receptor geometry are related to a change in relative orientation of the two NPR-A dimers: while there are no ligand-dependent orientation changes in the membrane-distal domains (ESI Fig. 10a†), the membraneproximal domains tend to close around ANP and cis-TOP271, but remain more open in the case of trans-TOP271 (Fig. 5d-g). This is in agreement with the crystal structures of the different apo- and ligand-bound natriuretic peptide receptors, which show up to 20 Å distance change between the two C-terminal/ membrane-proximal receptor domains upon ligand binding.^{55,56} trans-TOP271 thereby resembles the apo-form, in which fluctuating membrane-proximal distances shift the receptor towards an open state (Fig. 5c). These changes in receptor geometry can be assigned to isomeric differences in the bound conformation. Whereas the preferred conformations of *cis*-TOP271 are comparable to the crystal ring structure of ANP, the conformational ensemble of *trans*-TOP271 is narrower and more hairpin-like (Fig. 5e–g). The NPR-A-bound crystal structure of ANP also reveals a central pore in the 17 amino acid ring that is essential for ligand binding; only *cis*-TOP271 is able to adapt this donut-like conformation, while the *trans*-isomer forms a closed structure (Fig. 5e–g).

Lastly, we attempted to quantify the twist motion of the NPR-A membrane-proximal domains upon ligand binding, which is thought to initiate intracellular GC activation. Here, we detected a less prominent isomer dependency for the selected twist angle of the ligand-bound receptor domains compared to apo-NPR-A (see ESI Fig. 10b and ESI Table 3†). While the binding of ANP leads to a focusing of the twist angle distributions in NPR-A, bound *cis*- and *trans*-TOP271 show broader, shifted distributions and the apo-NPR-A inherits large angle fluctuations. This shows on the one hand the higher similarity of angle distributions of ANP and *cis*-TOP271 compared to *trans*-TOP271, but on the other hand also the flexibility of the apo-NPR-A membrane-proximal regions. Notably, crystal structures represent only a structural snapshot, while MD simulations cover a whole ensemble of structures and as a result we

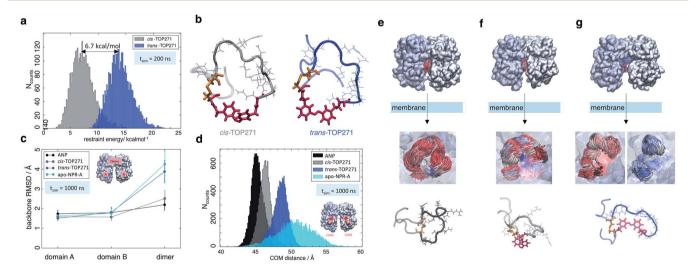


Fig. 5 MD simulations of ANP and TOP271. (a) Restraining energy distribution obtained from MD simulations (200 ns) of *cis*- and *trans*-TOP271 in solution, including restraints to keep the sampled conformations close to native ANP (see ESI†). (b) Representative restrained conformations of *cis*- and *trans*-TOP271 illustrating the steric hindrance of *trans*-azobenzene to fit into the native ring structure (red: azobenzene, orange: Cys7-Cys23 disulfide bridge). (c) Mean backbone-Cα RMSD and standard deviation of the NPR-A dimer and the both receptor domains A and B after 1 μs simulation, calculated for bound native ANP (black), bound *cis/trans*-TOP271 (gray and blue, respectively) and apo-NPR-A (cyan). The overall receptor RMSD clearly differs between native and *cis*-TOP271 *vs. trans*-TOP271 and apo-NPR-A. (d) Center-of-mass distance (orange balls and arrow) of the membrane-proximal NPR-A domains after 1 μs simulation. For bound native ANP (black) and *cis*-TOP271 (gray) the receptor domains tend to close compared to *trans*-TOP271 and the apo-NPR-A (blue and cyan). (e–g) Representative NPR-A and peptide conformations for bound native ANP (e) and bound *cis/trans*-TOP271 (f and g, respectively). Top row: isomer-dependent overall receptor geometry and binding site coverage visualised by time-superposition of the disulfide-connected isomer segment Cys7-Cys23 (red: ≤300 ns, white ≤600 ns, blue ≤1000 ns). Middle row: zoom into the time-superposition illustrates differences in the conformational ensemble of native ANP and *cis*-TOP271 *vs. trans*-TOP271 (azobenzene in line design). Bottom row: representative isomer conformations extracted from the simulated ensemble (red: azobenzene, orange: Cys7-Cys23 disulfide bridge).

conclude that the reduced distance between the membraneproximal domains in the ligand-bound state, and not the twisting motion of the NPR-A, is the major trigger for receptor activation.

In summary, our MD simulations showed a higher flexibility of the apo-NPR-A and the *trans*-TOP271-NPR-A receptor complex, whereas the membrane-proximal receptor domains tend to close around ANP and *cis*-TOP271. The simulations hence suggest an alternative regulation of guanylyl cyclase activity, in which the binding of ANP and *cis*-TOP271 to NPR-A suppresses dynamic fluctuations of the membrane-proximal domains of both receptor dimers, leading to defined ligand/receptor structures.

Conclusions

In this study, we present the design, synthesis, evaluation and application of TOP271, a peptidic hormone based on ANP with a photoresponsive azobenzene unit. Acting via the NPR-A receptor to generate cGMP, TOP271 allows the reversible photocontrol of contraction/dilation in aortic tissue, as well as Ca²⁺ oscillations in rodent islets of Langerhans. Although photodependent cGMP synthesis was described earlier, these approaches rely on the genetically-encoded photosensitive proteins EROS^{64,65} or BeCyclOps, ^{66,67} whereas TOP271 allows unprecedented photocontrolled cGMP synthesis in native tissue. EROS and BeCyclOps are based on bacterial flavin-containing photoreceptors and fungal rhodopsins, respectively, and can be used to induce penile erection in male rats or tactic behavior in C. elegans following illumination. A drawback of EROS is the residual cAMP activity, caused by the specific mutation of an adenylyl cyclase to an engineered guanylyl cyclase. BeCyclOps on the other hand utilizes native guanylyl cyclase activity and was shown to specifically engage only cGMP synthesis, but requires genetic introduction. Photochromic ligands like TOP271 avoid these issues and remain exogenously applied, thereby only targeting and activating the protein of interest. It should be noted that the stability of azobenzene compounds introduced ex vivo or in vivo has to be evaluated on a case-to-case basis as degradation of the diazene unit might occur depending on cell environment or azoreductase expression. While we have shown that this is not the case for polar compounds such as JB253 68 and JB558 69, careful evaluation has to be performed before any clinical applications can be envisioned.

With TOP271, we could selectively and reversibly manipulate the NPR-A/cGMP signalling pathway with high spatio-temporal precision. Interestingly, the 4-fold higher potency of *cis*-TOP271 for cGMP generation detected *in vitro* in transfected HEK293T cells is sufficient to trigger a more pronounced vasodilation *ex vivo* in aortic ring tissue. Although the exact intracellular cGMP concentration is an active source of research efforts, the changes observed are in agreement to prior findings, where small changes in the concentration of this second messenger provoke a significant amplification of downstream signals.⁷⁰ It also showcases the robustness and applicability of TOP271, which we believe will enable precise control of hemodynamic

processes, contributing to the dissection of vascular function in health and disease.

Moreover, TOP271 not only demonstrates the successful transformation of ANP into a photoswitchable peptide, but also extends the toolbox of photochromic ligands to all classes of transmembrane receptors. The incorporation of azobenzenes into peptides and proteins has been achieved in a multitude of systems in the last decade, i.e. in proteins of E. coli^{7,71} and in short peptides with specific secondary structures such as βsheet and β-hairpin motifs.^{27,29,30,72} Two major possibilities should be distinguished: (i) having an azobenzene as an amino acid residue and (ii) having an azoswitch in the peptide backbone. While the former has been used to gain optical control over binding affinity of transcription factor and cell adhesion molecules, 71,73 the latter was successfully applied to the optical control of muscle contraction31 and secondary structure formation.74 We envision our design herein, together with AzoChig28 and LirAzo,8 to be highly applicable to all kinds of peptides (e.g. neuropeptides, such as oxytocin, vasopressin, kisspeptin), as backbone substitution allows a larger conformational change upon isomerisation and therefore a larger change in affinity and/or efficacy. With recent synthetic efforts in mind,75 tetra-ortho-chloro-AMPP, exhibiting red-shifted switching wavelengths and high bistability, can be envisioned for the incorporation into target peptides. Such breadth already encompasses hairpin structures, α-helices and now macrocyclic structures, but can potentially be extended to antibodies, immunogens, peptidic hormones and receptors, where fine regulation of protein function by tertiary structure stabilization/ destabilization is necessary for function.76 Thus, the present findings set the stage for photochromic peptides to become a mainstay for optical control of biological processes using photopharmacology.

Methods/experimental section

Experimental procedures and chemical characterization can be found in the ESI.† Experimental protocols regarding live animals were approved by the University of Birmingham's Animal Welfare and Ethical Review Body (AWERB) and carried out in accordance with the Animals (Scientific Procedures) Act 1986 of the United Kingdom.

Acknowledgements

T. P. and C. F. were supported by the SFB749 project of the Deutsche Forschungsgemeinschaft and the Center for Integrated Protein Science Munich (CIPSM). J. B. was supported by a Studienstiftung des Deutschen Volkes PhD studentship. D. J. H. was supported by Diabetes UK R. D. Lawrence (12/0004431), EFSD/Novo Nordisk Rising Star and Birmingham Fellowships, as well as MRC Project (MR/N00275X/1), Imperial Confidence in Concept (ICiC), and Wellcome Trust Institutional Support Awards. This project has received funding from the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation programme (Starting Grant 715884 to D. J. H. and Advanced Grant 268795 to D. T.). A. H. R.

and M. Z. were supported by the SFB749 project of the Deutsche Forschungsgemeinschaft. We are very grateful to Dr Davor Pavlovic, Dr Fahima Syeda and Syeeda Nashitha Kabir for the planning and testing of Langendorff perfused heart experiments. We thank Dr Ruey-Bing Yang for providing the pCMV5_GC-A plasmid⁴¹ and Axel Schäfer for technical

Notes and references

Edge Article

assistance.

 \P All peptides are available for a cademic use from the Hoffmann-Röder lab upon request.

- 1 K. Deisseroth, Optogenetics, Nat. Methods, 2011, 8, 26-29.
- 2 W. A. Velema, W. Szymanski and B. L. Feringa, Photopharmacology: beyond proof of principle, *J. Am. Chem. Soc.*, 2014, **136**, 2178–2191.
- 3 J. Broichhagen, J. Frank and D. Trauner, A roadmap to success in photopharmacology, Acc. Chem. Res., 2015, 48, 1947–1960.
- 4 W. Costa, J. Liewald and A. Gottschalk, Photoactivated adenylyl cyclases as optogenetic modulators of neuronal activity, *Microarray Methods Protoc.*, 2014, **1148**, 161–175.
- 5 E. Reichhart, A. Ingles-Prieto and A. Tichy, A phytochrome sensory domain permits receptor activation by red light, *Angew. Chem., Int. Ed.*, 2016, **55**, 6339–6342.
- 6 M. Grusch, *et al.*, Spatio-temporally precise activation of engineered receptor tyrosine kinases by light, *EMBO J.*, 2014, 33, 1713–1726.
- 7 C. Hoppmann, V. Lacey and G. Louie, Genetically encoding photoswitchable click amino acids in *Escherichia coli* and mammalian cells, *Angew. Chem., Int. Ed. Engl.*, 2014, 53, 3932–3936.
- 8 J. Broichhagen, *et al.*, Optical control of insulin secretion using an incretin switch, *Angew. Chem., Int. Ed. Engl.*, 2015, 54, 15565–15569.
- 9 D.-R. Park, *et al.*, Arginine thiazolidine carboxylate stimulates insulin secretion through production of Ca²⁺-mobilizing second messengers NAADP and cADPR in pancreatic islets, *PLoS One*, 2015, **10**, 1–20.
- 10 B. Lee and S. G. Laychock, Atrial natriuretic peptide and cyclic nucleotides affect glucose-induced Ca^{2^+} responses in single pancreatic islet-cells correlation with $(\text{Ca}^{2^+} + \text{Mg}^{2^+})$ -ATPase activity, *Diabetes*, 1997, **46**, 1312–1318.
- 11 A. Ropero, *et al.*, The atrial natriuretic peptide and guanylyl cyclase-A system modulates pancreatic beta-cell function, *Endocrinology*, 2010, **151**, 3665–3674.
- 12 L. R. Potter, S. Abbey-Hosch and D. M. Dickey, Natriuretic peptides, their receptors, and cyclic guanosine monophosphate-dependent signaling functions, *Endocr. Rev.*, 2005, 27, 47–72.
- 13 B. Edwards, R. Zimmerman and T. Schwab, Atrial stretch, not pressure, is the principal determinant controlling the acute release of atrial natriuretic factor, *Circ. Res.*, 1988, 62, 191–195.
- 14 W. Yan, F. Wu, J. Morser and Q. Wu, Corin, a transmembrane cardiac serine protease, acts as a pro-

- atrial natriuretic peptide-converting enzyme, *Proc. Natl. Acad. Sci. U. S. A.*, 2000, **97**, 8525–8529.
- 15 C. Röcken, B. Peters, G. Juenemann and W. Saeger, Atrial amyloidosis an arrhythmogenic substrate for persistent atrial fibrillation, *Circulation*, 2002, **106**, 2091–2097.
- 16 S. John, J. Krege, P. Oliver and J. Hodgin, Genetic decreases in atrial natriuretic peptide and salt-sensitive hypertension, *Science*, 1995, **267**, 679–681.
- 17 M. Lopez, S. Wong, I. Kishimoto, S. Dubois and V. Mach, Salt-resistant hypertension in mice lacking the guanylyl cyclase-A receptor for atrial natriuretic peptide, *Nature*, 1995, 378, 65–68.
- 18 J. Burnett, P. Kao, D. Hu and D. Heser, Atrial natriuretic peptide elevation in congestive heart failure in the human, *Science*, 1986, 231, 1145–1147.
- 19 M. Cowie, A. Struthers, D. Wood and A. Coats, Value of natriuretic peptides in assessment of patients with possible new heart failure in primary care, *Lancet*, 1997, 350, 1347–1351.
- 20 C. Newton-Cheh, *et al.*, Association of common variants in NPPA and NPPB with circulating natriuretic peptides and blood pressure, *Nat. Genet.*, 2009, **41**, 348–353.
- 21 V. Cannone, *et al.*, A genetic variant of the atrial natriuretic peptide gene is associated with cardiometabolic protection in the general community, *J. Am. Coll. Cardiol.*, 2011, 58, 629–636.
- 22 A. Jujić, *et al.*, Atrial Natriuretic Peptide and Type 2 Diabetes Development – Biomarker and Genotype Association Study, *PLoS One*, 2014, **9**, e89201.
- 23 M. Coué, *et al.*, Defective Natriuretic Peptide Receptor Signaling in Skeletal Muscle Links Obesity to Type 2 Diabetes, *Diabetes*, 2015, **64**, 4033–4045.
- 24 H. You and S. G. Laychock, Atrial natriuretic peptide promotes pancreatic islet beta-cell growth and Akt/Foxo1a/cyclin D2 signaling, *Endocrinology*, 2009, **150**, 5455–5465.
- 25 J. McMurray, M. Packer and A. Desai, Angiotensinneprilysin inhibition *versus* enalapril in heart failure, *N. Engl. J. Med.*, 2014, 371, 993–1004.
- 26 Y. Saito, Roles of atrial natriuretic peptide and its therapeutic use, *J. Cardiol.*, 2010, **56**, 262–270.
- 27 A. Aemissegger, V. Kräutler, W. F. van Gunsteren and D. Hilvert, A photoinducible beta-hairpin, *J. Am. Chem. Soc.*, 2006, **127**, 2929–2936.
- 28 T. Podewin, *et al.*, Photocontrolled chignolin-derived β-hairpin peptidomimetics, *Chem. Commun.*, 2015, **51**, 4001–4004.
- 29 C. Renner and L. Moroder, Azobenzene as conformational switch in model peptides, *Chem. Bio. Chem.*, 2006, 7, 868– 878.
- 30 T. E. Schrader, *et al.*, Light-triggered beta-hairpin folding and unfolding, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 15729–15734.
- 31 C. Hoppmann, et al., Photocontrol of contracting muscle fibers, Angew. Chem., Int. Ed. Engl., 2011, 50, 7699–7702.
- 32 S. Bellotto, S. Chen, I. Rebollo, H. Wegner and C. Heinis, Phage Selection of Photoswitchable Peptide Ligands, *J. Am. Chem. Soc.*, 2014, **136**, 5880–5883.

33 K. Rück-Braun, *et al.*, Azobenzene-Based Amino Acids and Related Building Blocks: Synthesis, Properties, and Application in Peptide Chemistry, *Synthesis*, 2009, 24, 4256–4267.

Chemical Science

- 34 C. Hoppmann, *et al.*, Light-Directed Protein Binding of a Biologically Relevant β-Sheet, *Angew. Chem., Int. Ed. Engl.*, 2009, **48**, 6636–6639.
- 35 C. Renner, U. Kusebauch, M. Löweneck, A. G. Milbradt and L. Moroder, Azobenzene as photoresponsive conformational switch in cyclic peptides, *J. Pept. Res.*, 2005, **65**, 4–14.
- 36 H. Ogawa, Y. Qiu, C. M. Ogata and K. S. Misono, Crystal Structure of Hormone-bound Atrial Natriuretic Peptide Receptor Extracellular Domain, *J. Biol. Chem.*, 2004, 279, 28625–28631.
- 37 H. Ogawa, *et al.*, Structure of the atrial natriuretic peptide receptor extracellular domain in the unbound and hormone-bound states by single-particle electron microscopy, *FEBS J.*, 2009, **276**, 1347–1355.
- 38 P. R. Bovy, Structure activity in the atrial natriuretic peptide (ANP) family, *Med. Res. Rev.*, 1990, **10**, 115–142.
- 39 E. Talaty and J. Fargo, Thermal *cis-trans*-isomerization of substituted azobenzenes: a correction of the literature, *Chem. Commun.*, 1967, 65–66.
- 40 M. Mimeault, A. Lean, M. Lafleur, D. Bonenfant and A. Fournier, Evaluation of conformational and binding characteristics of various natriuretic peptides and related analogs, *Biochemistry*, 1995, 34, 955–964.
- 41 Y.-C. Chao, *et al.*, Guanylate cyclase-G, expressed in the Grueneberg ganglion olfactory subsystem, is activated by bicarbonate, *Biochem. J.*, 2010, **432**, 267–273.
- 42 A. Pfeifer, A. Kilić and L. Hoffmann, Regulation of metabolism by cGMP, *Pharmacol. Ther.*, 2013, **140**, 81–91.
- 43 E. Nisoli, E. Clementi and C. Tonello, Effects of nitric oxide on proliferation and differentiation of rat brown adipocytes in primary cultures, *Br. J. Pharmacol.*, 1998, **125**, 888–894.
- 44 B. Haas, P. Mayer, K. Jennissen, D. Scholz and M. Diaz, Protein kinase G controls brown fat cell differentiation and mitochondrial biogenesis, *Sci. Signaling*, 2009, **99**, 1–12.
- 45 M. Bordicchia, et al., Cardiac natriuretic peptides act via p38 MAPK to induce the brown fat thermogenic program in mouse and human adipocytes, J. Clin. Invest., 2012, 122, 1022–1036.
- 46 A. Kiemer, N. Weber, R. Fürst and N. Bildner, Inhibition of p38 MAPK Activation *via* Induction of MKP-1 Atrial Natriuretic Peptide Reduces TNF-α-Induced Actin Polymerization and Endothelial Permeability, *Circ. Res.*, 2002, **90**, 874–881.
- 47 C. Moro and M. Lafontan, Natriuretic peptides and cGMP signaling control of energy homeostasis, *Am. J. Physiol.: Heart Circ. Physiol.*, 2013, **304**, 358–368.
- 48 T. Kleppisch and R. Feil, *cGMP Signalling in the Mammalian Brain: Role in Synaptic Plasticity and Behaviour*, Springer, 2009, vol. 191, pp. 549–579.
- 49 S. H. Francis, M. A. Blount and J. D. Corbin, Mammalian Cyclic Nucleotide Phosphodiesterases: Molecular Mechanisms and Physiological Functions, *Physiol. Rev.*, 2011, 91, 651–690.

- 50 R. Feil, S. Lohmann, H. de Jonge and U. Walter, Cyclic GMP-dependent protein kinases and the cardiovascular system insights from genetically modified mice, *Circ. Res.*, 2003, 93, 907–916.
- 51 M. Biel and S. Michalakis, *Cyclic Nucleotide-Gated Channels*, Springer, 2009, vol. 191, pp. 111–136.
- 52 A. Alioua, Y. Tanaka, M. Wallner and F. Hofmann, The large conductance, voltage-dependent, and calcium-sensitive K⁺ channel, Hslo, is a target of cGMP-dependent protein kinase phosphorylation in vivo, *J. Biol. Chem.*, 1998, 273, 32950–32956.
- 53 J. Schlossmann, A. Ammendola, K. Ashman and X. Zong, Regulation of intracellular calcium by a signalling complex of IRAG, IP3 receptor and cGMP kinase Iβ, *Nature*, 2000, **404**, 197–201.
- 54 K. Koch and D. Dell'Orco, Protein and signaling networks in vertebrate photoreceptor cells, *Front. Mol. Neurosci.*, 2015, **8**, 67–81.
- 55 H. Ogawa, Y. Qiu, C. M. Ogata and K. S. Misono, Crystal structure of hormone-bound atrial natriuretic peptide receptor extracellular domain rotation mechanism for transmembrane signal transduction, *J. Biol. Chem.*, 2004, 279, 28625–28631.
- 56 X. L. He, A. Dukkipati and K. C. Garcia, Structural determinants of natriuretic peptide receptor specificity and degeneracy, *J. Mol. Biol.*, 2006, **361**, 698–714.
- 57 F. Marks, U. Klingmueller and K. Mueller-Decker, *Cellular signal processing*, Taylor & Francis, 2009, vol. 8.
- 58 C. M. Panayiotou, *et al.*, Resistance to endotoxic shock in mice lacking natriuretic peptide receptor A, *Br. J. Pharmacol.*, 2010, **160**, 2045–2054.
- 59 F. Ortola, B. Ballermann and S. Anderson, Elevated plasma atrial natriuretic peptide levels in diabetic rats. Potential mediator of hyperfiltration, *J. Clin. Invest.*, 1987, **80**, 670–674.
- 60 H. Matsubara, Y. Mori, J. Yamamoto and M. Inada, Diabetesinduced alterations in atrial natriuretic peptide gene expression in Wistar-Kyoto and spontaneously hypertensive rats, *Circ. Res.*, 1990, 67, 803–813.
- 61 M.-L. Lazo-de-la-Vega-Monroy and A. Vilches-Flores, The Role of NO-cGMP Signaling Pathway in Pancreatic Beta-cell Function, *Immunol., Endocr. Metab. Agents Med. Chem.*, 2014, 14, 8–14.
- 62 G. Gruden, A. Landi and G. Bruno, Natriuretic peptides, heart, and adipose tissue: New findings and future developments for diabetes research, *Diabetes Care*, 2014, 37, 2899–2908.
- 63 N. Matsuura, *et al.*, Nitric oxide-cyclic GMP system potentiates glucose-induced rise in cytosolic Ca²⁺ concentration in rat pancreatic beta-cells, *Life Sci.*, 1999, 65, 1515–1522.
- 64 M.-H. H. Ryu, O. V. Moskvin, J. Siltberg-Liberles and M. Gomelsky, Natural and engineered photoactivated nucleotidyl cyclases for optogenetic applications, *J. Biol. Chem.*, 2010, 285, 41501–41508.
- 65 T. Kim, M. Folcher, M. D.-E. Baba and M. Fussenegger, A synthetic erectile optogenetic stimulator enabling blue-

Edge Article

light-inducible penile erection, *Angew. Chem., Int. Ed. Engl.*, 2015, **54**, 5933–5938.

- 66 G. M. Avelar, et al., A rhodopsin-guanylyl cyclase gene fusion functions in visual perception in a fungus, Curr. Biol., 2014, 24, 1234–1240.
- 67 S. Gao, *et al.*, Optogenetic manipulation of cGMP in cells and animals by the tightly light-regulated guanylyl-cyclase opsin CyclOp, *Nat. Commun.*, 2015, **6**, 8046–8058.
- 68 Z. B. Mehta, *et al.*, Remote control of glucose homeostasis *in vivo* using photopharmacology, *Sci. Rep.*, 2017, 7, 291.
- 69 J. Broichhagen, *et al.*, A red-shifted photochromic sulfonylurea for the remote control of pancreatic beta cell function, *Chem. Commun.*, 2015, **51**, 6018–6021.
- 70 S. H. Francis, J. L. Busch, J. D. Corbin and D. Sibley, cGMP-dependent protein kinases and cGMP phosphodiesterases in nitric oxide and cGMP action, *Pharmacol. Rev.*, 2010, 62, 525–563.
- 71 M. Bose, D. Groff, J. Xie, E. Brustad and P. G. Schultz, The incorporation of a photoisomerizable amino acid into proteins in E. coli, *J. Am. Chem. Soc.*, 2006, **128**, 388–389.

- 72 T. Schrader, *et al.*, Folding and Unfolding of Light-Triggered beta-Hairpin Model Peptides, *J. Phys. Chem. B*, 2011, **115**, 5219–5226.
- 73 C. Hoppmann, I. Maslennikov, S. Choe and L. Wang, *In Situ* Formation of an Azo Bridge on Proteins Controllable by Visible Light, *J. Am. Chem. Soc.*, 2015, 137, 11218–11221.
- 74 S. Samanta, C. Qin, A. J. Lough and G. A. Woolley, Bidirectional Photocontrol of Peptide Conformation with a Bridged Azobenzene Derivative, *Angew. Chem., Int. Ed. Engl.*, 2012, 51, 6452–6455.
- 75 D. B. Konrad, J. A. Frank and D. Trauner, Synthesis of Redshifted Azobenzene Photoswitches by Late-Stage Functionalization, *Chem.–Eur. J.*, 2016, 22, 4364–4368.
- 76 A. V. Karginov, F. Ding, P. Kota, N. V. Dokholyan and K. M. Hahn, Engineered allosteric activation of kinases in living cells, *Nat. Biotechnol.*, 2010, 28, 743–747.